



UNIVERSITI PUTRA MALAYSIA

***APOPTOTIC-RELATED SIGNALLING PATHWAYS IN MCF-7 CELLS  
TREATED WITH ETHYL ACETATE EXTRACT OF *Dillenia suffruticosa*,  
AND ISOLATION OF ITS MAJOR COMPOUNDS***

**TOR YIN SIM**

**IB 2015 16**



**APOPTOTIC-RELATED SIGNALLING PATHWAYS IN MCF-7 CELLS  
TREATED WITH ETHYL ACETATE EXTRACT OF *Dillenia suffruticosa*,  
AND ISOLATION OF ITS MAJOR COMPOUNDS**

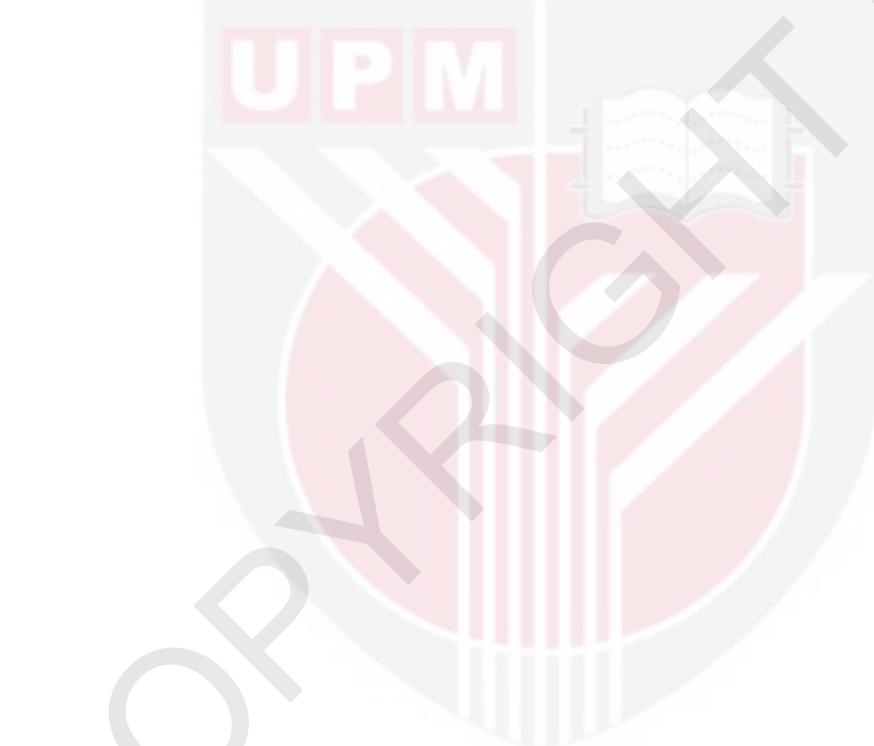
By

**TOR YIN SIM**



**Thesis Submitted to the School of Graduate Studies,  
Universiti Putra Malaysia, in Fulfillment of the Requirements for the  
Degree of Doctor of Philosophy**

**July 2015**



© COPYRIGHT UPM

## **COPYRIGHT**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia





Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of  
the requirements for the degree of Doctor of Philosophy

**APOPTOTIC-RELATED SIGNALLING PATHWAYS IN MCF-7 CELLS  
TREATED WITH ETHYL ACETATE EXTRACT OF *Dillenia suffruticosa*,  
AND ISOLATION OF ITS MAJOR COMPOUNDS**

By

**TOR YIN SIM**

**July 2015**

**Chairman : Latifah Saiful Yazan, PhD**

**Faculty : Institute of Bioscience**

Breast cancer is the most prevalent cancer among women worldwide. The trend for breast cancer treatment has shifted towards the use of natural product such as herbal medicine as an alternative and complementary medicine. *Dillenia suffruticosa* (Griff) Martelli that belongs to the family Dilleniaceae has been traditionally used to treat cancerous growth. In this study, the anti-cancer activity of ethyl acetate extract of *D. suffruticosa* (EADs) root was examined on breast cancer cells, MCF-7. EADs was prepared from the root of *D. suffruticosa* by using sequential solvent extraction. MTT assay was used to determine the cytotoxicity of EADs, which was demonstrated to be dose- and time-dependent, with  $IC_{50}$  of  $39 \pm 3.6 \mu\text{g/mL}$  at 72 hours. Flow cytometry cell cycle analysis displayed that EADs induced non-phase specific cell cycle arrest. EADs induced mainly apoptosis in MCF-7 cells in Annexin-FITC/PI analysis. The use of general caspase-inhibitor Z-VAD-FMK indicated that EADs-induced apoptosis was caspase-independent. EADs was found to promote oxidative stress that will lead to cell death because the pre-treatment with antioxidants  $\alpha$ -tocopherol and ascorbic acid significantly reduced the cytotoxicity of the extract ( $P<0.05$ ). DCFH-DA assay revealed that treatment with EADs attenuated the generation of intracellular ROS. The use of JC-1 dye reflected that EADs caused disruption in the mitochondrial membrane potential. Up-regulation of p53 and p21, is believed has led to EADs-induced non-phase specific cell cycle arrest ( $P<0.05$ ). Elevation of Bax/Bcl-2 ratio and the depolarization of mitochondrial membrane potential indicated that EADs-induced apoptosis was mitochondrial-dependent. The expression of oxidative stress-related proteins AKT, p-AKT, ERK, and p-ERK was downregulated with upregulation of JNK and p-JNK suggesting that induction of apoptosis by EADs is mediated by inhibition of AKT and ERK, and activation of JNK. The major compounds of EADs were then isolated using column chromatography and elucidated using nuclear magnetic resonance analysis producing a total of 6 compounds. The cytotoxicity of the isolated compound was determined using MTT assay. Gallic acid was found to be most cytotoxic against MCF-7 cell line compared to others, with  $IC_{50}$  of  $36 \pm 1.7 \mu\text{g/mL}$  ( $P<0.05$ ). In summary, EADs induced cell cycle arrest, oxidative stress and apoptosis in MCF-7 cells. Thus, EADs has the potential to be developed as an anti-cancer agent against breast cancer.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai  
memenuhi keperluan ijazah Doktor Falsafah

**LALUAN ISYARAT BERKAITAN APOPTOSIS DALAM SEL KANSER  
PAYUDARA MCF-7 DIRAWAT DENGAN EKSTRAK ETIL ASETAT *Dillenia suffruticosa*, DAN PENGASINGAN SEBATIAN UTAMANYA**

Oleh

**TOR YIN SIM**

**Julai 2015**

**Pengerusi** : **Latifah Saiful Yazan, PhD**

**Fakulti** : **Institut Biosains**

Kanser payudara adalah kanser paling lazim di kalangan wanita seluruh dunia. Kaedah rawatan kanser payudara telah beralih kepada penggunaan hasilan semulajadi seperti perubatan herba sebagai perubatan alternatif dan pelengkap. *Dillenia suffruticosa* (Griff Martelli) yang berasal dari keluarga Dilleniaceae telah digunakan untuk merawat pertumbuhan kanser secara tradisional. Dalam kajian ini, ciri-ciri anti-kanser ekstrak etil asetat akar *D. suffruticosa* (EADs) terhadap sel payudara, MCF-7, dinilai. EADs disediakan daripada akar *D. suffruticosa* menggunakan pengekstrakan berurutan pelarut. Asai MTT digunakan untuk menentukan kesitotoksikan EADs, yang ditunjukkan bersandar kepada dos dan masa, dengan  $IC_{50} = 39 \pm 3.6 \mu\text{g/mL}$  pada 72 jam. Analisis sitometri kitaran sel menunjukkan bahawa EADs mengaruh penahanan fasa tak spesifik kitaran sel. Mengikut analisa sitometri annexin-FITC/PI, EADs mengaruh apoptosis terhadap sel MCF-7. Penggunaan perencat umum caspase Z-VAD-FMK menunjukkan pengaruhan apoptosis oleh EADs adalah tidak bersandar kepada caspase. EADs didapati menggalakkan tekanan oksidatif yang menyebabkan kematian sel. Asai DCFH-DA mendedahkan bahawa rawatan dengan EADs melemahkan penjanaan spesies oksigen reaktif intrasel. Penggunaan JC-1 menunjukkan EADs mengakibatkan gangguan kepada potensi membran mitokondria. Peningkatan dalam pengekspresan p53 dan p21 dipercayai telah mendorong kepada penahanan fasa tidak spesifik kitaran sel aruhan EADs ( $P < 0.05$ ). Peningkatan nisbah Bax/Bcl-2 dan penyahkutuhan potensi membran mitokondria menunjukkan apoptosis aruhan EADs adalah bersandar kepada mitokondria. Pengekspresan protein yang berkaitan dengan tekanan oksidatif AKT, p-AKT, ERK, dan p-ERK yang diturunkan berikutan dengan peningkatan JNK dan p-JNK mencadangkan bahawa aruhan oleh EADs disebabkan oleh perencatan AKT dan ERK, dan pengaktifan JNK. Pengasingan sebatian utama EADs dijalankan dengan menggunakan kromatografi turus dan dikenalpasti dengan analisa resonans magnetik nuclear menghasilkan sejumlah enam sebatian. Kesitotoksikan sebatian yang diasangkan ditentukan dengan asai MTT. Asid galik adalah paling sitotoksik kepada sel MCF-7 berbanding dengan sebatian lain, dengan  $IC_{50} = 36 \pm 1.7 \mu\text{g/mL}$  ( $P < 0.05$ ). Secara ringkas, EADs mengaruh penahanan kitaran sel, tekanan oksidatif and apoptosis melalui pengawalan pelbagai gen dan protein yang terlibat dalam laluan isyarat apoptosis. Justeru itu, EADs mempunyai potensi dijadikan agen anti-kanser terhadap kanser payudara.

## **ACKNOWLEDGEMENT**

The completion of this process of learning is a joint effort from various groups of individual. I would like to take this opportunity to express my deepest appreciation to all who has supported and assisted me throughout my study. Without their undivided help, I would not be able to successfully accomplish this project.

In the first place, I would like to express my heartiest gratitude to my supervisor, Associate Professor Dr. Latifah Saiful Yazan for being such a dedicated mentor and is gracious enough to guide me along the whole process in completing my research. She is simply the best for being there whenever I have any doubts, offering time and valuable knowledge in her area of professional practices. Not to mention, my warm and sincere gratitude to my co-supervisors, Professor Rasedee Abdullah and Associate Professor Dr. Cheah Yoke Kqueen for their encouragement and constructive advises that lead to the success of this research.

I would like to extend my great appreciation to all staff including Mrs. Siti Muskinah, Ms. Norsharina, Mr. Chan Kim Wei, Ms. Norhayati, Mr. Abidin, Mrs. Mastura, Ms. Siti Aisyah, Mrs. Lina, Mrs. Nancy, Mrs. Norhafiza, Ms. Normahfuzah, Dr. Tan Sheau Wei and Dr. Yeap Swee Keong from the Laboratory of Molecular Biomedicine and the Laboratory of Vaccine Immunotherapeutic, Institute of Bioscience, Universiti Putra Malaysia; Mrs. Tommini and Mrs. Norlela from Laboratory of Cancer Research MAKNA, Institute of Bioscience, Universiti Putra Malaysia; Mrs. Ayuni and Mrs. Norhidayu from the Administration Office, Institute of Bioscience, Universiti Putra Malaysia; Mrs. Marsitah from the Department of Pathology, Faculty of Medicine and Health Sciences, Universiti Putra Malaysia; Mrs. Shareena from the Department of Chemistry, Faculty of Science, Universiti Putra Malaysia. Thank you for the technical assistance and advices.

Special thanks go to my lab partners and friends, Ms. Armania, Mrs. Aina, Mr. Foo Jhi Biau, Mr. Ng Wei Keat, Dr. How Chee Wun, Dr. Agustono Wibowo, Mrs. Wan Nor Hafiza, Mr. Tan Boon Keat, Ms. Ong Yong Sze, Mr. Ooi Der Jiun, Mrs. How Zhi Ping, Ms. Yap Huan Yong, Mrs. Kavitha, Mr. Kuan Wen Bin, Ms. Hanisah, Ms. Sarega, Mr. Hisyam and Mrs. Noreen for their support, and for sharing the moment of joy and hardship with me.

I would also like to acknowledge the financial support from Ministry of Education Malaysia for the Fundamental Research Grant Scheme (FRGS) and MyBrain15 (MyPhD) Scholarship for my study.

Last but not least, my heartfelt gratitude goes to my parents and my family members who shower me with their unconditional love and encouragement when I need them most.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirements for the degree of **Doctor of Philosophy**. The members of the Supervisory Committee were as follows:

**Latifah Saiful Yazan, PhD**

Associate Professor

Faculty of Medicine and Health Sciences

Universiti Putra Malaysia

(Chairman)

**Rasedee Abdullah, PhD**

Professor

Faculty of Veterinary Medicine

Universiti Putra Malaysia

(Member)

**Cheah Yoke Kqueen, PhD**

Associate Professor

Faculty of Medicine and Health Sciences

Universiti Putra Malaysia

(Member)

---

**BUJANG BIN KIM HUAT, PhD**

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date:

### **Declaration by graduate student**

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced; this thesis has not been submitted previously or concurrently for any other degree at any institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by university Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/ fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_ Date: 9 July 2015

Name and Matric No.: Tor Yin Sim (GS30894)

## **Declaration by Members of Supervisory Committee**

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature : \_\_\_\_\_

Name of Chairman of  
Supervisory  
Committee

: Assoc. Prof. Dr. Latifah Saiful Yazan

Signature : \_\_\_\_\_

Name of Chairman of  
Supervisory  
Committee

: Prof. Dr. Rasedee Abdullah

Signature : \_\_\_\_\_

Name of Chairman of  
Supervisory  
Committee

: Assoc. Prof. Dr. Cheah Yoke Kqueen

## TABLE OF CONTENTS

	<b>Page</b>
<b>ABSTRACT</b>	i
<b>ABSTRAK</b>	ii
<b>ACKNOWLEDGEMENTS</b>	iii
<b>APPROVAL</b>	iv
<b>DECLARATION</b>	vi
<b>LIST OF TABLES</b>	xii
<b>LIST OF FIGURES</b>	xiv
<b>LIST OF APPENDICES</b>	xv
<b>LIST OF ABBREVIATIONS</b>	xvi
 <b>CHAPTER</b>	
<b>1. INTRODUCTION</b>	1
<b>2. LITERATURE REVIEW</b>	5
2.1 Cancer	5
2.2 Normal breast	6
2.3 Breast cancer	7
2.3.1 Definition and type	7
2.3.2 Epidemiology	8
2.3.3 Classification of breast cancer	9
2.3.3.1 TNM classification	9
2.3.3.2 Molecular classification	13
2.3.4 Treatment of breast cancer	17
2.4 Natural products	19
2.4.1 Plant as a source of anticancer agent	19
2.4.2 Plant phytochemicals	22
2.4.2.1 Flavonoid	22
2.4.2.2 Phenolic acid	23
2.4.2.3 Triterpenoid	24
2.4.2.4 Phytosterol	24
2.4.3 Extraction, isolation and characterization of compound from plant	25
2.4.4 Challenges of plants as therapeutics agents	26
2.5 Cell death	28
2.5.1 Necrosis	28
2.5.2 Apoptosis	28
2.5.2.1 Apoptotic pathway	30
2.5.2.2 Apoptosis and cancer	32
2.6 Cell cycle and cancer	33
2.7 Oxidative stress and cancer	35
2.8 Cancer signaling pathway	37
2.8.1 MAPK signalling pathway	37
2.8.1.1 JNK signalling pathway	37
2.8.1.2 ERK signalling pathway	38
2.8.1.3 p38 MAPK signalling pathway	39
2.8.2 AKT signalling pathway	40
2.8.3 NF-κB signalling pathway	41

2.9	Genus <i>Dillenia</i>	41
2.9.1	<i>Dillenia suffruticosa</i>	45
<b>3.</b>	<b>DETERMINATION OF CYTOTOXICITY AND MODE OF CELL DEATH INDUCED BY ETHYL ACETATE EXTRACT OF <i>Dillenia suffruticosa</i> IN MCF-7 CELLS</b>	47
3.1	Introduction	47
3.2	Materials and Methods	48
3.2.1	Plant material	48
3.2.2	Chemical	48
3.2.3	Cell	49
3.2.4	Plant extraction	49
3.2.5	Sample preparation	50
3.2.6	Determination of cytotoxicity	51
3.2.7	Morphological analysis	51
3.2.8	Cell cycle analysis	51
3.2.9	Determination of mode of cell death	52
3.2.10	Statistical analysis	52
3.3	Results and discussion	53
3.3.1	Cytotoxicity of EADs	53
3.3.2	Cell cycle arrest in MCF-7 cells by EADs	56
3.3.3	Induction of apoptosis in MCF-7 cells by EADs	60
3.4	Conclusion	65
<b>4.</b>	<b>OXIDATIVE STRESS-INDUCED APOPTOSIS BY ETHYL ACETATE EXTRACT OF <i>Dillenia suffruticosa</i> IN MCF-7 CELLS</b>	67
4.1	Introduction	67
4.2	Materials and methods	68
4.2.1	Plant material	68
4.2.2	Chemical	68
4.2.3	Cell	68
4.2.4	Sample preparation	68
4.2.5	Evaluation of the effects of antioxidants on EADs-treated MCF-7 cells	69
4.2.6	Detection of ROS in EADs-treated MCF-7 cells	69
4.2.7	Determination of the effects of EADs on the expression of intracellular antioxidant enzymes genes of MCF-7 cells	69
4.2.7.1	RNA isolation	72
4.2.7.2	Reverse transcription and polymerase chain reaction	72
4.2.7.3	GeXP multiplex analysis	72
4.2.8	Statistical analysis	72
4.3	Results and discussion	72
4.3.1	Effects of antioxidants on EADs-treated MCF-7 cells	72
4.3.2	Level of reactive species in EADs-treated MCF-7 cells	75

4.3.3 Determination of the expression of intracellular enzymatic antioxidant genes in EADs-treated MCF-7 cells	77
4.4 Conclusion	79
<b>5. ELUCIDATION OF SIGNALLING PATHWAYS INVOLVED IN MCF-7 CELLS TREATED WITH ETHYL ACETATE EXTRACT OF <i>Dillenia suffruticosa</i></b>	81
5.1 Introduction	81
5.2 Materials and methods	82
5.2.1 Plant material	82
5.2.2 Chemical	82
5.2.3 Cell	83
5.2.4 Determination of the involvement of caspases in EADs-treated MCF-7 cells	83
5.2.5 Determination of the effect of EADs on mitochondrial membrane potential	84
5.2.6 Determination of the effects of EADs on apoptotic-related genes in EADs-treated MCF-7 cells	84
5.2.6.1 RNA isolation	84
5.2.6.2 Reverse transcription and polymerase chain reaction	84
5.2.6.3 GeXP multiplex analysis	84
5.2.7 Determination of the effects of EADs on apoptotic related proteins in EADs-treated MCF-7 cells	86
5.2.7.1 Preparation of protein lysates	86
5.2.7.2 Polyacrylamide gel preparation	86
5.2.7.3 Protein loading and gel electrophoresis	86
5.2.7.4 Transfer of protein from gel to membrane	87
5.2.7.5 Blocking and incubation with antibodies	87
5.2.7.6 Detection	87
5.2.8 Statistical analysis	88
5.3 Results and discussion	88
5.3.1 Involvement of caspases in EADs-treated MCF-7 cells	88
5.3.2 Depolarization of mitochondrial membrane potential in MCF-7 cells treated with EADs	92
5.3.3 Effects of EADs on the apoptotic related genes and proteins in EADs-treated MCF-7 cells	94
5.4 Conclusion	102
<b>6. ISOLATION AND CYTOTOXICITY OF THE ISOLATED COMPOUNDS FROM ETHYL ACETATE EXTRACT OF <i>Dillenia suffruticosa</i></b>	103
6.1 Introduction	103
6.2 Material and methods	104
6.2.1 Plant material	104
6.2.2 Material	104
6.2.3 Extraction of plant	104

6.2.4 Extraction, isolation and purification of the isolated compounds from EADs	104
6.2.4.1 Compound 1	105
6.2.4.2 Compound 2	105
6.2.4.3 Compound 3	105
6.2.4.4 Compound 4	105
6.2.4.5 Compound 5	106
6.2.4.6 Compound 6	106
6.2.5 Elucidation of structure	108
6.2.6 Determination of cytotoxicity of the isolated compounds	108
6.2.6.1 Cell	108
6.2.6.2 Sample preparation	108
6.2.6.3 Determination of the cytotoxicity of isolated compounds	108
6.2.7 Statistical analysis	109
6.3 Results and discussion	109
6.3.1 Yield of fractions and compounds	109
6.3.2 Elucidation of the structure of isolated compounds	110
6.3.3 Cytotoxicity of the isolated compounds from EADs	113
6.4 Conclusion	115
<b>7. GENERAL DISCUSSION, CONCLUSION AND RECOMMENDATION FOR FUTURE RESEARCH</b>	<b>117</b>
7.1 General discussion	117
7.2 Conclusion	121
7.3 Recommendation for future research	122
<b>REFERENCES</b>	<b>123</b>
<b>APPENDICES</b>	<b>167</b>
<b>BIODATA OF STUDENT</b>	<b>187</b>
<b>LIST OF PUBLICATIONS</b>	<b>188</b>

