



**UNIVERSITI PUTRA MALAYSIA**

***EFFECT OF DETERGENT, HERBS AND SPICES ON THE GROWTH OF  
Vibrio parahaemolyticus BIOFILM IN SEAFOOD***

***ELEXSON NILLIAN***

**FSKTM 2014 19**



**EFFECT OF DETERGENT, HERBS AND SPICES ON THE GROWTH OF  
*Vibrio parahaemolyticus* BIOFILM IN SEAFOOD**

**By**

**ELEXSON NILLIAN**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in  
Fulfilment of the Requirements for the Degree of Doctor of Philosophy**

**September 2014**

## **COPYRIGHT**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



**Dedicated to God for the love, wisdom and strength**

**Dedicated to my family and my relatives for their unconditional love and endless support**

**Dedicated to my friends for the wonderful friendship, love and joy**

**Dedicated to everyone whom have invested their lives in my life**

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the Degree of Doctor of Philosophy

**EFFECT OF DETERGENT, HERBS AND SPICES ON THE GROWTH OF  
*Vibrio parahaemolyticus* BIOFILM IN SEAFOOD**

By

**ELEXSON NILLIAN**

**September 2014**

**Chairman: Professor Son Radu, PhD**  
**Faculty: Food Science and Technology**

The aim of this study was to investigate the effects of detergent, herbs, and spices on antimicrobial activity and specificity to mitigate the growth of *V. parahaemolyticus* biofilm. A total of 394 samples were collected from wet markets (n=201) and hypermarkets (n=193), consisting of cockles (*Anadara granosa*), clams (*Mya arenaria*), shrimps (*Penaeus spp.*) and squids (*Loligo opalescens*). The prevalence of 9.14% (36/394) of positive *toxR* genes, 63.8% of positive isolates contained virulent genes of *V. parahaemolyticus* *tdh*+, 22.2% of *V. parahaemolyticus* *trh*+, and 5.56% (2/36) of positive isolates in both *V. parahaemolyticus* *tdh*+ and *trh*+ had been detected. The thirty six (n=36) isolates were further tested with antibiotic susceptibility test and they were found to be resistant to at least seven out of eighteen antibiotics. *V. parahaemolyticus* isolates (97.2%) showed Multiple Antibiotics Resistance index > 0.2, indicating that these isolates might have originated from high risk source. Next, RAPD-PCR was carried out to investigate the relationship between genetic diversity and clonal among the strains. As dendrogram was generated among the four groups, group Type B was the largest group that possessed 92.86% (13/14) of isolates from bivalve seafood, such as cockles and clams. The samples were derived from wet markets and they are closely related. RAPD-PCR fingerprinting and antibiotic resistance profiling indicated multiple antibiotics resistant *V. parahaemolyticus* were wide-spread among the groups. The isolates were examined in the assessment of biofilm producer at different temperatures. *V. parahaemolyticus* could form biofilm and presence of biofilm was detected at 37 °C (optimum temperature), followed by room temperature (25 °C), and 4°C (chiller temperature). The representatives of clonal from each group, which were resistant antibiotics of *V. parahaemolyticus*, were examined for effect of detergents. Out of the three detergents; Detergent 1 (Linear Alkylbenzene sulphonate based), Detergent 2 (Quaternary Ammonium based), and Detergent 3 (Sodium Hydroxide based) with minimum concentration of 100 mg/ml (100,000 µg / ml), Detergent 1 was found to be the most effective. As for Detergent 1, the Minimum Inhibition Concentrations (MICs) ranged from 97.656 - 1562.5 µg/ml, while the Minimal Bactericidal

Concentrations (MBCs) were at 781.25 - 3125 µg/ml. Meanwhile, inhibition time assay showed that the bactericidal activity of detergent was fast-acting against antibiotics resistant *V. parahaemolyticus* at 8× MIC within 1 hr and the reduction in CFU/ml was 3 log units (99.9%) with P value < 0.05 was considered statistically significant. The growth of antibiotics resistant *V. parahaemolyticus* biofilm was inhibited at 1562.5 – 6250 µg/ml and was eradicated at 3125 – 12500 mg/ml. Then, the study was furthered by using natural antimicrobial herbs and spices in inhibiting and in preventing the growth of *V. parahaemolyticus* biofilm. Out of the twenty herbs and spices, four of them, which were cloves, star anise, lemon leaves, and curry leaves, showed some activities towards the antibiotics resistant isolates. At a minimum of 10 mg/ml (1000 µg/ml), clove was the most effective as it showed MICs at 19.531-78.125 µg/ml and MBC at 78.125-625 µg/ml. The killing time activity with cloves was fast-acting against antibiotic resistant *V. parahaemolyticus* at 8× MIC in 0.5hr time, while 1 hr, 4 hr, and 8 hr with star anise, lemon leaves, and curry leaves extracts. The reduction in CFU/ml was 3 log units (99.9%). Besides, cloves demonstrated concentration that ranged from 78.125 – 156.25 µg/ml, and 312.5 – 625 µg/ml to inhibit and to totally eradicate the growth of antibiotic resistant *V. parahaemolyticus* biofilm. Thus, these findings can be used to mitigate the resistance of biofilm growth in any food supply chain related to seafood in future.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

**KESAN DETERGEN, HERBA DAN REMPAH RATUS KE ATAS  
PERTUMBUHAN *Vibrio parahaemolyticus* BIOFILEM DALAM MAKANAN  
LAUT**

Oleh

**ELEXSON NILLIAN**

**September 2014**

**Pengerusi : Profesor Son Radu, PhD**  
**Fakulti : Sains dan Teknologi Makanan**

Penyelidikan ini bertujuan untuk mengenalpasti kesan detergen, herba dan rempah ratus dengan aktiviti mikrobial yang tinggi dan spesifikasi khusus terhadap pengurangan pertumbuhan bio filem *V. parahaemolyticus*. Sejumlah 394 sampel telah di kumpul daripada pasar borong (n=201) dan pasaraya (n=193) yang terdiri daripada kerang (*Anadara granosa*), kepah (*Mya arenaria*), udang (*Penaeus spp.*) dan sotong (*Loligo opalescens*). Kelaziman sebanyak 9.14 % (36/394) positif gen *toxR*, 63.8% kehadiran gen virulen *V. parahaemolyticus* *tdh+*, 22.2% *V. parahaemolyticus* *trh+*, dan 5.56% (2/36) pencilan *V. parahaemolyticus* mempunyai kedua-dua *tdh+* and *trh+* gen. Sejumlah tiga puluh enam (n= 36) pencilan tersebut telah di uji terhadap ujian keberkesanan antibiotik dan kepelbagaian rintangan kepada sekurangnyanya terhadap tujuh daripada lapan belas antibiotik yang di kenalpasti. Pencilan *V. parahaemolyticus* (97.2%) menunjukkan kepelbagaian rintangan indek > 0.2, menunjukkan pencilan ini kemungkinan bersal daripada sumber yang berisiko tinggi. RAPD - PCR telah di lanjutkan dengan mengenal pasti kepelbagaian genetik dan pengkaitan klon di antara pencilan. Setelah dendogram di hasilkan, daripada empat kumpulan, kumpulan jenis B adalah kumpulan terbesar mengandungi 92.86% ( 13/14) pencilan terdiri daripada makanan laut seperti kerang dan kepah. Dimana diperolehi daripada pasar borong dan berkait rapat. Kaedah RAPD-PCR cap jari dan kepelbagaian rintangan antibiotik profil menunjukkan pengandaan kepelbagaian rintangan antibiotik *V. parahaemolyticus* tersebar secara meluas di antara kumpulan. Pencilan diperiksa dengan mengakses penghasilan bio filem di dalam suhu yang berbeza *V. parahaemolyticus* boleh di bentuk dan terdapat pertumbuhan bio filem di 37 ° C (suhu optimum), di ikuti oleh suhu bilik (25 °C) and with 4° C (suhu pendingin). Wakil dari setiap klon daripada setiap kumpulan yang mempunyai pengandaan rintangan antibiotik *V. parahaemolyticus* telah di periksa ke atas kesan detergen. Daripada tiga pencuci, detergen 1, (berasaskan Linear Alkylbenzene sulphonate), detergen 2 (berasaskan Quaternary Ammonium) dan detergen 3 (berasaskan Sodium Hydroxide) mempunyai kepekatan minimum

