

# ANTIOXIDANTS FROM MALAYSIAN AGARWOOD (AQUILARIA SPP.) LEAF EXTRACTS AND THEIR APPLICATIONS IN IN VITRO MODEL AND FOOD SYSTEM

**TAY PEI YIN** 

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### ANTIOXIDANTS FROM MALAYSIAN AGARWOOD (*AQUILARIA* SPP.) LEAVES EXTRACTS AND THEIR APPLICATIONS IN *IN VITRO* MODEL AND FOOD SYSTEM

By

TAY PEI YIN

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of Philosophy

August 2015

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

### ANTIOXIDANTS FROM MALAYSIAN AGARWOOD (AQUILARIA SPP.) LEAVES EXTRACTS AND THEIR APPLICATIONS IN IN VITRO MODEL AND FOOD SYSTEM

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#### TAY PEI YIN

August 2015

Chair : Tan Chin Ping, PhD Faculty: Food Science and Technology

Agriculture by-products are undervalued substrates that are normally removed and disposed from the food production line. In this study, young leaves from three commercial species of Malaysian agarwood (Aquilaria malaccensis, Aquilaria subintegra and Aquilaria crassna) were examined. These are by-products from agarwood plantations and are usually discarded during the cultivation of agarwood trees. The first part of the present study assessed the effects of the ethanol concentration (0-100%) (v/v), solid-to-solvent ratio (1:10-1:60) (w/v) and extraction time (30-180 min) on the extraction of polyphenols from young agarwood leaves. The total phenolic content (TPC), total flavonoid content (TFC) and DPPH radical-scavenging capacity were used for the quantification of polyphenols, and assessment of antioxidant capacity. The appropriate ranges of the extraction parameters were determined through single-factor experiments, and the optimal conditions for polyphenol extraction were established through response surface methodology (RSM). Among the three species of young agarwood leaves, the extract from A. subintegra young leaves (ASE) obtained through extraction with 64.64% (v/v) ethanol and a solid-to-solvent ratio of 1:70 (w/v) for 120 min was selected for the second part of the study. The antioxidative interactions (synergy, additive and antagonism) between ethanolic ASE and  $\alpha$ -tocopherol were evaluated through assays of the DPPH radical-scavenging activity, the  $\beta$ -carotene bleaching system and liposome peroxidation. Among all of the combinations of ASE and  $\alpha$ -tocopherol, only fraction 1/3 showed a synergistic interaction, which agrees well with the results from three different assays. In the third part of this study, ASE was incorporated into soybean oil-based mayonnaise at a concentration of 1/3 relative to the tocopherol content to prevent its deterioration during storage. The peroxide value (POV), total oxidation value (TOTOX) and conjugated trienes (CT) of ASE-enriched mayonnaise were significantly (p < 0.05) lower than the negative control mayonnaise samples at the end of the storage period. The addition of ASE to mayonnaise presented comparable ability to protect against deterioration compared with the addition of the commercially available synthetic ethylenediaminetetraacetic acid (EDTA). ASE was further tested for its toxicity via brine shrimp lethality and hemolytic analyses. Low brine shrimp lethality and low



hemolytic activity with IC<sub>50</sub> values of 96.6 µg/mL and 1864.7 µg/mL against brine shrimp and human erythrocytes, respectively, were observed. In addition, the major bioactive compound in ASE, namely iriflophenone 3-C- $\beta$ -glucoside (4.1%, w/w) with an IC<sub>50</sub> of 54.5 mg/L for scavenging DPPH radicals was identified and quantified based on 1D NMR, 2D-NMR, LC-MS and UV-vis spectral data. In summary, this study may serve as a reference for future natural polyphenol applications in the food industry.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

### MENOROKAI ANTIOKSIDAN DARIPADA PRODUK SAMPINGAN TIGA SPESIS KOMERSIL GAHARU MALAYSIA DAN APLIKASI MEREKA DI DALAM MODEL IN VITRO DAN SISTEM MAKANAN

Oleh

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Pengerusi: Profesor Tan Chin Ping, PhD Fakulti : Sains dan Teknologi Makanan

Produk sampingan pertanian adalah substrat bernilai rendah yang biasanya dikeluarkan dan dibuang daripada proses pengeluaran makanan. Dalam kajian ini, daun muda daripada tiga spesis komersial gaharu Malaysia (Aquilaria malaccensis, Aquilaria subintegra and Aquilaria crassna) telah dikaji. Ia adalah produk sampingan daripada ladang gaharu dan biasanya dibuang semasa penanaman pokok gaharu. Bahagian pertama kajian ini adalah mengenai pengaruh kepekatan etanol (0-100%) (v/v), nisbah pepejal-pelarut (1:10-1:60) (w/v) dan masa pengekstrakan (30-180 min) kepada pengekstrakan polifenol daripada daun muda gaharu. Hasil fenolik (jumlah kandungan fenolik; TPC dan jumlah kandugan flavonoid; TFC) telah digunakan untuk penentuan dan kuantifikasi polifenol, dan kapasiti pengikatan radikal 2,2-diphenyl-1-picrilhydrazil (DPPH) telah dinilai untuk menetukan kapasiti antioksidan. Julat parameter pengekstrakan yang sesuai telah ditentukan dengan menggunakan eksperimen-faktor tunggal dan keadaan pengekstrakan yang optimum untuk pengekstrakan polifenol telah diperolehi dengan cara pengoptimuman melalui metodologi respon permukaan (RSM). Antara tiga spesis daun muda gaharu, ekstrak daripada daun muda A. subintegra (ASE) yang diperolehi dengan menggunakan 64.64% (v/v) etanol, 1:70 (w/v) pepejal-pelarut nisbah dan 120 min masa pengekstrakan telah dipilih untuk bahagian kedua kajian. Interaksi (sinergim tambahan dan antagonistik) antara ASE dan α-tokoferol telah dinilai dengan menggunakan menggunakan kapasiti pengikatan radikal DPPH, sistem pelunturan  $\beta$ -karotena dan peroksidaan liposom. Di antara semua kombinasi ASE dan  $\alpha$ tokoferol, hanya pecahan 1/3 menunjukkan interaksi sinergim yang sesuai bagi tiga kaedah yang berbeza. Dalam bahagian ketiga kajian ini, ASE telah dimasukkan ke dalam mayones berasaskan minyak soya pada kepekatan 1/3 berbanding dengan kandungan tokoferol dalam minyak soya untuk mengelakkan kerosakan semasa penyimpanan. POV, TOTOX dan CT dari mayonis disediakan dengan menggunakan ASE lebih rendah daripada sampel kawalan negatif pada akhir tempoh penyimpanan. Mayones yang disediakan dengan menggunakan ASE mempunyai kemampuan yang standing dengan asid atelindiamintetrasetik (EDTA) sintetik dalam melindungi mayonis daripada kemerosotan. Seterusnya ketoksikan ASE telah diuji melalui analisis kadar kemautan udang brin dan hemolitik. Kadar kemautan udang brin dan aktiviti hemolitik yang rendah dengan nilai IC<sub>50</sub> sebanyak 96.6  $\mu$ g/mL dan 1864.7  $\mu$ g/mL terhadap udang brin dan eritrosit manusia telah diperhatikan. Akhirnya, sebatian bioaktif yang utama dalam ASE telah dipencilkan dan diperolehi dengan menggunakan kromatografi turus (CC) and kromatografi lapisan nipis (TLC). Iriflophenone 3-C- $\beta$ -glucoside telah dikenal pasti berdasarkan resonans magnet nukleus dan spektrometri jisim. Ia telah dikira sebagai polifenol utama (4.1%, w/w) di dalam ASE dengan nilai IC<sub>50</sub> sebanyak 54.5 mg/L dalam pengikatan radikal DPPH. Kesimpulannya, kajian ini boleh digunakan sebagai rujukan dalam aplikasi ekstrak polifenol semulajadi daripada gaharu dalam industri makanan.



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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

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3.2	$Y = \beta_0 + \sum_{i=1}^{k} \beta_i X_i + \sum_{i=1}^{k} \beta_{ii} X_i^2 + \sum_{i=1}^{k} \sum_{i=1}^{k-1} \beta_{ij} X_i X_j$	44
4.1	BCB inhibition capacity (%) = $[(DR_c - DR_s)/DR_c] \times 100\%$	73
4.2	$DR_c \text{ or } DR_s = \ln (a/b)/120)$	73
4.3	$ROO \bullet + \alpha$ -tocopherol $\rightarrow ROOH + TO \bullet$	79
5.1	FIC activity (%) = $[1 - (A_s/A_c)] \times 100\%$	83
5.2	Mortality (%) = $(N_d/N_t) \times 100\%$	86
5.3	Hemolysis (%) = $[(A_s - A_{nc})/(A_{pc} - A_{nc})] \times 100\%$	87

# LIST OF ABBREVIATIONS

Α	
a*	redness
AAPH	2,2' azobis (2-aminopropane) dihydrochloride
ABTS	2,2'-azinobis-(3-ethyl-benzothiazoline-6-sulphonic acid)
A. crassna	Aquilaria crassna
ACE	Aquilaria crassna extract
A. malaccensis	Aquilaria malacensis
AME	Aquilaria malacensis extract
ANOVA	analysis of variance
AR	analytical reagent
ArO•	aroxyl radical
ArOH	antioxidant
ASE	Aquilaria subintegra extract
AOCS	american oil chemists' society
APCI	atmospheric pressure chemical ionization
111 01	uniospherie pressure enemieur tomzuron
В	
b*	yellowness
BCB	β-carotene bleaching
BHA	butylated hydroxyanisole
BHT	butylated hydroxytoluene
2	outjined hjerohjtohene
С	
<sup>13</sup> C	carbon-13
CAT	catalase
CCC	conventional column chromatography
CCD	central composite design
CD	conjugated dienes
CD <sub>3</sub> OD	deuterated methanol
COSY	correlation spectroscopy
CT	conjugated trienes
CTC	condensed tannins content
CV	coefficient variation
0,	
D	
ā	doublet
dd	doublet doublet
1D	one dimensional
2D	two dimensional
DAD	diode array detector
DMSO	dimethyl sulfoxide
DPPH	2,2 <sup>'</sup> -diphenyl-1-picrylhydrazyl
DPPH-H	1,1-diphenyl-2-picryl hydrazine
DW	dry weight

G

E E° EDTA ESI Expt.	reduction potential ethylenediaminetetra-acetic acid electrospray ion source experimental data
F FAME FC FIC FID FRAP FTIR <i>F</i> -value	fatty acid methyl esters folin-Ciocalteu ferrous iron-chelating flame ionization detector ferric ion reducing antioxidant power fourier transform infrared statistical significance
G GAE GC GC-MS GPx GR GRAS GSH	gallic acid equivalent gas chromatography gas chromatography mass spectrometry glutathione peroxidase glutathione reductase generally recognized as safe glutathione
H $^{1}$ H H <sub>2</sub> O H <sub>2</sub> SO <sub>4</sub> HAT HMBC HO• H <sub>2</sub> O <sub>2</sub> HO <sub>2</sub> • <sup>-</sup> HPLC HSQC	proton water sulphuric acid hydrogen atom transfer heteronuclear multiple bond correlation hydroxyl radical hydrogen peroxide perhydroxyl radical high performance liquid chromatography heteronuclear single quantum coherence
I IC <sub>50</sub> J	50% inhibitory concentration
J L LC LC 50 LC-MS LDL	coupling constant lightness liquid chromatography 50% lethality concentration liquid chromatography mass spectrometry low density lipoproteins

 $\bigcirc$ 

m MeOH Meq MPOB MS	multiplet methanol milliequivalents of peroxide per kilogram malaysia palm oil board mass spectrometry
MUFA	monounsaturated fatty acids
N NMR NO•	nuclear magnetic resonance nitric oxide
O O₂• <sup>-</sup> -OH ORAC O/W	superoxide anion hydroxyl group oxygen radical absorbance capacity oil-in-water
P PBS PCA PDA PG PORIM POV PPO Pred. PUFA	phosphate-buffered saline plate count agar photodiode array/ potato dextrose agar propyl gallate palm oil research institute of malaysia peroxide values enzyme polyphenol oxidase predicted value polyunsaturated fatty acid
Q QE	quercetin equivalent
R R• R <sup>2</sup> R <sub>f</sub> RNS RO• ROO• ROO• ROOH ROS RSM	free radical coefficient of determination retention factor reactive nitrogen species alkoxyl radicals peroxyl radical lipid hydroperoxide reactive oxygen species response surface methodology
S SD SDS SET SOD	singlet standard deviation sodium dodecyl sulfate single electron transfer dismutase superoxide
T TAC	total anthocyanin content
	xxiii

6

xxiii

TBA TBHQ TFA TFC TLC TMS TO• TOTOX TPC TRAP TVC	thiobarbituric acid tertbutyl hydroquinone total flavanol assay total flavonoid content thin layer chromatography tetramethylsilane $\alpha$ -tocopheroxyl radical total oxidation values total phenolic content total radical trapping antioxidant parameter total viable counts
U UV-Vis	ultraviolet-visible
$     \begin{array}{c}       X \\       X_1 \\       X_2 \\       X_1^2 \\       X_2^2 \\       X_1 X_2     \end{array} $	linear coefficients linear coefficients quadratic coefficients quadratic coefficients interaction coefficient
$\mathbf{Y}$ $Y_n$	response variables

G

#### **CHAPTER I**

#### **INTRODUCTION**

#### Background

The health deterioration information is currently widespread, and the awareness of health problems is increasing. One of the crucial substances that cause problems to human health is free radicals. Free radicals are essential for human immune system responses and are produced in normal cell metabolism or physiological processes that utilize oxygen (Kalaivani & Mathew, 2010). However, the uncontrolled production of oxygen-derived free radicals leads to diseases, such as cancer, rheumatoid arthritis, cirrhosis and degenerative processes associated with aging (Tachakittirungrod, Ikegami, & Okonogi, 2007). Therefore, free radicals have been justified as harmful when they react with important cellular components, such as proteins, DNA and the cell membrane (Mantena, et al., 2008).

