PATHOGENESIS OF HAEMORRHAGIC SEPTICAEMIA IN ORGAN CULTURE, MICE AND CALVES

AMNA ELAMIN MOHAMED

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PATHOGENESIS OF HAEMORRHAGIC SEPTICAEMIA IN ORGAN CULTURE, MICE AND CALVES

By

AMNA ELAMIN MOHAMED

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of Philosophy

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DEDICATION

ALLAH you are my supreme love

ALHAMDULILLAH ….thank you for everything

To the memory of my late father

My beloved mother, sisters and brothers

Thanks for your doa’, love and care
Experimental intraperitoneal infection of mice with Pasteurella multocida serotype B:2 at the dose of $10^7$ cells caused death in all infected mice between 18 to 24 hours. Microscopy showed suppurative tracheitis. The alveoli lumina were infiltrated with numerous inflammatory cells mainly neutrophils. Pink-stained fibrin and red blood cells were also present in the alveoli lumina. Blue shade of bacterial colonies were observed in the alveolar space and pulmonary vessels. In the aorta, there was degeneration of the endothelium, disruption of the intima and focal myolytic changes in the smooth muscle.

In infected mice, scanning electron microscopy (SEM) showed cilial damage and deciliation of tracheal epithelial cells. In the lung extravasation of red blood cells in the alveolar lumina was observed. Rod-shaped bacteria were seen attached to the alveolar wall causing depression on the surfaces of the cells. Changes in the aortas were characterized by rounded endothelial cells. Some rounded endothelial cells were seen about to detach and expelled into the blood vessel lumen. Bacteria were
most evident at the intercellular junction of the endothelial cells that were beginning to slough.

In infected mice, transmission electron microscopy (TEM) of the trachea showed bacterial cells attached to deformed cilia. Necrotic and apoptotic endothelial cells were seen protruded into the lumen of the affected blood vessel. Several bacteria were attached to the plasma membrane of the endothelial cells and in the blood vessel lumen. Endothelial cells showed electron dense cytoplasmic extensions around the bacteria and as attaching and effacing lesions which were interpreted as the first steps in the phagocytosis process. In the lung, there was thickening of the alveolar septa. Bacteria were seen dividing inside the neutrophils and in the lumen of the blood vessels. Some bacteria were also seen in vacuolated macrophages. Endothelial cells showed apoptosis. Fibrin was present in the alveolar lumina. In the aorta, endothelial cells were fragmented and detached from the basement membranes. Bacteria were attached to the fragmented endothelium.

Calves inoculated intranasally with $3.0 \times 10^{10} \ P. \ multocida$ serotype B:2 following stressed with dexamethasone developed septicemia and fibrinous pleuropneumonia. All infected calves died between 24 to 48 hours. Thrombi were observed in the nasal cavity and lung. It is therefore believed that the $P. \ multocida$ had gained entry into the blood vessel of the nasal cavity and/or lung to various organs causing septicemia. This is validated by the isolation of pure growth of $P. \ multocida$ from various congested organs following intranasal infection.
The microscopic changes in infected calves were suppurative rhinitis, suppurative tracheitis, fibrinoid necrosis in the blood vessel walls, thrombosis, necrotic vasculitis, alveolar congestion and alveolitis with edema and fibrin deposition in the alveolar lumen. Scanning electron microscopy (SEM) showed severe loss of cilia and the presence of free red blood cells in the nasal mucosa and trachea. Bacteria were attached to the alveolar wall causing depression on the cell surface. In the aorta of infected calves, bacteria were most evident near the base of extruding cells.

Transmission electron microscopy (TEM) of infected calves showed spherical blebbing with fine granules on the tip of destructed cilia (bullae). Bacteria were closely adhered to the endothelial surface and some were connected by tiny surface strands to the plasma membranes. Some of the bacteria were engulfed by the endothelial cells. There was apoptotic and necrotic morphology of dying endothelial cells after contact with *P. multocida*. Some erythrocytes have passed through the vascular wall. On the basis of these findings, it is considered that endothelial necrosis and apoptosis may be related to the disruption of endothelial barrier which then lead to vascular leakage. This may therefore contribute to the hemorrhagic complication of hemorrhagic septicemia.

