



UNIVERSITI PUTRA MALAYSIA

***COMPARATIVE STUDY OF THE BIOLOGICAL PROPERTIES OF FOETAL
AND PLACENTAL DISTAL ENDS OF HUMAN UMBILICAL CORD
WHARTON'S JELLY MESENCHYMAL STEM CELLS***

ZURAIDAH YUSOFF

FPSK(m) 2015 55



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AND PLACENTAL DISTAL ENDS OF HUMAN UMBILICAL CORD
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By
ZURAIDAH YUSOFF

Thesis Submitted To the School of Graduates Studies, Universiti Putra Malaysia,
in Fullfilment of the Requirements for the Degree of Master of Science

August 2015

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of
the requirement for the degree of Master of Science

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Chairman : Associate Professor Rajesh Ramasamy, PhD
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Mesenchymal stem cells (MSC) are defined as a non-haematopoietic multipotent stem cells with the ability to self-renew; capable of differentiating into various mature cells and exert potent immunomodulatory activities on immune cells. These unique characteristics and properties of MSC had enabled them to be recognised as an attractive tool in stem cell based immunotherapy, gene therapy and regenerative medicine especially for fatal or incurable diseases. To date, many studies have demonstrated that MSC can be generated from various organs and tissues, such as birth-associated tissues including placenta, amnion and umbilical cord. Umbilical cord derived MSC (UC-MSC) has long been considered as an alternative to MSC that derived from bone marrow (BM-MSC) as they are regarded as medical waste with little ethical concern for research and easily culture expanded. The human umbilical cord (UC) comprises of various compartments such as two arteries, one vein, amnion, and Wharton's jelly (WJ). Cells isolated from these compartments were found to be plastic adherent and able to differentiate into many cell types such as osteoblasts, adipocytes, chondrocytes, hepatocytes, neural and cardiac cells. Since MSC are migrating from foetal liver to the bone marrow at last trimester of pregnancy, thus the chances of MSC to be deposited at various compartments of umbilical cord will principally different too. Thus, the present study is aimed to determine a better anatomical part of umbilical cord which potentially yield a greater number and quality of MSC. In this present study, two distinguished anatomical parts of human umbilical cord Wharton's jelly were consumed to generate MSC. These are foetal distal end (FE) that connected to the newborn baby and the placental distal end (PE) which is adjacent to the placenta. Mesenchymal stem cells were generated from the explants method where Wharton's jelly of umbilical cord were minced up to paste-like state and cultured in the same condition. The generated WJ-MSC were characterised using a standard panel of surface markers and their ability to differentiate into osteoblasts and adipocytes followed by quantification of PCR gene expression. The immunomodulatory effect of MSC that generated from both ends were tested with mitogen activated T cells along with Jurkat cells through tritiated thymidine assay, cell cycle assay and apoptosis assay. The

possibility of generating WJ-MSC from both distal ends of umbilical cord was about 37-50%. The adherent cells generated from FE and PE distal ends have exhibited similar morphological characteristics, presented as spindle-shaped fibroblast-like cells. As for immunophenotyping, comparable data were observed where MSC from both ends are positive for common MSC markers (CD29, CD73, CD90, CD106 and MHC-class I), negative expression for haematopoietic and immune markers (CD14, CD34, CD45, CD80, CD86 and MHC-class II). However, colony-forming unit assay of FE-WJMSC showed more number of colonies and bigger in size as compared to the PE-WJMSC. Moreover, FE-WJMSC showed rapid growth kinetics and shorter doubling time with an average of 34 hr. The predominant measurements in osteogenic and adipogenic induction assays via deposition of calcium and accumulation of lipid granules respectively demonstrated both MSC possessed an equivalent differentiation potency. Real time qPCR analysis on adipocytes and osteoblast differentiated from both ends showed up-regulation and down-regulation of specific set of genes, thus further confirmed the lineage specific differentiations. In term of immunomodulatory activity, MSC generated from both ends were able to suppress the proliferation of activated T cells in dose dependant manner and preserved cell viability simultaneously. Moreover, detention of T cells in G₀/G₁ phase of the cell cycle and arresting cells from entering into S phase were evident when FE-WJMSC were co-cultured with T cells. Furthermore, co-culture of FE-WJMSC with Jurkat cells has significantly increased the number of viable cells and rescued the cells from undergoing apoptosis. The current study displays the efficiency of generating MSC from different anatomical parts of human umbilical cord where FE-WJMSC can be regarded as a better source of MSC as compared to PE as they showed superior proliferation capacity, greater anti-proliferative effect on immune cells and cell preservation ability from apoptosis. The higher expansion potential and enhanced functionality of FE-WJMSC at *in vitro* promise a quick and uninterrupted supply of MSC to encounter the clinical demand and research purposes.

Abstrak tesis yang dikemukakan kepada Senate Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Sarjana Sains

**KAJIAN PERBANDINGAN TERHADAP CIRI-CIRI BIOLOGI SEL
MESENKIMA YANG DIPEROLEHI DARIPADA HUJUNG JANIN DAN
HUJUNG PLASENTA DARIPADA WHARTON'S JELLY TALI PUSAT
MANUSIA**

Oleh

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Pengerusi : Profesor Madya Rajesh Ramasamy, PhD
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Sel induk mesenkima (MSC) didefinisikan sebagai sel bukan hematopoisis (*non-haematopoietic*) multipotensi yang berkebolehan untuk membezakan dirinya menjadi pelbagai jenis sel matang yang lain dan menunjukkan kegiatan immunomodulatori ke atas pelbagai sel imun. Ciri-ciri yang unik ini telah menjadikan MSC diiktiraf sebagai agen yang menarik untuk rawatan berdasar kanser dalam bidang imunoterapi, gen terapi dan regenerasi terutama sekali untuk penyakit yang tidak boleh dirawat atau yang boleh menyebabkan kematian. Sehingga kini, banyak kajian telah menunjukkan bahawa MSC boleh dijana daripada pelbagai jenis organ dan tisu termasuklah tisu yang berkaitan dengan kelahiran seperti uri, amnion dan tali pusat. Sel induk mesenkima yang dijana daripada tali pusat (UC-MSC) sudah sekian lama dikenali sebagai alternatif kepada MSC yang dijana daripada sum-sum tulang (BM-MSC) kerana ianya dianggap sebagai sisa klinikal yang bebas dari etika perubatan dan senang diperkembangkan dalam kultur. Tali pusat manusia terdiri daripada beberapa bahagian iaitu dua arteri, satu vena, amnion dan *Wharton's jelly* (WJ). Sel yang diasingkan dari berlainan jenis bahagian tali pusat ini didapati boleh melekat pada permukaan plastik dan mampu membezakan diri menjadi sel lain seperti tulang, lemak, rawan, hati, neural dan jantung. Oleh kerana MSC semasa kehamilan bergerak dari hati janin ke sum-sum tulang pada *trimester* terakhir, ini bermakna secara prinsipnya, kebarangkalian untuk MSC terserap ke dalam pelbagai bahagian tali pusat juga berbeza. Oleh itu, kajian ini bertujuan untuk menentukan keberkesanan bahagian tali pusat yang berlainan anatomi untuk menjana MSC yang baik dari segi kuantiti dan kualiti. *Wharton's jelly* dari dua hujung tali pusat yang berbeza telah digunakan dalam kajian ini iaitu hujung janin (FE), bahagian yang bersambung dengan janin dan hujung plasenta (PE) iaitu bahagian yang bersambung kepada uri. Penjanaan MSC secara *explants* telah digunakan dimana WJ dipotong menjadi cebisan tisu yang kecil dan dikultur. Selepas penjanaan dan perkembangan WJ-MSC, ianya dicirikan mengikut *immunophenotyping* dan keupayaan pembezaan kepada osteosit dan adiposit, diikuti dengan kuantifikasi ekspresi gen

melalui analisa PCR. Kesan imunomodulatori MSC yang telah dijana dari kedua-dua hujung tali pusat diuji dengan sel T yang diaktifkan dengan mitogen melalui ujian proliferasi, kitaran sel dan apoptosis. Kebarangkalian untuk menjana WJ-MSC dari kedua-dua hujung tali pusat adalah sebanyak 37-50%. Penjanaan sel melekat dari FE dan PE menunjukkan morfologi fibroblas berbentuk gelondong yang sama cirinya. Bagi ujian penanda permukaan, kedua-dua hujung menunjukkan ekspresi positif untuk penanda yang biasa untuk MSC (CD29, CD73, CD90, CD106 and MHC-class I) ekspresi negatif untuk haematopoisis (CD14, CD34, CD45, CD80, CD86 and MHC-class II). Walaubagaimanapun, ujian *colony-forming unit* FE-WJMSC menunjukkan jumlah koloni yang lebih banyak dan saiz yang lebih besar dari PE. Tambahan lagi, FE menunjukkan kinetik pertumbuhan sel dan penggandaan populasi lebih pantas iaitu purata 34 jam. Induksi osteogenesis dan adipogenesis pula menunjukkan pengumpulan mineral kalsium dan granul lipid dimana kedua-duanya mempunyai potensi pembezaan yang sama. Analisa qPCR ke atas adiposit dan osteosit yang berjaya dibezakan dari kedua-dua hujung menunjukkan peningkatan dan penurunan regulasi bagi gen spesifik. Berhubung dengan aktiviti immunomodulatori, MSC dari kedua-dua hujung boleh merencat proliferasi sel T, berdasarkan nisbah MSC. Tambahan lagi, kehadiran FE-WJMSC dalam kultur sel T telah memerangkap sel-sel imun ini pada fasa G₀/G₁ pada kitaran sel dan menahan sel dari memasuki fasa S. Penemuan ini telah menunjukkan penjanaan MSC dari tisu yang sama tetapi bahagian anatomi yang berlainan mempunyai kecekapan yang berbeza dimana FE-WJMSC boleh dianggap sebagai sumber MSC yang lebih baik berbanding PE kerana telah menunjukkan kapasiti pertumbuhan lebih baik, kesan anti-pertumbuhan terhadap sel-sel imun yang lebih berkesan dan lebih cekap untuk memelihara sel dari apoptosis. Potensi perkembangan yang lebih tinggi dan peningkatan fungsi yang dipamerkan oleh FE-WJMSC secara *in vitro* boleh menjanjikan pembekalan MSC yang lebih pantas dan tanpa ganguan bagi memenuhi permintaan untuk kegunaan klinikal mahupun kajian.

