

UNIVERSITI PUTRA MALAYSIA

DEVELOPMENT OF A REAL-TIME PCR ASSAY FOR DIAGNOSIS OF BRUCELLA MELITENSIS INFECTION IN SHEEP

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By

TAKELE BELAY

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DEVELOPMENT OF A REAL-TIME PCR ASSAY FOR DIAGNOSIS OF BRUCELLA MELITENSIS INFECTION IN SHEEP

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Chairman: Siti Khairani Bejo, PhD

Faculty: Veterinary Medicine

In this study, a SYBR Green based real-time PCR assay using primers specific to IS711 of *Brucella melitensis* was developed and its diagnostic utility was evaluated by comparing it with conventional PCR method for detection of *Brucella* DNA from cultured bacteria and experimentally infected mice samples. The study was first undertaken with cultured bacteria, *B. melitensis* 16M as a reference and two *B. melitensis* Malysian isolates. *Brucella abortus* 544 and *Yersinia enterocolitica* O:9 were also used for cross reactivity testing. Three of the *B. melitensis* were successfully detected using both optimized SYBR Green I real-time and conventional PCR. No amplification products were produced with these primers from strains of *B. abortus* and *Y. enterocolitica* O:9 indicating that the primer sets are specific only for *B. melitensis*. The specificity of the PCR products amplified by the primers from Malysian strains of *B. melitensis* were confirmed by sequence analysis. The detection limit of the real-time PCR was compared with conventional PCR by preparing serially ten-fold dilutions of *B.*



melitensis DNA. The real-time PCR was found to be 10-fold more sensitive than the conventional PCR, detecting up to 50 fg of genomic DNA.

The viability of using conventional and real-time PCR assays together with culture methods as potential diagnostic tools for the detection of *B. melitensis* in experimentally infected mice was also assessed over an 8-week period of infection. It was evident from our result that regardless of duration of infection and sample type (liver, lymph node, blood, spleen and serum) the diagnostic yield (sensitivity) was 100%, 96.7% and 62.5% for real-time PCR, conventional PCR and culture methods respectively. This indicated that the sensitivity of the PCR assays was far higher than that of the culture method. The specificity of both conventional and real-time PCR was 100%. When serum was used as PCR template, *B. melitensis* specific amplicon was detected only until day 45 post inoculation (p.i) using conventional PCR unlike the real-time PCR method whereby positive results were detected through out the study period. In all samples with positive real-time PCR results, *B. melitensis* specific amplicons were distinguished by their characteristic melting temperature ranged between 86.4 °C to 87.4 °C in melting curve analysis.

Analytical sensitivities of the different tissue types used in the study were also compared using threshold cycle (Ct) values of the real-time PCR. For this purpose, samples collected at day 30 p.i. were used and the most sensitive tissue sample amongst the tested samples was estimated from the standard curve obtained from cloned *B. melitensis* 16M. Accordingly, the most sensitive tissue was found to be serum (mean Ct=15.07) followed by lymph node (mean Ct=19.50), spleen (mean Ct= 24.2), liver (mean Ct=26.4) and whole blood (mean Ct=26.2). These results indicate that serum samples



followed by lymph node are more sensitive for the real-time PCR assay than the other samples tested in this study.

In conclusion, this present results suggest that the diagnostic yield of real-time PCR that uses serum samples was at least as sensitive as a similar PCR protocol that uses other tested tissue samples. The speed and technical simplicity of real-time PCR coupled with minimal risk of handling serum samples make it a useful alternative to blood / other tested tissue PCR for animals with suspected brucellosis and negative or doubtful serological test results. However, although the use of this real-time PCR assay as a supplemental diagnostic tool for detection and identification of *Brucella* organisms in clinical specimens could be recommended, the need for the assay to be validated with confirmed positive and negative clinical samples of its primary host animals, and verify its ability to differentiate the vaccinal strains from field strains, is desirable to achieve the utmost benefits from the SYBR Green I based real-time PCR assay.

