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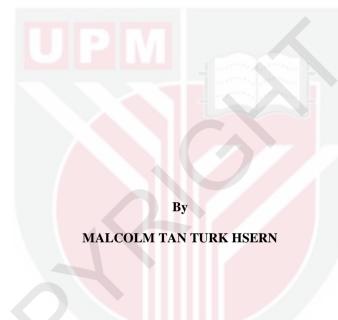
SURVEILLANCE OF VIBRIO PARAHAEMOLYTICUS IN SHELLFISH PURCHASED FROM SELECTED RETAILERS IN SELANGOR, MALAYSIA

MALCOLM TAN TURK HSERN

FSTM 2015 8



SURVEILLANCE OF *VIBRIO PARAHAEMOLYTICUS* IN SHELLFISH PURCHASED FROM SELECTED RETAILERS IN SELANGOR, MALAYSIA



Thesis submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfillment of the Requirements for the Degree of Master of Science

September 2015

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

SURVEILLANCE OF *VIBRIO PARAHAEMOLYTICUS* IN SHELLFISH PURCHASED FROM SELECTED RETAILERS IN SELANGOR, MALAYSIA

By

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September 2015

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The aims of this study were to optimize the multiplex polymerase chain reaction (mPCR) assay for the detection of *V. parahaemolyticus* and the respective pathogenic strains (tdh+ and/or trh+), and to compare with the loop-mediated isothermal amplification (LAMP) assay. These assays were not limited to detection only, and likewise coupled with the most probable number (MPN) to quantitate the bacterium in various shellfish samples obtained from wet markets and hypermarkets. The surveillance data were later adopted into a stochastic microbial risk assessment (MRA) model to evaluate the public health risk. In addition to these, a kitchen simulation was conducted to provide cross-contamination and decontamination data.

A total of 232 samples comprising of bloody clams, surf clams and shrimps were randomly purchased from the wet markets and hypermarkets in Selangor. 229 (98.7%) of the samples were positive for *V. parahaemolyticus* with counts ranging from 30 to >110, 000 MPN/g. Positive samples for tdh + V. *parahaemolyticus* were obtained in 77 out of 232 (33.1%) samples ranging from 30 to >110, 000 MPN/g. Meanwhile, positive samples for trh+ were identified in 16 out of 232 (6.9%) samples examined ranging from 30 to 9,600 MPN/g. In addition, we found that LAMP was rather more robust and sensitive compared to the multiplex PCR.

Bloody clam was selectively picked for a thorough MRA as the pathogenic strains of *V. parahaemolyticus* were prevalent in this sample. The study framework was established based on the currently available experimental and survey data in combination with the @RISK software to simulate the uncertainties based on the Monte Carlo simulation. The estimated risk was valued as 1.04E-4/daily serving/person. This translates to an estimate of 250 to 197,000 cases yearly.

The simulation of the handling of bloody clams in domestic kitchens was designed to imitate real events in domestic kitchens as much as possible to give a realistic quantitative data on how *V. parahaemolyticus* could cross-contaminate to other ready-to-eat (RTE) foods (eg, lettuce). It was found that 20.40 \pm 13.78% of the total *V. parahaemolyticus* population from the bloody clams were transferred to the hands and kitchen utensils through primary cross-contamination, and the cut lettuce had an average of 7432.5 MPN/g of *V. parahaemolyticus*. The attempted cleanings reduced the transferred population by 97.63 \pm 3.43% (water) and 96.00 \pm 5.03% (cloth), and the cut

lettuce had an average of 9.27 MPN/g of *V. parahaemolyticus*. Likewise, the bacterial transfer was minimal for secondary cross-contamination, which recorded 46.2 MPN/g (from contaminated water to lettuce) and 0.75 MPN/g (from contaminated plate to lettuce).

In conclusions, there is an immediate need for further investigation to look into the widespread of *V. parahaemolyticus* in Malaysia. Continued research, risk assessment and surveillance on the behaviour and characteristics of *V. parahaemolyticus* is very important in order to control, prevent and reduce the emerging problems caused by this interesting bacterium.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

PEMANTAUAN VIBRIO PARAHAEMOLYTICUS DALAM MAKANAN LAUT BERCENGKERANG YANG DIBELI DARI RUNCIT TERPILIH DI SELANGOR, MALAYSIA

Oleh

MALCOLM TAN TURK HSERN

September 2015

Pengerusi Fakulti : : Profesor Son Radu, PhD Sains dan Teknologi Makanan

Tujuan kajian ini adalah untuk mengoptimumkan assay reaksi polimerasi berantai multiplex (mPCR) untuk mengesan kehadiran *V. parahaemolyticus* dan strain patogenik (tdh+ dan/atau trh+) masing-masing, dan dibandingkan dengan assay amplifikasi bermediasi-gelung sesuhu (LAMP). Assay-assay ini turut dikombinasi dengan kaedah jumlah paling mungkin (MPN) untuk mengira kepekatan bakteria daripada pelbagai sampel makanan laut bercengkerang yang diperolehi dari pasar borong dan pasar raya besar. Data yang diperolehi sejurusnya diaplifikasikan dalam model penilaian risiko mikrob (MRA) untuk menilai risiko kesihatan umum. Tambahan pula, satu kajian simulasi dapur turut dijalankan untuk medapatkan data kontiminasi bersilang dan dekontaminasi.

Sejumlah 232 sample iaitu kerang, lala dan udang dibeli secara rawak daripada pasar borong dan pasar raya besar di Selangor. Sejumlah daripada 229 (98.7%) sample yang diuji adalah positif dengan *V. parahaemolyticus* berkepekatan dari 30 hingga >110, 000 MPN/g. Tujuh puluh tujuh dari 232 (33.1%) sampel yang diuji didapati positif dengan tdh+V. parahaemolyticus berkepekatan dari 30 hingga >110, 000 MPN/g. Manakala, 16 daripada 232 (6.9%) sampel yang diuji positif dengan trh+ berkepekatan dari 30 hingga 9,600 MPN/g. Di samping itu, kami mendapati bahawa LAMP adalah lebih robust dan sensitif daripada multiplex PCR.

Kerang dipilih secara selektif untuk MRA disebabkan prevalen strain *V. parahaemolyticus* yang patogenik adalah sangat tinggi dalam sample ini. Kajian ini berasakan experimentasi dan data survey semasa yang dikombinasi dengan @RISK software untuk mengsimulasikan kerawakan dengan mengunakan simulasi Monte Carlo. Estimasi berisiko divaluekan pada 1.04E-4/suapan sehari/seorang. Ini turut boleh ditranslasikan kepada 54 hingga 26,300 kes setahun.

Simulasi mengendalikan kerang dimodalkan berasakan event sebenar di tahap dapur untuk mendapatkan data quantifikasi *V. parahaemolyticus* kontaminasi bersilang kepada makanan yang sedia untuk dimakan (RTE; contohnya, sayur salad). Hasil daripada kajian ini 20.40 \pm 13.78% daripada jumlah populasi *V. parahaemolyticus* yang dipinda kepada tangan dan peralatan dapur, dan sayur salad mempunyai kepekatan *V.*

 \bigcirc

parahaemolyticus sebanyak 7432.5 MPN/g. Dekontaminasi berjaya mengurangkan populasi *V. parahaemolyticus* sebanyak 97.63 \pm 3.43% (air) and 96.00 \pm 5.03% (kain), dan sayur salad hanya mempunyai kepekatan *V. parahaemolyticus* sebanyak 9.27 MPN/g. Di samping itu, bakteria yang dipinda daripada konteminasi bersilang sekunder adalah minimal iaitu sebanyak 46.2 MPN/g (dari air kepada sayur salad) and 0.75 MPN/g (dari pinggan kepada sayur salad).

Konklusinya, siasatan lanjutan diperlukan segara untuk meninjau perubahan semasa V. *parahaemolyticus* di Malaysia. Hasil kajian berterusan, penilaian berisiko, peninjauan semasa dan penilaian karakter V. *parahaemolyticus* adalah sangat penting untuk megawal, mengelak dan mengurangkan masalah yang diakibatkan oleh bakteria oleh bakteria ini.