Other than health deterioration, free radicals have negative impacts in the food industry, including the induction of lipid oxidation by reacting with food lipids (Chan, Lee, Yap, Wan Aida, & Ho, 2009). Hence, to prolong the storage life of foods and to diminish damage to the human body, synthetic antioxidants, such as butylated hydroxyanisole (BHA) and butylated hydroxytoluene (BHT), are added to fresh or processed foods (Soong & Barlow, 2004). Nevertheless, a decline in the use of synthetic antioxidants has been observed due to their possible toxic and carcinogenic effects (Arabshahi-Delouee & Urooj, 2007). In some countries, such as Japan, Canada and Europe, BHA and tertbutylhydroquinone (TBHQ) are prohibited. Thus, the use of natural antioxidants that not only are low in toxicity but also possess pronounced antioxidant activity is encouraged in this health-conscious society.

Today, there are increased lines of evidence suggesting that phytochemicals from natural plant sources possess antioxidant properties associated with a lower risk of mortality from many diseases (Dixon, Xie, & Sharma, 2005; Rice-Evans, 2004). The first marketed natural antioxidant was rosemary extract, which was found to have antioxidant activity as effective as that of synthetic antioxidants (Martínez, Penci, Ixtaina, Ribotta, & Maestri, 2013). In addition, bamboo leaf extracts have been approved as a natural food additive by the Ministry of Health in China (Zhang, Jiao, Liu, Wu, & Zhang, 2008). The antioxidant properties of plant extracts are mainly attributed to the presence of polyphenols. Polyphenols of plants are hydroxylated derivatives of benzoic acid and cinnamic acids and have been reported to exert antioxidative and anticarcinogenic effects (Muanda, Kone, Dicko, Soulimani, & Younos, 2011).

Agarwood was recently categorized as one of the seven types of herbs that obtained approval from Terengganu to be developed into a high-value herbal plantation on 461 hectares of land in Pasir Raja, Terengganu (ETP 2011). In the last ten years, 1000 hectares of agarwood have been planted in Malaysia, as reported by Barden, Noordainie, Mullihen, and Song (2000). Malaysia is the third largest exporter in the world after India and Indonesia, and more than 200 products that contain *Aquilaria* are currently registered with the Ministry of Health (Lim & Noorainie, 2010). As a consequence, the widely available agarwood assures the availability of easy accessible and sustainable sources of its by-product, namely young leaves. Agarwood leaves were prepared as tea and used traditionally for the treatment of trauma-related illness. It has also been reported to have analgesic, anti-inflammatory, laxative and anti-diabetics activities (Zhou, Wang, Suolangjiba, Kou, & Yu, 2008). As a result, a comprehensive exploration of the young agarwood leaf would be an interesting research topic.

Solvent extraction is a common and popular approach used to obtain desired nutrients and nutraceuticals from plant materials (Gupta, Naraniwal, & Kothari, 2012). However, the extraction of bioactive compounds is complicated by the complexity of bioactive compounds and plant matrices. An effective solvent extraction system is a good way to achieve cost efficiency and economic feasibility in an industrial process. Various novel extraction techniques and parameters have been developed; however, there is no universal standardized optimal extraction condition for achieving maximal yields of antioxidant compounds and activity in different plants. Consequently, a systematic study of the efficiency of solvent extraction from young agarwood leaves to obtain polyphenols with high antioxidant activity at a reduced production cost is essential.

The application of natural antioxidants has not been widely adopted by the food industries due to their undesirable flavor, color, aroma, and, most importantly, higher production cost, which may not be affordable compared with conventional food processing. The addition of exogenous antioxidants from extracts to a food system may interact with each other or with the endogenous antioxidants present in the food, resulting in different overall effects as expected. Therefore, it is of special interest to search for synergistic combinations of natural antioxidants that would enhance the expected pronounced antioxidant effects and eventually reduce the production cost in the food industry. In general, the antioxidants present in plant extracts in various combinations and their interactions are found to be important for their overall effect. The interactions between these bioactive compounds have been categorized as synergistic, additive and antagonistic interactions. Synergism is defined as an interaction that exerts a larger overall effect compared with the effect expected from the simple addition of the effects of the individual bioactive compounds (Uri, 1961). To achieve synergism, the types of compounds, their concentration and the ratios at which they are mixed play important roles (Schwarz, Frankel, & German, 1996). For example, Thoo, et al. (2013) reported that polyphenols work synergistically with  $\alpha$ -tocopherol when mixes at a 2:1 ratio via the regeneration of  $\alpha$ -tocopherol by polyphenols to provide long-lasting tocopherol.

In food processing, including flavor, tex

In food processing, lipid oxidation is the major cause of food quality deterioration, including flavor, texture, aroma and nutritional value (Li, Kim, Li, Lee, & Rhee, 2014). Lipid oxidation is initiated in unsaturated fatty acids and causes the formation of hydroperoxides, which are susceptible to decomposition and results in the formation of

secondary reaction products, such as aldehydes, ketones, acids and alcohols (Kolakowska, 2003). These compounds are responsible for changes in the overall food quality. Mayonnaise is a semi-solid oil-in-water (O/W) emulsion with a high content of polyunsaturated fatty acids (PUFAs), which are very susceptible to oxidation (Alamed, McClements, & Decker, 2006). The oxidation in mayonnaise can be reduced by applying antioxidant agents to retard spoilage, extent shelf life and maintain quality and safety (Devatkal & Naveena, 2010). However, the study and use of natural herb extracts as natural food additives in mayonnaise has been limited. Therefore, the development of natural antioxidant-enriched mayonnaise is encouraged.

Over the last few decades, the methodologies used in the research of natural products have evolved significantly. Older methodologies or strategies, such as chemotaxonomic investigation, focused on the chemistry of compounds instead of their activity, and hence, chemical compounds with no activity were always identified from natural sources (Sarker, Latif, & Gray, 2006). In recent years, the isolation and identification of natural products were mainly based on biological activity testing. It is known that young agarwood leaf is an abundant source of antioxidants, and thus, it was interesting to isolate and identify the major constituents that contribute to its antioxidant activity using a bioassay-guided isolation technique. Moreover, an adequate understanding of the chemical components, chemical natures, solubility and glycosylation positions may provide consistent functional food applications (Ito, et al., 2012).

### **Scope and Objectives**

The first part of present study undertakes a systematic optimization of the solvent extraction of polyphenols from young A. malaccensis, A. subintegra and A. crassna leaves using single-factor experiments and response surface methodology (RSM) to gain a better understanding of the effects of the ethanol concentration, solid-to-solvent ratio and extraction time on the yield of polyphenols and the antioxidant capacity. After obtaining the maximal yields and antioxidant capacities of the extracts, the antioxidant properties of the extracts from three agarwood species were compared, and one species was selected for further study. The second part describes the effects of the interaction between agarwood extracts and  $\alpha$ -tocopherol on the antioxidant properties in various in *vitro* model systems. The combination ratio of extracts and  $\alpha$ -tocopherol that exhibited synergism in all of the model systems was used for the subsequent study described in Chapter V, which aimed to evaluate the capability of the extracts to inhibit lipid oxidation in a mayonnaise food system during eight weeks of storage. The toxicological risks of the extracts were evaluated before the incorporation of the extract into mayonnaise. The last part elucidates the structure of the potent antioxidant(s) in the extracts that were found to contribute to their antioxidant capacity using a detailed bioassay-guided isolation protocol.

The main objectives of this research study were:

- 1. to extract bioactive compounds (polyphenols) from young *A. malaccensis*, *A. subintegra* and *A. crassna* leaves (Chapter III);
- 2. to evaluate the synergism of the isolated antioxidant with α-tocopherol in *in vitro* model systems (Chapter IV);
- 3. to study their toxicological risks and ability to inhibit lipid oxidation in mayonnaise (Chapter V); and
- 4. to identify the major bioactive compound in the extracts (Chapter VI).



characterization of more bioactive compounds with pronounced biological activities present in young agarwood leaf extracts may be useful for their application in the nutraceutical and pharmaceutical industries.

#### REFERENCES

- Abdullah, M. K. B. (2008). Extraction of gaharu essential oil using enzymatic hydrodistillation University Malaysia Pahang, Malaysia.
- Abu-Salem, F. M., & Abou-Arab, A. A. (2008). Chemical, microbiological and sensory evaluation of mayonnaise prepared from ostrich eggs. *Grasas Aceites*, 59, 352-360.
- Abuja, P. M., & Albertini, R. (2001). Methods for monitoring oxidative stress, lipid peroxidation and oxidation resistance of lipoproteins. *Clinica Chimica Acta*, 306, 1-17.
- Adegoke, G. O., Vijay Kumar, M., G., G. K. A., Varadaraj, M. C., Sambaiah, K., & Lokesh, B. R. (1998). Antioxidants and lipid oxidation in foods - a critical appraisal. *Journal of Food Science and Technology*, 35, 283-298.
- Ahuja, S., & Dong, M. W. (2005). *Handbook of pharmaceutical analysis by HPLC*. UK: Elsevier.
- Akter, N., & Neelim, A. Z. (2008). Agarwood plantation at BRaC Tea estate: introduction, environmental factors and financial analysis. In, vol. 2011).
- Al-Farsi, M. A., & Lee, C. Y. (2008). Optimization of phenolics and dietary fibre extraction from date seeds. *Food Chemistry*, 108, 977-985.
- Alamed, J., McClements, D. J., & Decker, E. A. (2006). Influence of heat processing and calcium ions on the ability of EDTA to inhibit lipid oxidation in oil-in-water emulsions containing omega-3 fatty acids. *Food Chemistry*, 95, 585-590.
- Albayrak, S., Aksoy, A., Sagdic, O., & Hamzaoglu, E. (2010). Compositions, antioxidant and antimicrobial activities of Helichrysum (Asteraceae) species collected from Turkey. *Food Chemistry*, 119, 114-122.
- Ali, S. S., Kasoju, N., Luthra, A., Singh, A., Sharanabasava, H., Sahu, A., & Bora, U. (2008). Indian medicinal herbs as sources of antioxidants. *Food Research International*, 41, 1-15.
- Altunkaya, A., Becker, E. M., Gökmen, V., & Skibsted, L. H. (2009). Antioxidant activity of lettuce extract (Lactuca sativa) and synergism with added phenolic antioxidants. *Food Chemistry*, 115, 163-168.

- Alwerdt, J. L., Seigler, D. S., Gonzalez de Mejia, E., Yousef, G. G., & Lila, M. A. (2008). Influence of alternative liquid chromatography techniques on the chemical complexity and bioactivity of isolated proanthocyanidin mixtures. *Journal of Agricultural and Food Chemistry*, 56, 1896-1906.
- Anonymous. (1998). *Quality control methods for medicinal plant materials*. Geneva: World Health Organization.
- Antolovich, M., Prenzler, P. D., Patsalides, E., McDonald, S., & Robards, K. (2002). Methods for testing antioxidant activity. *Analyst*, 127, 183-198.
- AOCS. (2011). Official Methods of Analysis. In *Method Cd 18-90*). Champaign, Illinois, Washington DC: American Oil Chemists' Society.
- Apak, R., Güçlü, K., Demirata, B., Özyürek, M., Çelik, S., Bektaşoğlu, B., Berker, K., & Özyurt, D. (2007). Comparative evaluation of various total antioxidant capacity assays applied to phenolic compounds with the CUPRAC assay. *Molecules*, 12, 1496-1547.
- Arabshahi-Delouee, S., & Urooj, A. (2007). Antioxidant properties of various solvent extracts of mulberry (*Morus indica* L.) leaves. *Food Chemistry*, 102, 1233-1240.
- Arnao, M. B. (2000). Some methodological problems in the determination of antioxidant activity using chromogen radicals: a practical case. *Trends in Food Science & Technology*, 11, 419-421.
- Bajpai, V. K., Sharma, A., Kang, S. C., & Baek, K.-H. (2014). Antioxidant, lipid peroxidation inhibition and free radical scavenging efficacy of a diterpenoid compound sugiol isolated from *Metasequoia glyptostroboides*. Asian Pacific Journal of Tropical Medicine, 7, 9-15.
- Banik, R. M., & Pandey, D. K. (2008). Optimizing conditions for oleanolic acid extraction from Lantana camara roots using response surface methodology. *Industrial Crops and Products*, 27, 241-248.
- Barden, A., Noordainie, A. A., Mullihen, T., & Song, M. (2000). *Heart of the matter: agarwood use and trade and CITIES implementation for Aquilaria malaccensis.* Cambridge: TRAFFIC International
- Barros, L., Ferreira, M.-J., Queirós, B., Ferreira, I. C. F. R., & Baptista, P. (2007). Total phenols, ascorbic acid,  $\beta$ -carotene and lycopene in Portuguese wild edible mushrooms and their antioxidant activities. *Food Chemistry*, *103*, 413-419.
- Baş, D., & Boyacı, İ. H. (2007). Modeling and optimization I: usability of response surface methodology. *Journal of Food Engineering*, 78, 836-845.
- Becker, E. M., Nissen, L., & Skibsted, L. (2004). Antioxidant evaluation protocols: food quality or health effects. *European Food Research and Technology*, 219, 561-571.

