

UNIVERSITI PUTRA MALAYSIA

NUTRIENT COMPOSITION, ANTIOXIDANT ANDANTIPROLIFERATIVE PROPERTIES OF CLAUSENA EXCAVATA ANDMURRAYA KOENIGII

WAN NOR I'ZZAH BT. WAN MOHAMAD ZAIN

FPSK(M) 2005 29

3 SEP LUUU

NUTRIENT COMPOSITION, ANTIOXIDANT AND ANTIPROLIFERATIVE PROPERTIES OF CLAUSENA EXCAVATA AND MURRAYA KOENIGII

By

WAN NOR I'ZZAH BT. WAN MOHAMAD ZAIN

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia in Fulfilment of the Requirement for the Degree of Master of Science

November 2005



As I was writing this thesis, I realized that without my parents I would not be here today. It is with this thought that I dedicate this thesis to my parents,

Hj. Wan Mohamad Zain bin Wan Yaakob

Hjh. Nik Khairiah binti Nik Abdul Rahman

in remembrance of their loves, guidance and sacrifices.

May Allah bless both of you.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

NUTRIENT COMPOSITION, ANTIOXIDANT AND ANTIPROLIFERATIVE PROPERTIES OF CLAUSENA EXCAVATA AND MURRAYA KOENIGII

By

WAN NOR I'ZZAH BT. WAN MOHAMAD ZAIN

November 2005

Chairman	:	Associate Professor Asmah Rahmat, PhD
Faculty	:	Medical and Health Sciences

The proximate composition of Clausena excavata and Murraya koenigii leaves, together with the vitamin and mineral contents were investigated. Studies on the antioxidant and antiproliferative properties of the plant extracts and essential oils were also carried out. The proximate analysis showed that C. excavata leaves contained higher moisture, ash and crude fibre contents compared to M. koenigii. The contents of vitamins A, C and E in C. excavata were found to be 47.78 mg/100 g, 586.30 mg/100 g and 267.67 mg/100 g, while in *M. koenigii* the results were 1406.32 mg/100 g, 374.38 mg/100 g and 18.52 mg/100 g respectively. It seems that Murraya koenigii contained higher zinc (0.09 mg/100 g sample), copper (0.1 mg/100 g sample), sodium (0.4 mg/100 g sample) and potassium (0.91 mg/100 g sample) compared to Clausena excavata that showed 0.01 mg zinc and copper per 100 g sample, sodium (0.37 mg/100 g sample) and potassium (0.73 mg/100 g sample). Iron (0.32 mg/100 g sample), magnesium (0.96 mg/100 g sample) and calcium (5.46 mg/100 g sample) were found to be higher in *C. excavata* than



M. koenigii that possessed 0.14 mg iron per 100 g sample, 0.76 mg magnesium per 100 g sample and 5.28 mg calcium per 100 g sample.

The essential oils were obtained by hydrodistillation using fresh leaves and analysed using GC-MS spectrometry. The leaf oil of *C. excavata* was mainly made up of safrole (89.85%) while the leaf oil of *M. koenigii* was mainly made up of β -farnesene (42.85%). Other components that were present in appreciable amounts in *M. koenigii* oil were naphthalene (12.17%), α -caryophyllene (8.09%), caryophyllene (5.47%) and eudesmol (4.34%).

The methanol and water crude extracts together with the essential oils of *C. excavata* and *M. koenigii* leaves were investigated for their antioxidant capacities in two different assays, namely the β -carotene bleaching method and 1,1-diphenyl-2-picrylhydrazyl (DPPH) radical scavenging activity. The methanol extract of *M. koenigii* showed the most potent antioxidant activity in β -carotene bleaching assay, giving a percentage of 86.13 %, while *C. excavata* showed 76.60 % in the assay. On the other hand, *M. koenigii* methanol extract showed weak effect in scavenging DPPH radical with an EC₅₀ value of 2.14 mg/ml, compared to the methanol extract of *C. excavata* which exhibited 0.89 mg/ml. The water extract of *C. excavata* showed higher antioxidant activity in both β -carotene bleaching method (76.02 %) and DPPH radical scavenging method (EC₅₀ value = 2.53 mg/ml) as compared to *M. koenigii* water extract which possessed 62.26 % antioxidant activity in β - carotene bleaching method and 4.32 mg/ml EC₅₀ value in DPPH radical scavenging assay. Antioxidant activity of *M. koenigii* oil (91.01 %) was higher than *C. excavata* oil (66.3 %). Nevertheless, both of the essential oils did not present well in DPPH radical scavenging assay. The total phenolic content in the methanolic and water extracts of *C. excavata* and *M. koenigii* leaves, which were determined according to the Folin-Ciocalteu method, were expressed as gallic acid equivalents (GAEs); whereas the total phenolics in the methanolic extracts of *C. excavata* had the highest phenolic content (103.33 mg of GAEs/g of sample extract) while *M. koenigii* methanol extract showed 63.92 mg of GAEs/g of sample extract. The total phenolic content of *M. koenigii* water extract possessed 53.62 mg of GAEs/g of sample extract while *C. excavata* exhibited 53.46 mg of GAEs/g of sample extract respectively.

