



UNIVERSITI PUTRA MALAYSIA

**PREVALENCE OF MYCOPLASMA GALLISEPTICUM IN DOMESTIC
CHICKENS AND FREE FLYING BIRDS AND MOLECULAR
CHARACTERISATION OF THE ISOLATES**

MAHADEVAN JAGANATHAN

FPV 2006 8



**PREVALENCE OF *MYCOPLASMA GALLISEPTICUM* IN DOMESTIC CHICKENS AND
FREE FLYING BIRDS AND MOLECULAR CHARACTERISATION OF THE ISOLATES**

By

MAHADEVAN JAGANATHAN

**Thesis submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of Master of Veterinary Science**

May 2006



**This project paper is especially dedicated to
my beloved parents, my wife and
my son SUDEESSVAN,
for their patience, support, encouragement and understanding
of my interest in Veterinary Medicine.**



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Veterinary Science

PREVALENCE OF *MYCOPLASMA GALLISEPTICUM* IN DOMESTIC CHICKENS AND FREE FLYING BIRDS AND MOLECULAR CHARACTERISATION OF THE ISOLATES

By

MAHADEVAN JAGANATHAN, DVM

May 2006

Chairman: Professor Aini Ideris, PhD

Faculty: Veterinary Medicine

Chronic respiratory disease (CRD) and complicated chronic respiratory disease (CCRD) are caused by *Mycoplasma gallisepticum* (MG). Infected birds show respiratory and reproductive problems which lead to severe production losses in poultry industry. *Mycoplasma gallisepticum* has been isolated in chickens and free flying birds (FFB) in several parts of the world. Therefore the current study was carried out to determine the prevalence of MG infection, also to molecularly characterize MG isolated from commercial chickens (broilers and layers), multiflock birds (indigenous chickens, ducks, turkeys and guinea fowls) and FFB in Selangor.

This study showed that using fresh yeast extract in preparation of mycoplasma agar and taking samples from choanal site of birds have made isolation of MG possible especially in layer chickens and indigenous chickens. Twenty-seven (27) MG isolates were isolated



from layer birds and indigenous chickens. The recovery rate by culture method was lower compared to the detection rate by DNA profiling using Polymerase chain reaction (PCR). Excessive usage of antibiotics in broiler farms may have contributed to the failure to isolate MG from broiler chickens by culturing method. The attempt to isolate MG from FFB was unsuccessful due to the small anatomical structure of choanal cleft and high contamination of oral cavity of FFB. The latter may have hindered the growth of MG artificially. Therefore, this study also proved that PCR is a better tool for epidemiological study compared to culturing method.

Broiler chickens and FFB at farms showed a high prevalence of MG infection based on serology and DNA detection but isolation could not be isolated. Crows from non-farm area or were not in close contact with infected birds or farms, did not show evidence of MG infection. This study also shows that only clinically ill infected birds, excreted and spread the organism to other flocks or species, as observed in the crows from the infected farm. However, sub-clinically infected birds as in indigenous chickens did not transmit the organism to other chickens or birds.

Characterisations of the MG isolates were conducted to investigate the presence of one or more types of MG strain in this study. Sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) and restriction endonuclease analysis (REA) were used as characterisation tools. All MG isolates from commercial chickens showed a unique band at 75 kilo-Dalton in SDS-PAGE, which was an important characteristic of MG F strain, which suggested that the isolates from layer farms could have derived from MG F strain, whereas the isolates from indigenous chickens were not similar to MG F strain. Different environmental exposures by MG strain may have caused alternation in genotype and/or phenotype, which might vary from one farm to another farm. Thus, these MG strains



might reveal different patterns in REA and SDS-PAGE. It is possible that the genotypic and phenotypic heterogeneity of MG demonstrated in the present study may have adversely influenced the outcome of the serum agglutination serology and may be important to consider optimizing antibody and organism detection systems. However, the unique characteristic of MG strain was reported to be stable when tested by SDS-PAGE. Whereas, based on REA, only one strain of MG will circulate among the flock of a farm.

