



UNIVERSITI PUTRA MALAYSIA

***EVALUATION OF DIABETIC WOUND HEALING PROPERTIES OF  
Moringa oleiferaLAM USING In vitroAND In vivoWOUND MODELS***

ABUBAKAR MUHAMMAD AMALI

FPSK(p) 2015 14



**EVALUATION OF DIABETIC WOUND HEALING PROPERTIES OF  
*Moringa oleifera* LAM USING *In Vitro* AND *In Vivo* WOUND MODELS**

By  
**ABUBAKAR MUHAMMAD AMALI**

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in  
Fulfillment of the Requirements for the Degree of Doctor of Philosophy

September 2015

## **COPYRIGHT**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of University Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



## **DEDICATION**

*This thesis is dedicated to my late father, Alhaji Muhammad Amali. May Almighty Allah forgive him all his short comings and make Jannatul firdaus be his final abode amin.*



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirements for the Degree of Doctor of Philosophy

**EVALUATION OF DIABETIC WOUND HEALING PROPERTIES OF  
*Moringa oleifera* LAM USING *In Vitro* AND *In Vivo* WOUND  
MODELS**

By

**ABUBAKAR MUHAMMAD AMALI**

**September 2015**

**Chair:** Associate Professor Sharida Fakurazi, PhD  
**Faculty:** Medicine and Health Sciences

Diabetic wound is a common complication which affects significant number of people with diabetes. Its treatment is often very difficult which imposes burden and high cost on patients, family and society. Current treatments of diabetic wound are not sufficient enough with limited success in addition to non-affordability. *Moringa oleifera* Lam (*M. oleifera*) from the family *Moringaceae* (genus *Moringa*) commonly called drumstick or horseradish is a plant traditionally employed in the treatment of many ailments and has been scientifically proven to possess hepatoprotective, anti-inflammatory, antioxidant and hypoglycemic action in addition to other numerous activities.

The present study was undertaken to evaluate the potential of *M. oleifera* on wound healing in diabetic condition with a view to providing possible cost effective therapeutic alternative for treating diabetic wound topically. *In vitro* and *in vivo* wound models were utilized for our study.

The study initially demonstrated screening of crude extracts of methanol, ethanol and aqueous from *Moringa oleifera* leaves. Among these three solvent crude extracts, the methanolic crude extract was found to be the most active crude extract following the *in vitro* screening. The most active methanolic extract was then further subjected to bio-assay guided fractionation using hexane, dichloromethane, ethyl acetate, butanol and aqueous. The aqueous fraction was proven to be the most active fraction obtained from the results of *in vitro* screening and bio-guided assay fractionation.

The *in vitro* study included scratch test and proliferation assays using human dermal fibroblast cells (HDF), in which three different solvent crude extracts were screened and the most active methanolic crude extract was further subjected to differential bio graded assay fractionation. The most active aqueous fraction was finally obtained. HPLC, LC-MS/MS and UV spectroscopy were used for the identification and confirmation of bioactive compounds. Kaempferol and quercetin were identified in the crude methanolic extract while an active compound vicenin-2 was identified, confirmed and quantified in the bioactive aqueous fraction. Antioxidant and antibacterial assays were also conducted.

The *in vivo* study involved topical application of the formulated bioactive fraction using full thickness excision wound model in streptozotocin (STZ) and nicotinamide (NAD) induced diabetic rats. Healthy adult male Wistar rats weighing between 150-250g were used. Animals were grouped into six, consisting of six rats in each group (n=6): Two groups of normal and

diabetic controls, three groups of 0.5%, 1% and 2% w/w, aqueous fraction treated and one group of positive control that received 1% w/w silver sulfadiazine as standard drug. Treatments were applied topically in form of cream to the skin wounded area for 21 days. Biophysical, biochemical and histological parameters were evaluated. Proinflammatory cytokines analyses were performed using ELISA, Western blotting and immunohistochemistry techniques. Results were analyzed using SPSS version 20. Data were expressed as mean  $\pm$  standard deviation, and results were selected from at least three independent experiments performed in triplicate. *P*-values of 0.05 were considered to be statistically significant.

The *in vitro* test results demonstrated that, crude methanolic extract and aqueous fraction of *M. oleifera* significantly stimulated proliferation and migration of HDF cells ( $p < 0.05$ ) at 24, 48 and 72 hours after treatment to close the artificially wounded area when compared to untreated control cells. The distance was measured and analyzed quantitatively at time interval of 0, 24, 48 and 72 hrs after the scratch. The MTT assay results showed that, aqueous fraction was relatively non-toxic and did not affect the cellular activity of HDF cells even at concentrations of 800  $\mu$ g/mL after 72 hours. The aqueous fraction was tested and found to be effective in enhancing wound healing *in vitro* through proliferation and migration of human dermal fibroblast cells. In addition, antioxidant and antibacterial activities demonstrated by the bioactive aqueous fraction through radical scavenging and ferric reducing abilities as well inhibition of growth of *S. aureus*, *Ps. aeruginosa* and *E. coli* bacterial pathogens.

Following induction of diabetes by STZ-NAD in Wistar rats, hyperglycemia was maintained for 21 days and the reading of blood glucose level was more significant in diabetic groups compared to normal control group ( $p < 0.05$ ). There was also some form of partial destruction of Islet of Langerhans and some normal islets were seen to be preserved even after the administration of a low dose of STZ and NAD which mimics the type-2 diabetes seen in humans.

The *in vivo* topical applications of various doses (0.5%, 1% and 2%) of bioactive aqueous fractions was found to be effective in enhancing diabetic wound healing through overall decreased wound size, improved wound contraction, enhanced tissue regeneration and granulation tissue, the decrease wound size in diabetic treated groups was more significant compared to untreated diabetic control group ( $P < 0.05$ ) and there was also significant difference in contraction rate between diabetic treated groups compared to untreated diabetic control group ( $P < 0.05$ ).

The topical application of aqueous fraction to the wound of diabetic animals caused down regulation of inflammatory mediators (TNF- $\alpha$ , IL1- $\beta$ , IL-6, iNOS and COX-2) that was very significant in the diabetic treated groups as compared to non-treated diabetic control animals ( $p < 0.05$ ). The up regulation of VEGF protein in the diabetic treated groups was also found to be more significant ( $P < 0.05$ ) compared to untreated diabetic control group that had less expression of VEGF.

Down regulation of inflammatory mediators and up regulation of VEGF with *M. oleifera* aqueous fraction facilitates overall wound healing and closure in diabetic condition. The bioactive compounds present in aqueous fraction have also been successfully identified and confirmed by HPLC and LC-MS/MS using standard vicenin-2 compound. These findings suggested that, topical administration of bioactive aqueous fraction of *M. oleifera* containing Vicenin-2 compound may accelerate wound healing in hyperglycemic condition. Therefore, may serve as a lead in drug discovery for diabetic wound healing.

Abstrak tesis yang dikemukakan kepada senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

**MENILAI SIFAT PENYEMBUHAN *Moringa oleifera* LAM TERHADAP LUKA DIABETIK MENGGUNAKAN MODEL LUKA *In Vitro* DAN *In Vivo***

Oleh

**ABUBAKAR MUHAMMAD AMALI**

**September 2015**

**Pengerusi: Prof. Madya Sharida Fakurazi, PhD**

**Fakulti : Perubatan dan Kesihatan sains**

Rawatan selalunya sangat sukar dan membebankan pesakit, keluarga dan masyarakat dengan kos yang tinggi. Rawatan luka diabetik yang ada sekarang tidak mencukupi kerana kejayaan yang terhad ditambah oleh faktor ketidakmampuan. *Moringa oleifera* Lam (*M. oleifera*) daripada keluarga *Moringaceae* (genus *Moringa*) yang biasanya dipanggil kelor atau tumbuhan remunggai secara tradisi digunakan dalam rawatan pelbagai penyakit dan telah secara saintifiknya terbukti bahan aktif menghalang hati, antiradang, antioksidan dan tindakan hipoglisemik dan lain-lain.

Mula-mula kajian ini menunjukkan ujian ekstrak metanol, etanol dan air mentah daripada daun *Moringa oleifera*. Di antara tiga ekstrak pelarut mentah ini, ekstrak mentah etanol didapati sebagai ekstrak mentah paling aktif berdasarkan ujian *in vitro*. Ekstrak methanol mentah yang aktif ini kemudian menjalani pemeringkatan berpandu asai biologi menggunakan heksana, diklorometana, etil asetat, butanol dan air. Bahagian berair terbukti sebagai bahagian paling aktif yang didapatkan daripada hasil ujian *in vitro* dan pemeringkatan berpandu asai biologi.

Kajian *in vitro* termasuklah ujian calar dan pelbagai asai menggunakan sel-sel dermis fibroblas (SDF/HDF) manusia, yang tiga ekstrak pelarut diuji dan ekstrak paling aktif metanolik telah menjalani pemeringkatan asai gred biologi pembeza. Bahagian berair yang paling aktif akhirnya diperoleh. HPLC, LC-MS/MS dan spektroskopi UV digunakan bagi pengenalan dan pengesahan kompaun bioaktif. Kaempferol dan kuersetin telah dikenal pasti terdapat di dalam ekstrak mentah metanolik manakala kompaun aktif vicenin-2 telah dikenal pasti, disahkan dan didapati di dalam bahagian berair bioaktif. Pengesahan untuk antioksidan dan antibakteria juga dilakukan.

Kajian *in vivo* yang melibatkan aplikasi topikal bahagian bioaktif yang dilakukan menggunakan model eksisi luka penuh terhadap tikus diabetik yang diaruh oleh streptozotosin (STZ) dan nikotinamida (NAD). Tikus jantan Wistar dewasa yang sihat yang beratnya antara 150-250g telah digunakan. Haiwan ini dibahagiakan kepada enam kumpulan, yang mengandungi enam ekor tikus bagi setiap kumpulan (n=6): Dua kumpulan adalah kumpulan kawalan untuk normal dan menghidap diabetik, tiga kumpulan bagi 0.5%, 1% dan 2% w/w, dirawat dengan bahagian berair dan satu kumpulan kawalan positif yang menerima 1% w/w silver sulfadiazine sebagai dadah piawai. Rawatan dijalankan secara topikal dalam bentuk krim disapu pada kawasan yang luka selama 21 hari. Parameter biofizik, biokimia dan histologikal telah dinilai. Analisis pro keradangan sitokon telah

dilakukan menggunakan teknik ELISA, *Western blotting* dan imunohistokimia. Hasil kajian dianalisis menggunakan SPSS versi 20. Data dinyatakan dalam  $\text{min} \pm \text{sisihan piawai}$ , dan hasil telah dipilih daripada sekurang-kurangnya tiga eksperimen berasingan yang dilakukan sebanyak tiga kali. Nilai-*P* adalah 0.05 dianggap signifikan secara statistik.

Hasil ujian *in vitro* menunjukkan, ekstrak mentah metanolik dan bahagian berair *M. oleifera* secara signifikan merangsang peningkatan dan perpindahan sel (SDF/HDF) ( $p < 0.05$ ) dalam tempoh 24, 48 dan 72 jam selepas rawatan dan menutup kawasan luka secara artifisial berbanding sel kawalan yang tidak mendapat rawatan. Jarak telah diukur dan dianalsisi secara kuantitatif pada selang masa 0, 24, 48 dan 72 jam selepas tercalar. Hasil asai MTT menunjukkan, bahagian berair berkeadaan tidak toksid secara relatifnya dan tidak memberi kesan pada aktiviti selular sel SDF/HDF walaupun pada tahap kepekatan 800  $\mu\text{g/mL}$  selepas 72 jam. Bahagian berair telah diuji dan didapati berkesan dalam meningkatkan penyembuhan luka *in vitro* melalui peningkatan dan perpindahan sel dermis fibroblas manusia. Selain itu, aktiviti antioksidan dan antibakteria yang ditunjukkan oleh bahagian berair bioaktif melalui kebolehan hapus sisa radikal dan pengurangan ferik serta perencutan pertumbuhan patogen bakteria *S. aureus*, *Ps. aeruginosa* dan *E. Coli*.

Selepas tikus Wistar diaruh dengan diabetik menggunakan STZ-NAD, hiperglikemia dikekalkan selama 21 hari dan tahap glukos dalam darah meningkat dengan signifikan bagi kumpulan diabetik berbanding kumpulan kawalan yang normal ( $p < 0.05$ ). Selain itu berlaku sedikit kerosakan terhadap kelompok Langerhans dan sebahagian kelompok kelihatan terpelihara walaupun selepas dos rendah STZ dan NAD diberikan bagi menyerupai diabetik jenis-2 pada manusia.

Aplikasi topikal *in vivo* pelbagai dos (0.5%, 1% dan 2%) bahagian berair bioaktif didapati aktif dalam meningkatkan penyembuhan luka diabetik dengan secara keseluruhan mengurangkan saiz luka, pengecutan luka yang bertambah elok, peningkatan pertumbuhan semula tisu dan tisu granulasi, penurunan saiz luka pada kumpulan rawatan diabetik adalah lebih signifikan berbanding kumpulan diabetik tidak dirawat ( $P < 0.05$ ) dan juga terdapat perbezaan yang signifikan dalam kadar pengecutan antara kumpulan rawatan diabetik berbanding kumpulan kawalan tidak dirawat ( $P < 0.05$ ).

Aplikasi topikal bahagian berair terhadap luka haiwan diabetik menyebabkan pengawalaturan menurun pengantara keradangan (TNF- $\alpha$ , IL1- $\beta$ , IL-6, iNOS dan COX-2) yang didapati sangat signifikan dalam kumpulan kawalan diabetik berbanding kumpulan haiwan kawalan diabetik yang tidak dirawat ( $p < 0.05$ ). Pengawalaturan menaik bagi protein VEGF dalam kumpulan kawalan rawatan juga didapati lebih signifikan ( $P < 0.05$ ) berbanding kumpulan kawalan diabetik yang tidak dirawat yang mengandungi kurang VEGF.

Kesemua faktor ini menyumbang kepada keseluruhan penyembuhan dan penutupan luka pesakit diabetik. Kompaun bioaktif yang terdapat dalam bahagian berair juga Berjaya dikenal pasti dan disahkan oleh HPLC dan LC-MS/MS menggunakan piawai kompaun vicenin-2. Dapatkan ini menunjukkan, aplikasi topikal bahagian berair bioaktif *M. oleifera* yang mengandungi kompaun Vicenin-2 dapat mempercepatkan penyembuhan luka dalam keadaan hiperglikemik. Oleh itu, ia membawa kepada penemuan ubat untuk menyembuhkan luka diabetik.

## **ACKNOWLEDGEMENTS**

I wish to begin by thanking Almighty Allah, the Most Gracious and Most Merciful, Who bestow on me the blessings and wisdom to live up to the stage of completing this research project despite many challenges, I pray for His continuous guidance and blessings till the end of my last breath, amin. I will like to express my appreciation to all those who supported me during these years of struggle, in particular: Associate Prof Dr Sharida Binti Fakurazi, my supervisor and advisor, for fine tuning me to scientific research and experimental Pharmacology and letting me share your great knowledge and experience. Your warm and generous support, endless enthusiasm, great concern have been invaluable.

My appreciation goes to my co-supervisors, Associate Prof, Dr Faridah Abas, for excellent advice and support, and for giving me the freedom to combine my Pharmacological knowledge with Phytochemistry research. To Dr Zalinah Ahmad, I remain grateful for sharing exciting sources of inspiration and knowledge in your field of specialty. Dr Cheah Pike See, for excellent advice, professional and skillful technical assistance especially in areas related to histopathology and immunohistochemistry.

Many thanks to Dr Palanisamy Arulselvan for helping me straighten out my question marks in the field of Proteomics and Scientific writing. I wish to also thank all Staff of Laboratory of Vaccine and Immunotherapeutics, for the enormous help, and for creating such a positive and enabling atmosphere in the laboratory.

My colleagues and research mates; Abdulmalik, Aminu, Aimi, Shafinaz, Tina, Karthy, Atiqah, Sani, Aminu, Ashiru, Rabiu, Sani, UDUS Scholars and the Naija community members in UPM for the interactions, sharing of ideas and joyful moments, it's been a pleasure being part of you all. To all my friends and colleagues back home in Nigeria, too numerous to mention, for many years of friendship, I cherish all of you for the wonderful moments we had together.