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I certify that a Thesis Examination Committee has met on (07 August 2015) to conduct the final examination of Zuraidah Bt. Yusoff on her thesis entitled "Comparative Study of the Biological Properties of Foetal and Placental Distal Ends of Human Umbilical Cord Wharton's Jelly Mesenchymal Stem Cells" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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LIST OF ABBREVIATIONS

| | |
|--------------------|---|
| µl | Micro litre |
| ³ H-TdR | Tritiated thymidine |
| ABB | Annexin binding buffer |
| ALP | Alkaline phosphatase |
| AML | Acute myeloid leukaemia |
| APC | Antigen presenting cells |
| APC | Allophycoerythrin |
| B7-1 | CD80 |
| B7-2 | CD86 |
| bFGF | Basic fibroblast growth factor |
| BM | Bone marrow |
| BMP-2 | Bone morphogenetic protein-2 |
| bp | Base pair |
| BSP | Bone sialoprotein |
| c/EBP | CCAAT-enhancer-binding protein |
| CD | Cluster of differentiation |
| cDNA | Complementary deoxyribonucleic acid |
| CII | Type II collagen |
| CDKI | Cyclin-dependant kinase inhibitor |
| CFU | Colony forming unit |
| cm ² | Centimeter square |
| CO ₂ | Carbon dioxide |
| Col | Collagen type II |
| colla-1 | Collagen type 1 alpha-1 |
| cpm | Count per minute |
| CSCs | Cancer stem cells |
| CTL | Cytotoxic T lymphocytes |
| DC | Dendritic cells |
| DD | Death domain |
| DN | Double negative |
| DISC | Death inducing signaling complex |
| DMEM | Dulbecco's Modified Eagle's Medium |
| DMEMF12 | Dulbecco's Modified Eagle's Medium Ham's F-12 |
| DMSO | Dymethyl sulfoxide |
| DNA | Deoxyribonucleic acid |
| DNase | Deoxyribonuclease |
| DP | Double positive |
| E2F | Transcription factor E2F |
| EAA | Essential amino acid |
| ECM | Extra cellular matrix |
| EDTA | Ethylenediaminetetraacetic acid |
| EGF | Epidermal growth factor |
| ESC | Embryonic stem cells |
| FACS | Fluorescence-activated cell sorting |
| FADD | Fas-Associated protein with Death Domain |
| FBS | Fetal bovine serum |
| FITC | Fluorescein isothiocyanate |

| | |
|------------------|--|
| Flt-3 | Fms-like tyrosine kinase 3 |
| FoxP3 | Forkhead box P3 |
| G ₀ | Quiescence phase |
| G ₁ | Gap phase |
| GAPDH | Glyceraldehyde 3-phosphate dehydrogenase |
| GvHD | Graft versus host disease |
| HDMEM | High glucose Dulbecco's Modified Eagle's Medium |
| HLA-G | Human leucocyte antigen-G |
| HSC | Haematopoietic stem cells |
| ICAM-1 | Intercelllular adhesion molecule-1 |
| IDO | Indoleamine-2,3-dioxygenase |
| IGF | Insulin growth factor |
| IMDM | Iscove'c modified Dulbecco's medium |
| IFN- γ | Interferon gamma |
| IL | Interleukin |
| iPSC | Induced pluripotent stem cells |
| ISCT | International Society for Cellular Therapy |
| LIF | Leukaemia Inhibitory factor |
| LPL | Lipoprotein lipase |
| M | Mitotic phase |
| MAb | Monoclonal antibody |
| MHC | Major histocompatibility complex |
| MI | Myocardial infarction |
| ml | Mililiter |
| MLR | Mixed leukocyte reaction |
| mRNA | Messenger ribonucleic acid |
| MSC | Mesenchymal stem cells |
| ng | Nanogram |
| NK | Natural killer |
| NO | Nitric oxide |
| PBS | Phosphate buffer saline |
| PBMC | Peripheral blood mononuclear cell |
| PCR | Polymerase chain reaction |
| PD | Programmed death |
| PDT | Population doubling time |
| PDGF | Platelet-derived growth factor |
| PE | Phycoerythrin |
| PGE ₂ | Prostaglandin E ₂ |
| PHA-L | Phytohaemagglutinin-Leukocytes |
| PI | Propidium iodide |
| PPAR γ | Peroxisome proliferator-activated receptor gamma |
| PS | phosphatidylserine |
| rBM-MSC | Rat bone marrow mesenchymal stem cells |
| RNA | Ribonucleic acid |
| ROS | Reactive oxygen species |
| RPMI | Roswell park memorial institute |
| scf | Stem cell factor |
| SCID | Severe combined immunodeficient |
| SD | Standard deviation |
| SDF-1 | Stromal cell derived factor 1 |
| SLE | Systemic lupus erythematosus |

| | |
|---------------|------------------------------------|
| TCR | T cell receptor |
| TGF- β | Transforming growth factor-beta |
| Th 1 | T helper 1 |
| Th 2 | T helper 2 |
| TNF- α | Tumor necrosis factor α |
| TNFR | Tumor necrosis factor receptor |
| Tregs | T regulatory cells |
| VCAM-1 | Vascular cell adhesion molecule-1 |
| VEGF | Vascular endothelial growth factor |
| α | Alpha |
| β | Beta |
| γ | Gamma |
| δ | Delta |

CHAPTER 1

INTRODUCTION

A substantial amount of research over the past two decades has resulted in greater understanding of the human adult stem cell biology not only in the basic sciences but also their implication in therapeutic usage. Following the initial enthusiasm, it is now becoming clear that the use of stem cell transplantation is an important tool in the treatment of various malignancies, tissue repair and tissue regeneration. Although embryonic stem cell and inducible pluripotent stem cells (iPS) hold great potentials toward an unlimited differentiation, yet their ethical cues and high risk of cancer formation hindered their further application in clinical settings. This has allowed further exploration for a safe adult stem cells to be used in regenerative medicine. Adult stem cells are multipotent stem cells, which can be isolated from postnatal tissues, most commonly from bone marrow. Examples of adult stem cells are haematopoietic stem cells (HSC), mesenchymal stem cells (MSC) and other tissue resident stem cell types such as neural, epithelial, oval and satellite cells. Among them, mesenchymal stem cells appear to be an outstanding candidate of adult stem cells that possess multipotential differentiation ability and immuno-modulative activity.

Mesenchymal stem cell were first identified by a Russian scientist Friedenstein in 1970 as adherent cells that form fibroblasts-like colonies (Friedenstein *et al.* 1970). Since MSC were first discovered in bone marrow and co-exist with resident haematopoietic stem cells (HSC), it often address as non-haematopoietic stem cells. Besides that, MSC also known as well as mesenchymal stromal cells due to its contribution for stromal compartment to facilitate haematopoiesis. As per classical definition of adult stem cells, MSC comprise of a very rare population of stem cells that capable of self-renewal and lineage specific differentiation potential. It has been shown that MSC give rise to cells of mesodermal lineages such as bone, adipose, cartilage, tendon and skeletal muscle (Reyes *et al.* 2001; Pittenger *et al.* 1999; Fridenstein *et al.* 1974a). As a matter of fact, several reports have been shown that MSC also potentially differentiate into various non-mesodermal lineage tissues including pancreatic islet cell (Dang *et al.* 2015), cardiac muscle (Potdar and Prasannan 2013), hepatocyte (Kazemnejad *et al.* 2009) and neural cell (Kopen *et al.* 1999).

Various *in vitro* and preclinical studies based on normal and disease models had conferred the significant biological functions of MSC. Unlike other tissue specific adult stem cells, MSC hold a great potential to be considered as therapeutic agent due to its targeted homing capacity to the site of injuries and ability to differentiate into many different mesenchymal and parenchymal cell types thus, regenerate the damaged tissues or organs (Nagamura and He 2014; Kim *et al.* 2013). Mesenchymal stem cells considered as a ‘magic healing cells’ that secrete large amounts of pro-angiogenic, anti-inflammatory and anti-apoptotic cytokines/factors that exerts remarkable regenerative, reparative and immunosuppressive properties, which may play a role in the induction of transplantation tolerance and control of autoimmunity.

Historically, MSC derived from the bone marrow (BM) is considered as the most common source of MSC and designated as a gold standard as they have been used in most of the research and development for clinical therapies. However, the use of BM-MSC is not always acceptable or feasible due to many reasons such as high degree of viral exposures during BM aspiration, low yield of cell numbers and a significant reduction of proliferative and differentiation capacity with increasing age of the donor. Moreover, BM aspiration involves an invasive procedure and it may cause pain and discomfort to donors which may lead to infection, excessive bleeding and other complications. Undoubtedly, stem cell derived from adult bone marrow has very high successful rate for expansion, nonetheless many conditions may limit the accessibility. For instance, in some clinical cases such as bone marrow failure, aplastic anaemia, leukaemias and post-myeloblastic irradiation or chemotherapy, patients often encounter complications of inadequate cellular fractions in their bone marrow aspiration. For these patients, the only option for alternative sources of MSC will be a third party donor and most of the time it is difficult to find the right donor especially if they are volunteers. Furthermore, the number of cells required for clinical scale is large, which requires longer time period for *in vitro* culture expansion to obtain the adequate number of cells. These technical hitches can delay the treatment process and further affect the outcome of the treatment. This predicament has initiated an urge for finding the alternative source of MSC and umbilical cord MSC certainly could serve as an ideal candidate.