- Becker, E. M., Ntouma, G., & Skibsted, L. H. (2007). Synergism and antagonism between quercetin and other chain-breaking antioxidants in lipid systems of increasing structural organisation. *Food Chemistry*, 103, 1288-1296.
- Benson, E. E. (1990). *Free radical damage in stored plant germplasm*. Rome: International Board for Plant Genetic Resources.
- Berdahl, D. R., Nahas, R. I., & Barren, J. P. (2010). Synthetic and natural antioxidant additives in food stabilization: current applications and future research. In E. A. Decker, R. J. Elias & D. J. McClements (Eds.), Oxidation in foods and beverages and antioxidant applications volume 1 understanding mechanisms of oxidation and antioxidant activity, (pp. 273-300). Cambirdge: Woodhead Publishing Limited.
- Berezkin, V. G., & Buzaev, V. V. (1997). New thin-layer chromatography plate with a closed sorbent layer and details of its application. *Journal of Chromatography A*, 758, 125-134.
- Berthod, A., & Carda-Broch, S. (2004). Determination of liquid-liquid partition coefficients by separation methods. *Journal of Chromatography A*, 1037, 3-14.
- Bettaieb Rebey, I., Bourgou, S., Ben Slimen Debez, I., Jabri Karoui, I., Hamrouni Sellami, I., Msaada, K., Limam, F., & Marzouk, B. (2012). Effects of extraction solvents and provenances on phenolic contents and antioxidant activities of cumin (*Cuminum cyminum* L.) seeds. *Food and Bioprocess Technology*, 5, 2827-2836.
- Brown, J. E., Khodr, H., Hider, R. C., & Rice-Evans, C. A. (1998). Structural dependence of flavonoid interactions with Cu2+ ions: implications for their antioxidant properties. *Biochemical Journal*, *330* (*Pt 3*), 1173-1178.
- Byrdwell, W. C. (2001). Atmospheric pressure chemical ionization mass spectrometry for analysis of lipids. *Lipids*, *36*, 327-346.
- Cacace, J. E., & Mazza, G. (2003a). Mass transfer process during extraction of phenolic compounds from milled berries. *Journal of Food Engineering*, 59, 379-389.
- Cacace, J. E., & Mazza, G. (2003b). Optimization of Extraction of Anthocyanins from Black Currants with Aqueous Ethanol. *Journal of Food Science*, 68, 240-248.
- Cadenas, E., & Packer, L. (1996). Handbook of antioxidants. New York: Marcel Dekker.
- Cadenas, E., & Packer, L. (2001). *Handbook of antioxidants, revised and expanded* (2nd ed.). New York: Taylor and Francis e-Library.
- Campos, D., Chirinos, R., Barreto, O., Noratto, G., & Pedreschi, R. (2013). Optimized methodology for the simultaneous extraction of glucosinolates, phenolic compounds and antioxidant capacity from maca (*Lepidium meyenii*). *Industrial Crops and Products*, 49, 747-754.

- Canadanovic-Brunet, J. M., Djilas, S. M., Cetkovic, G. S., & Tumbas, V. T. (2005). Freeradical scavenging activity of wormwood (*Artemisia absinthium* L) extracts. *Journal of the Science of Food and Agriculture*, 85, 265-272.
- Carrasco-Pancorbo, A., Neusüß, C., Pelzing, M., Segura-Carretero, A., & Fernández-Gutiérrez, A. (2007). CE- and HPLC-TOF-MS for the characterization of phenolic compounds in olive oil. *Electrophoresis*, 28, 806-821.
- Carvalho, V. M., Nakamura, O. H., & Vieira, J. G. H. (2008). Simultaneous quantitation of seven endogenous C-21 adrenal steroids by liquid chromatography tandem mass spectrometry in human serum. *Journal of Chromatography B*, 872, 154-161.
- Castro, I. A., Moraes Barros, S. B., Lanfer Marquez, U. M., Motizuki, M., & Higashi Sawada, T. C. (2005). Optimization of the antioxidant capacity of a mixture of carotenoids and  $\alpha$ -tocopherol in the development of a nutritional supplement. *Food Research International*, *38*, 861-866.
- Cela, J., Falk, J., & Munné-Bosch, S. (2009). Ethylene signaling may be involved in the regulation of tocopherol biosynthesis in *Arabidopsis thaliana*. *FEBS Letters*, 583, 992-996.
- Chai, L. L., & Armarego, L. F. (2003). *Purification of laboratory chemicals*. UK: Elsevier.
- Chakrabarty, K., Kumar, A., & Menon, V. (1994). *Trade in agarwood*. New Delhi: TRAFFIC India and WWF-India.
- Chan, E. W. C., & Lim, Y. Y. (2006). Antioxidant activity of *Thunbergia Laurifolia* tea. *Journal of Tropical Forest Science*, 18, 130-136.
- Chan, S. W., Lee, C. Y., Yap, C. F., Wan Aida, W. M., & Ho, C. W. (2009). Optimization of extraction condition for phenolic compounds from limau purut (*Citrus hystrix*) peels. *International Food Research Journal*, *16*, 203-213.
- Chang, C. C., Yang, M. H., Wen, H. M., & Chern, J. C. (2002). Estimation of total flavonoid content in propolis by two complementary colorimetric methods. *Journal of Food and Drug Analysis*, 10, 178-182.
- Che Man, Y. B., & Tan, C. P. (1999). Effects of natural and synthetic antioxidants on changes in refined, bleached, and deodorized palm olein during deep-fat frying of potato chips. *Journal of the American Oil Chemists' Society*, *76*, 331-339.
- Chew, K. K., Ng, S. Y., Thoo, Y. Y., Khoo, M. Z., Wan Aida, W. M., & Ho, C. W. (2011). Effect of ethanol concentration, extraction time, and extraction temperature on the recovery of phenolic compounds and antioxidant capacity of *Centella asiatica* extracts. *International Food Research Journal*, 18, 566-573.

- Chirinos, R., Rogez, H., Campos, D., Pedreschi, R., & Larondelle, Y. (2007). Optimization of extraction conditions of antioxidant phenolic compounds from mashua (*Tropaeolum tuberosum* Ruíz & amp; Pavón) tubers. *Separation and Purification Technology*, 55, 217-225.
- Choi, Y., & Lee, J. (2009). Antioxidant and antiproliferative properties of a tocotrienolrich fraction from grape seeds. *Food Chemistry*, 114, 1386-1390.
- Cicco, N., Lanorte, M. T., Paraggio, M., Viggiano, M., & Lattanzio, V. (2009). A reproducible, rapid and inexpensive Folin–Ciocalteu micro-method in determining phenolics of plant methanol extracts. *Microchemical Journal*, 91, 107-110.
- Clemente, T. E., & Cahoon, E. B. (2009). Soybean oil: genetic approaches for modication of functionality and total content. *American Society of Plant Biologists, 151*, 1030-1040.
- Coupland, J. N., & McClements, D. J. (1996). Lipid oxidation in food emulsions. *Trends* in Food Science & Technology, 7, 83-91.
- Crozier, S., Preston, A., Hurst, J., Payne, M., Mann, J., Hainly, L., & Miller, D. (2011). Cacao seeds are a "super fruit": a comparative analysis of various fruit powders and products. *Chemistry Central Journal*, *5*, 5.
- Cullen, K. E. (2009). Encyclopedia of life science. New York: Facts On File.
- Dai, F., Chen, W.-F., & Zhou, B. (2008). Antioxidant synergism of green tea polyphenols with α-tocopherol and L-ascorbic acid in SDS micelles. *Biochimie*, *90*, 1499-1505.
- Dai, J., & Mumper, R. J. (2010). Plant phenolics: extraction, analysis and their antioxidant and anticancer properties. *Molecules*, 15, 7313-7352.
- De Beer, D., Schulze, A., Joubert, E., de Villiers, A., Malherbe, C., & Stander, M. (2012). Food ingredient extracts of *Cyclopia subternata* (Honeybush): variation in phenolic composition and antioxidant capacity. *Molecules*, 17, 14602-14624.
- Delgado, T., Malheiro, R., Pereira, J. A., & Ramalhosa, E. (2010). Hazelnut (*Corylus avellana* L.) kernels as a source of antioxidants and their potential in relation to other nuts. *Industrial Crops and Products*, 32, 621-626.
- Denyse, J. S., & Rodel, D. L. (2008). Smallholder tree growing for rural development and environmental services: lessons from asia. United Kingdom: Springer Science.
- Devatkal, S. K., & Naveena, B. M. (2010). Effect of salt, kinnow and pomegranate fruit by-product powders on color and oxidative stability of raw ground goat meat during refrigerated storage. *Meat Science*, *85*, 306-311.

Devindran, P. (2009). Malaysia want to be Asia's Gaharu trading center. In, vol. 2015).

- Dixon, R. A., Xie, D.-Y., & Sharma, S. B. (2005). Proanthocyanidins a final frontier in flavonoid research? *New Phytologist*, *165*, 9-28.
- Farrell, S. O., & Campbell, M. K. (2012). *Biochemistry*. United States: Cengage Learning.
- Feng, J., Yang, X.-W., & Wang, R.-F. (2011). Bio-assay guided isolation and identification of α-glucosidase inhibitors from the leaves of *Aquilaria sinensis*. *Phytochemistry*, 72, 242-247.
- Fenn, J. B., Mann, M., Meng, C. K., Wong, S. F., & Whitehouse, C. M. (1989). Electrospray ionization for mass spectrometry of large biomolecules. *Science*, 246, 64-71.
- Finney, D. J. (1971). Probit analysis (3rd ed.). Cambridge: Cambridge University Press.
- Fratkin, J. (1994). *Chinese herbal patent formulas: a practical guide*. Colorado: Shyaublications.
- Friebolin, H. (2005). *Basic one and two dimensional NMR spectroscopy* (4th ed.). New york: Wiley-VCH.
- Gani, R., Jiménez-González, C., & Constable, D. J. C. (2005). Method for selection of solvents for promotion of organic reactions. *Computers & Chemical Engineering*, 29, 1661-1676.
- Gardiner, W. P. (1997). *Statistical analysis of methods for chemists: a software-based approach.* Cambridge: The Royal Society of Chemistry.
- Gertz, C., Klostermann, S., & Kochhar, S. P. (2000). Testing and comparing oxidative stability of vegetable oils and fats at frying temperature. *European Journal of Lipid Science and Technology*, 102, 543-551.
- Gođevac, D., Vujisić, L., Mojović, M., Ignjatović, A., Spasojević, I., & Vajs, V. (2008). Evaluation of antioxidant capacity of *Allium ursinum* L. volatile oil and its effect on membrane fluidity. *Food Chemistry*, 107, 1692-1700.
- Gorinstein, S., Jastrzebski, Z., Leontowicz, H., Leontowicz, M., Namiesnik, J., Najman, K., Park, Y.-S., Heo, B.-G., Cho, J.-Y., & Bae, J.-H. (2009). Comparative control of the bioactivity of some frequently consumed vegetables subjected to different processing conditions. *Food Control*, 20, 407-413.
- Graversen, H., Becker, E., Skibsted, L., & Andersen, M. (2008). Antioxidant synergism between fruit juice and α-tocopherol. A comparison between high phenolic black chokeberry (Aronia melanocarpa) and high ascorbic blackcurrant (Ribes nigrum). *European Food Research and Technology*, 226, 737-743.
- Grill, J. M., Ogle, J. W., & Miller, S. A. (2006). An efficient and practical system for the catalytic oxidation of alcohols, aldehydes, and α,β-unsaturated carboxylic acids. *Journal of Organic Chemistry*, *71*, 9291-9296.

- Grinberg, L. N., Newmark, H., Kitrossky, N., Rahamim, E., Chevion, M., & Rachmilewitz, E. A. (1997). Protective effects of tea polyphenols against oxidative damage to red blood cells. *Biochemical Pharmacology*, 54, 973-978.
- Grotewold, E. (2008). *The science of flavonoids*. USA: Springer Science + Business Media.
- Grusak, M. A. (2002). Phytochemicals in plants: genomics-assisted plant improvement for nutritional and health benefits. *Current Opinion in Biotechnology*, *13*, 508-511.
- Gu, L., Wu, T., & Wang, Z. (2009). TLC bioautography-guided isolation of antioxidants from fruit of Perilla frutescens var. acuta. LWT - Food Science and Technology, 42, 131-136.
- Gunawan, S., Ismadji, S., & Ju, Y.-H. (2008). Design and operation of a modified silica gel column chromatography. *Journal of the Chinese Institute of Chemical Engineers*, 39, 625-633.
- Gupta, A., Naraniwal, W., & Kothari, V. (2012). Modern extraction methods for preparation of bioactive plant extracts. *International Journal of Applied and Natural Sciences*, 1, 8-26.
- Guyot, S., Marnet, N., & Drilleau, J.-F. (2000). Thiolysis-HPLC characterization of apple procyanidins covering a large range of polymerization states. *Journal of Agricultural and Food Chemistry*, 49, 14-20.
- Halliwell, B., & Whiteman, M. (2004). Measuring reactive species and oxidative damage in vivo and in cell culture: how should you do it and what do the results mean? *British Journal of Pharmacology*, 142, 231-255.
- Han, R.-M., Li, D.-D., Chen, C.-H., Liang, R., Tian, Y.-X., Zhang, J.-P., & Skibsted, L. H. (2011). Phenol acidity and ease of oxidation in isoflavonoid/β-carotene antioxidant synergism. *Journal of Agricultural and Food Chemistry*, 59, 10367-10372.
- Handique, J. G., & Baruah, J. B. (2002). Polyphenolic compounds: an overview. *Reactive and Functional Polymers*, 52, 163-188.
- Hanrahan, G., & Lu, K. (2006). Application of factorial and response surface methodology in modern experimental design and optimization. *Critical Reviews in Analytical Chemistry*, *36*, 141-151.
- Harrison, L. J., & Cunningham, F. E. (1985). Factors influencing the quality of mayonnaise: a review. *Journal of Food Quality*, 8, 1-20.
- Hashimoto, K., Nakahara, S., Inoue, T., Sumida, Y., Takahashi, M., & Masada, Y. (1985). A new chromone from agarwood and pyrolysis products of chromone derivatives. *Chemical and Pharmaceutical Bulletin*, 33, 5088-5091.

