

UNIVERSITI PUTRA MALAYSIA

PHARMACOLOGICAL PROPERTIES OF CHANNA SPP. EXTRACTS

AZLINA BINTI MOHD. JOHARI

FPV 2008 1



PHARMACOLOGICAL PROPERTIES OF CHANNA SPP. EXTRACTS

Ву

AZLINA BINTI MOHD. JOHARI

Thesis Submitted to the School of Graduates Studies, Universiti Putra Malaysia, In Fulfilment of the Requirement for the Degree of Master of Science

Mac 2008





Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

ANTI-INFLAMMATORY, ANTINOCICEPTIVE AND ANTIPYRETIC EFFECTS OF EXTRACTS FROM SNAKEHEAD FISH (Channa striatus AND C. lucius)

By

AZLINA BINTI MOHD. JOHARI

March 2008

Chairman: Associate Professor Arifah Abdul Kadir, PhD

Faculty : Veterinary Medicine

Channa spp. is a snakehead fish and widely consumed in Malaysia. A study was conducted to determine the effect of *Channa striatus* and *Channa lucius* extracts on carrageenan-induced synovitis in rabbits. The antinociceptive and antipyretic properties of both *Channa spp.* were also investigated in mice. The extracts of C. striatus and C. lucius were prepared as water and aqueous portion of chloroform: methanol extracts, respectively. Sixteen rabbits were randomly assigned into four groups. Each group of rabbits was treated orally 30 minutes before the induction of inflammation with C. striatus (60 mg/kg), C. lucius (60 mg/kg), ketoprofen (3 mg/kg) and saline solution (control), respectively. The right stifle joint was intra-articularly injected with 0.5 mL of 1% carrageenan. Whole blood was taken for serum thromboxane (TxB₂) assay before and at 1, 3, 6, 9, 12 and 24 hour (h) after the induction of inflammation. Synovial fluid and synovial membrane were collected during post-mortem for analysis and histopathology. The results indicated that TxB_2 synthesis was significantly (p<0.05) inhibited for 12 h after oral dosing of ketoprofen in rabbits. Serum TxB₂ for *C. striatus* was lower than that of the control group at 1, 3, 6 and 12 h. As for C. lucius, TxB₂



synthesis was inhibited at 9 and 12 h. However, the inhibition of TxB₂ for both Channa spp. was small and not significant as compared to the control group. As for *C. striatus* and ketoprofen treated groups, the total white blood cell (WBC) count was reduced compared to the control group but was not significant different. Histopathological results indicated a mild infiltration of leucocytes in the synovial membrane of ketoprofen treated rabbits. However in the control, C. striatus and C. lucius treated groups showed massive leucocyte infiltration, congestion in the blood vessels and fibrin exudation. As for the analgesic activity of Channa spp., twenty four mice were allocated equally into three treatment groups and one control group. The extracts of C. striatus (60 mg/kg), C. lucius (60 mg/kg) or ketoprofen (1 mg/kg) was administered intraperitoneally, 30 minutes before injection of acetic acid. Both extracts of the local Channa spp. and ketoprofen showed significant (p<0.05) reductions in the number of abdominal constriction and hind limb stretching as compared to the control group. As for the antipyretic effect of the Channa spp. extracts, twenty four mice were equally divided into three treatment groups and one control group. Mice were injected with 30% (w/v) suspension of yeast in saline at the dosage of 10 mL/kg subcutaneously. The temperature was recorded 18 hour before and measured every half an hour for 5 hours after dosing. C. striatus (60 mg/kg) and C. lucius (60 mg/kg) reduced hyperthermia significantly (p<0.05) at 2.5 to 5 h and 3 to 4 h, respectively. Ketoprofen (1 mg/kg) caused significant inhibition (p<0.05) at 0.5, 1, 1.5, 2, 3 and 3.5 h after dosing. In summary, C. striatus and C. lucius extracts may possess antinociceptive and antipyretic in mice but at 60 mg/kg, both fish extracts did not produce anti-inflammatory activity in rabbits.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

CIRI-CIRI ANTI-KERADANGAN, ANTINOSISEPTIF DAN ANTIPIRETIK DARIPADA EKSTRAK IKAN YANG KEPALANYA MENYERUPAI ULAR (Channa striatus DAN C. lucius)