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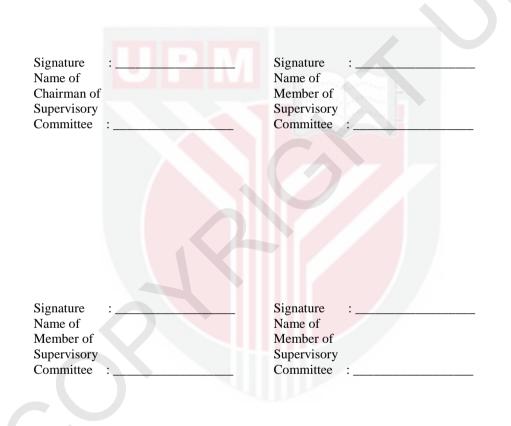


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LIST OF ABBREVIATIONS

μl	_	Microliter
ALOP	_	Appropriate Level of Protection
APW	-	Alkaline Peptone Water
ATCC	-	American Type Culture Collection
BAM	-	Bacteriological Analytical Manual
bp	_	base pair
CDC	_	Centre for Disease Control
Codex	_	Codex Alimentarius Commission
DNA	-	Deoxynucleic acid
dNTP	-	deoxyribonucleoside triphospate
EDTA	-	Ethylenediaminetetraacetic acid
FAO	-	Food and Agricultural Organization
FDA		Food and Drug Administration
FSO		Food Safety Objectives
GHP		Good Hygiene Practices
HACCP	-	
		Hazard Analysis Critical Control Point
ICMSF	-	International Commission on Microbiological Specifications
		for Food
LAMP	-	Loop-mediated isothermal amplification
MgCl ₂	-	Magnesium Chloride
mM	-	milliMolar
MOH	-	Ministry of Health
MPN	-	Most Probable Number
MRA	-	Microbial Risk Assessment
NaCl	-	Sodium Chloride
PCR/mPCR	-	Polymerase Chain Reaction/multiplex Polymerase Chain
Reaction		
RTE	-	Ready-to-eat
RPM	-	revolution per minute
SOP	-	Standard Operating Procedures
TBE	-	Tris-Boric acid-EDTA
TSA	-	Tryptic Soy Agar
U	-	Unit
U. S	-	United States
UV	-	Ultra violet
WHO	-	World Health Organization
WTO	-	World Trade Organization
α	-	Alpha
β	-	Beta

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CHAPTER 1

GENERAL INTRODUCTION

1.1 Introduction

Seafood is a nutritious food consumed all over the world. As a prime source of highquality protein, oil-rich and high in mineral, seafood is highly desirable in a healthy diet. Living in the marine environment, these sea creatures are constantly exposed to various pathogens which are detrimental to the publics' health. Thus, prior to the consumption of raw or undercooked seafood, one's life may be at risk. Jacxsens *et al.* (2009) have also reported that consumption of contaminated seafood has led to numerous foodborne outbreaks. Today, *Vibrio parahaemolyticus* is the leading causative pathogen often due to the consumption of raw or undercooked seafood, in which patients were reported to experience severe diarrhea, abdominal cramps, nausea, vomiting, headaches, fever and chills (Shimohata & Takahashi, 2010).

V. parahaemolyticus infection, nevertheless, is common in many Asian countries, inclining to their high seafood diets (Alam *et al.*, 2002). For instance, Japan, Taiwan, China, Korea and India are among the major countries that are struggling with this foodborne pathogen (Deepanjali *et al.*, 2005, Liu & Su, 2007 & Kim *et al.*, 2012). Likewise, cases of *V. parahaemolyticus* are also common in South East Asia, for example, in Thailand and Cambodia (FAO/WHO, 2011 & Vandy *et al.*, 2012). Nakaguchi (2011) also noted a high prevalence of *V. parahaemolyticus* in seafood investigated from Malaysia and Indonesia, in which supposed to signify a high infection rate, and thus, suggesting that cases of foodborne illness due to this pathogen are underreported. This is in agreement with the statement by Bilung *et al.* (2005), where infection cases for *V. parahaemolyticus* reported in Malaysia are infrequent. Creditably, the mild infections by *V. parahaemolyticus* are often neglected and pass unnoticed, hence contribute to underestimate the burden of *V. parahaemolyticus* disease, in which a major limitation in foodborne reporting in Malaysia, likewise, for other foodborne pathogens (Soon *et al.*, 2011).

Malaysian government acknowledges the rising issue of food safety, and efforts to intervene have been attempted, for instance, in 2002, the National Committee on Risk Analysis (NCRA) was established under the Food Safety and Quality Division, Ministry of Health Malaysia to look into food safety issues in the country. The NCRA initiated the first microbiological risk assessment in 2004 to estimate the risk posed by *V. parahaemolyticus* in black tiger prawn (Mohamad *et al.*, 2006). The project was attempted to request for expert advice due to the increased rejection of block frozen raw black tiger prawn by certain importing countries, and also, to issue a safety concern to the public (Sani *et al.*, 2013). Besides, attempts to improve the outbreak surveillance and monitoring system, likewise, implementing training (eg, sanitary food handling) and Hazard Analysis Critical Control Point (HACCP) were also part of the government's effort to prevent and/or reduce foodborne diseases. Also, some of the key agencies from the Malaysia's Ministry of Health, academia, industries and research institutions continue to strengthen their collaboration and networking in order to



coordinate the prevention and control of foodborne diseases, in attempts to improve the publics' health (Soon *et al.*, 2011).

Basically, this study evaluates the biosafety of *V. parahaemolyticus* in shellfishes, as an effort to provide an insight of Malaysia scenario and would warrant some degree of attention by public health authorities to better understand the potential risk posed by *V. parahaemolyticus*. Considering the fact that microorganisms are "dynamic", and may change drastically over time, constant monitoring is required to ensure our foods are safe for consumption. Therefore, the findings in this study would serve as a useful data and to further refine the food safety industries in Malaysia.

1.2 Objectives

The objectives of this study are:

- 1. To optimize the multiplex polymerase chain reaction (mPCR) assay for detection of *V. parahaemolyticus* and the respective pathogenic strains (*tdh*+ and/or *trh*+), and to compare with the loop-mediated isothermal amplification (LAMP) assay.
- 2. To detect and quantify the total and pathogenic strains of *V. parahaemolyticus* (*tdh*+ and/or *trh*+, respectively) in shellfishes from wet markets and hypermarkets, respectively.
- 3. To estimate the risk of acquiring Vibriosis (*V. parahaemolyticus* infection) from consumption of bloody clams.
- 4. To investigate the cross-contamination and de-contamination rate of *V*. *parahaemolyticus* during handling of bloody clams in domestic kitchen based on locals' practices.

REFERENCES

- Abbas, K. A., Saleh, A. M., Mohamed, A. & Lasekan, O. (2009). The relationship between water activity and fish spoilage during cold storage: A review. *Journal* of Food, Agricultural & Environmental 7: 86 – 90.
- Adam, M. & Moss, M. (2008). Bacterial agents of foodborne illness. In *Food Microbiology*, ed. Adam, M. & Moss, M., pp. 257-262. United Kingdom: RSC press.
- Alam, M. J., Tomochika, K., Miyoshi, S. & Shinoda, S. (2001). Analysis of seawaters for the recovery of culturable *Vibrio parahaemolyticus* and some other *Vibrio*. *Microbiology and Immunology* 45: 393-397.
- Alam, M. J., Tomochika, K., Miyoshi, S. & Shinoda, S (2002). Environmental investigation of potentially pathogenic Vibrio parahaemolyticus in the Seto-Island Sea, Japan. *FEMS Microbiology Letters* 208:83 – 87.
- Altekruse, S. F., Street, D. A., Fein, S.B. & Levy, A. S. (1996). Consumer knowledge of foodborne microbial hazards and food-handling practices. *Journal of Food Protection* 59: 287 294.
- Anderson, J. B., Shuster, T. A., Hansen, K. E., Levy, A. S. & Volk, A. (2004). A camera's view of consumer food-handling behaviors. *Journal of the American Dietetic Association* 104: 186 – 191.
- Bahk, G. J., Todd, E. C. D., Hong, C. H., Oh, D.H., & Ha, S. D., (2007). Exposure assessment for *Bacillus cereus* in ready-to-eat Kimbab selling at stores. *Food Control* 18: 682-688.
- Bassett, J., Nauta, M., Lindqvist, R., & Zwietering, M. (2012). Tool for microbiological risk assessment. International Life Science of Institute (ILSI) Europe Report Series.
- Bej, A. K., Patterson, D. P, Brasher, C. W., Vickery, M. C. L., Jones, D. D. & Kaysner, C. A. (1999). Detection of total and hemolysin-producing *Vibrio* parahaemolyticus in shellfish using multiplex PCR amplification of *tl*, *tdh* and *trh. Journal of Microbiological Methods* 36: 215-225.
- Beuchat (1973). Interacting effects of pH, temperature and salt concentration on the growth and survival of Vibrio parahaemolyticus. Applied Microbiology 25: 844 – 846.
- Bhunia A.K. (2008). Vibrio cholerae, Vibrio parahaemolyticus, Vibrio vulnificus. In: Foodborne Microbial Pathogens: Mechanisms and Pathogenesis, ed. Bhunia A. K., pp. 241 – 252. New York: Springer press