*In vitro* experiments conducted to study adhesion and colonization of *P. multocida* serotype B:2 to tracheal mucosa, lung and aorta explants of calf showed that *P. multocida* B:2 can adhere to the lung, trachea and aorta. The adherence of bacteria at different time (1, 2, 3, 4, and 6 hours), is significantly different, (P<0.0001). For the lung and aorta, the attachment of the bacteria in the first and second hour post incubation was high, while that of 3 hours was moderate. There was no colonization
at 4 and 6 hours post infection. Attachment of the bacterium to the trachea also showed significant difference (P<0.0089) where the attachment of the bacteria in the first hour post infection was high, compared to 2, 3, 4 and 6 hours post infection which was moderate to no bacteria. The results of this study revealed that *P. multocida* B:2 was more adhesive to the lung tissues which showed severe colonization at 1 and 2 hours after incubation while aorta showed mild *P. multocida* B:2 colonization at 1 and 2 hours after incubation and trachea showed less *P. multocida* B:2 colonization. This result showed that the *in vitro* colonization decreased with time. Transmission electron microscopy showed severe damage to the epithelium. This study showed that after 1 hour of incubation, *P. multocida* was lost from the surface of the cell. Therefore, transiet time between 10 to 30 minutes is needed to detect adhered bacteria to tracheal epithelial cells and aorta endothelial cells. These studies demonstrated that *P. multocida* type B:2 was able to colonize and invade the lung, trachea and aorta explants.

Transmission electron microscopy showed degenerative features of the lung *in vivo* and *in vitro*. Bacteria were attached to the surface of the endothelial and epithelial cells. There was evidence of invasion and the bacteria appear to be concentrated at certain locations along the cellular membrane of lung which induced dramatic morphological changes on the surface of the attached cells. Some endothelial and epithelial cells form blebbing on the site of the attachment. Some internalized bacteria appeared in membranous vacuoles.

*Pasteurella multocida* was able to attach to and invade epithelial and endothelial cells *in vivo* and *in vitro*. It is therefore believed that the *P. multocida* had gained
entry into the blood vessel of the nasal cavity, trachea and lung to various organs causing septicaemia. Thus the growth of bacteria to an overwhelming number and then colonization of different organs resulted in the development of lesions in various organs and death due to vital organ failure.
PATOGENESIS SEPTISEMIA BERDARAH DALAM KULTUR ORGAN, TIKUS DAN ANAK LEMBU

Oleh

AMNA ELAMIN MOHAMED

December 2007

Pengerusi: Profesor Madya. Jasni Sabri, PhD

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Pemeriksaan trakea tikus dengan mikroskop elektron transmisi menunjukkan bakteria terlekat pada silia yang cacat bentuk. Kromatin nukleus sel endotelium


Pemeriksaan dengan mikroskop cahaya menunjukkan rhinitis dan trakeatitis ber supuratif, nekrosis fibrinoid pada dinding salur darah, trombosis, nekrosis vaskulitis, kongesi, alveolitis serta edema dan deposisi fibrin. Mikroskopii elektron imbas anak lembu terjangkit menunjukkan kehilangan silia yang teruk dan kehadiran sel darah merah pada mukosa nasum dan trakea. Bakteria didapat terlekat pada dinding

Kajian *in vitro* yang dijalankan adalah untuk mengkaji perlekatan dan pengkolonian *P. multocida* serotip B:2 kepada eksplan segar permukaan mukosa paru-paru dan aorta. Perlekatan bakteria pada selang masa yang berlainan (1,2,3,4 dan 6 jam) menunjukkan perbedaan signifikan (P<0.0001) pada paru-paru dan aorta. Perlekatan bakteria dalam tempoh 1 dan 2 jam yang pertama pasca inokulasi adalah tinggi berbanding dengan 3 jam pasca inokulasi dengan perlekatan yang sederhana. Tidak terdapat pengkolonian pada 4 dan 6 jam pasca inokulasi. Perlekatan bakteria kepada mukosa trakea juga menunjukkan perbedaan yang signifikan (P<0.0089) pada selang masa berlainan dimana dalam tempoh 1 jam pertama pasca inokulasi adalah tinggi berbanding dengan 2,3,4 dan 6 jam pasca inokulasi yang sederhana atau ketiadaan bakteria. Meningkatkan tempoh inkubasi menurunkan insiden perlekatan bakteria. Cerapan ini menunjukkan kecenderungan *P. multocida* untuk melekat kepada tisu
paru-paru adalah lebih tinggi seperti yang ditunjukkan oleh pengkolonian bakteria yang tinggi dalam 1 dan 2 jam pasca inkubasi sementara pengkolonian oleh *P. multocida* B:2 pada aorta dalam tempoh yang sama adalah sederhana. Insiden pengkolonian *P. multocida* B:2 ke atas eksplan trakea adalah kurang berbanding dengan aorta. Kajian ini menunjukkan pengkolonian *in vitro* menurun dengan penurunan masa untuk menjadi sederhana dalam tempoh 3 jam pasca inkubasi dan selepas 4 dan 6 jam tidak terdapat sebarang bakteria pada tisu tersebut. Cerapan di atas memberi gambaran *P. multocida* lebih mudah menyerang endotelium trakea dan aorta berbanding dengan paru-paru.