sebanyak 100 mg/ml (100000 µg/ml), detergen 1 di dapati paling efektif. Untuk detergen 1, Perencatan kepekatan minimum (MICs) diantara 97.656 - 1562.5 µg/ml manakala minimum kepekatan penbunuhan (MBCs) di antara 781.25 - 3125 µg/ml. Assay masa membunuh menunjukkan aktiviti pembunuhan pencuci adalah bertindak pantas melawan rintangan antibiotik *V. parahaemolyticus* di 8× MIC di dalam 1 jam dan pengurangan di CFU/ml adalah 3 log units (99.9%) dengan nilai  $P < 0.05$  adalah di anggap tidak ada perbezaan signifikan statistik. Pertumbuhan bio filem rintangan antibiotik *V. parahaemolyticus* boleh di rencat di 1562.5 – 6250 µg/ml di basmi di 3125 – 12500 mg/ml. Kemudian, kajian telah di lanjutkan untuk memeriksa menggunakan herba dan rempah ratus untuk merencat dan menghalang pertumbuhan *V. parahaemolyticus* bio filem. Daripada dua puluh herba dan rempah ratus, empat termasuk cengkih, bunga lawang, daun limau dan daun kari menunjukkan aktiviti terhadap pencilan rintangan antibiotik. Di kepekatan minimum 10 mg/ml (1000 µg/ml), cengkih adalah paling efektif menunjukkan MICs di 19.531 - 78.125 µg/ml dan MBC 78.125 - 625 µg/ml. Aktiviti masa membunuh cengkih adalah pantas bertindak terhadap rintangan antibiotik *V. parahaemolyticus* di 8× MIC dalam 0.5 jam manakala 1 jam, 4 jam and 8 jam oleh ekstrak bunga lawang, daun limau dan daun kari, Pengurangan di CFU/ml adalah 3 log units (99.9%). Cengkih demonstrasi kepekatan di antara 78.125 – 156.25 µg/ml dan 312.5 – 625 µg/ml untuk merencat dan membasmi sepenuhnya pertumbuhan bio filem *V. parahaemolyticus*. Maka, penemuan ini boleh digunakan untuk mengurangkan pertumbuhan kerintangan bio filem di dalam rantai bekalan makanan berhubung dengan makanan laut di masa hadapan.



## ACKNOWLEDGEMENTS

I want to thank God for the wisdom that He has given to me and through His guidance and mercy; He had granted me with unconditional love that He had shown me. Although my name appears on the cover of this thesis as the author, those closest to me know that it never would have been written without the extraordinary godly men and women who invested in me over the years. Without all this, I never have the perseverance to pursue and complete Doctor of Philosophy.

I would like to dedicate my heartfelt thanks to Supervisory Committee Chairman, Professor Dr. Son Radu, who had contributed tremendously of his time and expertise in this research. His encouragement motivation, guidance even trust has always made me a visionary person.

My gratitude also goes to Associate Professor, Dr. Yaya Rukayadi as my Supervisory Committee for his comments, suggestion and support as well as reviewing my thesis. Thank you so much for his patience and for always believing in me and seeing my potential in this field. Thank you very much to him because he always with me when I need support and encouragement to continue my doctorate journey until the end. A million thank to Associate Professor, Dr. Nor Ainy Mahyudin for the continuous guidance, support and opportunity that's she give to me throughout the year. I would like to specially thank Professor Dr. Mitsuaki Nishibuchi and Dr. Yoshitsugu Nakaguchi from Kyoto University of Japan for their collaboration and support in this research. Not forgotten, Dr. Tuan Zainazor, who always motivate, encourage and guide me through all this study.

I would like to express my sincere appreciation to my friends Lye Ying Ling, Sylvester William, UbongAnyi, ElhamTaghavi, Aimi, Soopna, Najwa, Malcom, Ah Tong, Wei San, Nina, Dr. Natasha Lee and Dr. Noorlis Ahmad for sharing your experience and life with me in the Food Safety Research Centre. Thank you very much for your time, advice, guidance to me when I was is your friend.

My deepest appreciation with forever loves to my family: my wonderful mum, dad, brothers and sisters, uncles and aunties who really trust, caring and love me. Thank you so much when the world against me, you are always there to support me. I proud to have your in my life. To my little girl, Elisha, you always be my motivation, my love and purpose why I keep still finishing this journey.

A special appreciation goes to my members Brother Jhawn, Brother Mervin and wife, Dr. Tunung, Koko Wagner and wife, Kak Rose, Vella who teach me about life and who are always with me during the whole process of Doctorate study. UniCO, you are the inspirational, vision, passion and journey in my life to be History and Impact maker. You are awesome.

I would like to express my loving thanks to wonderful and great people in my life who really make my life more beautiful. I do not forget, although I was hospitalized; go in and out from the hospital due to my unexpected health problem, you always there to accompany me with prayer and support. That made me still continues my doctorate journey. This is for all of you.



I certify that a Thesis Examination Committee has met on 23 June 2014 to conduct the final examination of Elexson Nillian on his thesis entitled "Effect of Detergent, Herbs and Spices on the Growth of *Vibrio parahaemolyticus* Biofilm in Seafood" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

**Jinap binti Selamat, PhD**

Professor  
Faculty of Food Science and Technology  
Universiti Putra Malaysia  
(Chairman)

**Vasanth Kumari Neela, PhD**

Associate Professor  
Faculty of Medicine and Health Science  
Universiti Putra Malaysia  
(Internal Examiner)

**Wan Mohtar Wan Yusoff, PhD**

Professor  
Universiti Kebangsaan Malaysia  
Malaysia  
(External Examiner)

**I. Karunasagar, PhD**

Professor  
Karnataka Veterinary, Animal and Fisheries Sciences University  
India  
(External Examiner)



---

**ZULKARNAIN ZAINAL, PhD**  
Professor and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 9 December 2014

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

**Son Radu, PhD**

Professor  
Faculty of Food Science and Technology  
Universiti Putra Malaysia  
(Chairman)

**Yaya Rukayadi, PhD**

Associate Professor  
Faculty of Food Science and Technology  
Universiti Putra Malaysia  
(Member)

**Nor Ainy binti Mahyudin, PhD**

Associate Professor  
Faculty of Food Science and Technology  
Universiti Putra Malaysia  
(Member)

---

**BUJANG BIN KIM HUAT, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:

## Declaration by the student

I hereby confirm that:

- this thesis is my original work
- quotations, illustrations and citations have been duly referenced
- the thesis has not been submitted previously or concurrently for any other degree at any institutions
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be owned from supervisor and deputy vice –chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No: Elexson Nillian (GS33477)

## Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) were adhered to.



