



UNIVERSITI PUTRA MALAYSIA

***CYTOTOXICITY AND MECHANISMS OF ACTION OF
ZERUMBONE-LOADED NANOSTRUCTURED LIPID CARRIER IN
TRIPLE NEGATIVE BREAST CANCER CELLS, MDA-MB-231***

MAHNAZ HOSSEINPOUR

IB 2015 23



**CYTOTOXICITY AND MECHANISMS OF ACTION OF
ZERUMBONE-LOADED NANOSTRUCTURED LIPID CARRIER IN
TRIPLE NEGATIVE BREAST CANCER CELLS, MDA-MB-231**

By

MAHNAZ HOSSEINPOUR

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfilment of the Requirements for the Degree of Master of
Science**

April 2015

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



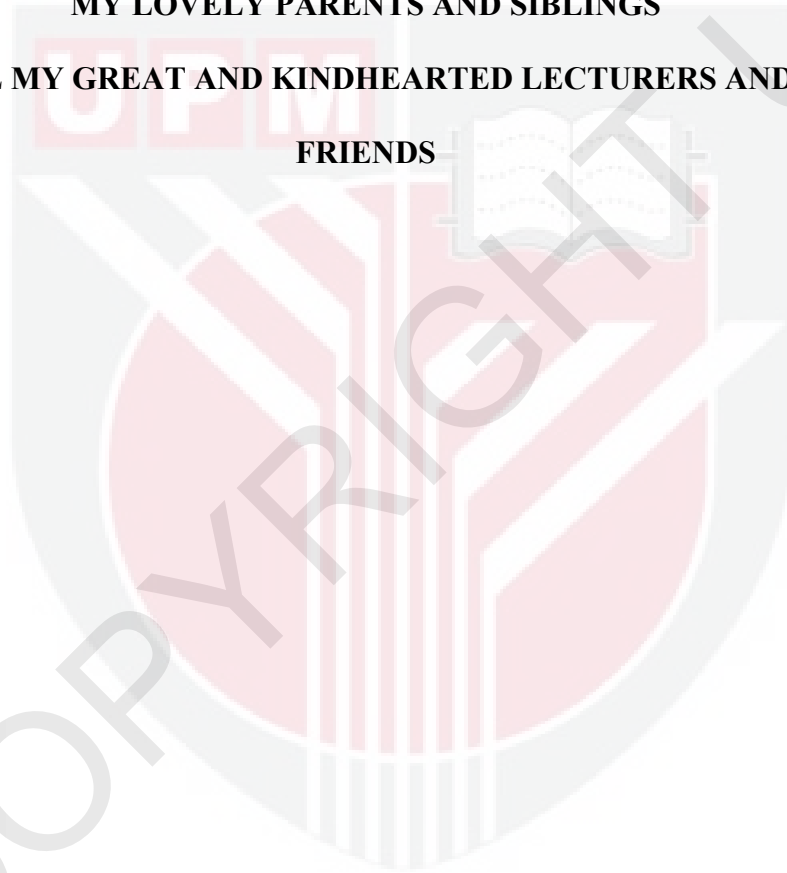
DEDICATION

THIS THESIS IS DEDICATED TO

MY BELOVED HUSBAND SAMAN MOHANDESAN

MY LOVELY PARENTS AND SIBLINGS

**ALL MY GREAT AND KINDHEARTED LECTURERS AND
FRIENDS**



Abstract of thesis presented to the Senate of University Putra Malaysia in
fulfilment of the requirement for the degree of Master of Science

**CYTOTOXICITY AND MECHANISMS OF ACTION OF ZERUMBONE-
LOADED NANOSTRUCTURED LIPID CARRIER IN TRIPLE NEGATIVE
BREAST CANCER CELLS, MDA-MB-231**

By

MAHNAZ HOSSEINPOUR

April 2015

Chairman: Ahmad Bustamam Abdul, PhD
Faculty: Institute of Bioscience

Breast cancer is a life-threatening disease mostly effecting women. According to the Malaysian National Cancer Registry (2006), breast cancer is one of the five most common cancers amongst Malaysian women. Currently, the most widely used breast cancer therapies include surgery, chemotherapy, radiotherapy, hormonal therapy and targeted therapy. Generally, most of these treatments can affect normal cells and tissues with several side effects. Therefore, scientists are applying new techniques and strategies, including the use of natural products in drug discovery for enhanced survival rates with fewer side effects. Zerumbone (ZER), a monocyclic, sesquiterpene crystalline compound, was isolated from a ginger herb, *Zingiber zerumbet* (L.) Smith, locally known as *lemboyang*. Several studies had demonstrated that ZER has the ability to inhibit abnormal proliferation of neoplastic colon, cervical, ovary, liver, breast and blood cells. In addition, ZER demonstrated selective antiproliferative effect towards cancer cells. Unfortunately, ZER is poorly soluble in water that leads to poor oral bioavailability and hence, delivery. For this reason, it is crucial to improve the delivery of this compound in blood by incorporation into nanostructured lipid carrier (ZER-NLC), which increases its water solubility. This study determined and compared the cytotoxicity of ZER-NLC with ZER towards triple negative breast cancer, MDA-MB-231, cells.

The IC₅₀ of ZER and ZER-NLC on MDA-MB-231 cells were determined by 3-(4,5-dimethylthiazol-2-yl) -2, 5-diphenyltetrazolium bromide assay. Mode of cell death was determined by microscopic observation of AO/PI double stained cells, annexin-V, TUNEL, caspases assays and cell cycle analysis. In addition, Western blot was used to determine the expression of Bcl-2, Bcl-xL, Bax, cytochrome c (Cyt-c) and proliferation cell nuclear antigen (PCNA) proteins.

ZER and ZER-NLC significantly ($P < 0.05$) decreased the proliferation of MDA-MB-231 cells in a time-dependent manner. The IC₅₀ of ZER and ZER-NLC on the MDA-MB-231 cells at 72 hours, were 5.96 ± 0.13 and 6.01 ± 0.11 $\mu\text{g/mL}$,

respectively. Both ZER and ZER-NLC caused development of apoptotic features in MDA-MDB-231 cells, particularly nuclear condensation, cell shrinkage, chromatin cleavage and membrane blebbing. Treatment with ZER and ZER-NLC caused MDA-MB-231 cell cycle arrest at G2/M phase. Furthermore, annexin-V and tunnel assay also demonstrated that both ZER and ZER-NLC induced apoptosis in MDA-MB-231 cells. ZER and ZER-NLC induced apoptosis via intrinsic (mitochondrial) pathway through the activation of caspase-3 and caspase-9. The western blot analysis provided evidence of up-regulation of Bax, Cyt-c with down-regulation of Bcl-2, Bcl-xL and PCNA proteins in both ZER- and ZER-NLC-treated MDA-MB-231 cells. Finally, ZER and ZER-NLC treatment did not affect the proliferation of non-cancerous breast epithelial cells (MCF-10A), suggesting that they are cancer-cell specific.

In conclusion, the current study showed that the cytotoxic effect of ZER-NLC in the induction of apoptosis similar to that of ZER. Thus, NLC can be used as a carrier for ZER in the treatment of breast cancers.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
Sebagai memenuhi keperluan untuk ijazah Master Sains

**KESITOTOKSIKAN DAN MEKANISME TINDAKAN PEMBAWA LIPID
NANOSTRUKTUR TERMUAT ZERUMBON TERHADAP SEL KANSER
PAYUDARA NEGATIF GANDA TIGA, MDA-MB-231**

Oleh

MAHNAZ HOSSEINPOUR

April 2015

Pengerusi: Ahmad Bustamam Abdul, PhD

Fakulti: Institut Biosains

Kanser payudara adalah penyakit yang paling banyak berlaku kepada wanita. Mengikut *Malaysian National Cancer Registry* (2006), kanser payudara merupakan antara lima kes kanser tertinggi dalam kalangan wanita Malaysia. Di kala ini, terapi kanser payudara yang kerap diguna termasuk pembedahan, kemoterapi, radioterapi, terapi hormon dan terapi sasaran. Secara umum, rawatan ini boleh mendatangkan kesan terhadap sel dan tisu normal dengan beberapa kesan sampingan. Zerumbon (ZER), suatu sebatian monosiklik, seskuiterpen hablur, telah dipencilkan daripada herba halia, *Zingiber zerumbet (L.) Smith*, yang juga dikenali sebagai lempoyang. Beberapa kajian menunjukkan ZER mempunyai keupayaan untuk menghalang pemroliferatan abnormal neoplasia kolon, serviks, ovari, hati, payudara dan sel darah. Tambahan, ZER mempamerkan kesan antipemroliferatan pilihan terhadap sel kanser. Malangnya, ZER kurang larut dalam air dan ini menyebabkan penurunan bioavailabiliti oral dan seterusnya penghantaran. Dengan sebab ini, adalah genting untuk meningkatkan penghantaran sebatian ini dalam darah dengan memuatkannya dalam pembawa lipid nanostruktur (ZER-NLC), yang meningkatkan kelarutannya. Kajian ini menentu dan membandingkan antara kesitoksikan ZER-NLC dengan ZER terhadap sel kanser payudara negatif ganda tiga, MDA-MB-231.

IC₅₀ ZER dan ZER-NLC pada sel MDA-MB-231 ditentukan melalui assai 3-(4, 5-dimetiltiazol-2-yl)-2, 5-difeniltetrazolium bromide. Cara kematian sel ditentukan melalui pencerapan mikroskopi terhadap sel terwarna berganda AO/PI, assai Annexin-V, TUNEL, dan kaspase dan analisis kitaran sel. Selain itu, sap Western digunakan untuk menentukan penyataan protein Bcl-2, Bcl-x1, Bax, cytochrome c (Cyt-c) dan antigen nucleus sel pemroliferatan (PCNA).