Keywords: Real-time PCR, SYBR Green, Brucella, Diagnosis, Mice, Culture



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia bagi memenuhi keperluan Master Sains

PEMBANGUNAN SATU UJIAN PCR MASA NYATA UNTUK DIAGNOIS JANGKITAN BRUCELLA MELITENSIS DI DALAM BIRI-BIRI

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Dalam kajian ini, assai PCR masa nyata berasaskan SYBR Green dengan menggunakan primer yang khusus kepada IS711 bagi *B. melitensis* telah dibangunkan dan utiliti diagnostiknya telah dinilai dengan membandingkannya dengan kaedah PCR konvensyenal bagi mengesan DNA *Brucella* daripada bakteria berkultur dan sampel mencit yang telah dijangkiti. Kajian berkenaan dijalankan terlebih dahulu dengan bakteria berkultur, *B. melitensis* 16M sebagai rujukan dan dua isolat *B. melitensis* setempat. *Brucella abortus* dan *Y. enterocolitica* O:9 juga digunakan untuk pengujian kereaktifan silang. Tiga daripada *B. melitensis* telah berjaya dikesan dengan menggunakan PCR masa nyata SYBR Green I teroptimum dan PCR konvensyenal. Tiada produk amplifikasi dihasilkan dengan primer ini daripada strain *B. abortus* dan *Y. enterocolitica* O:9 yang menunjukkan bahawa set primer adalah khusus hanya untuk *B .melitensis*. Spesifisiti produk PCR yang dihasilkan oleh primer daripada strain *B. melitensis* anyata



dibandingkan dengan PCR konvensional dengan menyediakan pencairan pada tahap sepuluh kali ganda secara bersiri bagi DNA *B. melitensis*. PCR masa nyata didapati 10 kali ganda lebih sensitif daripada PCR konvensional, mengesan sehingga 50 fg DNA genomik.

Kebolehhidupan menggunakan assai PCR konvensional dan masa nyata bersama-sama dengan kaedah kultur sebagai alat diagnostik berpotensi bagi pengesanan B. melitensis di dalam mencit yang dijangkiti secara uji kaji juga telah dinilai sepanjang tempoh 8minggu jangkitan. Jelas melalui keputusan kami bahawa tanpa menghiraukan tempoh jangkitan dan jenis sampel (hati, nodus limfa, darah, limpa dan serum), hasil diagnostik (kepekaan) masing-masing adalah 100%, 96.7% dan 62.5% untuk kaedah PCR masa nyata, PCR konvensional dan kultur. Ini menunjukkan kesensitifan assai PCR adalah jauh lebih tinggi daripada kaedah kultur. Spesifiti kedua-dua PCR konvensional dan masa nyata adalah 100%. Apabila serum digunakan sebagai templat PCR, amplikon khusus B. melitensis telah dikesan hanya sehingga 45 p.i dengan menggunakan PCR konvensional tidak seperti kaedah PCR masa nyata yang telah mengesan keputusan positif di sepanjang tempoh kajian berkenaan. Dalam semua sampel dengan keputusan PCR masa nyata yang positif, amplikon khusus B. melitensis telah dibezakan melalui suhu lebur yang jelas dengan julat antara 86.4 °C hingga 87.4 °C dalam analisis lengkung lebur.

Kesensitifan analisis bagi jenis tisu berlainan yang digunakan dalam kajian ini juga telah dibandingkan dengan menggunakan nilai kitaran ambang (Ct) PCR masa nyata. Untuk tujuan ini, sampel yang diambil pada hari 30 p.i. telah digunakan dan sampel tisu yang



paling sensitif daripada sampel yang diuji telah dianggarkan daripada lengkung standard yang diperoleh daripada 16M *B. melintesis* yang telah diklon. Seterusnya, tisu yang paling sensitif adalah serum (min Ct=15.07) diikuti dengan nodus limfa (min Ct=19.50), limpa (min Ct= 24.2), hati (min Ct=26.4) dan darah penuh (min Ct=26.2). Keputusan ini menunjukkan bahawa sampel serum dan diikuti dengan nodus limfa adalah lebih sensitif untuk assai PCR masa nyata daripada sampel lain yang diuji dalam kajian ini.