This study therefore shows a high evidence of MG infection in commercial birds and FFB in farm. All MG isolates recovered from layer chickens were identical and possess a unique 75 kilo-Dalton protein band which was specific for MG F. However, the MG isolates obtained from the indigenous chickens were different from MG F strain .Their origin could not be determined from this study.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan ijazah Master Sains Veterinar

PREVALENS OLEH *MYCOPLASMA GALLISEPTICUM* PADA AYAM TERNAKAN DOMESTIK DAN BURUNG-BURUNG TERBANG BEBAS SERTA PENCIRIAN MOLEKUL BAGI ISOLATNYA

Oleh

MAHADEVAN JAGANATHAN, DVM

May 2006

Pengerusi: Professor Aini Ideris, PhD

Fakulti: Perubatan Veterinar

Penyakit pernafasan kronik (CRD) dan penyakit komplikasi pernafasan kronik (CCRD) adalah disebabkan oleh *Mycoplasma gallisepticum* (MG). Ayam yang berinfeksi menunjukkan tanda-tanda masalah pernafasan dan reproduktif, yang memberi kerugian yang besar pada industri ayam. *Mycoplasma gallisepticum* telah diasingkan daripada ayam dan burung-burung terbang bebas di beberapa tempat di dunia. Oleh itu, kajian ini adalah untuk menentukan prevalens, dan membuat pencirian molekul bagi MG yang diasingkan daripada ayam komersial (ayam pedaging dan penelur), ladang ternakan kumpulan poltri berbagai (ayam kampung, itik, ayam belanda dan ayam mutiara) dan burung-burung terbang bebas di Selangor.

Kajian ini menunjukkan bahawa penggunaan yis ekstrak yang disediakan sebaiksebelum dalam penyediaan agar mycoplasma and pengambilan sampel dari bahagian choanal



telah berjaya mengasingkan MG, terutama dari ayam penelur and ayam kampung. Dua puluh tujuh (27) isolat MG telah berjaya diperolehi daripada ayam penelur dan ayam kampung. Kadar pengasingan organisma ini melalui pengkulturan adalah lebih rendah berbanding dengan kadar pengesanan melalui analisis profil DNA dengan menggunakan *Polymerase chain reaction* (PCR). Penggunaan antibiotic yang tidak menentu pada ayam pedaging mungkin telah menghindari pertumbuhan MG melalui pengkulturan. Manakala, bagi FFB, bentuk anatomi choanal kecil and ruang mulut yang tercemar merupakan faktor-faktor yang menyebabkan kegagalan pengasingan MG secara pengkulturan. Oleh yang demikian, untuk kajian epidemiologi, PCR telah membuktikan sebagai kaedah saintifik yang lebih sesuai berbanding dengan pengkulturan.

Berdasarkan pada ujian serologi dan pengesanan DNA, ayam komersial dan burung-burung terbang bebas (FFB) yang berada di kawasan ladang menunjukkan kadar prevalens yang tinggi, tetapi MG tidak berjaya diasingkan. Burung gagak yang bukan berada di ladang ayam, ataupun tidak mempunyai kontak dekat dengan ayam berpenyakit di ladang-ladang, tidak menunjukkan tanda infeksi oleh MG. Kajian ini juga menunjukkan hanya ayam-ayam yang berpenyakit mengeluarkan dan menyebarkan penyakit ini kepada kumpulan dan spesis lain seperti yang dilihat pada gagak daripada ladang berinfeksi. Walaubagaimanapun, burung yang berinfeksi secara sub-klinikal seperti yang dilihat pada ayam kampung, tidak menyebarkan organisma ini kepada ayam atau burung-burung lain.

Pencirian terhadap isolat-isolat MG telah dijalankan untuk mengenalpasti kehadiran satu atau lebih jenis strain dalam kajian ini. Kaedah-kaedah yang digunakan untuk teknik pencirian adalah *sodium dodecyl sulphate- polyacrylamide gel electrophoresis*