Mikroskopi elektron transmisi menunjukkan perubahan paru-paru *in vivo* dan *in vitro*. Bakteria kelihatan terlekat pada permukaan sel epitelium dan sel endotelium. Kelihatan terdapat bukti serangan oleh bakteria yang tertumpu pada lokasi tertentu di sepanjang membran sel paru-paru yang menyebabkan perubahan morfologi yang teruk di permukaan sel tersebut. Beberapa sel endotelium dan epitelium membentuk
bleb pada tapakan perlekan. Bakteria juga kelihatan di dalam vakuol bermembran sel-sel tersebut.

*P. multocida* berupaya menyerang sel epitelium dan endotelium dalam kajian *in vivo* dan *in vitro* tersebut di atas. Dengan ini adalah dipercayai *P. multocida* berjaya memasuki ke dalam salur darah rongga nasum, trakea, paru-paru dan pelbagai organ lain yang menyebabkan septisemia. Pembiakan bakteria kepada bilangan yang amat besar dan pengkoloniannya pada organ-organ tersebut berakhir dengan pembentukan lesi di beberapa organ yang berakhir dengan kematian haiwan disebabkan oleh kegagalan organ penting.
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All perfect praise is due to ALLAH; may ALLAH exalt the mention of Prophet Muhammad, his companions and those who follow in his footsteps.

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I certify that an Examination Committee has met on 14 December 2007 to conduct the final examination of Amna Elamin Mohamed on her Doctor of Philosophy thesis entitled “Experimental Pathogenesis of Haemorrhagic Septicemia in Organ Culture, Mice and Calves” in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The Committee recommends that the candidate be awarded the relevant degree. Members of the Examination Committee are as follows:

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Date:
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Date: 3 March 2008
DECLARATION

I declare that the thesis is my original work except for quotation and citation which have been duly acknowledged. I also declare that it has not been previously and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institution.

___________________
AMNA ELAMIN

Date: 20 February 2008
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3.13 Electron micrograph of the lung of a mouse from group 4 that died 20 hours p.i.. Note presence of bacteria entering into endothelial cells (arrows). The site where the bacteria attaches are depressed or invaginated to accommodate the size and shape of the bacteria. Inset show the magnification of the attaching bacteria. SEM. Bar1µm.

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3.17 Electron micrograph of the aorta of a mouse from group 4 that died 20 hours p.i.. Note a few bacteria on the eroded area with complete loss of endothelial cells (arrows). Infiltration of polymorphonuclear cells (PMN) (arrow heads), free red blood cells (black arrow) and massive deposition of fibrin (double arrow) is also observed. SEM. Bar 1 µm.

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3.19 Electron micrograph of the trachea of a mouse from group 3 that died 18 hours p.i.. Note bacterial cells are associated with broken cilia. Bacterial cell is undergoing division (arrow). Lead citrate & uranyl acetate. TEM X 8000

3.20 Electron micrograph of the trachea of a mouse from group 3 that died 18 hours p.i.. Note the endothelial cells are swollen with condense cytoplasm (big arrow). Erythrocytes filled the lumen and some are adhered to the endothelial cells. Many bacteria are seen in close association with the plasma membranes of the endothelial cells (small arrow). Invagination with finger like cytoplasmic extensions (double arrow) appear to engulf the attaching bacteria. Inset shows enlargement of attaching bacteria. Large halos are also seen around the bacteria. The tissues around the capillary contained edema fluid. Lead citrate & uranyl acetate. TEM X 1537

3.21 Electron micrograph of the trachea of a mouse from group 3 that died 18 hours p.i.. Note the degenerate endothelial cell with apoptotic nucleus. Bacteria (big arrows) are seen attached to the degenerate endothelial cells. Bacteria undergoing division (small arrow) is also seen in the lumen of the blood vessel and among erythrocytes. Lead citrate & uranyl acetate. TEM X 1356.
3.22 died 20 hours p.i.. Note the endothelial cells are severely swollen and protruding into the lumen (arrows). Bacteria cells were seen closely attached to the protruded endothelial cells. Lead citrate & uranyl acetate. TEM X 1356.