- Bilung, L. M., Radu, S., Bahaman, A. R., Rahim, R. A, Napis, S., Wong, C. V. L. M., Tanil, G. B. & Nishibuchi, M. (2005). Detection of *Vibrio parahaemolyticus* in clam (*Anadara granosa*) by PCR. *FEMS Microbiology Letters* 252: 85-88.
- Blodgett, R. (1998). Most probable number from serial dilutions. In U.S. Food and Drug Bacteriological analytical manual, rev. A, 8th ed. AOAC International, Gaithersburg, MD.
- Broberg, C. A., Calder, T. J. & Orth, K. (2011). *Vibrio parahaemolyticus* cell biology and pathogenicity determinants. *Microbes and Infection* 13(12-13): 992 1001.
- Brown, M., & Stringer, M. (2002). *Microbiological Risk Assessment in Food Processing.* England: Woodhead Publishing Limited.
- Burnham, V. E., Janes, M. E., Jakus, L. A., Supan, J., DePaola, A. & Bell, J. (2009). Growth and survival differences of *Vibrio vulnificus* and *Vibrio parahaemolyticus* strains during cold storage. *Journal of Food Science* 74: 314-318.
- Canizalez-Roman, A., Flores-Villasenor, H., Zazueta-Beltran, J., Muro-Amador, S. & Leon-Sicairos, N. (2011). Comparative evaluation of a chromogenic agar medium-PCR protocol with a conventional method for isolation of *Vibrio* parahaemolyticus strains from environmental and clinical samples. Canadian Journal of Microbiology 57: 136–142.
- Centers for Disease Control and Prevention (2013). Vibrio Illness (Vibriosis). Retrieved on February 2015, from *CDC*: <u>http://www.cdc.gov/vibrio</u>.
- Chamhuri, N. & Batt, P. J. (2013). Segmentation of Malaysian shoppers by store choice behaviour in their purchase of fresh meat and fresh produce. *Journal of Retail* and Consumer Services 20: 516 – 528.
- Chang, H. C., Chen, M. L., Su, Y. C., Pai, J. Y. & Chiu, T. H. (2011). Molecular characterizations of pathogenic *Vibrio parahaemolyticus* isolated from Southern Taiwan oyster-growing environment. *Food Control* 22: 245 251.
- Chen, S. & Ge, B. (2010). Development of a toxR-based loop-mediated isothermal amplification assay for detecting *Vibrio parahaemolyticus*. *BMC Microbiology* 10(41): 1-9.
- Chi, X. C., Wong, J., Yan, Q. P., & Su, Y. Q. (2007). Preparation and applications of monoclonal antibody against the pathogenic *Vibrio parahaemolyticus* from *Pseudosciaena crocea. Journal of Marine Science* 31: 1 – 5.

- Chitov, T., Kirikaew, P., Yungyune, P., Ruengprapan, N. & Sontikun, K. (2009). An incidence of large foodborne outbreak associated with *Vibrio mimicus*. *European Journal of Clinical Microbiology Infection Disease*. 28:421–424.
- Christian, J. H. B., Cole, M., Tomaska, L. 2003. Food safety and testing in perspective. In *Foodborne Microorganisms of Public Health* Significance, ed. Hocking, A. D., pp. 3-19. Waterloo DC: AIFST Inc.
- Cogan, T. A., Slader, J., Bloomfield, S. F. & Humphrey, T. J. (2002). Achieving hygiene in the domestic kitchen: the effectiveness of commonly used cleaning procedures. *Journal of Applied Microbiology* 92: 885–892.
- Cole, M. (2004). Food safety objectives concept and current status. *Mitteilungen aus Lebensmitteluntersuchung und Hygiene* 95: 13-20.
- Colwell, R. R. (2006). A global and historical perspective of the genus Vibrio. In The Biology of Vibrios, ed. Thompson F. L., Austin B., Swings J., pp. 3-11. Washington DC: ASM Press.
- Cook, D. W., Leary, P., Hunsucker, J. C., Sloan, E. M., Bowers, J. C., Blodgett, R. J. & DePaola, A. (2002). *Vibrio vulnificus* and *Vibrio parahaemolyticus* in U.S. retail shell oysters: a national survey from June 1998 to July 1999. *Journal of Food Protection* 65:79–87.
- Copin, S., Robert-Pillot, A., Malle, P., Quilici, M. L. & Gay, M. (2012). Evaluation of most-probable-number-PCR method with internal amplification control for the counting of total and pathogenic *Vibrio parahaemolyticus* in frozen shrimps. *Journal of Food Protection* 75(1): 150-153.
- Dai, J. H., Lee, Y. S., & Wong, H. C. (1992). Effects of iron limitation on production of a siderophore, outer membrane proteins, and hemolysin and on hydrophobicity, cell adherence, and lethality in mice of *Vibrio parahaemolyticus*. *Infection and Immunity*. 60: 2952-2956.
- Daniels, N. A. & Shafaie, A. (2000). A Review of pathogenic Vibrio infection for clinicans. Infections in Medicine Journal 17(10): 665-685.
- Daniels, N. A., Ray, B., Easton, A., Marano, N., Kahn, E., McShan, A. L., Del Rosario, L., Baldwin, T., Kingsley, M. A., Puhr, N. D., Wells, J. G. & Angulo, F. J. (2000). Emergence of a new Vibrio parahaemolyticus serotype in raw oysters: A prevention quandary. *Journal of the American Medical Association* 284 (12): 1541 – 1545.
- de Jong, A. E. I., Verhoeff-Bakkenes, L., Nauta, M. J. & de Jong, R. (2008). Crosscontamination in the kitchen: effect of hygiene measures. *Journal of Applied Microbiology* 105: 615 – 624.

- Deepanjali, A., Sanath Kumar, H., Karunasagar, I. & Karunasagar, I. (2005). Seasonal variation in abundance of total and pathogenic *Vibrio parahaemolyticus* bacteria in oysters along the southwest coast of India. *Applied and Environmental Microbiology* **71**(7): 3575 3580.
- DePaola, A., Nordstrom, J. L., Bowers, J. C., Wells, J. G. & Cook, D. W. (2003) Seasonal abundance of total and pathogenic *Vibrio parahaemolyticus* in Alabama oysters. *Applied and Environmental Microbiology* 69(3):1521-1526.
- Department of Statistics Malaysia (2013). Compedium of Environmental Statistic Malaysia 2013. Retrieved on January 2014, from Department of Statistics Malaysia: <u>https://www.statistics.gov.my/portal/index.php?option=com_content&view=article&id=413%253Aonline-publications-compendium-of-environment-statisticsmalaysia&catid=42&Itemid=169&lang=en</u>
- Desmarchelier, P. M. (2003). Pathogenic vibrios. In Chapter 11, Foodborne Microorganisms of Public Health Significance, ed. Hocking, A. D., pp. 333-358. Waterloo DC: AIFST Inc.
- Di Pinto, A., Terio, V., Novello, L. & Tantillo, G. (2010). Comparison between thiosulphate-citrate-bile salt sucrose (TCBS) agar and CHROMagar Vibrio for isolating Vibrio parahaemolyticus. Food Control 22(1): 124 – 127.
- Dickinson, G., Lim, K. Y. & Jiang, S. C. (2013). Quantitative microbial risk assessment of pathogeic vibrios in marine recreational waters of southern California. *Applied and Environmental Microbiology* 79(1): 294-302.
- Duff, S. B., Scott, E. A., Mafilios, M. S., Todd, E. C., Krilov, L. R., Geddes, A. M. & Ackerman, S. J. (2003). Cost-effectiveness of a targeted disinfection program in household kitchens to prevent foodborne illnesses in the United States, Canada, and the United Kingdom. *Journal of Food Protection* 66: 2103-2115.
- FAO/WHO (Food and Agriculture Organization / World Health Organization). Risk Management and food safety. FAO Food and Nutrition Paper 65: Rome. 1997.
- FAO/WHO (Food and Agriculture Organization / World Health Organization) *Risk* assessment of Vibrio parahaemolyticus in seafood: interpretative summary and technical report. Microbiological Risk Assessment Series No. 16: Rome, 2011.
- Fazil, A. M. (2005). A primer on risk assessment modeling: focus on seafood product. *FAO Fisheries Technical Paper* 462-456.
- Food Safety News (2013). CDC Progress Report: *Campylobacter* and *Vibrio* Rates Rose in 2012. Retrieved on February 2014, from Food Safety News: <u>http://www.foodsafetynews.com/2013/04/campylobacter-and-vibrio-rates-rose-in-2012-cdc-progress-report/#.UtFY79IW3xA</u>