Many thanks to my brothers and sisters back home in Nigeria, I salute all the encouragements and prayers offered me at all times for the great success, May Allah bless you all, amin.

To my departed father, of blessed memory, May Allah forgive him all his short comings and bless him with jannatul firdaus amin. To my living mother, who has been supportive all the way and gave me all the courage to pursue my interests and always being there for me during challenging moments of my life with non-stop prayers and admonishments, I owe you a debt that can never be paid back, all I will say therefore, May Almighty Allah reward you abundantly and continue to shower His blessings on You here in the world and the hereafter, amin.

To, my dear wife and partner, Hajiya Fatimah, Our children, Aisha and Muhammad, Your great care, invaluable support, patience, love and understanding will remain in my memory for ever and ever. I love you all!

I sincerely thank you all.

I certify that a Thesis Examination Committee has met on 2 September 2015 to conduct the final examination of Abubakar Muhammad Amali on his thesis entitled "Evaluation of Diabetic Wound Healing Properties of *Moringa oleifera* Lam Using *In Vitro* and *In Vivo* Wound Models" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

**Md Zuki bin Abu Bakar @ Zakaria, PhD**

Professor

Institute of Bioscience

Universiti Putra Malaysia

(Chairman)

**Mohamad Aris bin Mohd Moklas, PhD**

Senior Lecturer

Faculty of Medicine and Health Science

Universiti Putra Malaysia

(Internal Examiner)

**Norhaizan binti Mohd Esa, PhD**

Associate Professor

Faculty of Medicine and Health Science

Universiti Putra Malaysia

(Internal Examiner)

**Leung Ping - Chung, PhD**

Professor

Prince of Wales Hospital

Hong Kong

(External Examiner)



---

**ZULKARNAIN ZAINAL, PhD**

Professor and Deputy Dean

School of Graduate Studies

Universiti Putra Malaysia

Date: 5 November 2015

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

**Sharida Fakurazi, PhD**

Associate Professor

Faculty of Medicine and Health Sciences

Universiti Putra Malaysia

(Chairman)

**Faridah Abas, PhD**

Associate Professor

Faculty of Food Science and Technology

Universiti Putra Malaysia

(Member)

**Zalinah Ahmad, PhD**

Associate Professor

Faculty of Medicine and Health Sciences

Universiti Putra Malaysia

(Member)

**Cheah Pike See, PhD**

Senior Lecturer

Faculty of Medicine and Health Sciences

Universiti Putra Malaysia

(Member)

---

**BUJANG BIN KIM HUAT, PhD**

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date:

## **Declaration by graduate student**

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice –Chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity was upheld as according to the University Putra Malaysia (Graduate studies) Rules 2003 (Revision 2012-2013) and the University Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_

Date: \_\_\_\_\_

Name and Matric No: Abubakar Muhammad Amali GS28449

## **Declaration by Members of Supervisory Committee**

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.

Signature: \_\_\_\_\_

Name of  
Chairman of  
Supervisory  
Committee: Sharida Fakurazi, PhD

Signature: \_\_\_\_\_

Name of  
Member of  
Supervisory  
Committee: Faridah Abas, PhD

Signature: \_\_\_\_\_

Name of  
Member of  
Supervisory  
Committee: Zalinah Ahmad, PhD

Signature: \_\_\_\_\_

Name of  
Member of  
Supervisory  
Committee: Cheah Pike See, PhD

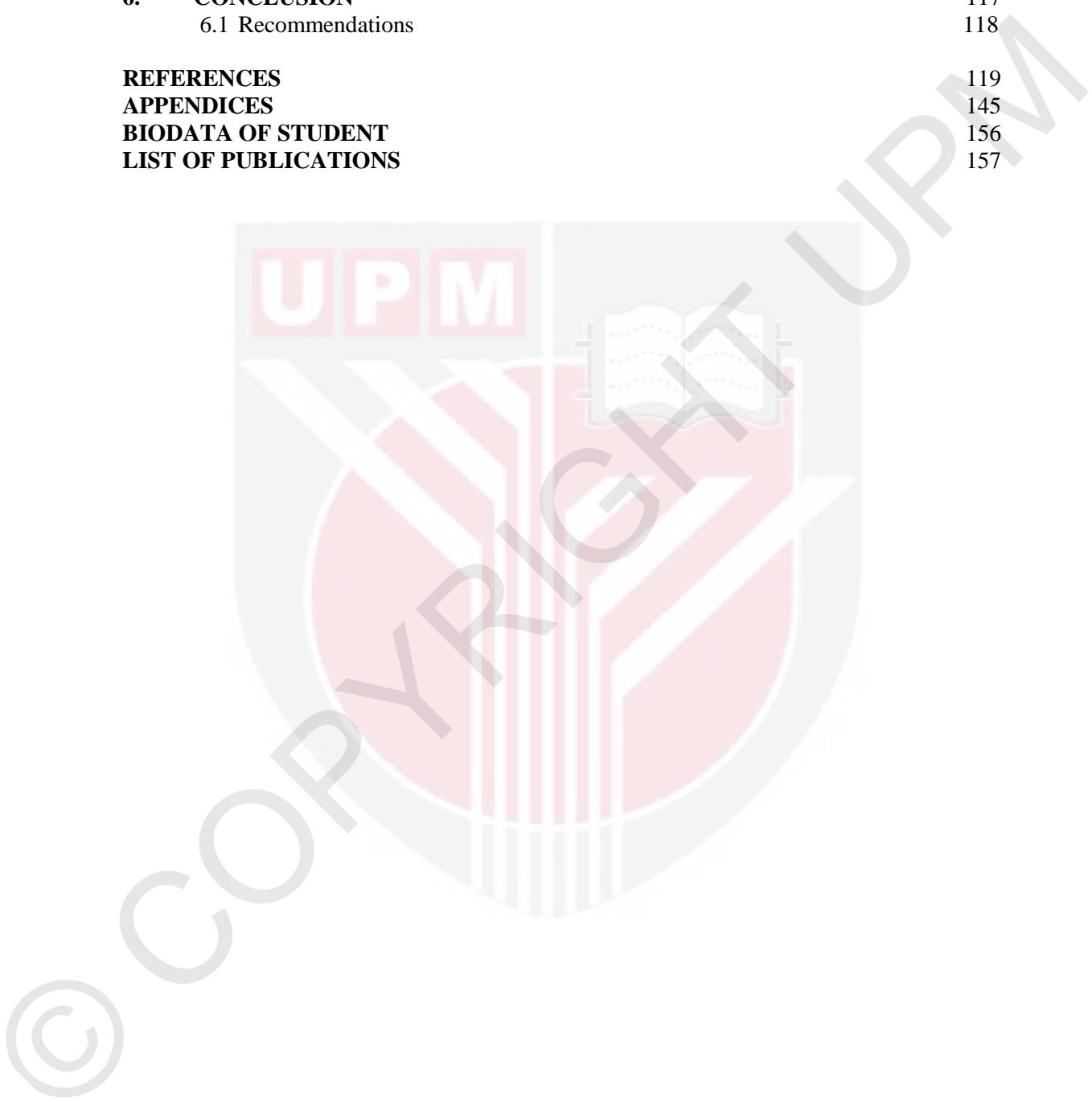
## TABLE OF CONTENTS

	Page
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	iii
<b>ACKNOWLEDGEMENTS</b>	v
<b>APPROVAL</b>	vi
<b>DECLARATION</b>	viii
<b>LIST OF TABLES</b>	xiv
<b>LIST OF FIGURES</b>	xv
<b>LIST OF ABBREVIATIONS</b>	xix
 <b>CHAPTER</b>	
<b>1. INTRODUCTION</b>	1
1.1 Background	1
1.2 Statement of Problem	2
1.3 Justification of the study	2
1.4 Research hypothesis	3
1.5 Objectives of the study	3
1.5.1 General objective	3
1.5.2 Specific objectives	3
1.6 Scope and relevance	4
<b>2. LITERATURE REVIEW</b>	5
2.1 Incidence and prevalence of diabetes	5
2.1.1 Facts and figures	5
2.1.2 Diabetes and its complications	5
2.2 The wound healing process	6
2.3. Wound healing process in diabetes	8
2.4 The role of bacteria in wound healing	9
2.5 The role of antioxidants in wound healing	10
2.6 The role of growth factors and cytokines in wound healing	10
2.7 The study of wounds in a diabetic induced animal model	11
2.8 Current therapy for diabetic wounds and its limitations	13
2.9 Natural products in diabetic wound treatment	16
2.10 <i>Moringa oleifera</i> and its medicinal properties	21
2.11 HPLC-DAD and LCMS/MS techniques in identification of bioactive compounds	25
2.11 Quercetin, kaempferol and Vicenin in diabetic wound healing	25
<b>3. MATERIALS AND METHODS</b>	28
3.1 Chemicals and reagents	29
3.2 Plant extraction and fractionation	29
3.2.1 Determination of yield of freeze dried extracts	30
3.3 Identification of bioactive compounds.	30
3.3.1 LC-MS/MS analysis	30
3.4 Confirmation and quantification of vicenin-2 from bioactive aqueous fraction	30
3.4.1 HPLC analytical method	30
3.4.2 UV/VIS (Visible) spectroscopy	31
3.5 <i>In vitro</i> studies	31

3.5.1	HDF cell lines and culture condition	31
3.5.2	Thawing and culturing of HDF cells	31
3.5.3	Trypsinizing and sub-culturing of HDF cells	31
3.5.4	Maintenance and cryopreservation of HDF cells	32
3.6	<i>In vitro</i> screening of different solvent crude extracts	32
3.6.1	Wound scratch test assay	32
3.6.2	Differential fractionation of active crude methanolic extract	34
3.7I	<i>n vitro</i> screening of different solvent fractions	34
3.7.1	Wound scratch test assay for different solvent fractions	34
3.7.2	HDF cell count assay	34
3.8	MTT colorimetric assay	34
3.9	Phytochemical screening of bioactive aqueous fractions	35
3.10	Antioxidant assay of bioactive aqueous fraction of <i>M. oleifera</i>	36
3.10.1	Determination of total antioxidant capacity of bioactive aqueous fraction	36
3.10.2	Phosphomolybdenum Assay	36
3.10.3	FRAP Assay	36
3.10.4	Free radical scavenging activity assay	37
3.10.5	DPPH radical scavenging activity assay	37
3.10.6	Nitric oxide radical scavenging activity assay	37
3.11	Determination of antibacterial activity of aqueous fraction of <i>M. oleifera</i>	38
3.11.1	Agar well diffusion method	38
3.11.2	Determination of Minimum inhibitory concentration (MIC) of aqueous fraction of <i>M. oleifera</i>	38
3.12	<i>In vivo</i> studies	38
3.12.1	Product formulation	38
3.12.2	Treatment of animals	39
3.12.3	Determination of optimum dosage for chemical induction of diabetes	39
3.12.4	Preparation of serum from whole blood (Thermo Fisher Scientific, USA)	40
3.12.5	Diabetic wound creation	40
3.12.6	Topical application of aqueous fraction to wounded area in animals	40
3.12.7	Assessment of wound area, wound contraction and epithelization period	41
3.12.8	Histological studies of wound healing	42
3.12.9	Molecular studies	42
3.12.9.1	Protein quantification from skin wound tissue	42
3.12.9.2	Evaluation of cytokines levels via ELISA	43
3.12.9.3	Evaluation of cytokines expression by Western blotting	43
3.12.9.4	Determination of cytokines expression by Immunohistochemistry.	45
3.13	Statistical analysis	45
<b>4.</b>	<b>RESULTS</b>	46
4.1	Percentage yields of freeze dried crude extracts and different fractions from active methanolic crude extract	46
4.2	Analysis of bioactive compounds	47
4.2.1	HPLC analysis	47
4.2.2	LCMS/MS analysis	49

4.2.3	Quantification and confirmation of Vicenin 2 from aqueous fraction of <i>M. oleifera</i>	50
4.2.3.1	HPLC analysis	50
4.2.3.2	UV-Vis spectroscopic analysis	52
4.3	<i>In vitro</i> study analysis	53
4.3.1	Wound scratch analysis of different crude extracts	53
4.3.2	Wound scratch test analysis for different solvent fractions from active crude methanolic extract.	58
4.3.3	HDF cells count analysis of different crude extracts of <i>M. oleifera</i>	64
4.3.4	HDF cells count analysis of different fractions from crude methanolic extract.	66
4.4	MTT proliferation assay.	67
4.5	Phytochemical compounds in crude methanolic extracts and aqueous fraction of <i>M. oleifera</i>	69
4.6	Antioxidant activity of aqueous fraction of <i>M. oleifera</i>	69
4.6.1	DPPH radical scavenging capacity	69
4.6.2	Phosphomolybdenum and Ferric reducing potential of aqueous fraction	70
4.6.3	Nitric oxide scavenging activity	71
4.6.4	Hydrogen peroxide scavenging activity	71
4.7	Antibacterial activity of aqueous fraction of <i>M. oleifera</i>	72
4.7.1	Antibacterial activity of aqueous fraction by zone of inhibition.	72
4.7.2	Minimum inhibitory concentration of aqueous fraction of <i>M. oleifera</i>	74
4.8	<i>Invivo</i> study analysis	74
4.8.1.	Blood glucose level following optimized doses of STZ and NAD induced diabetes	74
4.8.2.	Body weight changes following optimized doses of STZ and NAD induced diabetes	75
4.8.3.	Analysis of physiological and biochemical parameters following wound induction and treatment	76
4.8.3.1.	Blood glucose analysis in experimentally induced diabetes in animals	76
4.8.3.2.	Body weight changes following induction of experimental diabetes	77
4.8.3.3.	Serum insulin Level analysis in experimentally induced diabetes animals	78
4.8.3.4.	Rate of feed and water consumption following inductionof diabetes	79
4.9	Effect of aqueous fraction of <i>Moringa oleifera</i> on wound healing parameters	81
4.9.1.	Effect of aqueous fraction of <i>M. oleifera</i> on wound size	81
4.9.2.	Effect of aqueous fraction of <i>M. oleifera</i> on wound contraction	86
4.9.3.	Effect of aqueous fraction of <i>M. oleifera</i> on epithelization period	87
4.9.4.	Effect of aqueous fraction of <i>M. oleifera</i> on granulation tissue	88
4.9.5.	Histological observations of pancreatic tissue following induction of diabetes	89
4.9.6.	Histological observation of wound tissues	92
4.9.7.	Analysis of cytokines level in wound tissue by ELISA	99

4.9.8.	Analysis of cytokines expression by Western blot	101
4.9.9.	Analysis of VEGF protein expression by immunohistochemistry	105
<b>5.</b>	<b>DISCUSSION</b>	109
<b>6.</b>	<b>CONCLUSION</b>	117
	6.1 Recommendations	118
<b>REFERENCES</b>		119
<b>APPENDICES</b>		145
<b>BIODATA OF STUDENT</b>		156
<b>LIST OF PUBLICATIONS</b>		157



## LIST OF TABLES

Table	Page
2.1. Review of some medicinal plants with wound healing activity in diabetic condition.	17
2.2. Review of medicinal uses of <i>Moringa oleifera</i> lam	22
3.1. Animal grouping for determination of optimum dose for chemical induction of diabetes	39
3.2. Animal grouping, wound induction and treatment	41
3.3. Histological features of wound healing	42
4.1. LC-MS/MS spectral data of major compounds identified in methanolic crude extract and bioactive aqueous fraction of <i>M. oleifera</i> leaves detected with mass spectrometry in negative and positive ion modes	49
4.2. Quantitative amount of vicenin-2 contained in the ointment formulation of aqueous fraction of <i>M. oleifera</i>	51
4.3. Preliminary phytochemical screening of crude extracts and aqueou fraction of <i>M. oleifera</i> leaves	69
4.4. Ferric reducing antioxidant capacity (FRAP) and total antioxidant capacity of aqueous fraction of <i>M. oleifera</i> leaves by phosphomolybdenum assay	70
4.6. Antibacterial activity of aqueous fraction of <i>M. oleifera</i> against some bacterial isolates	73
4.6. Minimum inhibitory concentration (MIC) in µg/mL of aqueous fraction of <i>M. oleifera</i> against some bacterial isolates	74
4.7. Semi quantitative histopathological findings of 21 day wound in diabetic control and aqueous fraction treated Wistar rats	92