- Hassimotto, N. M. A., Genovese, M. I., & Lajolo, F. M. (2005). Antioxidant activity of dietary fruits, vegetables, and commercial frozen fruit pulps. *Journal of Agricultural and Food Chemistry*, 53, 2928-2935.
- He, G. Q., Xiong, H. P., Chen, Q. H., Ruan, H., Wang, Z. Y., & Traore, L. (2005). Optimization of conditions for supercritical fluid extraction of flavonoids from hops (*Humulus lupulus* L.). *Journal of Zhejiang University SCIENCE B*, 6, 999-1004.
- Herald, T. J., Gadgil, P., & Tilley, M. (2012). High-throughput micro plate assays for screening flavonoid content and DPPH-scavenging activity in sorghum bran and flour. *Journal of the Science of Food and Agriculture*, *92*, 2326-2331.
- Hernández-Hernández, E., Ponce-Alquicira, E., Jaramillo-Flores, M. E., & Guerrero Legarreta, I. (2009). Antioxidant effect rosemary (*Rosmarinus officinalis* L.) and oregano (*Origanum vulgare* L.) extracts on TBARS and colour of model raw pork batters. *Meat Science*, 81, 410-417.
- Herodež, Š. S., Hadolin, M., Škerget, M., & Knez, Ž. (2003). Solvent extraction study of antioxidants from Balm (*Melissa officinalis* L.) leaves. *Food Chemistry*, 80, 275-282.
- Hiramoto, K., Miura, Y., Ohnuki, G., Kato, T., & Kikugawa, K. (2002). Are watersoluble natural antioxidants synergistic in combination with α-Tocopherol ? *Journal of Oleo Science*, *51*, 569-576.
- Hismath, I., Wan Aida, W. M., & Ho, C. W. (2011). Optimization of extraction conditions for phenolic compounds from neem (*Azadirachta indica*) leaves. *International Food Research Journal*, 18, 931-939.
- Hoefler, A. C., & Coggon, P. (1976). Reversed-phase high performance liquid chromatography of tea constituents. *Journal of Chromatography*, 129, 460-463.
- Horn, A. F., Nielsen, N. S., & Jacobsen, C. (2009). Additions of caffeic acid, ascorbyl palmitate or γ-tocopherol to fish oil-enriched energy bars affect lipid oxidation differently. *Food Chemistry*, 112, 412-420.
- Huang, X., Powers, R., Tymiak, A., Espina, R., & Roongta, V. (2008). Introduction to NMR and its application in metabolite structure determination. In D. Zhang, M. Zhu & W. G. Humphreys (Eds.), *Drug metabolism in drug design and development*, (pp. 369-412). Hoboken: John Wiley and Sons, Inc.
- Huda, A. W. N., Munira, M. A. S., Fitrya, S. D., & Salmah, M. (2010). Antioxidant activity of *Aquilaria malaccensis* (thymelaeaceae) leaves. *Pharmacognosy Research*, *1*, 270-273.
- Ibrahim, M., & Jaafar, H. (2013). Abscisic acid induced changes in production of primary and secondary metabolites, photosynthetic capacity, antioxidant capability, antioxidant enzymes and lipoxygenase inhibitory activity of *Orthosiphon stamineus* Benth. *Molecules*, 18, 7957-7976.

- Ishihara, M., Tsuneya, T., & Uneyama, K. (1993). Fragrant sesquiterpenes from agarwood. *Phytochemistry*, 33, 1147-1155.
- Ito, T., Kakino, M., Tazawa, S., Watarai, T., Oyama, M., Maruyama, H., Araki, Y., Hara, H., & Iinuma, M. (2012). Quantification of polyphenols and pharmacological analysis of water and ethanol-based extracts of cultivated agarwood leaves. *Journal of Nutritional Science and Vitaminol (Tokyo)*, 58, 136-142.
- Jacobsen, C. (2010). Oxidation of fish oils and foods enriched with omega-3 polyunsaturated fatty acids sectors. In E. A. Decker, R. J. Elias & D. J. McClements (Eds.), Oxidation in foods and beverages and antioxidant applications Volume 2 management in different industry (pp. 156-163). Cambridge: Woodhead Publishing Limited.
- Jacobsen, C., Adler-Nissen, J., & Meyer, A. S. (1999). Effect of ascorbic acid on iron release from the emulsifier interface and on the oxidative flavor deterioration in fish oil enriched mayonnaise. *Journal of Agricultural and Food Chemistry*, 47, 4917-4926.
- Jacobsen, C., Hartvigsen, K., Lund, P., Adler-Nissen, J., Hølmer, G., & Meyer, A. S. (2000). Oxidation in fish oil enriched mayonnaise: 2. Assessment of the efficacy of different tocopherol antioxidant systems by discriminant partial least squares regression analysis. *European Food Research and Technology*, 210, 242-257.
- Jacobsen, C., Hartvigsen, K., Thomsen, M. K., Hansen, L. F., Lund, P., Skibsted, L. H., Hølmer, G., Adler-Nissen, J., & Meyer, A. S. (2001a). Lipid oxidation in fish oil enriched mayonnaise: calcium disodium ethylenediaminetetraacetate, but not gallic acid, strongly inhibited oxidative deterioration. *Journal of Agricultural and Food Chemistry*, 49, 1009-1019.
- Jacobsen, C., Let, M. B., Nielsen, N. S., & Meyer, A. S. (2008). Antioxidant strategies for preventing oxidative flavour deterioration of foods enriched with n-3 polyunsaturated lipids: a comparative evaluation. *Trends in Food Science & Technology*, 19, 76-93.
- Jacobsen, C., Timm, M., & Meyer, A. S. (2001b). Oxidation in fish oil enriched mayonnaise: ascorbic acid and low pH increase oxidative deterioration. *Journal of Agricultural and Food Chemistry*, 49, 3947-3956.
- Jafar, S. S., Hultin, H. O., Bimbo, A. P., Crowther, J. B., & Barlow, S. M. (1994).
   Stabilization by antioxidants of mayonnaise made from fish oil. *Journal of Food Lipids*, 1, 295-311.
- Jaswir, I., & Che Man, Y. (1999). Use optimization of natural antioxidants in refined, bleached, and deodorized palm olein during repeated deep-fat frying using response surface methodology. *Journal of the American Oil Chemists' Society*, 76, 341-348.

- Jeong, W. S., & Kong, A. N. T. (2004). Biological properties of monomeric and polymeric catechins: green tea catechins and procyanidins. *Pharmaceutical Biology*, 42, 84-93.
- Jiang, B., & Zhang, Z.-W. (2012). Comparison on phenolic compounds and antioxidant properties of cabernet sauvignon and merlot wines from four wine grapegrowing regions in china. *Molecules*, 17, 8804-8821.
- Jimenez-Alvarez, D., Giuffrida, F., Golay, P. A., Cotting, C., Lardeau, A., & Keely, B. J. (2008). Antioxidant activity of oregano, parsley, and olive mill wastewaters in bulk oils and oil-in-water emulsions enriched in fish oil. *Journal of Agricultural and Food Chemistry*, 56, 7151-7159.
- Joglekar, A. M., & May, A. T. (1987). Product excellence through design of experiments. *Cereal Foods World*, 32, 857-868.
- Kadir, A. A., Ng, L. T., & Chang, Y. S. (1997). A review on agar (gaharu) producing Aquilaria species. *Journal of Tropical Forest Products*, 2, 272-285.
- Kalaivani, T., & Mathew, L. (2010). Free radical scavenging activity from leaves of Acacia nilotica (L.) Wild. ex Delile, an Indian medicinal tree. Food and Chemical Toxicology, 48, 298-305.
- Kalaivani, T., Rajasekaran, C., Suthindhiran, K., & Mathew, L. (2011). Free radical scavenging, cytotoxic and hemolytic activities from leaves of *Acacia nilotica* (L.) Wild. ex. Delile subsp. *indica* (Benth.) Brenan. *Evidence-Based Complementary and Alternative Medicine*, 2011.
- Kanazawa, A., Sawa, T., Akaike, T., & Maeda, H. (2002). Dietary lipid peroxidation products and DNA damage in colon carcinogenesis. *European Journal of Lipid Science and Technology*, 104, 439-447.
- Karpińska, M., Borowski, J., & Danowska-Oziewicz, M. (2001). The use of natural antioxidants in ready-to-serve food. *Food Chemistry*, 72, 5-9.
- Karunasekara, T., & Poole, C. F. (2011). Models for liquid–liquid partition in the system formamide–organic solvent and their use for estimating descriptors for organic compounds. *Talanta*, 83, 1118-1125.
- Kazakevich, Y., & LoBrutto, R. (2007). *HPLC for pharmaceutical scientists*. New Jersey: John Wiley & Sons.
- Kchaou, W., Abbès, F., Blecker, C., Attia, H., & Besbes, S. (2013). Effects of extraction solvents on phenolic contents and antioxidant activities of Tunisian date varieties (*Phoenix dactylifera* L.). *Industrial Crops and Products*, 45, 262-269.
- Keokamnerd, T., Acton, J. C., Han, I. Y., & Dawson, P. L. (2008). Effect of commercial rosemary oleoresin preparations on ground chicken thigh meat quality packaged in a high-oxygen atmosphere. *Poultry Science*, 87, 170-179.

- Keshani, S., Luqman Chuah, A., Nourouzi, M. M., Russly, A. R., & Jamilah, B. (2010). Optimization of concentration process on pomelo fruit juice using response surface methodology (RSM). *International Food Research Journal*, 17, 733-742.
- Kochhar, S. P., & Roseel, J. B. (1990). Detection, estimation and evaluatoin of antioxidants in food systems. In B. J. F. Hudson (Ed.), *Food antioxidants*, (pp. 19-64). London: Elsevier Science Publishing LTD.
- Koketsu, M., & Satoh, Y.-I. (1997). Antioxidative activity of green tea polyphenols in edible oils. *Journal of Food Lipids*, 4, 1-9.
- Kokotkiewicz, A., Luczkiewicz, M., Sowinski, P., Glod, D., Gorynski, K., & Bucinski, A. (2012). Isolation and structure elucidation of phenolic compounds from *Cyclopia subternata* Vogel (honeybush) intact plant and *in vitro* cultures. *Food Chemistry*, 133, 1373-1382.
- Kolakowska, A. (2003). Lipid oxidation in food systems. In Z. E. Sikorski & A. Kolakowska (Eds.), *Chemical and functional properties of food lipids*). USA: CRC Press LLC.
- Koleva, I. I., van Beek, T. A., Linssen, J. P. H., Groot, A. d., & Evstatieva, L. N. (2002). Screening of plant extracts for antioxidant activity: a comparative study on three testing methods. *Phytochemical Analysis*, 13, 8-17.
- Kormin, S. B. (2005). The effect of heat processing on triterpene glycosides and antioxidant activity of herbal Pegaga. University Technology Malaysia, Malaysia.
- Kossah, R., Nsabimana, C., Zhang, H., & Chen, W. (2010). Optimization of extraction of polyphenols from Syrian sumac (*Rhus coriaria* L.) and Chinese sumac (*Rhus typhina* L.) fruits. *Research Journal of Phytochemistry*, *4*, 146-153.
- Kubola, J., & Siriamornpun, S. (2008). Phenolic contents and antioxidant activities of bitter gourd (*Momordica charantia* L.) leaf, stem and fruit fraction extracts in vitro. Food Chemistry, 110, 881-890.
- Kumeta, Y., & Ito, M. (2011). Genomic organization of delta-guaiene synthase genes in Aquilaria crassna and its possible use for the identification of Aquilaria species. *Journal of Natural Medicines*, 65, 508-513.
- Kyi, T. M., Daud, W. R. W., Mohammad, A. B., Wahid Samsudin, M., Kadhum, A. A. H., & Talib, M. Z. M. (2005). The kinetics of polyphenol degradation during the drying of Malaysian cocoa beans. *International Journal of Food Science & Technology*, 40, 323-331.
- Kylli, P. (2011). Berry phenolics: isolation, analysis, identification and antioxidant properties., University of Helsinki, Finland.

- Lagunes-Galvez, L., Cuvelier, M.-e., Ordonnaud, C., & Berset, C. (2002). Oxidative stability of some mayonnaise formulations during storage and daylight irradiation. *Journal of Food Lipids*, *9*, 211-224.
- Lampman, G. M., Pavia, D. L., Kriz, G. S., & Vyvyan, J. R. (2010). *Spectroscopy* (4th ed.). Canada: Brooks/ Cole, Cengage Learning.
- Lapornik, B., Prošek, M., & Golc Wondra, A. (2005). Comparison of extracts prepared from plant by-products using different solvents and extraction time. *Journal of Food Engineering*, 71, 214-222.
- Laranjinha, J., & Cadenas, E. (1999). Redox cycles of caffeic acid, alpha-tocopherol, and ascorbate: Implications for protection of low-density lipoproteins against oxidation. *IUBMB Life*, 48, 57-65.
- Laranjinha, J., Vieira, O., amp, x, lia, Madeira, V., amp, x, tor, & Almeida, L. (1995). Two related phenolic antioxidants with opposite effects on vitamin e content in low density lipoproteins oxidized by ferrylmyoglobin: Consumption vs regeneration. Archives of Biochemistry and Biophysics, 323, 373-381.
- Larson, R., & Edwards, B. (2014). *Calculus* (10th ed.). Boston, MA: Brooks/Cole, Cengage Learning.
- Lee, J., Koo, N., & Min, D. B. (2004). Reactive oxygen species, aging, and antioxidative nutraceuticals. *Comprehensive Reviews in Food Science and Food Safety, 3*, 21-33.
- Lee, L.-S., Lee, N., Kim, Y., Lee, C.-H., Hong, S., Jeon, Y.-W., & Kim, Y.-E. (2013). Optimization of ultrasonic extraction of phenolic antioxidants from green tea using response surface methodology. *Molecules*, 18, 13530-13545.
- Leopoldini, M., Russo, N., & Toscano, M. (2011). The molecular basis of working mechanism of natural polyphenolic antioxidants. *Food Chemistry*, 125, 288-306.
- Li, C.-Y., Kim, H.-W., Li, H., Lee, D.-C., & Rhee, H.-I. (2014). Antioxidative effect of purple corn extracts during storage of mayonnaise. *Food Chemistry*, *152*, 592-596.
- Lim, T. W., & Noorainie, A. A. (2010). *Wood for the trees: a review of the agarwood* (gaharu) trade in Malaysia. Malaysia: TRAFFIC Southeast Asia.
- Liu, W., Yu, Y., Yang, R., Wan, C., Xu, B., & Cao, S. (2010). Optimization of total flavonoid compound extraction from *Gynura medica* leaf using response surface methodology and chemical composition analysis. *International Journal of Molecular Sciences*, *11*, 4750-4763.
- Llobera, A. (2012). Study on the antioxidant activity of grape stems (*Vitis vinifera*). A preliminary assessment of crude extracts. *Food and Nutrition Sciences*, *3*, 500-504.