- Forsythe, S. J. (2002). *The microbiological risk assessment of food*. United Kingdom: Blackwell Science Ltd., Blackwell Publishing Company.
- Fuenzalida, L., Armijo, L., Zabala, B., Hernández, C., Rioseco, M. L., Riquelme, C. & Espejo, R. T. (2007). Vibrio parahaemolyticus strains isolated during investigation of the summer 2006 seafood related diarrhea outbreaks in two regions of Chile. International Journal of Food Microbiology 117: 270 – 275.
- Fujino, T., Okuno, Y., Nakada, D., Aoyama, A., Fukai K., Mukai, T., & Ueho T (1953). On the bacteriological examination of shirasu-food poisoning. *Medical Journal of Osaka University* 4: 299-304.
- Galan, J. E. & Wolf-Watz, H. (2006). Protein delivery into eukaryotic cells by type III secretion machines. *Nature* 444: 567–573.
- García, K., Torres, R., Uribe, P., Hernández, C., Rioseco, M. L., Romero, J. & Espejo R. T. (2009). Dynamics of clinical and environmental *Vibrio parahaemolyticus* strains during seafood-related summer diarrhea outbreaks in southern Chile. *Applied and Environmental Microbiology* 75(23): 7482-7487
- Gardner, I. A. (2004). An epidemiologic critique of current microbial risk assessment practices: The importance of prevalence and test accuracy data. *Journal of Food Protection* 67: 2000–2007.
- Gooch, J. A., DePaola, A., Bowers, J. & Marshall, D. L., (2002). Growth and survival of *Vibrio parahaemolyticus* in postharvest American oysters. *Journal of Food Protection* 65: 970–974.
- Gorman, R., Bloomfield, S. & Adley, C. C. (2002). A study of cross-contamination of food-borne pathogens in the domestic kitchen in the Republic of Ireland. *International Journal of Food Microbiology* 76: 143-150.
- Gough, N.L. & Dodd, C.E.R. (1998). The survival and disinfection of *Salmonella typhimurium* on chopping board surfaces of wood and plastic. *Food Control* 9: 363–368.
- Gurpreet, K., Tee, G. H., Amal, N. M., Paramesarvathy, R. & Karuthan, C. (2011). Incidence and determinants of acute diarrhoea in Malaysia: A population based study. *Journal of Health Population and Nutrition* 29(2): 103 – 112.
- Hara-Kudo, Y., Nishina, T., Nakagawa, H., Konuma, H., Hasegawa, J. & Kumagai, S. (2001). Improved method for detection of *Vibrio parahaemolyticus* in seafood. *Appl Environ. Microbiol.* 67: 5819 – 5823.

- Hara-Kudo, Y., Sugiyama, K., Nishibuchi, M., Chowdhury, A., Yatsuyanagi, J., Ohtomo, Y., Saito, A., Nagano, H., Nishina, T., Nakagawa, H., Konuma, H., Miyahara, M., & Kumagai, S. (2003). Prevalence of pandemic thermostable direct hemolysin-producing *Vibrio parahaemolyticus* O3:K6 in seafood and the coastal environment in Japan. *Appl Environ Microbiol* 69(7): 3883-3891.
- Hassan, N. A., Grim, C. J., Haley, B. J., Chun, J. Alam M., Taviani, E., Hoq, M., Munk, A.C., Saunders, E., Brettin, T. S., Bruce, D. C., Challacombe, J. F., Detter, J. C., Han, C. S., Xie, G., Nair, G. B., Huq, A. & Colwell, R.R. (2010). Comparative genomics of clinical and environmental *Vibrio mimicus*. *Proceedings of the National Academy of Sciences* 107: 21134 – 21139.
- Henegariu, O., Heerema, N. A., Dlouhy, S. R., Vance, G. H., & Vogt, P. H. (1997). Multiplex PCR: Critical parameters and step-by-step protocol. *BioTechniques* 23: 504-511.
- Holvoet, K., De Keuckelaere, A., Sampers, I., Van Haute, S., Stals, A. & Uytendaelle, M. (2014). Quantitative study of cross-contamination with *Escherichia coli*, *E. coli* O157, MS2 phage and murine norovirus in a simulated fresh-cut lettuce wash process. *Food Control* 37: 218 – 227.
- Honda, T., Ni, Y. X. & Miwatani, T. (1988). Purification and characterization of a hemolysin produced by a clinical isolate of Kanagawa phenomenon-negative *Vibrio parahaemolyticus* and related to the thermostable direct hemolysin. *Infection and Immunity* 56: 961-965.
- Honda, T., Ni, Y., Miwatani, T., Adachi, T. & Kim, J. (1992). The thermostable direct haemolysin of *Vibrio parahaemolyticus* is a poreforming toxin. *Canadian Journal of Microbiology* 38: 1175 1180.
- Iida, T., Park, K. S., Suthienkul, O., Kozawa, J., Yamaichi, Y., Yamamoto, K., & Honda, T. (1998). Close proximity of the *tdh*, *trh* and *ure* genes on the chromosome of *Vibrio parahaemolyticus*. *Microbiology* 144: 2517-2523.
- Infectious Agents Surveillance Report (2008). Bacterial food poisoning in Japan, 1998-2007, 29(8): 213-215. (<u>http://idsc.nih.go.jp/iasr/29/342/tpc342.html</u>).
- International Commission on Microbiological Specifications for Foods (1996). Microorganisms in Foods 5: Characteristics of Microbial Pathogens. Blackie Academic & Professional, London, UK.
- Iwahori, J., Yamamoto, A., Suzuki H., Yamamoto, T., Tsutsui, T., Motoyama, K., Sawada, M., Matsushita, T., Hasegawa, A., Osaka, K., Toyofuku, H. & Kasuga, F. (2010). Quantitative risk assessment of *Vibrio parahaemolyticus* in finfish: A model of raw horse mackerel consumption in Japan. *Risk Analysis* 30(12): 1817-1832.

- Jay, J. M., Loessner, M. J. & Golden, D. A. (2005). Modern Food Microbiology. In Foodborne gastroenteritis caused by Vibrio Yersinia and Campylobacter Species, ed. Jay, J. M., Loessner, M. J. & Golden, D. A., pp. 657 – 664. New York: Springer
- Kaysner, C. & DePaola, A. J. (2004). U.S. Food and Drug Administration: Bacteriological Analytical Manual: Methods for specific pathogens: Chapter 9: *Vibrio*. Retrieved on January 2015, from FDA: <u>http://www.fda.gov/Food/FoodScienceResearch/LaboratoryMethods/ucm07083</u> <u>0.htm</u>
- Kaysner, C. A. & DePaola, A. (2000). Outbreaks of *Vibrio parahaemolyticus* gastroenteritis from raw oyster consumption: Assessing the risk of consumption and genetic methods for detection of pathogenic strains. *Journal of shellfish* research 19: 657.
- Kaysner, C. A., Abeyta, C., Trost, P. A., Wetherington, J. H., Jinneman, K. C., Hill, W. E., & Wekell, M. M. (1994). Urea hydrolysis can predict the potential pathogenicity of *Vibrio parahaemolyticus* strains isolated in the Pacific Northwest. *Applied and Environmental Microbiology* 60: 3020-3022.
- Kim, S. Y., Li T., Heo, J. Y., Bae, Y. M., Hwang, I. K., Lee, S. Y. & Moon, B. (2012). Efficacies of cleaning methods for decontaminating *Vibrio parahaemolyticus* on the surfaces of cutting board cross-contaminated from grated fish fillet. *Journal* of Food Safety 32: 459 – 466.
- Kim, Y. W., Lee, S. H., Hwang, I. G. & Yoon, K. S. (2012). Effect of temperature on growth of Vibrio parahaemolyticus and Vibrio vulnificus in flounder, salmon sashimi and oyster meat. International Journal of Environmental Research Public Health 9: 4662 – 4675.
- Kim, Y.B., Okuda, J., Matsumoto, C., Takahashi, N., Hashimoto, S. & Nishibuchi, M. (1999). Identification of *Vibrio parahaemolyticus* strains at the species level by PCR targeted to the *toxR* gene. *Journal of Clinical Microbiology* 37: 1173-1177.
- Klontz, K.C., Timbo, B., Fein, S., & Levy, A. (1995). Prevalence of selected food consumption and preparation behaviors associated with increased risks of foodborne disease. *Journal of Food Protection* 58: 927 930.
- Kodama, T., Hiyoshi, H., Gotoh, K., Akeda, Y., Matsuda, S., Park K. S., Cantarelli, V. V., Iida, T. & Honda, T. (2008). Identification of two translocon proteins of *Vibrio parahaemolyticus* type III secretion system 2. *Infection and Immunity* 76: 4282 4289.

- Krachler, A. M., Ham, H. & Orth, K. (2011). The outer membrane adhesion factor MAM7 initiates host cell binding during infection by gram-negative pathogens. *Proceedings of the National Academy of Sciences* 108(28): 11614-11619.
- Kusumaningrum, H. D., Riboldi, G., Hazeleger, W. C. & Beumer, R. R. (2003). Survival of foodborne pathogens on stainless steel surfaces and crosscontamination to foods. *International Journal of Food Microbiology* 85: 227 – 236.
- Lake, R., Hudson, A. & Cressey, P. (2003). Risk Profile: Vibrio parahaemolyticus in seafood. Institute of Environmental Science and Research Limited. Christchurch Science Centre. New Zealand.
- Lammerding, A. M. & Fazil, A. (2000). Hazard identification and exposure assessment for microbial food safety risk assessment. *International Journal of Food Microbiology* 58: 147-157.
- Lammerding, A., (2007). Using microbiological risk assessment (MRA) in food safety. Summary report of a workshop in Prague, Czech Republic, ILSI Europe Report Series. ILSI Europe.
- Lee, K. K., Liu, P. S. & Huang, C. Y. (2003). Vibrio parahaemolyticus infectious for both humans and edible mollusk abalone. *Microbes and Infection* 5: 481–485.
- Letchumanan, V., Yin, W. F., Lee, L. H. & Chan, K. G. (2015). Prevalence and antimicrobial susceptibility of *Vibrio parahaemolyticus* isolated from retail shrimps in Malaysia. *Frontiers in Microbiology* 6:33. Doi: 10.3389/fmicb.2015.00033
- Li-Cohen, A.E. & Bruhn, C.M. (2002). Safety of consumer handling of fresh produce from the time of purchase to the plate: a comprehensive consumer survey. *Journal of Food Protection* 65: 1287 – 1296.
- Liew, W. S., Leisner, J. J., Rusul, G., Radu, S. & Rassip, A. (1998). Survival of Vibrio spp. including inoculated V. cholerae 0139 during heat-treatment of cockles (Anadara granosa). International Journal of Food Microbiol 42: 167-173.
- Lindqvist, R., Sylven, S., & Vagsholm, I., 2002. Quantitative microbial risk assessment exemplified by *Staphylococcus aureus* in unripened cheese made from raw milk. *International Journal of Food Microbiology* 78: 155–170.