## LIST OF FIGURES

<b>Figure</b>		<b>Page</b>
2.1.	Picture showing leaves of <i>M. oleifera</i> plant	21
3.1.	Representative workflow for methodology	28
3.2.	Representative workflow for wound scratch test assay	33
4.1 (a).	Percentage of yield of different solvent crude extracts of <i>M. oleifera</i>	46
4.1 (b).	Percentage of yield of different fractions from crude methanolic extract	47
4.2 (a).	HPLC-DAD chromatogram of crude methanolic extract of <i>M. oleifera</i>	48
4.2 (b).	HPLC-DAD chromatogram of aqueous fraction of <i>M. oleifera</i>	48
4.2 (c).	HPLC-DAD chromatogram for reference standard vicenin-2 with retention time of 11mins.	50
4.2 (c).	HPLC-DAD chromatogram for aqueous fraction showing vicenin-2 with retention time of 11mins.	51
4.2 (e).	UV-VIS absorption spectra of the aqueous fraction of <i>M. oleifera</i> compared to that of the standard vicenin-2 compound	52
4.3 (a).	Digital photographs of <i>in vitro</i> scratch assay in normal control HDF cells showing normal migration (x4 magnification).	54
4.3 (b).	Digital photographs of <i>in vitro</i> scratch assay in methanolic crude extract treated HDF cells with faster cell migration as compared with those of other crude extracts (x4 magnification).	55
4.3 (c).	Digital photographs of <i>in vitro</i> scratch assay in ethanolic crude extract treated HDF cells with slower migration as compared to methanolic crude extracts (x4 magnification).	56
4.3 (d).	Digital photographs of <i>in vitro</i> scratch assay in aqueous crude extract treated HDF cells with slower migration as compared to methanolic crude extracts (x4 magnification).	57
4.3 (e).	Graph of analysis of migration of HDF cells treated with different solvent crude extracts by wound scratch assay.	58
4.4 (a).	Digital photographs of <i>in vitro</i> scratch assay in hexane fraction treated HDF cells with slower cell migration as compared to aqueous fraction treated cells (x 4 magnification).	59

4.4 (b).	Digital photographs of <i>in vitro</i> scratch assay in dichloromethane fraction treated HDF with slower cell migration as compared to aqueous fraction treated cells (x 4 magnification).	60
4.4 (c).	Digital photographs of <i>in vitro</i> scratch assay in ethyl acetate fraction treated HDF cells with slower cell migration as compared to aqueous fraction treated cells (x 4 magnification).	61
4.4 (d).	Digital photographs of <i>in vitro</i> scratch assay in butanol fraction treated HDF cells with slower cell migration as compared to aqueous fraction cells (x 4 magnification).	62
4.4 (e).	Digital photographs of <i>in vitro</i> scratch assay in aqueous fraction treated HDF cells with faster cell migration as compared to those of other Fractions (x 4 magnification).	63
4.4 (f).	Graph of analysis of migration of HDF cells treated with different solvent fractions from active crude methanolic extract by wound scratch assay.	64
4.5 (a).	Effect of different different solvent crude extracts of <i>M. oleifera</i> on cell counts of HDF cells at 72 hours after treatment.	65
4.5 (b).	Effect of different different solvent fractions of <i>M. oleifera</i> on cell viability of HDF cells at 72 hours in tryphan blue exclusion assay	65
4.5 (c).	Effect of different different solvent fractions of <i>M. oleifera</i> on cell counts of HDF cells at 72 hours after treatment.	66
4.5 (d).	Effect of different different solvent fractions of <i>M. oleifera</i> on cell viability HDF cells 72 hours after treatment in tryphan blue exclusion assay	67
4.6 (a).	Effects of aqueous fraction of <i>M. oleifera</i> on cell viability of HDF cells after 48hrs using MTT assay	68
4.6 (b).	Effects of aqueous fraction of <i>M. oleifera</i> on cell viability of HDF after 72 hrs using MTT assay	68
4.7 (a).	Graph of DPPH radical scavenging activity of aqueous fraction of <i>M. oleifera</i>	70
4.7 (b).	Graph of nitric oxide scavenging activity of aqueous fraction of <i>M. oleifera</i>	71
4.7 (c).	Graph of hydrogen peroxide scavenging activity of aqueous fraction of <i>M. oleifera</i>	72
4.8 (a).	Graph of blood glucose level in normal control and diabetic induced rats for determination of optimum doses of STZ and NAD	75
4.8 (b).	Graph comparing body weight changes in normal and diabetic induced animals for determination of optimum doses of STZ and NAD	76

4.9 (a).	Glucose level measured at interval for 21 days tretment period .	77
4.9 (b).	Body weight changes following induction of diabetes for 21 days	78
4.9 (c).	Level of insulin measured at interval of 21 days treatment period	79
4.9 (d).	Pattern of feed intake in normal and diabetic rats at interval of 21 days	80
4.9 (e).	Pattern of water consumption in normal and diabetic rats at interval of 21 days.	80
4.10 (a).	Photographs of day 0 excisional wounds induced in treatment and control animals	81
4.10 (b).	Photographs of excisional wounds induced in treatment and untreated control animals on day 3 after wounding	82
4.10 (c).	Photographs of excisional wounds induced in treatment and untreated control animals on day 7 after wounding	83
4.10 (d ).	Photographs of excisional wounds induced in treatment and untreated control on day 14 after wounding	84
4.10 (e).	Photographs of excisional wounds induced in treatment and untreated control animals on day 21 after wounding	85
4.10 (f).	Graph showing decrease wound size caused by topical application of aqueous fraction of <i>M. oleifera</i>	86
4.11	Effects of different doses of aqueous fraction of <i>M. oleifera</i> on wound contraction rate.	87
4.12 (a).	Effects of aqueous fraction of <i>M. oleifera</i> on epithelization period.	88
4.12 (b).	Effects of aqueous fraction of <i>M. oleifera</i> on granulation tissue.	89
4.13 (a).	H & E stained histological sections of rat pancreatic tissue in normal control animal	90
4.13 (b).	H & E stained histological sections of rat pancreatic tissue in diabetic induced animals 21 days after wounding.	91
4.14 (a).	H & E stained sections of wound tissue in normal control rats without treatment	93
4.14 (b).	H & E stained sections of wound tissue in diabetic control rats without Treatment	94
4.14 (c).	H & E stained sections of wound tissue in rats treated with 0.5% aqueous Fraction	95
4.14 (d).	H & E stained sections of wound tissue in rats treated with 1% aqueous Fraction	96

4.14 (e).	H & E stained sections of wound tissue in rats treated with 2% aqueous Fraction	97
4.14 (f).	H & E stained sections of wound tissue in rats treated with 1% silver sulfadiazine (positive control)	98
4.15 (a).	Quantitative analysis of IL-1 $\beta$ production in wound tissue 21 days after treatment with various doses of aqueous fraction of <i>M. oleifera</i> .	99
4.15 (b).	Quantitative analysis of IL-6 production in wound tissue 21 days after treatment with various doses of aqueous fraction of <i>M. oleifera</i> .	100
4.15 (c).	Quantitative analysis of TNF $\alpha$ production in wound tissue 21 days after treatment with various doses of aqueous fraction of <i>M. oleifera</i> .	101
4.16 (a).	Western blot images of COX-2 expression in wound tissues of animals treated with various doses of aqueous fraction of <i>M. oleifera</i> for 21 days	102
4.16 (b).	Western blot images of iNOS expression in wound tissues of animals treated with various doses of aqueous fraction of <i>M. oleifera</i> .	102
4.16 (c).	Western blot images of iNOS expression in wound tissues of animals Treated with various doses of aqueous fraction of <i>M. oleifera</i> .	103
4.16 (d).	Quantitative analysis of COX-2 expression in wound tissue by western blot.	103
4.16 (e).	Quantitative analysis of iNOS expression in wound tissue by western blot.	104
4.16 (f).	Quantitative analysis of VEGF expression in wound tissue by western blot.	105
4.17 (a).	Immunohistochemical staining for VEGF antibody in normal and diabetic control wounds	106
4.17 (b).	Immunohistochemical staining for VEGF antibody in wounds treated with 0.5% (i) and 1 % (ii) aqueous fraction of <i>M. oleifera</i> .	107
4.17 (c).	Immunohistochemical staining for VEGF antibody in wounds treated with 1% aqueous fraction of (v) and 2% silver sulfadiazine (vi)	108

## LIST OF ABBREVIATIONS

ADA	American diabetic association
AGE	Advanced glycation end products
BSA	Bovine serum albumin
BMMSC	Bone marrow derived mesenchymal cells
COX-2	Cyclooxygenase -2
DAB	3,3-diaminobenzene
DMEM	Dulbecco's modified eagle's medium
DFU	Diabetic foot ulcer
ECM	Extracellular matrix;
EDTA	Ethylenediaminetetraacetic acid
EGF	Epidermal growth factor
ELISA	Enzyme-linked immunosorbent assay
EPC	Endothelial progenitor cell
FBS	Fetal bovine serum
FCS	Fetal calf serum
FGF	Fibroblast growth factor
GAG	Glycosaminoglycan
GSH	Reduced glutathione
H & E	Hematoxylin and Eosin
HCl	Hydrochloric acid
HPLC	High performance liquid chromatography
IGF	Insulin-like growth factor
IL-1	Interleukin
IDF	International diabetic federation
i.p	Intraperitoneal
IC <sub>50</sub>	Inhibitory concentration
IL-1 $\beta$	Interleukin 1-beta
IL-6	Interleukin six
KGF	Keratinocyte growth factor
LCMS/MS	Liquid Chromatography-Mass Spectrometry

MDA	Malaysian diabetic association
MIC	Minimum inhibitory concentration
MMP	Matrix metalloproteinase
min	Minute
MTT	(3-[4,5-dimethylthiazol-2-yl]-2,5 diphenyl tetrazolium bromide)
NAD.	Nicotinamide adenine dinucleotide
GF	Growth factor
NO	Nitric oxide
iNOS	Inducible nitric oxide synthase
PBS	Phosphate buffer saline
PDGF	Platelet-derived growth factor
PMN	Polymorphonuclear neutrophil
PVD	Peripheral vascular disease
PVDF	Polyvinylidene difluoride
RAGE	Receptors for Advanced Glycation End Products
ROS	Reactive oxygen specie
SDS	Sodium dodecyl sulfate
SDS-PAGE	Sodium dodecyl sulfate polyacrylamide gel electrophoresis
STZ	Streptozotocin
TEMED	Tetramethylethylenediamine
TGF $\beta$ -R	Transforming Growth Factor $\beta$ Receptor
TGF- $\beta$	Transforming Growth Factor $\beta$
TIMP	Tissue inhibitor of metalloproteinase
TMB	Tetramethylbenzidine
TNF $\alpha$	Tumour Necrosis Factor $\alpha$
UV/VIS	Ultraviolet visible
VEGF	Vascular endothelial growth factor
WHO	World health organization
w/w	Weight/weight
$\mu$ m	Micrometer
%	Percentage
$^{\circ}$ C	Degree celcius

# CHAPTER 1

## INTRODUCTION

### 1.1 Background

The Wound Healing Society defines wounds as physical injuries which result in an opening or break of the skin that disturb normal skin anatomy and function (Strodtbeck, 2001). It results in the loss of epithelium with or without loss of underlying connective tissue (Nagori and Solanki, 2011). Wound healing processes occur in a well-orchestrated combination of multifaceted biological and molecular events of cell migration, cell proliferation and extracellular matrix (ECM) deposition (Fulzele *et al.*, 2002).

As the world is facing epidemic of type 2 diabetes and an increasing incidence of type 1 diabetes (Wild *et al.*, 2004; Gale, 2002; Boulton *et al.*; 2005), the International Diabetes Federation (IDF) focused on global burden of diabetic foot disease as a global concern. People with diabetes have a 12– 25% lifetime risk of developing foot ulcer (Singh *et al.*, 2005; Falanga, 2005). High prevalence rates of diabetes in many countries of the world make foot ulcers a major and increasing public-health problem. Foot ulcers are known to cause substantial morbidity, impair quality of life, endangering high treatment costs (about US\$17 500– 27 987 / UK£9533 15 246) (Falanga, 2005). Since the lifetime risk of a person with diabetes developing a foot ulcer could be as high as 25%, and emergence of contributory pathogenic factors such as neuropathy and vascular disease that are present in more than 10% of people at the time of diagnosis of type 2 diabetes, then the burden of diabetic foot disease is set to increase in future (Group, 1998; Boulton *et al.*, 2005). Even the first year of diagnosis of diabetes could be a period of danger for foot ulcers and amputations (New *et al.*; 2000; Boulton *et al.*, 2005).

Persistent hyperglycemia may negatively affect wound healing and immune system of the body (Brem *et al.*, 2007). In diabetic patients, cell proliferation is often impaired, likely to undergo apoptosis, impairment of blood vessel growth and decreased deposition of collagen at the wound site. These factors contribute significantly to prolongation of injury thereby slowing the process of wound healing in diabetic condition.

Plants and chemical entities derived from plants have been identified and formulated for treatment and management of wounds. More of such herbal products are being investigated to date. Some of the plants that have been tested for diabetic wound healing include *Acalypha langiana* (Perez and Vargas, 2006), *Radix astragalis* and *Radix Rehmannia* (Lau *et al.*, 2009), *Rosmanis officinalis* L (Abu-Al-Basal., 2010), *Curcuma longa* (Sidhu *et al.*, 1999), *Sparassia crispa* (Kwon *et al.*, 2009), *Hylocereus undatus* (Perez *et al.*, 2005), *Momordica charantia* (Teoh *et al.*, 2009), *Lithospermum erythrorhizon* (Fujita *et al.*, 2003), *Aloe vera* (Atiba *et al.*, 2011).

Natural products have been widely used as source of therapeutic agents for the management of human diseases following their safety, efficacy and lesser side effects. The process of wound healing has been shown to be promoted by several natural products (Song and Salcido, 2011). Plants contain different bioactive principles such as alkaloids, flavonoids, tannins, and steroids. These agents may influence one or more phases of wound healing (Ponrasu and Suguna, 2012). Evaluation of various plant products according to their traditional uses and medicinal value based on their therapeutic efficacy leads to the discovery of newer and cost effective drugs for treating various ailments. This form the basis for our study to develop new agent from *M. oleifera* that may be useful in facilitating wound healing in hyperglycemic condition.

## 1.2 Statement of Problem

The underlying causes of risk factors leading to the onset of diabetic foot ulcers which include peripheral neuropathy (lack of sensation in poorly vascularized lower extremities) and can be motor, sensory and autonomic. Dry, stiff skin can crack easily and causes splits that can lead to infection, resulting in cellulitis or ulcerations which potentially leads to ultimate loss of the lower limb. (Tanenberg and Donofrio, 2008). Other factors such as environmental factors, peripheral vascular disease, a compromised immune system and poor metabolic control, in addition to social influences such as emotional, psychological and behavioural problems (Lyons, 2008) are all contributing risks that leads to diabetic ulcer.

One of the complications of diabetes is foot ulcer, known to be the main cause of prolonged hospital stays in developing countries, and are also known to cause substantial morbidity, impair quality of life and engender high treatment costs. The annual treatment cost of diabetic ulcer per patient stood at about US\$17 500– 27 987 / UK£9533– 15 246 (Falang, 2005; Martin *et al.*; (2009).

Despite holistic approach to treatment of diabetic foot ulcer (DFU) which involve tight glucose control and meticulous wound care, the prognosis is often quite poor. This results to amputation and physical disability that can occur even in first year of diagnosis of diabetes (New *et al.*; 2000; Boulton *et al.*, 2005). The lifetime risk of a person with diabetes developing DFU could be as high as 25%, and with the emergence of contributory pathogenic factors such as peripheral neuropathy and vascular disease, the burden of DFU is set to increase in the future as reported by the United kingdom prospective diabetic study (Group, 1998; Boulton *et al.*, 2005). Developing countries in Africa and Asia are more affected by DFU due to high prevalence of type 2 diabetes. For example it was estimated that, 15% of all deaths in South East Asia were attributable to diabetes (IDF, 2011). In Malaysia for example, a 2 year retrospective study (2003-2005) showed that, out of 203 amputated patients at University Sains Malaysia Hospital, 134 (66%) were diabetic related amputations (Yusof *et al.*; 2007).