Oleh

AZLINA BINTI MOHD. JOHARI

Mac 2008

Pengerusi : Profesor Madya Arifah Abdul Kadir, PhD

Fakulti : Perubatan Veterinar

Channa spp. adalah ikan yang kepalanya menyerupai ular dan secara amnya dimakan oleh masyarakat Malaysia. Satu kajian telah dijalankan untuk menentukan kesan ekstrak Channa striatus dan Channa lucius terhadap keradangan sinovial akibat karraginan pada arnab. Ciri-ciri antinosiseptif dan antipiretik daripada kedua-dua Channa spp. juga dikaji pada mencit. Channa striatus disediakan melalui pengekstrakan air manakala C. lucius melalui pengekstrakan bahagian akues menggunakan kloroform: metanol. Enam belas ekor arnab dibahagikan secara rawak kepada empat buah kumpulan. Setiap kumpulan arnab diberikan rawatan secara oral, iaitu C. striatus (60 mg/kg), C. lucius (60 mg/kg), ketoprofen (3 mg/kg) dan larutan saline (kawalan), masingmasing, 30 minit sebelum induksi keradangan dilakukan. Lutut kanan telah disuntik dengan 1% karraginan sebanyak 0.5 mL secara intra-rawan. Darah telah diambil untuk esei thromboxane B₂ (TxB₂) sebelum dan pada 1, 3, 6, 9, 12 dan 24 jam selepas induksi keradangan. Cecair sinovial dan membran sinovial



telah diambil ketika bedah siasat untuk analisis dan histopatologi. Keputusan menunjukkan penghasilan serum TxB_2 yang signifikan (p<0.05) telah direncat selama 12 jam selepas ketoprofen diberikan secara oral kepada arnab. Perencatan penghasilan serum TxB₂ daripada C. striatus pula adalah lebih rendah daripada kumpulan kawalan pada jam 1, 3, 6 dan 12. Untuk C. lucius pula, perencatan penghasilan TxB_2 adalah pada jam 9 dan 12. Walaupun begitu, perencatan TxB₂ yang dihasilkan oleh kedua-dua Channa spp. adalah kecil dan tidak signifikan jika dibandingkan dengan kumpulan kawalan. Untuk kumpulan rawatan C. striatus dan ketoprofen, jumlah kiraan sel darah putih adalah lebih rendah berbanding dengan kumpulan kawalan tetapi tidak Keputusan histopatologi bagi kumpulan rawatan yang telah signifikan. diberikan ketoprofen menunjukkan sedikit penyebaran leukosit di dalam membrane sinovial. Walaupun begitu, pada kumpulan kawalan, C. striatus dan C. lucius dapat diperhatikan penghasilan leukosit yang banyak, kesesakan oleh sel-sel darah pada saluran darah dan fibrin. Untuk aktiviti analgesik yang dihasilkan oleh Channa spp., dua puluh empat ekor mencit diagihkan sama rata kepada tiga kumpulan rawatan dan satu kumpulan kawalan. Channa striatus (60mg/kg), C. lucius (60mg/kg) dan ketoprofen (1 mg/kg) telah disuntik secara intra-peritoneal, 30 minit sebelum disuntik dengan asid asetik. Kedua-dua jenis Channa spp. tempatan dan ketoprofen menunjukkan penurunan yang signifikan (p<0.05) kepada jumlah pencerutan abdomen dan keregangan dikedua-dua belah kaki belakang jika dibandingkan dengan kumpulan kawalan. Untuk aktiviti antipiretik yang dihasilkan oleh Channa spp., dua puluh empat ekor mencit telah dibahagikan kepada tiga kumpulan rawatan dan satu kumpulan kawalan.



Mencit telah disuntik dengan 30% (w/v) larutan ragi sebanyak 10 mL/kg subkutis. Suhu telah direkodkan 18 jam sebelum dan setiap setengah jam selama lima jam selepas rawatan diberikan. *Channa striatus* (60 mg/kg) dan *C. lucius* (60 mg/kg) masing-masing telah dapat menurunkan suhu yang tinggi, secara signifikan (p<0.05) pada jam ke 2.5 hingga jam ke 5 dan jam ke 3 hingga jam ke 4. Ketoprofen (1 mg/kg) menyebabkan perencatan secara signifikan (p<0.05) pada jam 0.5, 1, 1.5, 2, 3 dan jam ke 3.5 selepas diberikan rawatan. Kesimpulannya, *C. striatus* dan *C. lucius* ekstrak telah menunjukkan ciri-ciri antinosiseptif dan antipiretik ke atas mencit tetapi pada 60mg/kg, kedua-dua jenis ikan tidak menunjukkan ciri-ciri anti-keradangan ke atas arnab.