ZER dan ZER-NLC telah mengurangkan pemroliferatan sel MDA-MB-231 *in-vitro* secara tererti ($P < 0.05$) dan bersandarkan masa. IC₅₀ untuk ZER dan ZER-NLC terhadap sel MDA-MB-231 pada 72 jam, masing-masing adalah 5.96 ± 0.13 dan 6.01 ± 0.11 $\mu\text{g/mL}$. Kedua-dua ZER dan ZER-NLC menyebabkan

perkembangan ciri apoptosis pada sel MDA-MB-231, terutamanya pegenapan nucleus, pengecutan sel, pembelahan kromatin dan pembleban membran. Perlakuan dengan ZER dan ZER-NLC menyebabkan hentian kitaran sel MDA-MB-231 pada fasa G2/M. Tambahan pula, assai Annexin-V dan TUNEL menunjukkan yang kedua-duanya, ZER dan ZER-NLC mengaruh apoptosis terhadap sel MDA-MB-231. ZER dan ZER-NLC mengaruh apoptosis melalui arah laluan intrinsik (mitokondrion) secara pengaktifan kaspase-3 dan kapase-9. Analisis sap Western menunjukkan bukti berlaku pengawalan-naik protein Bax, Cyt-c dan pengawalan-turun protein Bcl-2, Bcl-xL dan PCNA dalam sel MDA-MB-231 yang diperlakukan dengan ZER dan ZER-NLC. Akhir sekali, perlakuan ZER dan ZER-NLC terhadap sel epitelium normal bukan kanser (MCF-10A) tidak memberi kesan pada pemproliferasinya, menyaran bahawa kesan kedua-duanya adalah khusus sel kanser.

Kesimpulannya, kajian ini menunjukkan bahawa kesan sitotoksik ZER-NLC dalam pengaruh apoptosis adalah serupa dengan ZER. Dengan demikian, NLC boleh digunakan sebagai pembawa ZER dalam rawatan kanser payudara.

ACKNOWLEDGEMENTS

First of all, I would like to thank God for giving me the opportunity to complete my master study in this delightful and beautiful country, Malaysia. I would like to express my deepest respect, gratitude and admiration to my main supervisor, Dr. Ahmad Bustamam Abdul, for his excellent guidance, motivation, care, encouragement and patience. I thank him for his great help and support during each and all steps of this project. It was a great chance for me to have his endless guidance all the time and certainly without that I would not be able to complete my thesis. I would like to express my utmost gratitude for the commitment, kind treatment and enthusiasm of my co-supervisor, Professor Dr. Rasedee Abdullah. I truly appreciate his invaluable help and support.

I wish to express my appreciation to all staff in the UPM-MAKNA Cancer Research Laboratory, the Laboratory of Immunotherapeutic and Vaccines, Institute of Bioscience, Universiti Putra Malaysia for their invaluable assistance. In addition, I always feel blessed for having my helpful and super kind friends especially Dr. Heshu Sulaiman Rahman. It would not have been possible to finish my research project without their great support and help.

I also wish to express my deepest appreciation to my beloved parents and my precious husband, Saman Mohandesan. In every moment of my life, I feel my family is the greatest gift of God. They always provide the best situation for my success.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirements for the Degree of Master of Science. The members of the Supervisory Committee were as follows:

AHMAD BUSTAMAM ABDUL, PhD

Senior Lecturer
Faculty of Medicine and Health Science
Universiti Putra Malaysia
(Chairman)

RASEDEE ABDULLAH, PhD

Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Member)

BUJANG KIM HUAT, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 17 June 2015

DECLARATION

Declaration by the student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: Mahnaz Hosseinpour, GS36178

TABLE OF CONTENTS

		Page
	ABSTRACT	i
	ABSTRAK	iii
	ACKNOWLEDGEMENTS	v
	APPROVAL	vi
	DECLARATION	viii
	LIST OF TABLES	xiii
	LIST OF FIGURES	xiv
	LIST OF ABBREVIATIONS	xvi
CHAPTER		
1	INTRODUCTION	1
2	LITERATURE REVIEW	
	2.1 Cancer	4
	2.2 Breast Cancer	5
	2.2.1 Staging and Grading of Breast Cancer	5
	2.2.2 Types of Breast Cancer	6
	2.2.2.1 Ductal Carcinoma <i>In Situ</i>	6
	2.2.2.2 Lobular Carcinoma <i>In Situ</i>	6
	2.2.2.3 Infiltrating Ductal Carcinoma	6
	2.2.2.4 Invasive Lobular Carcinoma	6
	2.2.3 Molecular Categorisation of Breast Cancer	6
	2.2.3.1 Estrogen Receptor Positive Tumour (Luminal Subtype)	7
	2.2.3.2 Estrogen Receptor Negative Tumour	8
	2.2.3.2.1 HER-2/Neu Amplified Tumour	8
	2.2.3.2.2 Triple Negative Breast Cancer	8
	2.2.4 Risk Factors	9
	2.3 Cancer Treatment	10
	2.4 Natural Products in Cancer Drug Discovery	10
	2.5 <i>Zingiber zerumbet</i>	12
	2.6 Zerumbone	13
	2.6.1 Anti Cancer Activity of zerumbone	14
	2.7 Apoptosis	15
	2.7.1 Intrinsic Pathway	16
	2.7.2 Extrinsic Pathway	17
	2.8 Drug Solubility and Enhancement Methods	18
	2.9 Drug Delivery System in Cancer Therapy	19
	2.10 Novel generation of Lipid nanoparticles	19
	2.11 Nanostructured Lipid Carrier (nanoparticle) in Cancer Therapy	21
	2.12 Zerumbone-loaded Nanostructured Lipid Carrier	22

3	MATERIALS AND METHODS	24
3.1	Materials	24
3.2	Methods	25
3.2.1	Cell Culture Condition	25
3.2.1.1	MDA-MB-231 Cell Culture	25
3.2.1.2	MCF-10A Cell Culture	25
3.2.2	Cryopreservation	25
3.2.3	Thawing of Cell	26
3.2.4	Preparation of Zerumbone Stock Solution	26
3.2.5	Preparation of Zerumbone-loaded Nanostructured Lipid Carrier Stock Solution	25
3.2.6	Cytotoxicity Assay	27
3.2.7	Morphological Analysis of MDA-MB-231 Cells Treated with ZER and ZER-NLC	28
3.2.8	Analysis of Moth of Cell Death	28
3.2.8.1	Dual Fluorescence Viability Method	28
3.2.8.2	Annexin V-FITC Assay	28
3.2.9	Quantification of Apoptotic Cells	29
3.2.10	Cell Cycle Arrest Analysis	29
3.2.11	Colorimetric Assays of Caspase-3, Caspase-8 and Caspase-9	30
3.2.12	Western Blotting Analysis	30
3.2.12.1	Protein Extraction and Quantification	30
3.2.12.2	Western Blotting Assay	31
3.2.13	Statistical Analysis	32
4	RESULTS	
4.1	Anti-proliferative Activity of Zerumbone and Zerumbone-loaded Nanostructured Lipid Carrier on Triple Negative Breast Cancer Cell Line	33
4.2	Effect of Zerumbone and Zerumbone-loaded Nanostructured Lipid Carrier on Normal Epithelial Mammary Gland Cells	35
4.3	Morphological Changes of MDA-MB-231 Cells Treated with ZER and ZER-NLC	36
4.4	Apoptosis Features in MDA-MB-231 Cells Treated with ZER and ZER-NLC	39
4.5	Mode of Cell Death in MDA-MB-231 Cells Treated with ZER and ZER-NLC	41
4.6	Effect of Zerumbone and Zerumbone-loaded Nanostructured Lipid Carrier on MDA-MB-231 Cell Cycle	44
4.7	DNA Fragmentation Induction by ZER and ZER-NLC on MDA-MB-231 Cell	47
4.8	Stimulation of Caspase Activity by Zerumbone and Zerumbone- loaded Nanostructured in Lipid Carrier on MDA-MB- 231 cells	50
4.9	Western Blot	52

5	DISCUSSION	56
6	SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	61
	REFERENCES	63
	APPENDICES	81
	BIODATA OF STUDENT	91
	LIST OF CONFERENCE PROCEEDING	92
	LIST OF PUBLICATIONS	92



LIST OF TABLES

Table		Page
2.1	Breast cancer subtypes based on IHC and clinical characteristics.	7
2.2	The <i>in vitro</i> antiproliferative effect of zerumbone on various cancer cell lines.	15
2.3	Properties of ZER-NLC.	22
4.1	IC ₅₀ values of MDA-MB-231 cells treated with zerumbone (ZER), zerumbone-loaded nanostructured carrier (ZER-NLC), zerumbone free nanoparticle lipid carrier (NLC) and doxorubicin.	34
4.2	Proportion of MDA-MB-231 cells treated with zerumbone (ZER) and zerumbone-loaded nanostructured lipid carrier (ZER-NLC) at different stages of apoptosis.	43
4.3	Proportion of MDA-MB-231 cells treated with zerumbone (ZER) and zerumbone-loaded nanostructured lipid carrier (ZER-NLC) at different phase of cell cycle.	46
4.4	Proportion of apoptotic cells in zerumbone (ZER) treated and zerumbone loaded nanostructured lipid carrier (ZER-NLC) treated MDA-MB-231 cells.	49
4.5	Caspase activities in MDA-MB-231 cells treated with zerumbone (ZER) and zerumbone-loaded nanostructured lipid carrier (ZER-NLC).	51
4.6	Percent and relative density values of proteins extracted from MDA-MB-231 cells treated with zerumbone after 12, 24 and 48 hours.	54
4.7	Percent and relative density values of proteins extracted from MDA-MB-231 cells treated with ZER-NLC after 12, 24 and 48 hours.	55

LIST OF FIGURES

Figure		Page
2.1	<i>Zingiber zerumbet</i> .	12
2.2	Rhizome of <i>Zingiber zerumbet</i> .	13
2.3	Chemical structure of zerumbone.	13
2.4	Formation of a perfect and highly ordered crystalline structure in SLN and a less ordered particle matrix with many imperfections in NLC.	21
2.5	Selective accumulation of nanoparticle by enhanced permeability and retention.	22
4.3.1	Cell population reduction in MDA-MB-231 cells treated with ZER and ZER-NLC during 24, 48 and 72 hours incubation, as compared to untreated control. ($\times 100$).	37
4.3.2	Morphological changes of MDA-MB-231 cells treated with ZER and ZER-NLC at IC_{50} values after 24, 48 and 72 hours. Untreated cells are shown as control ($\times 400$).	38
4.4	Fluorescent micrograph of AO/PI-stained cells treated with ZER and ZER-NLC after 24, 48 and 72 hours. Untreated cells are viable cells (VC) with normal structure ($\times 400$).	40
4.5	Flow cytometric analysis showing apoptotic fraction (FITC-conjugated Annexin V and propidium iodide) in MDA-MB-231 cells after treatment with ZER and ZER-NLC at different times. Untreated cells are represented as control.	42
4.6	Cell cycle analysis of MDA-MB-231 cells treated with ZER and zerumbone-loaded nanostructured lipid carrier (ZER-NLC) after 12, 24 and 48 hours treatment.	45
4.7	Flow cytometric analysis of apoptosis induction in MDA-MB-231 cells treated with ZER and ZER-NLC after staining with rTdT.	48
4.8	Caspase activities in MDA-MB-231 cells treated with zerumbone (ZER) and zerumbone-loaded nanostructured lipid carrier (ZER-NLC) at different time point.	50
4.9.1	Protein expressions in MDA-MB-231 cells treated with zerumbone after 12, 24 and 48 hours treatment observed via western blotting assay. Untreated cells (control) were compared to treated cells.	53
4.9.2	Protein expressions in MDA-MB-231 cells treated with zerumbone-loaded nanostructured lipid carrier after 12, 24 and 48 hours treatment observed via western blotting assay.	53