- Luís, Â., Domingues, F., Gil, C., & Duarte, A. P. (2009). Antioxidant activity of extracts of Portuguese shrubs: *Pterospartum tridentatum*, *Cytisus scoparius* and *Erica* spp. *Journal of Medicinal Plants Research*, 3, 886-893.
- Luthria, D. L. (2008). Influence of experimental conditions on the extraction of phenolic compounds from parsley (Petroselinum crispum) flakes using a pressurized liquid extractor. *Food Chemistry*, 107, 745-752.
- Mabberley, D. J. (1997). *The plant book* (2nd ed.). UK: The Press Syndicate of the University of Cambridge.
- Madhujith, T., & Shahidi, F. (2006). Optimization of the extraction of antioxidative constituents of six barley cultivars and their antioxidant properties. *Journal of Agricultural and Food Chemistry*, 54, 8048-8057.
- Maleš, Ž., & Medić-Šarić, M. (2001). Optimization of TLC analysis of flavonoids and phenolic acids of Helleborus atrorubens Waldst. et Kit. *Journal of Pharmaceutical and Biomedical Analysis*, 24, 353-359.
- Man, Y. B. C., Liu, J. L., Jamilah, B., & Rahman, R. A. (1999). Quality changes of refined-bleached-deodorized (rbd) palm olein, soybean oil and their blends during deep-fat frying. *Journal of Food Lipids*, 6, 181-193.
- Mantena, R. K. R., Wijburg, O. L. C., Vindurampulle, C., Bennett-Wood, V. R., Walduck, A., Drummond, G. R., Davies, J. K., Robins-Browne, R. M., & Strugnell, R. A. (2008). Reactive oxygen species are the major antibacterials against Salmonella Typhimurium purine auxotrophs in the phagosome of RAW 264.7 cells. *Cellular Microbiology*, 10, 1058-1073.
- Marshak, D. R. (1996). *Strategies for protein purification and characterization: a laboratory course manual*. USA: Cold Spring Harbor Laboratory Press.
- Marston, A., & Hostettmann, K. (2006). Separation and quantification of flavonoids. In O. M. Anderson & K. R. Markham (Eds.), *Flavonoids: chemistry, biochemistry,* and applications, (pp. 1-36). New York: CRC Press.
- Martínez, M. L., Penci, M. C., Ixtaina, V., Ribotta, P. D., & Maestri, D. (2013). Effect of natural and synthetic antioxidants on the oxidative stability of walnut oil under different storage conditions. *LWT - Food Science and Technology*, *51*, 44-50.
- Mason, R. L., Gunst, R. F., & Hess, J. L. (1989). Statistical design and analysis of experiments with applications to engineering and science. London: Wiley & Sons.
- McClements, D. J., & Decker, E. A. (2000). Lipid oxidation in oil-in-water emulsions: impact of molecular environment on chemical reactions in heterogeneous food systems. *Journal of Food Science*, 65, 1270-1282.

- Michels, M. J. M., & Koning, W. (2000). Mayonnaise, dressings, mustard, mayonnaisebased salads, and acid sauces. In *The microbiological safety and quality of food* (pp. 812). Maryland: Aspen Publishers, Inc.
- Miliauskas, G., Venskutonis, P. R., & van Beek, T. A. (2004). Screening of radical scavenging activity of some medicinal and aromatic plant extracts. *Food Chemistry*, 85, 231-237.
- Mirhosseini, H., Tan, C. P., Hamid, N. S. A., Yusof, S., & Chern, B. H. (2009). Characterization of the influence of main emulsion components on the physicochemical properties of orange beverage emulsion using response surface methodology. *Food Hydrocolloids*, 23, 271-280.
- Montgomery, D. C. (2001). Design and analysis of experiments. New York: Wiley.
- Morrissey, P. A., Sheehy, P. J. A., Galvin, K., Kerry, J. P., & Buckley, D. J. (1998). Lipid stability in meat and meat products. *Meat Science*, 49, Supplement 1, S73-S86.
- Moshi, M. J., Innocent, E., Magadula, J. J., Otieno, D. F., Weisheit, A., Mbabazi, P. K., & Nondo, R. S. (2010). Brine shrimp toxicity of some plants used as traditional medicines in Kagera Region, north western Tanzania. *Tanzania Journal of Health Research*, 12, 63-67.
- Moure, A., Cruz, J. M., Franco, D., Domínguez, J. M., Sineiro, J., Domínguez, H., José Núñez, M. a., & Parajó, J. C. (2001). Natural antioxidants from residual sources. *Food Chemistry*, 72, 145-171.
- MPOB. (2005). A Compendium of Test on Palm Oil Products, Palm Kernel Products, Fatty Acids, Food Related Products and Others. In). Bangi: MPOB.
- Mriziq, K. S., & Guiochon, G. (2008). Column properties and flow profiles of a flat, wide column for high-pressure liquid chromatography. *Journal of Chromatography A*, 1187, 180-187.
- Muanda, F., Kone, D., Dicko, A., Soulimani, R., & Younos, C. (2011). Phytochemical composition and antioxidant capacity of three malian medicinal plant parts. *Evid Based Complement Alternat Med*, 2011, 674320.
- Myers, R. H., & Montgometry, D. C. (2002). *Response surface methodology: process* and product optimisation using designed experiments (2nd ed.). USA: John Wiley & Sons.
- Naczk, M., & Shahidi, F. (2004). Extraction and analysis of phenolics in food. *Journal of Chromatography A*, 1054, 95-111.
- Nagy, K., Courtet-Compondu, M. C., Holst, B., & Kussmann, M. (2007). Comprehensive analysis of vitamin E constituents in human plasma by liquid chromatography-mass spectrometry. *Analytical Chemistry*, 79, 7087-7096.

- Nakamura, Y., Ohto, Y., Murakami, A., & Ohigashi, H. (1998). Superoxide scavenging activity of rosmarinic acid from *Perilla frutescens* Britton Var. *acuta* f. *viridis*. *Journal of Agricultural and Food Chemistry*, 46, 4545-4550.
- Namal Senanayake, S. P. J. (2013). Green tea extract: chemistry, antioxidant properties and food applications a review. *Journal of Functional Foods*, *5*, 1529-1541.
- Nawaz, H., Shi, J., Mittal, G. S., & Kakuda, Y. (2006). Extraction of polyphenols from grape seeds and concentration by ultrafiltration. *Separation and Purification Technology*, 48, 176-181.
- Nekkanti, V., Muniyappan, T., Karatgi, P., Hari, M. S., Marella, S., & Pillai, R. (2009). Spray-drying process optimization for manufacture of drug-cyclodextrin complex powder using design of experiments. *Drug Development and Industrial Pharmacy*, 35, 1219-1229.
- Nelson, R. E., Grebe, S. K., DJ, O. K., & Singh, R. J. (2004). Liquid chromatographytandem mass spectrometry assay for simultaneous measurement of estradiol and estrone in human plasma. *Clinical Chemistry*, 50, 373-384.
- Nielsen, S. S. (2009). Food analysis (4th Edition ed.). USA: Springer.
- Nieto, G., Huvaere, K., & Skibsted, L. (2011). Antioxidant activity of rosemary and thyme by-products and synergism with added antioxidant in a liposome system. *European Food Research and Technology*, 233, 11-18.
- Nikhat, F., Satynarayana, D., & Subhramanyam, E. V. S. (2009). Isolation, characterization and screening of antioxidant activity of the roots of *Syzygium cuminii* (L) Skeel. *Asian Journal of Research in Chemistry*, *2*, 218-221.
- Niki, E. (2010). Assessment of Antioxidant Capacity *in vitro* and *in vivo*. Free Radical Biology and Medicine, 49, 503-515.
- Noh, B. Y., Lee, H. J., Do, J. R., & Kim, H. K. (2014). Antioxidant and ACE inhibitory activity of cultivated and wild *Angelica gigas* Nakai extracts prepared using different extraction conditions. *Preventive Nutrition and Food Science*, 19, 274-280.
- Nomura, T., Kikuchi, M., Kubodera, A., & Kawakami, Y. (1997). Proton-donative antioxidant activity of fucoxanthin with 1,1-diphenyl-2-picrylhydrazyl (DPPH). *Biochemistry and Molecular Biology International Journal*, *42*, 361-370.
- Norshazila, S., Jr., Syed Zahir, I., Mustapha Suleiman, K., Aisyah, M. R., & Kamarul Rahim, K. (2010). Antioxidant levels and activities of selected seeds of malaysian tropical fruits. *Malaysia Journal of Nutrition*, 16, 149-159.
- Noudeh, G. D., Sharififar, F., Khatib, M., Behravan, E., & Afzadi, M. A. (2009). Study of aqueous extract of three medicinal plants on cell membrane-permeabilizing and their surface properties. *African Journal of Biotechnology*, *9*, 110-116.

- Novak, I., Šeruga, M., & Komorsky-Lovrić, Š. (2010). Characterization of catechins in green and black teas using square-wave voltammetry and RP-HPLC-ECD. *Food Chemistry*, 122, 1283-1289.
- Nurdiyana, A. B. S. (2008). Comparison of gaharu (Aquilaria malaccensis) essential oil composition between each country. University Malaysia Pahang, Malaysia.
- Ohlsson, T., & Bengtsson, N. (2002). *Minimal processing technologies in the food industry*. North America: CRC Press LLC.
- Okudera, Y., & Ito, M. (2009). Production of agarwood constituents in *Aquilaria* calli and cell suspension cultures. *Plant biotechnology*, 26, 307-315.
- Osborn, H., & Akoh, C. (2003). Effects of natural antioxidants on iron-catalyzed lipid oxidation of structured lipid-based emulsions. *Journal of the American Oil Chemists' Society*, 80, 847-852.
- Oyen, L. P. A., & Dung, N. X. (1999). Prosea: plant resources of south-east asia 19, essential-oil plants. Indonesia: Bogor.
- Ozcelik, B., Lee, J. H., & Min, D. B. (2003). Effects of light, oxygen, and ph on the absorbance of 2,2-diphenyl-1-picrylhydrazyl. *Journal of Food Science*, 68, 487-490.
- Padda, M. S. (2006). Phenolic composition and antioxidant activity of sweetpotatoes (Ipomoea batatas (L.) Lam). Louisiana State University and Agricultural and Mechanical College, United States.
- Pandi-Perumal, S. R., & Cardinali, D. P. (2007). *Melatonin: from molecules to therapy*. New York: Nova Science.
- Payne, A. H., & Glish, G. L. (2005). Tandem mass spectrometry in quadrupole ion trap and ion cyclotron resonance mass spectrometers. *Methods in Enzymology*, 402, 109-148.
- Pedersen-Bjergaard, S., Rasmussen, K. E., & Grønhaug Halvorsen, T. (2000). Liquidliquid extraction procedures for sample enrichment in capillary zone electrophoresis. *Journal of Chromatography A*, 902, 91-105.
- Penumetcha, M., Khan, N., & Parthasarathy, S. (2000). Dietary oxidized fatty acids: an atherogenic risk? *Journal of Lipid Research*, *41*, 1473-1480.

Peterson, B. (1997). Thymelaceacea. Flora of Thailand, 6, 226-245.

Peyrat-Maillard, M. N., Cuvelier, M. E., & Berset, C. (2003). Antioxidant activity of phenolic compounds in 2,2 ' -azobis (2-amidinopropane) dihydrochloride (AAPH)-induced oxidation: Synergistic and antagonistic effects. *Journal of the American Oil Chemists' Society*, 80, 1007-1012.