3.23 High magnification of the previous section (Figure 4.27). Note the bacteria in close attachment (black arrow) with the disrupted plasma membrane of swollen endothelial cells. The endothelial cell membranes showed electron dense at the site of the attachment (white arrow). Cytoplasmic extensions around the bacteria are observed (arrow head). Lead citrate & uranyl acetate. TEM X 2140.

3.24 Electron micrograph of the trachea of a mouse from group 5 that died 22 hours p.i.. Note many bacteria are closely attached to necrotic endothelial cells. At the site of the attachment of the bacteria, cytoplasm of the host cell appeared electron dense and / or vesicular (blue arrows). Cytoplasmic extensions from the cell membrane which appears to surround a bacterium is evident (big arrow). The electron dense cytoplasm also is seen to extend into the lumen, like a pedestal which very closely attaches to an attaching bacterium (double arrow). Some of the bacteria are adhered to red blood cell. Lead citrate & uranyl acetate. TEM X 4000.

Electron micrograph of the trachea of a mouse from group 4 that died 20 hours p.i.. Note a bacterium attaches to a pedestal – like lesion of the cytoplasm. Note that the pedestal is not electron dense but a focus of cytoplasmic increased in electron density present adjacent to the pedestal (arrow). Lead citrate & uranyl acetate. TEM X 3074.

Electron micrograph of the trachea of mouse from group 5 that died 22 hours p.i.. Note that the bacteria (short arrow) is attached to the enlarged deformed and shortened cilia. The outlines of the deformed cilia are electron dense (long arrows). Lead citrate & uranyl acetate. TEM X 4500.

Electron micrograph of the lung of a mouse from group that died 18 hours p.i.. Note the bacteria is evident close to the surface of apoptotic cell. Cell membrane of the apoptotic endothelial adjacent to the attached bacteria appeared depressed / invaginated. On the surface of the cell on both sides of the attached bacterium, an electron dense fuzzy material is evident (arrow). Lead citrate & uranyl acetate. TEM X 3074.

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3.30 Electron micrograph of the lung of a mouse from group 5 that died 22 hours p.i.. Note that the neutrophil contained bacteria within vacuole (arrow). Lead citrate & uranyl acetate. TEM X 2459.

3.31 Electron micrograph of the lung of a mouse from group 4 that died 22 hours p.i.. Note the presence of bacterial vacuoles (arrows) in the cytoplasm of the alveolar macrophages. Lead citrate & uranyl acetate. TEM X 2459.

3.32 Electron micrograph of the lung of a mouse from group 5 that died 22 hours p.i.. Note that the endothelial cell (big arrow) is undergoing apoptosis with complete condensation of the nucleus. Bacteria within vacuoles are seen (small arrows) in the cytoplasm of neutrophil. Lead citrate & uranyl acetate. TEM X 1936.

3.33 Electron micrograph of the lung of a mouse from group 4 that died 20 hours p.i.. Note the presence of a bacterium (arrow) in the cytoplasm of an apoptotic pneumocyte. This apoptotic pneumocyte has margination of chromatin. Lead citrate & uranyl acetate. TEM X 1840.

3.34 Electron micrograph of the lung of a mouse from group 4 that died 20 hours p.i.. Note erythrocytes filled the lumen of the capillaries. The endothelial cells of the capillaries are swollen. Capillaries basement membranes are discontinued and are infiltrated with precipitate of fibrin (small arrow). Bacteria are also seen adhered to necrotic cell (big arrow). Sloughed necrotic cells were also seen in the alveolar lumina (double arrow). Lead citrate & uranyl acetate. TEM X 1537.

3.35 Electron micrograph of the lung of a mouse from group 5 that died 22 hours p.i. Note the fibrin was present throughout the lesions. The endothelial cells are disorganized and necrotic. Bacteria (arrow) are seen attached to necrotic endothelial cells. Lead citrate & uranyl acetate. TEM X 1667.

3.36 Electron micrograph of the lung of a mouse from group 6 that died 24 hours p.i.. Note swelling of endothelial cells with severe condensation of chromatin at the periphery of the nuclei and severe vacuolation of their cytoplasm (big arrows). Fragmentation of endothelial cell nucleus (short arrow) is also seen. Pneumocytes