D1	Viability of MDA-MB-231 cells treated with A: zerumbone (ZER), B: zerumbone-loaded nanostructured lipid carrier (ZER-NLC) after 24, 48 and 72 hours incubation.	85
D2	Viability of MDA-MB-231 cells treated with ZER-free (blank) NLC (A) and doxorubicin (B) after 24, 48 and 72 hours incubation.	86
D3	Viability of MCF-10A cells treated with A: zerumbone (ZER) and B: zerumbone-loaded nanostructured lipid carrier (ZER-NLC) after 24, 48 and 72 hours incubation.	87
E	The comparison between cell cycle analyses of MDA-MB-231 cells treated with ZER, ZER-NLC and untreated cells (control).	88
F	The molecular weight of B-action (A) and Bcl-2, Bcl-xL, PCNA, Bax and cytochrome-c (B) proteins expressed in MDA-MB-231 cells treated with ZER.	89
G	The molecular weight of B-action (A) and Bcl-2, Bcl-xL, PCNA, Bax and cytochrome-c (B) proteins expressed in MDA-MB-231 cells treated with ZER-NLC.	90

LIST OF ABBREVIATIONS

AI	Aromatase inhibitors
AIF	Apoptosis-inducing factor
AO	Acridine orange
Apaf-1	Apoptosis protease activating factor
ATCC	American Type Culture Collection
Bad	Bcl-2 associated death promoter
Bak	Bcl-2 homologous antagonist killer
Bax	Bcl ₂ associated x protein
B-cell	B lymphocyte
Bcl-2	B cell lymphoma 2
Bcl-xL	B cell lymphoma extra large
BCPT	Breast Cancer Prevention Trial
<i>BRCA1/2</i>	Breast cancer susceptibility genes 1 and 2
BSA	Bovine serum albumin
CAD	Caspase-activated DNase
CAM	Complementary and alternative medicine
Caspase	Cysteine-aspartic protease
c-FLIP	Cellular (FADD-like IL-1 β -converting enzyme)-inhibitory protein
COX-2	Cyclooxygenase -2
DCIS	Ductal carcinoma <i>in situ</i>
DISC	Death-inducing signalling complex
EBV	Epstein-Bar virus
EGF	Epidermal growth factor
EPR	Enhanced permeability and retention
ER	Estrogen receptor
FADD	Fas-associated protein with death domain
GSH	Glutathione
HER-2	Human Epidermal Growth Factor Receptor 2
IAP	Inhibitor of apoptosis protein
IDC	Invasive ductal carcinoma
ILC	Invasive lobular carcinoma
LCIS	Lobular carcinoma <i>in situ</i>
NADPH	Nicotinamide adenine dinucleotide phosphate
NCI	National Cancer Institute
NF- κ B	Nuclear factor kappa-light-chain-enhancer of activated B cells
NK	Natural killer
NLC	Nanostructured lipid carrier
p53	Tumour protein p53
PARP	Poly ADP ribose polymerase
PCNA	Proliferating cell nuclear antigen
PI	Propidium iodide
PR	Progesterone receptor
PUMA	p53 upregulated modulator of apoptosis

SERMs	Selective estrogen receptor modulators
SGM	Specific growth media for MCF-10A
SLN	Solid lipid nanoparticle
Smac	Second mitochondria-derived activator of caspases
TdT	Terminal deoxynucleotidyl transferase
TNBC	Triple negative breast cancer
TNF	Tumour necrosis factor
TNFR1	Tumour necrosis factor receptor 1
TPBC	Triple positive breast cancer
TRADD	TNFR1 associated death domain
TRAF	Tumour necrosis factor receptor-associated factor
TRAF1	TNF receptor-associated factor 1
TRAF2	TNF receptor-associated factor 2
TRAIL	TNF related apoptosis-including ligand
TUNEL	Tdt-mediated dUTP Nick End labelling
XO	Xanthine oxidase
ZER	Zerumbone
ZER-NLC	Zerumbone loaded nanostructured lipid carrier

CHAPTER 1

INTRODUCTION

A life-threatening disease and the most prevalent cancer amongst women is breast cancer. In 2002, this disease caused more than 400,000 deaths worldwide and accounted for 30.4% of all malignancies among women of all ethnic groups in Malaysia (Lim, 2003). The presence of genetic expressions of estrogen receptor (ER), progesterone receptor (PR) and amplification of human epidermal growth factor receptor 2 (HER-2 or HER 2/neu) provide three categorisations for breast cancer tumours, including estrogen receptor positive tumour (triple positive breast cancer, TPBC), estrogen receptor-negative tumour involving HER-2 amplified tumours and triple negative breast cancer (TNBC), in which there is no expression of ER, PR and HER-2 amplification (Lam *et al.*, 2013). In general, chemotherapy, hormonal therapy, radiotherapy and surgery are examples of cancer treatment methods, unfortunately, they have drawbacks of affecting the normal cells and infuriating side effects. Several studies demonstrated the important selective toxicity of drugs originated from natural sources such as ginger, red grapes, and tea in human disease treatment, especially for cancer and infectious diseases (Yang *et al.*, 2001; Mulakayala *et al.*, 2013).

Compounds extracted from microorganisms, plants and animal sources have played important roles for treatment of human ailments. Approximately 60% and 75% of new drugs were obtained from natural sources in the field of cancer and infectious disease, between 1981 and 2002 (Lam, 2007). Natural product compounds, which have been applied in most clinical trials, mostly originate from plant sources (Harvey, 2008). The use of herbal medicine has been famous as one of the oldest methods for human healthcare (Spiteri *et al.*, 2013). Phytomedicines are defined as herbal medicine that derived from all or part of plants due to their pharmacologically active compounds for disease treatment (Pribitkin, 2005). As reported by the World Health Organization (WHO), about 75% of the worldwide population have used plant products for medicinal purposes (Barrett *et al.*, 1999).

Several studies have demonstrated that herbal medicine plays a unique role in the developing field of complementary and alternative medicine (CAM). CAM can be defined as a category of various medical and health care techniques, methods, exercises and products for the purpose of prevention, diagnosis and treatment of illnesses. However, they are not recognised as part of conventional medicine (Issa *et al.*, 2006).

People in both developed and undeveloped countries have utilised plants for medicinal purposes since 6000 years ago. The Egyptian pharmacopoeia of Ebers Papyrus, in 1500 BC, described the application of plant extracts such as oil of Castor beans and poppy of Opium for treatment of diseases (Gossell-Williams *et al.*, 2006). It means that, botanical medicine has been used since ancient times. Seven of eight plant specie's pollen that were found from the 60000-year-old

Neanderthal burial site were conspicuous phytomedicinals (Barrett *et al.*, 1999). This provided clear evidence that plants demonstrated significant medicinal role since the birth.

In Malaysia, there seems to be an ever-growing number of herbal plants claimed to induce anti-proliferative effects on cancers (Mohan *et al.*, 2010a; Mohan *et al.*, 2010b; Mohan *et al.*, 2011). For example, several bioactive compounds isolated from *Boesenbergia rotunda*, a perennial herb found in Asian countries like Malaysia, showed the therapeutic potential in various cancers including Boesenbergin A for lung cancer (Isa *et al.*, 2012), and Panduratin A for colon cancer (Yun *et al.*, 2005) and prostate cancer (Yun *et al.*, 2006). It has been demonstrated that *Typhonium flagelliforme*, a local herbal plant known as rodent tuber, inhibited the proliferation of leukaemia cells (Mohan *et al.*, 2010a; Mohan *et al.*, 2010b). *Elephantopus scaber* and *Murraya koenigii* are other plants that possess anticancer properties towards breast (Ho *et al.*, 2011) and liver cancer (Syam *et al.*, 2011), respectively.

The rhizomes of the Zingiberaceae family, which consists of the most prolific plants in the tropical rainforests, have been widely applied in many Asian countries as traditional recipes with medical functions (Singh *et al.*, 2011). Approximately 161 species from 18 genera of this family could be found in Peninsular Malaysia. Members of Zingiberaceae are usually rich in terpenoids, because they are aromatic and used as spices (Chen *et al.*, 2008). *Zingiber zerumbet* (L.) Smith, locally known as wild edible ginger or 'lempanyang', is one of the important species of this family, has been traditionally applied for toothache, swelling, muscle sprain and sores (Sulaiman *et al.*, 2009). The milky, mucilaginous content of the pine cones is usually utilised for natural shampoo and hair conditioner production, while the rhizomes and leaves are used as flavouring agents and medicine (Sabu, 2003). The rhizome of *Zingiber zerumbet* (RZZ) has high medicinal values due to the presence of many bioactive compounds such as humulene, monoterpenes, and zerumbone. For this reason, recent several studies have been performed to isolate and identify its phytochemical content for use as a medicine (Yob *et al.*, 2011).

Zerumbone, which is the major sesquiterpenoid component of essential oil in *Zingiber zerumbet*, has been reported to inhibit the proliferation of the cervix, ovary, neoplastic colon, breast and liver cancer cells (Abdelwahab *et al.*, 2011). However, poor solubility characteristic of zerumbone and consequently, its poor oral bioavailability and delivery have been the main reason to find alternatives to overcome this problem. Thus, application of nanoparticles and nanocarriers including solid lipid nanoparticles (SLN) and nanostructured lipid carriers (NLC) were proposed to improve drug-delivery by increasing solubility of zerumbone (Jens and Rainer, 2008; Mistry *et al.*, 2011).