- Liu, F., Guan, W., Alam, M.J., Shen, Z., Zhang, S., Li, L., Shinoda, S., & Shi, L. (2009). Pulsed-field gel electrophoresis typing of multidrug-resistant *Vibrio* parahaemolyticus isolated from various sources of seafood. *Journal of Health Science* 55: 783-789.
- Luber, P., Brynestad, S., Topsch, D., Scherer, K. & Bartelt, A. (2006). Quantification of *Campylobacter* Species cross-contamination during handling of contaminated fresh chicken parts in kitchens. *Applied and Environmental Microbiology* 72(1): 66 – 70
- Makino, K., Oshima, K., Kurokawa, K., Yokoyama, K., Uda, T., Tagomori, K., Iijima, Y., Najima, M., Nakano, M., Yamashita, A., Kubota, Y., Kimura, S., Yasunaga, T., Honda, T., Shinagawa, H., Hattori, M. & Iida, T. (2003). Genome sequence of *Vibrio parahaemolyticus* a pathogenic mechanism distinct from that of *V. cholerae. The Lancet* 361:743-749.
- Malcolm, T. T. H., Cheah, Y. K., Mohamed Radzi, C. W. J. W., Abu Kasim, F., Kantilal, H. K., John, T. Y. H., Martinez-Urtaza, J., Nakaguchi, Y., Nishibuchi, M. & Son, R. (2015). Detection and quantification of pathogenic *Vibrio parahaemolyticus* in shellfish by using multiplex PCR and loop-mediated isothermal amplification assay. *Food Control* 47: 664-671.
- Martinez-Urtaza, J., Baker-Austin, C., Jones, J. L., Newton, A. E. & DePaolo, A., 2013. Spread of pacific northwest Vibrio parahaemolyticus strain. New England Journal of Medicine 369: 1573 - 1574
- Martinez-Urtaza, J., Lozano-Leon, A., DePaola, A., Ishibashi, M., Shimada, K., Nishibuchi, M., & Liebana, E. (2005). Characterization of pathogenic *Vibrio parahaemolyticus* isolates from clinical sources in Spain and comparison with Asian and North American pandemic isolates. *Journal of Clinical Microbiology* 42(10): 4672 4678.
- Martinez-Urtaza, J., Lozano-Leon, A., Varela-Pet, J., Trinanes, J., Pazos, Y. & Garcia-Martin, O. (2008). Environmental determinants of the occurrence and distribution of Vibrio parahaemolyticus in the rias of Galicia, Spain. Applied and Environmental Microbiology 74: 265–274.
- Massung, R. F. (2005). DNA Amplification: Current Technologies and Applications. *Emerg Infect Dis* 11(2): 357. Doi: 10.3201/eid1102.041049
- Miles, D.W., Ross, T., Olley, J. & McMeekin, T.A. (1997). Development and evaluation of a predictive model for the effect of temperature and water activity on the growth rate of *Vibrio parahaemolyticus*. *International Journal of Food Microbiology* 38: 133–142.

- Ministry of Health Malaysia (MOH) (2003). Food consumption statistics of Malaysia 2002/2003 for adult population aged 18 to 59 years. Putrajaya: Food Safety and Quality Division, Ministry of Health Malaysia.
- Miyamoto, Y., Kato, T., Obara, Y., Akiyama, S., Tazikawa, K. & Yamai, S. (1969). In vitro hemolytic characteristic of Vibrio parahaemolyticus : its close correlation with human pathogenicity. Journal of Bacteriology 100: 1147 – 1149.
- Miyamoto, Y., Obara, Y., Nikkawa, T., Yamai, S., Kato, T., Yamada, Y. & Ohashi, M. (1980). Simplified purification and biophysicochemical characteristics of Kanagawa phenomenon associated hemolysin of *Vibrio parahaemolyticus*. *Infection and Immunity* 28: 567 – 576.
- Mohamad, A. R., Hashim, J. K., Gunsalam, J., Radu, S. (2006). Microbiological Risk Assessment: Risk Assessment of Vibrio parahaemolyticus in Black Tiger Prawn (Penaeus monodon) – Technical Report. Joint Food Safety and Quality Division, Ministry of Health Malaysia/ National Food Safety Research Centre, Faculty of Food Science and Technology, Universiti Putra Malaysia Expert Consultation on Risk Assessment of Vibrio parahaemolyticus in Black Tiger Prawn (Penaeus monodon).
- Molenda, J. R., Johnson, W. G., Fishbein, M., Wentz, B., Mehlman, I. J., & Dadisman, T. A. (1972). *Vibrio parahaemolyticus* gastroenteritis in Maryland: laboratory aspects. *Journal of Applied. Microbiology*. 24: 444-448.
- Montville, T. J. & Matthews, K. R. (2005). *Vibrio* Species. In *Food Microbiology: An Introduction*, ed. Montville, T. J. & Matthews, K. R., pp. 164 – 169. Washington DC: ASM Press.
- Nagamine, K., Hase, T. & Notomi, T. (2002). Accelerated reaction by loop-mediated isothermal amplification using loop primers. *Molecular and Cell Probes* 16: 223 229.
- Nakaguchi, Y. (2013). Contamination by *Vibrio parahaemolyticus* and its Virulent strains in seafood marketed in Thailand, Vietnam, Malaysia and Indonesia. *Tropical medicine and health* 41(3): 95-102.
- Nauta, M. J., Litman, S., Barker, G. C. & Carlin, F. (2003). A retail and consumer phase model for exposure assessment of *Bacillus cereus*. *International Journal of Food Microbiology* 83: 205 218.
- Nemoto, J., Ikedo, M., Kojima, T., Momoda, T., Konuma, H. & Hara-Kudo, Y. (2011). Development and evaluation of a loop-mediated isothermal amplification assay for rapid and sensitive detection of *Vibrio parahaemolyticus*. *Journal of Food Protection* 74: 1462-1467.

- Nishibuchi, M. & Kaper, J.B. (1995) Thermostable direct hemolysin gene of *Vibrio* parahaemolyticus : a virulence gene acquired by a marine bacterium. *Infection* and *Immunity* 63: 2093-2099.
- Nishibuchi, M., Fasano, A., Russel, R. G. & Kaper, J. B. (1992). Enterotoxigenicity of *Vibrio parahaemolyticus* with and without genes encoding thermostable direct hemolysin. *Infection and Immunity* 60: 3539 – 3545.
- No, A.R., Okada, K, Kogure, K, & Park, K.S. (2011). Rapid detection of Vibrio parahaemolyticus by PCR targeted to the histone-like nucleoid structure (H-NS) gene and its genetic characterization. Letters in Applied Microbiology 53(2): 127-133.
- Noorlis, A., Ghazali, F. M., Cheah, Y. K., Tuan Zainazor, T. C., Ponniah, J., Tunung, R., Tang, J. Y. H., Nishibuchi, M., Nakaguchi, Y. & Son, R. (2011). Prevalence and quantification of *Vibrio species* and *Vibrio parahaemolyticus* in freshwater fish at hypermarket level. *International Food Research Journal* 18: 689-695.
- Nordstorm, J. L., Vickery, M. C. L., Blackstone, G. M., Murray, S. L. & DePaolo, A. (2007). Development of a multiplex real-Time PCR assay with an internal amplification control for the detection of total and pathogenic *Vibrio parahaemolyticus* bacteria in oysters. *Applied and Environmental microbiology* 73(18): 5840-5847.
- Notomi, T., Okayama H., Masubuchi H., Yonekawa T., Watanabe K., Amino N. & Hase T. (2000). Loop-mediated isothermal amplification of DNA. *Nucleic Acids Research* 28: E63.
- Ohnishi, K., Nakahira, K., Unzai, S., Mayanagi, K., Hashimoto, H., Shiraki, K., Honda, T. & Yanagihara, I. (2011). Relationship between heat-induced fibrillogenicity and hemolytic activity of thermostable direct hemolysin and a related hemolysin of Vibrio parahaemolyticus. FEMS Microbiology Letters. 318:10–17.
- Panicker, G., Call, D. R., Krug, M. J., & Bej, A. K. (2004). Detection of pathogenic Vibrio spp. in shellfish by using multiplex PCR and DNA microarrays. Applied and Environmental microbiology 70(12): 7436-7444.
- Parveen, S, Hettiarachchi, K. A., Bowers, J. C., Jones, J. L., Tamplin, M. L., McKay, R., Beatty, W., Brohawn, K., Dasilva, L. V. & Depaola, A. (2008). Seasonal distribution of total and pathogenic *Vibrio parahaemolyticus* in Chesapeake Bay oysters and waters. *International Journal of Food Microbiology* 128(2): 354-361.
- Paydar, M., The, C. S. J. & Thong, K. L. (2013). Prevalence and characterisation of potentially virulent *Vibrio parahaemolyticus* in seafood in Malaysia using conventional methods, PCR and REP-PCR. *Food Control* 32: 13 – 18.