HepG2 (hepatic cancer), MCF-7 (hormone-dependent breast cancer), MDA-MB-231 (non-hormone-dependent breast cancer), HeLa (cervical cancer) and CAOV3 (ovarian cancer) cell cultures were used to determine the antiproliferative activities of *C. excavata* and *M. koenigii*. The growth of viable cells was evaluated by using Microculture-tetrazolium[•] (MTT) assay. Clausine-B, which was isolated from *C. excavata* was found to inhibit 50% of HeLa cancer cells' proliferation at 22.90 μ g/ml, followed by *M. koenigii* methanol extract (25.00 μ g/ml), *M. koenigii* essential oil (31.10 μ g/ml) and *C. excavata* methanol extract (34.51 μ g/ml). The *Clausena excavata* methanol

extract, water extract and essential oil were found to cause 50% cell death of MCF-7 cancer cell line at concentrations of 36.50, 95.00 and 59.00 µg/ml respectively. Meanwhile, clausine-B and essential oil from *M. koenigii* were found to cause 50% cell death at 52.90 and 46.01 μ g/ml respectively. For HepG-2 liver cancer cell line, the highest mean total IC₅₀ value could be seen in *M. koenigii* methanol extract which possessed 23.90 μ g/ml. It was followed by clausine-B which was found to cause 50% cell death at a concentration of 28.94 µg/ml. The essential oil from M. koenigii and C. excavata methanol extract exhibited 48.00 and 53.03 µg/ml. Clausine-B and M. koenigii methanol extract were observed to inhibit the proliferation of MDA-MB-231 cell line at the concentrations of 21.50 and 37.98 µg/ml respectively. Three samples were found to cause 50% cell death of CAOV3 which is the ovarian cancer cell line. The samples are clausine-B ($IC_{50} = 27.00$ μ g/ml), *M. koenigii* methanol extract (IC₅₀ = 27.90 μ g/ml) and *C. excavata* methanol extract (IC₅₀ = $79.00 \,\mu\text{g/ml}$).

The findings of this study showed that the methanol extracts especially *M*. *koenigii* methanol extract have the great potential in antioxidant and antiproliferative activities. Clausine-B, was found to be active against all the cancer cell lines tested.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

KOMPOSISI NUTRIEN, CIRI-CIRI ANTIOKSIDAN DAN ANTIPROLIFERATIF CLAUSENA ECAVATA DAN MURRAYA KOENIGII

Oleh

WAN NOR I'ZZAH BT. WAN MOHAMAD ZAIN

November 2005

Pengerusi :	Profesor Madya Asmah Rahmat, I	?hD
-------------	--------------------------------	-----

Fakulti : Perubatan dan Sains Kesihatan

Komposisi proksimat serta kandungan vitamin dan mineral bagi Clausena excavata dan Murraya koenigii dikaji. Kajian ke atas kandungan antioksidan dan antiproliferasi bagi ekstrak dan minyak pati bagi kedua-dua tumbuhan turut dilaksanakan. Analisis proksimat menunjukkan daun C. excavata mempunyai kandungan air, abu dan serat kasar yang lebih tinggi berbanding M. koenigii. Vitamin A, C dan E yang diperolehi bagi C. excavata adalah 47.78 mg/100 g, 586.30 mg/100 g dan 267.67 mg/100 g, manakala bagi M. koenigii keputusannya adalah 1406.32 mg/100 g, 374.38 mg/100 g dan 18.52 mg/100 g. Ia dapat dilihat bahawa Murraya koenigii mengandungi kandungan zink (0.09 mg/100 g sampel), kuprum (0.1 mg/100 g sampel), sodium (0.4 mg/100 g sampel) dan potasium (0.91 mg/100 g sampel) yang lebih tinggi berbanding dengan Clausena excavata yang menunjukkan 0.01 mg zink dan kuprum per 100 g sampel, sodium (0.37 mg/100 g sampel) dan potasium (0.73 mg/100 g sampel). Ferum (0.32 mg/100 g sampel), magnesium (0.96 mg/100 g sampel) dan kalsium (5.46 mg/100 g sampel)