Kesimpulannya, keputusan ini menunjukkan bahawa hasil diagnostik PCR masa nyata yang menggunakan sampel serum adalah sekurang-kurangnya sama sensitif seperti protokol PCR serupa yang menggunakan sampel tisu lain yang diuji. Kelajuan dan keringkasan teknikal PCR masa nyata di samping risiko minimum yang terlibat dalam mengendalikan sampel serum menjadikannya alternatif berguna bagi PCR darah / tisu lain yang diuji untuk haiwan yang disyaki mempunyai brucellosis dan keputusan ujian serologi yang negatif atau meragukan. Namun, walaupun kami menyarankan penggunaan assai PCR masa nyata ini sebagai alat diagnostik tambahan untuk pengesanan dan pengenalpastian organisma *Brucella* dalam spesimen klinikal, keperluan untuk assai agar disahkan dengan sampel klinikal positif dan negatif bagi haiwan hos utamanya, dan menentusahkan kemampuannya untuk membezakan strain vaksin daripada strain lapangan, adalah dikehendaki untuk mencapai faedah tertinggi yang diingini daripada assai PCR masa nyata berasaskan SYB Green 1.

Kata kunci: PCR masa-nyata, SYBR-Green, Brucella, diagnostik, mencit, kultur



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I certify that Examination Committee has met on 16th October 2008 to conduct the final examination of Takele Belay Yazew on his Master of Science thisis entitled "Development of a Real-time PCR for Diagnosis of Brucella melitensis Infection in Sheep" in accordance with Universiti Pertanian Malaysia (higher Degree) Act 1980 and University Pertanian Malysia (Higher Degree) Regulations 1981. The Committee recommends that the candidate be awarded the relevant degree. Members of Examination Committee wer as follows:

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DECLARATION

I here declare that the thesis is based on my original work except for quotation and citations which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at UPM or other institutions.

Takele Belay

Date:



TABELE OF CONTENT

Page

| ABSTRACT | ERROR! BOOKMARK NOT DEFINED. |
|---------------------------------|------------------------------|
| ABSTRAK | ERROR! BOOKMARK NOT DEFINED. |
| ACKNOWLEDGEMENTS | Х |
| DECLARATION | XIII |
| LIST OF TABLES | XVI |
| LIST OF FIGURES | XVII |
| LIST OF ABBREVIATIONS AND SYMBO | DLS XXI |
| | |

CHAPTER

| 1 | INTRO | DUCTION | 1 | | |
|-----|--|--|-----|--|--|
| 2 | LITERATURE REVIEW | | | | |
| | 2.1 Small Ruminant Brucellosis | | | | |
| | 2.2 Classification of Brucella Species | | | | |
| | | ology of Brucella | 9 | | |
| | | blecular Genetics of Brucella | 10 | | |
| | 2.5 An | tigenic Characteristics | 12 | | |
| | | onotic Aspects of Brucella melitensis Infection | 14 | | |
| | | ucellosis in Sheep | 16 | | |
| | 2.7.1 | Pathogenesis and Immune Response in Sheep | 16 | | |
| | 2.7.2 | Sheep Brucellosis in Malaysia | 23 | | |
| | 2.8 La | boratory Techniques in the Diagnosis of Brucellosis in Sheep | 24 | | |
| | 2.8.1 | Direct Microscopic Examination | 24 | | |
| | 2.8.2 | Isolation | 25 | | |
| | 2.8.3 | Serological Tests | 26 | | |
| | | Tests Based on Cell Mediated Immunity | 33 | | |
| | 2.8.5 | 5 | 35 | | |
| 2.9 | Strategi | es for Control and Eradication of Brucellosis in Small Ruminants | 49 | | |
| 3 | DEVEL | OPMENT OF SYBR GREEN I BASED REAL-TIME PCR I | FOR | | |
| | DETEC | TION OF <i>B. MELITENSIS</i> | 54 | | |
| | 3.1 Int | roduction | 54 | | |
| | 3.2 Ma | aterials and Methods | 57 | | |
| | 3.2.1 | Bacterial Isolates | 57 | | |
| | 3.2.2 | The DNA Extractions | 57 | | |
| | 3.2.3 | The PCR Primers | 58 | | |
| | 3.2.4 | Determination of DNA Concentration and Purity | 59 | | |
| | 3.2.5 | Optimization of SYBR Green I Real-time PCR Assay | 60 | | |
| | 3.2.6 | • | 61 | | |
| | 3.2.7 | Melting Curve Analysis | 62 | | |
| | 3.2.8 | Detection of PCR Products by Agarose Gel Electrophoresis | 62 | | |