- Polymerase Chain Reaction. In *Wikipedia*. Retrieved on March 2015, from Wikipedia: <u>https://en.wikipedia.org/wiki/Polymerase chain reaction.</u>
- Purnell, G., Mattick, K. & Humphrey, T. (2004). The use of "hot wash" treatments to reduce the number of pathogenic and spoilage bacteria on raw retail poultry. *Journal of Food Engineering* 62: 29–36.
- Ravishankar, S., Zhu, L. & Jaroni, D. (2010). Assessing the cross contamination and transfer rates of *Salmonella enterica* from chicken to lettuce under different food-handling scenarios. *Food Microbiology* 27: 791 – 794.
- Ray, B. & Bhunia, A. K. (2008). Foodborne Infections. In *Fundamental Food Microbiology*, ed. Ray, B. & Bhunia, A. K., pp. 304 306. London: CRC Press.
- Redmond, E.C. & Griffith, C.J. (2003). Consumer food handling in the home: a review of food safety studies. *Journal of Food Protection* 66: 130 161.
- Ripabelli, G., Sammarco, M. L., Grasso, G. M., Fanelli, I., Capriolli, A., & Luzzi, I. (1999). Occurrence of *Vibrio* and other pathogens bacteria in *Mytilus* galloprovincialis (mussels) harvested from Andriatic Sea, Italy. *International* Journal of Food Microbiology 49: 43-48.
- Robert-Pillot, A., Guénolé, A., Lesne, J., Delesmont, R., Fournier, J. M., Quilici, M. L. (2004). Occurrence of the tdh and trh genes in *Vibrio parahaemolyticus* isolates from waters and raw shellfish collected in two French coastal areas and from seafood imported into France. *International Journal of Food Microbiology* 91: 319-325.
- Rodriguez-Castro, A., Ansede-Bermejo, J., Blanco-Abad, V., Varela-Pet, J., Garcia-Martin, O. & Martinez-Urtaza, J. (2009). Prevalence and genetic diversity of pathogenic populations of *Vibrio parahaemolyticus* in coastal waters of Galicia, Spain. *Environmental Microbiology Reports* 2(1): 58-66.
- Rosec, J. P., Simon, M., Causse, V. & Boudjemaa, M. (2009). Detection of total and pathogenic Vibrio parahaemolyticus in shellfish: Comparison of PCR protocols using pR72H or toxR targets with a culture method. International Journal of Food Microbiology 129: 136-145.
- Sambrook, J., Fritsch, E. F. & Maniatis. *Molecular cloning: A laboratory manual*. Cold Spring Harbor Laboratory, Cold Spring Harbor: New York. 1989.
- Sani, N. A., Ariyawansa, S., Babji, A. S., & Hashim, J. K. (2013). The risk assessment of Vibrio parahaemolyticus in cooked black tiger shrimps (*Penaeus mondon*) in Malaysia. Food Control 31: 546-552.

- Schott, E. (1996). Foodborne disease and other hygiene issues in the home. *Journal of Applied Bacteriology* 80: 5 9.
- Shamsudin, M. N., & Selamat, J. 2005. Changing retail food sector in Malaysia. PECC Pacific Food System Outlook 2005-06 Annual Meeting. Kun Ming. China 11-13 May.
- Shimohata T. & Takahashi A (2010). Diarrhea Induced by Infection of Vibrio parahaemolyticus. The Journal Medical Investigation 57(3,4): 179-182.
- Shinoda, S (2011). Sixty years from the discovery of *Vibrio parahaemolyticus* and some recollections. *Biocontrol Science* 16(1): 129-137.
- Sobrinho, P. S. C., Destro, M. T., Franco, B. D. G. M. & Landgraf, M. (2014). A quantitative risk assessment model for *Vibrio parahaemolyticus* in raw oysters in Sao Paulo State, Brazil. *International Journal of Food Microbiology* 180: 69-77.
- Soon, J. M., Singh, H., & Baines, R. (2011). Foodborne diseases in Malaysia: A review. *Food control* 22: 823-830.
- Su, Y.C., & Liu, C. (2007). Vibrio parahaemolyticus: A concern of seafood safety. Food Microbiology 24: 549-558.
- Sujeewa, A. K. W., Norrakiah, A. S. & Laina, M. (2009). Prevalence of toxic genes of Vibrio parahaemolyticus in shrimps (Penaeus monodon) and culture environment. International Food Research Journal 16: 89-95.
- Sutton, S. (2010). The most probable number method and its uses in enumeration, qualification, and validation. *Journal of Valid Technology* 16:35–38.
- Tada, J., Ohashi, T., Nishimura, N., Shirasaki, Y., Ozaki, H., Fukushima, S., Takano, J., Nishibuchi, M. & Takeda, Y. (1992). Detection of thermostable direct hemolysin gene (*tdh*) and thermostable direct-related hemolysin gene (*trh*) of *Vibrio parahaemolyticus* by polymerase chain reaction. *Molecular and Cellular Probes* 64: 477-487.
- Takahashi, A., Sato, Y., Shiomi, Y., Cantarelli, V. V., Iida, T., Lee, M. & Honda, T. (2000). Mechanisms of chloride secretion induced by thermostable direct haemolysin of *Vibrio parahaemolyticus* in human colonic tissue and a human intestinal epithelial cell line. *Journal of Medical Microbiology* 49: 801 - 810.

- Tanaka, N., Iwade, Y., Yamazaki, W., Gondaira, F., Vuddhakul, V., Nakaguchi, Y. & Nishibuchi, M. (2014). Most-probable-number loop-mediated isothermal amplification–based procedure enhanced with K antigen–specific immunomagnetic separation for quantifying tdh+ Vibrio parahaemolyticus in molluscan shellfish. Journal of Food Protection, 77(7): 1078-1085.
- Terzi, G., Buyuktanir, O. & Yurdusev, N. (2009). Detection of *tdh* and *trh* genes in *Vibrio parahaemolyticus* isolates in fish and muscles from Middle Black Sea Coast of Turkey. *Letters in Applied Microbiology* 49: 757 – 763.
- Tortora, G. J., Christine, L., Case, C. L., Funke, B.R., Funke, B. & Case, C. (2006) *Microbiology: An Introduction.* United States of America. Pearson Education.
- U.S Food and Drug Administration (2011). Fish and fishery products hazards and controls guidance Fourth Edition. Retrieved on February 2013, from FDA: <u>http://www.fda.gov/Food/GuidanceRegulation/GuidanceDocumentsRegulatoryI</u>nformation/Seafood/ucm2018426.htm
- U.S. Food and Drug Administration (2005). Quantitative Risk Assessment on the Public Health Impact of Pathogenic *Vibrio parahaemolyticus* in raw oysters. Center for Food Safety and Applied Nutrition, Food and Drug Administration, U.S. Department of Health and Human Services. <u>http://www.fda.gov/Food/ScienceResearch/ResearchAreas/RiskAssessmentSafet</u> <u>yAssessment/ucm184484.htm.</u>
- Urakawa, H. & Rivera, I. N. G. (2006). Aquatic Environment. In *The Biology of Vibrios*, ed. Thompson, F. L., Austin, B., Swings, J., pp. 175-189. Washington DC: ASM Press.
- van Asselt, E. D., de Jong, A. E. I, de Jonge, R. & Nauta, M. J. (2008). Crosscontamination in the kitchen: estimation of transfer rates from cutting boards, hand and knives. *Journal of Applied Microbiology* 105: 1392 – 1401.
- Vanderlinde, P. B. & Desmachelier, P. M. (2003). Microbial risk management and risk assessment. In *Foodborne microorganisms of public health significance*, ed. Hocking, A. D, pp. 21-38. Waterloo D. C: AIFST Inc.
- Vandy, S., Leakhann, S., Phalmony, H., Denny, J., & Roces, M. C. (2012). Vibrio parahaemolyticus enteritis outbreak following a wedding banquet in a rural village – Kampong Speu, Cambodia, April 2012. Outbreak Investigation Report. WPSAR 3(4).