didapati lebih tinggi di dalam *C. excavata* berbanding *M. koenigii* yang menunjukkan 0.14 mg ferum per 100 g sampel, 0.76 mg magnesium per 100 g sampel dan 5.28 mg kalsium per 100 g sampel.

Minyak pati diperolehi melalui teknik penyulingan hidro yang menggunakan daun segar dan dianalisis dengan menggunakan kaedah spektrometri GC-MS. Kandungan utama minyak pati daripada *C. excavata* adalah safrole (89.85%), manakala komponen utama bagi minyak pati daripada *M. koenigii* ialah β -farnesene (42.85%). Komponen-komponen lain yang diperolehi dalam amaun yang signifikan adalah naphthalene (12.17%), α -caryophyllene (8.09%), caryophyllene (5.47%) serta eudesmol (4.34%).

Kapasiti antioksidan bagi ekstrak kasar metanol dan air serta minyak pati daripada daun-daun *C. excavata* and *M. koenigii* dikaji melalui dua asai yang berbeza iaitu kaedah penurunan β-karotena dan aktiviti penghapusan radikal 1,1-difenil-2-pikrilhidrazil (DPPH). Ekstrak metanol *M. koenigii* menunjukkan aktiviti antioksidan yang paling berkesan di dalam asai penurunan β-karotena, dengan peratusan 86.13 %, manakala *C. excavata* menunjukkan 76.60 % di dalam asai tersebut. Namun begitu, ekstrak metanol *M. koenigii* adalah lemah di dalam penghapusan radikal DPPH dengan nilai EC₅₀ = 2.14 mg/ml, berbanding ekstrak metanol *C. excavata* yang menunjukkan 0.89 mg/ml. Ekstrak air *C. excavata* mempamerkan aktiviti antioksidan yang tinggi di dalam kedua-dua kaedah penurunan β-



karotena (76.02 %) serta penghapusan radikal DPPH (nilai $EC_{50} = 2.53$ mg/ml) berbanding ekstrak air M. koenigii. Namun, aktiviti antioksidan minyak pati M. koenigii (91.01 %) adalah lebih tinggi jika dibandingkan dengan minyak pati C. excavata (66.3 %). Namun demikian, kedua-dua minyak pati terbabit tidak menunjukkan aktiviti yang berkesan di dalam asai penghapusan radikal DPPH. Jumlah kandungan fenolik bagi ekstrak metanol dan air daun C. excavata dan M. koenigii ditentukan melalui kaedah Folin-Ciocalteu, serta ditunjukkan dalam gallic acid equivalent (GAE); yang mana jumlah fenolik di dalam ekstrak metanol bagi kedua-dua tumbuhan adalah lebih tinggi berbanding ekstrak air. Ekstrak metanol C. excavata mengandungi jumlah fenolik yang paling tinggi (103.33 mg GAE/g ekstrak sampel) manakala ekstrak metanol menunjukkan 63.92 mg of GAE/g ekstrak sampel. Jumlah kandungan fenolik bagi ekstrak air M. koenigii memperlihatkan 53.62 mg GAE/g ekstrak sampel manakala C. excavata menunjukkan 53.46 mg GAE/g sampel ekstrak sampel.