The human umbilical cord (UC) comprises of two arteries, one vein, cord lining and Wharton's jelly (WJ). Previous reports have described that each of these compartments can give rise to MSC (Conconi *et al.* 2011). Cells isolated from these compartments were found to be plastic adherent and able to differentiate into many cell types such as osteoblasts, adipocytes, chondrocytes, hepatocytes, neural and cardiac cells (Prasanna and Jahnava 2011). In fact, UC can be considered as a best alternative to BM as they are regarded as medical waste with little ethical concern for research (Hass *et al.* 2011). Moreover, a comparison study done on MSC derived from human umbilical cord revealed that they were better with respect to their proliferation capacity, rapid osteogenic differentiation and superior in transfection capacities when compared to the "gold standard" bone marrow mesenchymal stem cells (Baksh *et.al* 2007). However, several reports have indicated that MSC generated from various compartments of UC vary in term of initial stem cell composition and frequency; the ability to form colony forming units (CFU) (Majore *et al.* 2010; Karahuseyinoglu *et al.* 2007; Sarugaser *et al.* 2005;).

In this present study, two distinguished anatomical parts of human umbilical cord Wharton's jelly were consumed to generate MSC. These two anatomical parts are foetal distal end (FE) of UC that connected to the new-born baby and the placental distal end (PE) of UC which is connected to the placenta. The determination of an appropriate distal end of umbilical cord to generate MSC which potentially yield greater quantity and better quality of MSC will validate a fundamental character of WJ-MSC from different anatomical parts of human umbilical cord. Moreover, the findings from this study will be potentially useful for cryo-banking and also economical in term of space and cost. Hence, the early phase of this study is focused on generation of MSC derived from FE(FE-WJMSC) and PE(PE-WJMSC), followed by a standard characterisation using panel of surface markers and the ability to differentiate into osteocytes and adipocytes. The immunosuppressive activity of MSC also was investigated. Finally, the generated

WJ-MSC from two distal ends were co-culture with T cells and Jurkat cells to further explore the immunomodulatory effect and rescuing ability of WJ-MSC from serum-deprived cell death.

General objective

To generate, characterise and compare the mesenchymal stem cells that derived from foetal and placental distal ends of human umbilical cord Wharton's jelly.

Specific objectives:

1. To generate MSC from foetal distal end (FE) and placental distal end (PE) of human umbilical cord Wharton's jelly.
2. To compare the growth kinetics of MSC that derived from foetal and placental distal ends of human umbilical cord Wharton's jelly.
3. To characterise MSC that generated from both FE and PE by a standard immunophenotyping and differentiation potential.
4. To compare the immunosuppressive activity of MSC derived from FE and PE of human umbilical cord Wharton's jelly.
5. To evaluate the protective ability of FE-WJMSC and PE-WJMSC on serum-deprived Jurkat cells.

The hypotheses of this research are:

1. Colony forming adherent cells can be generated from both distal ends of human umbilical cord Wharton's jelly.
2. Adherent cells generated from both distal ends are able to express MSC specific surface makers and differentiate into mesodermal lineage cells.
3. Mesenchymal stem cell derived from FE of human umbilical cord Wharton's jelly will serve as a better source for mesenchymal stem cell with respect to their proliferation capability, differentiation potential and immunosuppressive activity as compared to PE.

REFERENCES

- Aggarwal S. and Pittenger M.F. (2005) Human mesenchymal stem cells modulate allogeneic immune cell responses. *Journal of The American Society of Hematology* 105:1815-1822.
- Aguilar S., Scotton C.J., McNulty K., Nye E., Stamp G., Laurent G., Bonnet D., Janes.M. (2009). Bone Marrow Stem Cells Expressing Keratinocyte Growth Factor via an Inducible Lentivirus Protects against Bleomycin-Induced Pulmonary Fibrosis. *Journal of Cell Therapy for Lung Fibrosis* 4:11-13.
- Almeida-P.G., Porada C.D., Tran N. and Zanjani E.D. (2000) Cotransplantation of Human stromal cell progenitors into preimmune fetal sheep results in early appearance of human donor cells in circulation and boosts cell levels in bone marrow at later time points after transplantation. *Blood* 95:3620-3627.
- Ankrum J. and Karp J.M. (2010) Mesenchymal stem cell therapy: Two steps forward, one step back. *Journal of Trends Mol Med* 16(5): 203–209.
- Aslan H., Zilberman Y., Kandel L., Libergall Meir., Ookouian R.J., Gazit D. and Gazit Z. (2006) Osteogenic Differentiation of Noncultured Immunoisolated Bone Marrow-Derived CD105 Cells. *Journal of Stem Cell* 24:1728–1737.
- Ayatollahi M., Salmani M.K., Geramizadeh B., Tabei S.Z., Soleimani M. and Sanati M.H. (2012) Conditions to improve expansion of human mesenchymal stem cells based on rat samples. *Journal of World J Stem Cells* 4(1):1-8.
- Ayuzawa R., Doi C., Rachakatla R.S., Pyle M.M., Maurya D.K., Troyer D. and Tamura M. (2009) Naïve human umbilical cord matrix derived stem cells significantly attenuate growth of human breast cancer cells in vitro and in vivo. *Journal of Cancer Lett.* 280(1):31–37.
- Baksh D., Yao R., Tuan R.S. (2007) Comparison of Proliferative and Multilineage Differentiation Potential of Human Mesenchymal Stem Cells Derived from Umbilical Cord and Bone Marrow. *Journal of Stem Cells* 25:1384–1392.
- Barberini D.J., Freitas N.P.P., Magnon M.S., Maia L., Listoni A.J., Heckler M.C., Sudano M.J., Golim M.A., Alvarenga F.C.L. and Amorim R.M. (2014) Equine mesenchymal stem cells from bone marrow, adipose tissue and umbilical cord: immunophenotypic characterization and differentiation potential. *Stem Cell Research & Therapy*, 5:25 <http://stemcellres.com/content/5/1/25>.
- Bartholomew A., Sturgeona C., Siatskasa M., Ferrera K., McIntosh K., Patil S., Hardy W., Devine S., Ucker D, Deans R., Moseley A. and Hoffman R. (2002) Mesenchymal stem cells suppress lymphocyte proliferation in vitro and prolong skin graft survival in vivo. *Journal of Experimental Hematology* 30:42–48.

Battula V.L., Treml S., Bareiss P.M., Gieseke F., Roelofs H., de Zwart P., Müller I., Schewe B., Skutella T., Fibbe W.E., Kanz L. and Bühring H-J (2009) Isolation of functionally distinct mesenchymal stem cell subsets using antibodies against CD56, CD271, and mesenchymal stem cell antigen-1. *Haematologica* 94:173-184. doi: 10.3324/haematol.13740.

Benvenuto F., Ferrari S., Gerdoni E., Gualandi F., Frassoni F., Pistoia V., Mancardi G., Uccelli A. (2007) Human Mesenchymal Stem Cells Promote Survival of T Cells in a Quiescent State. *Journal of Stem Cells* 25:1753–1760.

Blazsek I., Chagraoui J. and Peault B. (2000) Ontogenic emergence of the hematon, a morphogenetic stromal unit that supports multipotential hematopoietic progenitors in mouse bone marrow. *Journal of Blood* 96:3763-3771.

Bouffi C., Bony C., Courties G., Jorgensen C., Noe D. (2010) IL-6-Dependent PGE2 Secretion by Mesenchymal Stem Cells Inhibits Local Inflammation in Experimental Arthritis. *Plos One* 5(12): e14247. doi:10.1371/journal.pone.0014247.

Broere F., Apasov S.G., Sitkovsky M.V. and Van Eden W. (2011) T cell subsets and T cell-mediated immunity. DOI 10.1007/978-3-0346-0136-8_2.

Calvi L.M., Adams G.B., Weibrech K.W., Weber J.M., Olson D.P., Knight MC., Martin R. P., Schipani E., Divieti P., Bringhurst F. R., Milner L.A., Kronenberg H. M. & Scadden D.T. (2003) osteoblastic cells regulate the haematopoietic stem cell niche. *Journal of Nature* 425:841-846.

Caplan A.I. (1991) Mesenchymal Stem Cells. *Journal of Orthopaedic Research* 9:641-650.

Caplan A.I. and Westmichael D. (2014) Progressive Approval: A Proposal for a New Regulatory Pathway for Regenerative Medicine. *Journal of Stem Cells Translational Medicine* 3:560–563.

Cardoso T., Ferrari H., Garcia A., Novais J., Frade C.S., Ferrarezi M., Andrade A. and Gameiro R. (2012) Isolation and characterization of Wharton's jelly-derived multipotent mesenchymal stromal cells obtained from bovine umbilical cord and maintained in a defined serum-free three-dimensional system. *Journal of Cardoso et al. BMC Biotechnology* 12:18.

Chabannes D., Hill M., Merieau E., Rossignol J., Brion R., Soullou J.P., Anegon I. and Cuturi M.C. (2007) Role for heme oxygenase-1 in the immunosuppressive effect of adult rat and human mesenchymal stem cells. *Journal of The American Society* 110:3691-3694.

Chamberlain G., Fox J., Ashton B., Middleton J. (2007) Concise Review: Mesenchymal Stem Cells: Their Phenotype, Differentiation Capacity, Immunological Features, and Potential for Homing. *Journal of Stem Cells* 25:2739–2749.