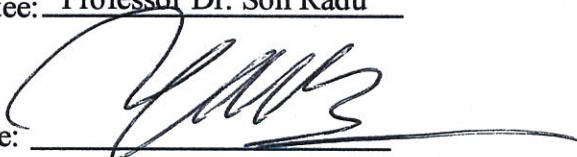
Signature: \_\_\_\_\_

Name of

Chairman of

Supervisory

Committee: Professor Dr. Son Radu



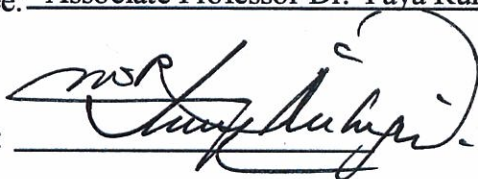
Signature: \_\_\_\_\_

Name of

Member of

Supervisory

Committee: Associate Professor Dr. Yaya Rukayadi



Signature: \_\_\_\_\_

Name of

Member of

Supervisory

Committee: Associate Professor Dr. Nor Ainy binti Mahyudin

## TABLE OF CONTENTS

|          |  |             |
|----------|--|-------------|
|          | <b>ABSTRACT</b>  | <b>Page</b> |
|          | <b>ABSTRAK</b>   | i           |
|          | <b>ACKNOWLEDGEMENTS</b>  | iii         |
|          | <b>APPROVAL</b>  | v           |
|          | <b>DECLARATION</b>   | vii         |
|          | <b>LIST OF TABLES</b>  | xiv         |
|          | <b>LIST OF FIGURES</b>   | xv          |
|          | <b>LIST OF ABBREVIATIONS</b>   | xvii        |
|          |  | xix         |
| <br>     | <b>CHAPTER</b>   |             |
| <b>1</b> | <b>GENERAL INTRODUCTION</b>  |             |
| 1.1      | Introduction   | 1           |
| 1.2      | Objectives   | 2           |
| <br>     | <b>2 LITERATURE REVIEW</b>   |             |
| 2.1      | The need to mitigate biofilm formation in seafood                    | 3           |
| 2.2      | Current researches on biofilm formation mitigation                   | 4           |
| 2.2.1    | Enzyme-based detergents  | 4           |
| 2.2.2    | Biofilm mitigation using phages                                      | 4           |
| 2.2.3    | Biofilm mitigation using microbial interactions/metabolite molecules | 5           |
| 2.2.4    | Quorum sensing cell-to-cell communication                            | 5           |
| 2.3      | Preventing biofilm formation   | 6           |
| 2.3.1    | Future researches on mitigation biofilm                              | 6           |
| 2.4      | Introduction to biofilm formation                                    | 7           |
| 2.5      | Food safety concern  | 9           |
| 2.5.1    | The importance of safe seafood                                       | 9           |
| 2.5.2    | Health benefits related to seafood consumption                       | 10          |
| 2.6      | World seafood industry   | 10          |
| 2.6.1    | Seafood industry in Malaysia   | 11          |
| 2.7      | Public health impact of <i>Vibrios</i>                               | 11          |
| 2.8      | Background of <i>Vibrios</i>   | 13          |
| 2.9      | Taxonomy of <i>Vibrio parahaemolyticus</i>                           | 14          |
| 2.10     | Bacteriology of <i>Vibrio parahaemolyticus</i>                       | 15          |
| 2.11     | Ecology of <i>Vibrio parahaemolyticus</i>                            | 17          |
| 2.12     | Epidemiology of <i>Vibrio parahaemolyticus</i>                       | 18          |
| 2.13     | Clinical manifestation   | 19          |
| 2.14     | Disease related to <i>Vibrio parahaemolyticus</i>                    | 20          |
| 2.15     | Isolation and identification of <i>V. parahaemolyticus</i>           | 20          |
| 2.16     | Rapid detection methods  | 22          |
| 2.16.1   | Polymerase Chain Reaction (PCR)                                      | 22          |
| 2.16.2   | Randomly Amplified Polymorphic DNA(RAPD)                             | 24          |
| 2.17     | Antibiotics susceptibility test                                      | 26          |
| 2.18     | <i>In vitro</i> methods  | 27          |
| 2.18.1   | Agar diffusion   | 27          |

|          |  |    |
|----------|--|----|
|          | 2.81.2 Agar and broth dilution   | 28 |
|          | 2.18.3 Inhibition or time kill curve   | 29 |
| 2.19     | Antibacterial properties in detergents   | 30 |
|          | 2.19.1 Acids   | 31 |
|          | 2.19.2 Alcohols  | 31 |
|          | 2.19.3 Alkalis   | 31 |
|          | 2.19.4 Oxidizing agent   | 31 |
|          | 2.19.5 Quaternary ammonium compounds   | 32 |
| 2.20     | Natural antimicrobial agents   | 32 |
|          | 2.20.1 Antimicrobial activities of herbs and spices  | 32 |
|          | 2.20.2 Cloves  | 33 |
|          | 2.20.3 Star Anise  | 35 |
|          | 2.20.4 Lemon leave   | 35 |
|          | 2.20.5 Curry leave   | 36 |
| 2.21     | Methods of extraction  | 36 |
|          | 2.21.1 Water   | 37 |
|          | 2.21.2 Acetone   | 37 |
|          | 2.21.3 Alcohol   | 37 |
|          | 2.21.4 Methanol  | 38 |
| <b>3</b> | <b>PREVALENCE OF <i>Vibrio parahaemolyticus</i> FROM SEAFOOD</b>   |    |
| 3.1      | Introduction   | 40 |
| 3.2      | Materials and Methods  | 42 |
|          | 3.2.1 Sampling and bacterial isolation   | 41 |
|          | 3.2.2 Culturing methods  | 41 |
|          | 3.2.3 Extraction of genomic DNA  | 41 |
|          | 3.2.4 Amplification of <i>Vibrio parahaemolyticus</i> <i>toxR</i> genes  | 42 |
|          | 3.2.5 Amplification for virulent genes <i>trh</i> <sup>+</sup> and <i>tdh</i> <sup>+</sup>                             | 42 |
|          | 3.2.6 Gel electrophoresis  | 42 |
| 3.3      | Results and discussion   | 43 |
| 3.4      | Conclusion   | 50 |
| <b>4</b> | <b>PROFILING OF <i>Vibrio parahaemolyticus</i> FROM SEAFOOD BASED ON ANTIBIOTIC RESISTANCE AND RAPD FINGERPRINTING</b> |    |
| 4.1      | Introduction   | 51 |
| 4.2      | Materials and Methods  | 52 |
|          | 4.2.1 <i>Vibrioparahaemolyticus</i> isolates   | 52 |
|          | 4.2.2 Antibiotic susceptibility test   | 52 |
|          | 4.2.3 Disc diffusion method  | 52 |
|          | 4.2.4 Multiple Antibiotic Resistance (MAR)   | 53 |
|          | 4.2.5 Randomly Amplified Polymorphic DNA (RAPD)  | 53 |
|          | 4.2.6 RAPD-PCR reaction  | 54 |
|          | 4.2.7 DNA Ladder   | 54 |
|          | 4.2.8 RAPD profile analysis  | 54 |
| 4.3      | Results and discussion   | 54 |
| 4.4      | Conclusion   | 65 |