ACKNOWLEDGEMENTS

In The Name of ALLAH, The Most Benevolent and Most Merciful

I would like to express my appreciation and sincere gratitude to my supervisor Associate Professor Dr. Arifah Abdul Kadir who gave her great invaluable advice, suggestion, encouragement, support and comment which enable me to complete this research.

My utmost gratitude express to other supervisory committee members, Associate Professor Dr. Muhammad Nazrul Hakim Abdullah and Professor Dr. Mohd. Zamri Saad for their guidance, encouragement and support to finish my research.

I would like to thank Professor Dr. Zaharah Abd.Rahman, Professor Dr. Rasedee Abdullah, Associate Professor Dr. Rosnina Hj. Yusoff and Associate Professor Dr. Mohd Roslan Sulaiman for allowing me to use the equipments in their laboratory, Puan Sairah, Dr. Hazilawati, Puan Hasiah, En. Hafandi, Dr. Dahlia and Puan Suhaila for their moral support and invaluable suggestions in preparing this thesis.

My sincere gratitude express to Puan. Zabedah Tumirin, En. Yap Keng Chee, En. Johari Ripin, En. Kufli Che Nor, En. Mohd. Halmi Othman, Puan Siti Muskinah, Puan Sapiah Jalal, Puan Zainab Nasri and Puan Rosmawati



Hanipah, for their willingness to provide information assistance, equipments and guidance throughout the period of the research.

I also want to thank my friends especially Solihah Hassan, Zetty Nadia, Dr. Shahida Ahmad, Cheong, Yunus Adam, Siti Sarah Udzir and Ahmad Shaiffuddin Abd Rahman for their kindness, assistance and co-operation during laboratory work.

Lastly, I wish to express my gratefulness to my beloved husband Haji Ahmad Faizal, my daughter and son, Aniqatul 'Iffah and Ahmad Muzakkir, my mother (Hajah Zun Khadijah), my brothers and sisters (Haji A'zlan Shah and Azrin Shah, Azian and Azimah) for their constant encouragement, inspiration, support and care to me along the study.



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

ARIFAH ABDUL KADIR, PhD

Associate Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Chairman)

MUHAMMAD NAZRUL HAKIM ABDULLAH, PhD

Associate Professor Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Member)

MOHD ZAMRI SAAD, PhD

Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Member)

AINI IDERIS, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date: 14 August 2008



I certify that an Examination Committee has met on 14th March 2008 to conduct the final examination of Azlina Haji Mohd Johari on her Master of Science thesis entitled "Anti-Inflammatory, Antinociceptive and Antipyretic Effects of Extracts from Snakehead Fish (*Channa striatus and C. lucius*" in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The Committee recommends that the student be awarded the Master of Science.

Members of the Examination Committee were as follows:

Mohamed Ali Rajion, PhD

Professor Faculty of Veterinary Medicine Universiti Putra Malaysia (Chairman)

Zuraini Ahmad, PhD

Associate Professor Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Internal Examiner)

Roslida Abd. Hamid @ Abd. Razak, PhD

Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Internal Examiner)

Chung Lip Yong, PhD

Associate Professor Department of Pharmacy Faculty of Medicine Universiti Malaya (External Examiner)

HASANAH MOHD. GHAZALI, PhD

Professor and Deputy Dean School and Graduate Studies Universiti Putra Malaysia

Date:



DECLARATION

I declare that the thesis is my original work except for quotations and citations, which have been duly acknowledge. I also declare that it has not been previously, and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institution.