## LIST OF TABLES

<b>Table</b>		<b>Page</b>
2.1	TNM classification for breast cancer	10
2.2	AJCC 7 <sup>th</sup> edition staging for breast cancer	12
2.3	Characteristics of the molecular subtype of breast cancer	16
2.4	Adverse effects caused by certain cancer chemotherapeutic drugs	18
2.5	Plant-derived anticancer agents in clinical use and clinical trials	20
2.6	Chromatography techniques used for the separation of compounds from plant extracts	25
2.7	Comparison between the apoptosis and necrosis	29
2.8	Traditional uses of <i>Dillenia</i> species	42
2.9	Pharmacological activities of <i>Dillenia</i> species	43
2.10	Compounds isolated from <i>Dillenia</i> species.	44
2.11	Traditional uses and pharmacological activites of <i>Dillenia suffruticosa</i>	46
3.1	Cytotoxicity of EADs towards breast cancer and non-breast cancer cell lines as determined using MTT assay at different time point reflected as IC <sub>50</sub> value	55
3.2	Cytotoxicity of EADs and tamoxifen towards MCF-7 cells as determined using MTT assay at different time point reflected as IC <sub>50</sub> value	56
4.1	List of genes with their respective primers and the accession number for GeXP multiplex analysis	71
5.1	List of genes with the respective primers and the accession number for GeXP multiplex analysis	85
6.1	Eight major fractions from the fractionation of EADs and the yield	109
6.2	Six isolated compounds from EADs and the yield	109
6.3	Cytotoxicity of the six compounds isolated from EADs on MCF-7 and MDA-MB-231 cells as determined by MTT assay reflected by the IC <sub>50</sub> value	113

## LIST OF FIGURES

<b>Figure</b>		<b>Page</b>
2.1	Incidence of the ten most common diagnosed cancer cases worldwide in 2012	5
2.2	Structure of the normal breast	6
2.3	Chemical structure of flavonoids	23
2.4	Chemical structure of phenolic acids	23
2.5	Chemical structure of phytosterols	24
2.6	Cell cycle	34
2.7	Flower, fruit and leaf of <i>D. suffruticosa</i>	45
3.1	Schematic diagram of the preparation of EADs by using solvents with increasing polarity in sequential solvent extraction	50
3.2	Viability of MCF-7 cells after treatment with EADs at different time point as determined using MTT assay	55
3.3	Cell cycle profile of MCF-7 cells treated with EADs at different time point	58
3.4	Population of MCF-7 cells at different phase of cell cycle after treatment with EADS for 24 hours (A) and 48 hours (B) as determined using flow cytometry analysis	59
3.5	Population of MCF-7 cells treated with EADs observed under an inverted light microscope at different time point	61
3.6	Morphological changes of MCF-7 cells treated with EADs observed under an inverted light microscope at different time point	62
3.7	Percentage of viable, apoptotic and late apoptotic cells in MCF-7 cells treated with EADs at different time point as determined using Annexin V-PI flow cytometric analysis	64
4.1	Effects of pre-treatment with antioxidants on the viability of MCF-7 cells treated with EADs at 24 hours (A) and 48 hours (B)	74
4.2	Level of ROS in MCF-7 cells after treatment with EADs and H <sub>2</sub> O <sub>2</sub> as determined using DCFH-DA assay	76

4.3	Expression level of the endogenous enzymatic antioxidants <i>SOD1</i> (A) <i>SOD2</i> (B) and <i>catalase</i> (C) in MCF-7 cells treated with EADs as determined using GeXP analysis system	78
5.1	Involvement of caspase in MCF-7 cells treated with EADs as determined by Annexin V/PI flow cytometric analysis	90
5.2	Percentage of apoptotic cells between caspase inhibitor negative and caspase inhibitor positive group in MCF-7 cells treated with EADs	91
5.3	Mitochondria membrane potential ( $\Delta\Psi_m$ ) of MCF-7 cells treated with EADs as determined using JC-1 fluorescent dye	93
5.4	Expression level of the apoptotic-related genes in MCF-7 cells treated with EADs as determined using GeXP analysis system	95
5.5	Expression level of the apoptotic-related proteins in MCF-7 cells treated with EADs at different time point as determined using Western blot	96
5.6	Bax/Bcl-2 ratio in MCF-7 cells treated with EADs at different time point	97
5.7	Expression level of the apoptotic-related proteins in MCF-7 cells treated with EADs at different time point as determined using Western blot	98
6.1	Extraction and isolation of compounds from EADs (Compound <b>1-6</b> )	107
6.2	Chemical structure of the compounds isolated from ethyl acetate extract of <i>Dillenia suffruticosa</i> , which are known as kaempferide ( <b>1</b> ), kaempferol ( <b>2</b> ), protocatechuic acid ( <b>3</b> ), gallic acid ( <b>4</b> ), 3-epimaslinic acid ( <b>5</b> ) and $\beta$ -sitosterol-3-O- $\beta$ -D-glucopyranoside ( <b>6</b> )	110
7.1	Proposed signaling pathway of EADs-induced apoptosis in MCF-7 cells	121

## LIST OF APPENDICES

A	TLC profile of EADs with different solvent systems under UV	167
B	TLC profile of EADs with different solvent systems detected using cerium (IV) sulphate	167
C	TLC profile of the fractions of EADs	167
D1	$^1\text{H}$ -NMR profile of compound 1	168
D2	$^{13}\text{C}$ -NMR profile of compound 1	169
E1	$^1\text{H}$ -NMR profile of compound 2	170
E2	$^{13}\text{C}$ -NMR profile of compound 2	171
F1	$^1\text{H}$ -NMR profile of compound 3	172
F2	$^{13}\text{C}$ -NMR profile of compound 3	173
G1	$^1\text{H}$ -NMR profile of compound 4	174
G2	$^{13}\text{C}$ -NMR profile of compound 4	175
H1	$^1\text{H}$ -NMR profile of compound 5	176
H2	$^{13}\text{C}$ -NMR profile of compound 5	177
I1	$^1\text{H}$ -NMR profile of compound 6	178
I2	$^{13}\text{C}$ -NMR profile of compound 6	179
J	Comparison of $^1\text{H}$ -NMR and $^{13}\text{C}$ -NMR data for compound 1 with the literature	180
K	Comparison of $^1\text{H}$ -NMR and $^{13}\text{C}$ -NMR data for compound 2 with the literature	181
L	Comparison of $^1\text{H}$ -NMR and $^{13}\text{C}$ -NMR data for compound 3 with the literature	181
M	Comparison of $^1\text{H}$ -NMR and $^{13}\text{C}$ -NMR data for compound 4 with the literature	182
N	Comparison of $^1\text{H}$ -NMR and $^{13}\text{C}$ -NMR data for compound 5 with the literature	183
O	Comparison of $^1\text{H}$ -NMR and $^{13}\text{C}$ -NMR data for compound 6 with the literature	184

## LIST OF ABBREVIATIONS

<sup>13</sup> C-NMR	Carbon nuclear magnetic resonance
1D-NMR	One dimensional NMR
<sup>1</sup> H-NMR	Proton nuclear magnetic resonance
2D-NMR	Two-dimensional NMR
AIF	Apoptosis inducing factor
AKT	Protein kinase B
APT	Attached proton test
CAT	Catalase
CDK	Cyclin-dependent kinase
DCFH-DA	Dichlorodihydrofluorescein diacetate
DCIS	Ductal carcinoma <i>in situ</i>
EGFR	Epidermal growth factor receptor
EMT	Epithelial to mesenchymal transition
ER	Oestrogen receptor
ERK	Extracellular signal-regulated kinase
FITC	Fluorescein isothiocyanate
GSH-Px	Glutathione peroxidase
HER-2	Human epidermal growth factor receptor-2
IARC	International Agency for Research on Cancer
IDC	Invasive ductal carcinoma
IKK	IκB kinase
IκB	Inhibitor of κB
JNK	c-Jun N-terminal kinase
MAPK	Mitogen-activated protein kinase
MPT	Membrane permeability transition
MPTP	Mitochondrial permeability transition pores
NF-κB	Nuclear factor-kappa B
NMR	Nuclear magnetic resonance
p38MAPK	p38 mitogen-activated protein kinase
PARP	Poly (ADP-ribose) polymerase
PgR	Progesterone receptor
PKC	Protein kinase C
RCS	Reactive chlorine species
RNS	Reactive nitrogen species
ROS	Reactive oxygen species
SAPK	Stress-activated protein kinase
SOD	Superoxide dismutase
TCM	Traditional Chinese medicine
TNF	Tumour necrosis factor
Z-VAD-FMK	Benzylloxycarbonyl-ValAla-Asp(O-methyl) fluoromethylketone
ΔΨm	Mitochondrial membrane potential

## CHAPTER 1

### INTRODUCTION

The burden of cancer is expanding at an alarming rate. As in 2012, approximately 14.1 million cancer cases and 8.8 million cancer deaths were reported worldwide. It is expected that in coming two decades, the cancer incidence will grow more than 70% (Ferlay *et al.*, 2015). Breast cancer is the most commonly diagnosed life-threatening cancer in women. Breast cancer is heterogeneous and associated with a combination of risk factors that includes hereditary, reproductive factors, hormonal and environmental factors (Althuis *et al.*, 2004; DeBruin and Josephy, 2002). There were approximately 522,000 deaths due to breast cancer in 2012 that ranked as the fifth cause of overall cancer cases (Ferlay *et al.*, 2015).

Apart from surgery, radiotherapy and hormonal therapy, chemotherapy is one of the most common treatments for breast cancer (ACS, 2014). Chemotherapy is a systemic treatment, which refers to the use of one or more anti-cancer drugs circulating throughout the body to eliminate cancer cells. Unfortunately, the treatment will also inevitably affect the surrounding normal cells, thus causing adverse effects to the breast cancer patients (Breakthrough-Breast-Cancer, 2013) that include cardiac toxicity, secondary cancers, hand-foot syndrome, premature menopause, altered cognitive function and neurotoxicity (ACS, 2014; Azim *et al.*, 2011). Thus, alternative medication with alleviated adverse effects is indeed crucial.

Cancer is a complex disease characterized by the alteration in dynamic signaling pathways that modulate cell growth, survival, differentiation and invasion. Occurrence of cancer often involves deregulation of multiple genes that confers growth advantages and unlimited proliferative ability to cancer cells. Altered genes are commonly involved in signaling pathways such as mitogen-activated protein kinase (MAPK), protein kinase B (AKT) (Paraiso *et al.*, 2010) and NF-κB pathway (Mauro *et al.*, 2009). Therefore, the emergence of anticancer agents that are aiming multiple pathways or targets has promising future to cure cancer (Sebolt-Leopold and English, 2006).

Currently, combination therapy or multi-target anticancer agent is used to target multiple cancer-related molecules simultaneously. The purpose of this approach is to activate or to suppress the diverse signaling processes responsible for the survival of tumor (Giordano and Petrelli, 2008). The same concept applies to phytotherapy, where plant extract contains a variety of bioactive compounds that could exert synergistic therapeutic effect by acting or targeting on different receptors of different signaling pathways (Bhatt *et al.*, 2010). Phytotherapy practitioners assert that a whole extract performs better than equivalent dosage of isolated compounds (Nelson and Kursar, 1999). The examples of popular phytotherapy are Indian Medicine (Ayurveda) and Traditional Chinese Medicine. The interaction of compounds within the extract is associated with the improvement in efficiency, declination of undesirable effect, increment in the bioavailability and stability, and sufficient therapeutic effect with less doses (Biavatti, 2009).

The paradigm of cancer therapy has been drifted towards the use of natural products particularly herbal medicine. It is evidenced through the sale of plant-derived chemotherapeutic drugs such as taxanes derivatives (paclitaxel and doxetacel) and camptothecin derivatives (topotecan and irinotecan), which accounted for one third of the total chemotherapeutic drug sales globally, reaching 4 billion dollar in year 2007 (Demain and Vaishnav, 2011).

Deregulation of molecules involved in apoptotic pathway renders unlimited replicative ability to the cancer cells. Many anticancer agents exploit the abnormalities to halt the tumor progression (Ghobrial *et al.*, 2005; Kasibhatla and Tseng, 2003). Plant extracts have been reported to modulate the apoptotic-related pathways in cancer cells. Therefore, they can be served as promising apoptosis-inducing cancer therapeutic agents (Röös, 2015). The apoptotic pathways can be classified into mitochondrial-dependent and mitochondria-independent pathway. The regulation of mitochondrial pathway is controlled by the family of Bcl-2 (Brunelle and Letai, 2009). Another important regulator of apoptosis is the tumor suppressor, p53, which can also incite DNA repair and cellular senescence (Fridman and Lowe, 2003). p53 can also control the transcription of the members of Bcl-2 family especially Bcl-2 and Bax. Literature has shown that many plant-derived extracts are capable to induce apoptosis in cancerous cells by governing p53, Bax and Bcl-2 in the apoptotic signaling pathway (Ryu *et al.*, 2012; Gao *et al.*, 2011; Cheng *et al.*, 2008).

*Dillenia suffruticosa* (Griffith ex Hook. F. and Thomson) Martelli (Family: Dilleniaceae), locally known as “Simpoh air”, is found abundantly in the secondary forest and swampy ground in Peninsular Malaysia to Papua New Guinea and Solomon Islands. The fruit of the plant is traditionally used for the treatment of cancerous growth (Ahmad and Holdsworth, 1995). Aside from that, the plant has been reported to possess antibacterial (Wiart *et al.*, 2004) and antiviral (Muliawan, 2008) activities. The hot water extract of the root of *D. suffruticosa* has anti-cervical (Said, 2010) and anti-colon (Husain, 2010) properties. This study was carried out as the ethyl acetate extract of *D. suffruticosa* was reported to be cytotoxic toward the breast cancer cell line, MCF-7 by Armania *et al.* (2013). In addition, the chemistry profile of ethyl acetate extract of *D. suffruticosa* has never been reported before.

The general objective of this study was to determine the mode of cell death of ethyl acetate extract of *D. suffruticosa* (EADs) and the signaling pathways involved in MCF-7 breast cancer cells.

The specific objectives of this study were:

1. To determine the mode of cell death and cell cycle profile in MCF-7 breast cancer cells treated with EADs.
2. To ascertain the involvement of oxidative stress in MCF-7 breast cancer cells treated with EADs.
3. To elucidate the signaling pathways related to apoptosis, survival and growth in MCF-7 breast cancer cells treated with EADs.
4. To isolate and identify the compounds in EADs.

It was hypothesized that:

1. EADs will show cytotoxic effect and induce cell cycle arrest in MCF-7 breast cancer cells.
2. EADs will induce oxidative stress in MCF-7 breast cancer cells.
3. EADs will induce apoptosis in MCF-7 breast cancer cells through the activation and inhibition of several apoptotic-related signaling pathways.
4. The compounds in EADs will be isolated and identified.



## REFERENCES

- Abdille, M. H., Singh, R. P., Jayaprakasha, G. K. and Jena, B. S. (2005). Antioxidant activity of the extracts from *Dillenia indica* fruits. *Food Chemistry*, 90(4): 891-896.
- ACS (2014). Breast cancer. <http://www.cancer.org/acs/groups/cid/documents/webcontent/003090-pdf.pdf> Retrieved 30th January, 2015.
- Afzal, S., Jensen, S., Sørensen, J., Henriksen, T., Weimann, A. and Poulsen, H. (2012). Oxidative damage to guanine nucleosides following combination chemotherapy with 5-fluorouracil and oxaliplatin. *Cancer Chemotherapy and Pharmacology*, 69(2): 301-307.
- Ahmad, F. B. and Holdsworth, D. K. (1995). Traditional medicinal plants of Sabah, Malaysia Part III. The Rungus people of Kudat. *International Journal of Pharmacognosy: A Journal of Crude Drug Research*, 33: 262–264.
- Ahmad, R. and Abdullah, N. (2013). *Antioxidant principles and in vitro evaluation methods*. Kuala Lumpur: UiTM Press.
- Akhand, R. (2012). *In vitro anti-microbial evaluation of different crude extracts from Dillenia pentagyna leaf and stem*. Bachelor Degree Thesis, East West University, Dhaka.
- Akihisa, T., Franzblau, S. G., Ukiya, M., Okuda, H., Zhang, F., Yasukawa, K., Suzuki, T. and Kimura, Y. (2005). Antitubercular activity of triterpenoids from Asteraceae flowers. *Biological and Pharmaceutical Bulletin*, 28(1): 158-160.
- Alam, M. B., Rahman, M. S., Hasan, M., Khan, M. M., Nahar, K. and Sultana, S. (2012). Antinociceptive and antioxidant activities of the *Dillenia indica* bark. *International Journal of Pharmacology*, 8: 243-251.
- Alessi, D. R., Saito, Y., Campbell, D. G., Cohen, P., Sithanandam, G., Rapp, U., Ashworth, A., Marshall, C. J. and Cowley, S. (1994). Identification of the sites in MAP kinase kinase-1 phosphorylated by p74raf-1. *EMBO Journal*, 13(7): 1610-1619.
- Alexandre, J., Batteux, F., Nicco, C., Chéreau, C., Laurent, A., Guillemin, L., Weill, B. and Goldwasser, F. (2006). Accumulation of hydrogen peroxide is an early and crucial step for paclitaxel-induced cancer cell death both *in vitro* and *in vivo*. *International Journal of Cancer*, 119(1): 41-48.
- Alexandre, J., Hu, Y., Lu, W., Pelicano, H. and Huang, P. (2007). Novel action of paclitaxel against cancer cells: bystander effect mediated by reactive oxygen species. *Cancer Research*, 67(8): 3512-3517.
- Ali, R. M. (2005). *Harnessing the cures from Malaysian rain-forest*. Proceedings of the National Congress on Genetics (12-14 May 2005), Kuala Lumpur, Malaysia.

- Alizart, M., Saunus, J., Cummings, M. and Lakhani, S. R. (2012). Molecular classification of breast carcinoma. *Diagnostic Histopathology*, 18(3): 97-103.
- Althuis, M. D., Fergenbaum, J. H., Garcia-Closas, M., Brinton, L. A., Madigan, M. P. and Sherman, M. E. (2004). Etiology of hormone receptor-defined breast cancer: a systematic review of the literature. *Cancer Epidemiology, Biomarkers and Prevention*, 13(10): 1558-1568.
- Altomare, D. A. and Testa, J. R. (2005). Perturbations of the AKT signaling pathway in human cancer. *Oncogene*, 24(50): 7455-7464.
- Alvarez, A., Lacalle, J., Cañavate, M. L., Alonso-Alconada, D., Lara-Celador, I., Alvarez, F. J. and Hilario, E. (2010). Cell death. A comprehensive approximation. Necrosis. *Microscopy: Science, Technology, Applications and Education*, 1: 1017-1024.
- Amiri-Kordestani, L., Wedam, S., Zhang, L., Tang, S., Tilley, A., Ibrahim, A., Justice, R., Pazdur, R. and Cortazar, P. (2014). First FDA approval of neoadjuvant therapy for breast cancer: pertuzumab for the treatment of patients with HER2-positive breast cancer. *Clinical Cancer Research*, 20(21): 5359-5364.
- Anderson, C. N. and Tolokovsky, A. M. (1999). A role for MAPK/ERK in sympathetic neuron survival: protection against a p53-dependent, JNK-independent induction of apoptosis by cytosine arabinoside. *Journal of Neuroscience*, 19(2): 664-673.
- Andre, F., O'Regan, R., Ozguroglu, M., Toi, M., Xu, B., Jerusalem, G., Masuda, N., Wilks, S., Arena, F., Isaacs, C., Yap, Y. S., Papai, Z., Lang, I., Armstrong, A., Lerzo, G., White, M., Shen, K., Litton, J., Chen, D., Zhang, Y., Ali, S., Taran, T. and Gianni, L. (2014). Everolimus for women with trastuzumab-resistant, HER2-positive, advanced breast cancer (BOLERO-3): a randomised, double-blind, placebo-controlled Phase 3 trial. *Lancet Oncology*, 15(6): 580-591.
- Ansenberger-Fricano, K., Ganini, D. d. S., Mao, M., Chatterjee, S., Dallas, S., Mason, R. P., Stadler, K., Santos, J. H. and Bonini, M. G. (2013). The peroxidase activity of mitochondrial superoxide dismutase (MnSOD/SOD2). *Free Radical Biology and Medicine*, 54: 116-124.
- Aoki, M., Nata, T., Morishita, R., Matsushita, H., Nakagami, H., Yamamoto, K., Yamazaki, K., Nakabayashi, M., Ogihara, T. and Kaneda, Y. (2001). Endothelial apoptosis induced by oxidative stress through activation of NF- $\kappa$ B: antiapoptotic effect of antioxidant agents on endothelial cells. *Hypertension*, 38(1): 48-55.
- Aouadi, M., Binetruy, B., Caron, L., Le Marchand-Brustel, Y. and Bost, F. (2006). Role of MAPKs in development and differentiation: lessons from knockout mice. *Biochimie*, 88(9): 1091-1098.