|          |  |    |
|----------|--|----|
| <b>5</b> | <b>ASSESSMENT BIOFILM FORMATION FOR <i>Vibrio parahaemolyticus</i> AT DIFFERENT TEMPERATURE</b>                                    |    |
| 5.1      | Introduction   | 56 |
| 5.2      | Materials and Methods  | 56 |
|          | 5.2.1 Bacterial strain and culture conditions  | 56 |
|          | 5.2.2 <i>In vitro</i> biofilm formation of <i>Vibrio parahaemolyticus</i> at different temperatures                                | 67 |
|          | 5.2.3 Quantification of <i>Vibrio parahaemolyticus</i> biofilm   | 67 |
|          | 5.2.4 Statistical analysis   | 68 |
| 5.3      | Results and discussion   | 68 |
| 5.4      | Conclusion   | 75 |
| <b>6</b> | <b>EFFECT OF DETERGENT AS AN ANTIBACTERIAL AGENT ON THE GROWTH OF ANTIBIOTICS RESISTANT <i>Vibrio parahaemolyticus</i> BIOFILM</b> |    |
| 6.1      | Introduction   | 76 |
| 6.2      | Materials and Methods  | 77 |
|          | 6.2.1 Bacterial strains and preparation of inoculums   | 77 |
|          | 6.2.2 Preparation of detergent assay as antibacterial agent  | 77 |
|          | 6.2.3 <i>In vitro</i> susceptibility test  | 77 |
|          | 6.2.4 The determination of Minimum inhibitory concentrations (MICs) and minimal bactericidal concentrations (MBCs)                 | 78 |
|          | 6.2.5 Times–kill assays  | 78 |
|          | 6.2.6 Sessile minimum inhibitory concentration (SMICs) and minimum bactericidal eradication concentration (MBECs)                  | 79 |
|          | 6.2.7 Statistical analysis   | 79 |
| 6.3      | Results and discussion   | 80 |
| 6.4      | Conclusion   | 92 |
| <b>7</b> | <b>EFFECT OF HERBS AND SPICES ON THE GROWTH OF ANTIBIOTICS RESISTANT <i>Vibrio parahaemolyticus</i> BIOFILM</b>                    |    |
| 7.1      | Introduction   | 93 |
| 7.2      | Materials and Methods  | 94 |
|          | 7.2.1 Preparation of extraction  | 94 |
|          | 7.2.2 Bacterial strains and preparation of inoculums   | 95 |
|          | 7.2.3 Assay of herbs and spices as antimicrobial agents  | 95 |
|          | 7.2.4 <i>In vitro</i> susceptibility tests   | 95 |
|          | 7.2.5 The determination of minimum inhibitory concentrations (MICs) and minimal bactericidal concentrations (MBCs)                 | 96 |
|          | 7.2.6 Times–kill assays  | 96 |
|          | 7.2.7 Assessment of <i>in vitro</i> biofilm produced by <i>V. parahaemolyticus</i> strains   | 87 |
|          | 7.2.8 Sessile minimum inhibitory concentration (SMICs) and minimum eradication bactericidal concentration (MBECs)                  | 97 |
|          | 7.2.9 Statistical analysis   | 98 |

|          |   |            |
|----------|---|------------|
| 7.3      | Results and discussion  | 98         |
| 7.4      | Conclusion  | 114        |
| <b>8</b> | <b>GENERAL DISCUSSION, CONCLUSION AND<br/>RECOMMENDATIONS</b> | <b>115</b> |
|          | <b>REFERENCES/BIBLIOGRAPHY</b>                                | <b>118</b> |
|          | <b>APPENDICES</b>   | <b>139</b> |
|          | <b>BIODATA OF STUDENT</b>                                     | <b>199</b> |
|          | <b>LIST OF PUBLICATIONS</b>                                   | <b>200</b> |



## LIST OF TABLES

| Table   | Page |
|---|------|
| 2.1 Growth limits and optimum conditions for <i>V. parahaemolyticus</i> in comparison to <i>V. cholera</i> and <i>V. vulnificus</i> .                 | 15   |
| 2.2 Biochemical traits used in the differentiation of some <i>Vibrio</i> species.   | 16   |
| 2.3 Flagellation patterns of <i>Vibrio</i> spp.   | 17   |
| 2.4 Symptoms of patients with gastroenteritis due to <i>V. parahaemolyticus</i> infection.  | 20   |
| 2.5 Comparison between TCBS agar and chrom agar   | 22   |
| 2.6 Terms used in antibacterial activity testing  | 29   |
| 2.7 Components of clove oil   | 34   |
| 3.1 Prevalence of <i>V. parahaemolyticus</i> in seafood samples   | 44   |
| 4.1 Distribution of antimicrobial resistance of <i>V. parahaemolyticus</i> isolates obtained from seafood.  | 56   |
| 4.2 The patterns of antibiotic resistance profile and Multiple Antibiotic Resistance (MAR) index of <i>V. parahaemolyticus</i> obtained from seafood. | 58   |
| 4.3 <i>V. parahaemolyticus</i> isolates recovered from seafood samples according to the sampling location   | 62   |
| 4.4 Random Amplified Polymorphism DNA (RAPD) types and subtypes of <i>V. parahaemolyticus</i> s isolates  | 63   |
| 5.1 The number of biofilm produced by <i>V. parahaemolyticus</i> at different temperatures analyzed in a number of samples                            | 69   |
| 6.1 Effect of detergents to antibiotics resistant <i>V. parahaemolyticus</i>  | 83   |
| 6.2 Comparison of <i>in vitro</i> activities for detergents against antibiotics resistant <i>V. parahaemolyticus</i>                                  | 84   |
| 6.3 Summary of time-kill curve using Fisher's Exact for <i>V. parahaemolyticus</i> ATCC 17802 and VP003   | 86   |
| 6.4 Comparative study of SMIC and MBEC activities of detergents against growth of <i>V. parahaemolyticus</i> biofilm                                  | 90   |
| 6.5 Effect of detergents against planktonic cells (MICs and MI and biofilm growth (SMICs and MBECs) of antibiotics resi <i>V. parahaemolyticus</i>    | 91   |
| 7.1 Susceptibility activities of antibiotics resistant <i>V. parahaemolyticus</i> against herbs and spices  | 101  |
| 7.2 Effect of herbs and spices on antibiotic resistant of <i>V. parahaemolyticus</i>  | 102  |
| 7.3 Comparative <i>in vitro</i> activities of cloves, star anise, lemon and curry leaves against antibiotics resistant <i>V. parahaemolyticus</i>     | 105  |
| 7.4 Summary of time-kill curve using Fisher's Exact for <i>V. parahaemolyticus</i> ATCC 17802 and VP003   | 110  |
| 7.5 Comparative study of SMIC and MBEC activities of cloves, star anise, lemon leaves and curry leaves against biofilm of <i>V. Parahaemolyticus</i>  | 113  |

|     |   |
|-----|---|
| 7.6 | Effect of herbs and spices against planktonic cells (MICs and MBCs) and biofilm formation (SMICs and MBECs) of antibiotics resistant <i>V. parahaemolyticus</i> |
|-----|---|