AZLINA MOHD JOHARI

Date: 13 July 2008



LIST OF TABLES

Та	ble	Page
1.	Fatty acid composition of two Channa spp. fish.	11
2.	Amino acid composition of Channa spp. fish.	12
3.	Lesion scores for microscopic changes in synovial membrane.	49
4.	Tracer dilution test for TxB ₂ radioimmunoassay.	54
5.	Antiserum dilution test for TxB ₂ radioimmunoassay.	54
6.	Radioimmunoassay parameters for reproducibility.	56
7.	Inter-assay and intra-assay variations of TxB_2 radioimmunoassay.	56
8.	Mean values for WBC differential count from the synovial fluid in rabbits treated with <i>Channa spp.</i> extracts.	64
9.	Results of lesion scoring in the synovial membrane in carrageenan induced-synovitis in rabbits treated with <i>Channa spp.</i> extracts.	66
10	. The antinociceptive activity of <i>C. striatus</i> and <i>C. lucius</i> extracts on acetic acid- induced writhing test in mice.	70
11	. Effect of Water Extract of <i>Channa striatus</i> and Aqueous Portion of Chloroform: methanol extract of <i>Channa lucius</i> extracts on Brewer's yeast-induced hyperthermia in mice.	74



LIST OF FIGURES

Figure		
rigure	P	Page
1.	Arachidonic acid metabolism via cyclooxygenase pathway.	16
2.	Cross-section of a cyclooxygenase enzyme in the lumen of the endoplasmic reticulum. COX are bifunctional enzymes with two different catalytic activities which are cyclooxygenase activity at cyclooxygenase catalytic sites (Cx) and peroxidase activity at peroxidase catalytic site (Px). Mb is the membrane binding part that attach to the endoplasmic recticulum.	19
3.	Competing reactions that form the basis of radioimmunoassay (RIA).	21
4.	Procedure of water extraction of Channa striatus.	39
5.	Procedure of aqueous portion of chloroform: methanol extraction of <i>Channa lucius</i> .	40
6.	Summary of the synovitis model	50
7.	A standard TxB ₂ curved plotted using 'four parameter logistic model'.	55
8.	Effect of control, <i>Channa spp</i> . and ketoprofen on serum TxB_2 synthesis in rabbits.	60
9.	Total WBC count in carrageenan-induced synovitis in rabbits treated with <i>C. striatus</i> and <i>C. lucius</i> extracts.	61
10.	Synovial fluid volume in carrageenan induced-synovitis in rabbits treated with <i>C. striatus</i> and <i>C. lucius</i> extracts.	62
11.	Protein concentration in carrageenan induced-synovitis in rabbits treated with <i>C. striatus</i> and <i>C. lucius</i> extracts.	63



LIST OF ABBREVIATIONS

C°	degree Celcius
AA	arachidonic acid
ANOVA	one-way analysis of variance
B ₀	blank
CNS	central nervous system
COX-1	cyclooxygenase-1
COX-2	cyclooxygenase-2
СРМ	count per minute
DHA	docosahexaenoic acid
DHETs	dihydroxyeicosatrienoic acids
ED ₂₀	dose of a drug required to produce 20% of the drug's maximal effect.
ED ₅₀	dose of a drug required to produce 50% of the drug's maximal effect.
ED ₈₀	dose of a drug required to produce 80% of the drug's maximal effect.
EETs	epoxyeicosatrienoic acids
EP	prostaglandin receptor
EPA	eicosapentaenoic acid
h	hour
HETES	monohydroxy eicosatetraenoic acids



IL-1	interleukin-1
IL-6	interleukin-6
IP	prostacyclin receptor
LPS	lipopolysaccharide
mPGES-1	microsomal PGE synthase-1
NOS	nitric oxide synthase
NSAIDs	non-steroidal anti- inflammatory drugs
NSB	non specific binding
PGD ₂	prostaglandin D ₂
PGE ₂	prostaglandin E ₂
PGF _{2α}	prostaglandin $F_{2\alpha}$
PGG ₂	prostaglandin G ₂
PGH ₂	prostaglandin H ₂
PGI ₂	prostacyclin
РКА	protein kinase A
PKC	protein kinase C
PMN	polymorphonuclear leucocytes
POPOP	[1,4-di-(2-(5- [phenyloxazoly benzene)]
PPO	2,5-diphenyloxazole
PUFA	polyunsaturated fatty acid
QC-H	quality control at high concentration



QC-L	quality control at low concentration
RIA	radioimmunoassay
S.D	standard deviation
S.E.M	standard error mean
тс	total count
TNF	tumour necrosis factor
TxA ₂	thromboxane A ₂
TxB ₂	thromboxane B ₂
v	volume
v/v	volume/volume
VCAM-1	vascular adhesion molecule-1
w	weight
w/v	weight/volume
WBC	white blood cell
β	beta
μΙ	microlitre