Kultur sel HepG2 (kanser hepar), MCF-7 (kanser payudara bergantung hormon), MDA-MB-231 (kanser payudara tidak bergantung hormon), HeLa (kanser pangkal rahim) dan CAOV3 (kanser ovari) telah digunakan untuk menentukan aktiviti antiproliferasi *C. excavata* dan *M. koenigii*. Pertumbuhan sel hidup ditentukan melalui asai Mikrokultur-tetrazolium (MTT). Clausine-B, yang diasingkan *C. excavata* telah didapati merencat 50% proliferasi sel kanser HeLa pada kepekatan 22.90 µg/ml, diikuti dengan ekstrak metanol *M. koenigii* (25.00 µg/ml), minyak pati *M. koenigii* (31.10 µg/ml) dan ekstrak metanol *C. excavata* (34.51 μ g/ml). Ekstrak metanol, ekstrak air dan minyak pati Clausena excavata, telah dilihat menyebabkan kematian 50% sel bagi titisan sel kanser MCF-7 pada kepekatan-kepekatan berikut; 36.50, 95.00 dan 59.00 µg/ml. Manakala clausine-B dan minyak pati M. koenigii didapati menyebabkan kematian 50% sel pada kepekatan 52.90 dan 46.01 μ g/ml . Bagi titisan sel kanser hepar HepG-2, jumlah purata nilai IC₅₀ dapat ditunjukkan di dalam ekstrak metanol *M. koenigii* yang mempamerkan 23.90 μ g/ml. Ini diikuti dengan clausine-B yang didapati menyebabkan kematian 50% sel pada kepekatan 28.94 µg/ml. Minyak pati *M. koenigii* dan ekstrak metanol *C. excavata* memperlihatkan 48.00 dan 53.03 μ g/ml. Clausine-B dan ekstrak metanol M. koenigii juga diperhatikan merencat proliferasi titisan sel kanser MDA-MB-231 pada kepekatan masing-masing 21.50 dan 37.98 µg/ml. Tiga sampel didapati menyebabkan kematian 50% sel CAOV3 yang mana merupakan titisan sel kanser ovari. Sampel-sampel terbabit adalah clausine-B (IC₅₀ = 27.00 μ g/ml), ekstrak metanol *M. koenigii* (IC₅₀ = 27.90 μ g/ml) dan ekstrak metanol *C. excavata* (IC₅₀ = $79.00 \,\mu\text{g/ml}$).

Hasil penemuan kajian ini mendapati ekstrak metanol terutamanya ekstrak metanol *M. koenigii* mempunyai potensi yang besar di dalam aktiviti antioxidan dan aktiviti antiproliferasi. Manakala clausine-B didapati aktif ke atas kesemua titisan sel kanser yang diuji.

ACKNOWLEDGEMENTS

Bismillahirrahmaanirrahiim.

In the name of Allah, The Most Gracious, The Most Merciful.

I thank many individuals, too numerous to mention, who have knowingly or unknowingly assisted me through my research.

I will always be grateful to Associate Professor Dr Asmah Rahmat for introducing me to this field and also for giving me personal attention, excellent advice, guidance, encouragement and support throughout my research. I also would like to acknowledge with sincere gratitude to Associate Professor Dr Fauziah Othman for her invaluable comments and suggestions. I would like to express my profound gratitude to Associate Prof. Dr Taufiq Yap Yun Hin for introducing me to this medicinal herbal plant, *Clausena excavata*, and also for his comments, advice and support. Thank you very much.

"Thank you very much for your guidance and for kindly providing invaluable professional assistance in reading, correcting, and improving specific portions of this thesis. They were priceless. I am indebted to all of you".

xi

Special thanks to K. Zi, Rafi, Aini, K. Idah and Sue for assisting me throughout my lab work at Microscopy and Microanalysis Unit. I am grateful to Dessy, Kapli, Hanim, Zet, Pah, Kak Normah and Kak Zana for assisting me in various ways in completing my lab work.

Appreciation also goes to Lim Gin Keat who had taught me in isolating essential oil. A simple thank is not sufficient to express the gratitude to what you have done in helping me. Not forgetting Kak Susi and Abang Herman. Thank you very much for your help, guidance and support.

I further wish to express my appreciation to my best colleagues Shikeen, Azah, Kak Abdah, Padly and Ze for having faith through our friendship and providing enthusiastic support. My lab partner and my best pal, Nida, we have come all this way and shared all the best and bad times of our long journey in completing our project and I am glad it has come to an end. Thank you for your patience and help, they were indescribable.

Finally, and most of all, I would like to express my heartiest acknowledge to my husband, Mohd Yasser bin Daud and all my beloved siblings, for their constant love, unfailing encouragement and sustaining interest in the progress of this thesis, and above all, patience.

Thank you.