| 3.2.9 | Evaluation of Real -time PCR | 63 |
|---------|--|----|
| 3.2.10 | Restriction Enzyme Confirmation of Recombinant Plasmids | 65 |
| 3.2.11 | Sequence Assembly and Analysis of Recombinant Plasmids | 65 |
| 3.3 Res | sults | 67 |
| 3.3.1 | Determination of DNA Concentration and Purity | 67 |
| 3.3.2 | Optimization of Real-time PCR Assay Using <i>B. melitensis</i> 16M | 67 |
| 3.3.3 | Sensitivity Test of SYBR Green I Based Real-time PCR | 69 |
| 3.3.4 | Melting Curve Analysis | 71 |
| 3.3.5 | Agarose Gel Electrophoresis Analysis | 72 |
| 3.3.6 | Evaluation of the Real-time PCR | 73 |
| 3.3.7 | Restriction enzyme confirmation of Recombinant Plasmids | 76 |
| 3.3.8 | Sequence Assembly and Analysis of Recombinant Plasmids | 77 |
| 3.4 Dis | cussion | 79 |

4 EVALUATION OF PERFORMANCE OF THE DEVELOPED REAL-TIME PCR ASSAY FOR DETECTION OF *BRUCELLA. MELITENSIS* USING CLINICAL SAMPLES OBTAINED FROM EXPERIMENTAL TRIALS AND SUSPECTED FIELD CASES OF SHEEP BRUCELLOSIS 84

| | AND SU | SPECTED FIELD CASES OF SHEEF BRUCELLOSIS | 04 |
|---------------|----------|--|-----|
| | 4.1 Int | roduction | 84 |
| | 4.2 Ma | terials and Methods | 87 |
| | 4.2.1 | Animals and Experimental Infection | 87 |
| | 4.2.2 | 1 | 88 |
| | 4.2.3 | | 88 |
| | 4.2.4 | Processing of Samples for PCR | 89 |
| | | The PCR Assays | 92 |
| | 4.2.6 | Determination of Most Sensitive Clinical Sample for Real-time PCR | 93 |
| | 4.2.7 | Application of the Developed Real-time PCR Assay on Suspected | |
| | | Clinical Samples of Sheep Brucellosis | 94 |
| | 4.3 Res | sults | 95 |
| | 4.3.1 | Experimental Infection in Mice | 95 |
| | 4.3.2 | DNA Extraction, Determination of DNA Concentration and Purity | 95 |
| | 4.3.3 | Detection of <i>B. melitensis</i> in Experimentally Infected Mice by | |
| | | Bacteriological, Conventional and Real-time PCR | 96 |
| | 4.3.4 | Application of the Developed Real-time PCR assay on Suspected | |
| | | Clinical Samples of Sheep Brucellosis | 104 |
| | 4.4 Dis | cussion | 105 |
| 5 | GENER | AL DISCUSSION AND CONCLUSION | 113 |
| REF | ERENCE | S | 117 |
| APPENDICES 13 | | | 134 |
| BIO | DATA OF | F STUDENT | 161 |
| LIST | Г OF PUB | LICATIONS | 162 |
| | | | |