- Pinelo, M., Arnous, A., & Meyer, A. S. (2006). Upgrading of grape skins: significance of plant cell-wall structural components and extraction techniques for phenol release. *Trends in Food Science & Technology*, 17, 579-590.
- Pinelo, M., Del Fabbro, P., Marzocco, L., Nunez, M. J., & Vicoli, M. C. (2005a). Optimization of continuous phenol extraction from *Vitis vinifera* byproducts. *Food Chemistry*, 92, 109-117.
- Pinelo, M., Rubilar, M., Jerez, M., Sineiro, J., & Nunez, M. J. (2005b). Effect of solvent, temperature, and solvent-to-solid ratio on the total phenolic content and antiradical activity of extracts from different components of grape pomace. *Journal of Agricultural and Food Chemistry*, 53, 2111-2117.
- Pitt, J. J. (2009). Principles and applications of liquid chromatography-mass spectrometry in clinical biochemistry. *Clinical Biochemistry Reviews*, 30, 19-34.
- Poiana, M. A. (2012). Enhancing oxidative stability of sunflower oil during convective and microwave heating using grape seed extract. *International Journal of Molecular Sciences*, 13, 9240-9259.
- Poole, C. F. (2003). Thin-layer chromatography: challenges and opportunities. *Journal* of Chromatography A, 1000, 963-984.
- PORIM. (1995). PORIM Test Methods. In). Ministry of Primary Industries: Malaysia: Palm Oil Research Institute of Malaysia.
- Powell, C., Clifford, M. N., Opie, S. C., & Gibson, C. L. (1995). Use of Porter's reagents for the characterization of thearubigins and other non-proanthocyanidins. *Journal of the Science of Food and Agriculture*, 68, 33-38.
- Prachakul, M. (1989). *Histology characteristic of normal and abnormal of agarwood* (Aquilaria crassna) heartwood. Kasetsart University, Bangkok.
- Pranakhon, R., Aromdee, C., & Pannangpetch, P. (2015). Effects of iriflophenone 3-Cbeta-glucoside on fasting blood glucose level and glucose uptake. *Pharmacognosy Magazine*, 11, 82-89.
- Prior, R. L., Lazarus, S. A., Cao, G., Muccitelli, H., & Hammerstone, J. F. (2001). Identification of procyanidins and anthocyanins in blueberries and cranberries (*Vaccinium Spp.*) using high-performance liquid chromatography/mass spectrometry. *Journal of Agricultural and Food Chemistry*, 49, 1270-1276.
- Prior, R. L., Wu, X., & Schaich, K. (2005). Standardized methods for the determination of antioxidant capacity and phenolics in foods and dietary supplements. *Journal* of Agricultural and Food Chemistry, 53, 4290-4302.
- Qaadan, F., Nahrstedt, A., Schmidt, M., & Mansoor, K. (2010). Polyphenols from Ginkgo biloba. Scientia Pharmaceutica, 78, 897-907.

- Qiu, H.-X., Liu, J., Kong, H., Liu, Y., & Mei, X.-g. (2007). Isobolographic analysis of the antinociceptive interactions between ketoprofen and paracetamol. *European Journal of Pharmacology*, 557, 141-146.
- Rafalowski, R., Zegarska, Z., Kuncewicz, A., & Borejszo, Z. (2008). Fatty acid composition, tocopherols and β-carotene content in polish commercial vegetable oils. *Pakistan Journal of Nutrition*, 7, 278-282.
- Rajalakshmi, D., & Narasimhan, S. (1996). Food antioxidants: sources and methods of evaluation. In D. L. Madhavi, S. S. Deshpande & D. K. Salunkhe (Eds.), *Food* antioxidants technological, toxicological, and health perspectives, (pp. 65-66). USA: Marcel Dekker, Inc.
- Ramarathnam, N., Osawa, T., Ochi, H., & Kawakishi, S. (1995). The contribution of plant food antioxidants to human health. *Trends in Food Science & Technology*, 6, 75-82.
- Rao, B. S., Shanbhoge, R., Upadhya, D., Jagetia, G. C., Adiga, S. K., Kumar, P., Guruprasad, K., & Gayathri, P. (2006). Antioxidant, anticlastogenic and radioprotective effect of Coleus aromaticus on Chinese hamster fibroblast cells (V79) exposed to gamma radiation. *Mutagenesis*, 21, 237-242.
- Rasmy, N. M., Hassan, A. A., Foda, M. I., & El-Moghazy, M. M. (2012). Assessment of the antioxidant activity of Sage (*Salvia officinalis* L.) extracts on the shelf life of mayonnaise. *World Journal of Dairy & Food Sciences*, 7, 28-40.
- Rassol, N., Rizwan, K., Zubaur, M., Naveed, K. U. R., Imran, I., & Ahmed, V. U. (2011). Antioxidant potential of different extracts and fractions of *Catharanthus roseus* shoots. *International Journal of Phytomedicine*, 3, 108-114.
- Ratnam, D. V., Ankola, D. D., Bhardwaj, V., Sahana, D. K., & Kumar, M. N. V. R. (2006). Role of antioxidants in prophylaxis and therapy: A pharmaceutical perspective. *Journal of Controlled Release*, 113, 189-207.
- Rice-Evans, C. (2004). Flavonoids and isoflavones: absorption, metabolism, and bioactivity. *Free Radical Biology and Medicine*, *36*, 827-828.
- Riemer, J., Hoepken, H. H., Czerwinska, H., Robinson, S. R., & Dringen, R. (2004). Colorimetric ferrozine-based assay for the quantitation of iron in cultured cells. *Analytical Biochemistry*, 331, 370-375.
- Rivière, C., Hong, V. N. T., Pieters, L., Dejaegher, B., Heyden, Y. V., Van, M. C., & Quetin-Leclercq, J. (2009). Polyphenols isolated from antiradical extracts of Mallotus metcalfianus. *Phytochemistry*, 70, 86-94.
- Robards, K. (2003a). Strategies for the determination of bioactive phenols in plants, fruit and vegetables. *Journal of Chromatography A*, 1000, 657-691.
- Robards, K. (2003b). Strategies for the determination of bioactive phenols in plants, fruit and vegetables. *Journal of Chromatography A*, 1000, 657-691.

- Rohman, A., Riyanto, S., Yuniarti, N., Saputra, W. R., Utami, R., & Mulatsih, W. (2010). Antioxidant activity, total phenolic, and total flavonoid of extracts and fractions of red fruit (*Pandanus conoideus* Lam.). *International Food Research Journal*, 17, 97-106.
- Romano, C. S., Abadi, K., Repetto, V., Vojnov, A. A., & Moreno, S. (2009). Synergistic antioxidant and antibacterial activity of rosemary plus butylated derivatives. *Food Chemistry*, 115, 456-461.
- Rosenberg, E. (2003). The potential of organic (electrospray- and atmospheric pressure chemical ionisation) mass spectrometric techniques coupled to liquid-phase separation for speciation analysis. *Journal of Chromatography A*, *1000*, 841-889.
- Routray, W., & Orsat, V. (2012). Microwave-assisted extraction of flavonoids: a review. *Food and Bioprocess Technology*, *5*, 409-424.
- Rozin, P., Kabnick, K., Pete, E., Fischler, C., & Shields, C. (2003). The ecology of eating: smaller portion sizes in France Than in the United States help explain the French paradox. *Psychological Science*, 14, 450-454.
- Sakanaka, S., Tachibana, Y., & Okada, Y. (2005). Preparation and antioxidant properties of extracts of Japanese persimmon leaf tea (kakinoha-cha). *Food Chemistry*, 89, 569-575.
- Santos-Buelga, C., Escribano-Bailon, M. T., & Lattanzio, V. (2010). *Recent advances in polyphenol research*. UK: Blackwell Publishing Ltd.
- Santos, J. P., Zaniquelli, M. E. D., De Giovani, W. F., & Galembeck, S. E. (2002). Aluminum ion complex formation with 3-hydroxyflavone in Langmuir and Langmuir–Blodgett films. *Colloids and Surfaces A: Physicochemical and Engineering Aspects*, 198–200, 569-576.
- Sarker, S. D., Latif, Z., & Gray, A. I. (2006). *Natural product isolation-an overview*. (2nd Edition ed.). New Jersey: Humana Press Inc.
- Schroeder, M. T., Becker, E. M., & Skibsted, L. H. (2006). Molecular mechanism of antioxidant synergism of tocotrienols and carotenoids in palm oil. *Journal of Agricultural and Food Chemistry*, 54, 3445-3453.
- Schwarz, K., Frankel, E. N., & German, J. B. (1996). Partition behaviour of antioxidative phenolic compounds in heterophasic systems. *Lipid / Fett, 98*, 115-121.
- Shahidi, F. (1997). *Natural antioxidant: chemistry, health effects and applications*. USA: AOCS Press.
- Shahidi, F., & Naczk, M. (1995). Food phenolics: sources, chemistry, effects and applications. Lancaster and Basel: Technomic.

- Shahidi, F., & Wanasundara, U. N. (2002). Methods for measuring oxidative rancidity in fats and oils. In C. C. Akoh & D. B. Min (Eds.), *Food lipids: chemistry*, *nutrition and biotechnology* 3rd ed., (pp. 387-395). Florida: CRC Press.
- Shahidi, F., & Zhong, Y. (2005). Lipid oxidation: measurement methods. In F. Shahidi (Ed.), *Bailey's industrial oil and fat products* 6th ed., (pp. 361-369). New Jersey: John Wiley & Sons, Inc.
- Shi, J., Nawaz, H., Pohorly, J., Mittal, G., Kakuda, Y., & Jiang, Y. (2005). Extraction of polyphenolics from plant material for functional foods—engineering and technology. *Food Reviews International*, 21, 139-166.
- Shi, J., Yu, J., Pohorly, J., Young, J. C., Bryan, M., & Wu, Y. (2003). Optimization of the extraction of polyphenols from grape seed meal by aqueous ethanol solution. *Journal of Food, Agriculture and Environment*, 1, 42-47.
- Shiao, Y. J., Chen, W. P., & Lin, Y. L. (2009). New polyphenols and triterpene from Pseudobulbs of *Pleione formosana*. Journal of the Chinese Chemical Society, 56, 828-833.
- Siems, W. G., Grune, T., & Esterbauer, H. (1995). 4-Hydroxynonenal formation during ischemia and reperfusion of rat small intestine. *Life Sciences*, *57*, 785-789.
- Sies, H. (1997). Oxidative stress: oxidants and antioxidants. *Experimental Physiology*, 82, 291-295.
- Silva, E. M., Rogez, H., & Larondelle, Y. (2007). Optimization of extraction of phenolics from *Inga edulis* leaves using response surface methodology. *Separation and Purification Technology*, 55, 381-387.
- Sin, H. N., Yusof, S., Abdul Hamid, N. S., & Abd. Rahman, R. (2006). Optimization of hot water extraction for sapodilla juice using response surface methodology. *Journal of Food Engineering*, 74, 352-358.
- Snyder, L. R., Kirkland, J. J., & Dolan, J. W. (2010). Introduction to modern liquid chromatography. USA: Wiley.
- Soong, Y.-Y., & Barlow, P. J. (2004). Antioxidant activity and phenolic content of selected fruit seeds. *Food Chemistry*, 88, 411-417.
- Sparr Eskilsson, C., & Björklund, E. (2000). Analytical-scale microwave-assisted extraction. *Journal of Chromatography A*, 902, 227-250.
- Spigno, G., Tramelli, L., & De Faveri, D. M. (2007). Effects of extraction time, temperature and solvent on concentration and antioxidant activity of grape marc phenolics. *Journal of Food Engineering*, 81, 200-208.
- Stadtman, E. R. (2004). Role of oxidant species in aging. *Current Medicinal Chemistry*, *11*, 1105-1112.

- Stalikas, C. D. (2007). Extraction, separation, and detection methods for phenolic acids and flavonoids. *Journal of Separation Science*, 30, 3268-3295.
- Stobiecki, M., & Kachlicki, P. (2006). The science of flavonoids. In E. Grotewold (Ed.), *Isolation and identification of flavonoids*, (pp. 47-70). New york: Springer.
- Suhara, Y., Kamao, M., Tsugawa, N., & Okano, T. (2004). Method for the determination of vitamin k homologues in human plasma using high-performance liquid chromatography-tandem mass spectrometry. *Analytical Chemistry*, 77, 757-763.
- Sukanya, S. L., Sudisha, J., Prakash, H. S., & Fathima, S. K. (2011). Isolation and characterization of antimicrobial compound from *Chromolaena odorata*. *Journal of Phytology*, 3, 26-32.
- Tachakittirungrod, S., Ikegami, F., & Okonogi, S. (2007). Antioxidant active principles isolated from Psidium guajava grown in Thailand. *Scientia Pharmaceutica*, 75, 179-193.
- Tallarida, R. J. (2001). Drug synergism: its detection and applications. *Journal of Pharmacology and Experimental Therapeutics*, 298, 865-872.
- Tan, P. W., Tan, C. P., & Ho, C. W. (2011). Antioxidant properties: effect of solid-tosolvent ratio on antioxidant compounds and capacities of Pegaga (*Centella* asiatica). International Food Research Journal, 18, 553-558.
- Tang, S. Z., Kerry, J. P., Sheehan, D., Buckley, D. J., & Morrissey, P. A. (2000). Dietary tea catechins and iron-induced lipid oxidation in chicken meat, liver and heart. *Meat Science*, 56, 285-290.
- Tang, Z. H., Qin, J. P., Xu, X. N., Shi, G. R., Yang, H., & Liang, Y. Z. (2011). Applying silica gel column chromatography purify resveratrol from extracts of Morus alba L. Leaf. 5, 3020-3027.
- Thomsen, M. K., Jacobsen, C., & Skibsted, L. H. (2000). Mechanism of initiation of oxidation in mayonnaise enriched with fish oil as studied by electron spin resonance spectroscopy. *European Food Research and Technology*, 211, 381-386.
- Thoo, Y. Y., Abas, F., Lai, O.-M., Ho, C. W., Yin, J., Hedegaard, R. V., Skibsted, L. H., & Tan, C. P. (2013). Antioxidant synergism between ethanolic *Centella asiatica* extracts and α-tocopherol in model systems. *Food Chemistry*, 138, 1215-1219.
- Thoo, Y. Y., Ho, S. K., Liang, J. Y., Ho, C. W., & Tan, C. P. (2010). Effects of binary solvent extraction system, extraction time and extraction temperature on phenolic antioxidants and antioxidant capacity from mengkudu (*Morinda citrifolia*). Food Chemistry, 120, 290-295.
- Tiengtum, P. (1995). *In-Vitro culture of agarwood trees (Aquilaria sp.)*. Kasetaart University, Bangkok.