- Apu, A. S., Muhit, M. A., Tareq, S. M., Pathan, A. H., Jamaluddin, A. T. M. and Ahmed, M. (2010). Antimicrobial activity and brine shrimp lethality bioassay of the leaves extract of *Dillenia indica* Linn. *Journal of Young Pharmacists*, 2(1): 50-53.
- Ariffin, O. Z. and NorSaleha, I. T. (2011). National Cancer Registry Report: Malaysia Cancer Statistics-Data and Figure 2001. *Ministry of Health, Malaysia*.
- Armania, N., Latifah, S. Y., Musa, S. N., Ismail, I. S., Foo, J. B., Chan, K. W., Noreen, H., Hisyam, A. H., Zulfahmi, S. and Ismail, M. (2013). *Dillenia suffruticosa* exhibited antioxidant and cytotoxic activity through induction of apoptosis and G2/M cell cycle arrest. *Journal of Ethnopharmacology*, 146(2): 525-535.
- ARS (2011). National genetic resources program USA: online searchable database <http://www.ars-grin.gov/cgi-bin/npgs/html/index.pl>. Retrieved 30th January, 2015.
- Auyeung, K., Law, P. and Ko, J. (2009). Astragalus saponins induce apoptosis via an ERK-independent NF- $\kappa$ B signaling pathway in the human hepatocellular HepG2 cell line. *International Journal of Molecular Medicine*, 23(2): 189-196.
- Azim, H. A., de Azambuja, E., Colozza, M., Bines, J. and Piccart, M. J. (2011). Long-term toxic effects of adjuvant chemotherapy in breast cancer. *Annals of Oncology*, 22(9): 1939-1947.
- Badertscher, M., Bühlmann, P. and Pretsch, E. (2009). *Structure determination of organic compounds* (Fourth edition). Heidelberg: Springer Berlin.
- Bai, L. and Zhu, W.-G. (2006). p53: structure, function and therapeutic applications. *Journal of Cancer Molecules*, 2(4): 141-153.
- Balasundram, N., Sundram, K. and Samman, S. (2006). Phenolic compounds in plants and agri-industrial by-products: antioxidant activity, occurrence, and potential uses. *Food Chemistry*, 99(1): 191-203.
- Balunas, M. J. and Kinghorn, A. D. (2005). Drug discovery from medicinal plants. *Life Sciences*, 78(5): 431-441.
- Barrera, G. (2012). Oxidative stress and lipid peroxidation products in cancer progression and therapy. *International Scholarly Research Notices Oncology*, 2012: 1-21.
- Barros, M. P. D., Lemos, M., Maistro, E. L., Leite, M. F., Sousa, J. P. B., Bastos, J. K. and Andrade, S. F. D. (2008). Evaluation of antiulcer activity of the main phenolic acids found in Brazilian green propolis. *Journal of Ethnopharmacology*, 120(3): 372-377.
- Basu, A. and Haldar, S. (1998). The relationship between Bcl2, Bax and p53: consequences for cell cycle progression and cell death. *Molecular Human Reproduction*, 4(12): 1099-1109.

- Bate-Smith, E. C. and Harborne, J. B. (1971). Differences in flavonoid content between fresh and herbarium leaf tissue in *Dillenia*. *Phytochemistry*, 10(5): 1055-1058.
- Bates, S. and Vousden, K. H. (1999). Mechanisms of p53-mediated apoptosis. *Cellular and Molecular Life Sciences*, 55(1): 28-37.
- Batra, P. and Sharma, A. (2013). Anti-cancer potential of flavonoids: recent trends and future perspectives. *3 Biotech*, 3(6): 439-459.
- Bayoumi, S. A. L., Rowan, M. G., Beeching, J. R. and Blagbrough, I. S. (2010). Constituents and secondary metabolite natural products in fresh and deteriorated cassava roots. *Phytochemistry*, 71(5-6): 598-604.
- Bhatt, A. N., Mathur, R., Farooque, A., Verma, A. and Dwarakanath, B. S. (2010). Cancer biomarkers-current perspectives. *Indian Journal of Medical Research*, 132: 129-149.
- Biavatti, M. W. (2009). Synergy: an old wisdom, a new paradigm for pharmacotherapy. *Brazilian Journal of Pharmaceutical Sciences*, 45(3): 371-378.
- Billecke, C., Finniss, S., Tahash, L., Miller, C., Mikkelsen, T., Farrell, N. P. and Bogler, O. (2006). Polynuclear platinum anticancer drugs are more potent than cisplatin and induce cell cycle arrest in glioma. *Journal of Neuro-Oncology*, 8(3): 215-226.
- Blagosklonny, M. V. (2000). Cell death beyond apoptosis. *Leukemia*, 14(8): 1502-1508.
- Bland, K. I., Menck, H. R., Scott-Conner, C. E., Morrow, M., Winchester, D. J. and Winchester, D. P. (1998). The national cancer data base: 10-year survey of breast carcinoma treatment at hospitals in the United States. *Cancer*, 83(6): 1262-1273.
- Blokhina, O., Virolainen, E. and Fagerstedt, K. V. (2003). Antioxidants, oxidative damage and oxygen deprivation stress: a review. *Annals of Botany*, 91(2): 179-194.
- Bond, M., Fabunmi, R. P., Baker, A. H. and Newby, A. C. (1998). Synergistic upregulation of metalloproteinase-9 by growth factors and inflammatory cytokines: an absolute requirement for transcription factor NF-kappa B. *FEBS Letters*, 435(1): 29-34.
- Bonni, A., Brunet, A., West, A. E., Datta, S. R., Takasu, M. A. and Greenberg, M. E. (1999). Cell survival promoted by the Ras-MAPK signaling pathway by transcription-dependent and -independent mechanisms. *Science*, 286(5443): 1358-1362.
- Boots, A. W., Haenen, G. R. M. M. and Bast, A. (2008). Health effects of quercetin: from antioxidant to nutraceutical. *European Journal of Pharmacology*, 585(2): 325-337.

- Bortner, C. D. and Cidlowski, J. A. (2007). Cell shrinkage and monovalent cation fluxes: role in apoptosis. *Archives of Biochemistry and Biophysics*, 462(2): 176-188.
- Bose, U., Gunasekaran, K., Bala, V. and Rahman, A. A. (2010). Evaluation of phytochemical and pharmacological properties of *Dillenia indica* Linn. leaves. *Journal of Pharmacology and Toxicology*, 5(5): 222-228.
- Bours, V., Bentires-Alj, M., Hellin, A.-C., Viatour, P., Robe, P., Delhalle, S., Benoit, V. and Merville, M.-P. (2000). Nuclear factor- $\kappa$ B, cancer, and apoptosis. *Biochemical Pharmacology*, 60(8): 1085-1089.
- Boyd, M. R. (1997). *The NCI in vitro anticancer drug discovery screen: concept, implementation, and operation*. B. Teicher (Ed.). New Jersey: Humana Press Totowa.
- Bradford, M. M. (1976). A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry*, 72: 248-254.
- Bradford, P. G. and Awad, A. B. (2007). Phytosterols as anticancer compounds. *Molecular Nutrition and Food Research*, 51(2): 161-170.
- Brancho, D., Tanaka, N., Jaeschke, A., Ventura, J.-J., Kelkar, N., Tanaka, Y., Kyuuma, M., Takeshita, T., Flavell, R. A. and Davis, R. J. (2003). Mechanism of p38 MAP kinase activation *in vivo*. *Genes and Development*, 17(16): 1969-1978.
- Breakthrough-Breast-Cancer (2013). The best treatment. <http://www.breakthrough.org.uk/sites/default/files/media/About%20Breast%20Cancer/Breast%20cancer%20publications/Best%20treatment%20guide%20England%20and%20Wales.pdf>. Retrieved 15th January, 2015.
- Bröker, L. E., Kruyt, F. A. E. and Giaccone, G. (2005). Cell death independent of caspases: a review. *Clinical Cancer Research*, 11(9): 3155-3162.
- Brunelle, J. K. and Letai, A. (2009). Control of mitochondrial apoptosis by the Bcl-2 family. *Journal of Cell Science*, 122(4): 437-441.
- Brusotti, G., Cesari, I., Dentamaro, A., Caccialanza, G. and Massolini, G. (2014). Isolation and characterization of bioactive compounds from plant resources: the role of analysis in the ethnopharmacological approach. *Journal of Pharmaceutical and Biomedical Analysis*, 87: 218-228.
- Bucar, F., Wubea, A. and Schmid, M. (2013). Natural product isolation—how to get from biological material to pure compounds. *Natural Product Reports*, 30: 525-545.
- Buettner, G. R. (1993). The pecking order of free radicals and antioxidants: lipid peroxidation,  $\alpha$ -tocopherol, and ascorbate. *Archives of Biochemistry and Biophysics*, 300(2): 535-543.

- Burlacu, A. (2003). Regulation of apoptosis by Bcl-2 family proteins. *Journal of Cellular and Molecular Medicine*, 7(3): 249-257.
- Burstein, H. J., Polyak, K., Wong, J. S., Lester, S. C. and Kaelin, C. M. (2004). Ductal carcinoma *in situ* of the breast. *New England Journal of Medicine*, 350(14): 1430-1441.
- Butler, M. S. (2004). The role of natural product chemistry in drug discovery. *Journal of Natural Products*, 67(12): 2141– 2153.
- Butterfield, D. A., Koppal, T., Howard, B., Subramaniam, R. A. M., Hall, N., Hensley, K., Yatin, S., Allen, K., Aksenov, M., Aksenova, M. and Carney, J. (1998). Structural and functional changes in roteins induced by free radical-mediated oxidative stress and protective action of the antioxidants N-tert-butyl- $\alpha$ -phenylnitro and vitamin E. *Annals of the New York Academy of Sciences*, 854(1): 448-462.
- Buttke, T. M. and Sandstrom, P. A. (1994). Oxidative stress as a mediator of apoptosis. *Immunology Today*, 15(1): 7-10.
- Cagnol, S. and Chambard, J. C. (2010). ERK and cell death: mechanisms of ERK-induced cell death--apoptosis, autophagy and senescence. *FEBS Journal*, 277(1): 2-21.
- Cailleau, R., Young, R., Oliv é, M. and Reeves, W. J. (1974). Breast tumor cell lines from pleural effusions. *Journal of the National Cancer Institute*, 53(3): 661-674.
- Calderon-Montano, J. M., Burgos-Moron, E., Perez-Guerrero, C. and Lopez-Lazaro, M. (2011). A review on the dietary flavonoid kaempferol. *Mini-Reviews in Medicinal Chemistry*, 11(4): 298-344.
- Calixto, J. B. (2000). Efficacy, safety, quality control, marketing and regulatory guidelines for herbal medicines (phytotherapeutic agents). *Brazilian Journal of Medical and Biological Research*, 33: 179-189.
- Cargnello, M. and Roux, P. P. (2011). Activation and function of the MAPKs and their substrates, the MAPK-activated protein kinases. *Microbiology and Molecular Biology Reviews*, 75(1): 50-83.
- Carrick, S., Parker, S., Thornton, C. E., Ghersi, D., Simes, J. and Wilcken, N. (2009). Single agent versus combination chemotherapy for metastatic breast cancer. *Cochrane Database Systematic Reviews*, (2): 1-104.
- cBioPortal (2013). cBioPortal for cancer genomics. <http://www.cbioportal.org/public-portal/>. Retrieved 29th January, 2015
- Cerveira, N., Bizarro, S. and Teixeira, M. R. (2012). Cancer cell cycle. *Canal BQ*, 1(9): 1-8.

- Cesari, I., Hoerle, M., Simoes-Pires, C., Grisoli, P., Queiroz, E. F., Dacarro, C., Marcourt, L., Moundipa, P. F., Carrupt, P. A., Cuendet, M., Caccialanza, G., Wolfender, J. L. and Brusotti, G. (2013). Anti-inflammatory, antimicrobial and antioxidant activities of *Diospyros bipindensis* (Gurke) extracts and its main constituents. *Journal of Ethnopharmacology*, 146(1): 264-270.
- Chang, L. and Karin, M. (2001). Mammalian MAP kinase signalling cascades. *Nature*, 410(6824): 37-40.
- Chang, Y. J., Huang, Y. P., Li, Z. L. and Chen, C. H. (2012). GRP78 knockdown enhances apoptosis via the down-regulation of oxidative stress and Akt pathway after epirubicin treatment in colon cancer DLD-1 cells. *PLoS ONE*, 7(4): 1-11.
- Chen, Y. R., Meyer, C. F. and Tan, T. H. (1996). Persistent activation of c-Jun N-terminal kinase 1 (JNK1) in gamma radiation-induced apoptosis. *Journal of Biological Chemistry*, 271(2): 631-634.
- Cheng, J. Q., Lindsley, C. W., Cheng, G. Z., Yang, H. and Nicosia, S. V. (2005). The Akt/PKB pathway: molecular target for cancer drug discovery. *Oncogene*, 24(50): 7482-7492.
- Cheng, Y.-L., Lee, S.-C., Harn, H.-J., Huang, H.-C. and Chang, W.-L. (2008). The extract of *Hibiscus syriacus* inducing apoptosis by activating p53 and AIF in human lung cancer cells. *American Journal of Chinese Medicine*, 36(01): 171-184.
- Chew, H. K. (2001). Adjuvant therapy for breast cancer: who should get what? *Western Journal of Medicine*, 174(4): 284-287.
- Chia, Y. C., Rajbanshi, R., Calhoun, C. and Chiu, R. H. (2010). Anti-neoplastic effects of gallic acid, a major component of *Toona sinensis* leaf extract, on oral squamous carcinoma cells. *Molecules*, 15(11): 8377-83389.
- Chong, K. Y., Tan, H. T. W. and Corlett, R. T. (2009). A checklist of the total vascular plant flora of Singapore: native, naturalised and cultivated species. [http://rmbr.nus.edu.sg/raffles\\_museum\\_pub/flora\\_of\\_singapore\\_tc.pdf](http://rmbr.nus.edu.sg/raffles_museum_pub/flora_of_singapore_tc.pdf). Retrieved 10th February, 2015
- Chowdhury, T. A., Halder, S., Guha, S. K. and Mosihuzzaman, M. (1998). Isolation and characterization of three triterpenoids and adicarboxylic acid from the fruit pulp of *Dillenia indica*. *Journal of the Bangladesh Chemical Society*, 11: 89-96.
- Chugh, C. A., Mehta, S. and Dua, H. (2012). Phytochemical screening and evaluation of biological activities of some medicinal plants of Phagwara, Punjab. *Asian Journal of Chemistry*, 24(12): 5903-5905.

- Chung, P. (2004). *Screening of Malaysian plants for antimicrobial activity and isolation and identification of antimicrobial compounds of Callicarpa farinosa*. Master of Medical Science Thesis, University of Malaya, Malaysia.
- Citoğlu, G. S. and Acıkara, Ö. B. (2012). Column chromatography for terpenoids and flavonoids. S. Dhanarasu (Ed.), *Chromatography and its applications*. Croatia: InTech.
- Clark, A. S., West, K., Streicher, S. and Dennis, P. A. (2002). Constitutive and inducible Akt activity promotes resistance to chemotherapy, trastuzumab, or tamoxifen in breast cancer cells. *Molecular Cancer Therapeutics*, 1(9): 707-717.
- Collins-Burow, B. M., Burow, M. E., Duong, B. N. and McLachlan, J. A. (2000). Estrogenic and antiestrogenic activities of flavonoid phytochemicals through estrogen receptor binding-dependent and -independent mechanisms. *Nutrition and Cancer*, 38(2): 229-244.
- Collins, K., Jacks, T. and Pavletich, N. P. (1997). The cell cycle and cancer. *Proceedings of the National Academy of Sciences*, 94(7): 2776-2778.
- Constantinou, C., Papas, K. A. and Constantinou, A. I. (2009). Caspase-independent pathways of programmed cell death: the unraveling of new targets of cancer therapy? *Current Cancer Drug Targets*, 9(6): 717-728
- Cossarizza, A., Baccaranicontri, M., Kalashnikova, G. and Franceschi, C. (1993). A new method for the cytofluorometric analysis of mitochondrial membrane potential using the J-aggregate forming lipophilic cation 5,5,6,6-tetrachloro-1,1,3,3-tetraethylbenzimidazolcarbocyanine iodide (JC-1). *Biochemical and Biophysical Research Communications*, 197(1): 40-45.
- Cossarizza, A. and Salvioli, S. (2001). Flow cytometric analysis of mitochondrial membrane potential using JC-1. *Current protocols in cytometry*. United States of America: John Wiley and Sons, Inc.
- Costantini, D. L., Villani, D. F., Vallis, K. A. and Reilly, R. M. (2010). Methotrexate, paclitaxel, and doxorubicin radiosensitize HER2-amplified human breast cancer cells to the auger electron-emitting radiotherapeutic agent <sup>111</sup>In-NLS-trastuzumab. *Journal of Nuclear Medicine*, 51(3): 477-483.
- Cragg , G. M. and Newman, D. J. (2005). Plants as a source of anti-cancer agents. *Journal of Ethnopharmacology*, 100: 72-79.
- Crews, C. M., Alessandrini, A. and Erikson, R. L. (1992). The primary structure of MEK, a protein kinase that phosphorylates the ERK gene product. *Science*, 258(5081): 478-480.
- Crowell, J. A., Steele, V. E. and Fay, J. R. (2007). Targeting the AKT protein kinase for cancer chemoprevention. *Molecular Cancer Therapeutics*, 6(8): 2139-2148.

- Cuendet, M. and Pezzuto, J. M. (2003). Antitumor activity of bruceantin: an old drug with new promise. *Journal of Natural Products*, 67(2): 269-272.
- Cueva, C., Moreno-Arribas, M. V., Martí-Álvarez, P. J., Bills, G., Vicente, M. F., Basilio, A., Rivas, C. L., Requena, T., Rodríguez, J. M. and Bartolomé, B. (2010). Antimicrobial activity of phenolic acids against commensal, probiotic and pathogenic bacteria. *Research in Microbiology*, 161(5): 372-382.
- Cullen, S. P. and Martin, S. J. (2006). Caspase activation pathways: some recent progress. *Cell Death and Differentiation*, 16(7): 935-938.
- Cushnie, T. P. and Lamb, A. J. (2005). Antimicrobial activity of flavonoids. *International Journal of Antimicrobial Agents*, 26(5): 343-356.
- Da Silva, L., Clarke, C. and Lakhani, S. R. (2007). Demystifying basal-like breast carcinomas. *Journal of Clinical Pathology*, 60(12): 1328-1332.
- Damayanthi, Y. and Lown, J. W. (1998). Podophyllotoxins: current status and recent developments. *Current Medicinal Chemistry*, 5(3): 205-252.
- Dan, S. and Dan, S. S. (1980). Triterpenoids of Indian Dilleniaceae. *Journal of the Indian Chemical Society*, 57: 760-765.
- Das, M. and Sarma, B. P. (2013). Evaluation of hypoglycemic effect of an Indian fruit: *Dillenia indica*. *International Journal of Research in Ayurveda and Pharmacy*, 4(4): 545-546.
- De Laurentiis, M., Cianniello, D., Caputo, R., Stanzione, B., Arpino, G., Cinieri, S., Lorusso, V. and De Placido, S. (2010). Treatment of triple negative breast cancer (TNBC): current options and future perspectives. *Cancer Treatment Reviews*, 36(3): S80-S86.
- Deacon, K., Mistry, P., Chernoff, J., Blank, J. L. and Patel, R. (2003). p38 Mitogen-activated protein kinase mediates cell death and p21-activated kinase mediates cell survival during chemotherapeutic drug-induced mitotic arrest. *Molecular Biology of the Cell*, 14(5): 2071-2187.
- DeBruin, L. S. and Josephy, P. D. (2002). Perspectives on the chemical etiology of breast cancer. *Environmental Health Perspectives*, 110(1): 119-128.
- Demain, A. L. and Vaishnav, P. (2011). Natural products for cancer chemotherapy. *Microbial Biotechnology*, 4(6): 687-699.
- Deraniyagala, S. A., Ratnasooriya, W. D. and Wijesekara, P. S. (2014). Antinociceptive activity of aqueous fruit extract of *Dillenia retusa* T. in rats. *International Journal of Advances in Pharmacy, Biology and Chemistry*, 3(2): 371-377.