114



## LIST OF FIGURES

| Figure  |  | Page |
|---------|--|------|
| 2.1     | Stages in biofilm formation  | 8    |
| 3.1 A   | Representative amplification of <i>toxR</i> gene for the detection of <i>V. parahaemolyticus</i> (368 bp). Lane M: 100 bp DNA ladder; Lane 1 and 2: representative for positive samples at 368 bp; Lane 3: positive control at 368 bp and Lane 4: negative sample. | 43   |
| 3.1 B   | Representative amplification for the detection of <i>V. parahaemolyticus tdh<sup>+</sup></i> gene (251 bp). Lane M: 100 bp DNA ladder; Lanes 1 and 2: representatives of positive samples; Lane 3: positive control and Lane 4: negative sample                    | 45   |
| 3.1 C   | Representative amplification for the detection of <i>V. parahaemolyticus trh</i> gene (250 bp). Lane 1 and 2: representatives of positive samples; Lane 3: positive control; Lane M: 100 bp DNA ladder and Lane 4: negative sample.                                | 45   |
| 3.2     | Percentage of comparison between Wet market (PBS) (n=201) and Hypermarket (HMG)(n=193)   | 47   |
| 3.3     | Percentage of <i>V. parahaemolyticus toxR</i> gene in seafood sample   | 48   |
| 3.4     | Percentage of <i>V. parahaemolyticus trh</i> and <i>tdh</i> genes in seafood sample  | 49   |
| 4.1     | Dendrogram of RAPD profiles for <i>V. parahaemolyticus</i> strains using OPA8 and OPA10  | 94   |
| 5.1     | Formation of <i>V. parahaemolyticus</i> biofilm from seafood samples in 37 °C, Note : Lane 1-36 : <i>V. parahaemolyticus</i> and Lane 37 : <i>V. parahaemolyticus</i> ATCC 17802, Lane 38 : Negative control(fresh TSB)  | 70   |
| 5.2     | Formation of <i>V. parahaemolyticus</i> biofilm from seafood samples in room temperature(25 °C), , Note : Lane 1-36 : <i>V. parahaemolyticus</i> , Lane 37 : <i>V. parahaemolyticus</i> ATCC 17802 and Lane 38 : Negative control(fresh TSB)                       | 71   |
| 5.3     | Formation of <i>V. parahaemolyticus</i> biofilm from seafood samples at 4 °C, Note : Lane 1-36 : <i>V. parahaemolyticus</i> and Lane 37 : <i>V. parahaemolyticus</i> ATCC 17802, Lane bar 38 : Negative control(fresh TSB)   | 72   |
| 6.1     | Representative of inhibition zone for disk diffusion using D1 on antibiotics resistant <i>V. parahaemolyticus</i>  | 81   |
| 6.2 (A) | Activity of D1 (alkylbenzene sulphonate based) against <i>V. parahaemolyticus</i> ATCC 17802   | 67   |
| 6.2 (B) | Activity of antibiotics resistant <i>V. parahaemolyticus</i> VP003 against detergent 1   | 68   |
| 7.1     | Steps of extraction  | 94   |

|         |  |     |
|---------|--|-----|
| 7.2     | Representative of inhibition zone for disk diffusion test using cloves on antibiotics resistant <i>V. parahaemolyticus</i> | 99  |
| 7.3 (A) | The activity of cloves against control ATCC 17082  | 107 |
| (B)     | Activity of antibiotics resistant <i>V. parahaemolyticus</i>   | 108 |
| (C)     | VP003 strain against extracts cloves, star anise, lemon  |     |
| (D)     | leaves and curry leaves  |     |
| (E)     |  |     |



## LIST OF ABBREVIATIONS

|                   |   |   |
|-------------------|---|---|
| µl                | - | Microliter  |
| µm                | - | Micrometer  |
| µg                | - | Microgram   |
| ADI               | - | Acceptable daily intake                             |
| ATCC              | - | American Type Culture Collection                    |
| BAM               | - | Bacteriological Analytical Manual                   |
| bp                | - | base pair   |
| CDC               | - | Centre for Disease Control                          |
| D1                | - | Detergent 1   |
| D2                | - | Detergent 2   |
| D3                | - | Detergent 3   |
| DNA               | - | Deoxynucleic acid                                   |
| dNTP              | - | deoxyribo nucleoside triphosphate                   |
| EDTA              | - | Ethylene diamine tetra-acetic acid                  |
| FAO               | - | Food and Agricultural Organization                  |
| FDA               | - | Food and Drug Administration                        |
| HIV               | - | Human immunodeficiency virus                        |
| ICMSF             | - | International Commission on Microbiological         |
| MAR               | - | Multiple Antibiotic Resistance                      |
| mg                | - | milligram   |
| MgCl <sub>2</sub> | - | Magnesium Chloride                                  |
| MIC               | - | Minimum inhibitory concentration                    |
| MBC               | - | Minimum inhibitory concentration                    |
| MBEC              | - | Minimum bactericidal eradication concentration      |
| mM                | - | milli Molar   |
| MOH               | - | Ministry of Health                                  |
| MPN               | - | Most probable Number                                |
| MRO               | - | Multiple resistant organism                         |
| NaCl              | - | Sodium chloride                                     |
| NCCLS             | - | National Committee for Clinical Laboratory Standard |
| OD                | - | Optical Density                                     |
| PCR               | - | Polymerase Chain Reaction                           |
| RAPD              | - | Randomly Amplified Polymorphic DNA                  |
| SPB               | - | Salt Polymyxin Broth                                |
| SMIC              | - | Sessile Minimum inhibitory concentration            |
| <i>Taq</i>        | - | <i>Thermus aquaticus</i>                            |
| TBE               | - | Tris-Boric acid-EDTA                                |
| TSA               | - | Trypticase Soy Agar                                 |
| TSB               | - | Tryptic soy broth                                   |
| U                 | - | Unit  |
| U. S              | - | United States                                       |
| UV                | - | Ultra violet  |
| V                 | - | Volt  |
| WHO               | - | World Health Organization                           |
| WTO               | - | World Trade Organization                            |
| x g               | - | unit gravity  |

## CHAPTER 1

### GENERAL INTRODUCTION

#### 1.1 Introduction

In nature, most bacteria exist as plankton (free floating) cells or suspended cells (sessile cells), and some exist on surfaces. According to the National Institute of Health (NIH) in the United States, 80 % of microbial can form biofilm and this includes food borne pathogens, such as *Salmonella* spp., *Klebsiella* spp., *Pseudomonas* spp., *Campylobacter* spp., *Escherichia coli*, and *Listeria* spp. These bacteria are of special significance in ready-to-eat and minimally processed food products (Kim et al., 2006). Bacteria have the capability to attach themselves and to colonize surfaces in many natural and artificial surfaces, which are called biofilms.

Biofilm is known as a complex community that consists of microorganisms embodied in a matrix of extracellular polymeric material that they have generated. Thus, a mass or biofilm is established when bacteria adhere to the surface and produce extracellular polysaccharides (EPS). The EPS matrix consists of not only polysaccharides, but also proteins, which may be the major component in environmental and waste water biofilms and nucleic acids. This environment increases the resistance of bacteria to detergents and antibiotics as the dense extracellular matrix and the outer layer of cells protect the interior of the community (Merill, 2010).

*Vibrio parahaemolyticus* is a bacterium that is dangerous because it is one of the main causes of contamination in seafood, which can cause gastroenteritis worldwide (Lee, & Pogliano, 2007). *V. parahaemolyticus* is marine seafood borne pathogen, which causes gastroenteritis in humans (Miwatani, & Takeda, 1976). It has been recognized as a common cause of food borne illnesses in many Asian countries, including Japan, Taiwan, and China. According to Su and Liu (2007), it is the leading cause of human gastroenteritis in the United States and it is an unsafe seafood borne pathogen throughout the world.

Polymerase Chain Reaction (PCR) was adopted due to its specificity, easily automated, and its ability to amplify the amount of sample within few minutes if food poisoning outbreak occurs. Besides, in order to plan and to monitor targeted intervention strategies at preserving the therapeutic efficacy of antimicrobial agents, surveillance of antibiotic resistance is a key element to provide the latest information on the magnitude and the trends in resistance. On top of that, RAPD-PCR analysis was used to determine the relationship between antibiotic and isolates that were recovered from different habitats based on their RAPD-PCR fingerprints. Antibiotic resistance and RAPD proved to be



effective tools in characterizing and in differentiating the *V. parahaemolyticus* strains (Lesley et al., 2005; Tunung et al., 2007).

In addition, food processing environments provide a variety of conditions, which might favor the formation of biofilm with the presence of moisture, nutrients, and inocula of microorganism from raw materials. Bacterial colonization of food processing equipment and facilities is the main concern and is a potential source of contamination of foods that may lead to spoilage or transmission of food borne pathogens. Moreover, when biofilm detaches from the surfaces, individual microorganisms can easily spread.