- Toma, M., Vinatoru, M., Paniwnyk, L., & Mason, T. J. (2001). Investigation of the effects of ultrasound on vegetal tissues during solvent extraction. *Ultrasonics Sonochemistry*, 8, 137-142.
- Tozuka, Z., Kaneko, H., Shiraga, T., Mitani, Y., Beppu, M., Terashita, S., Kawamura, A., & Kagayama, A. (2003). Strategy for structural elucidation of drugs and drug metabolites using (MS)n fragmentation in an electrospray ion trap. *Journal* of Mass Spectrometry, 38, 793-808.
- Trombino, S., Serini, S., Di Nicuolo, F., Celleno, L., Ando, S., Picci, N., Calviello, G., & Palozza, P. (2004). Antioxidant effect of ferulic acid in isolated membranes and intact cells: synergistic interactions with alpha-tocopherol, beta-carotene, and ascorbic acid. *Journal of Agricultural and Food Chemistry*, 52, 2411-2420.
- Tsen, S. Y., Ameri, F., & Smith, J. S. (2006). Effects of rosemary extracts on the reduction of heterocyclic amines in beef patties. *Journal of Food Science*, 71, C469-C473.
- Tubaro, F., Ghiselli, A., Rapuzzi, P., Maiorino, M., & Ursini, F. (1998). Analysis of plasma antioxidant capacity by competition kinetics. *Free Radical Biology and Medicine*, 24, 1228-1234.
- Ullah, M. O., Haque, M., Urmi, K. F., Zulfiker, A. H., Anita, E. S., Begum, M., & Hamid, K. (2013). Anti-bacterial activity and brine shrimp lethality bioassay of methanolic extracts of fourteen different edible vegetables from Bangladesh. *Asian Pacific Journal of Tropical Biomedicine*, 3, 1-7.
- Uma, D. B., Ho, C. W., & Wan Aida, W. M. (2010). Optimization of extraction parameters of total phenolic compounds from henna (*Lawsonis intermis*) leaves. *Sains Malaysia*, 39, 119-128.
- Uri, N. (1961). Mechanisms of autoxidation. New York: John Wiley & Sons.
- Valko, M., Rhodes, C. J., Moncol, J., Izakovic, M., & Mazur, M. (2006). Free radicals, metals and antioxidants in oxidative stress-induced cancer. *Chemico-Biological Interactions*, 160, 1-40.
- Van der Ven, C., Gruppen, H., de Bont, D. B. A., & Voragen, A. G. J. (2002). Optimization of the angiotensin converting enzyme inhibition by whey protein hydrolysates using response surface methodology. *International Dairy Journal*, *12*, 813-820.
- Vázquez, G., Fernández-Agulló, A., Gómez-Castro, C., Freire, M. S., Antorrena, G., & González-Álvarez, J. (2012). Response surface optimization of antioxidants extraction from chestnut (*Castanea sativa*) bur. *Industrial Crops and Products*, 35, 126-134.
- Vázquez, G., Fontenla, E., Santos, J., Freire, M. S., González-Álvarez, J., & Antorrena, G. (2008). Antioxidant activity and phenolic content of chestnut (*Castanea*

*sativa*) shell and eucalyptus (*Eucalyptus globulus*) bark extracts. *Industrial Crops and Products*, 28, 279-285.

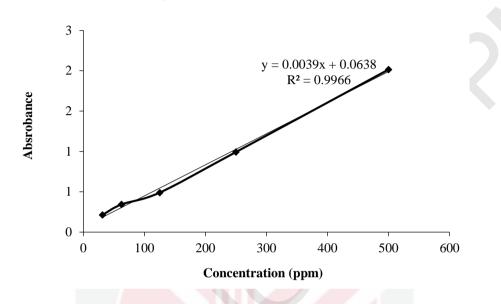
- Vermerris, W., & Nicholson, R. (2006). *Phenolic compound biochemistry*. USA: Springer Science + Business Media B. V.
- Vieira, F. G., Borges Gda, S., Copetti, C., Gonzaga, L. V., Nunes Eda, C., & Fett, R. (2009). Activity and contents of polyphenolic antioxidants in the whole fruit, flesh and peel of three apple cultivars. *Arch Latinoam Nutr*, 59, 101-106.
- Vlase, L., Benedec, D., Hanganu, D., Damian, G., Csillag, I., Sevastre, B., Mot, A., Silaghi-Dumitrescu, R., & Tilea, I. (2014). Evaluation of antioxidant and antimicrobial activities and phenolic profile for *Hyssopus officinalis*, *Ocimum basilicum* and *Teucrium chamaedrys*. *Molecules*, 19, 5490-5507.
- Vulic, I., Vitarelli, G., & Zenner, J. M. (2002). Structure–property relationships: phenolic antioxidants with high efficiency and low colour contribution. *Polymer Degradation and Stability*, 78, 27-34.
- Vuong, Q. V., Golding, J. B., Nguyen, M., & Roach, P. D. (2010). Extraction and isolation of catechins from tea. *Journal of Separation Science*, 33, 3415-3428.
- Wan, C., Yu, Y., Zhou, S., Tian, S., & Cao, S. (2011). Isolation and identification of phenolic compounds from Gynura divaricata leaves. *Pharmacogn Mag*, 7, 101-108.
- Wang, J., Liu, H., Zhao, J., Gao, H., Zhou, L., Liu, Z., Chen, Y., & Sui, P. (2010). Antimicrobial and antioxidant activities of the root bark essential oil of *Periploca sepium* and its main component 2-hydroxy-4-methoxybenzaldehyde. *Molecules*, 15, 5807-5817.
- Wang, J., Sun, B., Cao, Y., Tian, Y., & Li, X. (2008). Optimization of ultrasound-assisted extraction of phenolic compounds from wheat bran. *Food Chemistry*, 106, 804-810.
- Wang, T., Jónsdóttir, R., & Ólafsdóttir, G. (2009). Total phenolic compounds, radical scavenging and metal chelation of extracts from Icelandic seaweeds. *Food Chemistry*, 116, 240-248.
- Warade, A., Gaikwad, R., Sapkal, R., & Sapkal, V. (2011). Simulation of multistage countercurrent liquid-liquid extraction. *Leonardo Journal of Sciences*, 20, 79-94.
- Weiwitayaklung, P., Thavanapong, N., & Charoenteeraboon, J. (2009). Chemical constituents and antimicrobial activity of essential oil and extracts of Heartwood of Aquilaria crassna obtained from water distillation and supercritical fluid carbon dioxide extraction. Silpakorn University Science & Technology Journal, 3, 25-33.

Wilkinson, J. (2003). Revise in a month VCE chemistry. Sydney: Pascal Press.

- Wilson, I. D., & Lewis, S. (1985). Contemporary developments in thin-layer chromatography. *Journal of Pharmaceutical and Biomedical Analysis*, 3, 491-501.
- Wong Paz, J. E., Muñiz Márquez, D. B., Martínez Ávila, G. C. G., Belmares Cerda, R. E., & Aguilar, C. N. (2015). Ultrasound-assisted extraction of polyphenols from native plants in the Mexican desert. *Ultrasonics Sonochemistry*, 22, 474-481.
- Wrona, M., Korytowski, W., Różanowska, M., Sarna, T., & Truscott, T. G. (2003). Cooperation of antioxidants in protection against photosensitized oxidation. *Free Radical Biology and Medicine*, 35, 1319-1329.
- Wu, L.-c., Hsu, H.-W., Chen, Y.-C., Chiu, C.-C., Lin, Y.-I., & Ho, J.-a. A. (2006). Antioxidant and antiproliferative activities of red pitaya. *Food Chemistry*, 95, 319-327.
- Xu, B. J., & Chang, S. K. C. (2007). A comparative study on phenolic profiles and antioxidant activities of legumes as affected by extraction solvents. *Journal of Food Science*, 72, S159-S166.
- Yan, L. Y., Teng, L. T., & Jhi, T. J. (2006). Antioxidant properties of guaca fruit: comparison with some local fruits. *Sunway academic journal*, 3, 2006.
- Yang, M.-H., & Schaich, K. M. (1996). Factors affecting DNA damage caused by lipid hydroperoxides and aldehydes. *Free Radical Biology and Medicine*, 20, 225-236.
- Yang, Z.-Y., Zeng, E. Y., Xia, H., Wang, J.-Z., Mai, B.-X., & Maruya, K. A. (2006). Application of a static solid-phase microextraction procedure combined with liquid–liquid extraction to determine poly(dimethyl)siloxane–water partition coefficients for selected polychlorinated biphenyls. *Journal of Chromatography A*, 1116, 240-247.
- Yang, Z., Tu, Y., Baldermann, S., Dong, F., Xu, Y., & Watanabe, N. (2009). Isolation and identification of compounds from the ethanolic extract of flowers of the tea (*Camellia sinensis*) plant and their contribution to the antioxidant capacity. *LWT - Food Science and Technology*, 42, 1439-1443.
- Yildirim, A., Gumus, M., Dalga, S., Sahin, Y. N., & Akcay, F. (2003). Dehydroepiandrosterone improves hepatic antioxidant systems after renal ischemia-reperfusion injury in rabbits. *Ann Clin Lab Sci, 33*, 459-464.
- Yim, H. S., Fook, Y. C., Ho, S. K., & Ho, C. W. (2009). Phenolic profiles of selected edible wild mushrooms as affected by extraction solvent, time and temperature. *Asian Journal of Food and Agro-Industry*, *2*, 392-401.
- Yin, J., Becker, E. M., Andersen, M. L., & Skibsted, L. H. (2012). Green tea extract as food antioxidant. Synergism and antagonism with α-tocopherol in vegetable oils and their colloidal systems. *Food Chemistry*, 135, 2195-2202.

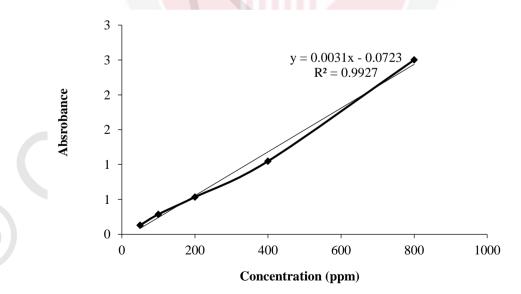
- Yong, Y. S. (2007). Determination of synthetic phenolic antioxidants in food items using HPLC and total antioxidants using FIA approaches. University Sains Malaysia, Malaysia.
- Young, I. S., & Woodside, J. V. (2001). Antioxidants in health and disease. *Journal of Clinical Pathology*, 54, 176-186.
- Zhang, G., Hu, M., He, L., Fu, P., Wang, L., & Zhou, J. (2013). Optimization of microwave-assisted enzymatic extraction of polyphenols from waste peanut shells and evaluation of its antioxidant and antibacterial activities *in vitro*. *Food and Bioproducts Processing*, 91, 158-168.
- Zhang, L., Wang, Y., Wu, D., Xu, M., & Chen, J. (2011a). Microwave-assisted extraction of polyphenols from Camellia oleifera fruit hull. *Molecules*, *16*, 4428-4437.
- Zhang, L., Yang, X., Zhang, Y., Wang, L., & Zhang, R. (2011b). In vitro antioxidant properties of different parts of pomegranate flowers. Food and Bioproducts Processing, 89, 234-240.
- Zhang, Q.-A., Zhang, Z.-Q., Yue, X.-F., Fan, X.-H., Li, T., & Chen, S.-F. (2009). Response surface optimization of ultrasound-assisted oil extraction from autoclaved almond powder. *Food Chemistry*, 116, 513-518.
- Zhang, Q., Zhang, J., Shen, J., Silva, A., Dennis, D., & Barrow, C. (2006). A simple 96well microplate method for estimation of total polyphenol content in seaweeds. *Journal of Applied Phycology*, 18, 445-450.
- Zhang, S. Q., Bi, H. M., & Liu, C. J. (2007a). Extraction of bio-active components from *Rhodiola sachalinensis* under ultrahigh hydrostatic pressure. *Separation and Purification Technology*, 57, 277-282.
- Zhang, Y., Jiao, J., Liu, C., Wu, X., & Zhang, Y. (2008). Isolation and purification of four flavone C-glycosides from antioxidant of bamboo leaves by macroporous resin column chromatography and preparative high-performance liquid chromatography. *Food Chemistry*, 107, 1326-1336.
- Zhang, Z.-S., Li, D., Wang, L.-J., Ozkan, N., Chen, X. D., Mao, Z.-H., & Yang, H.-Z. (2007b). Optimization of ethanol–water extraction of lignans from flaxseed. *Separation and Purification Technology*, 57, 17-24.
- Zhang, Z., Pang, X., Ji, Z., & Jiang, Y. (2001). Role of anthocyanin degradation in litchi pericarp browning. *Food Chemistry*, 75, 217-221.
- Zhou, B., Wu, L.-M., Yang, L., & Liu, Z.-L. (2005). Evidence for α-tocopherol regeneration reaction of green tea polyphenols in SDS micelles. *Free Radical Biology and Medicine*, 38, 78-84.
- Zhou, M., Wang, H., Suolangjiba, Kou, J., & Yu, B. (2008). Antinociceptive and antiinflammatory activities of *Aquilaria sinensis* (Lour.) Gilg. leaves extract. *Journal of Ethnopharmacology*, 117, 345-350.