- Devita, V. T., Jr., Serpick, A. A. and Carbone, P. P. (1970). Combination chemotherapy in the treatment of advanced Hodgkin's disease. *Annals of Internal Medicine*, 73(6): 881-895.
- Dhanasekaran, D. N. and Reddy, E. P. (2008). JNK signaling in apoptosis. *Oncogene*, 27(48): 6245-6251.
- Dhillon, A. S., Hagan, S., Rath, O. and Kolch, W. (2007). MAP kinase signalling pathways in cancer. *Oncogene*, 26(22): 3279-3290.
- Diehl, J. A., Cheng, M., Roussel, M. F. and Sherr, C. J. (1998). Glycogen synthase kinase-3 beta regulates cyclin D1 proteolysis and subcellular localization. *Genes and Development*, 12(22): 3499-3511.
- Ding, W. X., Shen, H. M. and Ong, C. N. (2000). Critical role of reactive oxygen species and mitochondrial permeability transition in microcystin-induced rapid apoptosis in rat hepatocytes. *Hepatology*, 32(3): 547-555.
- Dolado, I. and Nebreda, A. R. (2008). AKT and oxidative stress team up to kill cancer cells. *Cancer Cell*, 14(6): 427-429.
- Dreosti, I. E. (2000). Antioxidant polyphenols in tea, cocoa, and wine. *Nutrition*, 16(7-8): 692-694.
- Dunnwald, L., Rossing, M. and Li, C. (2007). Hormone receptor status, tumor characteristics, and prognosis: a prospective cohort of breast cancer patients. *Breast Cancer Research*, 9(1): 1-10.
- Egger, L., Schneider, J., Rheme, C., Tapernoux, M., Hacki, J. and Borner, C. (2003). Serine proteases mediate apoptosis-like cell death and phagocytosis under caspase-inhibiting conditions. *Cell Death and Differentiation*, 10(10): 1188-1203.
- Eldahshan, O. A. (2011). Isolation and structure elucidation of phenolic compounds of Carob leaves grown in Egypt. *Current Research Journal of Biological Sciences*, 3(1): 52-55.
- Elmore, S. (2007). Apoptosis: a review of programmed cell death. *Toxicologic Pathology*, 35(4): 495-516.
- Erin, L. S. H., Mun, P. P., Ling, N. S., Ping, O. C., Jie, S. X., Ying, N. S., Yng, L. T., Buru, A. S. and Pichika, M. R. (2013). Evaluation of four extracts from *Dillenia ovata* stem bark and leaves for antibacterial and antifungal activity. *International Journal of Pharmacy and Pharmaceutical Sciences*, 5(3): 471-474.
- Eroles, P., Bosch, A., Alejandro Pérez-Fidalgo, J. and Lluch, A. (2012). Molecular biology in breast cancer: intrinsic subtypes and signaling pathways. *Cancer Treatment Reviews*, 38(6): 698-707.

- Fabiani, R., Rosignoli, P., De Bartolomeo, A., Fuccelli, R. and Morozzi, G. (2008). Inhibition of cell cycle progression by hydroxytyrosol is associated with upregulation of cyclin-dependent protein kinase inhibitors p21WAF1/Cip1 and p27Kip1 and with induction of differentiation in HL60 cells. *Journal of Nutrition*, 138(1): 42-48.
- Ferlay, J., Soerjomataram, I., Dikshit, R., Eser, S., Mathers, C., Rebelo, M., Parkin, D. M., Forman, D. and Bray, F. (2015). Cancer incidence and mortality worldwide: Sources, methods and major patterns in GLOBOCAN 2012. *International Journal of Cancer*, 136(5): E359-E386.
- Ferreira, C. G., Epping, M., Kruyt, F. A. E. and Giaccone, G. (2002). Apoptosis: target of cancer therapy. *Clinical Cancer Research*, 8(7): 2024-2034.
- Fey, D., Croucher, D. R., Kolch, W. and Kholodenko, B. N. (2012). Crosstalk and signalling switches in mitogen-activated protein kinase cascades. *Frontiers in Physiology*, 3: 1-14.
- Fiuza, S. M., Gomes, C., Teixeira, L. J., Girao da Cruz, M. T., Cordeiro, M. N., Milhazes, N., Borges, F. and Marques, M. P. (2004). Phenolic acid derivatives with potential anticancer properties--a structure-activity relationship study. Part 1: methyl, propyl and octyl esters of caffeic and gallic acids. *Bioorganic and Medicinal Chemistry*, 12(13): 3581-3589.
- Foghsgaard, L., Wissing, D., Mauch, D., Lademann, U., Bastholm, L., Boes, M., Elling, F., Leist, M. and Jäätelä, M. (2001). Cathepsin B acts as a dominant execution protease in tumor cell apoptosis induced by tumor necrosis factor. *Journal of Cell Biology*, 153(5): 999-1010.
- Foo, J. B., Yazan, L. S., Tor, Y. S., Armania, N., Ismail, N., Imam, M. U., Yeap, S. K., Cheah, Y. K., Abdullah, R. and Ismail, M. (2014). Induction of cell cycle arrest and apoptosis in caspase-3 deficient MCF-7 cells by *Dillenia suffruticosa* root extract via multiple signalling pathways. *BMC Complementary and Alternative Medicine*, 14: 1-16.
- Frank, D. A. (2012). *Signaling pathways in cancer: twenty-first century approaches to cancer therapy*. D. A. Frank (Ed.), (Vol. 7). New York: Springer.
- Franke, T. F., Hornik, C. P., Segev, L., Shostak, G. A. and Sugimoto, C. (2003). PI3K/Akt and apoptosis: size matters. *Oncogene*, 22(56): 8983-8998.
- Fresno Vara, J. A., Casado, E., de Castro, J., Cejas, P., Belda-Iniesta, C. and Gonzalez-Baron, M. (2004). PI3K/Akt signalling pathway and cancer. *Cancer Treatment Reviews*, 30(2): 193-204.
- Fridman, J. S. and Lowe, S. W. (2003). Control of apoptosis by p53. *Oncogene*, 22(56): 9030-9040.

- Fuloria, N. K. and Fuloria, S. (2013). Structural elucidation of small organic molecules by 1D, 2D and multi dimensional-solution NMR spectroscopy. *Analytical and Bioanalytical Techniques*, S11: 1-8.
- Funk, J. O. (2001). Cell cycle checkpoint genes and cancer. *eLS*. United States of America: John Wiley and Sons Ltd.
- Gandhi, D. and Mehta, P. (2013). *Dillenia indica* Linn. and *Dillenia pentagyna* Roxb.: pharmacognostic, phytochemical and therapeutic aspects. *Journal of Applied Pharmaceutical Science*, 3(11): 134-142.
- Gao, J., Morgan, W. A., Sanchez-Medina, A. and Corcoran, O. (2011). The ethanol extract of *Scutellaria baicalensis* and the active compounds induce cell cycle arrest and apoptosis including upregulation of p53 and Bax in human lung cancer cells. *Toxicology and Applied Pharmacology*, 254(3): 221-228.
- Gartel, A. L. and Tyner, A. L. (2002). The role of the cyclin-dependent kinase inhibitor p21 in apoptosis. *Molecular Cancer Therapeutics*, 1(8): 639-649.
- Geddes, D. T. (2007). Inside the lactating breast: the latest anatomy research. *Journal of Midwifery and Women's Health*, 52(6): 556-563.
- Ghobrial, I. M., Witzig, T. E. and Adjei, A. A. (2005). Targeting apoptosis pathways in cancer therapy. *CA: A Cancer Journal for Clinicians*, 55(3): 178-194.
- Gibbons, S. (2004). Anti-staphylococcal plant natural products. *Natural Product Reports*, 21: 263-277.
- Giordano, S. and Petrelli, A. (2008). From single- to multi-target drugs in cancer therapy: when aspecificity becomes an advantage. *Current Medicinal Chemistry*, 15(5): 422-432.
- Goldstein, D. (1998). *Inverted microscopes*. United Kingdom: Microscopy UK.
- Gomes, C. A., Girão da Cruz, T., Andrade, J. L., Milhazes, N., Borges, F. and Marques, M. P. M. (2003). Anticancer activity of phenolic acids of natural or synthetic origin: a structure-activity study. *Journal of Medicinal Chemistry*, 46(25): 5395-5401.
- Gordaliza, M. (2007). Natural products as leads to anticancer drugs. *Clinical and Translational Oncology*, 9(12): 767-776.
- Gorrini, C., Harris, I. S. and Mak, T. W. (2013). Modulation of oxidative stress as an anticancer strategy. *Nature Reviews Drug Discovery*, 12(12): 931-947.
- Gospodarowicz, M. K., Miller, D., Groome, P. A., Greene, F. L., Logan, P. A. and Sabin, L. H. (2004). The process for continuous improvement of the TNM classification. *Cancer*, 100(1): 1-5.

- Gottlieb, E., Armour, S. M., Harris, M. H. and Thompson, C. B. (2003). Mitochondrial membrane potential regulates matrix configuration and cytochrome c release during apoptosis. *Cell Death and Differentiation*, 10(6): 709-717.
- Gratton, J. P., Morales-Ruiz, M., Kureishi, Y., Fulton, D., Walsh, K. and Sessa, W. C. (2001). Akt down-regulation of p38 Signaling provides a novel mechanism of vascular endothelial growth factor-mediated cytoprotection in endothelial cells. *Journal of Biological Chemistry*, 276(32): 30359-30365.
- Greene, F. L. and Sabin, L. H. (2009). A worldwide approach to the TNM staging system: collaborative efforts of the AJCC and UICC. *Journal of Surgical Oncology*, 99(5): 269-272.
- Grosvenor, P. W., Supriono, A. and Gray, D. O. (1995). Medicinal plants from Riau province, Sumatra, Indonesia. Part 2: antibacterial and antifungal activity. *Journal of Ethnopharmacology*, 45(2): 97-111.
- Guarneri, V., Frassoldati, A., Bottini, A., Cagossi, K., Bisagni, G., Sarti, S., Ravaioli, A., Cavanna, L., Giardina, G., Musolino, A., Untch, M., Orlando, L., Artioli, F., Boni, C., Generali, D. G., Serra, P., Bagnalasta, M., Marini, L., Piacentini, F., D'Amico, R. and Conte, P. (2012). Preoperative chemotherapy plus trastuzumab, lapatinib, or both in human epidermal growth factor receptor 2-positive operable breast cancer: results of the randomized Phase II CHER-LOB study. *Journal of Clinical Oncology*, 30(16): 1989-1995.
- Guicciardi, M. and Gores, G. (2003). The death receptor family and the extrinsic pathway. X.-M. Yin & Z. Dong (Eds.). *Essentials of apoptosis* (pp 67-84). New York: Humana Press.
- Gurni, A. A. and Kubitzki, K. (1981). Flavonoid chemistry and systematics of the Dilleniaceae. *Biochemical Systematics and Ecology*, 9(2-3): 109-114.
- Gutierrez-Uzquiza, A., Arechederra, M., Bragado, P., Aguirre-Ghiso, J. A. and Porras, A. (2011). p38 $\alpha$  mediates cell survival in response to oxidative stress via induction of antioxidant genes: effect on the p70S6K pathway. *Journal of Biological Chemistry*, 287(4): 2632-2642.
- Guzi, T. (2004). CYC-202 Cyclacel. *Current Opinion in Investigational Drugs*, 5(12): 1311-1318.
- Halestrap, A. P., McStay, G. P. and Clarke, S. J. (2002). The permeability transition pore complex: another view. *Biochimie*, 84(2-3): 153-166.
- Halliwell, B. (1993). The role of oxygen radicals in human disease, with particular reference to the vascular system. *Pathophysiology of Haemostasis and Thrombosis*, 23(1): 118-126.
- Halliwell, B. and Whiteman, M. (2004). Measuring reactive species and oxidative damage *in vivo* and in cell culture: how should you do it and what do the results mean? *British Journal of Pharmacology*, 142(2): 231-255.

- Han, J. and Sun, P. (2007). The pathways to tumor suppression via route p38. *Trends in Biochemical Sciences*, 32(8): 364-371.
- Han, S. I., Kim, Y. S. and Kim, T. H. (2008). Role of apoptotic and necrotic cell death under physiologic conditions. *BMB Reports*, 41(1): 1-10.
- Hanahan, D. and Weinberg, Robert A. (2011). Hallmarks of cancer: the next generation. *Cell*, 144(5): 646-674.
- Hanson, J. R. (2003). The classes of natural product and their isolation. *Natural products: the secondary metabolites* (pp 1–34). Cambridge, UK: Royal Society of Chemistry.
- Hanum, F. and Hamzah, N. (1999). The use of medicinal plant species by the Temuan tribe of Ayer Hitam Forest, Selangor, Peninsular Malaysia. *Pertanika Journal of Tropical Agriculture*, 22(2): 85-94.
- Haque, M. E., Islam, M. N., Hossain, M., Mohamad, A. U., Karim, M. F. and Rahman, M. A. (2008). Antimicrobial and cytotoxic activities of *Dillenia pentagyna*. *Journal of Pharmacology Science*, 7(1): 103-105.
- Hashimoto, K., Mori, N., Tamesa, T., Okada, T., Kawauchi, S., Oga, A., Furuya, T., Tangoku, A., Oka, M. and Sasaki, K. (2004). Analysis of DNA copy number aberrations in hepatitis C virus-associated hepatocellular carcinomas by conventional CGH and array CGH. *Modern Pathology*, 17(6): 617-622.
- Helbig, G., Christopherson, K. W., Bhat-Nakshatri, P., Kumar, S., Kishimoto, H., Miller, K. D., Broxmeyer, H. E. and Nakshatri, H. (2003). NF-kappaB promotes breast cancer cell migration and metastasis by inducing the expression of the chemokine receptor CXCR4. *Journal of Biological Chemistry*, 278(24): 21631-21638.
- Hennessy, B. T., Smith, D. L., Ram, P. T., Lu, Y. and Mills, G. B. (2005). Exploiting the PI3K/AKT pathway for cancer drug discovery. *Nature Reviews Drug Discovery*, 4(12): 988-1004.
- Herbst, R. S. and Frankel, S. R. (2004). Oblimersen sodium (GenaSense bcl-2 antisense oligonucleotide): a rational therapeutic to enhance apoptosis in therapy of lung cancer. *Clinical Cancer Research*, 10(12): 4245-4248.
- Herschkowitz, J. I., Simin, K., Weigman, V. J., Mikaelian, I., Usary, J., Hu, Z., Rasmussen, K. E., Jones, L. P., Assefnia, S., Chandrasekharan, S., Backlund, M. G., Yin, Y., Khramtsov, A. I., Bastein, R., Quackenbush, J., Glazer, R. I., Brown, P. H., Green, J. E., Kopelovich, L., Furth, P. A., Palazzo, J. P., Olopade, O. I., Bernard, P. S., Churchill, G. A., Van Dyke, T. and Perou, C. M. (2007). Identification of conserved gene expression features between murine mammary carcinoma models and human breast tumors. *Genome Biology*, 8(5): R76-R76.

- Hortobagyi, G. N., de la Garza Salazar, J., Pritchard, K., Amadori, D., Haidinger, R., Hudis, C. A., Khaled, H., Liu, M.-C., Martin, M., Namer, M., O'Shaughnessy, J. A., Shen, Z. Z. and Albain, K. S. (2005). The global breast cancer burden: variations in epidemiology and survival. *Clinical Breast Cancer*, 6(5): 391-401.
- Howard, A. and Pelc, S. R. (1986). Synthesis of desoxyribonucleic acid in normal and irradiated cells and its relation to chromosome breakage. *International Journal of Radiation Biology*, 49(2): 207-218.
- Hsu, J.-D., Kao, S.-H., Ou, T.-T., Chen, Y.-J., Li, Y.-J. and Wang, C.-J. (2011). Gallic acid induces G2/M phase arrest of breast cancer cell MCF-7 through stabilization of p27Kip1 attributed to disruption of p27Kip1/Skp2 complex. *Journal of Agricultural and Food Chemistry*, 59(5): 1996-2003.
- Huang, K. F., zhang, G. D., Huang, Y. Q. and Diao, Y. (2012). Wogonin induces apoptosis and down-regulates survivin in human breast cancer MCF-7 cells by modulating PI3K–AKT pathway. *International Immunopharmacology*, 12(2): 334-341.
- Hudes, G., Carducci, M., Tomczak, P., Dutcher, J., Figlin, R., Kapoor, A., Staroslawska, E., Sosman, J., McDermott, D., Bodrogi, I., Kovacevic, Z., Lesovoy, V., Schmidt-Wolf, I. G. H., Barbarash, O., Gokmen, E., O'Toole, T., Lustgarten, S., Moore, L. and Motzer, R. J. (2007). Temsirolimus, interferon alfa, or both for advanced renal-cell carcinoma. *New England Journal of Medicine*, 356(22): 2271-2281.
- Hung, H. (2004). Inhibition of estrogen receptor alpha expression and function in MCF-7 cells by kaempferol. *Journal of Cellular Physiology*, 198(2): 197-208.
- Hunziker, J. and Berk, J. (1976). Device for holding a variety of tissue culture vessels during microscopy. *In Vitro*, 12(3): 263-264.
- Husain, N. (2010). *Anti-colorectal cancer properties of Dillenia suffruticosa (Griffith ex. Hook. f. & Thomson) Martelli water extract in in vitro and in vivo models*. Master Thesis, Universiti Putra Malaysia, Malaysia.
- Ibrahim, N. I., Dahlui, M., Aina, E. N. and Al-Sadat, N. (2012). Who are the breast cancer survivors in Malaysia? *Asian Pacific Journal of Cancer Prevention*, 13: 2213-2218.
- Ichimura, K., Vogazianou, A. P., Liu, L., Pearson, D. M., Backlund, L. M., Plant, K., Baird, K., Langford, C. F., Gregory, S. G. and Collins, V. P. (2007). 1p36 is a preferential target of chromosome 1 deletions in astrocytic tumours and homozygously deleted in a subset of glioblastomas. *Oncogene*, 27(14): 2097-2108.
- Igney, F. H. and Krammer, P. H. (2002). Death and anti-death: tumour resistance to apoptosis. *Nature Reviews Cancer*, 2(4): 277-288.

- Ito, N., Hakamata, H. and Kusu, F. (2010). Simultaneous determination of betasitosterol, campesterol, stigmasterol, and brassicasterol in serum by high-performance liquid chromatography with electrochemical detection. *Analytical Methods*, 2(2): 174-179.
- Itokawa, H., Wang, X. and Lee, K. H. (2005). Homoharringtonine and related compounds. G. M. Cragg, D. G. I. Kingston & D. J. Newman (Eds.). *Anticancer agents from natural products* (pp 47). Boca Raton: Brunner-Routledge Psychology Press.
- Jada, S. R., Subur, G. S., Matthews, C., Hamzah, A. S., Lajis, N. H., Saad, M. S., Stevens, M. F. G. and Stanslas, J. (2007). Semisynthesis and *in vitro* anticancer activities of andrographolide analogues. *Phytochemistry*, 68(6): 904-912.
- Jager, S., Trojan, H., Kopp, T., Laszczyk, M. N. and Scheffler, A. (2009). Pentacyclic triterpene distribution in various plants-rich sources for a new group of multi-potent plant extracts. *Molecules*, 14(6): 2016-2031.
- Jänicke, R. U. (2009). MCF-7 breast carcinoma cells do not express caspase-3. *Breast Cancer Research and Treatment*, 117(1): 219-221.
- Jänicke, R. U., Sprengart, M. L., Wati, M. R. and Porter, A. G. (1998). Caspase-3 is required for DNA fragmentation and morphological changes associated with apoptosis. *Journal of Biological Chemistry*, 273(16): 9357-9360.
- Janssen-Heininger, Y. M. W., Poynter, M. E. and Baeuerle, P. A. (2000). Recent advances towards understanding redox mechanisms in the activation of nuclear factor- $\kappa$ b. *Free Radical Biology and Medicine*, 28(9): 1317-1327.
- Jassem, J., Pienkowski, T., Pluzanska, A., Jelic, S., Gorbunova, V., Mrsic-Krmpotic, Z., Berzins, J., Nagykalnai, T., Wigler, N., Renard, J., Munier, S. and Weil, C. (2001). Doxorubicin and paclitaxel versus fluorouracil, doxorubicin, and cyclophosphamide as first-line therapy for women with metastatic breast cancer: final results of a randomized phase III multicenter trial. *Journal of Clinical Oncology*, 19(6): 1707-1715.
- Jatoi, I., Anderson, W. F., Jeong, J.-H. and Redmond, C. K. (2011). Breast cancer adjuvant therapy: time to consider its time-dependent effects. *Journal of Clinical Oncology*, 29(17): 2301-2304.
- Jemal, A., Bray, F., Melissa, M., Ferlay, M. J., Ward, E. and Forman, D. (2011). Global Cancer Statistics, *CA: A Cancer Journal for Clinicians*, 61(2): 69-90.
- Jiang, G., Lin, S., Wen, L., Jiang, Y., Zhao, M., Chen, F., Prasad, K. N., Duan, X. and Yang, B. (2013). Identification of a novel phenolic compound in litchi (*Litchi chinensis* Sonn.) pericarp and bioactivity evaluation. *Food Chemistry*, 136(2): 563-568.