- Velazquez-Roman, J., Leon-Sicairos, N., Flores-Villasenor, H., Villafana-Rauda, S. & Canizales-Roman, A. (2012). Association of Pandemic Vibrio parahaemolyticus O3:K6 Present in the coastal environment of northwest Mexico with cases of recurrent diarrhea between 2004 and 2010. Applied and Environmental Microbiology 78(6): 1794 – 1803.
- Velusamy, V., Arshak, K., Korostynska, O., Oliwa, K. & Adley, C. (2010). An overview of foodborne pathogen detection: In the perspective of biosensors. *Biotechnology Advances* 28: 232 – 254.
- Wachtel, M.R., McEvoy, J.L., Luo, Y., Williams-Campbell, A.M. & Solomon, M.B. (2003). Cross-contamination of lettuce (*Lactuca sativa* L.) with *Escherichia coli* O157:H7 via contaminated ground beef. *Journal of Food Protection* 66: 1176 – 1183.
- Wadstrom T. & Ljungh A. (2014). Vibrio as a foodborne pathogen. In Food Associated Pathogens, ed. Tham, W. & Danielsson-Tham, M. L., pp.269-277. London: CRC Press.
- Wang, L., Zhang, J., Bai, H., Li, X., Lv, P. & Guo, A. (2014). Specific detection of Vibrio parahaemolyticus by fluorescence quenching immunoassay based on quantum dots. Applied Biochemistry and Biotechnology 173(5): 1073 – 1082.
- Whyte, P., McGill, K. & Collins, J. D. (2003). An assessment of steam pasteurization and hot water immersion treatments for the microbiological decontamination of broiler carcasses. *Food Microbiology* 20: 111–117.
- Wong, H. C. & Lee, Y.S. (1994). Regulation of iron on bacterial growth and production of thermostable direct hemolysin by *Vibrio parahaemolyticus* in intraperitoneal infected mice. *Microbiology and Immunology* 38: 367-371.
- Wooldrige M. (2009). Qualitative Risk Asssessment. In *Microbial Risk Analysis of Food* ed. Schaffner D. W., pp 1- 27. Washington D. C.: ASM Press.
- Yamamoto, A., Nishibuchi, M., Kasuga, F., Yamamoto, S., Toyofuku, H., Shigematsu, M., Osaka, K., Vose, D., Charernjiratragul, W., Vuddhakul, V., & Iwahori, J. (2008). Quantitative modeling for risk assessment of *Vibrio parahaemolyticus* in bloody clams in southern Thailand. *International Journal of Food Microbiology* 124(1): 70-78.
- Yamazaki, W., Kumeda, Y., Misawa, N., Nakaguchi, Y. & Nishibuchi, M. (2010). Development of loop-mediated isothermal amplification assay for sensitive and rapid detection of the *tdh* and *trh* genes of *Vibrio parahaemolyticus* and related *Vibrio* species. *Applied and Environmental Microbiology* 76(3): 820-828.

- Yamazaki, W., Ishibashi, M., Kawahara, R. & Inoue, K. (2008). Development of a loop mediated isothermal amplification assay for sensitive and rapid detection of *Vibrio parahaemolyticus*. *BMC Microbiolology* 8: 163.
- Yang, Z. Q., Jiao, X. A., Zhou, X. H., Gao, G. X., Fang, W.M. & Gu, R.X. (2008). Isolation and molecular characterization of *Vibrio parahaemolyticus* from fresh, low-temperature preserved, dried and salted seafood products in two coastal areas of eastern China. *International Journal of Food Microbiology* 125(3): 279 – 285.
- Yeung, P. S. & Boor, K.J. (2004). Epidemiology, pathogenesis, and prevention of foodborne Vibrio parahaemolyticus infections. Foodborne Pathogen and Disease 1: 74–88.
- Yoh, M., Tang, G. Q. & Honda, T. (1999). Is type strain of Vibrio parahaemolyticus TRH producer? The 33rd Symposium of Vibrio parahaemolyticus. Naha, Okinawa.
- Zimmerman, A. M., DePaola, A., Bowers, J. C., Krantz, J. A., Nordstrom, J. L., Johnson, C. N. & Grimes, D. J. (2007). Variability of total and pathogenic Vibrio parahaemolyticus densities in northern gulf of Mexico Applied and Environmental Microbiology 73: 7589–7596.

APPENDICES

Appendix A: Media and solution preparation

<u>Media</u>

Alkaline Peptone Water (APW)	
Peptone	10.0 g
Sodium Chloride	10.0 g

Suspend 20.0 g of the APW powder in 1 L of purified water and mix thoroughly. Autoclave at 121° C for 15 minutes. Final pH 8.3 – 8.7.

Tryptic Soy Agar 3% sodium chloride (TSA 3% NaCl)

Pancreatic Digest of Casein	15.0 g
Papaic Digest of Soybean	5.0 g
Sodium chloride	5.0 g
Agar	15.0 g

Suspend 40.0 g of the TSA powder and 30.0 g of sodium chloride in 1 L of purified water and mix thoroughly. Heat with frequent agitation and boil for 1 minute to completely dissolve the powder. Autoclave at 121°C for 15 minutes. Final pH 7.3 \pm 0.2.

Gel electrophoresis

10X Tris-Borate-EDTA (10X TBE)

Tris-Base	108.0 g
Boric acid	55.0 g
EDTA	9.3 g
Distilled water	1000 ml

For routine electrophoresis, 10X TBE were diluted to 0.5X TBE, with the pH 8.3. Solution can be kept at room temperature.

1.2% Agarose Gel

Agarose	1.2 g
0.5X TBE	100 ml

Heat until the medium boils. Do not overheat.

Ethidium Bromide solution

C

Ethidium bromide	1.0 g
Distilled water	100 ml



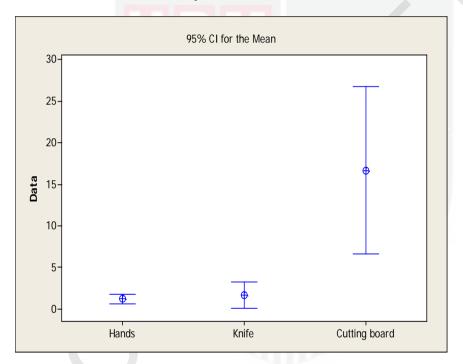
Appendix B: Statistical Analysis

<u>Statistical Analysis of Kitchen Simulation Data (Working Chapter 3)</u> <u>Transfer rate (primary cross-contamination)</u>

Kruskal-Wallis Test: Transfer rate versus surface transferred to						
Treatment	Ν	Median		Ave	Rank	Ζ
Cutting board	10	14.8000		24.6	4.00	
Hands		10	0.8950		11.4	-1.80
Knife		10	0.8950		10.5	-2.20
Overall		30			15.5	

 $H = 16.08 \ DF = 2 \ P = 0.000$

H = 16.09 DF = 2 P = 0.000 (adjusted for ties)

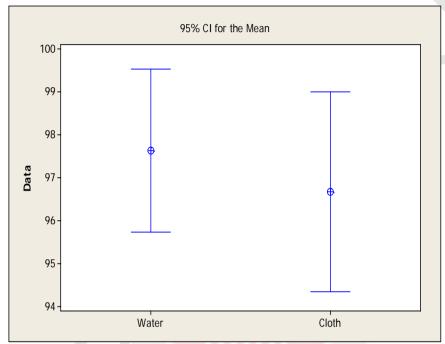


Conclusion: Difference is significant.

<u>Decontamination rate</u> Mann-Whitney Test: Decontamination rate versus type of treatment

Treatment	Ν	Median
Water	15	98.790
Cloth	16	98.340

Point estimate for ETA1-ETA2 is 0.430 95.4 Percent CI for ETA1-ETA2 is (-0.679,2.018) W = 255.0 Test of ETA1 = ETA2 vs ETA1 not = ETA2 is significant at 0.5665



Conclusion: There is no difference between treatments.

Kruskal-Wallis Test: Treatment on hands

Treatment	Ν	Median	A	ve	Rank	Ζ
No treatment		10	0.89500		34.6	4.13
Water		15	0.06000		12.9	-3.30
Cloth		16	0.14000		20.1	-0.37
Overall		41			21.0	

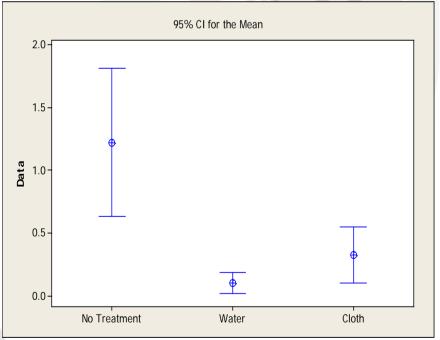
 $H = 19.89 \ DF = 2 \ P = 0.000$

H = 19.96 DF = 2 P = 0.000 (adjusted for ties)

Mann-Whitney Test: Water versus table cloth

Treatment	Ν	Median
Water	15	0.0600
Table Cloth	16	0.1400

Point estimate for ETA1-ETA2 is -0.0700 95.4 Percent CI for ETA1-ETA2 is (-0.3400,-0.0002) W = 192.0Test of ETA1 = ETA2 vs ETA1 not = ETA2 is significant at 0.0604 The test is significant at 0.0594 (adjusted for ties)



Conclusion: Difference is significant.