LIST OF PLATES

Plate		Page
1.	Channa striatus (haruan).	8
2.	Channa lucius (bujuk).	8
3.	Right stifle joint; normal and non-induced. Normal synovial membrane (S).	66
4.	Left stifle joint-inflamed and swollen synovial membrane (S).	66
5.	Section of normal synovial membrane from non-induced leg of the rabbit.	67
6.	Section of synovial membrane from control rabbit.	67
7.	Section of synovial membrane from C.striatus treated rabbit.	68
8.	Section of synovial membrane from <i>C.lucius</i> treated rabbit.	68
9.	Synovial membrane section from ketoprofen treated rabbit.	69



Appe	Appendix		
A.	Reagent preparations	110	
C1.	Calculation of TxB ₂ Standard Values	113	
Table			
13:	Thromboxane B ₂ standard preparation	111	
14:	Thromboxane B ₂ standard protocol	111	
15:	Typical radioimmunoassay data	114	
16:	Time course of serum TxB ₂ synthesis in control rabbits.	115	
17:	Time course of serum TxB ₂ synthesis in <i>C. striatus</i> treated rabbits.	115	
18:	Time course of serum TxB ₂ synthesis in <i>C. lucius</i> treated rabbits.	115	
19:	Time course of serum TxB_2 synthesis in ketoprofen treated rabbits.	116	
20:	The sequential tissue processing procedure in an automatic tissue processor.	117	
21:	The H&E staining procedures.	118	

LIST OF APPENDICES



TABLE OF CONTENTS

	Page
ABSTRACT	ü
ABSTRAK	iv
ACKNOWLEDGEMENTS	vii
APPROVAL	ix
DECLARATION	xi
LIST OF TABLES	XV
LIST OF FIGURES	xvi
LIST OF PLATES	xvii
LIST OF APPENDICES	xviii
LIST OF ABBREVIATIONS	xix

CHAPTER

1	INTRODUC	TION	1
2	LITERATUR	REREVIEW	5
	2.1	Local Channa species	5
	2.2	Description of Channa spp.	6
		2.2.1 Channa striatus	6
		2.2.2 Channa lucius	6
	2.3	Constituents of Channa striatus and Channa lucius	9
	2.4	Properties of Channa striatus and Channa lucius	13
	2.5	Inflammation	15
		2.5.1 The Inflammatory Process	15
		2.5.2 Inflammatory Mediators (Eicosanoids)	15
		2.5.3 Synovitis	18
	2.6	Pain	26
		2.6.1 Nociceptors	26
		2.6.2 Prostaglandins and Pain Sensitization	27
		2.6.3 Peripheral Mechanisms	27
		2.6.4 Central Mechanisms	29
	2.7	Fever	29
		2.7.1 Pathogenesis of Fever	30
		2.7.2 Mechanisms of Fever	32
	2.8	Non-steroidal Anti-inflammatory Drugs (NSAIDs)	34
		2.8.1 Ketoprofen	34
3	MATERIAL	S AND METHODS	36
	3.1	Chemicals and Drugs	36
	3.2	Apparatus	36
	3.3	Preparation of Fish Extracts	36
		3.3.1 Water Extract of <i>C. striatus</i>	37
		3.3.2 Aqueous Portion of Chloroform: Methanol Extract (C. Jucius)	38
	3.4	Experimental Animals	41



	3.4.1 Mice	41
o =	3.4.2 Rabbits	41
3.5	Validation of Thromboxane B ₂ Radioimmunoassay	41
	3.5.1 Standard	42
	3.5.2 Antiserum	42
	3.5.3 Iracer	42
	3.5.4 Butter	42
	3.5.5 Scintillation Cocktail	43
	3.5.6 Tracer Dilution Test	43
	3.5.7 Antiserum Dilution Test	43
	3.5.8 Standard Assay Procedure	44
	3.5.9 Analysis of Data	44
	3.5.10 Characterization of the Assay	45
3.6	Carrageenan-induced Synovitis in Rabbits	46
	3.6.1 Animals and Treatment	46
	3.6.2 Preparation of 1% Carrageenan Solution	46
	3.6.3 Saline Solution (0.9%)	46
	3.6.4. Acute Synovitis Model	47
	3.6.5 Serum Collection	47
	3.6.6 Synovial Fluid Collection and Analysis	48
-	3.6.7 Histopathology	48
3.7	Analgesic Activity	51
	(Abdominal Constriction Test)	
3.8	Antipyretic Activity	52
	(Brewer's Yeast Induced Pyrexia)	
3.9	Statistical analysis	52
RESULTS		53
4.1	Validation and Characterization of Thromboxane B ₂	53
4.0	Assay	
4.2	Entropy on Correspondence induced Sympositics in Debbits	57
	A 2.1 Decreased upperiod of the Water Extract	57
	4.2.1 Pharmacouynamics of the Water Extract	57
	Of C. Striatus and the Aqueous Pontion of	
	After Oral Administration	
	Aner Oral Administration	E0
	4.2.2 Synovial Fluid Parameters	00 50
	4.2.3 GIUSS NECIOPSY FINDINGS	20
	4.2.4 Enstopathology Findings	59
4.0	4.2.3 Lesion Sconny	59 70
4.3	Anunociceptive Activity of Channa Striatus and	70
A A	Channa Lucius III Milee	70
4.4	Channa Lucius In Mico	12