- Johnson, G. L. and Nakamura, K. (2007). The c-Jun kinase/stress-activated pathway: regulation, function and role in human disease. *Biochimica et Biophysica Acta*, 1773(8): 1341-1348.
- Jones, C., Nonni, A. V., Fulford, L., Merrett, S., Chaggar, R., Eusebi, V. and Lakhani, S. R. (2001). CGH analysis of ductal carcinoma of the breast with basaloid/myoepithelial cell differentiation. *British Journal of Cancer*, 85(3): 422-427.
- Joshi, C., Karumuri, B., Newman, J. J. and DeCoster, M. A. (2012). Cell morphological changes combined with biochemical assays for assessment of apoptosis and apoptosis reversal. *Current Microscopy Contributions to Advances in Science and Technology*: 756-762.
- Joza, N., Susin, S. A., Daugas, E., Stanford, W. L., Cho, S. K., Li, C. Y., Sasaki, T., Elia, A. J., Cheng, H. Y., Ravagnan, L., Ferri, K. F., Zamzami, N., Wakeham, A., Hakem, R., Yoshida, H., Kong, Y. Y., Mak, T. W., Zuniga-Pflucker, J. C., Kroemer, G. and Penninger, J. M. (2001). Essential role of the mitochondrial apoptosis-inducing factor in programmed cell death. *Nature*, 410(6828): 549-554.
- Kajstura, M., Halicka, H. D., Pryjma, J. and Darzynkiewicz, Z. (2007). Discontinuous fragmentation of nuclear DNA during apoptosis revealed by discrete “sub-G1” peaks on DNA content histograms. *Cytometry Part A*, 71A(3): 125-131.
- Kaltschmidt, B., Kaltschmidt, C., Hofmann, T. G., Hehner, S. P., Dröge, W. and Schmitz, M. L. (2000). The pro- or anti-apoptotic function of NF-κB is determined by the nature of the apoptotic stimulus. *European Journal of Biochemistry*, 267(12): 3828-3835.
- Kamal-Eldin, A. and Appelqvist, L. Å. (1996). The chemistry and antioxidant properties of tocopherols and tocotrienols. *Lipids*, 31(7): 671-701.
- Kamal-Eldin, A., Frank, J., Razdan, A., Tengblad, S., Basu, S. and Vessby, B. (2000). Effects of dietary phenolic compounds on tocopherol, cholesterol, and fatty acids in rats. *Lipids*, 35(4): 427-435.
- Kampa, M., Alexaki, V.-I., Notas, G., Nifli, A.-P., Nistikaki, A., Hatzoglou, A., Bakogeorgou, E., Kouimtzoglou, E., Blekas, G., Boskou, D., Gravanis, A. and Castanas, E. (2004). Antiproliferative and apoptotic effects of selective phenolic acids on T47D human breast cancer cells: potential mechanisms of action. *Breast Cancer Research*, 6(2): R63-R74.
- Kanduc, D., Mittelman, A., Serpico, R., Sinigaglia, E., Sinha, A. A., Natale, C., Santacroce, R., Di Corcia, M. G., Lucchese, A., Dini, L., Pani, P., Santacroce, S., Simone, S., Bucci, R. and Farber, E. (2002). Cell death: apoptosis versus necrosis *International Journal of Oncology*, 21(1): 165-170.

- Kang, G.-Y., Lee, E.-R., Kim, J.-H., Jung, J. W., Lim, J., Kim, S. K., Cho, S.-G. and Kim, K. P. (2009). Downregulation of PLK-1 expression in kaempferol-induced apoptosis of MCF-7 cells. *European Journal of Pharmacology*, 611(1–3): 17-21.
- Kang, K. S., Wang, P., Yamabe, N., Fukui, M., Jay, T. and Zhu, B. T. (2010). Docosahexaenoic acid Induces apoptosis in MCF-7 cells *in vitro* and *in vivo* via reactive oxygen species formation and caspase 8 activation. *PLoS ONE*, 5(4): 1-13.
- Karamaæ, M., Kosiñska, A. and Pegg, R. B. (2006). Content of galic acid in selected plant extracts. *Polish Journal of Food and Nutrition Sciences*, 15(1): 55-58.
- Karin, M. (2006). Nuclear factor-[kappa]B in cancer development and progression. *Nature*, 441(7092): 431-436.
- Karin, M. and Ben-Neriah, Y. (2000). Phosphorylation meets ubiquitination: the control of NF-[kappa]B activity. *Annual Review of Immunology*, 18: 621-663.
- Kashiwada, Y., Nonaka, G.-i., Nishioka, I., Chang, J.-J. and Lee, K.-H. (1992). Antitumor agents, 129. Tannins and related compounds as selective cytotoxic agents. *Journal of Natural Products*, 55(8): 1033-1043.
- Kasibhatla, S. and Tseng, B. (2003). Why target apoptosis in cancer treatment? *Molecular Cancer Therapeutics*, 2(6): 573-580.
- Kaufmann, S. H. and Gores, G. J. (2000). Apoptosis in cancer: cause and cure. *BioEssays*, 22(11): 1007-1017.
- Kaur, R., Kapoor, K. and Kaur, H. (2011). Plants as a source of anticancer agents. *Journal of Natural Product Plant Resources*, 1(1): 119-124.
- Kawai, M., Hirano, T., Higa, S., Arimitsu, J., Maruta, M., Kuwahara, Y., Ohkawara, T., Hagiwara, K., Yamadori, T., Shima, Y., Ogata, A., Kawase, I. and Tanaka, T. (2007). Flavonoids and related compounds as anti-allergic substances. *Allergology International*, 56(2): 113-123.
- Keen, J. C. and Davidson, N. E. (2003). The biology of breast carcinoma. *Cancer*, 97(S3): 825-833.
- Kennecke, H., Yerushalmi, R., Woods, R., Cheang, M. C. U., Voduc, D., Speers, C. H., Nielsen, T. O. and Gelmon, K. (2010). Metastatic behavior of breast cancer subtypes. *Journal of Clinical Oncology*, 28(20): 3271-3277.
- Kerr, J. F., Wyllie, A. H. and Currie. (1972). Apoptosis: a basic biological phenomenon with wide-ranging implications in tissue kinetics. *British Journal of Cancer*, 26(4): 239-257.
- Kerrigan, R. A., Craven, L. A. and Dunlop, C. R. (2011). *Dilleniaceae*. P. S. Short & I. D. Cowie (Eds.). (Vol. 1). Australia: Northern Territory Herbarium.

- Khandelwal, N., Simpson, J., Taylor, G., Rafique, S., Whitehouse, A., Hiscox, J. and Stark, L. A. (2011). Nucleolar NF-[kappa]B/RelA mediates apoptosis by causing cytoplasmic relocalization of nucleophosmin. *Cell Death and Differentiation*, 18(12): 1889-1903.
- Khlebnikov, A. I., Schepetkin, I. A., Domina, N. G., Kirpotina, L. N. and Quinn, M. T. (2007). Improved quantitative structure–activity relationship models to predict antioxidant activity of flavonoids in chemical, enzymatic, and cellular systems. *Bioorganic and Medicinal Chemistry*, 15(4): 1749-1770.
- Kholodenko, B. N., Hancock, J. F. and Kolch, W. (2010). Signalling ballet in space and time. *Nature Reviews Molecular Cell Biology*, 11: 414-426.
- Khosravi-Far, R. and Esposti, M. D. (2004). Death receptor signals to mitochondria. *Cancer Biology and Therapy*, 3(11): 1051-1057.
- Kim, A. H., Khursigara, G., Sun, X., Franke, T. F. and Chao, M. V. (2001). Akt phosphorylates and negatively regulates apoptosis signal-regulating kinase 1. *Molecular and Cellular Biology*, 21(3): 893-901.
- Kim, J. H., Kang, M. J., Park, C. U., Kwak, H. J., Hwang, Y. and Koh, G. Y. (1999). Amplified CDK2 and cdc2 activities in primary colorectal carcinoma. *Cancer*, 85(3): 546-553.
- Kim, K. H., Choi, S. U. and Lee, K. R. (2010). Bioactivity-guided isolation of cytotoxic triterpenoids from the trunk of *Berberis koreana*. *Bioorganic and Medicinal Chemistry Letters*, 20(6): 1944-1947.
- Kim, S. J., Ju, J. W., Oh, C. D., Yoon, Y. M., Song, W. K., Kim, J. H., Yoo, Y. J., Bang, O. S., Kang, S. S. and Chun, J. S. (2002). ERK-1/2 and p38 kinase oppositely regulate nitric oxide-induced apoptosis of chondrocytes in association with p53, caspase-3, and differentiation status. *Journal of Biological Chemistry*, 277(2): 1332-1339.
- Kinnula, V. L. and Crapo, J. D. (2004). Superoxide dismutases in malignant cells and human tumors. *Free Radical Biology and Medicine*, 36(6): 718-744.
- Kogel, D. and Prehn, J. H. M. (2003). *Caspase-independent cell death mechanisms*. W. LosM (Ed.). New York: Kluwer Academic Press.
- Kohno, M. and Pouyssegur, J. (2006). Targeting the ERK signaling pathway in cancer therapy. *Annals of Medicine*, 38(3): 200-211.
- Kolomeichuk, S. N., Terrano, D. T., Lyle, C. S., Sabapathy, K. and Chambers, T. C. (2008). Distinct signaling pathways of microtubule inhibitors-vinblastine and Taxol induce JNK-dependent cell death but through AP-1-dependent and AP-1-independent mechanisms, respectively. *FEBS Journal*, 275(8): 1889-1899.
- Kong, Q., Beel, J. A. and Lillehei, K. O. (2000). A threshold concept for cancer therapy. *Medical Hypotheses*, 55(1): 29-35.

- Konishi, H., Matsuzaki, H., Tanaka, M., Takemura, Y., Kuroda, S., Ono, Y. and Kikkawa, U. (1997). Activation of protein kinase B (Akt/RAC-protein kinase) by cellular stress and its association with heat shock protein Hsp27. *FEBS Letters*, 410(2-3): 493-498.
- Koroleva, O., Torkova, A., Nikolaev, I., Khrameeva, E., Fedorova, T., Tsentalovich, M. and Amarowicz, R. (2014). Evaluation of the antiradical properties of phenolic acids. *International Journal of Molecular Sciences*, 15(9): 16351-16380.
- Kroemer, G., Dallaporta, B. and Resche-Rigon, M. (1998). The mitochondrial death/life regulator in apoptosis and necrosis. *Annual Review of Physiology*, 60: 619-642.
- Kroemer, G., El-Deiry, W. S., Golstein, P., Peter, M. E., Vaux, D., Vandenabeele, P., Zhivotovsky, B., Blagosklonny, M. V., Malorni, W., Knight, R. A., Piacentini, M., Nagata, S. and Melino, G. (2005). Classification of cell death: recommendations of the Nomenclature Committee on Cell Death. *Cell Death and Differentiation*, 12 (2): 1463-1467.
- Kroemer, G., Galluzzi, L. and Brenner, C. (2007). Mitochondrial membrane permeabilization in cell death. *Physiological Reviews*, 87(1): 99-163.
- Kroemer, G., Galluzzi, L., Vandenabeele, P., Abrams, J., Alnemri, E. S., Baehrecke, E. H., Blagosklonny, M. V., El-Deiry, W. S., Golstein, P., Green, D. R., Hengartner, M., Knight, R. A., Kumar, S., Lipton, S. A., Malorni, W., Nunez, G., Peter, M. E., Tschopp, J., Yuan, J., Piacentini, M., Zhivotovsky, B. and Melino, G. (2009). Classification of cell death: recommendations of the Nomenclature Committee on Cell Death 2009. *Cell Death and Differentiation*, 16(1): 3-11.
- Kumar, S., Kumar, V. and Prakash, O. (2011). Antidiabetic and antihyperlipidemic effects of *Dillenia indica* (L.) leaves extract. *Brazilian Journal of Pharmaceutical Sciences*, 47(2): 373-378.
- Kung, A. L., Zetterberg, A., Sherwood, S. W. and Schimke, R. T. (1990). Cytotoxic effects of cell cycle phase specific agents: result of cell cycle perturbation. *Cancer Research*, 50(22): 7307-7317.
- Kuttan, G., Kumar, K. B., Guruvayoorappan, C. and Kuttan, R. (2007). Antitumor, anti-invasion, and antimetastatic effects of curcumin. *Advances in Experimental Medicine and Biology*, 595: 173-184.
- Kyriakis, J. M. and Avruch, J. (2001). Mammalian mitogen-activated protein kinase signal transduction pathways activated by stress and inflammation. *Physiological Reviews*, 81(2): 807-869.
- Lahkar, M., Thakuria, B. and Pathak, P. (2010). A study of the anxiolytic-like activity of *Dillenia Indica* Linn. leaves in experimental models of anxiety in mice. *International Journal of Pharmacology*, 9(2): 1-3.

- Lamoral-Theys, D., Pottier, L., Dufrasne, F., Neve, J., Dubois, J., Kornienko, A., Kiss, R. and Ingrassia, L. (2010). Natural polyphenols that display anticancer properties through inhibition of kinase activity. *Current Medicinal Chemistry*, 17(9): 812-825
- Lawlor, M. A. and Alessi, D. R. (2001). PKB/Akt: a key mediator of cell proliferation, survival and insulin responses? *Journal of Cell Science*, 114(16): 2903-2910.
- Lee, E., Moon, B.-H., Park, Y., Hong, S., Lee, S., Lee, Y. and Lim, Y. (2008). Effects of hydroxy and methoxy substituents on NMR data in flavonols. *Bulletin of the Korean Chemical Society*, 29(2): 507-510.
- Lee, H., Lee, J., Kim, S., Nho, C., Jung, S., Song, D.-G., Kim, C. and Pan, C.-H. (2010). Inhibitory effects of dicaffeoylquinic acids from *Artemisia dubia* on aldo-keto reductase family 1b10. *Journal of the Korean Society for Applied Biological Chemistry*, 53(6): 826-830.
- Leek, R. D., Kaklamani, L., Pezzella, F., Gatter, K. C. and Harris, A. L. (1994). Bcl-2 in normal human breast and carcinoma, association with oestrogen receptor-positive, epidermal growth factor receptor-negative tumours and *in situ* cancer. *British Journal of Cancer*, 69(1): 135-139.
- Leist, M. and Jaattela, M. (2003). *Caspase-independent cell death*. S. Grimm (Ed.). United Kingdom: BIOS Scientific Publishers Ltd.
- Levenson, A. S. and Jordan, V. C. (1995). MCF-7: the first hormone-responsive breast cancer cell line. *Cancer Research*, 57: 3071-3078.
- Li, N. and Karin, M. (1999). Is NF- $\kappa$ B the sensor of oxidative stress? *FASEB Journal*, 13(10): 1137-1143.
- Li, X., Wang, X., Chen, D. and Chen, S. (2011). Antioxidant activity and mechanism of protocatechuic acid *in vitro*. *Functional Foods in Health and Disease*, 7: 232-244.
- Lima, C. C., Lemos, R. P. L. and Conserva, L. M. (2014). Dilleniaceae family: an overview of its ethnomedicinal uses, biological and phytochemical profile. *Journal of Pharmacognosy and Phytochemistry*, 3(2): 181-204.
- Lin, A. and Dibling, B. (2002). The true face of JNK activation in apoptosis. *Aging Cell*, 1(2): 112-116.
- Ling, W. H. and Jones, P. J. H. (1995). Dietary phytosterols: a review of metabolism, benefits and side effects. *Life Sciences*, 57(3): 195-206.
- Liu, C., Gong, K., Mao, X. and Li, W. (2011). Tetrandrine induces apoptosis by activating reactive oxygen species and repressing Akt activity in human hepatocellular carcinoma. *International Journal of Cancer*, 129(6): 1519-1531.

- Liu, P., Begley, M., Michowski, W., Inuzuka, H., Ginzberg, M., Gao, D., Tsou, P., Gan, W., Papa, A., Kim, B. M., Wan, L., Singh, A., Zhai, B., Yuan, M., Wang, Z., Gygi, S. P., Lee, T. H., Lu, K.-P., Toker, A., Pandolfi, P. P., Asara, J. M., Kirschner, M. W., Sicinski, P., Cantley, L. and Wei, W. (2014). Cell-cycle-regulated activation of Akt kinase by phosphorylation at its carboxyl terminus. *Nature*, 508(7497): 541-545.
- Lobo, V., Patil, A., Phatak, A. and Chandra, N. (2010). Free radicals, antioxidants and functional foods: impact on human health. *Pharmacognosy Reviews*, 4(8): 118-126.
- Lodish, H., Berk, A., Zipursky, S. L., Matsudaira, P., Baltimore, D. and Darnell, J. (2000). *Molecular Cell Biology*. W. H. Freeman (Ed.), (Fourth edition). New York: NCBI Bookshelf.
- Loeffler, M., Daugas, E., Susin, S. A., Zamzami, N., Metivier, D., Nieminen, A. L., Brothers, G., Penninger, J. M. A. and Kroemer, G. (2001). Dominant cell death induction by extramitochondrially targeted apoptosis-inducing factor. *Journal of Federation of American Societies for Experimental Biology*, 15(3): 758-767.
- Losa, J. H., Cobo, C. P., Viniegra, J. G., Lobo, V. J. S.-A., Cajal, S. R. Y. and Sanchez-Prieto, R. (2003). Role of the p38 MAPK pathway in cisplatin-based therapy. *Oncogene*, 22(26): 3998-4006.
- Lowe, S. W. and Lin, A. W. (2000). Apoptosis in cancer. *Carcinogenesis*, 21(3): 485-495.
- Lu, C. X., Fan, T. J., Hu, G. B. and Cong, R. S. (2003). Apoptosis-inducing factor and apoptosis. *Sheng Wu Hua Xue Yu Sheng Wu Wu Li Xue Bao (Shanghai)*, 35(10): 881-885.
- Lu, S. and Wang, J. (2014). Homoharringtonine and omacetaxine for myeloid hematological malignancies. *Journal of Hematology and Oncology*, 7(2): 1-10.
- Lu, Z. and Xu, S. (2006). ERK1/2 MAP kinases in cell survival and apoptosis. *IUBMB Life*, 58(11): 621-631.
- Luo, H., Daddysman, M., Rankin, G., Jiang, B.-H. and Chen, Y. (2010). Kaempferol enhances cisplatin's effect on ovarian cancer cells through promoting apoptosis caused by down regulation of cMyc. *Cancer Cell International*, 10(1): 1-16.
- Lüpertz, R., Wäjen, W., Kahl, R. and Chovolou, Y. (2010). Dose- and time-dependent effects of doxorubicin on cytotoxicity, cell cycle and apoptotic cell death in human colon cancer cells. *Toxicology*, 271(3): 115-121.
- Ly, J. D., Grubb, D. R. and Lawen, A. (2003). The mitochondrial membrane potential in apoptosis; an update. *Apoptosis*, 8(2): 115-128.

- Ma, Q. (2010). Transcriptional responses to oxidative stress: pathological and toxicological implications. *Pharmacology and Therapeutics*, 125(3): 376-393.
- Macahig, R. A., Matsunami, K. and Otsuka, H. (2011). Chemical studies on an endemic Philippine plant: sulfated glucoside and seco-A-ring triterpenoids from *Dillenia philippinensis*. *Chemical and Pharmaceutical Bulletin*, 59(3): 397-401.
- Malumbres, M. and Barbacid, M. (2007). Cell cycle kinases in cancer. *Current Opinion in Genetics and Development*, 17(1): 60-65.
- Manning, B. D. and Cantley, L. C. (2007). AKT/PKB signaling: navigating downstream. *Cell*, 129(7): 1261-1274.
- Marnett, L. J. (2000). Oxyradicals and DNA damage. *Carcinogenesis*, 21(3): 361-370.
- Martin, A. G. (2010). NFkB anti-apoptotic or pro-apoptotic, maybe both. *Cell Cycle*, 9(16): 3131-3132.
- Martindale, J. L. and Holbrook, N. J. (2002). Cellular response to oxidative stress: signaling for suicide and survival. *Journal of Cellular Physiology*, 192: 1-15.
- Martins, N. M., Santos, N. A., Curti, C., Bianchi, M. L. and Santos, A. C. (2008). Cisplatin induces mitochondrial oxidative stress with resultant energetic metabolism impairment, membrane rigidification and apoptosis in rat liver. *Journal of Applied Toxicology*, 28(3): 337-344.
- Massaoka, M. H., Matsuo, A. L., Figueiredo, C. R., Farias, C. F. and Girola, N. (2012). Jacaranone induces apoptosis in melanoma cells via ROS-mediated downregulation of Akt and p38 MAPK activation and displays antitumor activity *in vivo*. *PLoS ONE*, 7(6): 1-11.
- Mat Salleh, K. and Latiff, A. (2002). *Tumbuhan ubatan Malaysia*: Bangi: Universiti Kebangsaan Malaysia.
- Matsuo, M., Sasaki, N., Saga, K. and Kaneko, T. (2005). Cytotoxicity of flavonoids toward cultured normal human cells. *Biological and Pharmaceutical Bulletin*, 28(2): 253-259.
- Mauro, C., Zazzeroni, F., Papa, S., Bubici, C. and Franzoso, G. (2009). The NF-kappaB transcription factor pathway as a therapeutic target in cancer: methods for detection of NF-kappaB activity. *Methods in Molecular Biology*, 512: 169-207.
- McCafferty, M. P. J., Healy, N. A. and Kerin, M. J. (2009). Breast cancer subtypes and molecular biomarkers. *Diagnostic Histopathology*, 15(10): 485-489.
- McGill, G. and Fisher, D. E. (1997). Apoptosis in tumorigenesis and cancer therapy. *Frontiers in Bioscience*, 2: 353-379.