3 SEP ZUUN

TABLE OF CONTENTS

Page

DEDICATION	ii
ABSTRACT	iii
ABSTRAK	vii
ACKNOWLEDGEMENTS	xi
APPROVAL	xiii
DECLARATION	xv
LIST OF TABLES	xix
LIST OF FIGURES	xx
LIST OF ABBREVIATIONS	xxiv

CHAPTER

1	INTRODUCTION	
	1.1.Background	1
	1.2.Objectives of study	9
2	LITERATURE REVIEW	
	2.1.Nutrient and cancer	10
	2.1.1. Carbohydrate	12
	2.1.2. Fibre	13
	2.1.3. Fats	13
	2.1.4. Protein	15
	2.1.5. Vitamin	15
	2.1.6. Minerals	19
	2.2. Antioxidant and cancer	21
	2.3.Cancer	26
	2.3.1. Human cancer cell lines	29
	2.3.2. Liver cancer	29
	2.3.3. Breast cancer	31
	2.3.4. Cervical cancer	33
	2.3.5. Ovarian cancer	35
	2.4.Essential oil	37
	2.5. Clausena excavata species	41
	2.5.1. Previous works on Clausena excavata	43
	2.5.2. Essential oil of C. excavata	49
	2.6. Murraya koenigii species	50
	2.6.1. Previous works on Murraya koenigii	51
	2.6.2. Essential oil of M. koenigii	54
	2.7. Leaf surface morphology	56



MATERIALS AND METHODS	
3.1. Plant materials	58
3.2. Nutrient composition	58
3.2.1. Chemicals	59
3.2.2. Proximate analysis	59
3.2.2.1. Moisture (air-oven method)	59
3.2.2.2. Ash	60
3.2.2.3. Crude fibre	60
3.2.2.4. Protein	61
3.2.2.5. Carbohydrate	63
3.2.2.6. Fat (ether extract)	64
3.2.3. Minerals determination	64
3.2.4. Determination of vitamins A, C and E	65
3.2.4.1. Analysis of vitamin A	65
. 3.2.4.2. Analysis of vitamin C	66
3.2.4.3. Analysis of vitamin E	66
3.2.4.4. HPLC analysis	67
3.3. Antioxidant activities	68
3.3.1. Chemicals	68
3.3.2. Extraction of samples	68
3.3.3. β -carotene bleaching assay	69
3.3.4. Free radical scavenging assay	70
3.3.5. Determination of total phenolic content	71
3.4. Isolation of essential oil	71
3.4.1. GC-MS analysis	75
3.4.2. Identification of the oil constituents	75
3.5. Antiproliferative assay	75
3.5.1. Cell cultures and chemicals	76
3.5.2. Extraction of samples	76
3.5.2.1. Hot water extraction	77
3.5.2.2. Organic extraction	77
3.5.3. Cells culturing	78
3.5.4. Subculturing cells	78
3.5.5. Cells bahaviour	79
3.5.6. MTT assav	79
3.5.6.1. Growth inhibitory effect	80
3.6. Leaf surface morphology	80
3.6.1. Chemicals	81
3.6.2. Scanning electron microscopy	81
3.7. Elemental analysis	82
3.8. Statistical analysis	83
RESULTS	
4.1. Nutrient composition	84
4.2. Isolation of essential oils	
4.2.1 Essential oil from <i>Clausena excavata</i> leaves	89

3

4



xvii

	4.2.2. Essential oil from <i>Murraya koenigii</i> leaves	92
	4.3. Antioxidant activities	07
	4.3.1. β -carotene bleaching assay	97
	4.3.2. Free radical scavenging assay	104
	4.3.3. Determination of total phenolic content	110
	4.4. Antiproliferative assay	
	4.4.1. MTT assay	112
	4.4.1.1. Growth inhibitory effect	116
	4.5. Leaf surface morphology	118
	4.6. Elemental analysis	128
5	DISCUSSION	
	5.1. Nutrient composition	130
	5.2. Isolation of essential oils	134
	5.3. Antioxidant activities	
	5.3.1. β -carotene bleaching assay	137
	5.3.2. Free radical scavenging assay	141
	5.3.3. Determination of total phenolic content	143
	5.4. Antiproliferative assay	
	5.4.1. MTT assay	146
	5.4.1.2. Growth inhibitory effect	150
	5.5. Leaf surface morphology	151
	5.6. Elemental analysis	152
6	CONCLUSIONS	154
BIBI	LIOGRAPHY	156
APF	PENDICES	179
BIO	DATA OF THE AUTHOR	198