- Chen L., Tredget E.E., Wu P.Y.G., Wu Y. (2008) Paracrine Factors of Mesenchymal Stem Cells Recruit Macrophages and Endothelial Lineage Cells and Enhance Wound Healing. *Plos One* 3(4): e1886. doi:10.1371/journal.pone.0001886.
- Colter D.C., Class R., Di Girolamo C.M. and Prockop D.J. (2000) Rapid expansion of recycling stem cells in cultures of plastic-adherent cells from human bone marrow. *Journal of PNAS* 97(7):3213–3218.
- Conconi M.T., Di Liddo R., Tommasini M., Calore C. and Parnigotto P.P. (2011) Phenotype and Differentiation Potential of Stromal Populations Obtained from Various Zones of Human Umbilical Cord: An Overview. *The Open Tissue Engineering and Regenerative Medicine Journal* 4:6-20.
- Corcione A., Benvenuto F., Ferretti E., Giunti D., Cappiello V., Cazzanti F., Rissi M., Gualandi F., Mancardi G.L., Pistoia V. and Uccelli A. (2006) Human mesenchymal stem cells modulate B-cell functions. *Journal of The American Society of Hematology* 107:367-372.
- Cortes Y., Ojeda M., Araya D., Dueñas F., Fernández M. and Peralta O. (2013) Isolation and multilineage differentiation of bone marrow mesenchymal stem cells from abattoir-derived bovine fetuses. *Journal of BMC Veterinary Research* 9:133.
- Cottet S. and Schorderet D.F. (2009) Mechanisms of Apoptosis in Retinitis Pigmentosa. *Current Molecular Medicine* 9:375-383.
- Crisostomo P., Wang M., Wairiuko G., Morrell E., Terrell A., Seshadri P., Nam U.H. and Meldrum D. (2006) High Passage Number of Stem Cells Adversely Affects Stem Cell Activation and Myocardial Protection. *Journal of Shock Society* 26(6):575-580.
- Cutler A.J., Limbani V., Girdlestone J. and Navarrete C. (2010). Umbilical Cord-Derived Mesenchymal Stromal Cells Modulate Monocyte Function to Suppress T Cell Proliferation. *The Journal of Immunology* 185:6617– 6623.
- Dae-W.K., Staples M., Shinozuka K., Pantcheva P., Sung-D.K and Borlongan C.V. (2013) Wharton's Jelly-Derived Mesenchymal Stem Cells: Phenotypic Characterization and Optimizing Their Therapeutic Potential for Clinical Applications. *Int. J. Mol. Sci.* 14:11692-11712; doi:10.3390/ijms140611692.
- Dalous J., Larghero J. and Baud O. (2012) Transplantation of umbilical cord-derived mesenchymal stem cells as a novel strategy to protect the central nervous system:technical aspects, preclinical studies, and clinical perspectives. *Journal of Pediatric Research* 71(4):482-488.
- Dang L.T-T., Bui A.N-T., Pham V.M., Phan N.K., Pham P.V (2015) Production of islet-like insulin-producing cell clusters in vitro from adipose-derived stem cells. *Biomedical Research and Therapy* 2(1):184-192.

- Dardalhon V., Korn T., Kuchroo V. and Anderson A. (2008) Role of Th1 and Th17 cells in organ-specific autoimmunity. *Journal of Autoimmun* 3:252–256.
- da Silva M.L., Chagastelles P.C and Nardi N.B. (2006) Mesenchymal stem cells reside in virtually all post-natal organs and tissues. *Journal of Cell Science* 119(11):2204-2213.
- Deng J., Petersen B., Steindler D., Jorgensen M., Laywell E. (2006) Mesenchymal Stem Cells Spontaneously Express Neural Proteins in Culture and Are Neurogenic after Transplantation. *Journal of Stem Cells* 24:1054 –1064.
- Di Nicola M., Stella C.Carlo., Magni M., Milanesi M., Longoni P.D., Matteucci P., Grisanti S. and Gianni A. (2002) Human bone marrow stromal cells suppress T-lymphocyte proliferation induced by cellular or nonspecific mitogenic stimuli. *Journal of The American Society of Hematology* 99:3838-3843.
- D'Ippolito G., Schiller P., Ricordi C., Roos B. and Howard G. (1999) Age-Related Osteogenic Potential of Mesenchymal Stromal Stem Cells from Human Vertebral Bone Marrow. *Journal of Bone and Mineral Research* 14:1115–1122.
- Divya M., Roshin G., Divya T., Abdul R.V., Santhoshkumar T., Elizabeth K., James J. and Pillai R. (2012) Umbilical cord blood-derived mesenchymal stem cells consist of a unique population of progenitors co-expressing mesenchymal stem cell and neuronal markers capable of instantaneous neuronal differentiation. *Stem Cell Research & Therapy* 3(57):1-16.
- Dominici M., Le Blanc K., Mueller I., Cortenbach S., Marini F.C., Deans K., Keating A., Prockop and Horwitz E.M. (2006) Minimal criteria for defining multipotent mesenchymal stromal cells. The International Society for Cellular Therapy position statement. *Journal of Cytotherapy* 8(4):315-317.
- Duff S., Li C., Garland J. and Kumar S. (2003) CD105 is important for angiogenesis: evidence and potential applications. *Journal of FASEB J.* 17:984–992.
- Eliopoulos N., Stagg J., Lejeune L., Pomme S. and Galipeau J. (2005) Allogeneic marrow stromal cells are immune rejected by MHC class I and class II mismatched recipient mice. *Journal of The American Society of Hematology* 106:4057-4065.
- Elmore S. (2007) Apoptosis: A Review of Programmed Cell Death. *Journal of Toxicol Pathol* 35(4): 495–516.
- English K., French A. and Wood K.J. (2010) Mesenchymal Stromal Cells: Facilitators of Successful transplantation?. *Cell Stem Cell* 7:431-442.
- English K., Ryan J.M., Tobin L., Murphy M.J., Barry F.P. and Mahon B.P. (2009) Cell contact, prostaglandin E2 and transforming growth factor beta 1play non-redundant roles in human mesenchymal stem cell induction of

- CD4+CD25High forkhead box P3+ regulatory T cells. Clinical and Experimental Immunology, 156: 149–160.
- Fan G., Wen L., Li M., Li C., Luo B., Wang F., Zhou L. and Liu L. (2011) Isolation of mouse mesenchymal stem cells with normal ploidy from bone marrows by reducing oxidative stress in combination with extracellular matrix. BMC Cell Biology 12:30 <http://www.biomedcentral.com/1471-2121/12/30>.
- Ferrer S.M., Michurina T., Ferraro F., Mazloom A., MacArthur B., Lira S., Scadden D.T., Ma'ayan A., Enikolopov G.N. and Frenette P.S. (2010) Mesenchymal and haematopoietic stem cells form a unique bone marrow niche. Journal of Nature 466:829-833.
- Fleming H.E., Janzen V., Lo Celso C., Guo J., Leahy K.M., Kronenberg H.M. and Scadden D.T. (2008) Wnt signaling in the niche enforces hematopoietic stem cell quiescence and is necessary to preserve self-renewal in vivo. Journal of Cell Stem Cell 2(3):274–283.
- Friedenstein A.J., Chailakhyan R.K. and Lalykina K.S. (1970) The development of fibroblast colonies in monolayers cultures of guinea-pig bone marrow and spleen cells. Cell Tissue Kinet 3:393-403.
- Friedenstein A.J., Chailakhyan R.K., Latsinik N.V., Panasyuk A.F and Keiliss-Borok I.V. (1974a) Stromal cells responsible for transferring the microenvironment of the haemopoietic tissues. Cloning in vitro and retransplantation in vivo. Transplantation 17:331-340.
- Fuchs E., Tumbar T. and Guasch G. (2004) Socializing with the Neighbors: Review Stem Cells and Their Niche. Journal of Cell Press 116:769–778.
- Gao Y., Yao A., Zhang W., Lu S., Yu Y., Deng L., Yin A., Xia Y., Sun B. and Wang X. (2010) Human mesenchymal stem cells overexpressing pigment epithelium-derived factor inhibit hepatocellular carcinoma in nude mice. Journal of Oncogene 29:2784–2794.
- Gao Y., Bai C., Xiong H., Li Q., Shan Z., Huang L., Ma Y. and Guan W. (2013) Isolation and Characterization of Chicken Dermis-Derived Mesenchymal Stem/Progenitor Cells. BioMed Research International. Article ID 626258, 8 pages <http://dx.doi.org/10.1155/2013/626258>.
- Geske FJ., Lieberman R., Strange R. and Gerschenson LE. (2001) Early stages of p53 induced apoptosis are reversible. Journal of Cell Death and Differentiation 8:182-191.
- Gieseke F., Bochner J., Bussolari R., Dominici M., Handgretinger R. and Muller I. (2010) Human multipotent mesenchymal stromal cells use galectin-1 to inhibit immune effector cells. Blood 116(19):3770-3779.

Gorcynski R.M. (2012) CD200:CD200R-Mediated Regulation of Immunity. International Scholarly Research Network ISRN Immunology, Article ID 682168, 18 pages, doi:10.5402/2012/682168.

Goyeneche A.A., Harmon J.M. and Telleria C.M. (2006) Cell death induced by serum deprivation in luteal cells involves the intrinsic pathway of apoptosis. Journal of Reproduction 131:103–111.

Gronthos S., Zannettino A.C.W., Hay S.J., Shi S., Graves S.E., Kortesidis A. and Simmons P.J. (2003) Molecular and cellular characterisation of highly purified stromal stem cells derived from human bone marrow. Journal of Cell Science 116:1827-1835.

Gu Y., Xue Q., Chen Y., Yu G.H., Qing M., Shen Y., Wang M., Shi Q., Zhang X.G. (2013) Different roles of PD-and FasL in immunomodulation mediated by human placenta-derived mesenchymal stem cells. Journal of Human Immunology 74: 267–276.

Hackett C.H., Flaminio M.J.B. F. and Fortier L. (2011) Analysis of CD14 Expression Levels in Putative Mesenchymal Progenitor Cells Isolated from Equine Bone Marrow. Journal Mary Ann Liebert, Inc. 20:721-735.

Härkönen P.L. and Laine J.K. (2001) Cell Proliferation Assay by Using MicroBeta 3 H-Thymidine Incorporation. Perkin Elmer Life Sciences, Inc.

Harris D.T. (2014) Stem Cell Banking for Regenerative and Personalized Medicine. Journal of Biomedicines 2:50-79.

Hayward C.J., Fradette J., Galbraith T., Rémy M., Guignard R., Gauvin R., Germain L., Auger F.A. (2012) Harvesting the Potential of the Human Umbilical Cord: Isolation and Characterisation of Four Cell Types for Tissue Engineering Applications. Cells Tissues Organs DOI: 10.1159/000341254.

Hass R., Kasper C., Böhm S. and Jacob R. (2011) Different populations and sources of human mesenchymal stem cells (MSC): A comparison of adult and neonatal tissue-derived MSC. Cell Communication and Signaling, 9:12http://www.biosignaling.com/content/9/1/12.