- Micheli, A., Baili, P., Quinn, M., Mugno, E., Capocaccia, R. and Grosclaude, P. (2003). Life expectancy and cancer survival in the EUROCARE-3 cancer registry areas. *Annals of Oncology*, 14(5): 28-40.
- Middleton, E., Jr., Kandaswami, C. and Theoharides, T. C. (2000). The effects of plant flavonoids on mammalian cells: implications for inflammation, heart disease, and cancer. *Pharmacological Reviews*, 52(4): 673-751.
- Migliato, K. F., Chiosini, M. A., Mendonça, F. A., Esquisatto, M. A., Salgado, H. R. and Santos, G. M. T. (2011). Effect of glycolic extract of *Dillenia indica* L. combined with microcurrent stimulation on experimental lesions in Wistar rats. *Wounds*, 23(5): 111-120.
- Miller-Keane and O'Toole, M. (2005). *Miller-Keane encyclopedia and dictionary of medicine, nursing and allied health* (Seventh edition). New Jersey: Saunders, Elsevier Incorporation.
- Miller, T. M., Moulder, K. L., Knudson, C. M., Creedon, D. J., Deshmukh, M., Korsmeyer, S. J. and Johnson, E. M., Jr. (1997). Bax deletion further orders the cell death pathway in cerebellar granule cells and suggests a caspase-independent pathway to cell death. *Journal of Cell Biology*, 139(1): 205-217.
- Mitra, S. K. and Kannan, R. (2007). A note on unintentional adulterations in ayurvedic herbs. *Ethnobotanical Leaflets*, 2007(11): 11-15.
- Miyamoto, K., Murayama, T., Hatano, T., Yoshida, T. and Okuda, T. (1999). *Host-mediated anticancer activities of tannins*. G. Gross, R. Hemingway, T. Yoshida & S. Branham (Eds.). (Vol. 66). New York: Springer US.
- Moll, R., Franke, W. W., Schiller, D. L., Geiger, B. and Krepler, R. (1982). The catalog of human cytokeratins: patterns of expression in normal epithelia, tumors and cultured cells. *Cell*, 31(1): 11-24.
- Montaigne, D., Hurt, C. and Neviere, R. (2012). Mitochondria death/survival signaling pathways in cardiotoxicity induced by anthracyclines and anticancer-targeted therapies. *Biochemistry Research International*, 2012: 1-12.
- Mooney, L. M., Al-Sakkaf, K. A., Brown, B. L. and Dobson, P. R. (2002). Apoptotic mechanisms in T47D and MCF-7 human breast cancer cells. *British Journal of Cancer*, 87(8): 909-917.
- Moreau, R. A., Whitaker, B. D. and Hicks, K. B. (2002). Phytosterols, phytostanols, and their conjugates in foods: structural diversity, quantitative analysis, and health-promoting uses. *Progress in Lipid Research*, 41(6): 457-500.
- Mori, K., Shibanuma, M. and Nose, K. (2004). Invasive potential induced under long-term oxidative stress in mammary epithelial cells. *Cancer Research*, 64(20): 7464-7472.

- Moriguchi, T., Kawasaki, H., Matsuda, S., Gotoh, Y. and Nishida, E. (1995). Evidence for multiple activators for stress-activated protein kinase/c-Jun amino-terminal kinases. Existence of novel activators. *Journal of Biological Chemistry*, 270(22): 12969-12972.
- Mosmann, T. (1983). Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *Journal of Immunological Methods*, 65(1-2): 55-63.
- Motzer, R. J., Escudier, B., Oudard, S., Hutson, T. E., Porta, C., Bracarda, S., Grünwald, V., Thompson, J. A., Figlin, R. A., Hollaender, N., Urbanowitz, G., Berg, W. J., Kay, A., Lebwohl, D. and Ravaud, A. (2008). Efficacy of everolimus in advanced renal cell carcinoma: a double-blind, randomised, placebo-controlled Phase III trial. *Lancet*, 372(9637): 449-456.
- Muliawan, S. Y. (2008). Effect of *Dillenia suffruticosa* extract on dengue virus type 2 replication. *Universa Medicina*, 27(1): 1-5.
- Müller, M., Wilder, S., Bannasch, D., Israeli, D., Lehlbach, K., Li-Weber, M., Friedman, S. L., Galle, P. R., Stremmel, W., Oren, M. and Krammer, P. H. (1998). p53 activates the CD95 (APO-1/Fas) gene in response to DNA damage by anticancer drugs. *Journal of Experimental Medicine*, 188(11): 2033-2045.
- Mundhe, K. S., Kale, A. A., Gaikwad, S. A., Deshpande, N. R. and Kashalkar, R. V. (2011). Evaluation of phenol, flavonoid contents and antioxidant activity of *Polyalthia longifolia*. *Journal of Chemical and Pharmaceutical Research*, 3(1): 764-769.
- Munirah, M. A., Siti-Aishah, M. A., Reena, M. Z., Sharifah, N. A., Rohaizak, M., Norlia, A., Rafie, M. K., Asmiati, A., Hisham, A., Fuad, I., Shahrun, N. S. and Das, S. (2011). Identification of different subtypes of breast cancer using tissue microarray. *Romanian Journal of Morphology and Embryology*, 52(2): 669-677.
- Murai, H., Hiragami, F., Kawamura, K., Motoda, H., Koike, Y., Inoue, S., Kumagishi, K., Ohtsuka, A. and Kano, Y. (2010). Differential response of heat-shock-induced p38 MAPK and JNK activity in PC12 mutant and PC12 parental cells for differentiation and apoptosis. *Acta Medica Okayama*, 64(1): 55-62.
- Musa, S. N. (2012). *In vitro evaluation of antioxidant and cytotoxic properties of Dillenia suffruticosa extract*. Bachelor Thesis, Universiti Putra Malaysia, Malaysia.
- Mustafa, R. A., Abdul Hamid, A., Mohamed, S. and Bakar, F. A. (2010). Total phenolic compounds, flavonoids, and radical scavenging activity of 21 selected tropical plants. *Journal of Food Science*, 75(1): C28-C35.

- Muttarak, M., Lerttumnongtum, P., Somwangjaroen, A. and Chaiwun, B. (2006). Phyllodes tumour of the breast. *Biomedical Imaging and Intervention Journal*, 2(2): 1-5.
- Nascimento, G. G. F., Locatelli, J., Freitas, P. C. and Silva, G. L. (2000). Antibacterial activity of plant extracts and phytochemicals on antibiotic-resistant bacteria. *Brazilian Journal of Microbiology*, 31: 247-256.
- Nashed, B., Yeganeh, B., HayGlass, K. T. and Moghadasian, M. H. (2005). Antiatherogenic effects of dietary plant sterols are associated with inhibition of proinflammatory cytokine production in Apo E-KO mice. *Journal of Nutrition*, 135(10): 2438-2444.
- Nazarewicz, R. R., Zenebe, W. J., Parihar, A., Larson, S. K., Alidema, E., Choi, J. and Ghafourifar, P. (2007). Tamoxifen induces oxidative stress and mitochondrial apoptosis via stimulating mitochondrial nitric oxide synthase. *Cancer Research*, 67(3): 1282-1290.
- Nazaruk, J. and Borzym-Kluczyk, M. (2014). The role of triterpenes in the management of diabetes mellitus and its complications. *Phytochemistry Reviews*: 1-16.
- Nelson, A. C. and Kursar, T. A. (1999). Interactions among plant defense compounds: a method for analysis. *Chemoecology*, 9(2): 81-92.
- Newcomb, E. W. (2004). Flavopiridol: pleiotropic biological effects enhance its anti-cancer activity. *Anticancer Drugs*, 15(5): 411-419.
- Newman, D. J., Cragg, G. M. and Snader, K. M. (2003). Natural products as sources of new drugs over the period 1981-2002. *Journal of Natural Products*, 66(7): 1022-1037.
- Nguyen-Pouplin, J., Tran, H., Tran, H., Phan, T. A., Dolecek, C., Farrar, J., Tran, T. H., Caron, P., Bodo, B. and Grellier, P. (2007). Antimalarial and cytotoxic activities of ethnopharmacologically selected medicinal plants from South Vietnam. *Journal of Ethnopharmacology*, 109(3): 417-427.
- Nguyen, D.-M.-C., Seo, D.-J., Kim, K.-Y., Park, R.-D., Kim, D.-H., Han, Y.-S., Kim, T.-H. and Jung, W.-J. (2013). Nematicidal activity of 3,4-dihydroxybenzoic acid purified from *Terminalia nigrovenulosa* bark against *Meloidogyne incognita*. *Microbial Pathogenesis*, 59: 52-59.
- Nick, A., Rali, T. and Sticher, O. (1995a). Biological screening of traditional medicinal plants from Papua New Guinea. *Journal of Ethnopharmacology*, 49(3): 147-156.
- Nick, A., Wright, A. D., Rali, T. and Sticher, O. (1995b). Antibacterial triterpenoids from *Dillenia papuana* and their structure-activity relationships. *Phytochemistry*, 40(6): 1691-1695.

- Nick, A., Wright, A. D., Sticher, O. and Rali, T. (1994). Antibacterial triterpenoid acids from *Dillenia papuana*. *Journal of Natural Product* 57: 1245-1250.
- Niki, E., Noguchi, N., Tsuchihashi, H. and Gotoh, N. (1995). Interaction among vitamin C, vitamin E, and beta-carotene. *American Journal of Clinical Nutrition*, 62(6): 1322S-1326S.
- Noble, R. L. (1990). The discovery of the vinca alkaloids-chemotherapeutic agents against cancer. *Biochemistry and Cell Biology*, 68(12): 1344-1351.
- Noda, N. and Wakasugi, H. (2001). Cancer and oxidative stress. *Journal of the Japan Medical Association*, 124(11): 1571-1574.
- Norbury, C. and Nurse, P. (1992). Animal cell cycles and their control. *Annual Review of Biochemistry*, 61: 441-468.
- Nordberg, J. and Arnér, E. S. J. (2001). Reactive oxygen species, antioxidants, and the mammalian thioredoxin system. *Free Radical Biology and Medicine*, 31(11): 1287-1312.
- Nunez, R. (2001). DNA measurement and cell cycle analysis by flow cytometry. *Current Issues in Molecular Biology*, 3(3): 67-70.
- Nylandsted, J., Rohde, M., Brand, K., Bastholm, L., Elling, F. and Jäättelä, M. (2000). Selective depletion of heat shock protein 70 (Hsp70) activates a tumor-specific death program that is independent of caspases and bypasses Bcl-2. *Proceedings of the National Academy of Sciences*, 97(14): 7871-7876.
- Nyman, U., Joshi, P., Madsen, L. B., Pedersen, T. B., Pinstrup, M., Rajasekharan, S., George, V. and Pushpangadan, P. (1998). Ethnomedical information and *in vitro* screening for angiotensin-converting enzyme inhibition of plants utilized as traditional medicines in Gujarat, Rajasthan and Kerala (India). *Journal of Ethnopharmacology*, 60(3): 247-263.
- O'Brien, M. E., Talbot, D. C. and Smith, I. E. (1993). Carboplatin in the treatment of advanced breast cancer: a Phase II study using a pharmacokinetically guided dose schedule. *Journal of Clinical Oncology*, 11(11): 2112-2117.
- Ohashi, T., Mizutani, A., Murakami, A., Kojo, S., Ishii, T. and Taketani, S. (2002). Rapid oxidation of dichlorodihydrofluorescein with heme and hemoproteins: formation of the fluorescein is independent of the generation of reactive oxygen species. *FEBS Letters*, 511(1-3): 21-27.
- Oida, K., Matsuda, A., Jung, K., Xia, Y., Jang, H., Amagai, Y., Ahn, G., Nishikawa, S., Ishizaka, S., Jensen-Jarolim, E., Matsuda, H. and Tanaka, A. (2014). Nuclear factor-κB plays a critical role in both intrinsic and acquired resistance against endocrine therapy in human breast cancer cells. *Scientific Reports*, 4: 1-8.

- Ortega, S., Malumbres, M. and Barbacid, M. (2002). Cyclin D-dependent kinases, INK4 inhibitors and cancer. *Biochimica et Biophysica Acta*, 1602(1): 73-87.
- Ostlund, R. E., Racette, S. B., Okeke, A. and Stenson, W. F. (2002). Phytosterols that are naturally present in commercial corn oil significantly reduce cholesterol absorption in humans. *American Journal of Clinical Nutrition*, 75(6): 1000-1004.
- Ostrakhovitch, E. A. and Cherian, M. G. (2005). Inhibition of extracellular signal regulated kinase (ERK) leads to apoptosis inducing factor (AIF) mediated apoptosis in epithelial breast cancer cells: the lack of effect of ERK in p53 mediated copper induced apoptosis. *Journal of Cellular Biochemistry*, 95(6): 1120-1134.
- Otake, Y. and Walle, T. (2002). Oxidation of the flavonoids galangin and kaempferide by human liver microsomes and CYP1A1, CYP1A2, and CYP2C9. *Drug Metabolism and Disposition*, 30(2): 103-105.
- Øverby, A., Zhao, C.-M. and Chen, D. (2014). Plant phytochemicals: potential anticancer agents against gastric cancer. *Current Opinion in Pharmacology*, 19: 6-10.
- Ozsaran, Z. and Alanyali, S. D. (2013). Staging of breast cancer. A. Haydaroglu & G. Ozigit (Eds.). *Principles and practice of modern radiotherapy techniques in breast cancer*. New York: Springer Science and Business Media New York
- Paraiso, K. H. T., Der Kooi, K. V., Messina, J. L. and Smalley, K. S. M. (2010). Measurement of constitutive MAPK and PI3K/AKT signaling activity in human cancer cell lines. E. Cadena & L. Packer (Eds.). *Methods in enzymology* (Vol 484, pp 549-567). Netherlands: Academic Press.
- Park, J. S., Carter, S., Reardon, D. B., Schmidt-Ullrich, R., Dent, P. and Fisher, P. B. (1999). Roles for basal and stimulated p21(Cip-1/WAF1/MDA6) expression and mitogen-activated protein kinase signaling in radiation-induced cell cycle checkpoint control in carcinoma cells. *Molecular Biology of the Cell*, 10(12): 4231-4246.
- Park, K.-R., Nam, D., Yun, H.-M., Lee, S.-G., Jang, H.-J., Sethi, G., Cho, S. K. and Ahn, K. S. (2011).  $\beta$ -caryophyllene oxide inhibits growth and induces apoptosis through the suppression of PI3K/AKT/mTOR/S6K1 pathways and ROS-mediated MAPKs activation. *Cancer Letters*, 312(2): 178-188.
- Park, M.-T. and Lee, S.-J. (2002). Cell cycle and cancer. *Journal of Biochemistry and Molecular Biology*, 36(11): 60-65.
- Patra, R. C., Rautray, A. K. and Swarup, D. (2011). Oxidative stress in lead and cadmium toxicity and its amelioration. *Veterinary Medicine International*, 2011: 1-9.

- Patra, R. C., Swarup, D. and Dwivedi, S. K. (2001). Antioxidant effects of alpha tocopherol, ascorbic acid and L-methionine on lead induced oxidative stress to the liver, kidney and brain in rats. *Toxicology*, 162(2): 81-88.
- Pavanarasivam, G. and Sultanbawa, M. U. S. (1974). Chemical investigation of ceylonese plants. IX. Betulinic acid in the Dilleniaceae and a review of its natural distribution. *Phytochemistry*, 13: 2002-2006.
- Pavanarasivam, G. and Sultanbawa, M. U. S. (1975). Flavonoids of some Dilleniaceae species. *Phytochemistry*, 14(44): 1127-1128.
- Peddi, P. F., Ellis, M. J. and Ma, C. (2012). Molecular basis of triple negative breast cancer and implications for therapy. *International Journal of Breast Cancer*, 2012: 1-7.
- Peek, H. W., Halkes, S. B. A., Mes, J. J. and Landman, W. J. M. (2013). *In vivo* screening of four phytochemicals/extracts and a fungal immunomodulatory protein against an *Eimeria acervulina* infection in broilers. *Veterinary Quarterly*, 33(3): 132-138.
- Peng, J., Lapolla, S. M., Zhang, Z. and Lin, J. (2009). The Bax BH3 peptide H2-H3 promotes apoptosis by inhibiting Bcl-2's pore-forming and anti-Bax activities in the membrane. *Sheng Wu Yi Xue Gong Cheng Xue Za Zhi*, 26(4): 829-835.
- Pentikainen, V., Suomalainen, L., Erkkila, K., Martelin, E., Parvinen, M., Pentikainen, M. O. and Dunkel, L. (2002). Nuclear factor-kappa B activation in human testicular apoptosis. *American Journal of Pathology*, 160(1): 205-218.
- Pereira, D. F., Zanon, R. B., dos Santos, M., Boligon, A. A. and Athayde, M. L. (2012). Antioxidant activities and triterpenoids isolated from *Alternanthera brasiliiana* (L.) Kuntze leaves. *Natural Product Research*, 27(18): 1660-1663.
- Perou, C. M., Sorlie, T., Eisen, M. B., van de Rijn, M., Jeffrey, S. S., Rees, C. A., Pollack, J. R., Ross, D. T., Johnsen, H., Akslen, L. A., Fluge, O., Pergamenschikov, A., Williams, C., Zhu, S. X., Lonning, P. E., Borresen-Dale, A.-L., Brown, P. O. and Botstein, D. (2000). Molecular portraits of human breast tumours. *Nature*, 406(6797): 747-752.
- Petit, P. X., Lecoeur, H., Zorn, E., Dauguet, C., Mignotte, B. and Gougeon, M. L. (1995). Alterations in mitochondrial structure and function are early events of dexamethasone-induced thymocyte apoptosis. *Journal of Cell Biology*, 130(1): 157-167.
- Petronelli, A., Pannitteri, G. and Testa, U. (2009). Triterpenoids as new promising anticancer drugs. *Anticancer Drugs*, 20(10): 880-892.
- Piccolo, M. T. and Crispi, S. (2012). The dual role played by p21 may influence the apoptotic or anti-apoptotic fate in cancer. *Journal of Cancer Research Updates*, 1: 189-202.

- Polivka, J., Jr., Polivka, J., Rohan, V., Topolcan, O. and Ferda, J. (2012). New molecularly targeted therapies for glioblastoma multiforme. *Anticancer Research*, 32(7): 2935-2946.
- Polivka, J. J. and Janku, F. (2014). Molecular targets for cancer therapy in the PI3K/AKT/mTOR pathway. *Pharmacology and Therapeutics*, 142(2): 164-175.
- Poljsak, B., Suput, D. and Milisav, I. (2013). Achieving the balance between ROS and antioxidants: when to use the synthetic antioxidants. *Oxidative Medicine and Cellular Longevity*, 2013: 1-11.
- Polster, B. M., Basañez, G., Etxebarria, A., Hardwick, J. M. and Nicholls, D. G. (2005). Calpain I induces cleavage and release of apoptosis-inducing factor from isolated mitochondria. *Journal of Biological Chemistry*, 280(8): 6447-6454.
- Polyak, K. (2011). Heterogeneity in breast cancer. *Journal of Clinical Investigation*, 121(10): 3786-3788.
- Porter, A. G. and Jänicke, R. U. (1999). Emerging roles of caspase-3 in apoptosis. *Cell Death and Differentiation*, 6(2): 99-104.
- Potter, A. J., Gollahon, K. A., Palanca, B. J. A., Harbert, M. J., Choi, Y. M., Moskovitz, A. H., Potter, J. D. and Rabinovitch, P. S. (2002). Flow cytometric analysis of the cell cycle phase specificity of DNA damage induced by radiation, hydrogen peroxide and doxorubicin. *Carcinogenesis*, 23(3): 389-401.
- Pozarowski, P. and Darzynkiewicz, Z. (2004). Analysis of cell cycle by flow cytometry. A. Schönthal (Ed.). *Checkpoint controls and cancer* (Vol 281, pp 301-311). New York: Humana Press.
- Pozo-Guisado, E., Merino, J. M., Mulero-Navarro, S., Lorenzo-Benayas, M. J., Centeno, F., Alvarez-Barrientos, A. and Salguero, P. M. F. (2005). Resveratrol-induced apoptosis in MCF-7 human breast cancer cells involves a caspase-independent mechanism with downregulation of Bcl-2 and NF-κB. *International Journal of Cancer*, 115(1): 74-84.
- Pradhan, B. K. and Badola, H. K. (2008). Ethnomedicinal plant use by Lepcha tribe of Dzongu valley, bordering Khangchendzonga Biosphere Reserve, in North Sikkim, India. *Journal of Ethnobiology and Ethnomedicine*, 4: 1-18.
- Prasad, P. R. C., Reddy, C. S., Raza, S. H. and Dutt, C. B. S. (2008). Folklore medicinal plants of North Andaman Islands, India. *Fitoterapia*, 79(6): 458-464.
- Puglia, C. D. and Powell, S. R. (1984). Inhibition of cellular antioxidants: a possible mechanism of toxic cell injury. *Environmental Health Perspectives*, 57: 307-311.