## 1.3 Justification of the study

The future of diabetic wound treatment lies in research and development of effective agents that facilitates wound healing. Therefore, researchers, medical practitioners,

manufacturers of medicines and most importantly, diabetic patients, would all want to see a breakthrough in this area. The basic understanding of pathogenesis of diabetic wounds and its impaired healing may pave way for the development of novel and cost effective agents that enhances timely healing and closure of wound.

The present study may serve as a lead in discovering agent that promotes wound healing in diabetes. This may help reduce cost of hospitalization and save patients from amputations arising from complications. The study may also lead to better understanding of influence of natural products on chronic wound which forms a platform for further studies that open up possibility of finding alternative therapy for diabetic wound.

It is in the hope of improving these outcomes; we conduct an investigation into bioactive compounds from *Moringa oleifera* which may provide treatment alternatives for diabetic wound and contribute to an increased database of knowledge of phytomedicines in health and diseases.

#### **1.4 Research hypothesis**

The hypothesis to be tested in this study is that, topical application of *Moringa oleifera* enhances wound healing in hyperglycemic condition using *in vitro* and *in vivo* wound models.

#### **1.5 Objectives of the study**

##### **1.5.1 General objective**

The objective of the present study is to evaluate the wound healing efficacy of topical administration of *Moringa oleifera* in diabetes using *in vitro* and *in vivo* experimental wound models.

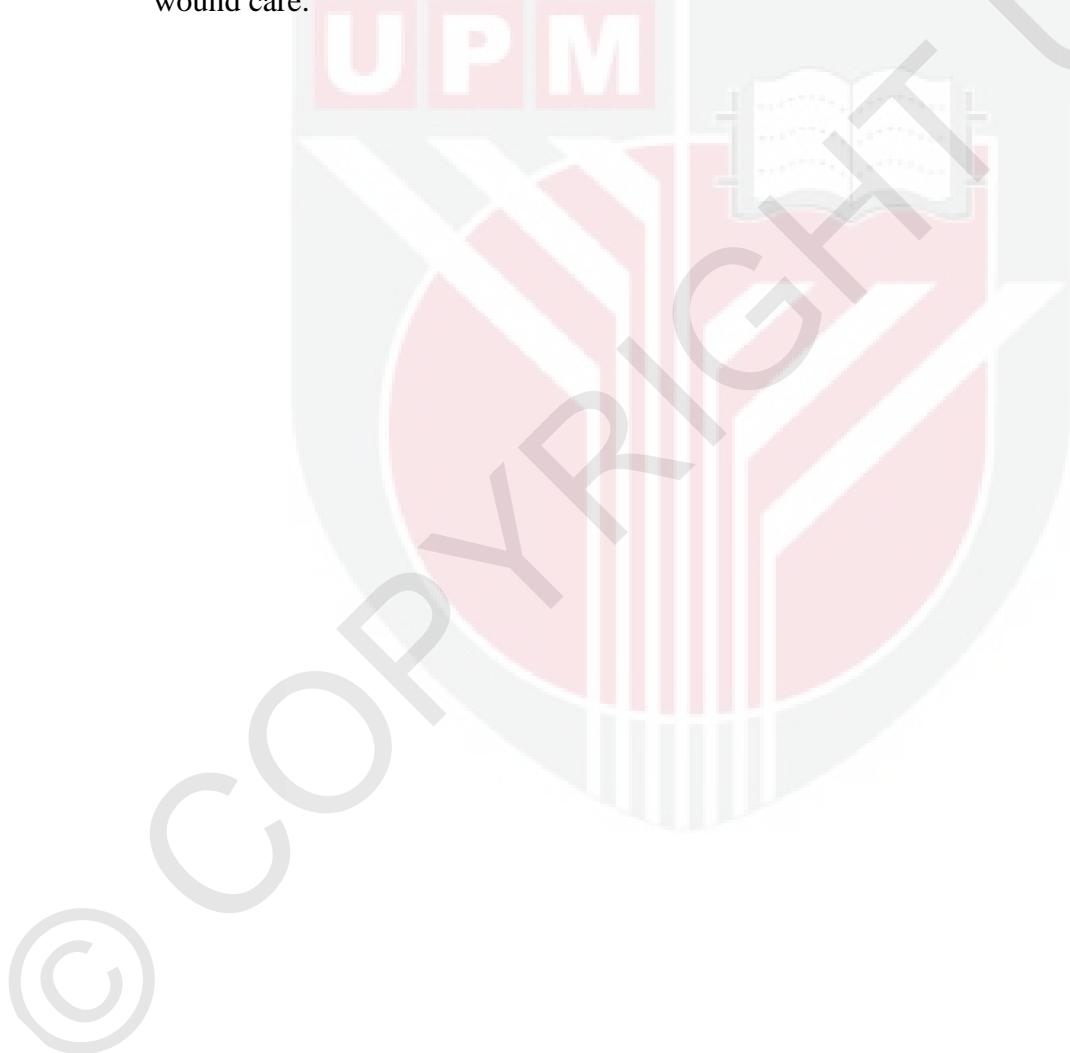
##### **1.5.2 Specific objectives**

- 1) To determine the most active crude extract and fraction of *M. oleifera* using *in vitro* bio-guided assay fractionation.
- 2) To identify and confirm bioactive compounds present in active crude extract and fraction using HPLC-DAD and LC-MS/MS analysis.
- 3) To evaluate the antibacterial activities of bioactive fraction of *M. oleifera*.
- 4) To evaluate the antioxidant activities of bioactive fraction of *M. oleifera*.
- 5) To evaluate wound healing efficacy of bioactive fraction of *M. oleifera* topically in hyperglycemic induced animal model.
- 6) To examine and evaluate histological changes of wound tissue samples in rats, following treatment with bioactive fraction of *M. oleifera*.
- 7) To evaluate expression levels of selected pro-inflammatory cytokines via analysis of wound tissue samples using ELISA, Western blotting and immunohistochemistry techniques.

## **1.6 Scope and relevance**

There is controversy over how to treat poorly healing wounds like diabetic wounds depending on the stage of the wound, topical agents could be beneficial as supported by some evidences like the use of topical antibiotics and a state-of-the-art topical dressing (Sweitzer *et al.*, 2006).

However, the management of diabetic foot is complex, requiring a multifactorial approach and therefore, use of topical application alone may have its own limitations because it is not effective for highly exuding wounds; rapidly absorbs fluid, loses its rheological characteristics, and becomes mobile as it remains on wounds for longer periods of time (Boateng *et al.*, 2008). In these situations topical application has to be considered as an adjunctive therapy to systemic therapy. Although topical therapy is important, it is often insufficient and therefore DFU treatment requires additional wound care.



## REFERENCES

- Abas, F., Shaari, K., Israf, D., Syafri, S., Zainal, Z., Lajis, N. H. (2010). LC– DAD– ESI-MS analysis of nitric oxide inhibitory fractions of *Tenggek burung melicope ptelefolia champ. ex benth.* *Journal of Food Composition and Analysis*, 23(1), 107-112.
- Abbas, Z. G. (2013). The Global Burden of Diabetic Foot. *Contemporary Management of the Diabetic Foot, 1*.
- Abu-Al-Basal, M. A. (2010). Healing potential of *Rosmarinus officinalis* L. on full-thickness excision cutaneous wounds in alloxan-induced-diabetic BALB/c mice. *Journal of Ethnopharmacology*, 131(2), 443-450.
- Adetutu, A., Morgan, W. A., Corcoran, O. (2011). Antibacterial, antioxidant and fibroblast growth stimulation activity of crude extracts of *Bridelia ferruginea* leaf, a wound-healing plant of Nigeria. *Journal of ethnopharmacology*, 133(1), 116-119.
- Agyare, C., Dwobeng, A. S., Agyepong, N., Boakye, Y. D., Mensah, K. B., Ayande, P. G., Adarkwa-Yiadom, M. (2013). Antimicrobial, Antioxidant, and Wound Healing Properties of *Kigelia africana* (Lam.) Beneth. and *Strophanthus hispidus* DC. *Advances in pharmacological sciences*, 2013.
- Ahmad, H. Z. (2014). Diabetes and Diabetic Foot in Malaysia, A presentation made at the 11<sup>th</sup> Asia Pacific conference on diabetic limbs. Monash University Parkville Campus, Malaysia.
- Ajay, M., Achike, F. I., Mustafa, A. M., Mustafa, M.R. (2006). Effect of quercetin on altered vascular reactivity in aortas isolated from streptozotocin-induced diabetic rats. *Diabetes Research and Clinical Practice* 73: 1– 7
- Akkol, E. K., Acikara, O. B., Suntar, I., Enhancement of wound healing by topical application of *Scorzonera* species: Determination of the constituents by HPLC with new validated reverse phase method. *Journal of ethnopharmacology*, 137(2), 1018-1027.
- Ambiga, S., Narayanan, R., Gowri, D., Sukumar D., Madhavan, S. (2007) “Evaluation of wound healing activity of Ipomoea carnea tivit jacq,” *Ancient Science of Life*, 26(3), 45.
- Ambiga, S., Narayanan, R., Gowri, D., Sukumar, D., Madhavan, S. (2007). Evaluation of wound healing activity of flavonoids from *Ipomoea carnea* jacq. *Ancient science of Life*, 26(3), 45.
- American Diabetes Association. (2008). Diagnosis and classification of diabetes mellitus. *Diabetes care*, 31(Supplement 1), S55-S60.

Animal models for wound repair. *Archives of dermatological Research*. 290 Suppl: S1-11.

Anokwuru, C. P., Anyasor, G. N., Ajibaye, O., Fakoya, O., Okebugwu, P. (2011). Effect of Extraction Solvents on Phenolic, Flavonoid and Antioxidant activities of Three Nigerian Medicinal plants. *Nature and Science*, 9(7), 53-61.

Ashok Kumar, N., Pari, L. (2003). Antioxidant action of *Moringa oleifera* Lam.(drumstick) against antitubercular drugs induced lipid peroxidation in rats. *Journal of Medicinal Food*, 6(3), 255-259.

Association, A. D. (2011). Standards of medical care in diabetes. *Diabetes Care*, 34(Suppl 1), S11-S61.

Atiba, A., Ueno, H., Uzuka, Y. (2011). The effect of *Aloe vera* oral administration on cutaneous wound healing in type 2 diabetic rats. *The Journal of Veterinary Medical Science / the Japanese Society of Veterinary Science*, 73(5), 583-589.

Atsukwei, D., Eze, E. D., Adams, M. D., Adinoyi, S. S., Ukpabi, C. N. (2014). Hypolipidaemic Effect of Ethanol Leaf Extract of *Moringa oleifera* Lam. in experimentally induced Hypercholesterolemic Wistar Rats. *International Journal of Nutrition and Food Sciences*, 3(4), 355.

Bahado-Singh, P. S., Riley, C. K., Lowe, H. I., Watson, C. T., Wheatley, A. O. B., Morrison, E. S. A. Y. (2014). Wound healing potential of *Tillandsia recurvata* and *Guaiacum officinale* in streptozotocin induced Type 1 Diabetic rats. *American Journal of Biomedical and Life Sciences*, 2(6), 146-149.

Balasubramani, M., Kumar, T. R., Babu, M. (2001). Skin substitutes: A review. *Burns*, 27(5), 534-544.

Bao, P., Kodra, A., Tomic-Canic, M., Golinko, M. S., Ehrlich, H. P., Brem, H. (2009). The role of vascular endothelial growth factor in wound healing. *Journal of Surgical Research*, 153(2), 347-358.

Basso, F. G., Pansani, T. N., Turrioni, A. P. S., Bagnato, V. S., Hebling, J., de Souza Costa, C. A. (2012). In vitro wound healing improvement by low-level laser therapy application in cultured gingival fibroblasts. *International journal of dentistry*, 2012.

Bastaki, S. (2005). Review diabetes mellitus and its treatment, *International Journal of Diabetes and Metabolism*, 13(10), 111– 134.

Basu, S., Stuckler, D., McKee, M., Galea, G. (2012). Nutritional determinants of worldwide diabetes: An econometric study of food markets and diabetes prevalence in 173 countries. *Public Health Nutr*, 13, 1-8.

- Belliraj, T. S., Nanda, A., Ragunathan, R. (2015). *In-vitro hepatoprotective activity of Moringa oleifera mediated synthesis of gold nanoparticles*. *Journal of Chemical & Pharmaceutical Research*, 7(2).
- Benzie, I. F., Strain, J. (1996). The ferric reducing ability of plasma (FRAP) as a measure of “antioxidant capacity” in *Biochemistry*, 239(1), 70-76.
- Bergin, S., Wright, P. (2006). Silver based wound dressings and topical agents for treating diabetic foot ulcers. *Cochrane Database of Systematic Reviews*, (1), Art. No.: CD005082. DOI: 10.1002/14651858.CD005082.
- Berkovich, L., Earon, G., Ron, I., Rimmon, A., Vexler, A., Lev-Ari, S. (2013). *Moringa oleifera* aqueous leaf extract down-regulates nuclear factor-kappaB and increases cytotoxic effect of chemotherapy in pancreatic cancer cells. *BMC complementary and alternative medicine*, 13(1), 212.
- Besse, J. L., Leemrijse, T., Deleu, P. A. (2011). Diabetic foot: the orthopaedic surgery angle. *Orthopaedics & Traumatology: Surgery & Research*, 97(3), 314-329.
- Bhraa l i , R. , Tabassum, J. , Azad Moringa ( 2 0 0 3 ) oleifera Lam, on hepatic carcinogen metabolizing enzymes, antioxidant parameters and skin Asian Pacific Journal of n e s i s Cancer Prevention, 4(2), 131-139.
- Bishop, J. B., Phillips, L. G., Mustoe, T. A., VanderZee, A. J., Wiersema, L., Roach, D. E., Robson, M. C. (1992). A prospective randomized evaluator-blinded trial of two potential wound healing agents for the treatment of venous stasis ulcers. *Journal of vascular surgery*, 16(2), 251-257.
- Biswas, T. K., Maity, L. N., Mukherjee, B. (2004). Wound healing potential of *Pterocarpus santalinus* Linn: a pharmacological evaluation. *The international journal of lower extremity wounds*, 3(3), 143-150.
- Blair, S. D., Backhouse, C. M., Wright, D. D. I., Riddle, E., McCollum, C. N. (1988). Do dressings influence the healing of chronic venous ulcers? *Phlebology*, 3(2), 129-134. Blois, M.S. (1958). Antioxidant determinations by the use of a stable free radical, *Nature*, 181, 1199- 1200.
- Boateng, J. S., Matthews, K. H., Stevens, H. N., and Eccleston, G. M. (2008). Wound healing dressings and drug delivery systems: A review. *Journal of Pharmaceutical Sciences*, 97(8), 2892-2923.
- Boulton A (2005). The diabetic foot: Epidemiology, risk factors, and the status of care. *Diabetes VOICE*, 50 (SI):5-7
- Boulton, A. J. M., Cavanagh, P. R., Rayman, G., (eds). *The Foot in Diabetes*, 4th edn. Wiley and Sons Ltd, Chichester, UK, 2006.