Hsiao S.T.Feng., Asgari A., Lokmic Z., Sinclair R., Dusling G.J., Lim S.Y. and Dilley R.J. (2012) Comparative Analysis of Paracrine Factor Expression in Human Adult Mesenchymal Stem Cells Derived from Bone Marrow, Adipose, and Dermal Tissue. Journal of Mary Ann Liebert, Inc 21(12):2189-2203.

Hsieh J-Y., Wang H-W., Chang S-J., Liao K-H., Lee H., Lin W-S., Wu C-H., Lin W-Y., Cheng S-M. (2013) Mesenchymal Stem Cells from Human Umbilical Cord Express Preferentially Secreted Factors Related to Neuroprotection, Neurogenesis, and Angiogenesis. Plos One 8(8): e72604. doi:10.1371/journal.pone.0072604.

- Hung S-C., Pochampally R.R., Hsu S-C., Sanchez C., Chen S-C., Spees J., Prockop D.J. (2007) Short-Term Exposure of Multipotent Stromal Cells to Low Oxygen Increases Their Expression of CX3CR1 and CXCR4 and Their Engraftment In Vivo. *Plos One* 2(5): e416. doi:10.1371/journal.pone.0000416.
- Itahana K., Dimri G. and Campisi J. (2001) Regulation of cellular senescence by p53. *Journal of Eur. J. Biochem.* 268:2784-2791.
- Jaenisch R. and Young R. (2008) Stem cells, the molecular circuitry of pluripotency and nuclear reprogramming. *Journal of Cell* 132(4):567–582.
- Jiang X-X., Zhang Y., Liu B., Zhang S-X., Wu Y., Yu X-D. and Mao N. (2005) Human mesenchymal stem cells inhibit differentiation and function of monocyte-derived dendritic cells. *Journal of The American Society of Hematology* 105:4120-4126.
- Jin H.J., Bae Y.K., Kim M., Kwon S-J., Jeon H.B., Choi S.J., Kim S.W., Yang Y.S., Oh W. and Chang J.W. (2013) Comparative Analysis of Human Mesenchymal Stem Cells from Bone Marrow, Adipose Tissue, and Umbilical Cord Blood as Sources of Cell Therapy. *International Journal of Molecular Sciences* 14:17986-18001.
- Jing W., Chen Y., Lu L., Hu X., Shao C., Zhang Y., Zhou X., Zhou Y., Wu L., Liu R., Fan K., Jin G. (2014) Human umbilical cord blood- derived mesenchymal stem cells producing IL-15 eradicate established pancreatic tumour in syngeneic mice. DOI: 10.1158/1535-7163.MCT-14-0175.
- Kalaszczynska I. and Ferdyn K. (2015) Wharton's Jelly Derived Mesenchymal Stem Cells: Future of Regenerative Medicine? Recent Findings and Clinical Significance. *BioMed Research International*, Article ID 430847, 11 pages, <http://dx.doi.org/10.1155/2015/430847>.
- Kavanagh H. & Mahon B. P. (2011) Allogeneic mesenchymal stem cells prevent allergic airway inflammation by inducing murine regulatory T cells. *Journal of Allergy* 66:523–531.
- Karahuseyinoglu S., Cinar O., Kilic E., Kara F., Akay G.G., Demiralp D., Tukun A., Uckan D., Can A. (2007) Biology of Stem Cells in Human Umbilical Cord Stroma: *In Situ* and *In Vitro* Surveys. *Journal of Stem Cells* 25:319–331.
- Kazemnejad S., Allameh A., Soleimani M., Gharehbaghian A., Mohammadi Y., Amirizadeh N. and Jazayery M. (2009) Biochemical and molecular characterization of hepatocyte-like cells derived from human bone marrow mesenchymal stem cells on a novel three-dimensional biocompatible nanofibrous scaffold. *J Gastroenterol Hepatol* 24:278-287.
- Kern S., Eichler H., Stoeve J., Kluther H., Bieback K. (2006) Comparative Analysis of Mesenchymal Stem Cells from Bone Marrow, Umbilical Cord Blood, or Adipose Tissue. *Journal of Stem Cells* 24:1294–1301.

- Kim D.H., Yoo K.H., Yim Y.S., Choi J., Lee S.H., Jung H.L., Sung K.W., Yang S-E., Oh W.I., Yang Y-S., Kim S-H., Choi S-Y., Koo H.H. (2006) Cotransplanted Bone Marrow Derived Mesenchymal Stem Cells (MSC) Enhanced Engraftment of Hematopoietic Stem Cells in a MSC-dose Dependent Manner in NOD/SCID Mice. Journal of The Korean Academy of Medical Sciences 21:1000-1004.
- Kim D-W., Staples M., Shinozuka K., Pantcheva P., Kang S-D. and Borlongan C. (2013) Wharton's Jelly-Derived Mesenchymal Stem Cells: Phenotypic Characterization and Optimizing Their Therapeutic Potential for Clinical Applications. International Journal of Molecular Sciences 14:11692-11712.
- Kita K., Gauglitz G.G., Phan T.T., Herndon D.N. and Jeschke M.G. (2010) Isolation and Characterization of Mesenchymal Stem Cells From the Sub-Amniotic Human Umbilical Cord Lining Membran. Journal of Mary Ann Liebert, Inc. 19(4):491-501.
- Kluth S.M., Radke T.F. and Kögler G. (2013) Increased Haematopoietic Supportive Function of USSC from Umbilical Cord Blood Compared to CB MSC and Possible Role of DLK-1. Article ID 985285, 12 pages <http://dx.doi.org/10.1155/2013/985285>.
- Kögler G., Sensken S., Airey J., Trapp T., Müschen M, Feldhahn,Niklas, Liedtke S., Sorg Rudiger., Johannes F., Rosenbaum C., Greschat S., Knipper A., Bender J., Degistirici Ö., Gao J., Caplan A., Colletti E.J., Porada G.A., Müller H.W., Zanjani E. and Weernet P. (2004) A New Human Somatic Stem Cell from Placental Cord Blood with Intrinsic Pluripotent Differentiation Potential. The Journal of Experimental Medicine 200(2):123–135.
- Kopen G.C., Prockop D.J. and Phinney D. (1999) Marrow stromal cells migrate throughout forebrain and cerebellum, and they differentiate into astrocytes after injection into neonatal mouse brains. Journal of Proc. Natl. Acad. Sci. USA 96:10711–10716.
- Krampera M., Cosmi L., Angeli R., Pasini A., Liotta F., Andreini A., Santarasci V., Mazzinghi B., Pizzolo G., Vinnate F., Romagni P., Maggi E., Romagni S., Annunziato F. (2006) Role for Interferon in the Immunomodulatory Activity of Human Bone Marrow Mesenchymal Stem Cells. Journal of Stem Cells 24:386–398.
- Krysko DV., Denecker G., Festjens N., Gabriels S., Parthoens E., D'Herde K. and Vandenabeele P. (2006) Macrophages use different internalization mechanisms to clear apoptotic and necrotic cells. Journal of Cell Death and Differentiation 13:2011–2022.
- Kurokawa T., Fischer K., Bertz H., Hoegerle S., Finke J. and Mackensen A. (2002) In vitro and in vivo charaterisation of Graft-Versus-Tumor responses in melanoma patients after allogenic peripheral blood stem cell transplantation. Journal of Int.J.Cancer 101:52–58.

- Le Blanc K., Tammik L., Sundberg B., Haynesworth S.E. Ringden O. (2003) Mesenchymal Stem Cells Inhibit and Stimulate Mixed Lymphocyte Cultures and Mitogenic Responses Independently of the Major Histocompatibility Complex. Scandinavian Journal of Immunology 57:11-20.
- Lee J.W., Fang X., Krasnodembskaya A., Howard J.P. and Matthay M.A. (2011) Concise Review: Mesenchymal Stem Cells for Acute Lung Injury: Role of Paracrine Soluble Factors. Journal of Stem Cells 29(6):913–919.
- Li D., Wang N., Zhang L., Hanyu Z., Xueyuan B., Fu B., Shaoyuan C., Zhang W., Xuefeng S., Li R. and Chen X. (2013) Mesenchymal stem cells protect podocytes from apoptosis induced by high glucose via secretion of epithelial growth factor. Stem Cell Research & Therapy, 4:103, <http://stemcellres.com/content/4/5/103>.
- Liechty K.W., Mackenzie T., Shaaban A., Radu A., Moseley A., Deans R., Marshak D.R. & Flake A.W. (2000) Human mesenchymal stem cells engraft and demonstrate sites specific differentiation after in utero transplantation in sheep. Journal of Nature America 6(11):1282-1286.
- Li W.W., Wei Y.H., Li H., Lai D.M. and Lin T.N. (2013) Isolation and Characterization of a Novel Strain of Mesenchymal Stem Cells from Mouse Umbilical Cord: Potential Application in Cell-Based Therapy. Plos One 8(8): e74478. doi:10.1371/journal.pone.0074478.
- Locke M., Windsor J. and Dunbar P.R. (2009) Human adipose-derived stem cells: isolation, characterisation and applications in surgery. ANZ J Surg 79:235-244.
- Loeb C.R.K., Harris J.L. and Craik C.S. (2006) Granzyme B Proteolyzes Receptors Important to Proliferation and Survival, Tipping the Balance toward Apoptosis. The Journal of Biological Chemistry 281(38):28326–28335.
- Lucas M., Daniel L., Tomasello E., Guia S., Horschowski N., Aoki N., Branger D.F., Gomez S. and Vivier E. (2002) Massive inflammatory syndrome and lymphocytic immunodeficiency in KARAP/DAP12-transgenic mice. Journal of Eur. J. Immunol 32:2653–2663.
- Lu L-L., Liu Y-J., Yang S-G., Zhao Q-J., Wang X., Gong W., Han Z-B., Xu Z-S., Lu Y-X., Liu D., Chen Z-Z., Han Z-C. (2006) Isolation and characterization of human umbilical cord mesenchymal stem cells with hematopoiesis supportive function and other potentials. Journal of Haematologica 91:1017-1026.
- Lund R.D., Wang S., Lu B., Girman S., Holmes T., Sauv'e Y., Mwssina D.J., Harris I.R., Kihm A.J., Harmon A.M., Chin F-Y., Gosiewska A., Mistry S.K. (2007) Cells Isolated from Umbilical Cord Tissue Rescue Photoreceptors and Visual Functions in a Rodent Model of Retinal Disease. Journal of Stem Cells 25:602–611.