- Ragasa, C. Y., Alimboyoguen, A. B. and Shen, C.-C. (2009). Antimicrobial triterpenes from *Dillenia philippinensis*. *Philippine Agricultural Scientist*, 4: 78-87.
- Rahal, A., Kumar, A., Singh, V., Yadav, B., Tiwari, R., Chakraborty, S. and Dhama, K. (2014). Oxidative stress, prooxidants, and antioxidants: the interplay. *BioMed Research International*, 2014: 1-19.
- Rahman, K. (2007). Studies on free radicals, antioxidants, and co-factors. *Journal of Clinical Interventions in Aging*, 2: 219-236.
- Rao, K. V. K., Schwartz, S. A., Nair, H. K., Aalinkeel, R., Mahajan, S., Chawda, R. and Nair, M. P. N. (2004). Plant derived products as a source of cellular growth inhibitory phytochemicals on PC-3M, DU-145 and LNCaP prostate cancer cell lines. *Current Science*, 87: 1585-1588.
- Rathee, P., Chaudhary, H., Rathee, S., Rathee, D., Kumar, V. and Kohli, K. (2009). Mechanism of action of flavonoids as anti-inflammatory agents: a review. *Inflammation and Allergy Drug Targets*, 8(3): 229-235.
- Ravishankar, D., Rajora, A. K., Greco, F. and Osborn, H. M. I. (2013). Flavonoids as prospective compounds for anti-cancer therapy. *International Journal of Biochemistry and Cell Biology*, 45(12): 2821-2831.
- Reed, J. C. (1999). Dysregulation of apoptosis in cancer. *Journal of Clinical Oncology*, 17(9): 2941-2953.
- Reers, M., Smiley, S. T., Mottola-Hartshorn, C., Chen, A., Lin, M. and Chen, L. B. (1995). Mitochondrial membrane potential monitored by JC-1 dye. A. C. Giuseppe M. Attardi (Ed.), *Methods in enzymology* (Vol 260, pp 406-417). Netherlands: Academic Press.
- Reichert, J. M. (2003). Trends in development and approval times for new therapeutics in the United States. *Nature Reviews Drug Discovery*, 2(9): 695– 702.
- Reis-Filho, J. S., Simpson, P. T., Gale, T. and Lakhani, S. R. (2005). The molecular genetics of breast cancer: the contribution of comparative genomic hybridization. *Pathology-Research and Practice*, 201(11): 713-725.
- Reskin, S. J., Bellizzi, A., Cardone, R. A., Tommasino, M., Casavola, V. and Paradiso, A. (2003). Paclitaxel induces apoptosis via protein kinase A- and p38 mitogen-activated protein-dependent inhibition of the Na<sup>+</sup>/H<sup>+</sup> exchanger (NHE) NHE isoform 1 in human breast cancer cells. *Clinical Cancer Research*, 9(6): 2366-2373.
- Reuter, S., Gupta, S. C., Chaturvedi, M. M. and Aggarwal, B. B. (2010). Oxidative stress, inflammation, and cancer: how are they linked? *Free Radical Biology and Medicine*, 49(11): 1603-1616.

- Rice-Evans, C. A., Miller, N. J. and Paganga, G. (1996). Structure-antioxidant activity relationships of flavonoids and phenolic acids. *Free Radical Biology and Medicine*, 20(7): 933-956.
- Ridnour, L. A., Oberley, T. D. and Oberley, L. W. (2004). Tumor suppressive effects of MnSOD overexpression may involve imbalance in peroxide generation versus peroxide removal. *Antioxidants and Redox Signaling*, 6(3): 501-512.
- Riley, P. A. (1994). Free radicals in biology: oxidative stress and effects of ionizing radiation. *International Journal of Radiation Biology*, 65: 27-33.
- Rincón, S. V. D., Rousseau, Ratna, S. and Miller, W. H. (2002). Retinoic acid-induced growth arrest of MCF-7 cells involves the selective regulation of the IRS-1/PI 3-kinase/AKT pathway. *Oncogene*, 22: 3353-3360.
- Ríos, J.-L. (2015). Apoptotic activities of Mediterranean plant species. V. R. Watson & R. R. Preedy (Eds.). *Mediterranean diet* (pp 611-620). San Diego: Academic Press.
- Robbins, D. and Zhao, Y. (2012). Oxidative stress induced by MnSOD-p53 interaction: pro- or anti-tumorigenic? *Journal Signal Transduction*, 2012: 1-13.
- Robbins, R. J. (2003). Phenolic acids in foods: an overview of analytical methodology. *Journal of Agricultural and Food Chemistry*, 51(10): 2866-2887.
- Roberts, P. J. and Der, C. J. (2007). Targeting the Raf-MEK-ERK mitogen-activated protein kinase cascade for the treatment of cancer. *Oncogene*, 26(22): 3291-3310.
- Robertson, F. M., Bondy, M., Yang, W., Yamauchi, H., Wiggins, S., Kamrudin, S., Krishnamurthy, S., Le-Petross, H., Bidaut, L., Player, A. N., Barsky, S. H., Woodward, W. A., Buchholz, T., Lucci, A., Ueno, N. and Cristofanilli, M. M. (2010). Inflammatory breast cancer the disease, the biology, the treatment. *CA: A Cancer Journal for Clinicians*, 60: 351-375.
- Robinson, M. M. and Zhang, X. (2011). *The world medicines situation 2011. Traditional medicines: global situation, issues and challenges* (Third edition). Geneva: World Health Organization.
- Rodrigues, J. R., Charris, J., Camacho, J., Barazarte, A., Gamboa, N. and Antunes, F. (2012). Cytotoxic effects of N'-formyl-2-(5-nitrothiophen-2-yl) benzothiazole-6-carbohydrazide in human breast tumor cells by induction of oxidative stress. *Anticancer Research*, 32(7): 2721-2726.
- Romashkova, J. A. and Makarov, S. S. (1999). NF-κB is a target of AKT in anti-apoptotic PDGF signalling. *Nature*, 401: 86-90.

- Romond, E. H., Perez, E. A., Bryant, J., Suman, V. J., Geyer, C. E., Davidson, N. E., Tan-Chiu, E., Martino, S., Paik, S., Kaufman, P. A., Swain, S. M., Pisansky, T. M., Fehrenbacher, L., Kutteh, L. A., Vogel, V. G., Visscher, D. W., Yothers, G., Jenkins, R. B., Brown, A. M., Dakhil, S. R., Mamounas, E. P., Lingle, W. L., Klein, P. M., Ingle, J. N. and Wolmark, N. (2005). Trastuzumab plus adjuvant chemotherapy for operable HER2-positive breast cancer. *New England Journal of Medicine*, 353(16): 1673-1684.
- Rouzier, R., Perou, C. M., Symmans, W. F., Ibrahim, N., Cristofanilli, M., Anderson, K., Hess, K. R., Stec, J., Ayers, M., Wagner, P., Morandi, P., Fan, C., Rabiul, I., Ross, J. S., Hortobagyi, G. N. and Pusztai, L. (2005). Breast cancer molecular subtypes respond differently to preoperative chemotherapy. *Clinical Cancer Research*, 11(16): 5678-5685.
- Roy, S., Srivastava, R. and Shankar, S. (2010). Inhibition of PI3K/AKT and MAPK/ERK pathways causes activation of FOXO transcription factor, leading to cell cycle arrest and apoptosis in pancreatic cancer. *Journal of Molecular Signaling*, 5(1): 1-10.
- Rushworth, S. A., Murray, M. Y., Barrera, L. N., Heasman, S.-A., Zaitseva, L. and MacEwan, D. J. (2012). Understanding the role of miRNA in regulating NF- $\kappa$ B in blood cancer. *American Journal of Cancer Research*, 2(1): 65-74.
- Ryan, K. M., Ernst, M. K., Rice, N. R. and Vousden, K. H. (2000). Role of NF-[ $\kappa$ ]B in p53-mediated programmed cell death. *Nature*, 404(6780): 892-897.
- Ryan, M. J., Dudash, H. J., Docherty, M., Geronilla, K. B., Baker, B. A., Haff, G. G., Cutlip, R. G. and Alway, S. E. (2010). Vitamin E and C supplementation reduces oxidative stress, improves antioxidant enzymes and positive muscle work in chronically loaded muscles of aged rats. *Experimental Gerontology*, 45(11): 882-895.
- Ryu, D.-S., Lee, H.-S., Lee, G.-S. and Lee, D.-S. (2012). Effects of the ethyl acetate extract of *Orostachys japonicus* on induction of apoptosis through the p53-mediated signaling pathway in human gastric cancer Cells. *Biological and Pharmaceutical Bulletin*, 35(5): 660-665.
- Sahoo, N., Manchikanti, P. and Dey, S. (2010). Herbal drugs: standards and regulation. *Fitoterapia*, 81(6): 462-471.
- Said, Z. (2010). *In vitro cytotoxicity and in vivo anti-neoplastic properties of Dillenia suffruticosa water extract on cervical carcinogenesis*. Master Thesis, Universiti Putra Malaysia, Malaysia.
- Salakou, S., Kardamakis, D., Tsamandas, A. C., Zolota, V., Apostolakis, E., Tzelepi, V., Papathanasopoulos, P., Bonikos, D. S., Papapetropoulos, T., Petsas, T. and Dougenis, D. (2007). Increased Bax/Bcl-2 ratio up-regulates caspase-3 and increases apoptosis in the thymus of patients with myasthenia gravis. *In Vivo*, 21(1): 123-132.

- Salganik, R. I. (2001). The benefits and hazards of antioxidants: controlling apoptosis and other protective mechanisms in cancer patients and the human population. *Journal of the American College of Nutrition*, 20(5): 464S–472S.
- Salvioli, S., Ardizzone, A., Franceschi, C. and Cossarizza, A. (1997). JC-1, but not DiOC6(3) or rhodamine 123, is a reliable fluorescent probe to assess  $\Delta\Psi$  changes in intact cells: implications for studies on mitochondrial functionality during apoptosis. *FEBS Letters*, 411(1): 77-82.
- Sandal, T. (2002). Molecular aspects of the mammalian cell cycle and cancer. *Oncologist*, 7(1): 73-81.
- Sandhu, R., Parker, J. S., Jones, W. D., Livasy, C. A. and Coleman, W. B. (2010). Microarray-based gene expression profiling for molecular classification of breast cancer and identification of new targets for therapy. *Lab Medicine*, 41(6): 364-372.
- Sarker, S. D. and Nahar, L. (2012). *Natural products isolation*. J. M. Walker (Ed.), (Third edition). New York: Humana Press.
- Saroya, A. S. (2011). *Herbalism, phytochemistry and ethnopharmacology*. Florida: CRC Press.
- Sasidharan, S., Chen, Y., Saravanan, D., Sundram, K. M. and Yoga Latha, L. (2011). Extraction, isolation and characterization of bioactive compounds from plants' extracts. *African Journal of Traditional, Complementary, and Alternative Medicines*, 8(1): 1-10.
- Savill, J. (1997). Recognition and phagocytosis of cells undergoing apoptosis. *British Medical Bulletin*, 53(3): 491-508.
- Sax, J. K. and El-Deiry, W. S. (2003). p53 downstream targets and chemosensitivity. *Cell Death and Differentiation*, 10(4): 413-417.
- Saxena, M., Saxena, J., Nema, R., Singh, D. and Gupta, A. (2013). Phytochemistry of medicinal plants. *Journal of Pharmacognosy and Phytochemistry*, 1(6): 168-182.
- Schaeffer, H. J. and Weber, M. J. (1999). Mitogen-activated protein kinases: specific messages from ubiquitous messengers. *Molecular Cell Biology*, 19(4): 2435-2444.
- Schieven, G. L. (2005). The biology of p38 kinase: a central role in inflammation. *Current Topics in Medicinal Chemistry*, 5(10): 921-928.
- Schrader, K., Huai, J., Jockel, L., Oberle, C. and Borner, C. (2010). Non-caspase proteases: triggers or amplifiers of apoptosis? *Cellular and Molecular Life Sciences*, 67(10): 1607-1618.

- Sebolt-Leopold, J. S. and English, J. M. (2006). Mechanisms of drug inhibition of signalling molecules. *Nature*, 441(7092): 457-462.
- Sen, R. and Baltimore, D. (1986). Inducibility of kappa immunoglobulin enhancer-binding protein NF-kappa B by a post-translational mechanism. *Cell*, 47(6): 921-928.
- Seo, S., Tomita, Y. and Tori, K. (1981). Biosynthesis of oleanene- and ursine-type triterpenes from [4-<sup>13</sup>C] mevalonolactone and [1,2-<sup>13</sup>C<sub>2</sub>] acetate in tissue cultures of Isodon japonica Hara. *Journal of the American Chemical Society*, 101: 2075-2080.
- Sethi, G., Sung, B. and Aggarwal, B. B. (2008). Nuclear factor-κB activation: from bench to bedside. *Experimental Biology and Medicine*, 233(1): 21-31.
- Shah, U., Shah, R., Acharya, S. and Acharya, N. (2013). Novel anticancer agents from plant sources. *Chinese Journal of Natural Medicines*, 11(1): 16-23.
- Sharma, H. K., Chhangte, L. and Dolui, A. K. (2001). Traditional medicinal plants in Mizoram, India. *Fitoterapia*, 72(2): 146-161.
- Sherr, C. J. (1994). G1 phase progression: cycling on cue. *Cell*, 79(4): 551-555.
- Shi, X., Mao, Y., Saffiotti, U., Wang, L., Rojanasakul, Y., Leonard, S. S. and Vallyathan, V. (1995). Antioxidant activity of tetrandrine and its inhibition of quartz-induced lipid peroxidation. *Journal of Toxicology and Environmental Health*, 46(2): 233-248.
- Shim, H. Y., Park, J. H., Paik, H. D., Nah, S. Y., Kim, D. S. and Han, Y. S. (2007). Acacetin-induced apoptosis of human breast cancer MCF-7 cells involves caspase cascade, mitochondria-mediated death signaling and SAPK/JNK1/2-c-Jun activation. *Molecules and Cells*, 24(1): 95-104.
- Shishodia, S. and Aggarwal, B. B. (2004). Nuclear factor-κB: a friend or a foe in cancer? *Biochemical Pharmacology*, 68(6): 1071-1080.
- Shoeb, M. (2006). Anticancer agents from medicinal plants. *Bangladesh Journal of Pharmacology*, 1(2): 35-41.
- Shome, U., Khanna, R. K. and Sharma, H. P. (1979). Pharmacognostic studies on *Dillenia indica* Linn. I. Leaf. *Proceedings of Indian Academy of Science*, 88(1): 35-48.
- Shrivastava, A., Tiwari, M., Sinha, R. A., Kumar, A., Balapure, A. K., Bajpai, V. K., Sharma, R., Mitra, K., Tandon, A. and Godbole, M. M. (2006). Molecular iodine induces caspase-independent apoptosis in human breast carcinoma cells involving the mitochondria-mediated pathway. *Journal of Biological Chemistry*, 281(28): 19762-19771.

- Sies, H. (1991). Oxidative stress: from basic research to clinical application. *American Journal of Medicine*, 91(3): S31-S38.
- Simstein, R., Burow, M., Parker, A., Weldon, C. and Beckman, B. (2003). Apoptosis, chemoresistance, and breast cancer: insights from the MCF-7 cell model system. *Experimental Biology and Medicine*, 228(9): 995-1003.
- Singha, A. K., Bhattacharjee, B., Ghosh, R., De, U. C. and Maiti, D. (2013). Antibacterial, anti-alpha glucosidase and antioxidant properties of *Dillenia pentagyna* Roxb. *Asian Journal of Pharmaceutical and Clinical Research*, 6(4): 173-177.
- Singletary, S. E. and Connolly, J. L. (2006). Breast cancer staging: working with the sixth edition of the AJCC cancer staging manual. *CA: A Cancer Journal for Clinicians*, 56(1): 37-47.
- Skibola, C. F. and Smith, M. T. (2000). Potential health impacts of excessive flavonoid intake. *Free Radical Biology and Medicine*, 29(3-4): 375-383.
- Skinner, K. A. and Silverstein, M. J. (2001). The management of ductal carcinoma *in situ* of the breast. *Endocrine Related Cancer*, 8(1): 33-45.
- Solarity, E., Droin, N. and Sordet, O. (2002). Cell death pathways as targets for anticancer drugs. B. C. Baguley & D. J. Kerr (Eds.). *Anticancer drug development* (pp 55–76). San Diego: Academic Press.
- Solowey, E., Lichtenstein, M., Sallon, S., Paavilainen, H., Solowey, E. and Lorberboum-Galski, H. (2014). Evaluating medicinal plants for anticancer activity. *Scientific World Journal*, 2014: 1-12.
- Song, G., Ouyang, G. and Bao, S. (2005). The activation of Akt/PKB signaling pathway and cell survival. *Journal of Cellular and Molecular Medicine*, 9(1): 59-71.
- Sørlie, T., Tibshirani, R., Parker, J., Hastie, T., Marron, J. S., Nobel, A., Deng, S., Johnsen, H., Pesich, R., Geisler, S., Demeter, J., Perou, C. M., Lønning, P. E., Brown, P. O., Børresen-Dale, A.-L. and Botstein, D. (2003). Repeated observation of breast tumor subtypes in independent gene expression data sets. *Proceedings of the National Academy of Sciences*, 100(14): 8418-8423.
- Sosa, V., Moliné T., Somoza, R., Paciucci, R., Kondoh, H. and Leonart, M. E. (2013). Oxidative stress and cancer: an overview. *Ageing Research Reviews*, 12(1): 376-390.
- Soule, H. D., Maloney, T. M., Wolman, S. R., Peterson, W. D. J., Brenz, R., McGrath, C. M., Russo, J., Pauley, R. J., Jones, R. F. and Brooks, S. C. (1990). Isolation and characterization of a spontaneously immortalized human breast epithelial cell line, MCF-10. *Cancer Research*, 50(15): 6075-6086.

- Soule, H. D., Vazquez, J., Long, A., Albert, S. and Brennan, M. (1973). A human cell line from a pleural effusion derived from a breast carcinoma. *Journal of the National Cancer Institute*, 51(5): 1409-1416.
- Soung, Y. H., Lee, J. W., Nam, S. W., Lee, J. Y., Yoo, N. J. and Lee, S. H. (2006). Mutational analysis of *AKT1*, *AKT2* and *AKT3* genes in common human carcinomas. *Oncology*, 70(4): 285-289.
- Sousa, M. C., Varandas, R., Santos, R. C., Santos-Rosa, M., Alves, V. and Salvador, J. A. R. (2014). Antileishmanial activity of semisynthetic lupane triterpenoids betulin and betulinic acid derivatives: synergistic effects with miltefosine. *PLoS ONE*, 9(3): 1-12.
- Spitale, A., Mazzola, P., Soldini, D., Mazzucchelli, L. and Bordoni, A. (2009). Breast cancer classification according to immunohistochemical markers: clinicopathologic features and short-term survival analysis in a population-based study from the South of Switzerland. *Annals of Oncology*, 20(4): 628-635.
- Sriram, D., Yogeeswari, P., Thirumurugan, R. and Bal, T. R. (2005). Camptothecin and its analogues: a review on their chemotherapeutic potential. *Natural Product Research*, 19(4): 393-412.
- Srithi, K., Balslev, H., Wangpakapattanawong, P., Srisanga, P. and Trisonthi, C. (2009). Medicinal plant knowledge and its erosion among the Mien (Yao) in northern Thailand. *Journal of Ethnopharmacology*, 123(2): 335-342.
- Srivastava, B. K. and Pande, C. S. (1981). Chemical examination of the bark of *Dillenia indica*. *Acta Ciencia Indica*, 7: 170-174.
- Srivastava , S. D. (1981). Flavonoids from the stem of *Dillenia pentagyna*. *Phytochemistry*, 20: 2445.
- Srivastava, S. K. and Srivastava, S. D. (1984). A new diterpene, dipoloic acid from the stem of *Dillenia pentagyna*. *Current Science*, 53: 646-647.
- Staaf, J., Ringnér, M., Vallon-Christersson, J., Jönsson, G., Bendahl, P.-O., Holm, K., Arason, A., Gunnarsson, H., Hegardt, C., Agnarsson, B. A., Luts, L., Grabau, D., Fernö, M., Malmström, P.-O., Johannsson, O. T., Loman, N., Barkardottir, R. B. and Borg, Å. (2010). Identification of subtypes in human epidermal growth factor receptor 2-positive breast cancer reveals a gene signature prognostic of outcome. *Journal of Clinical Oncology*, 28(11): 1813-1820.
- Stählin, H. (1973). Activity of a new glycosidic lignan derivative (VP 16-213) related to podophyllotoxin in experimental tumors. *European Journal of Cancer*, 9(3): 215-221.
- Stevenson, D. E. and Hurst, R. D. (2007). Polyphenolic phytochemicals- just antioxidants or much more? *Cellular and Molecular Life Sciences*, 64(22): 2900-2916.