Kruska-Wallis: Treatment on knife

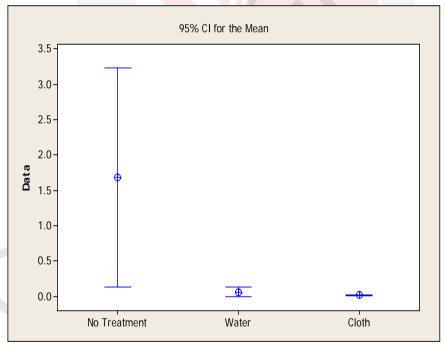
Treatment	Ν	Median	Ave	Rank	Ζ
No Treatment	10	0.89500	35.3	4.33	
Water		15 (0.01000	18.2	-1.15
Cloth		16 (0.01500	14.8	-2.67
Overall		41		21.0	

 $\begin{array}{l} H = 19.35 \quad DF = 2 \quad P = 0.000 \\ H = 19.80 \quad DF = 2 \quad P = 0.000 \quad (adjusted \ for \ ties) \end{array}$

Mann-Whitney: Water versus cloth

Treatment	Ν	Median
Water	15	0.0100
Cloth	16	0.0150

Point estimate for ETA1-ETA2 is 0.010095.4 Percent CI for ETA1-ETA2 is (-0.0100, 0.0400)W = 264.0 Test of ETA1 = ETA2 vs ETA1 not = ETA2 is significant at 0.3529 The test is significant at 0.3413 (adjusted for ties)



Conclusion: Difference is significant.

Kruskal-Wallis: Treatments on cutting board

Treatment	Ν	Medi	an	Ave	Rank	Z
No Treatment	10	0.895	500	35.3	4.33	
Water		15	0.01000)	18.2	-1.15
Cloth		16	0.01500)	14.8	-2.67
Overall		41			21.0	

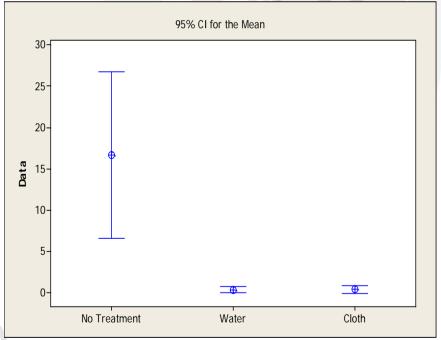
H = 19.35 DF = 2 P = 0.000

H = 19.80 DF = 2 P = 0.000 (adjusted for ties)

Mann-Whitney: Water versus table cloth

Treatment	Ν	Median
Water	15	0.100
Cloth	16	0.035

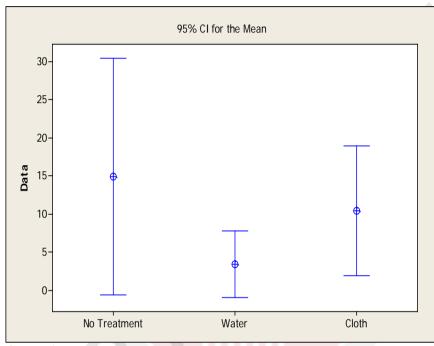
Point estimate for ETA1-ETA2 is 0.040 95.4 Percent CI for ETA1-ETA2 is (-0.050,0.180) W = 278.0Test of ETA1 = ETA2 vs ETA1 not = ETA2 is significant at 0.1383 The test is significant at 0.1361 (adjusted for ties)





Kruskal-Wallis: Transfer rate to lettuce						
Treatment	Ν	Media	n	Ave	Rank	Ζ
No Treatment	4	14.385	5	10.3	1.56	
Water		5	2.860		4.2	-2.20
Cloth		5	11.580		8.6	0.73
Overall		14			7.5	

$H=5.19 \ DF=2 \ P=0.075$



Conclusion: There is no difference between transfer rates.

Transfer efficiency (secondary cross-contamination)

Kruskal-Wallis Test: Primary vs secondary cross-contamination					
Treatment	N	Median	Ave	Rank	Ζ
Cloth	11	0.01000	8.0	-3.17	
Primary	10	18.23000	21.5	4.22	
Water	5	0.02000	9.7	-1.24	
Overall	26		13.5		

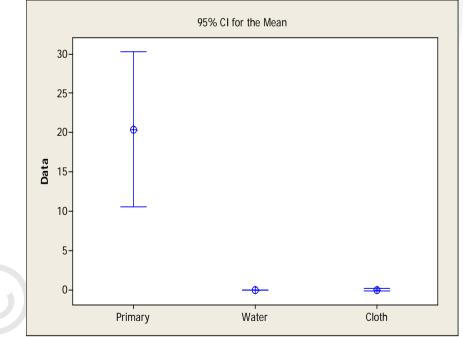
 $H = 17.96 \ DF = 2 \ P = 0.000$

H = 18.25 DF = 2 P = 0.000 (adjusted for ties)

Mann-Whitney Test: Secondary cross-contamination (water versus cloth)

Treatment	Ν	Median
Water 5	0.0200	
Cloth	11	0.0100

Point estimate for ETA1-ETA2 is 0.010095.9 Percent CI for ETA1-ETA2 is (-0.0299, 0.0200)W = 48.5 Test of ETA1 = ETA2 vs ETA1 not = ETA2 is significant at 0.5332 The test is significant at 0.5184 (adjusted for ties)



Conclusion: Difference is significant

BIODATA OF THE STUDENT

Malcolm Tan Turk Hsern was born in Selangor, Malaysia on November 24th, 1989. He went for his primary education at Sekolah Rendah Kebangsaan SS17 Subang Jaya, Subang Jaya. After that, he continued her secondary education at Sekolah Menengah Kebangsaan SS17 Subang Jaya, Subang Jaya, He went to foundation in 2007 and pursued his Bachelor of Science (Biotechnology) in 2008 at University Tunku Abdul Rahman (UTAR) and graduated in 2011. He then decided to further study at Universiti Putra Malaysia (UPM) and was offered to pursue a Master's Degree by the School of in 2013. The followings are the publications that he and his colleagues have published and conferences that he has attended during the course of her study.



LIST OF PUBLICATIONS

- Malcolm, T. T. H., Cheah, Y. K., Mohamed Radzi, C. W. J. W., Abu Kasim, F., Kantilal, H. K., John, T. Y. H., Martinez-Urtaza, J., Nakaguchi, Y., Nishibuchi, M. & Son, R. (2015). Detection and quantification of pathogenic *Vibrio parahaemolyticus* in shellfish by using multiplex PCR and loop-mediated isothermal amplification assay. *Food Control* 47: 664-671.
- New, C. Y., Kantilal, H. K., Tan, M. T. H., Nakaguchi, Y., Nishibuchi, M., & Son, R. (2014). Consumption of raw oysters: a risk factor for Vibrio parahaemolyticus infection. International Food Research Journal 21(6): 2459 – 2472.
- Najwa, M.S., Rukayadi, Y., Ubong, A., Loo, Y.Y., Chang, W.S., Lye, Y.L., Thung, T.Y. Aimi, S.A., Malcolm, T.T.H, Goh, S.G., Kuan, C.H., Yoshitsugu, N., Nishibuchi, M. and Son, R. (2015). Quantifiation and antibiotic susceptibility of Salmonella spp., Salmonella Enteritidis and Salmonella Typhimurium in raw vegetables (ulam). International Food Research Journal 22(5): 1761-1769.

Proceedings

- Malcolm T. T. H., Che Wan Jasimah W. M. R., Farihah A.K. Azirah H. Nakaguchi Y., Nischibuchi M. & Son R. Incidence of tdh-positive and trh-positive Vibrio parahaemolyticus in cockles determined by the Most probable number-Loop mediated isothermal amplification (MPN-LAMP). South African Association for Food Science & Technology Biennial International Congress and Exhibition 7-9 October, 2013. Pretoria, South Africa., South African Association for Food Science & Technology Biennial International Congress and Exhibition, 07 Oct 2013 to 09 Oct 2013, South African Association for Food Science and Technology, (International)
- Malcolm T. T. H., Cheah Y. K., Che Wan Jasimah W. M. R., Haresh K. K., Nakaguchi Y., Nischibuchi M., and Son R. Incidence of Pathogenic Vibrio parahaemolyticus in Cockles Determined by the Most Probable Number-Multiplex Polymerase Chain Reaction (MPN-mPCR). 2nd International Food Safety Conference 2-3 December, 2013. Royale Chulan Hotel, Kuala Lumpur.