- Ma S., Xie N., Li W., Yuan B., Shi Y., and Wang Y. (2014) Immunobiology of mesenchymal stem cells. *Cell Death and Differentiation* 21:216–225.
- Majore I., Moretti P., Stahl F., Hass R., Kasper C. (2010) Growth and Differentiation Properties of Mesenchymal Stromal Cell Populations Derived from Whole Human Umbilical Cord. *Stem Cell Rev and Rep DOI* 10.1007/s12015-010-9165-y.
- Marmotti A., Mattia S., Bruzzone M., Buttiglieri S., Risso A., D. Bonasia E., Blonna D., Castoldi F., Rossi R., Zanini C., Ercole E., Defabiani E., Tarella C. and Peretti G.M.(2012) Minced Umbilical Cord Fragments as a Source of Cells for Orthopaedic Tissue Engineering: An In Vitro Study. *Stem Cells International*, Article ID 326813, 13 pages, doi:10.1155/2012/326813.
- Maslova O., Shuvalova N.S., Sukhorad O.M., Zhukova S.M., Deryabina O.G., Makarenko M.V., Govseiev D.O. and Kordium V.A. (2013) Heterogeneity of Umbilical Cords as a Source for Mesenchymal Stem Cells. *Dataset Papers in Biology*, Article ID 370103, 4 pages, <http://dx.doi.org/10.7167/2013/370103>.
- McClory S., Hughes T., Freud A.G., Briercheck E.L., Martin C., Trimboli A.J., Yu J., Zhang X., Leone G., Nuovo G. and Caligiuri M. (2012) Evidence for a stepwise program of extrathymic T cell development within the human tonsil. *The Journal of Clinical Investigation* 122(4):1403–1415.
- Mellor A.L. and Munn D.H. (2004) IDO Expression by dendritic cells: Tolerance and tryptophan catabolism. *Journal of Nature Publishing Group* 4:762-774.
- Mennan C., Wright K., Bhattacharjee A., Balain B., Richardson J. and Roberts S. (2013) Isolation and Characterisation of Mesenchymal Stem Cells from Different Regions of the Human Umbilical Cord. *BioMed Research International*, Article ID 916136, 8 pages <http://dx.doi.org/10.1155/2013/916136>.
- Menssen A., Häupl T., Sittinger M., Delorme B., Charbord P. and Ringe J. (2011) Differential gene expression profiling of human bone marrow-derived mesenchymal stem cells during adipogenic development. *BMC Genomics* 12:461.<http://www.biomedcentral.com/1471-2164/12/461>.
- Mochizuki T., Muneta T., Sakaguchi Y., Nimura A., Yokoyama A., Koga H. and Sekiya I. (2006) Higher Chondrogenic Potential of Fibrous Synovium- and Adipose Synovium-Derived Cells Compared With Subcutaneous Fat-Derived Cells. *Journal of Arthritis & Rheumatism* 54(3):843–853.
- Morando S., Vigo T., Esposito M., Casazza S., Novi G., Principato M.C., Furlan R. and Uccelli A. (2012) The therapeutic effect of mesenchymal stem cell transplantation in experimental autoimmuneEncephalomyelitis is mediated by peripheral and central mechanisms. *Journal of Morando et al. Stem Cell Research & Therapy* 3(3):1-7.

- Mou X-Z., Lin J., Chen J-Y., Li Y-F., Wu X-X., Xiang B-Y., Li C-Y., Ma J-M., Xiang C. (2013) Menstrual blood-derived mesenchymal stem cells differentiate into functional hepatocyte-like cells. *Biomed & Biotechnol* 14(11):961-972.
- Mougiakakos D., Jitschin R., Johansson C., Okita R., Kiessling R. and Le Blanc K. (2011) The impact of inflammatory licensing on heme oxygenase-1-mediated induction of regulatory T cells by human mesenchymal stem cells. *Journal of Blood* 117(18):4826-4835.
- Muguruma Y., Yahata T., Miyatake H., Sato T., Uno T., Itoh J., Kato S., Ito M., Hotta T. and Ando K. (2006) Reconstitution of the functional human hematopoietic microenvironment derived from human mesenchymal stem cells in the murine bone marrow compartment. *Journal of Blood* 107:1878-1887.
- Nagamura T. and He H. (2014) Umbilical cord-derived mesenchymal stem cells: Their advantages and potential clinical utility. *World Journal Stem Cells* 6(2):195-202.
- Navabazam A.R., Nodoshan F.S., Sheikhha M.H., Miresmaeli S.M., Soleimani M., Fesahat F. (2013) Characterization of mesenchymal stem cells from human dental pulp, preapical follicle and periodontal ligament. *Iran Journal Reprod Med* 11(3):235-242.
- Nbaheen M., Vishnubalaji R., Ali D., Bouslimi A., Jassir F., Megges M., Prigione A., Adjaye J., Kassem M. and Aldahmash A. (2013) Human Stromal (Mesenchymal) Stem Cells from Bone Marrow, Adipose Tissue and Skin Exhibit Differences in Molecular Phenotype and Differentiation Potential. *Journal of Stem Cell Rev and Rep* 9:32-43.
- Németh K., Leelahavanichku A., Yuen P., Mayer B., Parmelee A., Doi K., Robey P., Leelahavanichkul K., Koller B.H., Brown J., Hu X., Jelinek I., Star R. and Mezey É. (2009) Bone marrow stromal cells attenuate sepsis via prostaglandin E2-dependent reprogramming of host macrophages to increase their interleukin-10 production. *Journal of Nat Med* 15(1):42-49.
- Orkin S.H. and Zon L.I. (2008) Hematopoiesis: An Evolving Paradigm for Stem Cell Biology. *Journal of Cell* 132(4):631-644.
- Ortiz L.A., Gambelli F., McBride C., Gaupp D., Baddoo M., Kaminski N. and Phinney D.G. (2003) Mesenchymal stem cell engraftment in lung is enhanced in response to bleomycin exposure and ameliorates its fibrotic effects. *Journal of PNAS* 100(14):8407-8411.
- Oswald J., Boxberger S., Jorgensen B., Feldmann S., Ehninger G., Bornhauser M., Werner C. (2004) Mesenchymal Stem Cells Can Be Differentiated Into Endothelial Cells In Vitro. *Stem Cells* 22:377-384.
- Park M., Kim Y-H., Woo S-Y., Lee H.J., Yu Y., Kim H.S., Park Y.S., Jo I., Park J-W., Jung S-C., Lee H., Jeong B., Ry K-H. (2014) Tonsil-derived Mesenchymal

Stem Cells Ameliorate CCl4-induced Liver Fibrosis in Mice via Autophagy Activation. Scientific Reports 5:8616 DOI:10.1038/srep08616.

Parkin J. and Cohen B. (2001) An overview of the immune system. *The Lancet* 357:1777–1789.

Patterson M.K. (1979) Measurement of growth and viability of cells in culture. *Methods Enzymol.* 58:141-152.

Pirjali T, N. Azarpira, M. Ayatollahi, M. H. Aghdaie, B. Geramizadeh, T. Talai (2013) Isolation and Characterization of Human Mesenchymal Stem Cells Derived from Human Umbilical Cord Wharton's Jelly and Amniotic Membrane. *Int J Org Transplant Med* 4(3):111-116.

Pittenger M.F, Mackay A.M, Beck S.C, Jaiswal R.K, Douglas R, Mosca J.D, Moorman M.A., Simonetti D.W, Craig S, Marshak D.R (1999) Multilineage potential of adult human mesenchymal stem cells. *Science* 284:143-147.

Plotnikov E.Y., Pulkova N.V., Pevzner I.B., Zorova L.D., Silachev D.N., Morosova M.A., Sukhikh G.T. & Zorov D.B. (2013) Inflammatory pre-conditioning of mesenchymal multipotent stromal cells improves their immunomodulatory potency in acute pyelonephritis in rats. *Cyotherapy* 15:679-689.

Potdar P.D. and Prasannan P. (2013) Differentiation of Human Dermal Mesenchymal Stem Cells into Cardiomyocytes by Treatment with 5-Azacytidine: Concept for Regenerative Therapy in Myocardial Infarction. *ISRN Stem Cells*, Article ID 687282, 9 pages <http://dx.doi.org/10.1155/2013/687282>.

Prasanna S.J., Gopalakrishnan D., Shankar S.R., Vasandan A.B. (2010) Pro-Inflammatory Cytokines, IFN γ and TNF α , Influence Immune Properties of Human Bone Marrow and Wharton Jelly Mesenchymal Stem Cells Differentially Stem Cell and Inflammation. *Plos One* 5(2): e9016. doi:10.1371/journal.pone.0009016.

Prasanna S.J. and Jahnavi V.S. (2011) "Wharton's jelly mesenchymal stem cells as off-the-shelf cellular therapeutics: a closer look into their regenerative and immunomodulatory properties," *The Open Tissue Engineering and Regenerative Medicine Journal* 4(1):28–38.

Preston S.L., Alison M.R., Forbes S.J., Direkze N.C., Poulsom R., Wright N.A. (2003) The new stem cell biology: something for everyone. *J Clin Pathol: Mol Pathol* 56:86–96.

RafeiM., BirmanE., FornerK and GalipeauJ. (2009) Allogeneic Mesenchymal Stem Cells for Treatment of Experimental Autoimmune EncephalomyelitisMolecular Therapy 17(10):1799–1803.