(SDS-PAGE) dan *restriction endonuclease analysis* (REA), Semua isolat MG daripada ayam komersial menunjukkan gegelang unik pada 75 kilo-Dalton (kD) dalam ujian SDS-PAGE, yang merupakan satu ciri penting bagi strain MG F, menunjukkan bahawa kemungkinan besar MG yang diisolat daripada ayam penelur berasal daripada strain MG F, manakala isolat daripada ayam kampung tidak sama dengan strain MG F. Pendedahan persekitaran yang berlainan oleh strain MG boleh menyebabkan pertukaran dalam genotip dan/atau fenotip yang mungkin berlainan antara satu ladang dengan ladang yang lain. Oleh itu, strain MG ini berkemungkinan mendedahkan corak REA dan SDS-PAGE yang berbeza. Barangkali, kewujudan perbezaan dalam genotip dan fenotip MG dalam kajian ini secara tak langsung boleh mempengaruhi keputusan ujian serologi *serum agglutination test* dan adalah dicadangkan agar untuk mengoptimasikan antibodi dan sistem pengesanan orgnisma. Walaubagaimana pun, ciri strain MG yang unik dilaporkan stabil bila diuji dengan menggunakan SDS-PAGE. Manakala ,berdasarkan dari keputusan REA , hanya satu jenis strain MG akan berlegar di kelompok ayam dalam ladang tertentu.

Kajian ini telah menunjukkan prevalen infeksi MG yang tinggi pada ayam komersial dan FFB di ladang ayam. Semua isolat dari ayam penelur adalah sama dan memiliki satu gegelang di 75 kilo-Dalton yang merupakan ciri spesifik bagi MG F, manakala bagi ayam kampung ia adalah berbeza dari MG F. Asal isolat ini tidak dapat ditentukan dalam kajian ini.

ACKNOWLEDGEMENTS

This thesis was an ambitious project from the start and would have never been completed without the skills and talents of many people. Though only my name appears on the cover, much credit and my heartfelt thanks are owed to the following;

To GOD, who had made me possible in getting closer to good caring people who had made this challenge a VICTORY.

Especially to my dearest supervisor, Professor Dr. Aini Ideris, who has been there for me at all times where I was lost in deep pitch of darkness. Her motivation and concerns made me get up and move forward reaching to the finishing line. Thank YOU for being there for me.

I would also like to thank members of the supervisory committee, Professor Dato' Dr. Sheikh Omar Abdul Rahman, Associate Professor Dr. Abdul Rahim Mutalib and Dr. Nadzri Salim for their suggestions and guidance.

To my father, mother and my beloved wife, whom I owe their support throughout this long and demanding project.

Finally to all the staffs and academicians of the Faculty of Veterinary Medicine, Assoc. Prof. Dr. A. Rahman, Dr. Zunita, Dr. Ungku Chulan, Dr. Goh Yong Meng, Miss Yap May Lin and Encik Hajaraih, for their support during the course of the project.



To my dearest friends, especially Dr. Tan Chin Giap, Dr. Raguram, Dr. Saravanan, Dr. Gunalan, Mr. Lee Ee Liang, Mr.Beh Boon Cong and Mr.Beh Boon Kee, thank you for everything. Also to staff of Veterinary Research Institute, Ipoh, Perak, Madam Tan Lin Gee whom improvise my culturing and cloning techniques of MG. Last but not least, Dr. Jayashankar from Gym-tech Feed mill for being a very supportive and understanding manager.



I certify that an Examination Committee has met on 5th May 2006 to conduct the final examination of Mahadevan Jaganathan on his Master of Veterinary Science thesis entitled “Prevalence of *Mycoplasma gallisepticum* in Domestic Chickens and Free Flying Birds and Molecular Characterisation of the Isolates” in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The Committee recommends that the candidate be awarded the relevant degree. Members of the Examination Committee are as follows:

Mohd. Hair Bejo, PhD

Associate Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Chairman)

Zunita Zakaria, PhD

Lecturer
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Internal Examiner)

Jasni Sabri, PhD

Associate Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Internal Examiner)

Sharifah Syed Hassan, PhD

Veterinary Research Institute
(External Examiner)

HASANAH MOHD. GHAZALI, PhD

Professor/Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:



This thesis submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Veterinary Science. The members of the Supervisory Committee are as follow:

Aini Ideris, PhD

Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Chairman)

Sheikh Omar b. Abdul Rahman, PhD

Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Member)

Abdul Rahim Mutalib, PhD

Associate Professor
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Member)

Nadzri Salim

Lecturer
Faculty of Veterinary Medicine
Universiti Putra Malaysia
(Member)

AINI IDERIS, PhD

Professor/Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 10 th AUGUST 2006



DECLARATION

I hereby declare that the thesis is based on my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at UPM or other institutes.