Hypothesis

Zerumbone and zerumbone-loaded nanostructured lipid carrier induce death of human breast cancer line (MDA-MB-231) through the intrinsic apoptosis pathway.

Objectives of the Study

General Objective

To evaluate the *in vitro* anti-breast cancer activity of zerumbone and zerumbone-loaded nanostructured lipid carrier on the triple negative breast cancer, MDA-MB-231 cells.

Specific Objectives

To determine the;

1. Effect of zerumbone and zerumbone-loaded nanostructured lipid carrier on MDA-MB-231 breast cancer cells.
2. Apoptosis mechanism in MDA-MB-231 cells treated with zerumbone and zerumbone-loaded nanostructured lipid carrier.
3. Expression of Bax, Bcl-2, Cytochrome c, Bcl-xL, and PCNA protein in MDA-MB-231 cells treated with zerumbone and zerumbone-loaded nanostructured lipid carrier.

REFERENCES

- Abbasalipourkibir, R., Salehzada, A., & Rasedee, A. (2011). Solid lipid nanoparticles as new drug delivery system. *International Journal of Biotechnology and Molecular Biology Research*, 2: 252-261.
- Abdelwahab, S. I., Abdul, A. B., Alzubairi, A. S., Mohamed Elhassan, M., & Mohan, S. (2009). *In vitro* ultramorphological assessment of apoptosis induced by zerumbone on (HeLa). *Journal of biomedicine & biotechnology*, 2009: 769568-769578.
- Abdelwahab, S. I., Abdul, A. B., Devi, N., Ehassan, T. M., Al-Zubairi, A. S., Mohan, S., & Mariod, A. A. (2010). Regression of cervical intraepithelial neoplasia by zerumbone in female Balb/c mice prenatally exposed to diethylstilboestrol: Involvement of mitochondria-regulated apoptosis. *Experimental Toxicology and Pathology*, 62: 461-469.
- Abdelwahab, S. I., Abdul, A. B., Mohan, S., Taha, M. M., Syam, S., Ibrahim, M. Y., & Mariod, A. A. (2011). Zerumbone induces apoptosis in T-acute lymphoblastic leukemia cells. *Leukemia Research*, 35: 268-271.
- Abdelwahab, S. I., Abdul, A. B., Zain, Z. N. M., & Hadi, H. (2012). Zerumbone inhibits interleukin-6 and induces apoptosis and cell cycle arrest in ovarian and cervical cancer cells. *International immunopharmacology*, 12: 594-602.
- Abdul, A., Al-Zubairi, A., Tailan, N., Wahab, S., Zain, Z., Ruslay, S., & Syam, M. M. (2008). Anticancer activity of natural compound (zerumbone) extracted from *Zingiber zerumbet* in human HeLa cervical cancer cells. *International Journal of Pharmacology*, 4: 160-168.
- Abdul, A. B., Abdelwahab, S. I., Jalinas, J. B., Al-Zubairi, A. S., & Taha, M. M. (2009). Combination of zerumbone and cisplatin to treat cervical intraepithelial neoplasia in female balb/c mice. *International Journal of Gynecological Cancer*, 19: 1004-1010.
- Aggarwal, B. B. (2004). Nuclear factor-kappaB: the enemy within. *Cancer Cell*, 6: 203-208.
- Aggarwal, B. B., & Shishodia, S. (2006). Molecular targets of dietary agents for prevention and therapy of cancer. *Biochemical Pharmacology*, 71: 1397-1421.
- Alhaj, N. A., Abdullah, R., Ibrahim, S., & Bustamam, A. (2008). Tamoxifen drug loading solid lipid nanoparticles prepared by hot high pressure homogenisation techniques. *American Journal of Pharmacology and Toxicology*, 3: 219-224.

- Alwi, S. S., Nallappan, M., & Pihie, A. H. L., (2007). Zerumbone Exerts Antiproliferative Activity via Apoptosis on HepG2 Cells. *Malaysian Journal of Biochemistry and Molecular Biology*, 15: 19–23.
- Anassamy, T., Ahmad Bustamam, A., Mohd Aspollah, S., Siddig, I. A., Syam, M., Behnam, K., Aziz, M. Z., Nadzir, N. M., Andas, A. R. J., Beng, N. K., Hadi, A. H. A., & Rahman, H. S. (2013). A Phenylbutenoid Dimer, cis-3-(3', 4'-Dimethoxyphenyl)-4-[(E)-3''', 4'''-Dimethoxystyryl] Cyclohex-1-ene, Exhibits Apoptogenic Properties in T-Acute Lymphoblastic Leukemia Cells via Induction of p53-Independent Mitochondrial Signalling Pathway. *Evidence-Based Complementary and Alternative Medicine*, 2013: 14 pages.
- Asano, H., Fukunaga, S., Deguchi, Y., Kawamura, S., & Inaba, M. (2012). Bcl-xL and Mcl-1 are involved in prevention of in vitro apoptosis in rat late-stage erythroblasts derived from bone marrow. *The Journal of Toxicological Sciences*, 37: 23-31.
- Ashkenazi, A., & Dixit, V. M. (1998). Death receptors: signaling and modulation. *Science*. 281: 1305–8.
- Australian Cancer Research Foundation (2014). Overview of Breast Cancer. Sydney: Australian Cancer Research Foundation.
- Baba, A. I. (2009). Apoptosi and necrosis. *Lucrari Stintifice Medicina Veterinara*, 42: 3–5.
- Bachmeier, B., Fichtner, I., Killian, P. H., Kronschi, E., Pfeffer, U., & Efferth, T. (2011). Development of resistance towards artesunate in MDA-MB-231 human breast cancer cells. *PLoS One*. 6: e20550.
- Barrett. B., Kiefer, D., & Rabago, D. (1999). Assessing the risks and benefits of herbal medicine: an overview of scientific evidence. *Alternative Therapies in Health and Medicine*, 5: 40–49.
- Basu, A., & Haldar, S. (1998). The relationship between Bcl2 , Bax and p53 : consequences for cell cycle progression and cell death, *Molecular Human Reproduction*, 4: 1099–1109.
- Beaglehole, R., Bonita, R., & Magnusson, R. (2011). Global cancer prevention: an important pathway to global health and development. *Public Health*, 125: 821–831.
- Beg, A. A., & Baltimore, D. (1996). An essential role for NF- κ B in preventing TNF- α -induced cell death. *Science*. 274: 782–784.
- Bertram, J. S. (2000). The molecular biology of cancer. *Moleuclar Aspects of Medicine*, 21:167–223.
- Bharti, A. C., & Aggarwal, B. B. (2002). Nuclear factor- κ B and cancer (Its role in prevention and therapy). *Biochemical Pharmacology*. 64: 883–888.

- Bhattacharya, S., Prasanna, A., & Haldar, P. K. (2011). Evaluation of antiproliferative activity of *Trichosanthes dioica* root against Ehrlich ascites carcinoma cells. *Academic Journal of Cancer Research*, 4: 38-42.
- Bhuiyan, M. N. I., Chowdhury, J. U., & Begum, J. (2008). Chemical investigation of the leaf and rhizome essential oils of *Zingiber zerumbet* (L.) Smith from Bangladesh. *Bangladesh Journal of Pharmacology*, 4: 9-12.
- Bissonauth, V., Shatenstein, B., & Ghadirian, P. (2008). Nutrition and breast cancer among sporadic cases and gene mutation carriers: An overview. *Cancer Detection and Prevention*, 32:52–64.
- Boon, H., Stewart, M., Kennard, M. A., Gray, R., Sawka, C., Brown, J. B., McWilliam, C., Gavin, A., Baron, R. A., & Aaron, D. (2000). The use of complementary/alternative medicine by breast cancer survivors in Ontario: Prevalence and perceptions. *Journal of Clinical Oncology*, 18: 2515-2521.
- Bortner, C. D., Oldenburg, N. B., & Cidlowski, J. a. (1995). The role of DNA fragmentation in apoptosis. *Trends in Cell Biology*, 5: 21–26.
- Bosman, F. T., Visser, B. C., & van Oeveren, J. (1996). Apoptosis: pathophysiology of programmed cell death. *Pathology, Research and Practice*, 192: 676–683.
- Brahmachari, G. (2011). Natural Products in Drug Discovery: Impacts and Opportunities—An Assessment. In *Bioactive Natural Products*, ed. Brahmachari, G., pp. 1-199. Singapore: World Scientific Publishing Company.
- Bray, K. (2011). Manipulation of cell death pathways in cancer. *PhD Thesis*. University of Medicine and Dentistry of New Jersey.
- Brien, K. M. O., Cole, S. R., & Millikan, R. C. (2010). Intrinsic breast tumor subtypes, race, and long-term survival in the Carolina Breast Cancer Study Abstract Purpose Statement of Translational Relevance Methods Study Population. *Clinical Cancer Research*, 16: 6100–6110.
- Bushrab, N., & Müller, R. (2003). Nanocrystals of poorly soluble drugs for oral administration. *NewDrugs*, 20–22.
- Butler, M. S. (2008). Natural products to drugs: natural product-derived compounds in clinical trials. *Natural Product Reports*, 25: 475-516.
- Cancer Research UK (2014). Statistics and outlook for breast cancer. London: Cancer Research UK.
- Chan, W., Cheung, K., & Schorge, J. (2000). Bcl-2 and p53 Protein Expression, Apoptosis, and p53 Mutation in Human Epithelial Ovarian Cancers. *The American Journal of Pathology*, 156: 409–417.