Raffaghelli L., Bianchi G., Bertolotto M., Montecucco F., Busca A., Dallegrì F., Otonello L., PistolaV. (2008) Human Mesenchymal Stem Cells Inhibit

- Neutrophil Apoptosis: A Model for Neutrophil Preservation in the Bone Marrow Niche. *Stem Cells* 26:151–162.
- Ramasamy R., Lam EW-F., Soeiro I., Tisato V., Bonnet D. and Dazzi F. (2007) Mesenchymal stem cells inhibit proliferation and apoptosis of tumor cells: impact on in vivo tumor growth. *Leukemia* 21:304–310.
- Ranganath S.H., Levy O., Inamdar M.S., and Karp J.M. (2012) Harnessing the Mesenchymal Stem Cell Secretome for the Treatment of Cardiovascular Disease. *Cell Stem Cell*. 10(3): 244–258.
- Rasmussen I., Uhlin M., Le Blanc K., and Levitsky V. (2007) Mesenchymal stem cells fail to trigger effector functions of cytotoxic T lymphocytes. *J. Leukoc. Biol.* 82:887–893.
- Rastogi R.P., Richa and Sinha R.P. (2009) Apoptosis: Molecular Mechanism and Pathogenicity. *EXCLI Journal* 8:155–181.
- Ratliff B.B., Singh N., Yasuda K., Park H-C., Addabbo F., Ghaly T., Rajdev M., Jasmin J-F., Plotkin M., Lisanti M.P. and Goligorsky M.S. (2010) Mesenchymal Stem Cells, Used As Bait, Disclose Tissue Binding Sites Tool in the Search for the Niche?. *Am J Pathol* 177:873–883.
- Ren G., Zhang L., Zhao X., Xu G., Zhang Y., Roberts A.I., Zhao R.C. and Shi Y. (2008) Mesenchymal Stem Cell-Mediated Immunosuppression Occurs via Concerted Action of Chemokines and Nitric Oxide. *Cell Stem Cell* 2:141–150.
- Ren G., Su J., Zhang L., Zhao X., Ling W., L'huillie A., Zhang, Jimin L., Yongqin., Roberts A.I., Ji W., Zhang H., Rabson A.B. and Shi Y. (2009) Species Variation in the Mechanisms of Mesenchymal Stem Cell-Mediated Immunosuppression. *Stem Cells* 27:1954–1962.
- Reyes M., Lund T., Lenvik T., Aguiar D., Koodie L. and Verfaillie C.M. (2001) Purification and ex vivo expansion of postnatal human marrow mesodermal progenitor cells. *Blood* 98:2615–2625.
- Ribeiro C.A., Salgado A.J., Fraga J.S., Silva N.A., Reis R.L. and Sousa N. (2011) The secretome of bone marrow mesenchymal stem cells-conditioned media varies with time and drives a distinct effect on mature neurons and glial cells (primary cultures). *J Tissue Eng Regen Med* DOI: 10.1002/term.365.
- Romanov Y.A., Svintsitskaya V.A., Smirnov V.N. (2003) Searching for Alternative Sources of Postnatal Human Mesenchymal Stem Cells: Candidate MSC-Like Cells from Umbilical Cord. *Stem Cells* 21:105–110.
- Roura S., Farre J., Soler-Botija C., Llach A., Hove-Madsen L., Cairo J., Go`dia F., Cinca J., Bayes-Genis A. (2006) Effect of aging on the pluripotential capacity

- of human CD105+ mesenchymal stem cells. European Journal of Heart Failure 8:555–563.
- Ruster B., Gottig S., Ludwig R.J., Bistrian, Roxana, Muller S., Seifried E., Gille J. and Henschler R. (2006) Mesenchymal stem cells display coordinated rolling and adhesion behavior on endothelial cells. Blood 108:3938-3944.
- Sacchetti B., Funari A., Michienzi S., Di Cesare S., Piersanti S., Saggio I., Tagliafico E., Ferrari S., Robey P.G., Riminiucci M. and Bianco P. (2007) Self-Renewing Osteoprogenitors in Bone Marrow Sinusoids Can Organize a Hematopoietic Microenvironment. Cell 131:324–336.
- Sandrasaigaran P., Vidyadarshan S., Ahmad A.R and Ramasamy R. (2014) Generation and Characterisation of Mesenchymal Stem Cells Derived from Human Cartilage Tissue. Regenerative Research 3(2):2014-2144.
- Sarugaser R., Hanoun L., Keating A., Stanford W.L., Davies J.E. (2009) Human Mesenchymal Stem Cells Self-Renew and Differentiate According to a Deterministic Hierarchy. Plos One 4(8): e6498. doi:10.1371/journal.pone.0006498.
- Sarugaser R., Lickorish D., Baksh D., Hosseini M.M., Davies J.E. (2005) Human Umbilical Cord Perivascular (HUCPV) Cells: A Source of Mesenchymal Progenitors. Stem Cells 23:220–229.
- Sato K., Ozaki K., Oh L., Meguro A., Hatanaka K., Nagai T., Muroi K. and Ozawa K. (2007) Nitric oxide plays a critical role in suppression of T-cell proliferation by mesenchymal stem cells. Blood 109:228-234.
- Scheers I., Lombard C., Najimi M. and Sokal E.M. (2011) Cell Therapy for the Treatment of Metabolic Liver Disease: An Update on the Umbilical Cord Derived Stem Cells Candidates. The Open Tissue Engineering and Regenerative Medicine Journal 4:48-53.
- Secco M., Zucconi E., Vieria N.M., Fogaca L.L.Q., Cerquira A., Carvalho M.D.F., Jazedje T., Okamoto O.K., Muotri A.R., Zatzmayana. (2008) Multipotent Stem Cells from Umbilical Cord: Cord Is Richer than Blood Stem Cells 26:146–150.
- Seo M.S., Jeong Y.H., Park J.R., Park S.B., Rho K.H., Kim H.S., Yu K.R., Lee S.H., Jung J.W., Lee Y.S., Kang K.S. (2009) Isolation and characterization of canine umbilical cord blood-derived mesenchymal stem cells. J. Vet. Sci. 10(3):181-187.
- Seshi B., Kumar S. and Sellers D. (2000) Human Bone Marrow Stromal Cell: Coexpression of Markers Specific for Multiple Mesenchymal Cell Lineages. Blood Cells, Molecules, and Diseases 26(3):234–246.

- Shi M., Li J., Liao L., Chen B., Li B., Chen L., Jia H., Zhao R.C. (2007) Regulation of CXCR4 expression in human mesenchymal stem cells by cytokine treatment: role in homing efficiency in NOD/SCID mice. *Haematologica* 92:897-904.
- Shi Y., Hu G., Su J., Li W., Chen Q., Shou P., Xu C., Chen X., Huang Y., Zhu Z., Huang X., Han X., Xie N., Ren G. (2010) Mesenchymal stem cells: a new strategy for immunosuppression and tissue repair. *Cell Research* 20:510-518.
- Shimizu S. and Tsujimoto Y. (2000) Proapoptotic BH3-only Bcl-2 family members induce cytochrome c release, but not mitochondrial membrane potential loss, and do not directly modulate voltage-dependent anion channel activity. *Nat. Academy of Sciences* 2:577-582.
- Sioud M., Mobergslien A., Boudabous A. & Floisand Y. (2010) Evidence for the Involvement of Galectin-3 in Mesenchymal Stem Cell Suppression of Allogeneic T-Cell Proliferation. *Scandinavian Journal of Immunology* 71:267-274.
- Smeets R.L., Fleuren W.W.M., He X., Vink P.M., Wijnands F., Gorecka M., Klop H., Bauerschmidt S., Garritsen A., Koenen H.J.P.M., Joosten I., Boots A.M.H. and Alkema W. (2012) Molecular pathway profiling of T lymphocyte signal transduction pathways; Th1 and Th2 genomic fingerprints are defined by TCR and CD28-mediated signaling. Smeets et al. *BMC Immunology*, 13:12, <http://www.biomedcentral.com/1471-2172/13/12>.
- Smith C. (2003) Hematopoietic Stem Cells and Hematopoiesis. *Cancer Control* 10(1):9-16. Sotiropoulou P.A., Perez S.A., Gritzapis A.D., Baxevanis C.N., Papamichail M. (2006) Interactions Between Human Mesenchymal Stem Cells and Natural Killer Cells. *Stem Cells* 24:74-85.
- Spaeth E., Klopp A., Dembinski J., Andreeff M. and Marini F. (2008) Inflammation and tumor microenvironments: defining the migratory itinerary of mesenchymal stem cells. *Gene Therapy* 15:730-738.
- Staykova M.A., Berven L.A., Cowden W.B., Willenborg D.O. and Crouch M.F. (2003) Nitric oxide induces polarization of actin in encephalitogenic T cells and inhibits their in vitro transendothelial migration in a p70S6 kinase-independent manner. *The FASEB Journal* express article10.1096/fj.02-0577fje.
- Stolzing A., Coleman N. and Scutt A. (2006) Glucose-Induce Replicative Senescence in Mesenchymal Stem Cells. *Rejuvenation Research* 9(1):31-35.
- Studenty M., Marini F.C., Champlin R.E., Zompetta C., Fidler I.J. and Andreeff M. (2002) Bone Marrow-derived Mesenchymal Stem Cells as Vehicles for Interferon- β Delivery into Tumors. *Cancer Research* 62:3603-3608.
- Sugiyama T., Kohara H., Noda M. and Nagasawa T. (2006) Maintenance of the Hematopoietic Stem Cell Pool by CXCL12-CXCR4 Chemokine Signaling in Bone Marrow Stromal Cell Niches. *Immunity* 25:977-988.

Sun L., Wang D., Liang J., Zhang H., Feng X., Wang H., Hua B., Liu B., Ye S., Hu X., Xu W., Zeng X., Hou Y., Gary S. Gilkeson,⁶ Richard M. Silver,⁶ Lu L. and Shi S. (2010) Umbilical Cord Mesenchymal Stem Cell Transplantation in Severe and Refractory Systemic Lupus Erythematosus. *Arthritis Rheumatism* 62(8):2467–2475.

Sung H.J., Hon S.C., Yoo J.H., Oh J.H., Shin H.J., Choi I.Y., Ahn K.H., Kim S.H., Park Y. and Kim B.S (2010) Stemness Evaluation of Mesenchymal Stem Cells from Placentas According to Developmental Stage: Comparison to Those from Adult Bone Marrow. *J Korean Med Sci* 25:1418-1426.