LIST OF TABLES

| Table | P | age |
|-------|--|----------|
| 3.1 | Primers employed for amplification and sequencing of <i>B. melitensis</i> specific DNA segment | 59 |
| 3.2 | Performance of conventional and real-time PCR on serially diluted <i>B</i> . <i>melitensis</i> DNA template | 70 |
| 3.3 | Reproducibility tests. Ct, Copy number, S.D and C.V for triplicate standard samples of 10-fold serial dilutions | 75 |
| 3.4 | Reproducibility tests. Ct, Copy number, S.D and C.V obtained for the same dilutions over four different days | 76 |
| 4.1 | Total samples collected from experimentally infected mice based on day of infection | 97 |
| 4.2 | Diagnostic yield of three different diagnostic methods with different samples of experimental mice (Regardless of sampling date) | s 98 |
| 4.3 | Bacteriological diagnostic yield of different samples collected from <i>B</i> . <i>melitensis</i> infected mice | 99 |
| 4.4 | Diagnostic yield of three different diagnostic methods with samples collected from experimental mice (Regardless of sampling date and sample type) 1 | d 00 |
| 4.5 | Determination of the absolute copy number of <i>B. melitensis</i> IS711 in unknow experimental samples | wn 03 |
| 4.6 | Biochemical tests for B. melitensis, B. abortus and Y. enterocoliticaIdentification1 | 36 |



LIST OF FIGURES

| Figure | Pa | age |
|--------|---|------------|
| 2.1 | The different steps (a) and the exponential amplification in PCR (b) | 38 |
| 2.2 | The SYBR Green I assay | 42 |
| 2.3 | The molecular beacon assay | 44 |
| 2.4 | Hybridization Probes | 46 |
| 2.5 | TaqMan Sequence Detection Chemistry | 48 |
| 3.1 | Optimization of real-time PCR using serial dilution of SYBR Green I dye | 68 |
| 3.2 | Optimization of real time PCR temperature prior to fluorescence reading | 68 |
| 3.3 | Real-time PCR amplification (A) and melting curve analysis (B) with DI obtained from <i>B. melitensis</i> 16M using <i>B. melitensis</i> specific primers un optimized PCR conditions | |
| 3.4 | Sensitivity of the real-time (A) and conventional (B) PCR assays for detect of <i>B. melitensis</i> DNA from pure culture | tion 70 |
| 3.5 | Melting curve analysis of serially diluted B. melitensis DNA template | 72 |
| 3.6 | Specificity of real-time PCR in detecting <i>B. melitensis</i> using <i>B. meliter</i> IS711 specific primers | ısis 74 |
| 3.7 | Confirmation of real-time PCR amplification specificities of <i>B. meliter</i> specific IS711 primers | ısis 74 |
| 3.8 | Reproducibility and real-time PCR efficiency tests | 76 |
| 3.9 | Restriction endonuclease (<i>Eco</i> RI) analysis of the plasmid DNAs extrac from PCR positive transformant colonies | ted 77 |
| 3.10 | The BLAST matches of the recombinant plasmid nucleotide sequences of two Malysian isolates of <i>B. melitensis</i> with the published sequences | the 78 |
| 4.1 | Representative ethidium bromide stained gel analysis for conventional P amplification products of <i>B. melitensis</i> from serum samples of experimenta infected mice collected at different dates of p.i. | |



| 4.2 | Representative melting curve analysis of the amplification curve of live spleen, whole blood and lymph node samples collected from mice in group and E (45 days p.i) together with positive and negative control samples 10 | |
|------|---|----------|
| 4.3 | A standard curve generated from 10-fold dilutions of a known concentration of template DNA 10 | ns)3 |
| 4.4 | Representative amplification curve of serum samples collected from suspects field cases of sheep brucellosis 10 | ed)4 |
| 4.5 | Representative amplification curve of liver, spleen, whole blood and lymp node samples collected from mice in group A (3 mice) and E (1 mouse), (day p.i) 14 | |
| 4.6 | Melting curve analysis of Figure 4.5 | 15 |
| 4.7 | Representative amplification curve of liver, spleen, whole blood and lymp node samples collected from mice in group A (3 mice) and E (1 mouse), (3 days p.i) 14 | |
| 4.8 | Melting curve analysis of Figure 4.7 14 | 16 |
| 4.9 | Representative amplification curve of liver, spleen, whole blood and lymp node samples collected from mice in group A (3 mice) and E (1 mouse), (6 days p.i) 14 | |
| 4.10 | Melting curve analysis of Figure 4.9 14 | 17 |
| 5.11 | Representative amplification curve of liver, spleen, whole blood and lymp node samples collected from mice in group B (2 mice) and E (1 mouse), (day p.i) 14 | (1 |
| 4.12 | Melting curve analysis of Figure 4.11 14 | 18 |
| 4.13 | Representative amplification curve of liver, spleen, whole blood and lymp node samples collected from mice in group B (2 mice) and E (1 mouse), (3 days p.i) 14 | |
| 4.14 | Melting curve analysis of Figure 4.13 | 19 |
| 4.15 | Representative amplification curve of liver, spleen, whole blood and lymp node samples collected from mice in group B (2 mice) and E (1 mouse), (6 days p.i) 15 | |
| 4.16 | Melting curve analysis of Figure 4.15 | 50 |