- Chane-Ming, J., Vera, R., & Chalchat, J.-C. (2003). Chemical composition of the essential oil from rhizomes, leaves and flowers of *Zingiber zerumbet* Smith from Reunion Island. *Journal of Essential Oil Research*, 15: 202-205.
- Chavez, K. J., Garimella, S. V., & Lipkowitz, S. (2010). Triple negative breast cancer cell lines: one tool in the search for better treatment of triple negative breast cancer. *Breast Disease*, 32: 35–48.
- Cheang, M. C. U., Chia, S. K., Voduc, D., Gao, D., Leung, S., Snider, J., Watson, M., Davies, S., Bernard, P. S., Ellis, M. J., & Nielsen, T. O. (2009). Ki67 index, HER2 status, and prognosis of patients with luminal B breast cancer. *Journal of the National Cancer Institute*, 101:736–350.
- Chechina, O. E., Ryazantseva, N. V., & Novitsky, V. V. (2011). The proteins of Bcl-2 family are molecular targets of IL-2 and IL-4 apoptotic effects. *Immunology*, 32: 127-130.
- Chen, H., Kim, S., Li, L., Wang, S., Park, K., & Cheng, J. (2008). Release of hydrophobic molecules from polymer micelles into cell membranes revealed by Förster resonance energy transfer imaging. *Proceedings of the National Academy of Science of the United State of America*, 105: 6596–6601.
- Chen, I., Chang, C. h., Ng, C. C., Wang, C. Y., Shyu, Y. T., & Chang, T. L. (2008). Antioxidant and antimicrobial activity of Zingiberaceae plants in Taiwan. *Plant Foods for Human Nutrition*, 63: 15–20.
- Cheung, H., Liu, X., & Rennert, O. M. (2012). Apoptosis: Reprogramming and the fate of mature cells. *International Scholarly Research Notices Cell Biology*, 2012: 8 pages.
- Cho, K., Wang, X. U., Nie, S., & Shin, D. M. (2008). Therapeutic nanoparticles for drug delivery in cancer. *Clinical Cancer Research*, 14: 1310-1316.
- Coughlin, S. S., & Ekwueme, D. U. (2009). Breast cancer as a global health concern. *Cancer Epidemiology*, 33: 315–318.
- Chrystal, K., Allan, S., Forgeson, G., & Isaacs, R. (2003). The use of complementary/alternative medicine by cancer patients in a New Zealand regional cancer treatment centre. *The New Zealand Medical Journal*, 116: 80-89.
- Collins, K., Jacks, T., & Pavletich, N. P. (1997). The cell cycle and cancer. *Proceedings of the National Academy of Sciences of the United States of America*, 94: 2776–2778.
- Chow, A. (2010). Cell Cycle Control by Oncogenes and Tumor Suppressors: Driving the Transformation of Normal Cells into Cancerous Cells. *Nature Education*, 3: 1–6.

- Coultas, L., & Strasser, A. (2003). The role of the Bcl-2 protein family in cancer. *Seminars in Cancer Biology*, 13: 115–123.
- Cummings, S. R., Eckert, S., Krueger, K. A., Grady, D., Powles, T. J., Cauley, J. A., Norton, L., Nickesen, T., Black, D., & Jordan, V. C. (1999). The Effect of Raloxifene on Risk of Breast Cancer in Postmenopausal Women. *The Journal of American Medical Association*, 281: 2189–2198.
- Dahlui, M., Ramli, S., & Bulgiba, A. M. (2011). Breast cancer prevention and control programs in Malaysia. *Asian Pacific Journal of Cancer Prevention*, 12: 1631–1634.
- Dandekar, D. S., Lopez, M., Carey, R. I., & Lokeshwar, B. L. (2005). Cyclooxygenase-2 inhibitor celecoxib augments chemotherapeutic drug-induced apoptosis by enhancing activation of caspase-3 and-9 in prostate cancer cells. *International journal of cancer*, 115: 484-492.
- Davis, M. E. (2008). Nanoparticle therapeutics: an emerging treatment modality for cancer. *Nature Reviews Drug Discovery*, 7: 771-782.
- Deorukhkar, A., Ahuja, N., Mercado, A., Diagaradjane, P., Mohindra, P., Guha, S., Aggarwal, B., & Krishnan, S. (2010). Zerumbone, A Sesquiterpene from Southeast Asian Edible Ginger Sensitizes Colorectal Cancer Cells to Radiation Therapy. *International Journal of Radiation Oncology, Biology, Physics*, 78: S654.
- Diaz-Moralli, S., Tarrado-Castellarnau, M., Miranda, A., & Cascante, M. (2013). Targeting cell cycle regulation in cancer therapy. *Pharmacology and Therapeutics*, 138: 255–271.
- Dong, Y., & Feng, S. (2007). *In vitro* and *in vivo* evaluation of methoxy polyethylene glycol-polylactide (MPEG-PLA) nanoparticles for small-molecule drug chemotherapy. *Biomaterials*, 28: 4154–4160.
- Du, C., Fang, M., Li, Y., Li, L., & Wang, X. (2000). Smac, a mitochondrial protein that promotes cytochrome c-dependent caspase activation by eliminating IAP inhibition. *Cell*, 102: 33–42.
- Duve, R. (1980). Highlights of the chemistry and pharmacology of wild ginger *Zingiber zerumbet* Smith. *Agricultural Journal*, 42: 41-43.
- Edward, K. H., & Li, D. (2008). *Drug Like Properties: Concept, Structure, Design and Methods, from ADME to Toxicity Optimization*. 1st ed. United States of America: Elsevier Inc. pp. 56-85.
- Elmore, S. (2007). Apoptosis: a review of programmed cell death. *Toxicologic Pathology*, 35: 495–516.

- Elumalai, P., Gunadharini, D. N., Senthilkumar, K., Banudevi, S., Arunkumar, R., Benson, C. S., Sharmila, G., & Arunakaran, J. (2012). Induction of apoptosis in human breast cancer cells by nimbolide through extrinsic and intrinsic pathway. *Toxicology Letters*, 215: 131–142.
- Ekambaram, P., Sathali, A. H., & Priyanka, K. (2012). Solid lipid nanoparticles: A review. *Scientific Reviews and Chemical Communication*, 2: 80-102.
- Eroles, P., Bosch, A., Pérez-Fidalgo, J. A., & Lluch, A. (2012). Molecular biology in breast cancer: intrinsic subtypes and signaling pathways. *Cancer Treatment Reviews*, 38: 698–707.
- Foglieni, C., Meoni, C., & Davalli, A. M. (2001). Fluorescent dyes for cell viability: an application on prefixed conditions. *Histochemistry and Cell Biology*, 115: 223-229.
- Foo, J. B., Yazan, L. S., Chan, K. W., Tahir, P. M., & Ismail, M. (2011). Kenaf seed oil from supercritical carbon dioxide fluid extraction induced G1 phase cell cycle arrest and apoptosis in leukemia cells. *African Journal of Biotechnology*, 10: 5389-5397.
- Foulkes, W.D., Smith, I.E. & Reis-Filho, J. (2010) Triple-negative breast cancer. *The New England Journal of Medicine*, 363: 1938–1948.
- Freedman, A. N., Yu, B., Gail, M. H., Costantino, J. P., Graubard, B. I., Vogel, V. G., & Anderson, G. L. (2011). Benefit/risk assessment for breast cancer chemoprevention with raloxifene or tamoxifen for women age 50 years or older. *Journal of Clinical Oncology: Official Journal of the American Society of Clinical Oncology*, 29: 2327–2333.
- Gao, L., Zhang, D., & Chen, M. (2008). Drug nanocrystals for the formulation of poorly soluble drugs and its application as a potential drug delivery system. *Journal of Nanoparticle Research*, 10: 845-862.
- Gaurav, A. (2011). Comparative analysis of apoptotic function between humans, chimpanzees and macaques. *PhD Thesis*, Georgia University, USA.
- Gerber, D. E. (2008). Targeted therapies: a new generation of cancer treatments. *American Family Physician*, 77: 311–9.
- Golbeck, J., Fragoso, G., Hartel, F., Hendler, J., Oberthaler, J., & Parsia, B. (2011). The National Cancer Institute's thesaurus and ontology. *Web Semantics: Science, Services and Agents on the World Wide Web*.
- Gossell-Williams, M., Simon, O. R., & West, M. E. (2006). The past and present use of plants for medicines. *The West Indian medical journal*, 55: 217–8.
- Guicciardi, M. E., & Gores, G. J. (2009). Life and death by death receptors. *The Federation of American Societies for Experimental Biology Journal*, 23: 1625-1637.

- Gullo, V. P., & Hughes, D. E. (2005). Exploiting new approaches for natural product drug discovery in the biotechnology industry. *Drug Discovery Today: Technologies*, 2: 281–286.
- Güney, G., Genc, L., & Dikmen, G. (2011). Use of Nanocarrier Systems in Cancer Therapy. *Journal of Materials Science*, 5: 577–582.
- Hafidh, R.R., Abas, F., Abdulmir, A., Jahanshiri, F., Bakar, F., & Sekawi, Z. (2009). Cancer research of natural products in Asia. *International Journal of Cancer Research*, 5: 69-82.
- Harvey, A. L. (2008). Natural products in drug discovery. *Drug discovery today*, 13: 894–901.
- Hayashi, R., Ito, Y., Matsumoto, K., Fujino, Y., & Otsuki, Y. (1998). Quantitative Differentiation of Both Free 3'-OH and 5'-OH DNA Ends Between Heat-induced Apoptosis and Necrosis. *Journal of Histochemistry & Cytochemistry*, 46: 1051–1059.
- Heiser, D., Labi, V., Erlacher, M., & Villunger, A. (2004). The Bcl-2 protein family and its role in the development of neoplastic disease. *Experimental Gerontology*, 39: 1125–35.
- Ho, W. Y., Yeap, S. K., Ho, C. L., Raha, A. R., Suraini, A. A., & Alitheen, N. B. (2011). Elephantopus scaber induces cytotoxicity in MCF-7 human breast cancer cells via p53-induced apoptosis. *Journal of Medicinal Plants Research*, 5: 5741-5749.
- Hoffman, A., Spetner, L., & Burke, M. (2002). Redox-regulated mechanism may account for zerumbone's ability to suppress cancer-cell proliferation. *Carcinogenesis*, 23: 1961-1962.
- Hosemann, S. (2013). Compass : Breast Cancer Risk Reduction. *Oncology*, 58: 1–7.
- Huang, G. C., Chien, T. Y., Chen, L. G., & Wang, C. C. (2005) Antitumor effects of zerumbone from Zingiber zerumbet in P-388D1 cells *in vitro* and *in vivo*. *Journal of Medical Plant and Natural Product Research*, 71:219–224.
- Hudis, C. & Gianni, L. (2011). Triple-negative breast cancer: an unmet medical need. *The Oncologist*, 16: 1–11.
- IARC. (2002). Some traditional herbal medicines, some mycotoxins, naphthalene and styrene. *Monographs on the Evaluation of Carcinogenic Risks to Human*, 82: 41–70.
- Isa, N. M., Abdul, A. B., Abdelwahab, S. I., Abdullah, R., Sukari, M. A., Kamalidehghan, B., Hadi, A. H. A., & Mohan, S (2012). Boesenbergin A, a chalcone from *Boesenbergia rotunda* induces apoptosis via mitochondrial dysregulation and cytochrome c release in A549 cells *in vitro*: Involvement