- Stewart, Z. A., Westfall, M. D. and Pietenpol, J. A. (2003). Cell-cycle dysregulation and anticancer therapy. *Trends in Pharmacological Sciences*, 24(3): 139-145.
- Stoka, V., Turk, B., Schendel, S. L., Kim, T.-H., Cirman, T., Snipas, S. J., Ellerby, L. M., Bredesen, D., Freeze, H., Abrahamson, M., Brömme, D., Krajewski, S., Reed, J. C., Yin, X.-M., Turk, V. and Salvesen, G. S. (2001). Lysosomal protease pathways to apoptosis: cleavage of Bid, not pro-caspases, is the most likely route. *Journal of Biological Chemistry*, 276(5): 3149-3157.
- Storz, P. (2005). Reactive oxygen species in tumor progression. *Frontiers in Bioscience*, 10: 1881-1896.
- Sucher, N. J. and Carles, M. C. (2008). Genome-based approaches to the authentication of medicinal plants. *Planta Medica*, 74(6): 603-623.
- Sudheerkumar, P., Shiras, A., Das, G., Jagtap, J. C., Prasad, V. and Shastry, P. (2008). Independent activation of Akt and NF-kappaB pathways and their role in resistance to TNF-alpha mediated cytotoxicity in gliomas. *Molecular Carcinogenesis*, 47(2): 126-136.
- Sun, Y., Huang, L., Mackenzie, G. G. and Rigas, B. (2011). Oxidative stress mediates through apoptosis the anticancer effect of phospho-nonsteroidal anti-inflammatory drugs: implications for the role of oxidative stress in the action of anticancer agents. *Journal of Pharmacology and Experimental Therapeutics*, 338(3): 775-783.
- Talib, W. H. and Mahasneh, A. M. (2010). Antiproliferative activity of plant extracts used against cancer in traditional medicine. *Scientia Pharmaceutica*, 78: 33-45.
- Tan, R. (2001). Simpoh Air *Mangrove and wetland wildlife at Sungei Buloh Nature Park*. [http://www.naturia.per.sg/buloh/plants/simpoh\\_air.html](http://www.naturia.per.sg/buloh/plants/simpoh_air.html) Retrieved 5th December, 2015
- Tanaka, T., Tanaka, T. and Tanaka, M. (2011). Potential cancer chemopreventive activity of protocatechuic acid. *Journal of Experimental and Clinical Medicine*, 3(1): 27-33.
- Tang, D., Wu, D., Hirao, A., Lahti, J. M., Liu, L., Mazza, B., Kidd, V. J., Mak, T. W. and Ingram, A. J. (2002). ERK activation mediates cell cycle arrest and apoptosis after DNA damage independently of p53. *Journal of Biological Chemistry*, 277(15): 12710-12717.
- Tanwani, A. K. and Majeed, M. (2009). Pattern of invasive ductal carcinoma of breast according to nottingham prognostic index. *Annals of Pakistan Institute of Medical Sciences*, 5(4): 251-254.
- Tao, Z., Jones, E., Goodisman, J. and Souid, A.-K. (2008). Quantitative measure of cytotoxicity of anticancer drugs and other agents. *Analytical Biochemistry*, 381(1): 43-52.

- Terrasse, V. (2013). *Latest world cancer statistics. Global cancer burden rises to 14.1 million new cases in 2012: marked increase in breast cancers must be addressed*. Geneva: International Agency for Research on Cancer, WHO.
- Tesso, H. (2005). *Isolation and structure elucidation of natural products from plants*. PhD Thesis, University of Hamburg, Germany
- Testa, J. R. and Bellacosa, A. (2001). AKT plays a central role in tumorigenesis. *Proceedings of the National Academy of Sciences*, 98(20): 10983-10985.
- Thompson, A. M. and Moulder-Thompson, S. L. (2012). Neoadjuvant treatment of breast cancer. *Annals of Oncology*, 23(10): 231-236.
- Thornton, T. M. and Rincon, M. (2009). Non-classical p38 MAP kinase functions: cell cycle checkpoints and survival. *International Journal of Biological Sciences*, 5(1): 44-52.
- Tiwari, K. P. and Srivastava, S. D. (1979). Pigments from the stem bark of *Dillenia indica*. *Planta Medica*, 35: 188-190.
- Tiwari, K. P., Srivastava, S. D. and Srivastava, S. K. (1980). Antibacterial triterpenoids from *Dillenia papuana* and their structure activity relationships. *Phytochemistry*, 19: 980-981.
- Toker, A. and Yoeli-Lerner, M. (2006). Akt signaling and cancer: surviving but not moving on. *Cancer Research*, 66(8): 3963-3966.
- Tournier, C., Hess, P., Yang, D. D., Xu, J., Turner, T. K., Nimnual, A., Bar-Sagi, D., Jones, S. N., Flavell, R. A. and Davis, R. J. (2000). Requirement of JNK for stress-induced activation of the cytochrome c-mediated death pathway. *Science*, 288(5467): 870-874.
- Tournier, C., Whitmarsh, A. J., Cavanagh, J., Barrett, T. and Davis, R. J. (1997). Mitogen-activated protein kinase kinase 7 is an activator of the c-Jun NH<sub>2</sub>-terminal kinase. *Proceedings of the National Academy of Sciences of the United States of America*, 94(14): 7337-7342.
- Tovey, F. I., Bardhan, K. D. and Hobsley, M. (2013). Dietary phospholipids and sterols protective against peptic ulceration. *Phytotherapy Research*, 27(9): 1265-1269.
- Tran, S. E., Holmstrom, T. H., Ahonen, M., Kahari, V. M. and Eriksson, J. E. (2001). MAPK/ERK overrides the apoptotic signaling from Fas, TNF, and TRAIL receptors. *Journal of Biological Chemistry*, 276(19): 16484-16490.
- Trudeau, M. E., Eisenhauer, E. A., Higgins, B. P., Letendre, F., Lofters, W. S., Norris, B. D., Vandenberg, T. A., Delorme, F. and Muldal, A. M. (1996). Docetaxel in patients with metastatic breast cancer: a Phase II study of the National Cancer Institute of Canada-Clinical Trials Group. *Journal of Clinical Oncology*, 14(2): 422-428.

- Tsai-Turton, M., Luong, B. T., Tan, Y. and Luderer, U. (2007). Cyclophosphamide-induced apoptosis in COV434 human granulosa cells involves oxidative stress and glutathione depletion. *Toxicological Sciences*, 98(1): 216-230.
- UICC (2007). *TNM history, evolution and milestones*. Geneva: International Union Against Cancer.
- Vachon, P. H., Harnois, C., Grenier, A., Dufour, G., Bouchard, V., Han, J., Landry, J., Beaulieu, J. F., Vezina, A., Dydensborg, A. B., Gauthier, R., Cote, A., Drolet, J. F. and Lareau, F. (2002). Differentiation state-selective roles of p38 isoforms in human intestinal epithelial cell anoikis. *Gastroenterology*, 123(6): 1980-1991.
- Valko, M., Rhodes, C. J., Moncol, J., Izakovic, M. and Mazur, M. (2006). Free radicals, metals and antioxidants in oxidative stress-induced cancer. *Chemico-Biological Interactions*, 160(1): 1-40.
- Van 't Veer, L. J., Dai, H., van de Vijver, M. J., He, Y. D., Hart, A. A. M., Mao, M., Peterse, H. L., van der Kooy, K., Marton, M. J., Witteveen, A. T., Schreiber, G. J., Kerkhoven, R. M., Roberts, C., Linsley, P. S., Bernards, R. and Friend, S. H. (2002). Gene expression profiling predicts clinical outcome of breast cancer. *Nature*, 415: 530-536.
- Van Acker, S. A. B. E., Van Den Berg, D. j., Tromp, M. N. J. L., Griffioen, D. H., Van Bennekom, W. P., Van Der Vijgh, W. J. F. and Bast, A. (1996). Structural aspects of antioxidant activity of flavonoids. *Free Radical Biology and Medicine*, 20(3): 331-342.
- Van Antwerp, D. J., Martin, S. J., Kafri, T., Green, D. R. and Verma, I. M. (1996). Suppression of TNF- $\alpha$ -induced apoptosis by NF- $\kappa$ B. *Science*, 274(5288): 787-789.
- Vanags, D. M., Pörn-Ares, M. I., Coppola, S., Burgess, D. H. and Orrenius, S. (1996). Protease involvement in fodrin cleavage and phosphatidylserine exposure in apoptosis. *Journal of Biological Chemistry*, 271(49): 31075-31085.
- Vergarajauregui, S., San Miguel, A. and Puertollano, R. (2006). Activation of p38 mitogen-activated protein kinase promotes epidermal growth factor receptor internalization. *Traffic*, 7(6): 686-698.
- Verma, S. and Singh, S. P. (2008). Current and future status of herbal medicines. *Veterinary World*, 1(11): 347-350.
- Vermeulen, K., Van Bockstaele, D. R. and Berneman, Z. N. (2003). The cell cycle: a review of regulation, deregulation and therapeutic targets in cancer. *Cell Proliferation*, 36(3): 131-149.
- Veronesi, U., Viale, G., Rotmensz, N. and Goldhirsch, A. (2005). Rethinking TNM: breast cancer TNM classification for treatment decision-making and research. *Breast*, 15(1): 3-8.

- Veskoukis, A. S., Tsatsakis, A. M. and Kouretas, D. (2012). Dietary oxidative stress and antioxidant defense with an emphasis on plant extract administration. *Cell Stress and Chaperones*, 17(1): 11-21.
- Villa, P. G., Henzel, W. J., Sensenbrenner, M., Henderson, C. E. and Pettmann, B. (1998). Calpain inhibitors, but not caspase inhibitors, prevent actin proteolysis and DNA fragmentation during apoptosis. *Journal of Cell Science*, 111(6): 713-722.
- Vousden, K. H. and Lu, X. (2002). Live or let die: the cell's response to p53. *Nature Reviews Cancer*, 2(8): 594-604.
- Wada, T. and Penninger, J. M. (2004). Mitogen-activated protein kinases in apoptosis regulation. *Oncogene*, 23(16): 2838-2849.
- Walker, A. R. P., Adam, F. I. and Walker, B. F. (2004). Breast cancer in black African women: a changing situation. *Journal of the Royal Society for the Promotion of Health*, 124(2): 81-85.
- Walker, D. H. and Maller, J. L. (1991). Role for cyclin A in the dependence of mitosis on completion of DNA replication. *Nature*, 354(6351): 314-317.
- Wallace-Brodeur, R. R. and Lowe, S. W. (1999). Clinical implications of p53 mutations. *Cellular and Molecular Life Sciences*, 55(1): 64-75.
- Walsh, C. M. (2014). Grand challenges in cell death and survival: apoptosis vs. necroptosis. *Frontiers in Cell and Developmental Biology*, 2: 1-3.
- Wang, K., Zhu, X., Zhang, K., Zhu, L. and Zhou, F. (2014). Investigation of gallic acid induced anticancer effect in human breast carcinoma MCF-7 cells. *Journal of Biochemical and Molecular Toxicology*, 28(9): 387-393.
- Wang, L., Liu, L., Shi, Y., Cao, H., Chaturvedi, R., Calcutt, M. W., Hu, T., Ren, X., Wilson, K. T., Polk, D. B. and Yan, F. (2012). Berberine induces caspase-independent cell death in colon tumor cells through activation of apoptosis-inducing factor. *PLoS ONE*, 7(5): 1-12.
- Wang, Q., Somwar, R., Bilan, P. J., Liu, Z., Jin, J., Woodgett, J. R. and Klip, A. (1999). Protein kinase B/Akt participates in GLUT4 translocation by insulin in L6 myoblasts. *Molecular and Cellular Biology*, 19(6): 4008-4018.
- Wang, S. and El-Deiry, W. (2005). *P53, cell cycle arrest and apoptosis*. P. Hainaut & K. Wiman (Eds.). Netherlands: Springer
- Wang, S., Konorev, E. A., Kotamraju, S., Joseph, J., Kalivendi, S. and Kalyanaraman, B. (2004). Doxorubicin induces apoptosis in normal and tumor cells via distinctly different mechanisms: intermediacy of H<sub>2</sub>O<sub>2</sub>- and p53-dependent pathways. *Journal of Biological Chemistry*, 279(24): 25535-25543.

- Wang, X., Martindale, J. L., Liu, Y. and Holbrook, N. J. (1998). The cellular response to oxidative stress: influences of mitogen-activated protein kinase signalling pathways on cell survival. *Biochemical Journal*, 333(2): 291-300.
- Wani, M. C., Taylor, H. L., Wall, M. E., Coggon, P. and McPhail, A. T. (1971). Plant antitumor agents. VI. Isolation and structure of taxol, a novel antileukemic and antitumor agent from *Taxus brevifolia*. *Journal of the American Chemical Society*, 93(9): 2325-2327.
- Wardman, P., Burkitt, M., Patel, K., Lawrence, A., Jones, C., Everett, S. and Vojnovic, B. (2002). Pitfalls in the use of common luminescent probes for oxidative and nitrosative stress. *Journal of Fluorescence*, 12(1): 65-68.
- Webber, C., Gospodarowicz, M., Sabin, L. H., Wittekind, C., Greene, F. L., Mason, M. D., Compton, C., Brierley, J. and Groome, P. A. (2014). Improving the TNM classification: findings from a 10-year continuous literature review. *International Journal of Cancer*, 135(2): 371-378.
- Weigelt, B., Mackay, A., A'Hern, R., Natrajan, R., Tan, D. S. P., Dowsett, M., Ashworth, A. and Reis-Filho, J. S. (2010). Breast cancer molecular profiling with single sample predictors: a retrospective analysis. *Lancet Oncology*, 11(4): 339-349.
- Wen, J., You, K.-R., Lee, S.-Y., Song, C.-H. and Kim, D.-G. (2002). Oxidative stress-mediated apoptosis: the anticancer effect of the sesquiterpene lactone parthenolide. *Journal of Biological Chemistry*, 277(41): 38954-38964.
- Whitaker-Worth, D. L., Carbone, V., Susser, W. S., Phelan, N. and Grant-Kels, J. M. (2000). Dermatologic diseases of the breast and nipple. *Journal of the American Academy of Dermatology*, 43(5): 733-754.
- WHO (2014, November 2014). Cancer. <http://www.who.int/mediacentre/factsheets/fs297/en/>. Retrieved 8th January, 2015
- Wiart, C., Mogana, S., Khalifah, S., Mahan, M., Ismail, S., Buckle, M., Narayana, A. K. and Sulaiman, M. (2004). Antimicrobial screening of plants used for traditional medicine in the state of Perak, Peninsular Malaysia. *Fitoterapia*, 75(1): 68-73.
- Wijeratne, S. S., Cuppett, S. L. and Schlegel, V. (2005). Hydrogen peroxide induced oxidative stress damage and antioxidant enzyme response in Caco-2 human colon cells. *Journal of Agricultural and Food Chemistry*, 53(22): 8768-8774.
- Williamson, G. (2003). The use of HPLC with coulometric array detection in the analysis of flavonoids in complex matrices *Methods in polyphenol analysis*. London: The Royal Society of Chemistry.
- Willmann, M., Wacheck, V., Buckley, J., Nagy, K., Thalhammer, J., Paschke, R., Triche, T., Jansen, B. and Selzer, E. (2009). Characterization of NVX-207, a

- novel betulinic acid-derived anti-cancer compound. *European Journal of Clinical Investigation*, 39(5): 384-394.
- Wolf, B. B., Schuler, M., Echeverri, F. and Green, D. R. (1999). Caspase-3 is the primary activator of apoptotic DNA fragmentation via DNA fragmentation factor-45/inhibitor of caspase-activated DNase inactivation. *Journal of Biological Chemistry*, 274(43): 30651-30656.
- Wood, D. E., Thomas, A., Devi, L. A., Berman, Y., Beavis, R. C., Reed, J. C. and Newcomb, E. W. (1998). Bax cleavage is mediated by calpain during drug-induced apoptosis. *Oncogene*, 17(9): 1069-1078.
- Wyllie, A. H. (1997). Apoptosis: an overview. *British Medical Bulletin*, 53(3): 451-465.
- Xiang, J., Chao, D. T. and Korsmeyer, S. J. (1996). BAX-induced cell death may not require interleukin 1 beta-converting enzyme-like proteases. *Proceedings of the National Academy of Sciences USA*, 93(25): 14559-14563.
- Xu, N., Lao, Y., Zhang, Y. and Gillespie, D. A. (2012). Akt: a double-edged sword in cell proliferation and genome stability. *Journal of Oncology*, 2012: 1-15.
- Yang, C., Pei, W., Zhao, J., Cheng, Y., Zheng, X. and Rong, J. (2014). Bornyl caffeate induces apoptosis in human breast cancer MCF-7 cells via the ROS- and JNK-mediated pathways. *Acta Pharmacologica Sinica*, 35: 113-123.
- Yang, D. D., Kuan, C. Y., Whitmarsh, A. J., Rincon, M., Zheng, T. S., Davis, R. J., Rakic, P. and Flavell, R. A. (1997). Absence of excitotoxicity-induced apoptosis in the hippocampus of mice lacking the JNK3 gene. *Nature*, 389(6653): 865-870.
- Yang, E. and Korsmeyer, S. J. (1996). Molecular thanatopsis: a discourse on the Bcl-2 Family and cell death. *Blood*, 2: 386-401.
- Yeshwante, S. B., Juvekar, A. R., Nagmoti, D. M., Wankhede, S. S., Shah, A. S., Pimprikar, R. B. and Saindane, D. S. (2009). Anti-inflammatory activity of methanolic extracts of *Dillenia indica* L. leaves. *Journal of Young Pharmacists*, 1(1): 63-66.
- Yu, H. G., Yu, L. L., Yang, Y., Luo, H. S., Yu, J. P., Meier, J. J., Schrader, H., Bastian, A., Schmidt, W. E. and Schmitz, F. (2003). Increased expression of RelA/nuclear factor-kappa B protein correlates with colorectal tumorigenesis. *Oncology*, 65(1): 37-45.
- Zarubin, T. and Han, J. (2005). Activation and signaling of the p38 MAP kinase pathway. *Cell Research*, 15: 11-18.
- Zeni, A. L., Zomkowski, A. D., Maraschin, M., Rodrigues, A. L. and Tasca, C. I. (2012). Ferulic acid exerts antidepressant-like effect in the tail suspension test in mice: evidence for the involvement of the serotonergic system. *European Journal of Pharmacology*, 679(1-3): 68-74.

- Zhang, G.-J., Kimijima, I., Onda, M., Kanno, M., Sato, H., Watanabe, T., Tsuchiya, A., Abe, R. and Takenoshita, S. (1999). Tamoxifen-induced apoptosis in breast cancer cells relates to down-regulation of bcl-2, but not bax and bcl-XL, without alteration of p53 protein levels. *Clinical Cancer Research*, 5(10): 2971-2977.
- Zhang, H.-M., Zhao, L., Li, H., Xu, H., Chen, W.-W. and Tao, L. (2014). Research progress on the anticarcinogenic actions and mechanisms of ellagic acid. *Cancer Biology and Medicine*, 11(2): 92-100.
- Zhang, H., Chen, S., Qin, F., Huang, X., Ren, P. and Gu, X. (2008). Simultaneous determination of 12 chemical constituents in the traditional Chinese Medicinal Prescription Xiao-Yao-San-Jia-Wei by HPLC coupled with photodiode array detection. *Journal of Pharmaceutical and Biomedical Analysis*, 48(5): 1462-1466.
- Zhang, W. and Liu, H. T. (2002). MAPK signal pathways in the regulation of cell proliferation in mammalian cells. *Cell Research*, 12(1): 9-18.
- Zhou, G.-B., Zhang, J., Wang, Z.-Y., Chen, S.-J. and Chen, Z. (2007). Treatment of acute promyelocytic leukaemia with all-trans retinoic acid and arsenic trioxide: a paradigm of synergistic molecular targeting therapy. *Philosophical Transactions of the Royal Society of London B: Biological Sciences*, 362(1482): 959-971.
- Zhu, Y., Mao, X. O., Sun, Y., Xia, Z. and Greenberg, D. A. (2002). p38 mitogen-activated protein kinase mediates hypoxic regulation of Mdm2 and p53 in neurons. *Journal of Biological Chemistry*, 277(25): 22909-22914