In seafood food supply chain from pre harvest to marketable level, safe and effective detergent applied on the food contact surfaces and food facilities or equipment will ensure an acceptable decrease in microbial levels without the presence of toxic residuals or causing any damage for humans and useful forms of life. Natural antimicrobials nowadays are favorable concern to several issues pertaining to microorganisms. These include those that reduce the need of antibiotics and monitor microbial contamination in food. Due to the generally recognized as safe (GRAS) properties, herbs and spices have appeared to be natural antimicrobial, which could meet consumer demands for safe and healthier food consumption without any undesirable effect when applied in the food system.

In seafood industrial practice, it is necessary to develop a complete and a cost-effective cleaning program, which will inhibit the accumulation of both particulates and sessile bacterial cells present on the surfaces of food and processing line. Thus, in order to get rid of this problem, this study was carried out to investigate the effects of detergent, herbs, and spices on the growth of *V. parahaemolyticus* biofilm. Therefore, the objectives of this study were:

1. To determine the prevalence of *Vibrio parahaemolyticus*, regulator gene *toxR* gene, and virulence gene (*trh*, *tdh*) from seafood using polymerase chain reaction.
2. To profile *V. parahaemolyticus* based on antibiotics resistance and Random Amplified Polymorphism DNA–PCR fingerprinting.
3. To compare the formation of *V. parahaemolyticus* biofilms with varying temperatures: chiller temperature (4 °C), room temperature (25 °C), and optimum temperature (37 °C).
4. To determine the effect of detergents against growth of antibiotic-resistant *V. parahaemolyticus* biofilm.
5. To determine the effects of herbs and spices against growth of antibiotic-resistant *V. parahaemolyticus* biofilm.

## REFERENCES

- Abdullah, M. (2010). Biopotency role of culinary spices and herbs and their chemical constituents in health and commonly used spices in Nigerian dishes and snacks. *African Journal of Food Science* 5(3):111-124
- Abraham, T. J., Manly, R., Palaniappan, R. and Dhevendaran, K. (1997). Pathogenicity and antibiotic sensitivity of luminous *Vibrio harveyi* isolated from diseases penaeid shrimp. *Journal of Aquaculture Tropical* 3(12): 1-8.
- Adams, M. R. and Moss, M. O. (2002). Food Microbiology. United Kingdom: *The Royal Society of Chemistry* 6: 142-144.
- Adeleye, A., Vivian, E., Rita, N., Stella, S. and Emmanuel, O. (2008). Antimicrobial susceptibility of potentially pathogenic halophilic *Vibrio* species isolated from seafoods in Lagos, Nigeria. *African Journal of Biotechnology* 7 (20): 3791-3794
- Aditya, M., Vishhal, G., Hemant, N. and Vishal, S. (2013). Protective Effect of (Curry Leaf) Leaves Extract Against Genotoxicity Induced by Cyclophosphamide in Mouse Bone Marrow Cells. *Global Veterinaria* 10 (2): 128-133
- Alam, M. J., Tomochika, K., Miyoshi, S., Shinoda, S. (2002). Environmental investigation of potentially pathogenic *Vibrio parahaemolyticus* in the Seto-Inland Sea, Japan. *Federation of European Microbiological Societies (FEMS) Microbiology Letters* 20: 83-87
- American Heart Association (2013). Accessed on 26<sup>th</sup> June 2013. [http://www.heart.org/HEARTORG/GettingHealthy/NutritionCenter/Fish-101\\_UCM\\_305986\\_Article.jsp](http://www.heart.org/HEARTORG/GettingHealthy/NutritionCenter/Fish-101_UCM_305986_Article.jsp)
- Amit, P. and Parul, S. (2011). Antibacterial activity of *Syzygium aromaticum* (clove) with metal ion effect against food borne pathogens. *Asian Journal of Plant Science and Research* (2):69-80
- Anderson, D. L. and Gibb, A. J. (1994). Identification and phylogeny of spore-cyst fungi (*Ascosphaera* spp.) using ribosomal DNA sequences. *Mycological Research* (102)5: 541-547
- Annous, B. A., Fratomico, P. M., and Smith, J. L. (2009). Scientific status summary. *Journal of Food Science* 74, R24-37
- Aqil, F., Beg, A. and Ahmad, I. (2000). *in vitro* testing of essential oil against soil fungi, *Journal Medicine Aromatic Plant Science* 22(1): 44-46

- Araujo, P., Lemos, M., Mergulhao, F., Melo, L. and Simoes, M. (2011). *In antimicrobial resistance to disinfectants in biofilm* (Ed) Mendez, V. Science against Microbial pathogens: communicating current research and technological advances
- Armstrong, J., Gibbs, A., Peakall, R., and Weiller, G. (1990). RAPD Distance Programme. Australian National University, Canberra, Australia
- Augustin, M., Ali-Vehmas, T., and Atroschi, F. (2004). Assessment of enzymatic cleaning agents and disinfectants against bacterial biofilms. *Journal of Pharmacy and Pharmaceutical Sciences* 18, 55–64
- Austin, B. (2009). Vibrios as causal agents of zoonoses. *Veterinary Microbiology*, doi:10.1016/j.vetmic.2009.03.015 (In Press)
- Australian and New Zealand Food safety, (2014), Accessed on 25th June 2014. <http://www.foodstandards.gov.au/Pages/default.aspx>
- Baggaley, (2008). Antimicrobial Resistance in Developing Countries.. Springer New York Dordrecht Heidelberg, London
- Barry, A. L. (1976). The Antimicrobial Susceptibility Test: Principles and Practices. In V. Lorian, ed. Antibiotics in Laboratory Medicine, 2nd ed. William & Wilkins, Baltimore. Cited P.M. Davidson and M.E. Parish. 1989. Methods for Testing the Efficacy of Food Antimicrobials. *Food Technology* (1): 148-155
- Bartolini, F., Gallerani V., Raggi M., Viaggi D (2005). Contact design and targeting for the production of public goods in agriculture: the impact of the 2003 CAP reform. Proceeding of 11th EAAE congress: “The future of Rural Europe in the global agrifood system”. Copenhagen. 24-27 August 2005
- Bauer, A. W., Kirby, W. M. M., Sherris, J. C., Turck, M. (1966). Antibiotics susceptibility testing by standardized single disk method. *American Journal of Clinical Pathology* 45: 493-496
- Bhattacharya, M., Choudhury, P. and Kumar, R. (2008). Antibiotic and metal-resistant strains of *Vibrio parahaemolyticus* isolated from shrimps. *Microbiology of Drug Resistant* 6: 171-172
- Birkenhauer, J.M., and Oliver, J.D. (2003). Use of diacetyl to reduce the load of *Vibrio vulnificus* in the Eastern oyster, *Crassostrea virginica*. *Journal of Food Protection* 66: 38–43
- Bina, L. J., Tista, P., Anjana, S. and Kayo, D. Y. (2010). Study of antimicrobial activity of lime juice against *Vibrio cholera*. *Scientific World* Vol. 8, No. 8: 7