- of HSP70 and Bcl2/Bax signalling pathways. *Journal of Functional Foods*, 5: 87-97.
- Ismail-Khan, R., & Bui, M. M. (2010). A review of triple-negative breast cancer. *Cancer Control : Journal of the Moffitt Cancer Center*, 17: 173–176.
- Issa, A. Y., Volate, S. R., & Wargovich, M. J. (2006). The role of phytochemicals in inhibition of cancer and inflammation: New directions and perspectives. *Journal of Food Composition and Analysis*, 19: 405–419.
- Jantan, I., Yassin, M. S., Chin, C. B., Chen, L., & Sim, N. L. (2003). Antifungal activity of the essential oils of nine Zingiberaceae species. *Pharmaceutical Biology*, 41: 392–397.
- Jens, U. A., & Rainer, H. M. (2008). Nanocrystals technology, drug delivery and clinical applications. *International Journal of Nanomedicine*, 3: 295-309.
- Jaspers, J. E., Rottenberg, S., & Jonkers, J. (2009). Therapeutic options for triple-negative breast cancers with defective homologous recombination. *Biochimica et Biophysica Acta*, 1796: 266–280.
- Jia, T., Zhang, L., Duan, Y., Zhang, M., Wang, G., Zhang, J., & Zhao, Z. (2014). The differential susceptibilities of MCF-7 and MDA-MB-231 cells to the cytotoxic effects of curcumin are associated with the PI3K/Akt-SKP2-Cip/Kips pathway. *Cancer Cell International*, 14: 126.
- Jin, Z., & El-deiry, W. S. (2005). Overview of Cell Death Signaling Pathways. *Cancer biology & therapy*, 4: 139–163.
- Joensuu, H., & Gligorov, J. (2012). Adjuvant treatments for triple-negative breast cancers. *Annals of Oncology : Official Journal of the European Society for Medical Oncology*, 23: 40-45.
- Joo, W. D., Visintin, I., & Mor, G. (2013). Targeted cancer therapy-Are the days of systemic chemotherapy numbered. *Maturitas*, 76: 308–314.
- Junghanns, J.-U. A. H., & Müller, R. H. (2008). Nanocrystal technology, drug delivery and clinical applications. *International Journal of Nanomedicine*, 3: 295–309.
- Kajstura, M., Halicka, H. D., Pryjma, J., & Darzynkiewicz, Z. (2007). Discontinuous Fragmentation of Nuclear DNA During Apoptosis Revealed by Discrete “ Sub-G 1 ” Peaks on DNA Content Histograms, *Cytometry*, 71:125–131.
- Keck, C. M., & Müller, R.H. (2006). “Drug nanocrystals of poorly soluble drugs produced by high pressure homogenisation,” *European Journal of Pharmaceutics and Biopharmaceutics*, 62: 3–16.

- Khan, N., Adhami, V. M., & Mukhtar, H. (2008). Apoptosis by dietary agents for prevention and treatment of cancer. *Biochemical Pharmacology*, 76: 1333–1339.
- Kitayama, T., Okamoto, T., Hill, R. K., Kawai, Y., Takahashi, S., Yonemori, S., Yamamoto, Y., Ohe, K., & Sawada, S. (1999). Chemistry of zerumbone. 1. Simplified isolation, conjugate addition reactions, and a unique ring contracting transannular reaction of its dibromide. *The Journal of Organic Chemistry*, 64: 2667-2672.
- Kitayama, T., Yamamoto, K., Utsumi, R., Takatani, M., Hill, R. K., Kawai, Y., Sawada, S., & Okamoto, T. (2001). Chemistry of Zerumbone. 2. Regulation of Ring Bond Cleavage and Unique Antibacterial Activities of Zerumbone Derivatives. *Bioscience Biotechnology and Biochemistry*, 65: 2193-2199.
- Kitayama, T., Yokoi, T., Kawai, Y., Hill, R. K., Morita, M., Okamoto, T., Yamamoto, Y., Fokin, V. V., Sharpless, K. P., & Sawada, S. (2003). The chemistry of zerumbone. Part 5: Structural transformation of the dimethylamine derivatives. *Tetrahedron*, 59: 4857-4866.
- Kitayama, T., Furuya, A., Moriyama, C., Masuda, T., Fushimi, S., Yonekura, Y., Kubo, H., & Sawada, S. (2006). Elucidation of the sharpless epoxidation of zerumbol. *Tetrahedron: Asymmetry*, 17: 2311-2316.
- Komarova, E. & Gudkov, V. (2001). Chemoprotection from p53-dependent apoptosis: potential clinical applications of the p53 inhibitors. *Biochemical Pharmacology*, 62: 657–667.
- Krysko, D.V., Vanden Berghe, T., Parthoens, E., D’Herde, K., & Vandenabeele, P. (2008). Methods for distinguishing apoptotic from necrotic cells and measuring their clearance. *Methods in Enzymology*. 442: 307–341.
- Lam, K. S. (2007). New aspects of natural products in drug discovery. *Trends in microbiology*, 15: 279–89.
- Lam, S. W., Jimenez, C. R., & Boven, E. (2013). Breast cancer classification by proteomic technologies: Current state of knowledge. *Cancer Treatment Reviews*, 40: 129-138.
- Lampe, J. W. (1999). Health effects of vegetables and fruit: assessing mechanisms of action in human experimental studies. *The American Journal of Clinical Nutrition*, 70: 475-490.
- Lee, E., McKean-Cowdin, R., Ma, H., Spicer, D. V., Berg, D. V. D., Bernstein, L & Ursin, G. (2011). Characteristics of triple-negative breast cancer in patients with a BRCA1 mutation: results from a population-based study of young women. *Journal of Clinical Oncology*, 29: 4373–4380.

- Lee, H.-P., Li, T.-M., Tsao, J.-Y., Fong, Y.-C., & Tang, C.-H. (2012). Curcumin induces cell apoptosis in human chondrosarcoma through extrinsic death receptor pathway. *International Immunopharmacology*, 13: 163–169.
- Lee, J. S., Kim, H. S., Jung, J. J., Kim, Y. B., Park, C. S., & Lee, M. C. (2001). Correlation between angiogenesis, apoptosis and cell proliferation in invasive ductal carcinoma of the breast and their relation to tumor behavior. *Analytical and Quantitative Cytology and Histology*, 23: 161–168.
- Li, C. I., 2010. *Breast Cancer Epidemiology*, 1st ed. New York: Springer Science, Business Media.
- Liedtke, C., & Kiesel, L. (2012). Breast cancer molecular subtypes--modern therapeutic concepts for targeted therapy of a heterogeneous entity. *Maturitas*, 73: 288–294.
- Lim, G. C. C. (2002). Overview of cancer in Malaysia. *Japanese Journal of Clinical oncology*, 32: S37-S42.
- Lim, G.C.C., (2003) Cancer in Malaysia - There is Light at the End of the Tunnel. *Medical Journal of Malaysia*, 58: 632-635.
- Lim, G. C. C., & Halimah, Y. (2004). Cancer Incidence in Malaysia 2003. National Cancer Registry Kuala Lumpur.
- Liu, F. T., Newland, & A. C., Jia, L. (2003). Bax conformational change is a crucial step for PUMA-mediated apoptosis in human leukemia. *Biochemical and Biophysical Research Communications*, 310: 956–62.
- Macdonald, F., Ford, C. H. J., & Casson, A. G. (2004). *Molecular Biology of Cancer*. 2nd ed. New York: Garland Science/BIOS Scientific Publishers.
- Malkas, L. H., Herbert, B. S., Abdel-Aziz, W., Dobrolecki, L. E., Liu, Y., Agarwal, B., Hoelz, D., Badve, S., Schnaper, L. Mechref, Y., Goulet, R. J., & Hickey, R. J. (2006). A cancer-associated PCNA expressed in breast cancer has implications as a potential biomarker. *Proceedings of the National Academy of Sciences of the United States of America*, 103: 19472–19477.
- Malladi, S., Challa-Malladi, M., Bratton, S. B., & Charlene, A. M. (Eds.). (2010). *Apoptosis*. Oxford: Elsevier.
- Martins, S., Sarmiento, B., Ferreira, D. C., & Souto, E. B. (2007). Lipid-based colloidal carriers for peptide and protein delivery--liposomes versus lipid nanoparticles. *International Journal of Nanomedicine*, 2: 595–607.
- Mathur, V., Satrawala, Y., Rajput, M. S., Kumar, P., Shrivastava, P., & Vishvkarma, A. (2011). Solid lipid nanoparticles in cancer therapy. *International Journal of Drug Delivery*, 2: 192–199.