| 4.17 | Representative amplification curve of liver, spleen, whole blood and lymp node samples collected from mice in group C (2 mice) and E (1 mouse), (days p.i) 15 |
|------|---|
| 4.18 | Melting curve analysis of the Figure 4.17 15 |
| 4.19 | Representative amplification curve of liver, spleen, whole blood and lymp node samples collected from mice in group C (2 mice) and E (1 mouse), (3 days p.i) 15 |
| 4.20 | Melting curve analysis of the Figure 4.19 15 |
| 4.21 | Representative amplification curve of liver, spleen, whole blood and lymp node samples collected from mice in group D (2 mice) and E (1 mouse), (days p.i) 15 |
| 4.22 | Melting curve analysis of the Figure 4.21 15 |
| 4.23 | Representative amplification curve of liver, spleen, whole blood and lymp node samples collected from mice in group D (2 mice) and E (1 mouse) ,(3 days p.i) 15 |
| 4.24 | Melting curve analysis of the Figure 4.23 15 |
| 4.25 | Representative amplification curve of serum samples collected from mice is group A at 1, 7, 15, 30, 45 and 60 days p.i (total 6 samples) 15 |
| 4.26 | Melting curve analysis of Figure 4.25 |
| 4.27 | Representatives of conventional PCR amplification products of <i>B. melitens</i> from liver samples of experimentally infected mice collected at different date of p.i. |
| 4.28 | Representative conventional PCR amplification products of <i>B. melitensis</i> from whole blood samples of experimentally infected mice collected at different dates of p.i. 15 |
| 5.29 | Representative conventional PCR amplification products of <i>B. melitensis</i> from serum samples of experimentally infected mice collected at different dates of p.i. 15 |
| 4.30 | Representative conventional PCR amplification products of <i>B. melitensis</i> from spleen samples of experimentally infected mice collected at different dates of p.i. 15 |



- 4.31 Representative conventional PCR amplification products of *B. melitensis* from lymph node samples of experimentally infected mice collected at different dates of p.i. 158
- 4.32 Representative conventional PCR amplification products of *B. abortus* from liver samples of experimentally infected mice collected at different dates of p.i. 158
- 4.33 Representative conventional PCR amplification products of *B. abortus* from whole blood samples of experimentally infected mice collected at different dates of p.i 159
- 4.34 Representative conventional PCR amplification products of *Y. enterocolitica* O:9 from lymph node samples of experimentally infected mice collected at different dates of p.i. 159
- 4.35 Representative conventional PCR amplification products of *Y. enterocolitica* O:9 from whole blood samples of experimentally infected mice collected at different dates of p.i. 160



LIST OF ABBREVIATIONS AND SYMBOLS

| % | Percentage |
|-------------------|---|
| μg | Microgram |
| μΜ | Micromolar |
| A ₂₆₀ | Absorbance at 260 nm |
| A ₂₈₀ | Absorbance at 260 nm |
| AMOS-PCR | Abortus-melitensis-ovis-suis PCR |
| BLAST | Basic Local Alignment Search Tool |
| bp | Base pair |
| BSA | Bovine serum albumine |
| CDC | Charge Coupled Device |
| cDNA | Complementary deoxyribonucleic acid |
| CFT | Complement Fixation Test |
| cfu | Colony forming units |
| CO ₂ | Carbon dioxide |
| Ct | Threshold cycle |
| CV | Coefficient of variation |
| dATP | Deoxyadenosine triphosphate |
| dCTP | Deoxycytosine triphosphate |
| df/dt | Derivations of fluorescence with respect to temperature |
| dGTP | Deoxyguanosine triphosphate |
| dH ₂ O | Distilled water |
| DNA | Deoxyribonucleic acid |