MAHADEVAN JAGANATHAN

Date: 10 th AUGUST 2006



TABLE OF CONTENTS

		Page
DEDICATION		ii
ABSTRACT		iii
ABSTRAK		vi
ACKNOWLEDGEMENTS		ix
APPROVAL		xi
DECLARATION		xvii
LIST OF TABLES		xvii
LIST OF FIGURES		xix
LIST OF PLATES		xxi
LIST OF ABBREVIATIONS		xxiii
 CHAPTERS		
I	INTRODUCTION	1
II	LITERATURE REVIEW	7
	Chronic Respiratory Disease and Complicated Chronic Respiratory Disease	7
	Morphological and Bacteriological Characteristics of <i>Mycoplasma gallisepticum</i>	8
	Pathogenesis	10
	Immunity	11
	Epidemiology	12
	Clinical signs	15
	Commercial birds	15
	Broiler	15
	Layer	15
	Other Species of Birds	16
	Histopathology	17
	Laboratory Diagnosis	18
	Indirect Fluorescence Immunoassay (IFA)	18
	Molecular Techniques	19
	Polymerase Chain Reaction (PCR)	19
	Sodium Dodecyl Sulphate Polyacrylamide Gel Electrophoresis (SDS-PAGE)	20
	Restriction Endonuclease Analysis (REA)	21
	Serology Test	22
	Rapid Serum Agglutination Test (RSAT)	22
	Treatment	23
	Antibiotics	23
	Antimicrobial Amphipathic Peptide and Non-Antibiotic	24



	Psychotropic Drugs	
	Control	25
	Management	26
	Vaccination	26
III	PREVALENCE OF <i>MYCOPLASMA GALLISEPTICUM</i> IN DOMESTIC CHICKENS AND FREE FLYING BIRDS	
	Introduction	30
	Materials and Methodology	31
	Study Populations	31
	Categories of birds	32
	Commercial birds	32
	History of Layer farms	32
	Farm A	33
	Farm B	33
	Farm C	33
	History of Broiler farms	34
	History of Multi-flock Farm	34
	History of Free flying birds (FFB)	34
	Free flying birds in farms	34
	Free flying birds from out-side farms	35
	Analysis of Samples	35
	Rapid Serum Agglutination Test (RSAT)	35
	Conventional Method	36
	Culturing by <i>Mycoplasma media</i> (agar and broth)	36
	Confirmation by indirect immunofluorescence antibody (IFA) test	37
	Polymerase Chain Reaction (PCR)	38
	Preparation of DNA template	38
	Amplification of DNA	38
	Data Analysis	39
	Results	39
	Discussion	45
IV	MOLECULAR CHARACTERISATION OF <i>MYCOPLASMA GALLISEPTICUM</i> ISOLATES	
	Introduction	53
	Materials and Methodology	54
	<i>Mycoplasma gallisepticum</i> strain	54
	Cloning of <i>Mycoplasma gallisepticum</i> local and reference isolates	54
	SDS-PAGE for <i>Mycoplasma</i> protein	54
	Preparation of MG DNA for REA	55
	Digestion with restriction enzyme	56
	Electrophoresis	57
	Interpretation of Restriction Enzyme Analysis (REA)	57