- Mehnert, W., & Mader, K. (2001). Solid lipid nanoparticles: Production, characterization and applications. *Advance Drug Delivery Reviews*, 47:165–196.
- Mistry, S. N., Patel, P. K., Bharadia, P. D., Pandya, V. M., & Modi, D. A. (2011). Novel Drug delivery system for lipophilic agents: Solid Lipid Nanoparticles. *Journal of Pharmaceutics and Cosmetology*, 1: 76-89.
- Mohan, S., Abdul, A. B., Abdelwahab, S. I., Al-Zubairi, A. S., Aspollah, S. M., Abdullah, R., Taha, M. M., Beng, N. K., & Isa, N. M. (2010a). Typhonium flagelliforme inhibits the proliferation of murine leukemia WEHI-3 cells in vitro and induces apoptosis in vivo. *Leukemia Research*, 34: 1483-1492.
- Mohan, S., Bustamam, A. A., Ibrahim, S., Al-Zubairi, A. S., Aspollah, M., Abdullah, R., & Elhassan, M. M. (2010b). In Vitro Ultramorphological Assessment of Apoptosis on CEMss Induced by Linoleic Acid-Rich Fraction from Typhonium flagelliforme Tuber. *Evidence-Based Complementary and Alternative Medicine*, 2011: 421894-421894.
- Mohan, S., Bustamam, A. A., Ibrahim, S., Al-Zubairi, A. S., Aspollah, M., Abdullah, R., & Elhassan, M. M. (2011). In Vitro Ultramorphological Assessment of Apoptosis on CEMss Induced by Linoleic Acid-Rich Fraction from Typhonium flagelliforme Tuber. *Evidence-Based Complementary and Alternative Medicine*, 2011: 421894-421894.
- Moorthi, C., Manavalan, R., & Kathiresan, K. (2011). Nanotherapeutics to overcome conventional cancer chemotherapy limitations. *Journal of Pharmacy and Pharmaceutical Sciences*, 14: 67–77.
- Mosmann, T. (1983). Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *Journal of Immunological Methods*. 65: 55-63.
- Mulakayala, C., Babajan, B., Madhusudana, P., Anuradha, C. M., Rao, R. M., Nune, R., Manna, S. K., Munakalaya, N., & Kumar, C. S. (2013). Synthesis and evaluation of resveratrol derivatives as new chemical entities for cancer. *Journal of Molecular Graphics & Modelling*, 41: 43–54.
- Mullan, P. B., Quinn, J. E., Gilmore, P. M., McWilliams, S., Andrews, H., Gervin, C., McCabe, N., McKenna, S., White, P., Song, Y. H., Maheswaran, S., Liu, E. Haber, D. A., & Harkin, D. P. (2001). BRCA1 and GADD45 mediated G2/M cell cycle arrest in response to antimicrotubule agents. *Oncogene*, 20: 6123–6131.
- Müller, R., Misund, K., Holien, T., Bachke, S., Gilljam, K. M., Våtsveen, T. K., Bellacchio, E., Sundan, A., & Otterlei, M. (2013). Targeting proliferating cell nuclear antigen and its protein interactions induces apoptosis in multiple myeloma cells. *PloS One*, 8: e70430.

- Müller, R. H., Radtke, M., & Wissing, S. A. (2002). Solid lipid nanoparticles (SLN) and nanostructured lipid carriers (NLC) in cosmetic and dermatological preparations. *Advanced Drug Delivery Reviews*, 54: 131–155.
- Murakami, A., Takahashi, D., Kinoshita, T., Koshimizu, K., Kim, H. W., Yoshihiro, A., Nakamura, Y., Jiwajinda, S., Terao, J., & Ohigashi, H. (2002). Zerumbone, a Southeast Asian ginger sesquiterpene, markedly suppresses free radical generation, proinflammatory protein production, and cancer cell proliferation accompanied by apoptosis: the α , β -unsaturated carbonyl group is a prerequisite. *Carcinogenesis*, 23: 795-802.
- Murakami, A., Miyamoto, M., & Ohigashi, H. (2004). Zerumbone, an anti-inflammatory phytochemical, induces expression of proinflammatory cytokine genes in human colon adenocarcinoma cell lines. *Biofactors*, 21: 95-101.
- Nadege, B., Patrick, L., & Rodrigue, R. (2009). Mitochondria: from bioenergetics to the metabolic regulation of carcinogenesis. *Frontiers in Bioscience*, 14: 4015-4034.
- Nagata, S., Nagase, H., Kawane, K., Mukae, N., & Fukuyama, H. (2003). Degradation of chromosomal DNA during apoptosis. *Cell Death and Differentiation*, 10: 108-116.
- Nakamura, Y., Yoshida, C., Murakami, A., Ohigashi, H., Osawa, T., & Uchida, K. (2004). Zerumbone, a tropical ginger sesquiterpene, activates phase II drug metabolizing enzymes. *Federation of European Biochemical Societies*, 572: 245–250.
- Namiki, Y., Fuchigami, T., Tada, N., Kawamura, R., Matsunuma, S., Kitamoto, Y., & Nakagawa, M. (2011). Nanomedicine for cancer: lipid-based nanostructures for drug delivery and monitoring. *Accounts of Chemical Research*, 44: 1080-1093.
- Newmeyer, D. D., Bossy-Wetzler, E., Kluck, R. M., Wolf, B. B., Beere, H. M., & Green, D. R. (2000). Bcl-xL does not inhibit the function of Apaf-1. *Cell Death and Differentiation*, 7: 402–407.
- O'Donovan, N., Crown, J., & Stunell, H. (2003). Caspase 3 in Breast Cancer. *Clinical Cancer Research*. 9: 738-742.
- Pardeike, J., Hommoss, A., & Müller, R. H. (2009). Lipid nanoparticles (SLN, NLC) in cosmetic and pharmaceutical dermal products. *International Journal of Pharmaceutics*, 366: 170–84.
- Parkin, D. (2001). Global cancer statistics in the year 2000. *The Lancet Oncology*, 2: 533–543.

- Patidar, A., & Thakur, D. (2010). A REVIEW ON NOVEL LIPID BASED NANOCARRIERS. *International Journal of Pharmacy and Pharmaceutical Sciences*, 2: 30–35.
- Phipps, A. I., & Li, C. I. (2010). Breast Cancer Biology and Clinical Characteristics. In *Breast cancer epidemiology*, ed: Li, C. I. pp. 21-47. Springer.
- Pietenpol, J. A., & Stewart, Z. A. (2002). Cell cycle checkpoint signaling: cell cycle arrest versus apoptosis. *Toxicology*, 182: 475–481.
- Pribitkin, A. E. (2005). Herbal Medicine and Surgery. *Seminars in Integrative Medicine*, 3: 17–23.
- Proskuryakov, S. Y. ., Konoplyannikov, A. G., & Gabai, V. L. (2003). Necrosis: a specific form of programmed cell death. *Experimental Cell Research*, 283: 1–16.
- Quinn, J. E., Kennedy, R. D., Mullan, P. B., Gilmore, P. M., Carty, M., Johnston, P. G., & Harkin, D. P. (2003). BRCA1 Functions as a Differential Modulator of BRCA1 Functions as a Differential Modulator of Chemotherapy-induced Apoptosis 1. *Cancer Research*, 63: 6221–6228.
- Rahman, H. S., Rasedee, A., How, C. W., Abdul, A. B., Zeenathul, N. A., Othman, H. H., Seed, M. I., & Yeap, S. K. (2013). Zerumbone-loaded nanostructured lipid carriers: preparation, characterization, and antileukemic effect. *International Journal of Nanomedicine*, 8: 2769–2781.
- Rahman, H. S., Rasedee, A., Abdul, A. B., Zeenathul, N. A., Othman, H. H., Yeap, S. K., & Ghani, W. N. H. A. (2014). Zerumbone-loaded nanostructured lipid carrier induces G2/M cell cycle arrest and apoptosis via mitochondrial pathway in a human lymphoblastic leukemia cell line. *International Journal of Nanomedicine*, 9: 1–12.
- Rahman, M. A., Sultan, M. T., & Islam, M. R. (2012). Apoptosis and cancer: insights molecular mechanisms and treatments. *International Journal of Biomolecule and Biomedicine*, 2: 1-16.
- Rai, N. K., Tripathi, K., Sharma, D., & Shukla, V. K. (2005). Apoptosis: a basic physiologic process in wound healing. *The International of Journal of Lower Extremity Wounds*. 4: 138–44.
- Rakha, E. a, Reis-Filho, J. S., & Ellis, I. O. (2008). Basal-like breast cancer: a critical review. *Journal of Clinical Oncology: Official Journal of the American Society of Clinical Oncology*, 26: 2568–2581.
- Roy, A., Baliga, M., & Katiyar, S. (2005). Epigallocatechin-3-gallate induces apoptosis in estrogen receptor-negative human breast carcinoma cells via modulation in protein expression of p53 and Bax and caspase-3 activation. *Molecular Cancer Therapeutics*, 4: 81–90.

- Ruslay, S., Abas, F., Shaari, K., Zainal, Z., Sirat, H., Israfi, D. A., & Lajis, N. H. (2007). Characterization of the components present in the active fractions of health gingers (*Curcuma xanthorrhiza* and *Zingiber zerumbet*) by HPLC–DAD–ESIMS. *Food Chemistry*, 104: 1183-1191.
- Sabu, M. (2003). Revision of the genus *Zingiber* in South India. *Folia Malaysiana*, 4: 25-52.
- Sahu, M. K., Soni, G. C., & Prajapati, S. K. (2012). Formulation and characterization of topical nanostructured lipid carrier gel of flurbiprofen and its composition with micellar gel preparation. *World Journal of Pharmaceutical Sciences*, 1: 1235-1247.
- Sakinah, S. A., Handayani, S. T., & Hawariah, L. P. (2007). Zerumbone induced apoptosis in liver cancer cells via modulation of Bax/Bcl-2 ratio. *Cancer Cell International*, 7: 1-11.
- Sarkar, F. H., & Li, Y. (2004). Cell signaling pathways altered by natural chemopreventive agents. *Mutation Research*, 555: 53–64.
- Savjani, K. T., Gajjar, A. K., and Savjani, J. K. (2012). Drug solubility: importance and enhancement techniques. *International Scholarly Research Notices Pharmaceutics*, 2012: 195727.
- Schiller, M., Bekeredjian-Ding, I., Heyder, P., Blank, N., Ho, A. D., & Lorenz, H. M. (2007). Autoantigens are translocated into small apoptotic bodies during early stages of apoptosis. *Cell Death Differentiation*, 15: 183-191.
- Schimmer, A. D. (2004). Inhibitor of apoptosis proteins: translating basic knowledge into clinical practice. *Cancer Research*, 64: 7183–7190.
- Seenivasan, A., Panda, T., & Théodore, T. (2011). Lovastatin Nanoparticle Synthesis and Characterization for Better Drug Delivery. *Open Biotechnology Journal*, 6: 28-32.
- Sehrawat, A., Arlotti, J. a, Murakami, A., & Singh, S. V. (2012). Zerumbone causes Bax- and Bak-mediated apoptosis in human breast cancer cells and inhibits orthotopic xenograft growth in vivo. *Breast cancer research and treatment*, 136: 429–441.
- Selvamuthukumar, S., & Velmurugan, R. (2012). Nanostructured Lipid Carriers: A potential drug carrier for cancer chemotherapy. *Lipids in Health and Disease*, 11: 1-8.
- Singh, C. B., Nongalleima, K., Brojendrosingh, S., Ningombam, S., Lokendrajit, N., & Singh, L. W. (2011). Biological and chemical properties of *Zingiber zerumbet* Smith: a review. *Phytochemistry Reviews*, 11: 113–125.
- Skommer, J., Wlodkowic, D., & Deptala, A. (2007). Larger than life: Mitochondria and the Bcl-2 family. *Leukemia Research*, 31: 277–286.