| dNTP | Deoxynucleotide triphosphate |
|-------------------|---|
| dsDNA | Double-stranded DNA |
| DVS | Department of Veterinary Service |
| EDTA | Ethylenediaminetetraacetic acid disodium salt |
| ELISA | Enzyme-linked immunosorbent assay |
| EtdBr | Ethidium bromide |
| fg | Femto gram (10^{-15}) |
| fig. | Figure |
| FOA | Food and Agriculture Organization of the United Nations |
| FRET | Fluorescence resonance energy transfer |
| g/mol | Gram per mole |
| h | Hour |
| IHK | Institue Haiwan Kluang |
| IMR | Institute of Medical Research |
| INF | Interferon |
| Ip | Intraperitonieally |
| IS | Insertional sequence |
| IU | International unit |
| L | Litter |
| LPS | Lipopolysaccharide |
| Mb | Mega base |
| MgCl ₂ | Magnisium chloride |
| min | Minute |
| mL | Millilitre |



| mM | Millimolar |
|-------|--|
| mol | Mole |
| MZCP | Mediterranean Zoonoses Control Programme |
| ng | Nanogram (10 ⁻⁹) |
| NTC | No template control |
| O.D | Optical density |
| OIE | International Epizootic Office |
| omp | Outer membrane protein |
| ORF | Open reading frame |
| p.i | Post inoculation |
| PBS | Phosphate buffer saline |
| PCR | Polymerase chain reaction |
| pg | Pico gram |
| рр | Pages |
| RBPT | Rose Bengal plate test |
| RNA | Ribonucleic acid |
| Rnase | Ribonuclease |
| rpm | Revolution per minute |
| S | Seconds |
| SD | Standard deviation |
| ssDNA | Single stranded DNA |
| TAE | Tris-acetate-EDTA |
| Taq | Thermus aquaticus |
| TBE | Tris-base EDTA buffer |



| Tm | Melting temperature |
|--------|-------------------------------|
| UPM | Universiti Putra Malaysia |
| UV | Ultraviolet |
| V | Volt |
| v/v | Volume to volume ratio |
| VRI | Veterinary Research Institute |
| w/v | Weight to volume ratio |
| WHO | World Health Organization |
| H_2S | Hydrogen sulfoxide |



CHAPTER 1

INTRODUCTION

Brucellosis, caused by Gram-negative facultative organisms belonging to the genus *Brucella*, is an infectious zoonotic disease that is associated with chronic debilitating infections in humans and reproductive failure in domestic animals (Corbel, 1997). Although brucellosis in livestock and transmission of infection to the human population has been significantly decreased following the instigation of effective vaccination-based control and slaughter programs in parts of the world, it remains an uncontrolled problem in regions of high endemicity such as the Mediterranean, Middle East, Africa, Latin America and parts of Asia. The most recent concern focuses on the potential use of *Brucella* species, primarily *B. melitensis*, as an agent of biological warfare because of the debilitating disease it causes. Widespread dispersal of aerosolized *B. melitensis* would pose a biological, agricultural, as well as an economical threat to all countries involved (Refai, 2002; Kaufmann *et al.*, 1997).

The disease is caused by several *Brucella* species, which are aerobic, non-motile, Gramnegative, non-spore forming, and facultative intracellular coccobacilli. On the basis of 16S rRNA gene sequence comparisons, the genus *Brucella* is grouped in alpha-2 subdivision of the class *Proteobacteria* (Moreno *et al.*, 1990)[•] The genus is divided according to cultural, metabolic, antigenic properties and host specificity in to six species: *B. abortus, B. canis, B. melitensis, B. neotomae, B. ovis*, and *B. suis* (Corbel and Morgan, 1984). In addition, a new strain affecting marine mammals was recently