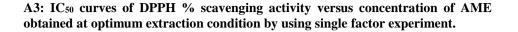
## APPENDICES

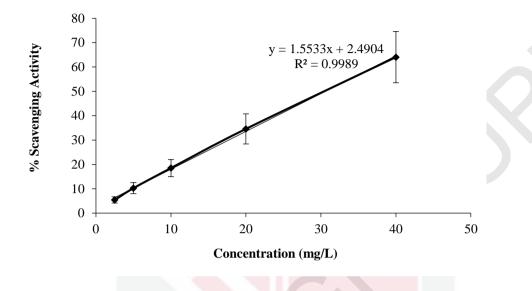


### A1: Calibration curves for gallic acid for determination of TPC.

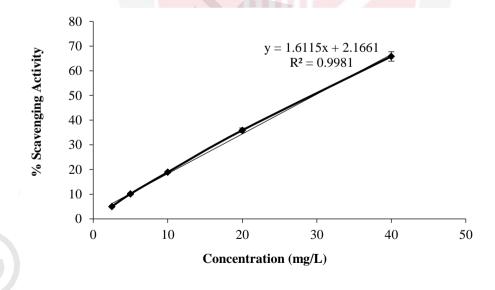
A2: Calibration curves for quercetin for determination of TFC.



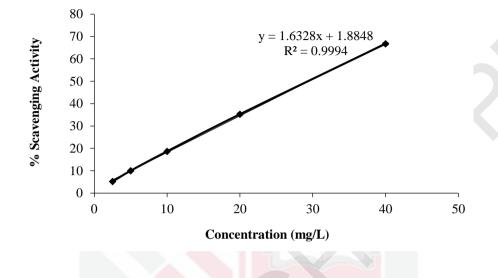




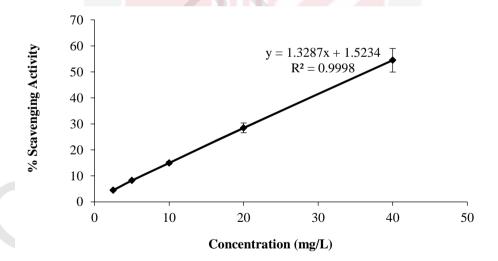
A4: IC<sub>50</sub> curves of DPPH % scavenging activity versus concentration of ASE obtained at optimum extraction condition by using single factor experiment.



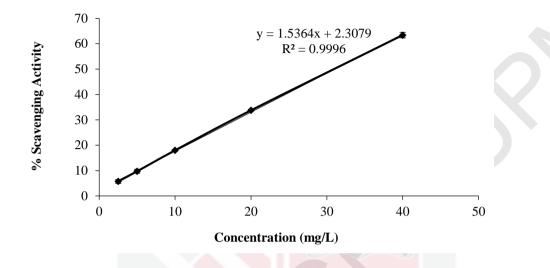
A5: IC<sub>50</sub> curves of DPPH % scavenging activity versus concentration of ACE obtained at optimum extraction condition by using single factor experiment.



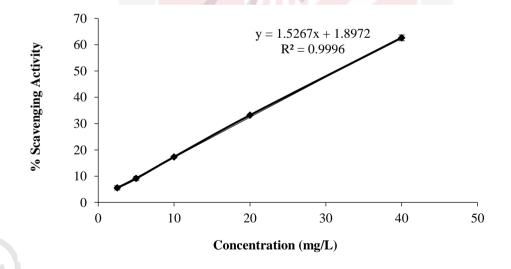
A6: IC<sub>50</sub> curves of DPPH % scavenging activity versus concentration of AME obtained at optimum extraction condition by using RSM.



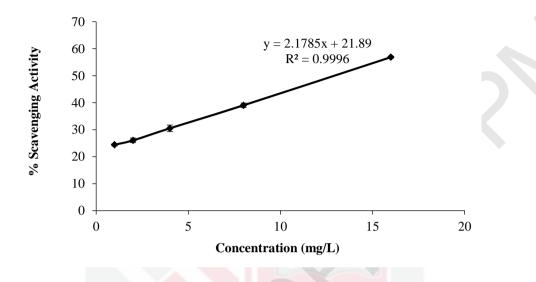
A7: IC<sub>50</sub> curves of DPPH % scavenging activity versus concentration of ASE obtained at optimum extraction condition by using RSM.



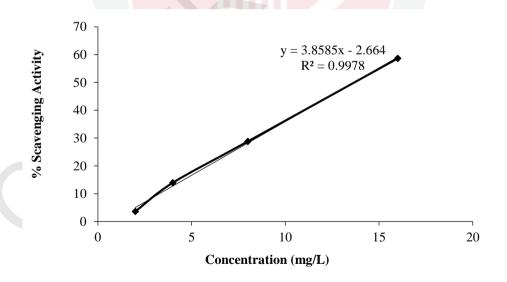
A8: IC<sub>50</sub> curves of DPPH % scavenging activity versus concentration of ACE obtained at optimum extraction condition by using RSM.

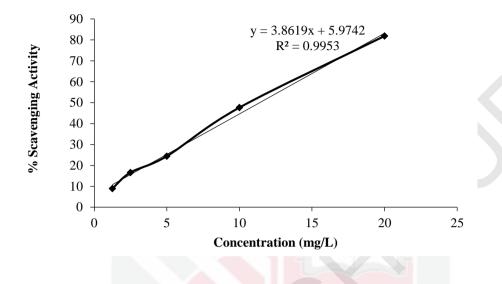


A9: IC  $_{50}$  curves of DPPH % scavenging activity versus concentration of ascorbic acid.



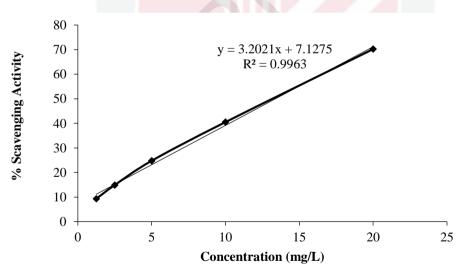
A10: IC<sub>50</sub> curves of DPPH % scavenging activity versus concentration of αtocopherol.





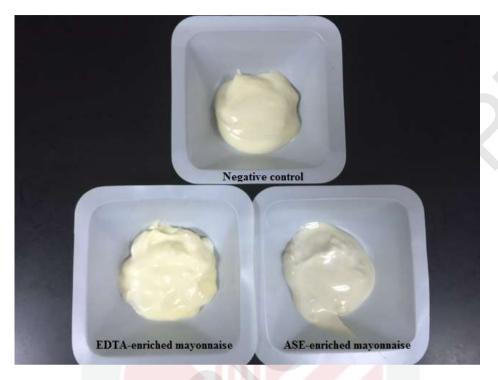
A11: IC  $_{50}$  curves of DPPH % scavenging activity versus concentration of (+)-catechin hydrate.

A12: IC<sub>50</sub> curves of DPPH % scavenging activity versus concentration of BHA.



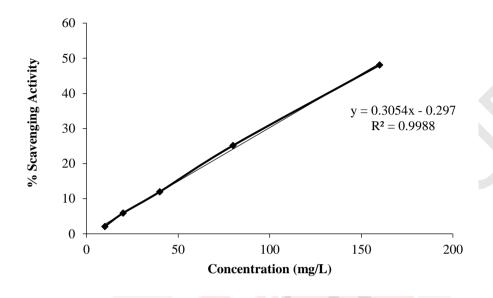
 $\bigcirc$ 

# A13: Color of mayonnaises



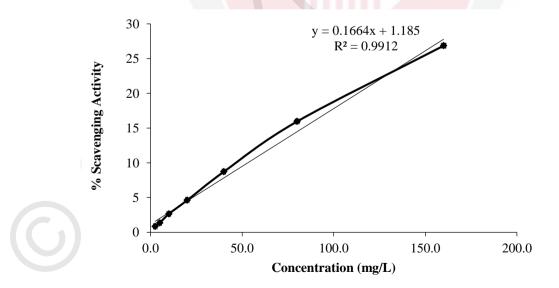
A14: Yield of *Aquilaria subintegra* young leaves extracted from different solvent fractions.

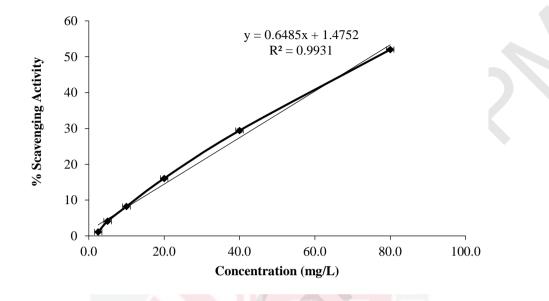
Fractions	Yield of samples (g)	Recovery rate (%)
Hexane	1.7822	2.31
Chloroform	3.7520	4.86
Ethyl acetate	7.0238	9.09
Water	59.3425	76.82
Total	71.9005	93.07
Total weight of the sample before liquid-liquid extraction = 77.2529 g		



A15: IC  $_{50}$  curves of DPPH % scavenging activity versus concentration of hexane fraction.

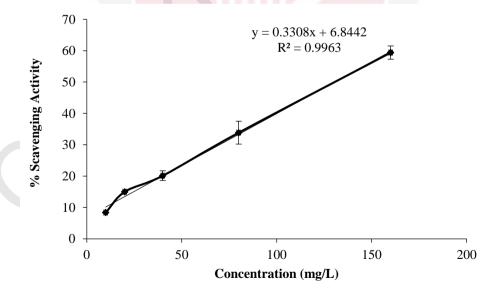
A16: IC<sub>50</sub> curves of **DPPH %** scavenging activity versus concentration of chloroform fraction.

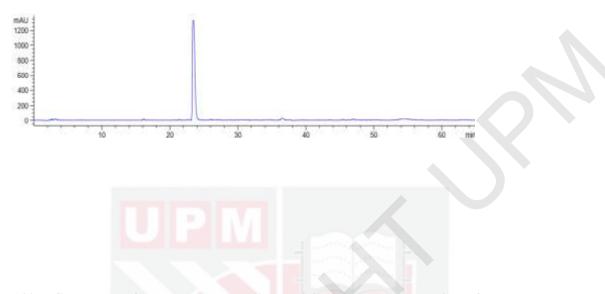




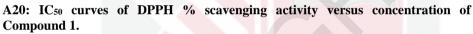
A17: IC  $_{50}$  curves of DPPH % scavenging activity versus concentration of ethyl acetate fraction.

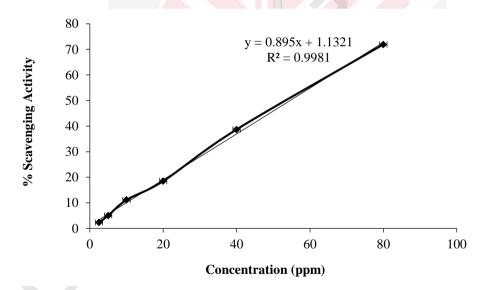
A18: IC<sub>50</sub> curves of DPPH % scavenging activity versus concentration of water fraction.





# A19: HPLC chromatogram of iriflopheone-3-*C*-β-glucoside.





### **BIODATA OF STUDENT**

Tay Pei Yin was born on 17<sup>th</sup> February 1989 and raised in a small town, Muar in a state of Johor. She comes from a family of six and is the second daughter to her parents. She completed her primary and secondary school education at SJK(C) Chung Hwa Presbyterian and Chung Hwa High School, respectively.

After graduating from secondary school, she enrolled UCSI University in the subsequent year and successfully completed with First Class Honours in Bachelor of Science in Food Science and Technology in 2011. Her honour thesis entitled "Effects of ethanol concentration, solid-to-solvent ratio and extraction time on antioxidant properties of gaharu (*Aquilaria crassna*) young shoots (3.0mm)" was presented in August 2011 under the supervision of Assoc. Prof. Dr. Ho Chun Wai.

After the graduation, she had worked as a research assistant at the Faculty of Applied Sciences, UCSI for one year. In September 2012, she entered Universiti Putra Malaysia (UPM) as a PhD candidate under the supervision of Prof. Dr. Tan Chin Ping. As indicated in the present thesis, the principle objectives of her project were to extract the polyphenols from three commercial species of Malaysian agarwood by-product, followed by the evaluation of antioxidant synergism with  $\alpha$ -tocopherol in various *in vitro* model system and its ability to inhibit lipid peroxidation in mayonnaise, as well as the identification and quantification of the major polyphenol constituent presented in agarwood by-product.

## LIST OF PUBLICATIONS

- Tay, P. Y., Tan, C. P., Abas, F., Yim, H. S., and Ho, C. W. (2014). Assessment of extraction parameters on antioxidant capacity, polyphenol content, epigallocatechin gallate (EGCG), ppicatechin gallate (ECG) and iriflophenone 3-C-β-glucoside of agarwood (*Aquilaria crassna*) young leaves. Molecules, 19 (8), 12304-12319.
- 2. Tay, P. Y., Tan, C. P., Abas, F., Yim, H. S.,and Ho, C. W. (In preparation). Effect of processing parameters on antioxidant capacity and polyphenol content of *Aquilaria malaccensis* leaves.
- 3. Tay, P. Y., Tan, C. P., Abas, F., Yim, H. S., and Ho, C. W. (In preparation). Effects of binary solvent extraction system, solid-to-solvent ratio, and extraction time on polyphenol content, antioxidant capacity, epigallocatechin gallate (EGCG), epicatechin gallate (ECG), and iriflophenone  $3-C-\beta$ -glucoside from agarwood (*Aquilaria subintegra*) young leaves.
- 4. Tay, P. Y., Abas, F., Ho, C. W., Thoo, Y. Y., and Tan, C. P. (In preparation). Antioxidant activity of an ethanolic extract from young agarwood (*Aquilaria subintegra*) leaves and its synergism with added α-tocopherol in model systems.
- 5. Tay, P. Y., Abas, F., Ho, C. W., and Tan, C. P. (In preparation). Antioxidative activity and chemical safety of agarwood (*Aquilaria subintegra*) young leaves extract during storage of mayonnaise.
- 6. Tay, P.Y., Tan, C. P., Abas, F., and Ho, C. W. Influence of processing parameters on antioxidant capacity and polyphenol content of agarwood (*Aquilaria malaccensis*) young leaves, Proceedings of 6<sup>th</sup> International Conference and Exhibition on Nutraceuticals & Functional Foods, Taipei, Taiwan, Nov. 5-9, 2013.