- Smith, M., & Boon, H. S. (1999). Counseling cancer patients about herbal medicine. *Patient Education and Counseling*, 38: 109–120.
- Somchit, M. N., Mak, J. H., Ahmad Bustamam, A., Zuraini, A., Arifah, A. K., Adam, Y., & Zakaria, Z. A. (2012). Zerumbone isolated from *Zingiber zerumbet* inhibits inflammation and pain in rats. *Journal of Medicinal Plant Research*, 6: 177-180.
- Souto, E. B., Wissing, S. A., Barbosa, C. M., & Müller, R. H. (2004). Development of a controlled release formulation based on SLN and NLC for topical clotrimazole delivery. *International Journal of Pharmaceutics*, 278: 71-77.
- Spiteri, M., Attard, E., Serracino-inglott, A., & Azzopardi, L. M. (2013). Compilation of a herbal medicine formulary for herbal substances in Malta and its usefulness amongst healthcare professionals. *Journal of Young Pharmacists*, 5: 22–25.
- Sporn, M. B., & Suh, N. (2000). Chemoprevention of cancer. *Carcinogenesis*, 21: 525–230.
- Sørli T, Perou CM, Tibshirani R, Aas, T., Geisler, S., Johnsen, H., Hastie, T., Elsen, M. B., Jeffrey, S. S., Thorsen, T., Quist, H., MAtese, J. C., Brown, P. O., & Botstein, D. (2001). Gene expression patterns of breast carcinomas distinguish tumor subclasses with clinical implications. *Proceeding of the National Academy of Sciences*. 98: 10869-10874.
- Srujana, T. S., Babu, K. R., & Rao, B. S. S. (2012). Phytochemical Investigation and Biological Activity of Leaves Extract of Plant *Boswellia Serrata*. *The Pharma Innovation*, 1: 25-52.
- Sulaiman, M. R., Perimal, E. K., Zakaria, Z. A, Mokhtar, F., Akhtar, M. N., Lajis, N. H., & Israf, D. A. (2009). Preliminary analysis of the antinociceptive activity of zerumbone. *Fitoterapia*, 80: 230–232.
- Sulaiman, M. R., TengkuMohamad, T. A. S., Shaik Mossadeq, W. M., Moin, S., Yusof, M., Mokhtar, A. F., Zakaria, Z. A., Israf, D. A., & Lajis, N. (2010). Antinociceptive activity of the essential oil of *Zingiber zerumbet*. *Planta Medica*, 762: 107–112.
- Sun, Y., Sheng, Q., Cheng, Y., Xu, Y., Han, Y., Wang, J., Shi, L., Zhao, H., & Cu, C. (2013). Zerumbone induces apoptosis in human renal cell carcinoma via Gli-1/Bcl-2 pathway. *Die Pharmazie - An International Journal of Pharmaceutical Sciences*, 5: 141-145.
- Surh, Y.-J. (2003). Cancer chemoprevention with dietary phytochemicals. *Nature Reviews Cancer*, 3: 768–780.
- Susin, S. a., Daugas, E., Ravagnan, L., Samejima, K., Zamzami, N., Loeffler, M., Costantini, P., Ferri, K. F., Mak, T. W., Earnshaw, W. C., & Kroemer, G.

- (2000). Two Distinct Pathways Leading to Nuclear Apoptosis. *Journal of Experimental Medicine*, 192: 571–580.
- Syam, S., Abdul, A. B., Sukari, M. A., Mohan, S., Abdelwahab, S. I., & Wah, T. S. (2011). The growth suppressing effects of girinimbine on HepG2 involve induction of apoptosis and cell cycle arrest. *Molecules*, 16: 7155-7170.
- Takada, Y., Andreeff, M., & Aggarwal, B. B. (2005). Indole-3-carbinol suppresses NF-kappaB and IkappaBalpha kinase activation, causing inhibition of expression of NF-kappaB-regulated antiapoptotic and metastatic gene products and enhancement of apoptosis in myeloid and leukemia cells. *Blood*, 106: 641–649.
- Tiwari, M. (2012). Nano cancer therapy strategies. *Journal of Cancer Research and Therapeutics*, 8: 19-22.
- Varier, N. S. (1944). Chemical examination of the rhizomes of *Zingiber zerumbet*, Smith. *Proceedings of the Indian Academy of Sciences A*, 20: 257–260.
- Velmurugan, R., Selvamuthukumar, S., & Manavalan, R. (2011). Multi criteria decision making to select the suitable method for the preparation of nanoparticles using an analytical hierarchy process. *Die Pharmazie*, 66:836–842.
- Vemula, V. R., Lagishetty, V., & Lingala, S. (2010). “Solubility enhancement techniques”. *International Journal of Pharmaceutical Sciences Review and Research*, 5: 41–51.
- Vimala, S., Norhanom, A. W., & Yadav, M. (1999). Anti-tumour promoter activity in Malaysian ginger rhizobia used in traditional medicine. *British Journal of Cancer*, 80: 110.
- Vogel, V. G., Costantino, J. P., Wickerham, D. L., Cronin, W. M., & Wolmark, N. (2002). The study of tamoxifen and raloxifene: preliminary enrollment data from a randomized breast cancer risk reduction trial. *Clinical Breast Cancer*, 3:153–159.
- Walsh, J. G., Cullen, S. P., Sheridan, C., Lüthi, A. U., Gerner, C., & Martin, S. J. (2008). Executioner caspase-3 and caspase-7 are functionally distinct proteases. *Proceedings of the National Academy of Sciences of the United States of America*, 105: 12815–12819.
- Wang, C. Y., Cusack, J. C. Jr., Liu, R., & Baldwin, A. S. Jr. (1999). Control of inducible chemoresistance (Enhanced anti-tumor therapy through increased apoptosis by inhibition of NF-κB). *Nature Medicine*. 5: 412–417.

- Weng, H.-Y., Hsu, M.-J., Wang, C.-C., Chen, B.-C., Hong, C.-Y., Chen, M.-C., Chlu, W. T., & Lin, C. H. (2012). Zerumbone suppresses IKK α , Akt, and FOXO1 activation, resulting in apoptosis of GBM 8401 cells. *Journal of Biomedical Science*, 19: 1-11.
- Willson, A. (2012). AO/PI Viability. Innovation and Expertise in the Science of Cell Counting. Available from: <http://www.nexcelom.com/Nexcelom-Blog/aopi-viability/> [Accessed on 15 April 2014].
- Witko-Sarsat, V., Mocek, J., Bouayad, D., Tamassia, N., Ribeil, J.-A., Candalh, C., Davezac, N., Reuter, N., Mouthon, L., Hermine, O., & Cassatelli, M. A. (2010). Proliferating cell nuclear antigen acts as a cytoplasmic platform controlling human neutrophil survival. *The Journal of Experimental Medicine*, 207: 2631–2645.
- Xian, M., Ito, K., Nakazato, T., Shimizu, T., Chen, C. K., Yamato, K., Murakami, A., Ohigashi, H., Ikeda, Y., & Kizaki, M. (2007). Zerumbone, a bioactive sesquiterpene, induces G2/M cell cycle arrest and apoptosis in leukemia cells via a Fas and mitochondria mediated pathway. *Cancer Science*, 98: 118-126.
- Yang, F., Oz, H. S., Barve, S., de Villiers, W. J., McClain, C. J., & Varilek, G. W. (2001). The green tea polyphenol (-)-epigallocatechin-3-gallate blocks nuclear factor-kappa B activation by inhibiting I kappa B kinase activity in the intestinal epithelial cell line IEC-6. *Molecular Pharmacology*, 60: 528–33.
- Yang S, Zhu J, Lu Y, Liang, B., & Yang, C. (1999). Body Distribution of camptothecin solid lipid nanoparticles after oral administration. *Pharmaceutical Research*, 16:751–757.
- Yip, C., Taib, N., & Mohamed, I. (2006). Epidemiology of Breast Cancer in Malaysia. *Asian Pacific Journal of Cancer*, 7: 369–374.
- Yob, N., Jofry, S. M., Affandi, M., Teh, L., Salleh, M., & Zakaria, Z. (2011). *Zingiber zerumbet* (L.) Smith: a review of its ethnomedicinal, chemical, and pharmacological uses. *Evidence-Based Complementary and Alternative Medicine*, 2011, 12 pages.
- Yu, J., & Zhang, L. (2008). PUMA, a potent killer with or without p53. *Oncogene*, 27: 1–22.
- Yun, J. M., Kwon, H., Mukhtar, H., & Hwang, J. K. (2005). Induction of apoptosis by Panduratin A isolated from *Kaempferia pandurata* in human colon cancer HT-29 cells. *Planta Medica-Natural Products and Medicinal Plant Research*, 71: 501–507.

- Yun, J. M., Kweon, M. H., Kwon, H., Hwang, J. K., & Mukhtar, H. (2006). Induction of apoptosis and cell cycle arrest by a chalconepanduratin A isolated from *Kaempferia pandurata* in androgen-independent human prostate cancer cells PC3 and DU145. *Carcinogenesis*, 27: 1454–1464.
- Zainal, A., Zainudin, M., & Saleha, I. N. (2006). *Malaysian cancer statistics-data and figure peninsular Malaysia 2006*. National Cancer Registry. Malaysia.
- Zainal, A. O., & Nor Saleha, I. T. (2011). National Cancer Registry Report 2007. *Malaysia: Ministry of Health*.
- Zakaria, Z. A., Mohamad, A. S., Chear, C. T., Wong, Y. Y., Israf, D. A., & Sulaiman, M. R. (2010). Antiinflammatory and antinociceptive activities of *Zingiber zerumbet* methanol extract in experimental model systems. *Medical Principles and Practice*, 19: 287–294.
- Zepeda-castilla, E. J., Recinos-money, E., Cuéllar-hubbe, M., Robles-vidal, C. D., & Maafs-molina, E. (2008). Molecular classification of breast cancer. *Cirugia y Cirujanos*, 76: 87–93.
- Zhao, H., Lo, Y.-H., Ma, L., Waltz, S. E., Gray, J. K., Hung, M.-C., & Wang, S.-C. (2011). Targeting tyrosine phosphorylation of PCNA inhibits prostate cancer growth. *Molecular Cancer Therapeutics*, 10: 29–36.