	Results	58
	Discussion	85
V	DISCUSSION AND CONCLUSION	90
	BIBLIOGRAPHY	94
	APPENDICES	115
	BIODATA OF THE AUTHOR	120



LIST OF TABLES

Table		Page
3.0	Farm management system on various criteria based on observations	32
3.1	Score for antibody against MG using Rapid Serum Agglutination Test	42
3.2	Isolation of MG organism from domestic birds on mycoplasma media, percentage of MG antibody and percentage of MG DNA detection using PCR	43
3.3	Percentage of MG DNA detection using PCR and antibody using RSAT between layer and broiler farms	43
3.4	Isolation of MG organism from free flying birds on mycoplasma media, percentage of MG antibody and percentage of MG DNA detection using PCR	43
4.0	Criteria for interpreting pulsed field gel electrophoresis patterns	56
4.1	Restriction sites and number of cleavage bands produced by restriction endonuclease on MG DNA	58
5.0	F Values of the RE analyses among MG isolates from farm C digested with <i>Bam</i> HI	115
5.1	F Values of the RE analyses among MG isolates from farm C digested with <i>Eco</i> RI.	115
5.2	F Values of the RE analyses among MG isolates from farm C digested with <i>Hind</i> III	116
5.3	F Values of the RE analyses among MG isolates from farm B digested with <i>Bam</i> HI	116
5.4	F Values of the RE analyses among MG isolates from farm B digested with <i>Eco</i> RI	117
5.5	F Values of the RE analyses among MG isolates from farm B digested with <i>Hind</i> III	117
5.6	F Values of the RE analyses among MG isolates from indigenous birds digested with <i>Bam</i> HI	118
5.7	F Values of the RE analyses among MG isolates from indigenous chicken digested with <i>Eco</i> RI	118



5.8	F Values of the RE analyses among MG isolates from indigenous chicken digested with <i>Hind</i> III	118
5.9	F Values of the RE analyses among MG isolates of respective farm digested with <i>Eco</i> RI.	119
5.10	F Values of the RE analyses among MG isolates from respective farms digested with <i>Bam</i> HI	119
5.11	F Values of the RE analyses among MG isolates from respective farms digested with <i>Hind</i> III	119



LISTS OF FIGURES

Figure		Page
1	A schematic representative of Plate 6 of Restriction endonuclease patterns obtained with Farm C isolates with <i>Bam</i> HI	67
2	A schematic representative of Plate 7 of Restriction endonuclease patterns obtained with Farm C isolates with <i>Eco</i> RI	68
3	A schematic representative of Plate 8 of Restriction endonuclease patterns obtained with Farm C isolates with <i>Hind</i> III	69
4	A schematic representative of Plate 9 of Restriction endonuclease patterns obtained with Farm B isolates with <i>Bam</i> HI	71
5	A schematic representative of Plate 10 Restriction endonuclease patterns obtained with Farm B isolates with <i>Eco</i> RI	73
6	A schematic representative of Plate 11 of Restriction endonuclease patterns obtained with Farm B with <i>Hind</i> III	74
7	A schematic representative of Plate 12 of Restriction endonuclease patterns obtained with isolates obtained from indigenous chickens with <i>Bam</i> HI	76
8	A schematic representative of Plate 13 of Restriction endonuclease patterns obtained with isolates obtained from indigenous chickens with <i>Eco</i> RI	77
9	A schematic representative of Plate 14 of Restriction endonuclease patterns obtained with isolates obtained from indigenous chicken with <i>Hind</i> III	79
10	A schematic representative of Plate 15 Restriction endonuclease patterns obtained with <i>Bam</i> HI. M- Marker. R-Reference, A-representative isolate of Farm A, B-representative isolate of Farm B, C-representative isolate of Farm C, D-representative isolate of indigenous chicken, E- isolate from eye of diseased chicken from Farm B	80
11	A schematic representative of Plate 16. Restriction endonuclease patterns obtained with <i>Eco</i> RI. M- Marker. R-Reference, A-representative isolate of Farm A, B-representative isolate of Farm C, C-representative isolate of Farm B, D-representative isolate of indigenous chicken, E- isolate from eye of diseased chicken from Farm B	83



- 12 A schematic representative of Plate 16 of Restriction endonuclease patterns obtained with *Hind* III. M- Marker. R-Reference, A-representative isolate of Farm A, B-representative isolate of Farm C, C-representative isolate of Farm B, D-representative isolate of indigenous chicken, E- isolate from eye of diseased chicken from Farm B