- Blascek H. P., Wang. H. A. (2007). Biofilm in the Food environment (Eds) Willey Blackwell, Ames, Oxford, Carlton
- Bostock J. C. (2009). Use of information technology in aquaculture. In New technologies in aquaculture: improving production efficiency, quality and environmental management, ch. 35 (eds Burnell G., Allan G., editors.), 1064–1118 Oxford, UK: Woodhead Publishing
- Brooks, J.D. and Flint, S.H. (2008). Biofilms in the food industry: problems and potential solutions. *International Journal of Food Science and Technology* Vol.43, pp. 2163-2176
- Burt, S.A. (2004). Essential oils: Their antibacterial properties and potential applications in foods: A review. *International Journal Food Microbiology* 94: 223-253
- Bhaskar, N., Setty, T.M.R., Mondal, S., Joseph, M.A., Raju, C.V., Raghunath, B.S. and Anantha, C.S. (1998). Prevalence of bacteria of public health significance in the cultured shrimp (*Penaeus monodon*). *Food Microbiology* 15: 511-519
- Boeije, G. M., Schowanek, D. R. and Vanrolleghem, P. A. (2000). Incorporation of biofilm activity in river biodegradation modeling: a case study for linear alkylbenzene sulphonate (LAS). *Water Research* 34(5): 1479-1486
- Bradley, D. (2005). Star role for bacteria in controlling flu pandemic. *Nature reviews. Drug discovery* 4 (12): 945–946
- Broberg, C. A., Calder, T. J. and Orth, K. (2011). *Vibrio parahaemolyticus* cell biology and pathogenicity determinants. *Microbes and Infection* 13: 992-1001
- Brooks, J.D. and Flint, S.H. (2008). Biofilms in the food industry: problems and potential solutions. *International Journal of Food Science and Technology*, Vol.43, pp. 2163-2176
- Canillac, N. and Mourey, A. (2001). Antibacterial activity of the essential oil of *Picea excelsa* on *Listeria*, *Staphylococcus aureus* and coliform bacteria. *Food Microbiology* 18: 261–2
- Carson, C.F., Cookson, B.D., Farrelly, H.D. and Riley, T.V. (1995). Susceptibility of methicillin-resistant *Staphylococcus aureus* to the essential oil of *Melaleuca alternifolia*. *Journal of Antimicrobial Chemotherapy* 35: 421–424
- Cassell, G. H. (1995). ASM Task Force urges broad program on antimicrobial resistance. *ASM News* 61: 116–120

- Centers for Disease Control and Prevention (CDC) (2005). Outbreak of *Vibrio parahaemolyticus* Infection Associated with Eating Raw Oysters and Clams Harvested from Long Island Sound, Connecticut, New Jersey, and New York, Morbidity and Mortality Weekly Report 47: 129-131
- Centers for Disease Control and Prevention (CDC), (2013). *Vibrio* illnesses after Hurricane Katrina multiple states. *Morbidity Mortality Week Report* 54: 928-931
- Chai, L. C., Tunung, R., Usha, M. R., Jurin, W.G., Fatimah, A.B. Farinazleen, M. G., Son, R., and Malakar, P. R. (2007): Thermophilic *Campylobacter* spp. in salad vegetables in Malaysia. *International Journal of Food Microbiology* 117: 106-111
- Chai, L. C., Ghazali, F.M., Bakar, F.A., Lee, H. Y., Suhaimi, R. L., Talib, S. L., Nakaguchi, Y. Nishibuchi, and Son, R. (2009). Occurrence of thermophilic *Campylobacter* spp. contamination on vegetable farms in Malaysia. *Journal of Microbiology and Biotechnology* 19 (11): 1415-1420
- Chaignon, P., Sadovskaya, I., Ragunah, Ch., Ramasubbu, N., Kaplan, J. B., and Jabbouri, S. (2007). Susceptibility of *staphylococcal* biofilms to enzymatic treatments depends on their chemical composition. *Applied Microbiology and Biotechnology* 75, 125–132
- Chakraborty, M. A. (2008). The antioxidant and antimicrobial properties of the methanolic extract from *Cocos nuciferamesocarp*. *Food Chemical* 107: 994-999
- Chang, C. M., Chiang, M.L. and Chou, C. C. (2000). Responses of heat-shocked *Vibrio parahaemolyticus* to subsequent physical and chemical stresses. *Journal of Food Protection* 67: 2183-2188
- Chao, G. X., Jiao, X. N., Zhou, X. H., Wang, F., Yang, Z.Q., Huang, J. L., Pan, Z. M., Zhou, L. P. and Qian, X. Q. (2009). Distribution of genes encoding four pathogenicity Islands VPals., T6SS, Biofilm, and type I Pilus in food and clinical strains of *Vibrio parahaemolyticus* in China. *Foodborne Pathogen Discovery* 76: 649-658
- Chapman, J. S. (2003). Biocide resistance mechanisms. *International Biodeterioration and Biodegradation* 51(2): 133-138
- Chen, Y. T., Chang, H. Y., Lai, Y. C., Pan, C. C., Tsai, S. F. and Peng, H. L. (2004). Sequencing and analysis of the large virulence plasmid pLVPK of *Klebsiella pneumoniae* CG43. *Gene* 337: 189–198



- Chmielewsky, R.A. and Frank, J. F. (2006). Biofilm formation and control in food processing facilities. *Comprehensive Reviews in Food Science and Food Safety* 2: 22 - 32
- Clinical and Laboratory Standards Institute (CLSI) (2009). Document M7- A6, Methods for Dilution Antimicrobial Susceptibility Tests for Bacteria that Grow Aerobically, Approved Guideline, Wayne, PA
- Clinical and Laboratory Standards Institute (2012). Document M7- A6, Methods for Dilution Antimicrobial Susceptibility Tests for Bacteria that Grow Aerobically, Approved Guideline, Wayne, PA.
- Colwell, R. R. (1984). Polyphasic taxonomy of the genus *Vibrio*: numerical taxonomy of *Vibrio cholerae*, *Vibrio parahaemolyticus* and related *Vibrio* species. *Journal of Bacteriology* 104: 410-433
- Colwell, R. R. (2006). A global and historical perspective of the genus *Vibrio*. In *The Biology of Vibrios*, ed. F. L. Thompson, B. Austin, and J. Swings, pp. 3-11. Washington D. C.: ASM Press
- Commission of European Committees (1994). Report of the scientific committee for food.11<sup>th</sup> series.
- Conte, M. P., Longhi, C., Polidoro, M., Petrone, G., Buonfiglio, V., Di Santo, S. (1996). Iron availability affects entry of *Listeria monocytogenes* into the enterocyte like cell line Caco-2. *Infection and Immunity* 64, 3925–3929
- Cook, W. H. and Canning, P. (1990). Inheritance is not subtyping. In *Proc. ACM Symp. on Principles of Programming Languages*.
- Cowan, M. M. (1999). Plant products as antimicrobial agents. *Clinical Microbiology Review* 12: 564-582.
- Cox, J. M. and Fleet, G. H. (2003). New directions in the microbiological analysis of foods. In, *Foodborne Microorganisms of Public Health Significance*, 6th Edition (eds) A.D. Hocking, et al. AIFST (NSW) Food Microbiology Group, Sydney pp.103-162
- Cutler, H., G. (1995). Natural product flavor compounds as potential antimicrobials, insecticides, and medicinals. *Agro Food Industry Hi- Technology* 6; 19–23
- Cressy, H.K., A.R. Jerrett, C.M. Osborne and P.J. Bremer. (2003). A novel method for the reduction of numbers of *Listeria monocytogenes* cells by freezing in combination with an essential oil on bacteriological media. *Journal Food Protection* 66 (3): 390-395