- Boulton, A. J. M., Vileikyte, L., Ragnarson-Tennvall, G., Apelqvist, J. (2005). The global burden of diabetic foot disease. *The Lancet*, 366(9498), 1719-1724.
- Breksa III, A. P., Hidalgo, M. B., Yuen, M. L. (2009). Liquid chromatography-electrospray ionisation mass spectrometry method for the rapid identification of *Citrus limonoid* glucosides in citrus juices and extracts. *Food Chemistry*, 117(4), 739-744.
- Brem, H., Balleux, J., Bloom, T., Kerstein, M. D., Hollier, L. (2000). Healing of diabetic foot ulcers and pressure ulcers with human skin equivalent: a new paradigm in wound healing. *Archives of surgery*, 135(6), 627-634.
- Brem, H., Tomic-Canic, M. (2007). Cellular and molecular basis of wound healing in diabetes. *Journal of Clinical Investigation*, 117(5), 1219-1222.
- Bryan, D., Walker, K. B., Ferguson, M., Thorpe, R. (2005). Cytokine gene expression in a murine wound healing model. *Cytokine*, 31(6), 429-438.
- Buckley, S. C., Scott, S., Das, K. (2000). Late review of the use of silver sulphadiazine dressings for the treatment of fingertip injuries. *Injury*, 31(5), 301-304.
- Bukar, A., Uba, A., Oyeyi, T. (2010). Antimicrobial profile of *Moringa oleifera* Lam. extracts against some food- borne microorganisms *Bayero Journal of Pure and Applied Sciences*, 3(1).
- Chaby, G., Senet, P., Vaneau, M., Martel, P., Guillaume, J. C., Meaume, S., Chosidow, O. (2007). Dressings for acute and chronic wounds: a systematic review. *Archives of dermatology*, 143(10), 1297-1304.
- Chan, J. C., Malik, V., Jia, W., Kaduwaki, T., Yajnik, C. S., Yoon, K. H., Hu, F. B. (2009). Diabetes in Asia: epidemiology, risk factors, and pathophysiology. *Jama*, 301(20), 2129-2140.
- Chatzigeorgiou, A., Halapas, A., Kalafatakis, K., Kamper, E. (2009). The use of animal models in the study of diabetes mellitus. *In Vivo*, 23(2): 245-258.
- Chithra, P., Sajithlal, G. B., Chandrakasan, G. (1998). Influence of *Aloe vera* on the healing of dermal wounds in diabetic rats. *Journal of ethnopharmacology*, 59(3), 195-201.
- Cho, H. J., Chang, Y. C. (2014). Extract of Moringa Root Inhibits PMA-induced Invasion of Breast Cancer Cells. *Journal of Life Science*, 24(1), 8-13.
- Choi, J. S., Leong, K. W., Yoo, H. S. (2008). *In vivo* wound healing of diabetic ulcers using electrospun nanofibers immobilized with human epidermal growth factor (EGF). *Biomaterials*, 29(5), 587-596.

- Choudhary, M. K., Bodakhe, S. H., Gupta, S. K. (2013). Assessment of the antiulcer potential of *Moringa oleifera* root-bark extract in rats. *Journal of acupuncture and meridian studies*, 6(4), 214-220.
- Chuang, P. H., Lee, C. W., Chou, J. Y., Murugan, M., Shieh, B. J., Chen, H. M. (2007). Anti-fungal activity of crude extracts and essential oil of *Moringa oleifera* Lam. *Bioresource Technology*, 98(1), 232-236.
- Chulani., H.L. (1996) In: The law of medical negligence 1st edn. *Radhakrishnan Medical and Educational Trust: Mumbai*, 51-83.
- Cirico, T., Omaye S. T (2006) Additive or synergistic effects of phenolic compounds on human low density lipoprotein oxidation. *Food and Chemical Toxicology* 44: 510– 516
- Clakar R. A. (2001) . “ *Annals of the New York Academy of Sciences*, 936(1), 355-367.
- Clericuzio M, Tinello S, Burlando B, Ranzato E, Martinotti S, Cornara L, La Rocca A (2012). Flavonoid oligoglycosides from *Ophioglossum vulgatum* L. having wound healing properties. *Planta Medica* 78(15), 1639-1644.
- Colagiuri, R., Eigenmann, C. A. (2009). A national consensus on outcomes and indicators for diabetes patient education. *Diabetic Medicine*, 26 (4), 442-446.
- Cooper, R. (2004). A review of the evidence for the use of topical antimicrobial agents in wound care. *Worldwide wounds*, 1-11.
- Cruciani, M., Lipsky, B. A., Mengoli, C., De Lalla, F. (2005). Are granulocyte colony- stimulating factors beneficial in treating diabeticfoot infections? A meta-analysis. *Diabetes Care*, 28(2), 454-460.
- Dabai, Y., Kawo, A., Aliyu, R. (2012). Phytochemical screening and antibacterial activity of the leaf and root extracts of *Senna italic*. *African Journal of Pharmacy. Pharmacology*, 6(12), 914-918.
- Dahanukar, S.A., Kulkarni, R.A., Rege, N.N (2000). Pharmacology of medicinal plants and natural products. *Indian Journal of Pharmacology*. 32, S81-S118.
- Danaei, G., Finucane, M. M., Lu, Y., Singh, G. M., Cowan, M. J., Paciorek, C. J., et al., (2011). National, regional, and global trends in fasting plasma glucose and diabetes prevalence since 1980: Systematic analysis of health examination surveys and epidemiological studies with 370 country-years and 2· 7 million participants. *The Lancet*, 378(9785), 31-40.
- Dash, G. K., Murthy, P. N. (2011). Studies on wound healing activity of *Heliotropium indicum* Linn. leaves on rats. *ISRN pharmacology*, 2011. Davidson JM (1998).

Department of Statistics, Malaysia (2012). Population projections, Malaysia. 2010-2040. *Department of statistics, Malaysia*.

Deribe, B., Woldemichael, K., Nemera, G. (2014). Prevalence and Factors Influencing Diabetic Foot Ulcer among Diabetic Patients Attending Arbaminch Hospital, South Ethiopia. *J Diabetes Metab* 2 (322): 2

do Nascimento P. M, Pinheiro A. L, Salgado M. A, Ramalho L. M. (2004). A preliminary report on the effect of laser therapy on the healing of cutaneous surgical wounds as a consequence of an inversely proportional relationship between wavelength and intensity: histological study in rats. *Photomedicine and Laser Surgery*. 22 (6), 513-518.

Dorresteijn, J. A., Kriegsman, D. M., Assendelft, W. J., Valk, G. D. (2010). Patient education for preventing diabetic foot ulceration. *The Cochrane Library*.

Dorsett-Martin, W. A. (2004). Rat models of skin wound healing: a review. *Wound Repair and Regeneration*, 12(6), 591-599.

Doupijs VeJves A (2008). Classification, dia...  
ulcers. *Wounds*. 20:117– 26.

Dow, G., Browne A., Sibbald, R.G. (1999). Infection in chronic wounds: controversies in diagnosis and treatment of Ostomy *Wound Management* 45, 23– 40.

Drosou, A., Falabella A., Kirsner R.S. (2003). Antiseptics on wounds: an area of controversy. *Wounds*. 15(5), 149-66.

Dunford, C., Cooper R., Molan, P., (2000). Using honey as a dressing for infected skin lesions. *Nursing Times*; 96(14 Suppl), 7-9.

Edwards, R., and Harding, K. G. (2004). Bacteria and wound healing. *Current opinion in infectious diseases*, 17(2), 91-96.

Eming, S. A., Krieg, T., Davidson, J. M. (2007). Inflammation in wound repair: molecular and cellular mechanisms. *Journal of Investigative Dermatology*, 127(3), 514-525.

Enoch, S., and Price, P. (2004). Cellular, molecular and biochemical differences in the pathophysiology of healing between acute wounds, chronic wounds and wounds in the aged. *World Wide Wounds*, 1-16.

Evans, W. C. (2009). *Trease and evans' Pharmacognosy*, Elsevier Health Sciences.

Ezealisiji, K. M., Omotosho, A. E., Udo, R. U. T. H., Agbo, M. O. (2014). Wound healing activity of n-hexane and methanol extracts of *Tetracarpidium conophorum* (Mull. Arg) Hutch (African walnut) in Wistar rats. *Malays J Pharm Sci. Malay*, 12(1), 79-88.

- Fabry W, Okemo, PO, Ansorg R. (1998). Antibacterial activity of East African medicinal plants. *Journal of Ethnopharmacology*. 60, 79-84.
- Fahey, J. W., Zalcmann, A.T., Talalay P. (2001). The chemical diversity and distribution of glucosinolates and isothiocyanates among plants. *Phytochemistry* 56:5– 51.
- Fahey, J. W., (2005). *Moringa oleifera*: A review of the medical evidence for its nutritional, therapeutic, and prophylactic properties. Part 1. *Phytochemistry*, (47), 123,157.
- Faizi, S., Siddiqui, B. S., Saleem, R., Aftab, K., Shaheen, F., Gilani, A. U. H. (1998). Hypotensive constituents from the pods of *Moringa oleifera*. *Planta Medica*, 64(3), 225-228.
- Fakhry S. M., Alexander J., Smith, D. (1995). Regional and institutional variations in burn care. *Journal Burn Care and Rehabilitation*, 16, 8690.
- Fakurazi, S., Sharifudin, S. A., Arulselvan, P. (2012). *Moringa oleifera* hydroethanolic extracts effectively alleviate acetaminophen-induced hepatotoxicity in experimental rats through their antioxidant nature. *Molecules*, 17(7), 8334-8350.
- Falanga, V. (2005). Wound healing and its impairment in the diabetic foot. *The Lancet*, 366(9498), 1736-1743.
- Faries, P. L., Teodorescu, V. J., Morrissey, N. J., Hollier, L. H., Marin, M. L. (2004). The role of surgical revascularization in the management of diabetic foot wounds. *Am J Surg*, 187:34S-37S.
- Faries, P. L., Brophy, D., LoGerfo, F. W., Akbari, C. M., Campbell, D. R., Spence, L. D., Pomposelli, F. B. (2001). Combined iliac angioplasty and infrainguinal revascularization surgery are effective in diabetic patients with multilevel arterial disease. *Annals of vascular surgery*, 15(1), 67-72.
- Faries, P. L., Teodorescu, V. J., Morrissey, N. J., Hollier, L. H., Marin, M. L. (2004). The role of surgical revascularization in the management of diabetic foot wounds. *The American journal of surgery*, 187(5), S34-S37.
- Farjana, N., Saud, Z. A., Rahman, M. H., Haque., M. E. (2003). *In vitro* antimicrobial activity of the compound isolated from chloroform extract of *Moringa oleifera* Lam.
- Ferrara, N., and Henzel, W. J. (1989). Pituitary follicular cells secrete a novel heparin-binding growth factor specific for vascular endothelial cells. *Biochemical and Biophysical Research Communications*, 161(2), 851-858.
- Frankel, Y. M., Melendez, J. H., Wang, N.Y., Price, L.B., Zenilman, J. M., Lazarus, G. S. (2009). Defining wound microbial flora: molecular microbiology opening new horizons. *Archives of Dermatology*. 145, 1193– 1195.

- Fujita, N., Sakaguchi, I., Kobayashi, H., Ikeda, N., Kato, Y., Minamino, M., Ishii, M. (2003). An extract of the root of *Lithospermum erythrorhizon* accelerates wound healing in diabetic mice. *Biological and Pharmaceutical Bulletin*, 26(3), 329-335.
- Fulzele, S., Satturwar, P., Joshi, S., Dorle, A. (2002). Wound healing activity of *Hingvadya ghrita* in rats. *Indian Drugs*, 39(11), 606-609.
- Gal, P., Kilik, R., Mokry, M., Vidinsky, B., Vasilenko, T., Mozes, S., Lenhardt, L. (2008). Simple method of open skin wound healing model in corticosteroid-treated and diabetic rats: standardization of semi-quantitative and quantitative histological assessments. *Veterinarni Medicina*, 53(12), 652-659.
- Gale, E. A. (2002). The rise of childhood type 1 diabetes in the 20th century. *Diabetes*, 51(12), 3353-3361.
- Galiano, R. D., Michaels, V., Dobryansky, M., Levine, J. P., Gurtner, G. C. (2004). Quantitative and reproducible murine model of excisional wound healing. *Wound repair and regeneration*, 12 (4), 485-492.
- Gao, J., Bai, J., Man, Q., Liu, G. (2000). [Mechanism of silk hydrates on modulating the blood glucose metabolism in rats with experimental diabetes]. *Wei sheng yan jiu= Journal of hygiene research*, 29(6), 379-382.8.
- Garg, H. G. (2000) Scarless wound healing. New York: Marcel Dekkar Inc. Electronic book.
- Giacco, F., Brownlee, M. (2010). Oxidative stress and diabetic complications. *Circulation research*, 107(9), 1058-1070.
- Giacco, F., Brownlee, M. (2010). Oxidative stress and diabetic complications. *Circulation research*, 107(9), 1058-1070.
- Gibbons, S., (2005). Plants as a source of bacterial resistance modulators and antiinfective agents. *Phytochemical Reviews* 4, 63– 78.
- Gonsalves, C. F. Venous leg ulcers (2003). Techniques in vascular and interventional radiology. 6: 132-136.
- Gottrup, F., Ågren, M. S., Karlsmark, T. (2000). Models for use in wound healing research: A survey focusing on *in vitro* and *in vivo* adult soft tissue. *Wound Repair and Regeneration*, 8(2), 83-96.
- Gowda, A., Shanbhag, V., Shenoy, S., Bangalore, E. R., PrabhuK, M. R. (2013). Wound healing property of topical application of ethanolic extract of *Michelia champaca* flowers in diabetic rats. *International Journal of Pharmacology and Clinical Sciences*, 2(3), 67-74.
- Grayer, R. J., Kite, G. C., Veitch, N. C., Eckert, M. R., Marin, P. D., Senanayake, P., Paton, A. J. (2002). Leaf flavonoid glycosides as chemosystematic characters in *Ocimum*. *Biochemical systematics and ecology*, 30(4), 327-342.

- Grebe, S. K., Singh, R. J. (2011). LC-MS/MS in the Clinical Laboratory – Where to From Here? *The Clinical Biochemist Reviews*, 32(1), 5– 31.
- Grinnell, F., Zhu, M. (1994). Identification of neutrophil elastase as the proteinase in burn wound fluid responsible for degradation of fibronectin. *Journal of investigative dermatology*. 103(2), 155-161.
- Group, U. K., (1998). *Prospective Diabetes Study*: Tight blood pressure control and risk of macrovascular and microvascular complications in type 2 diabetes: UKPDS 38. *BMJ: British Medical Journal*, 703-713.
- Guo, S., DiPietro, L. A. (2010). Factors affecting wound healing. *Journal of Dental Research*, 89(3), 219-229.
- Gupta, R., Mathur, M., Bajaj, V. K., Katariya, P., Yadav, S., Kamal, R., Gupta, R. S. (2012). Evaluation of antidiabetic and antioxidant activity of *Moringa oleifera* in experimental diabetes. *Journal of diabetes*, 4(2), 164-171.
- Gutierrez, R. P. (2006). Evaluation of the wound healing properties of *Acalypha langiana* in diabetic rats. *Fitoterapia*, 77(4), 286-289.
- Haghpanah, S., Bogie, K., Wang, X., Banks, P.G., Ho, C.H. (2006). Reliability of electronic versus manual wound measurement techniques. *Arch Phys Med Rehabil.* 87(10):1396– 1402
- Hamza, A. A. (2010). Ameliorative effects of *Moringa oleifera* lam seed extract on liver fibrosis in rats. *Food and Chemical Toxicology*, 48(1), 345-355.
- Harborne, J. B. (1998). Phytochemical methods a guide to modern techniques of plant analysis. *Springer Science & Business Media*.
- Hashim, N. H. N., Abas, F., Shaari, K., Lajis, N. H. (2012). LC- DAD- ESIMS/MS characterization of antioxidant and anticholinesterase constituents present in the active fraction from *Persicaria hydropiper*. *LWT-Food Science and Technology*, 46(2), 468-476.
- Hayashi, K., Kojima, R., Ito, M. (2006). Strain differences in the diabetogenic activity of streptozotocin in mice. *Biological and Pharmaceutical Bulletin*, 29(6), 1110-1119.
- Hayashi, T., Ishida, Y., Kimura, A., Takayasu, T., Eisenmenger, W., Kondo, T. (2004). Forensic application of VEGF expression to skin wound age determination. *International Journal of Legal Medicine*, 118(6), 320-325.
- Helm, C. L. E., Fleury, M. E., Zisch, A. H., Boschetti, F., Swartz, M. A. (2005). Synergy between interstitial flow and VEGF directs capillary morphogenesis *in vitro* through a gradient amplification mechanism. *Proceedings of the National Academy of Sciences of the United States of America*, 102(44), 15779-15784.