xviii

Table		Page
3.1.	Conditions for HPLC separation of antioxidant vitamins.	67
4.1.	Nutrients composition of <i>Clausena excavata</i> and <i>Murraya koenigii</i> .	85
4.2.	Minerals determined in <i>C. excavata</i> and <i>M. koenigii</i> Leaves through Atomic Absorption Spectrophotometry (AAS).	88
4.3.	Composition of the essential oil of <i>Clausena excavata</i> leaves.	90
4.4.	Composition of the essential oil of <i>Murraya koenigii</i> leaves.	93
4.5.	Free radical scavenging activity (EC ₅₀) of <i>Clausena</i> <i>excavata</i> and <i>Murraya koenigii</i> methanol and water extract, essential oil and standards (BHT, ascorbic acid and α -tocopherol).	109
4.6.	Total phenolic content of methanol and water extracts from <i>Clausena excavata</i> and <i>Murraya koenigii</i> leaves.	111
4.7.	IC50 values of samples from <i>Clausena excavata</i> and <i>Murraya koenigii</i> .	114
4.8.	Elements detected in <i>C. excavata</i> and <i>M. koenigii</i> leaves through elemental analysis by using EDX attached to the VPSEM.	129



•

xix

LIST OF FIGURES

Figure		Page
2.1.	Clausena excavata species.	43
2.2.	Murraya koenigii species.	51
3.1.	Dean and Stark Apparatus.	73
3.2.	Flow chart of the isolation of essential oil.	74
3.3.	SEM JEOL 6400.	82
3.4.	LEO 1455 Variable Pressure-EDX.	83
4.1.	Safrole.	89
4.2.	Gas Chromatogram of the essential oil of <i>Clausena excavata</i> .	91
4.3.	β-farnesene.	92
4.4.	Gas Chromatogram of the essential oil of <i>Murraya koenigii.</i>	96
4.5.	Antioxidant activity of <i>C. excavata</i> and <i>M. koenigii</i> methanolic extracts compared with BHT (<i>BHT50</i>), α -tocopherol (<i>TOC50</i>) and ascorbic acid (<i>Ascorbic acid50</i>) at 50mg/l using β -carotene bleaching assay.	98
4.6.	Antioxidant activity of <i>C. excavata</i> and <i>M. koenigii</i> water extracts compared with BHT (<i>BHT50</i>), α -tocopherol (<i>TOC50</i>) and ascorbic acid (<i>Ascorbic acid50</i>) at 50 mg/l using β -carotene bleaching assay.	99
4.7.	Antioxidant activity of <i>C. excavata</i> and <i>M. koenigii</i> essential oil compared with BHT (<i>BHT50</i>), α -tocopherol (<i>TOC50</i>) and ascorbic acid (<i>Ascorbic acid50</i>) at 50 mg/l using β -carotene bleaching assay.	100



4.8.	Mean total antioxidant activity of <i>C. excavata</i> and <i>M. koenigii</i> methanol extracts, water extracts and essential oils and standards: BHT (<i>BHT50</i>), α -tocopherol (<i>TOC50</i>) and ascorbic acid (<i>Ascorbic acid50</i>). Superscripts of the same letter for each data were not significantly different at the level of p>0.05. Antioxidant activity was measured using β -carotene bleaching assay.	102
4.9.	Scavenging activity of <i>C. excavata</i> (<i>CE</i>) and <i>M. koenigii</i> (<i>MK</i>) methanolic extracts, BHT, α -tocopherol (<i>TOC</i>) and ascorbic acid on DPPH radical. Values are expressed as mean ± standard deviation of six replicate measurements.	105
4.10.	Scavenging activity of <i>C. excavata</i> (<i>CE</i>) and <i>M. koenigii</i> (<i>MK</i>) water extracts, BHT, α -tocopherol (<i>TOC</i>) and ascorbic acid on DPPH radical. Values are expressed as mean \pm standard deviation of six replicate measurements.	106
4.11.	Scavenging activity of <i>C. excavata</i> and <i>M. koenigii</i> essential oil, BHT, α -tocopherol (<i>TOC</i>) and ascorbic acid on DPPH radical. Values are expressed as mean \pm standard deviation of six replicate measurements.	107
4.12.	Phase contrast micrograph of antiproliferative activity of Clausine-B on HeLa cell line at 24, 48 and 72 hours incubation period ($IC_{50} = 22.90 \ \mu g/ml$). A. Untreated HeLa cell line (control) after 24 hours incubation. x4. B. Treated HeLa cell line after 24 hours incubation. x4. C. Untreated HeLa cell line (control) after 48 hours incubation. x4. D. Treated HeLa cell line after 48 hours incubation. x4. E. Untreated HeLa cell line (control) after 72 hours incubation. x4. F. Treated HeLa cell line after 72 hours incubation. x4.	117
4.13.	Scanning electron micrographs of the upper surfaces of fresh <i>C. excavata</i> leaves. A. (a) Thorny structure can be seen distributed in clusters on the upper surface, (b) New thorn originated from the surface. x25. B. Distance between one thorn to another thorn was equal forming a triangle. x30. C. Leaf cells surrounding thorny structures varied	120