Svobodova E., Krulova M., Zajicova A., Pokorna K., Prochazkova J., Trosan P. and Holan V. (2012) The Role of Mouse Mesenchymal Stem Cells in Differentiation of Naive T Cells into Anti-Inflammatory Regulatory T-Cell or Proinflammatory Helper T-Cell 17 Population. *Stem Cells and Development* 21(6). DOI: 10.1089/scd.2011.0157.

Tabera S., Simón J.A.P., Campelo M.D., Luis I.A.S., Blanco B., López A., Benito A., Ocio., Guijo F.M.S., Cañizo C. and Miguel J.F.S. (2008) The effect of mesenchymal stem cells on the viability, proliferation and differentiation of B-lymphocytes. *Haematologica* 93:1301-1309.

Taichman R.S., Reilly M.J., Verma R.S. and Emerson S.G. (1997) Augmented Production of Interleukin-6 by Normal Human Osteoblasts in Response to CD34⁺ Hematopoietic Bone Marrow Cells In Vitro. *Blood* 89(4):1165-1172.

Takahashi K. and Yamanaka S. (2006) Induction of Pluripotent Stem Cells from Mouse Embryonic and Adult Fibroblast Cultures by Defined Factors. *Cell* 126:663–676.

Thevenot P., Nair A., Shen J., Lotfi P., Ko C.Yu. and Tang L. (2010) The Effect of Incorporation of SDF-1 α into PLGA Scaffolds on Stem Cell Recruitment and the Inflammatory Response. *Biomaterials* 31(14):3997–4008.

Tian L.L.H., Yue W., Zhu F., Li S. and Li W. (2011) Human Mesenchymal Stem Cells Play a Dual Role on Tumor Cell Growth In Vitro and In Vivo. *J. Cell. Physiol.* 226:1860–1867.

Tipnis S., Viswanathan C. and Majumdar A.S. (2010) Immunosuppressive properties of human umbilical cord-derived mesenchymal stem cells: role of B7-H1 and IDO. *Immunology and Cell Biology* 88:795–806.

Tondreau T., Meuleman N., Delforge A., Dejeneffe M., Leroy R., Massy M., Mortier C., Bron D. and Lagneaux L. (2005) Mesenchymal Stem Cells Derived from CD133- Positive Cells in Mobilized Peripheral Blood and Cord Blood: Proliferation, Oct4 Expression, and Plasticity. *Stem Cells* 23:1105–1112.

Tong C.K., Vellasamy S., Tan B.C., Abdullah M., Vidyadarshan S., Seow H.F. and Ramasamy R. (2011) Generation of mesenchymal stem cell from human

- umbilical cord tissue using a combination enzymatic and mechanical disassociation method. *Cell Biol. Int.* 35:221–226.
- Tyson J.J., Nagy A.C. and Novak B. (2002) The dynamics of cell cycle regulation. *BioEssays* 24:1095–1109.
- Uccelli A. and Moretta L. (2008) Mesenchymal stem cells in health and disease. *Net Rev Immunol* 8(9):726–736.
- van Poll D., Parekkadan B., Cho C., Berthiaume F., Nahmias Y., Tilles A. and Yarmush M.L. (2008) Mesenchymal Stem Cell-Derived Molecules Directly Modulate Hepatocellular Death and Regeneration In Vitro and In Vivo. *Hepatology* 47:1634–1643.
- VellasamyS., SandrasaigaranP., VidyadarshanS., GeorgeE., RamasamyR. (2012) Isolation and characterisation of mesenchymal stem cells derived from human placenta tissue. *World J Stem Cells* 4(6):53-61.
- VellasamyS., VidyadarshanS., GeorgeE., Mahenderan A., RamasamyR. (2013) Immunosuppressive activity of Human Umbilical Cord and Placenta derived Mesenchymal Stem Cells on Lymphocytes' Proliferation. *Regenerative Research* 2(1):41-49.
- Vermulenn K, Van B.D.R. and Berneman Z.N. (2003) The cell cycle: a review of regulation, deregulation and therapeutic targets in cancer. *Cell Proliferation* 36(3):131–149.
- Vermes, I.n., C. Haanen, H. Steffens-Nakken and C. Reutellingsperger (1995) A novel assay for apoptosis flow cytometric detection of phosphatidylserine expression on early apoptotic cells using fluorescein labelled annexin V. *Journal of immunological methods* 184:39-51.
- Vidal M.A., Kilroy G.E., Lopez M.J., Johnson J.R., Moore R.M. and Gimble J.M. (2007) Characterization of Equine Adipose Tissue-Derived Stromal Cells: Adipogenic and Osteogenic Capacity and Comparison with Bone Marrow-Derived Mesenchymal Stromal Cells. *Veterinary Surgery* 36:613–622.
- Vieira N. M., Brandalise V., Zucconi E., Secco M., Strauss B. E. and Zatz M. (2010) Isolation, Characterization, and Differentiation Potential of Canine Adipose-Derived Stem Cells. *Cell Transplantation* 19:279–289.
- Wan Y.Y. and Flavell R.A. (2009) How Diverse—CD4 Effector T Cells and their Functions. *Journal of Molecular Cell Biology* 1:20–36.
- Wang S., Qu X. and Zhao R.C. (2012) Clinical applications of mesenchymal stem cells. *Journal of Hematology & Oncology* 5:19. <http://www.jhoonline.org/content/5/1/19>.

Wang, Y., Z. Zhang, et.al (2013) Long-term cultured mesenchymal stem cells frequently develop genomic mutations but do not undergo malignant transformation. *Cell Death Dis* 4:e950.

Williams A.R., Hatzistergos K.E., Addicott B., McCall F., Carvalho D., Suncion V., Morales A., Da Silva J., Sussman M.A., Heldman A.W. and Hare J.M. (2013) Enhanced Effect of Human Cardiac Stem Cells and Bone Marrow Mesenchymal Stem Cells to Reduce Infarct Size and Restore Cardiac Function after Myocardial Infarction. *Circulation* 127(2):213–223.

Williams A.R. and Hare J.M. (2011) Mesenchymal stem cells: Biology, pathophysiology, translational findings, and therapeutic implications for cardiac disease. *Circ Res* 109(8):923–940.

Wu Y., Wang Z., Cao Y., Xu L., Li X., Liu P., Yan P., Liu Z., Zhao D., Wang J., Wu X., Gao C., Da W., Han Z. (2013) Cotransplantation of haploidentical hematopoietic and umbilical cord mesenchymal stem cells with a myeloablative regimen for refractory/relapsed hematologic malignancy 92(12):1675-1684.

Wu Y., Cao Y., Li X., Xu L., Wang Z., Liu P., Yan P., Liu Z., Wang J., Jiang S., Wu X., Gao C., Da W., Han Z. (2014) Cotransplantation of haploidentical hematopoietic and umbilical cord mesenchymal stem cells for severe aplastic anemia: Successful engraftment and mild GVHD. *Stem Cell Research* 12:132–138.

Xiong Y., Mahmood A. and Chopp M. (2010) Angiogenesis, neurogenesis and brain recovery of function following injury. *Curr Opin Investig Drugs* 11(3):298–308.

Xu G., Zhang Y., Zhang L., Ren G. and Shi Y. (2007) The Role of IL-6 in Inhibition of Lymphocyte Apoptosis by Mesenchymal Stem Cells. *Biochem Biophys Res Commun.* 361(3):745–750.

Yin Y., Danliang C., Xuan C., Hongwei S. and Shulin H. (2014) Human umbilical cord-derived mesenchymal stem cells inhibit proliferation but maintain survival of Jurkat leukemia cells in vitro by activating Notch signaling. *J South Med Univ.* 34(4):441-447.

Yu Q., Chen L., You Y., Zou C., Zhang Y., Liu Q. and Cheng F. (2011) Erythropoietin combined with granulocyte colony-stimulating factor enhances MMP-2 expression in mesenchymal stem cells and promotes cell migration. *Molecular Medicine Reports* 4:31-36.

Yueying M., Shuolong Y., Yue Z., Liangwei X., Weiwei G., Lidong Z., Suoqiang Z., Shiming Y. (2014) Isolation and induction of differentiation of seine adipose-derived mesenchymal stem cells. *Journal of Otology* 9(2):101-105.

Zappia E., Casazza S., Pedemonte E., Benvenuto F., Bonanni I., Gerdoni E., Giunti D., Ceravolo A., Cazzanti F., Frassoni F., Mancardi G. and Uccelli A. (2005)

- Mesenchymal stem cells ameliorate experimental autoimmune encephalomyelitis inducing T-cell anergy. Blood 106:1755-1761.
- Zaragozi L.E., Ailhaud G., Dani C. (2006) Autocrine Fibroblast Growth Factor 2 Signaling Is Critical for Self-Renewal of Human Multipotent Adipose-Derived Stem Cells. Stem Cells 24:2412–2419.
- Zhang B., Liu R., Shi D., Liu X., Chen Y., Dou X., Zhu X., Lu C., Liang W., Liao L., Zenke M. and Zhao R.C.H. (2009) Mesenchymal stem cells induce mature dendritic cells into a novel Jagged-2-dependent regulatory dendritic cell population. Blood 113:46-57.
- Zhou R., Li Z., He C., Li R., Xi H., Li C., Xiao J. and Che Z.Y. (2014) Human Umbilical Cord Mesenchymal Stem Cells and Derived Hepatocyte-Like Cells Exhibit Similar Therapeutic Effects on an Acute Liver Failure Mouse Model. Plos One 9(8): e104392. doi:10.1371/journal.pone.0104392.

Zhu Y., Yang Y., Zhang Y., Hao G., Liu T., Wang L., Yang T., Qiong W., Guangyi Z., Jun W. and Yukui L. (2014) Placental mesenchymal stem cells of fetal and maternal origins demonstrate different therapeutic potentials. Stem Cell Research & Therapy 5:48. <http://stemcellres.com/content/5/2/48>.