- Hinchliffe, R. J., Valk, G. D., Apelqvist, J., Armstrong, D. G., Bakker, K., Game, F. L., Jeffcoate, W. J. (2008). Specific guidelines on wound and wound-bed management. *Diabetes/metabolism research and reviews*, 24(S1), S188-S189.
- Hirsch, T., Spielmann, M., Zuhaili, B., Koehler, T., Fossum, M., Steinau, H. U., Eriksson, E. (2008). Enhanced susceptibility to infections in a diabetic wound healing model. *BMC surgery*, 8(1), 5.
- Ho, E., Chen, G., Bray, T. M. (2000). Alpha-Phenyl-tertbutylnitrone (PBN) inhibits NF κ B activation protects against chemically induced diabetes. *Free Radical Biology and Medicine*, 28(4): 604-614.
- Hoffman, S (1984). Hoffmann, S. (1984). Silver sulfadiazine: an antibacterial agent for topical use in burns. *Scandinavian Journal of Plastic and Reconstructive Surgery and Hand Surgery*, 18(1), 119-126.
- Hong, Y. K., Lange-Asschenfeldt, B., Velasco, P., Hirakawa, S., Kunstfeld, R., Brown, L. F., Detmar, M. (2004). VEGF-A promotes tissue repair-associated lymphatic vessel formation via VEGFR-2 and tandem  $\alpha$ 1 $\beta$ 1 integrins. *The FASEB journal*, 18(10), 1111-1113.
- Houghton, P.J., Hylands, P.J., Mensah, A.Y., Hensel, A., Deters, A.M., (2005). *In vitro* tests and ethnopharmacological investigations: wound healing as an example. *Journal of Ethnopharmacology* 100, 100– 107.
- Howell-Jones, R., Wilson, M., Hill, K., Howard, A., Price, P., Thomas, D. (2005). A review of the microbiology, antibiotic usage and resistance in chronic skin wounds. *Journal of Antimicrobial Chemotherapy*, 55(2), 143-149.
- Hsu, A, Mustoe TA. The Principles of Wound Healing (2010). In: Weinzweig J (ed). *Plastic Surgery Secrets*, Mosby Elsevier, China, 1-43.  
<http://www.inriodulce.com/images/moringa-oleifera.jpg>.
- Hu, F. B. (2011). Globalization of Diabetes The role of diet, lifestyle, and genes. *Diabetes care*, 34(6), 1249-1257.
- Hukkeri, V. I., Nagathan, C. V., Karadi, R. V., Patil, B. S. (2006). Antipyretic and wound healing activities of *Moringa oleifera* Lam. in rats. *Indian journal of pharmaceutical sciences*, 68(1), 124.
- Icks, A., Haastert, B., Trautner, C., Giani, G., Glaeske, G., Hoffmann, F. (2009). Incidence of lower-limb amputations in the diabetic compared to the non-diabetic population. Findings from nationwide insurance data, Germany, 2005-2007. *Experimental and Clinical Endocrinology and Diabetes*. 117(9):500-4.
- International Diabetes Federation (2009). *IDF Diabetes Atlas, 2<sup>nd</sup> edn, International Diabetes Federation*.

International Diabetes Federation (2011). *IDF Diabetes Atlas, 3rd edn, International Diabetes Federation.*

International Diabetes Federation (2013). *IDF Diabetes Atlas, 4th edn.* Brussels, Belgium: *International Diabetes Federation.*

International Diabetes Federation (2014). *IDF Diabetes Atlas, 6th edn.* Brussels, Belgium: *International Diabetes Federation.*

International Diabetes Federation Diabetes (2012). *IDF Diabetes Atlas, 5th ed* Brussels, Belgium, *International Diabetes Federation.*

Jain, P. G., Patil, S. D., Haswani, N. G., Girase, M. V., Surana, S. J. (2010). Hypolipidemic activity of *Moringa oleifera* Lam., Moringaceae, on high fat diet induced hyperlipidemia in albino rats. *Revista Brasileira de Farmacognosia*, 20(6), 969-973.

Jaiswal, D., Rai, P. K., Kumar, A., Mehta, S., Watal, G. (2009). Effect of *Moringa oleifera* Lam. leaves aqueous extract therapy on hyperglycemic rats. *Journal of ethnopharmacology*, 123(3), 392-396.

James, G. A., Swogger, E., Wolcott, R., Secor, P., Sestrich, J., Costerton, J. W., Stewart, P. S. (2008). Biofilms in chronic wounds. *Wound Repair and regeneration*, 16(1), 37-44.

Jorge, M. P., Madjarof, C., Ruiz, A. L. T. G., Fernandes, A. T., Rodrigues, R. A. F., de Oliveira Sousa, I. M., de Carvalho, J. E. (2008). Evaluation of wound healing properties of *Arrabidaea chica Verlot* extract. *Journal of Ethnopharmacology*, 118(3), 361-366.

Kansara, S. S., Singhal, M. (2013). Evaluation of antiulcer activity of *Moringa oleifera* seed extract. *J. Pharmaceut. Sci. Biosci. Res*, 3(1), 20-25.

Kansara, S.S., Singhal, M. (2013). Evaluation of antiulcer activity of *Moringa oleifera* seed extract. *J Pharm 286 Sci Biosci Res*. 3: 20-25.

Kant, V., Gopal, A., Kumar, D., Pathak, N. N., Ram, M., Jangir, B. L., Kumar, D. (2015). Curcumin-induced angiogenesis hastens wound healing in diabetic rats. *Journal of Surgical Research*, 193(2), 978-988.

Kapoor, M., Howard, R., Hall, I., Appleton, I. (2004). Effects of *epicatechin gallate* on wound healing and scar formation in a full thickness incisional wound healing model in rats. *The American journal of pathology*, 165(1), 299-307.

Kar, A., Choudhary, B. K., Bandyopadhyay, N. G. (2003). Comparative evaluation of hypoglycaemic activity of some Indian medicinal plants in alloxan diabetic rats. *Journal of ethnopharmacology*, 84(1), 105-108.

Karthivashan G, Tangestani Fard M, Arulselvan P, Abas F, Fakurazi S. (2013). Identification of bioactive candidate compounds responsible for oxidative

- challenge from hydro-ethanolic extract of *Moringa oleifera* leaves. *J Food Sci*, 78:C1368– C75.
- Karthivashan, G., Arulselvan, P., Tan, S. W., & Fakurazi, S. (2015). The molecular mechanism underlying the hepatoprotective potential of *Moringa oleifera* leaves extract against acetaminophen induced hepatotoxicity in mice. *Journal of Functional Foods*, 17, 115-126.
- Kawakami, Y., Hosokawa, T., Morinaka, T., Irino, S., Hirano, S., Kobayashi, H. Takahashi, Y. (2012). Antiatherogenic effect of guava leaf extracts inhibiting leucocyte-type 12-lipoxygenase activity. *Food Chemistry*, 131(4), 1069-1075.
- Kekuda, T.P. Mallikarjun, N. Swathi, D. Nayana, K. Aiyar M.B and Rohini, T ( 2 0 1 0 ) . “ Ant i b a c t e r i a l a n d aMoringa f u n g a l oleifera l a m , ” *Journal of Pharmaceutical Science R2s2A37*.
- Keong, L. C., Halim, A. S. (2009). *In vitro* models in Biocompatibility. Assessment for biomedical-grade chitosan derivatives in wound management. *International Journal of Molecular Sciences*, 10(3), 1300-1313.
- Kerr, M. (2012). Foot care for people with diabetes: the economic case for change. *NHS Diabetes and Kidney Care*.
- Khorshid, F., Ali, S. S., Alsofyani, T., Albar, H. A. (2010). *Plectranthus tenuiflorus* (Shara) promotes wound healing: *In vitro* and *in vivo* studies. *International Journal of Botany*, 6(2). 169-180.
- King, A. (2012). The use of animal models in diabetes research. *British Journal of Pharmacology*.
- Klasen HJ (2000b) A historical review of the use of silver in the treatment of burns. Renewed interest for silver. *Burns* 26: 1318.
- Konturek, C.T., Brzozowski, S.J., Konturek, S., Kwiecien, A., Dembinski, E.G., Hahn, P. (2001). Influence of bacterial lipopolysaccharide on healing of chronic experimental ulcer in rat. *Scandinavian journal of gastroenterology*, 36(12): 1239-1247.
- Kravitz, S. R., McGuire J., Shanahan S. D. (2003). Physical assessment of the diabetic foot. *Adv Skin Wound Care*. 16:68– 75.
- Krueger, R. J. (2007). Flavonoids. Chemistry, biochemistry and applications. *Economic Botany*, 61(1), 101-101.
- Kuchmerovska, T., Shymanskyy, I., Bondarenko, L., & Klimenko, A. (2008). Effects of nicotinamide supplementation on liver and serum. *Eur J Med Res*, 13, 275-280.
- Kuehn, B. M. (2007). Chronic wound care guidelines issued. *JAMA* 297:938– 939

- Kumar N., Gupta A. K. (2010). Wound-healing activity of *Onosma hispidum* (ratanjot) in normal and diabetic rats. *Journal of Herbs, Spices & Medicinal Plants*, 15(4), 342-351.
- Kura, A.U., Al Ali, S. H., Hussein, M. Z., Fakurazi, S and Arulselvan, P. (2013). Development of a controlled-release anti-parkinsonian nanodelivery system using levodopa as the active agent. *International Journal of Nanomedicine*, 8, 1103.
- Kwon, A; Qiu, Z., Hashimoto, M., Yamamoto, K., and Kimura, T. (2009). Effects of medicinal mushroom (*Sparassis crispa*) on wound healing in streptozotocin-induced diabetic rats. *The American Journal of Surgery*, 197(4), 503-509.
- Ladwig, G. P., Robson, M. C., Liu, R. A. N., Kuhn, M., Muir, D. F., Schultz, G. S. (2002). Ratios of activated matrix metalloproteinase-9 to tissue inhibitor of matrix metalloproteinase-1 in wound fluids are inversely correlated with healing of pressure ulcers. *Wound repair and regeneration*, 10(1), 26-37.
- Lau, T. W., Lam, F. F. Y., Lau, K. M., Chan, Y. W., Lee, K. M., Sahota, D. S., Lau, C. B. S. (2009). Pharmacological investigation on the wound healing effects of *Radix Rehmanniae* in an animal model of diabetic foot ulcer. *Journal of ethnopharmacology*, 123(1), 155-162.
- Lauer, G., Sollberg, S., Cole, M., Flamme, I., Stürzebecher, J., Mann, K., Eming, S. A. (2000). Expression and proteolysis of vascular endothelial growth factor is increased in chronic wounds. *Journal of investigative dermatology*, 115(1), 12-18.
- Lee, H. J., Jeong, Y. J., Lee, T. S., Park, Y. Y., Chae, W. G., Chung, I. K., Chang, Y. C. (2013). *Moringa* fruit inhibits LPS-induced NO/iNOS expression through suppressing the NF- $\kappa$ B activation. *The American journal of Chinese medicine*, 41(05), 1109-1123.
- Lee, S., Kim, Y. J., Kwon, S., Lee, Y., Choi, S. Y., Park, J. and Kwon, H. J. (2009) Inhibitory effects of flavonoids on TNF- $\alpha$ -induced IL-8 gene expression in HEK 293 cells. *BMB Rep.* 42, 265-270.
- Lee, Y. H., Chang, J. J., Chien, C. T., Yang, M. C., & Chien, H. F. (2012). Antioxidant sol-gel improves cutaneous wound healing in streptozotocin-induced diabetic rats. *Experimental diabetes research*, 2012. Article ID 504693, 11 pages.
- Leite Pereira, M., David de Oliveira, H., Tadeu Abreu de Oliveira, J., Menezes Gifoni, J., de Oliveira Rocha, R., de Oliveira Bezerra de Sousa, D., Maria Vasconcelos, I. (2011). Purification of a chitin-binding protein from *Moringa oleifera* seeds with potential to relieve pain and inflammation. *Protein and peptide letters*, 18(11), 1078-1085.
- Leite Pereira, M., David de Oliveira, H., Tadeu Abreu de Oliveira, J., Menezes Gifoni, J., de Oliveira Rocha, R., de Oliveira Bezerra de Sousa, D., Maria

- Vasconcelos, I. (2011). Purification of a chitin-binding protein from *Moringa oleifera* seeds with potential to relieve pain and inflammation. *Protein and peptide letters*, 18(11), 1078-1085.
- Lenzen, S. (2008). The mechanisms of alloxan- and streptozotocin-induced diabetes, *Diabetologia*, 51(2), 216– 226.
- Letchuman, G. R., Wan Nazaimoon, W. M., Wan Mohamad, W. B., Chandran, L. R., Tee, G. H., Jamaiyah, H., Ahmad Faudzi, Y. (2010). Prevalence of diabetes in the Malaysian national health morbidity survey III 2006. *Med J Malaysia*, 65(3), 180-6.
- Leung, D. W., Cachianes, G., Kuang, W., Goeddel, D. V., & Ferrara, N. (1989). Vascular endothelial growth factor is a secreted angiogenic mitogen. *Science*, 246(4935), 1306-1309.
- Li, J., Chen, J., Kirsner, R. (2007). Pathophysiology of acute wound healing. *Clinics in dermatology*, 25(1), 9-18.
- Li, S., Zhao, J., Liu, J., Xiang, F., Lu, D., Liu, B., Chen, B. (2011). Prospective randomized controlled study of a Chinese herbal medicine compound *Tangzu Yuyang* Ointment for chronic diabetic foot ulcers: a preliminary report. *Journal of ethnopharmacology*, 133(2), 543-550.
- Liang, C.C. Park, A.Y and Guan, J.L. (2007) *In vitro* scratch assay: A convenient and inexpensive method for analysis of cell migration *in vitro*, *Nature Protocols*, vol. 2, no. 2, pp. 329-333.
- Lipsky, B. A., Berendt, A. R., Cornia, P. B., Pile, J. C., Peters, E. J., Armstrong, D. G., Senneville, E. (2012). 2012 Infectious Diseases Society of America clinical practice guideline for the diagnosis and treatment of diabetic foot infections. *Clinical infectious diseases*, 54(12), 132-173.
- Lipsky, B. A., Hoey, C. (2009). Topical antimicrobial therapy for treating chronic wounds. *Clinical infectious diseases*, 49(10), 1541-1549.
- Liu, H., Lin, S., Xiao, D., Zheng, X., Gu, Y., Guo, S. (2013). Evaluation of wound healing potential of *resina draconis* (*Dracaena cochinchinensis*) in animal models. *Evidence-Based Complementary and Alternative Medicine*. 2013, 709865, 10 pages.
- Lobmann, R., Ambrosch, A., Schultz, G., Waldmann, K., Schiweck, S., Lehnert, H. (2002). Expression of matrix-metalloproteinases and inhibitors in wounds of diabetic and non-diabetic patients. *Diabetologia*, 45(7), 1011-1016.
- Lodhi, S. Pawaar, R. S. Jain A. P and Singh, Tephrosia purpurea (linn.) *Journal of Ethnopharmacology*, 108 (2): 204-210.