in shape either square, angular or hexagonal, very closely packed and crowded. x100. D. (c) Thorns on the upper surface of *C. excavata* showing bulbous base, shaft, and swollen distal tip. x200.

- 4.14. Scanning electron micrographs showing numerous 121 stomatas on the lower surfaces of fresh C. excavata leaves. A. (a) Thorns seen on the lower surface. (b) Certain parts of the surfaces appeared in flowerlike shape. x100. B. Structure of the cells on the lower surface. x150. C. (c) Newly-developed thorn grew from the flower-like structure. x200. D. (d) The new thorn grew from this swollen base which originated from the flower-like structures. x500.
- 4.15. Scanning electron micrographs illustrate the upper 122 surfaces of processed C. excavata leaves. A. Thorns were seen distributed in cluster on the upper surface. x500. B. Irregular lines on some parts of the upper surface. x800. C. Parallel striations on the upper surface. x1,000. D. Hairy cells structure on the upper surface. x1,500.
- 4.16. Scanning electron micrographs of the lower 123 surfaces of processed C. excavata leaves. A. Curving thorn appeared at the middle of leaf lower surface; stomata could be seen closed to it. x500. B. Tapering thorn could be observed which was also located at the middle part of leaf lower surface. x550. C. Lower surface showing numerous scattering stomata. x600. D. Newly grown thorn could be found with opened stomata appeared adjacent to it. x900.
- 4.17. Scanning electron micrographs of the upper 124 surfaces of fresh M. koenigii leaves. A. Middle part of *M. koenigii* leaf. x500. B. Structure of the upper surface. x500. C. (a) Flower-like structure located within irregular lines on *M. koenigii* upper surface. x400. Close-up view of the flower-like structure which was located within the irregular lines. x_{3} , 000.
- 4.18. Scanning electron micrographs illustrates the lower 125 surfaces of fresh M. koenigii leaves. A. Middle part of M. koenigii lower surface. x100. B. Thorny



structure could be seen on the middle part. x500. C. Stomata located within irregular lines on rough surface of *M. koenigii*. x400. D. Close-up view of opened stomata which were located within the irregular lines. X2, 000.

4.19. Scanning electron micrographs illustrate the upper 126 surfaces of processed *M. koenigii* leaves.
A. (a) Flower-like structure could be found on the upper surface of *M. koenigii*. x300. B. Flower-like structure were seen on hairy surface. x500.
B. Upper surface showing irregular lines structure. x1, 000. Parallel striation could be seen on the upper surface. x1, 500.

4.20. Scanning electron micrographs showing the lower 127 surfaces of processed *M. koenigii* leaves. A. Lower surface of *M. koenigii*. x100. B. Stomata could be observed on rough surface. x300. C. Stomata located within irregular lines on rough surface of *M. koenigii* lower surface. x500.
D. Close-up view of opened stomata which were located within the irregular lines. x1, 500.





LIST OF ABBREVIATIONS

AAS	Atomic Absorption Spectrophotometry
AOAC	Association of Official Analytical Chemists
ATCC	American Type of Culture Collection
BHT	butylated hyroxytoluene
CO ₂	carbon dioxide
DPPH	α, α-diphenyl-β-picrylhydrazyl
ECCC	European Collection of Cell Culture
EDX	Energy Dispersive X-ray
GAE	Gallic Acid Equivalent
g	gram
GC-MS	Gas Chromatography-Mass Spectroscopy
HC1	hydrochloric acid
HPLC	High Performance Liquid Chromatography
μg	micro gram
μl	micro litre
μm	micro meter
1	litre
М	molar
ml	milli litre
mM	milli Molar
MeOH	methanol
min	minute
MTT	microculture tetrazolium
N	nitrogen
nm	nano meter
Na ₂ SO ₄	natrium sulfate
ppm	part per million
SEM	Scanning Electron Microscope
VPSEM	Variable Pressure Scanning Electron Microscope