84



LISTS OF PLATES

Plate		Page
1	Agarose gel electrophoresis of PCR products of samples using MG species specific primers	44
2	SDS-PAGE of proteins from MG isolates from Farm C in 10 % gel: Lane M= molecular mass standard (kD=kilo-Dalton); 1=MG F, 1-8=isolated from Farm C. Arrows indicating band pattern at 75 k D	63
3	SDS-PAGE of proteins from MG isolates from Farm B in 10 % gel: Lane M=molecular mass standard (K=kilo Dalton); No:1-9=MG isolates from Farm B. Arrows indicating band pattern at 75 k D	64
4	SDS-PAGE of proteins from MG isolates from indigenous chicken in 10 % gel: Lane. No.1-5=isolate from indigenous chicken	65
5	SDS-PAGE of proteins from MG isolates from respective farms in 10 % gel: Lane M= molecular mass standard (K=kilo Dalton); R-representative isolate of MG F strain (reference), A- representative isolate of Farm A, B-representative isolate of Farm B, C - representative isolate of Farm C, D-representative isolate of indigenous chicken, E-isolate of eye of a diseased chicken from Farm B	66
6	Restriction endonuclease patterns obtained with MG isolates digested with <i>Bam</i> HI. M-Marker, No.1-11-isolates from Farm C	67
7	Restriction endonuclease patterns obtained with MG isolates digested with <i>Eco</i> RI. M-Marker, No.1-11- isolates from Farm C	68
8	Restriction endonuclease patterns obtained with MG isolates digested with <i>Hind</i> III M-Marker, 1-isolate from infected eye of chicken from Farm B. No. 2-12 -isolates from Farm C	69
9	Restriction endonuclease patterns obtained with MG isolates digested with <i>Bam</i> HI. M-Marker, No. 1-10-isolates from farm B	70
10	Restriction endonuclease patterns obtained with MG isolates with <i>Eco</i> RI. M- Marker, No.1- isolate from farm A, No. 2-11 -isolates from farm B	72
11	Restriction endonuclease patterns obtained with MG isolates with <i>Hind</i> III M-Marker, R-Reference strain MG F, No.1-10-isolates from farm B	74
12	Restriction endonuclease patterns obtained with MG isolates from indigenous chicken <i>Bam</i> HI. M-Marker, R-MGF reference strain. No. 1-5-isolates from indigenous chicken	75
13	Restriction endonuclease patterns obtained with MG isolates from	



	indigenous chicken <i>Eco</i> RI M-Marker, R-MG F reference strain, No.1-5-isolates from indigenous chicken	77
14	Restriction endonuclease patterns obtained with MG isolates from indigenous chicken <i>Eco</i> RI. M-Marker, R- MG F strain, No.1-5-isolates from indigenous chicken	78
15	Restriction endonuclease patterns obtained with <i>Bam</i> HI. M- Marker, R-representative isolate of MG F strain (reference), A- representative isolate of Farm A, B-representative isolate of Farm B, C - representative isolate of Farm C, D-representative isolate of indigenous chicken, E-isolate of eye of a diseased chicken from Farm B	80
16	Restriction endonuclease patterns obtained with MG digested with <i>Eco</i> RI (A-E) and <i>Hind</i> III (A-E). M- Marker. R-representative isolate of MG F strain (reference), A- representative isolate of Farm A, B-representative isolate of Farm B, C - representative isolate of Farm C, D-representative isolate of indigenous chicken, E-isolate of eye of a diseased chicken from Farm B	82



LIST OF ABBREVIATIONS

B cell	Bursal cell
BRL	Bethesda Research Laboratories
CFU	Colony forming units
CRD	Chronic respiratory disease
CCRD	Complicated chronic respiratory disease
CPZ	Chlorpromazine
DNA	Deoxyribonucleic acid
EDTA	Ethylenediaminetetra acetic acid
ELISA	Enzyme-link immunosorbent assay
FLHS	Fatty liver hemorrhagic syndrome
HA	Hemagglutination
HI	Hemagglutination inhibition
IFA	Indirect Fluorescence Antibody
Ig A	Immunoglobulin A
IBD	Infectious bursal disease
Ig M	Immunoglobulin M
Ig G	Immunoglobulin G
IM	Intra-muscularly
IT	Intra-tracheally
kD	kilo Daltons
M	Molar
MG	<i>Mycoplasma gallisepticum</i>
MS	<i>Mycoplasma synoviae</i>
mm	millimeter



ml	milliliter
nm	nanometer
PCR	Polymerase chain reaction
pH	Logarithm 10 {H}
RAPD	Random amplified polymerase DNA
REA	Restriction Endonuclease Analysis
SDS-PAGE	Sodium dodecyl sulphate- polyacrylamide gel electrophoresis
SPA	Serum plate agglutination
T cell	Thymus cell
TBE	Tris-Borate
USA	United States of America
s	Seconds
°C	Degree Celsius
%	Percentage
μm	micrometer