- Lodhi, S., Singhai, A. K. (2013). Wound healing effect of flavonoid rich fraction and luteolin isolated from *Martynia annua* linn. on streptozotocin induced diabetic rats. *Asian Pacific Journal of Tropical Medicine*, 6(4), 253-259.
- Lyon, K. C. (2008). The case for evidence in wound care: investigating advanced treatment modalities in healing chronic diabetic lower extremity wounds. *Journal of Wound Ostomy & Continence Nursing*, 35(6), 585-590.
- M. Sargent (Ed.), Guide to achieving reliable quantitative LC-MS measurements, RSC Analytical Methods Committee, 2013. ISBN 978-0-948926-27-3.
- Mafauzy, M. (2006) Diabetes mellitus in Malaysia. *Medical Journal of Malaysia*, 61(4), 397
- Mahajan, S. G., Mali, R. G., and Mehta, A. A. (2007) Protective effect of ethanolic extract of seeds of *Moringa oleifera* lam against inflammation associated with development of arthritis in rats. *Journal of Immunotoxicology*, 4(1), 39-47.
- Manaheji, H., Jafari, S., Zaringhalam, J., Rezazadeh, S., Taghizadfarid, R. (2011). Analgesic effects of methanolic extracts of the leaf or root of *Moringa oleifera* on complete Freund's adjuvant-induced arthritis in rats. *Zhong xi yi jie he xue bao= Journal of Chinese integrative medicine*, 9(2), 216-222.
- Mansbridge, J. N., Liu, K., Pinney, R. E., Patch, R., Ratcliffe, A., Naughton, G. K. (1999). Growth factors secreted by fibroblasts: Role in healing diabetic foot ulcers. *Diabetes, Obesity and Metabolism*, 1(5), 265-279.
- Margolis, D. J., Malay, D. S., Hoffstad, O. J., Leonard, C. E., MacCurdy, T., de Nava, K. L., Siegel, K. L. (2011). Incidence of diabetic foot ulcer and lower extremity amputation among Medicare beneficiaries. *Data Points Publication Series*. 2006 to 2008.
- Marrassini, C., Davicino, R., Acevedo, C., Anesini, C., Gorzalczany, S., Ferraro, G. (2011). Vicenin-2, a potential anti-inflammatory constituent of *Urtica circularis*. *Journal of natural products*, 74(6), 1503-1507.
- Martin, J. M., Zenilman, J. M., and Lazarus, G. S. (2009). Molecular microbiology: New dimensions for cutaneous biology and wound healing. *Journal of Investigative Dermatology*, 130(1), 38-48.
- Masiello, P., Broca, C., Gross, R., Roye, M., Manteghetti, M., Hillaire-Buys, D., Ribes, G. (1998). Experimental NIDDM: development of a new model in adult rats administered streptozotocin and nicotinamide. *Diabetes*, 47(2), 224-229.
- Matsumura, Y. (2007). A histopathological and immunohistochemical study of wound healing mechanism and biological effect of low-energy laser irradiation in type 2 diabetic model mice. *International Journal of Oral-Medical Sciences*, 6(1), 1-13.

- Mayne, S. T (2003). Antioxidant nutrients and chronic disease: use of biomarkers of exposure and oxidative stress status in epidemiologic research *Journal of Nutrition.*, 133, 3, 33S-940S.
- McMaster, S. K., Paul-Clark, M. J., Walters, M., Fleet, M., Anandarajah, J., Sriskandan, S., Mitchell, J. A. (2008). Cigarette smoke inhibits macrophage sensing of Gram-negative bacteria and lipopolysaccharide: relative roles of nicotine and oxidant stress. *British journal of pharmacology*, 153(3), 536-543.
- Mekkes J. R., Loots M. A. M., Van Der Wal A. C., Bos, J. D. (2003). Causes, investigation and treatment of leg ulceration. *Br J Dermatol.* 148: 388-401.
- Melo, S. S., Arantes, M. R., Meirelles, M. S., Jordao Jr, A. A., & Vannucchi, H. (2000). Lipid peroxidation in nicotinamide deficient and nicotinamide supplemented rats with Streptozotocin induced diabetes. *Acta diabetologica*, 37(1), 33-39.
- Mendez, M. V., Raffetto, J. D., Phillips, T., Menzoian, J. O., and Park, H. (1999). The proliferative capacity of neonatal skin fibroblasts is reduced after exposure to venous ulcer wound fluid: A potential mechanism for senescence in venous ulcers. *Journal of Vascular Surgery*, 30(4), 734-743.
- Menke, N. B., Ward, K. R., Witten, T. M., Bonchev, D.G., Diegelmann, R.F. (2007). Impaired wound healing. *Clin Dermatol* 25:19-25
- Mensah, A. Y., Sampson, J., Houghton, P. J., Hylands, P. J., Westbrook, J., Dunn, M., Cherry, G. W. (2001). Effects of *Buddleja globosa* leaf and its constituents relevant to wound healing. *Journal of Ethnopharmacology*, 77(2), 219-226.
- Ministry of Health (M.O.H) Malaysia (2011). The National Health and Morbidity Survey 2011 (NHMS 2011).
- Molan, P. C. (1999). The role of honey in the management of wounds. *Journal of Wound Care*, 8(8): 415-8.
- Mollering, R.C. (1995). Past, present and future of antimicrobial agents. *American Journal of Medicine*. 99, (Suppl 6A), 11S-18S.
- Momoh, M. A., Chime, S. A., Kenechukwu, F. C. (2013). Novel drug delivery system of plant extract for the management of diabetes: an antidiabetic study. *Journal of dietary supplements*, 10(3), 252-263.
- Moseley, R., Hilton, J. R., Waddington, R. J., Harding, K.G., Stephens, P., Thomas, D.W. (2004). Comparison of oxidative stress biomarker profiles between acute and chronic wound environments. *Wound Repair Regeneration*. 12(4): 419-29.
- Moura, L. I., Dias, A. M., Carvalho, E., de Sousa, H. C. (2013). Recent advances on the development of wound dressings for diabetic foot ulcer treatment—A

- review. *Acta biomaterialia*, 9(7), 7093-7114.
- Murthy, S., Gautam, M. K., Goel, S., Purohit, V., Sharma, H., and Goel, R. K. (2013). Evaluation of *in Vivo* Wound Healing Activity of *Bacopa monniera* on Different Wound Model in Rats. *BioMed research international*, 2013, 972028.
- Nagori, B. P. and Solanki, R. (2011). “Research Journal of Medicinal Plant, 5(4), 392-405.
- Nandave, M., Ojha, S. K., Joshi, S., Kumari, S., Arya, D. S. (2009). *Moringa oleifera* leaf extract prevents isoproterenol-induced myocardial damage in rats: Evidence for an antioxidant, antiperoxidative, and cardioprotective intervention. *Journal of Medicinal Food*, 12(1), 47-55.
- Nayak, B., Isitor, G., Maxwell, A., Bhogadi, V., Ramdath, D. (2007). Wound-healing activity of *morinda citrifolia* fruit juice on diabetes-induced rats. *Journal of Wound Care*, 16(2), 83-86.
- Nayak, S. (2006). Influence of ethanol extract of *vinca rosea* on wound healing in diabetic rats. *Online Journal of Biological Sciences*, 6(2), 51-55.
- Ndong, M., Uehara, M., Katsumata, S. I., Suzuki, K. (2007). Effects of oral administration of *Moringa oleifera* Lam on glucose tolerance in Goto-Kakizaki and Wistar rats. *Journal of clinical biochemistry and nutrition*, 40(3), 229.
- New, J. P., Hollis, S., Campbell, F., McDowell, D., Burns, E., Dornan, T. L., Young, R. J. (2000). Measuring clinical performance and outcomes from diabetes information systems: an observational study. *Diabetologia*, 43(7), 836-843.
- Nganlasom, J., Suttitum, T., Jirakulsomchok, D., Puapairoj, A. (2008). Effects of *Centella asiatica* linn. leaves and *Garcinia mangostana* linn. hull on the healing of dermal wounds in diabetic rats. *Srinagarind Medical Journal (SMJ)*, 23(4), 402-407.
- Nikkon, F., Saud, Z. A., Rahman, M. H., Haque, M. E. (2003). In vitro Antimicrobial Activity of the Compound Isolated from Chloroform Extract of *Moringa oleifera* Lam. *Pakistan journal of biological Sciences*, 6(22), 1888-1890.
- Nisar, M., Khan, S., Dar, A., Rehman, W., Khan, R., Jan, I. (2013). Antidepressant screening and flavonoids isolation from *Eremostachys laciniata* (L) Bunge. *African Journal of Biotechnology*, 10(9), 1696-1699.
- Nogami, M., Hoshi, T., Kinoshita, M., Arai, T., Takama, M., Takahashi, I. (2007). Vascular endothelial growth factor expression in rat skin incision wound. *Medical Molecular Morphology*, 40(2), 82-87.
- Ola, S. S., Catia, G., Marzia, I., Francesco, V. F., Afolabi, A. A., and Nadia, M. (2009). HPLC/DAD/MS characterisation and analysis of flavonoids and

- cynnamoil derivatives in four Nigerian green-leafy vegetables. *Food Chemistry*, 115(4): 1568-1574.
- O'Loughlin, A and O'Brien, T. (2011). *Topical stem and progenitor cell therapy for diabetic foot ulcers*. INTECH Open Access Publisher.
- O'Meara SM, Cullum NA, Majid M, Sheldon TA. (2000) Systematic review of antimicrobial agents used for chronic wounds. *British Journal of Surgery*, 88(1), 4-21.
- Panchatcharam, M., Miriyala, S., Gayathri, V. S., Suguna, L. (2006). Curcumin improves wound healing by modulating collagen and decreasing reactive oxygen species. *Molecular and Cellular Biochemistry*, 290(1-2), 87-96.
- Paocharoen, V. (2010). The efficacy and side effects of oral *Centella asiatica* extract for wound healing promotion in diabetic wound patients. *J Med Assoc Thai*, 93(Suppl 7), S166-S170.
- Papanas N, Papatheodorou K, Papazoglou D, Kotsiou S, Maltezos E (2010). Association between foot temperature and sudomotor dysfunction in type 2 diabetes. *J Diabetes Sci Technol*. 4:803– 7.
- Park, B. K., Lee, S. H., Seo, J. N., Rhee, J. W., Park, J. B., Kim, Y. S., Kwon, H. J. (2010). Protection of burn-induced skin injuries by the flavonoid kaempferol. *BMB reports*, 43(1), 46-51.
- Parvathy M., Umamaheshwari, A(2007). “ Cytotoxicity of *Moringa oleifera* leaf oil extracts on human mulberry Trepel in Medicolegal Research, 2 (1), 44-50.
- Pascoe, W. S., Storlien, L. H. (1990). Inducement by fat feeding of basal hyperglycemia in -cell function: model for study of normal etiology and pathogenesis of NIDDM. *Diabetes*, 39(2), 226-233.
- Pastar, I., Nusbaum, A. G., Gil, J., Patel, S. B., Chen, J., Valdes, J., Davis, S. C. (2013). Interactions of methicillin resistant *S aureus* USA300 and *Pseudomonas aeruginosa* in polymicrobial wound infection. *PLoS One*, 8(2), e56846.
- Pattanayak, S., Nayak, S.S., Dinda, S.C., Panda, D and Navale, K.P. (2011). Evaluation of Herbal Ointments Formulated with Methanolic Extract of *Cajanus scarabaeoides*. *Journal of Pharmacy and Allied Health Sciences*, 1, 49-57.
- Pauzi, N. A. S., Muhammad, A. A., Fakurazi, S., Arulselvan, P., Ahmad, Z. (2013). Preliminary Study of the Optimization of Protocol for Development of Type 2 Diabetic Model in Rats. *Indian Journal of Science and Technology*, 6(7), 4960-4965.

- Pauzi, N. S., Govindarajan K., Faridah A., Arulselvan, P., Fakurazi, S. (2015). Wound healing potential of *spirulina platensis* extracts on human dermal fibroblast cells. *Exp and Clin Sci Journal* (14), 385-393.
- Pendsey, S. P. (2010). Understanding diabetic foot. *International journal of diabetes in developing countries*, 30(2), 75.
- Perez, G., Vargas, S., Ortiz, H. (2005). Wound healing properties of *Hylocereus undatus* on diabetic rats. *Phytotherapy Research*, 19(8), 665-668.
- Perry N. B., Anderson, R.E., Brennan, N.J., Douglas, M.H., Heaney, A.J., McGimpsey, A., Smallfield, B.M., (1999). Essential oils from *Dalmatian sage (Salvia officinalis l.)*: variations among individuals, plant parts, seasons, and sites. *Journal of Agricultural Food Chemistry*. 47(5), 2048-54.
- Pessoa E.S., Melhado, R.M., Theodoro, L.H., Garcia, V.G (2004). A histologic assessment of the influence of low-intensity laser therapy on wound healing in steroid-treated animals. *Photomedicine and Laser Surger*. 22(3), 199-204.
- Pimple, B., Kadam, P., and Patil, M., (2012). Ulcer healing properties of different extracts of *Origanum majorana* in streptozotocin-nicotinamide induced diabetic rats. *Asian Pacific Journal of Tropical Disease*, 2(4), 312-318.
- Pirbalouti, A. G., Shahrzad, A., Abed, K., Hamed, B. (2010). Wound healing activity of *Malva sylvestris* and *Punica granatum* in alloxan-induced diabetic rats. *Acta Poloniae Pharmaceutica - Drug Research*, 67(5), 511-516.
- Ponrasu, T and Suguna, L. (2012). Efficacy of *Annona squamosa* on wound healing in streptozotocin-induced diabetic rats. *International wound Journal*, (6), 613-623.
- Posten, W. Wrone, D.A. Dover, J.S. Arndt, K.A. Silapunt S., and Alam, M. (2005). "Low-level laser therapy for wound *Dermatologic Surgery*, 31(3), 334– 340.
- Prasad, S. K., Kumar, R., Patel, D. K., Hemalatha, S. (2010). Wound healing activity of *Withania coagulans* in streptozotocin-induced diabetic rats. *Pharmaceutical Biology*, 48(12), 1397-1404.
- Prieto, P., Pineda, M., Aguilar, M. (1999). Spectrophotometric quantitation of antioxidant capacity through the formation of a phosphomolybdenum complex: specific application *Analytical Biochemistry*, 269, 337– 341.
- Punithavathi VR, Prince PSM (2009) Combined effects of quercetin and α-tocopherol on lipids and glycoprotein components in isoproterenol induced myocardial infarcted Wistar rats. *Chemico-Biological Interactions* 181: 322– 327

- Rahman, M. M., Sheikh, M. M. I., Sharmin, S. A., Islam, M. S., Rahman, M. A., Rahman, M. M., Alam, M. F. (2009). Antibacterial activity of leaf juice and extracts of *Moringa oleifera* Lam. against some human pathogenic bacteria. *CMU J Nat Sci*, 8(2), 219.
- Ramkumar, K. M., Manjula, C., Sankar, L., Suriyanarayanan, S., Rajaguru, P. (2009). Potential *in vitro* antioxidant and protective effects of *gymnema montanum* H. on alloxan-induced oxidative damage in cells, pancreatic cancer cells, and HIT-T15. *Food and Chemical Toxicology*, 47(9), 2246-2256.
- Rathi, B. S., Bodhankar, S. L., Baheti, A. M. (2006). Evaluation of aqueous leaves extract of *Moringa oleifera* Linn for wound healing in albino rats. *Indian journal of experimental biology*, 44(11), 898.
- Rebolledo, F. A., Soto, J. T., de la Peña, J. E. (2011). *The pathogenesis of the diabetic foot ulcer: prevention and management*. INTECH Open Access Publisher.
- Riedel, F., Philipp, K., Sadick, H., Goessler, U., Hörmann, K., Verse, T. (2005). Immunohistochemical analysis of radiation-induced non-healing dermal wounds of the head and neck. *In vivo*, 19(2), 343-350.
- Robson, M. C., Hill, D. P., Smith, P. D., Wang, X., Meyer-Siegler, K., Ko, F., Robson, L. E. (2000). Sequential cytokine therapy for pressure ulcers: clinical and mechanistic response. *Annals of surgery*, 231(4), 600.
- Rogers, L. C., Bevilacqua, N. J., Armstrong, D. G., Andros, G. (2010). Digital planimetry results in more accurate wound measurements: a comparison to standard ruler measurements. *Journal of diabetes science and technology*, 4(4), 799-802.
- Russell, A.D. (2002) Antibiotic and biocide resistance in bacteria: comments and conclusions. *Journal of Applied Microbiology*. 92, 171S-173S.
- Saadabi, A. M., Zaid, I. A. (2011). An *in vitro* antimicrobial activity of *Moringa oleifera* L. seed extracts against different groups of microorganisms. *Australian Journal of Basic and Applied Sciences*, 5(5), 129-134.
- Saaristo, A., Tammeela, T., Färkkilä, A., Herttuala, S., Alitalo, K. (2006). Vascular endothelial growth factor-C accelerates diabetic wound healing. *The American journal of pathology*, 169(3), 1080-1087.
- Sashidhara, K. V., Rosaiah, J. N., Tyagi, E., Shukla, R., Raghbir, R., Rajendran, S. M. (2009). Rare dipeptide and urea derivatives from roots of *Moringa oleifera* as potential anti-inflammatory and antinociceptive agents. *European Journal of Medicinal Chemistry*, 44(1), 432-436.