4



5	DISC 5.1 5.2 5.3 5.4	USSION Validation and Characterization of Thromboxane B ₂ Assay The Effect of <i>Channa Striatus</i> and <i>Channa Lucius</i> Extract on Carrageenan-induced Synovitis In Rabbits Antinociceptive Activity of <i>Channa Striatus</i> and <i>Channa Lucius</i> In Mice Antipyretic Activity of <i>Channa Striatus</i> and <i>Channa Lucius</i> In Mice	75 75 78 83 89
6	SUMI FUTU	MARY, CONCLUSION AND RECOMMENDATIONS FOR IRE RESEARCH	92
BIBLIOGRAPHY APPENDICES BIODATA OF THE STUDENT			96 110 120



CHAPTER 1

INTRODUCTION

Inflammatory diseases which are associated with pain such as arthritis are among the most common health problem in the western countries (Horrocks and Yeo, 1999). In Malaysia, two million people have problems associated with synovitis and later it developed into arthritis, where 600,000 people are more than 60 years old (Najibah, 2002). Inflammation is part of the body's immediate response to infection or injury. It is represented by redness, swelling, heat and pain (Galbraith *et al.*, 2001). Inflammation can also contribute to the rise of temperature in the body and produce other symptoms such as weakness, unable to concentrate and decrease activity (Crocetti *et al.*, 2002). Furthermore, high fever can be harmful to the body and may trigger brain disturbance such as convulsion in infants (Griffith, 2006).

Acute synovial inflammation include increasing of synovial fluid with distension of synovial capsule, increasing skin temperature over the joint and hyperplasia of the synovial membrane. During synovitis, permeability of the synovial membrane increases and proteins accumulates in the joint. Eicosanoids and other inflammatory mediators contribute to the inflammatory response by increasing the permeability of synovial vessels and increasing the movement of leukocytes from the blood stream into the surrounding tissues (Owens *et al.*, 1996). The earliest cells appearing at inflamed sites are granulocytes, with



1

monocytes/macrophages and lymphocytes appearing later. Granulocytes and monocytes/macrophages are involved in pathogen killing, clearing up cellular and tissue debris and tissue repair (Calder, 2006).

Sensory and emotional experiences associated with injury or inflammation are known as pain (Almeida *et al.*, 2004). Pain can be divided into acute and chronic pain. Acute pain ceases when the injury recovered. As for chronic pain, it occurs for a longer time compared to acute pain and may cause mental depression and decrease motor activity (Shipton, 1999).

As with inflammation, there could be an increase in the body temperature. This is due to the production of cytokines such as interleukins, tumor necrosis factor and others (Werner *et al.*, 2006). These cytokines will trigger the production of prostaglandin E_2 in the thermoregulatory centre at the anterior of the hypothalamus to produce fever (Sehic *et al.*, 1996).

Nonsteroidal anti-inflammatory drugs (NSAIDs) are widely used for treatment of inflammation, pain and fever. The main mechanism of action of these drugs is believed to be the inhibition of the cyclooxygenase enzymes and leads to the conversion of arachidonic acid to prostaglandins (Lee *et al.*, 2003). Even though some of the drugs have a good effect in reducing the symptoms of the diseases, they have various side effects such as gastrointestinal ulcer, bleeding and renal damage (Coppelli *et al.*, 2004). This leads to a major interest from the



2