- Sasidharan, S., Nilawatyi, R., Xavier, R., Latha, L. Y., Amala, R. (2010). Wound healing potential of *Elaeis guineensis jacq* leaves in an infected albino rat model. *Molecules*, 15(5), 3186-3199.
- Sato, N., Kashima, K., Tanaka, Y., Shimizu, H., Mori, M. (1997). Effect of granulocyte-colony stimulating factor on generation of oxygen-derived free radicals and myeloperoxidase activity in neutrophils from poorly controlled NIDDM patients. *Diabetes*, 46(1), 133-137.
- Schäfer, M., Werner, S. (2008). Oxidative stress in normal and impaired wound repair. *Pharmacological Research*, 58(2), 165-171.
- Scherer, S. S., Pietramaggiori, G. Mathews, J. C., Chan, R., Fiorina, P., Orgrill, D. P. (2008). Wound healing kinetics of the genetically diabetic mouse. *Wounds*. 20:18– 28.
- Schierle, C.F., De la Garza, M., Mustoe, T. A., Galiano, R. D. (2009). Staphylococcal biofilms impair wound healing by delaying re-epithelialization in a murine cutaneous wound model. *Wound Repair Regeneration*. 17, 354– 359.
- Schultz, G. S., Sibbald, R. G., Falanga, V., Ayello, E. A., Dowsett, C., Harding, K., Vanscheidt, W. (2003). Wound bed preparation: a systematic approach to wound management. *Wound repair and regeneration*, 11(s1), S1-S28.
- Shah, D. M., Darling 3rd, R. C., Chang, B. B., Fitzgerald, K. M., Paty, P. S., Leather, R. P. (1995). Long-term results of in situ saphenous vein bypass. Analysis of 2058 cases. *Annals of surgery*, 222(4), 438.
- Sharifudin, S. A., Fakurazi, S., Hidayat, M. T., Hairuszah, I., Aris Mohd Moklas, M., Arulselvan, P. (2013). Therapeutic potential of *M oleifera* against acetaminophen-induced hepatotoxicity in rats. *Pharm Biology*, 51(3), 279-288.
- Sharma, Y., Jeyabalan, G., Singh, R. (2013). Potential wound healing agents from medicinal plants: a review. *Pharmacologia*, 4, 349.
- Shaw, T. J., Martin, P. (2009). Wound repair at a glance. *Journal of Cell Science*. 122: 18, 3209-3213.
- Sheehan P, Jones P, Caselli A, Giurini JM, Veves A (2003). Percent change in wound area of diabetic foot ulcers over a 4-week period is a robust predictor of complete healing in a 12-week prospective trial. *Diabetes Care*. 26(6):1879– 1882.
- Shoskes, D. A., Nickel, J. C. (2011). Quercetin for chronic prostatitis/chronic pelvic pain syndrome. *Urologic Clinics of North America*, 38: 279– 284
- Shukla, A., Rasik, A., Jain, G., Shankar, R., Kulshrestha, D., Dhawan, B. (1999). *In vitro and in vivo* wound healing activity of *Asiaticoside* isolated from *Centella asiatica*. *Journal of Ethnopharmacology*, 65(1), 1-11

- Sibbald, R.G., and Woo, K. Y. (2008). The biology of chronic foot ulcers in persons with diabetes. *Diabetes/metabolism Research and Reviews*, 24(S1), S25-S30.
- Siddhuraju, P., Becker, K. (2003). Antioxidant properties of various solvent extracts of total phenolic constituents from three different agro climatic origins of drumstick tree (*Moringa oleifera* Lam.) leaves. *Journal of agricultural and food chemistry*, 51(8), 2144-2155.
- Siddiqui, M. J. A., and Ismail, Z. (2011). Simultaneous analysis of bioactive markers from *Orthosiphon stamineus* benth leaves extracts by reverse phase high performance liquid chromatography. *Tropical Journal of Pharmaceutical Research*, 10(1), 97-103.
- Sidhu, G. S., Singh, A. K., Thaloor, D., Banaudha, K. K., Patnaik, G. K., Srimal, R. C., Maheshwari, R. K. (1998). Enhancement of wound healing by Curcumin in animals. *Wound Repair and Regeneration*, 6(2), 167-177.
- Simirgiotis, M. J. (2013). Antioxidant Capacity and HPLC-DAD-MS profiling of Chilean peumo (*Cryptocarya alba*) fruits and comparison with German Peumo (*Crataegus monogyna*) from Southern Chile. *Molecules*, 18(2), 2061-2080
- Singh, A., Bajpai, S., Singh, N., Kumar, V., Gour, J. K., Singh, P. K., Singh, R. K. (2014). Wound healing activity of standardized extract of *Curculigo orchoides* in streptozotocin- induced diabetic mice. *Asian Pacific Journal of Tropical Disease*, 4, S48-S53.
- Singh, D., Arya, P. V., Aggarwal, V. P., Gupta, R. S. (2014). Evaluation of Antioxidant and Hepatoprotective Activities of *Moringa oleifera* Lam. Leaves in Carbon Tetrachloride-Intoxicated Rats. *Antioxidants*, 3(3), 569-591.
- Singh, D., Singh, D., Choi, S. M., Zo, S. M., Painuli, R. M., Kwon, S. W., Han, S. S. (2014). Effect of extracts of *Terminalia chebula* on proliferation of keratinocytes and fibroblasts cells: An alternative approach for wound healing. *Evidence-based Complementary and Alternative Medicine*, 2014. 701656, 13 pages.
- Singh, N., Armstrong, D. G., Lipsky, B. A. (2005). Preventing foot ulcers in patients with diabetes. *Jama*, 293(2), 217-228.
- Singh, R. G., Negi, P. S., Radha, C. (2013). Phenolic composition, antioxidant and antimicrobial activities of free and bound phenolic extracts of *Moringa oleifera* seed flour. *Journal of Functional Foods*, 5(4), 1883-1891.
- Slavin, J. (1996). The role of cytokines in wound healing. *The Journal of Pathology*, 178(1), 5-10.
- Song, J. J., and Salcido, R. (2011). Use of honey in wound care: An update. *Advances in Skin and Wound Care*, 24(1), 40.

- Souza, E. L. D., Lima, E. D. O., Freire, K. R. D. L., Sousa, C. P. D. (2005). Inhibitory action of some essential oils and phytochemicals on the growth of various moulds isolated from foods. *Brazilian archives of Biology and Technology*, 48(2), 245-250.
- Sreejayan, N., Rao, M. N. A. (1997). Nitric oxide scavenging by Curcuminoids. *Journal of Pharmacy and Pharmacology*, 49(1), 105-107.
- Srinivas, R. B., KiranKumar, R. R., Naidu, V., Madhusudhana, K., Agwane, S.B., Ramakrishna, S., Parakash, B. D. (2008). Evaluation of antimicrobial, antioxidant and wound-healing potentials of *Holoptelea integrifolia*. *Journal of Ethnopharmacol.* 115, 249-56.
- Srinivasan, K., Ramarao, P. (2007). Animal model in type 2 diabetes research: An overview. *Indian Journal of Medical Research*, 125(3), 451.
- Sriyani, K. A., Wasalathanthri, S., Hettiarachchi, P., Prathapan, S. (2013). Predictors of Diabetic Foot and Leg Ulcers in a Developing Country with a Rapid Increase in the Prevalence of Diabetes Mellitus. *PloS one*, 8(11), e80856.
- Stalikas, C. D. (2007). Extraction, separation, and detection methods for phenolic acids and flavonoids. *Journal of separation science*, 30(18), 3268-3295.
- Steed, D. L. (1995). Clinical evaluation of recombinant human platelet- derived growth factor for the treatment of lower extremity diabetic ulcers. *Journal of Vascular Surgery*, 21(1), 71-81.
- Stern, H. S. (1989) Silver sulphadiazine and the healing of partial-thickness burns: a prospective clinical trial. *British Journal of Plastic Surgery* 42, 5815.
- Strodtbeck, F. (2001). Physiology of wound healing. *Newborn and infant nursing Reviews*. 1(1), 43-52.
- Sultana, B., Anwar, F., Ashraf M. (2009). Effect of extraction solvent/technique on the antioxidant activity of selected medicinal plant extracts. *Molecules*, 14(6), 2167-2180.
- Sweitzer, S.M., Fann, S. A., Borg, T. K., Baynes, J. W., Yost, M. J. (2006). What is the future of diabetic wound care? *The Diabetes Educator*, 32(2), 197-210.
- Syarina, P. N. A., Karthivashan, G., Abas, F., Arulselvan, P., Fakurazi, S (2015). Wound healing potential of *spirulina platensis* extracts on human dermal fibroblast cells. EXCLI Journal. 14:385-393
- Talreja, T. (2010). Screening of crude extract of flavonoids of *Moringa oleifera* against bacterial and fungal pathogen. *Journal of Phytology*. 2(11): 31-35.
- Tam, J. C. W., Lau, K. M., Liu, C. L., To, M. H., Kwok, H. F., Lai, K. K., San Lau, C. B. (2011). The *in vivo* and *in vitro* diabetic wound healing effects of a 2-herb formula and its mechanisms of action. *Journal of ethnopharmacology*, 134(3), 831-838.

- Tanenberg, R. J., & Donofrio, P. D. (2008). Neuropathic problems of the lower limbs in diabetic patients. *The Diabetic Foot, 7th ed.* Philadelphia: Mosby Elsevier.
- Taylor, P. J. (2005). Matrix effects: the Achilles heel of quantitative high-performance liquid chromatography– electrospray tandem mass spectrometry. *Clinical biochemistry*, 38(4), 328-334.
- Teoh, S. L., Latiff, A. A., and Das, S. (2009). The effect of topical extract of *Momordica charantia* (bitter gourd) on wound healing in non-diabetic rats and in rats with diabetes induced by streptozotocin. *Clinical and Experimental Dermatology*, 34(7), 815-822.
- Thakur, R., Jain, N., Pathak, R., and Sandhu, S. S. (2011). Practices in wound healing studies of plants. *Evidence-Based Complementary and Alternative Medicine*, 438056, 17 pages.
- Tiloke, C., Phulukdaree, A., Chuturgoon, A. A. (2013). The antiproliferative effect of *Moringa oleifera* crude aqueous leaf extract on cancerous human alveolar epithelial cells. *BMC complementary and alternative medicine*, 13(1), 226.
- Tiwari, V., Tiwari A., Madhavan, V. (2010). Preliminary phytochemical analysis, HPTLC studies and antipyretic activity of alcohol and aqueous extract of *Helicteres isora* L. Root. *International Journal of Pharmacy and Pharmaceutical Science*; 2(2): 74-79.
- Tonks A, Cooper R. A., Price A. J., Molan P. C, Jones K. P. (2001). Stimulation of TNF-alpha release in monocytes by honey. *Cytokine*; 14(4): 240-2.26.
- Tonks, A. J., Cooper, R.A., Jones, K. P., Blair, S., Parton, J., Tonks, A (2003). Honey stimulates inflammatory cytokine production from monocytes. *Cytokines*. 21(5), 242-7.
- Tonnesen, M. G., Feng, X., Clark, R. A., (2000, December). Angiogenesis in wound healing. In *Journal of Investigative Dermatology Symposium Proceedings* (Vol. 5, No. 1, pp. 40-46). Nature Publishing Group.
- Tran, N. Q., Joung, Y. K., Lih, E., Park, K. (2011). In situ forming and rutin-releasing chitosan hydrogels as injectable dressings for dermal wound healing. *Biomacromolecules* 12(8), 2872– 2880.
- Trengove, N. J., Bielefeldt-Ohmann, H., Stacey, M. C. (2000). Mitogenic activity and cytokine levels in non-healing and healing chronic leg ulcers. *Wound Repair and Regeneration*, 8(1), 13-25.
- Tribolo, S., Lodi, F., Connor, C., Suri, S., Wilson, V. G., Taylor, M. A., Hughes, D. A. (2008). Comparative effects of quercetin and its predominant human metabolites on adhesion molecule expression in activated human vascular endothelial cells. *Atherosclerosis*, 197(1), 50-56.
- Tsala, D. E., Amadou, D., Habtemariam, S. (2013). Natural wound healing and

- bioactive natural products. *Phytopharmacology*, 4(3), 532-60.
- Tsourdi, E., Barthel, A., Rietzsch, H., Reichel, A., Bornstein, S. R. (2013). Current aspects in the pathophysiology and treatment of chronic wounds in diabetes mellitus. *BioMed research international*, 2013.
- Udupa, S.L; Shaila, HP; Udupa, AL; Ramesh, K.V., Kulkarni, D.R., (1991). *Biochemistry Archives*, 7, 207-212.
- Uma, N., Fakurazi, S., Hairuszah, I. (2010). *Moringa oleifera* enhances liver antioxidant status via elevation of antioxidant enzymes activity and counteracts paracetamol-induced hepatotoxicity. *Malaysian Journal of Nutrition*, 16(2), 293-307.
- Urquiaga, I., leighton, F. (2000). Plant polyphenol antioxidants and oxidative stress. *Biological Research*, 33(2), 55-64.
- Viswanathan, V., Kesavan, R., Kavitha, K., Kumpatla, S. (2011). A pilot study on the effects of a Polyherbal formulation cream on diabetic foot ulcers. *The Indian Journal of Medical Research*, 134(2), 168-173.
- Werner, S., Grose, R. (2003). Regulation of wound healing by growth factors and cytokines. *Physiological reviews*, 83(3), 835-870.
- White, R., Cooper, R. (2005). Silver sulphadiazine: A review of the evidence. *Wounds Uk*, 1(2), 51.
- Whiting, D. R., Guariguata, L., Weil, C., and Shaw, J. (2011). IDF diabetes atlas: Global estimates of the prevalence of diabetes for 2011 and 2030. *Diabetes Research and Clinical Practice*, 94(3), 311-321.
- Wild, S., Roglic, G., Green, A., Sicree, R., King, H. (2004). Global prevalence of diabetes estimates for the year 2000 and projections for 2030. *Diabetes care*, 27(5), 1047-1053.
- Williams, D. T., Harding, K. G., Price, P. ( 2 0 0 5 ) . A n e v a l u a t i o n methods used in screening for lower limb arterial disease in diabetes. *Diabetes Care*. 28:2206– 10.
- Wolfender, J. L., Maillard, M., Hostettmann, K. (1994). Thermospray liquid chromatography-mass spectrometry in phytochemical analysis. *Phytochemical Analysis*, 5(4), 153-182.
- Wong, V. W., Sorkin M, Glotzbach JP, Longaker MT, Gurtner GC. 2011. Surgical approaches to create murine models of human wound healing. *J Biomed Biotechnol* 96:9618.
- Woo, K., Ayello, E. A., and Sibbald, R. G. (2007). The edge effect: Current therapeutic options to advance the wound edge. *Advances in Skin & Wound Care*, 20(2), 99-117.

- Wu, S., Armstrong, D. (2005): Risk assessment of the diabetic foot and wound. *International Wound Journal*, 2:17– 24.
- Yang, J., Klaidman, L. K., Nalbandian, A., Oliver, J., Chang, M. L., Chan, P. H., & Adams, J. D. (2002). The effects of nicotinamide on energy metabolism following transient focal cerebral ischemia in Wistar rats. *Neuroscience letters*, 333(2), 91-94.
- Yusof, M. I., Sulaiman, A. R., & Muslim, D. A. (2007). Diabetic foot complications: a two-year review of limb amputation in a Kelantanese population. *Singapore medical journal*, 48(8), 729-732.
- Zaine, N. H., Burns, J., Vicaretti, M., Fletcher, J. P., Begg, L., & Hitos, K. (2014). Characteristics of diabetic foot ulcers in Western Sydney, Australia. *Journal of foot and ankle research*, 7(1), 39.