- Daniels, N. A., Ray, B., Easton, A., Marano, N., Kahn, E., McShan, A. L. 2<sup>nd</sup>, Del Rosario, L., Baldwin, T., Kingsley, M. A., Puhr, N. D., Wells, J. G., Angulo, F. J. (2000). Emergence of a new *Vibrio parahaemolyticus* serotype in raw oysters: A prevention quandary. *Journal of the American Medical Association (JAMA)* 284 (12): 1541-1545
- Dao, H. N., Rayner, D. S. and Alvin, W. G. (1999). Marketing and export of marinebased food products. Department of Fisheries, Sabah
- Das, A. K., Rajkumar, V. and Dwivedi, D. K. (2010). Antioxidant effect of curry leaf (*Murraya koenigii*) powder on quality of ground and cooked goat meat. *International Food Research Journal* 18: 563-569
- Davidson, F. M (1997). Chemical preservatives and natural antimicrobial compounds. In: Doyle, M., L.R. Beuchat, T. J. Montville, (Eds). *Food Microbiology: Fundamental and Frontiers*, ASM Press, Washington, D. C. 6: 520-556
- Deans, S. G. and Ritchie, G. (1987). Antibacterial properties of plant essential oils. *International Journal of Food Microbiology* 5: 165– 180
- Delaquis, P. J. K. Stanich, B., Girard, K. and Mazza, G. (2002). Antimicrobial activity of individual and mixed fractions of dill, cilantro, coriander and eucalyptus essential oils. *International Journal Food Microbiology* 74 (1/2): 101-109
- Deepanjali, A., Kumar, H. S., and Karunasagar, I. (2005). Seasonal variation in abundance of total and pathogenic *Vibrio parahaemolyticus* bacteria in oysters along the southwest coast of India. *Applied Environment Microbiology* 71: 3575–3580
- DePaola, A., Capers, G. M., Alexander, D. (1994). Densities of *Vibrio vulnificus* in the intestines of fish from the US Gulf Coast. *Applied and Environmental Microbiology* 60: 984-988
- Desmarchelier, P. M. (1997). Pathogenic Vibrios. In: *Foodborne Microorganisms of Public Health Significance*. 5<sup>th</sup> ed., AIFST (NSW Branch), Food Microbiology Group, North Sydney, NSW: 285-312
- Desmarchelier, P. M. (2003). Pathogenic vibrios. In *Foodborne Microorganisms of Public Health Significance*, ed. Hocking, A.D., pp: 333-358
- Dewanti, R., and Wong, A. C. (1995). Influence of culture conditions on biofilm formation by *Escherichia coli*. *International Journal Food Microbiology* 26:147-164

- Djordjevic, D, Wiedmann, M. and Sborough, L. A. (2002). Microtiter plate assay for assessment of *Listeria monocytogenes* biofilm formation. *Journal Applied Microbiology* 68: 2950-2958
- Dileep, V., Kumar, H. S., Kumar, Y., Nishibuchi, M., Karunasagar, I. and Karunasagar, I. (2003). Application of polymerase chain reaction for detection of *Vibrio parahaemolyticus* associated with tropical seafoods and coastal environment. *Letters in Applied Microbiology* 36: 423-427
- Dong, Y.-H., Gusti, A. R., Zhang, Q., Xu, J.-L., and Zhang, L.-H. (2002). Identification of quorum-sensing N-acyl homoserine lactonases from *Bacillus* species. *Applied and Environmental Microbiology* 68, 1754–1759
- Donlan, R. M. (2002). Biofilms: microbial life on surfaces. *Emerging infectious diseases* 8 (9): 881
- Dorman, H. J. D. and Deans, S. G. (2009). Antimicrobial agents from plants: antibacterial activity of plant volatile oils. *Journal Applied Microbiology* 88: 308- 316
- Duan, J. and Su, Y. C. (2005). Occurrence of *Vibrio parahaemolyticus* in two Oregon oyster-growing bays. *Journal of Food Science* 70: 58-63
- Dvorak, G. (2008). Disinfection 101. *Journal of Food Microbiology* 9; 110-111
- Dwivedi, B. K., Pardley, R. L., Pardley, G., Pant, H. L., and Logani, L. (2002): Evaluation of angiospermic plant extracts against *Rhizopusstolonifer* and *Gloesporiumsidii* of guava. *Biodiversity* 13: 129-134.
- Eckert, K. A., Kunkel, T. A. (1990). High fidelity DNA synthesis by the *Thermusaquaticus* DNA polymerase. *Nucleic Acids Research* 18 (13): 3739-3744
- Eloff, J. N. (1998). Which extractant should be used for the screening and isolation of antimicrobial components from plants. *Journal of Ethnopharmacology* 60: 1–8
- Endzt, H., Ang, C. W., Van den Braak, N., Duim, B., Rigter, A., Price, L. J. (2000). Molecular characterization of *Campylobacter jejuni* from patients with Guillain-Barré and Miller Fisher syndromes. *Journal of Clinical Microbiology* 38: 2297-2301
- European Commission, (2000). Scientific committee on veterinary measures relating to Public Health on *V. vulnificus* and *V. parahaemolyticus* in raw or undercooked seafood.



- Faruque, S. M., Nair, G. B. (2006). Epidemiology. In *The Biology of Vibrios*, ed. F.L. Thompson, B. Austin, and J. Swings, pp. 385-398. Washington D. C.: ASM Press
- Fett, W. F. (2010). Naturally occurring biofilm on alfalfa and other types of sprouts. . *Journal of Food Protection* 63 : 624-632
- Fleming, T. H. (1998). The short-tailed fruit bat: a study of plant-animal interactions. University of Chicago Press, Chicago
- French G. L. J. (2006). Inhibitory and Bactericidal Activities of Daptomycin, Vancomycin, and Teicoplanin against Methicillin-Resistant *Staphylococcus aureus* Isolates Collected. *Antimicrobiology Chemotherapy* 58:1107—1117
- Food and Agriculture Organization of the United Nations (FAO)(2005). A Regional Survey of the Aquaculture Sector in West Asia. Aquaculture Development and Coordination Programme, Rome. ADCP/REP/89/35: 4-5
- Food and Agriculture Organization of the United Nations (FAO). (2010). A Regional Survey of the Aquaculture Sector in West Asia. Aquaculture Development and Coordination Programme, Rome. ADCP/REP/89/35: 4-5
- Food Safety and Inspection isolation method (FSIS) (2012), Microbiology Laboratory Guidebook. <http://www.fsis.usda.gov/wps/portal/ffis/topics/science/>
- Forsythe, S. J., and Hayes, P. R. (1998). Food hygiene, microbiology and HACCP (3rd ed.). Aspen Publishers
- Fukushima, H., and Seki, R. (2003). Ecology of *Vibrio vulnificus* and *Vibrio parahaemolyticus* in brackish environments of the Sada River in Shimane Prefecture, Japan. *FEMS Microbiology Ecology* 48: 221-229
- Gopal, S. Otta, S. k., Kumar, S., Karunasagar, I., Nishibuchi, M. (2005): The occurrence of *Vibrio* species in tropical shrimp culture environments; implications for food safety. *International Journal of Food Microbiology* 102: 151-159
- Greg, R. (2009). Controlling Contamination in Water Treatment Equipment. *CWS-VI Water Conditioning and Purification*, Vol (1) 667-668
- Guardabassi, L., Andreas, P., John, E., and Anders, D. (1998). Antibiotic Resistance in *Acinetobacter* spp. Isolated from Sewers Receiving Effluent from a Hospital and a Pharmaceutical Plant. *Applied and Environmental Microbiology* 64: 3499-3502

- Goutam, M. P. and Purohit, R. M. (1974). Antimicrobial activity of the essential oil of the leaves of *Murraya koenigii Spreng* (Indian Curry Leaf), *Indian Journal of pharmacology* 36 (1), 11
- Gwendelynn, B. T., Son, R., Nishibuchi, M., Raha, A. R., Suhaimi, N., Lesley, M., Jurin, W. G. (2003). Characterization of *Vibrio parahaemolyticus* isolated from coastal seawater in Peninsular Malaysia. *The Southeast Asian Journal of Tropical Medicine and Public Health* 36 (4): 940-945
- Hafidh, R., Ahmed, S., Law, S.W., Fatimah, A. B., Faridah, A., Fatemeh, J. and Zamberi S. (2011). Inhibition of Growth of Highly Resistant Bacterial and Fungal Pathogens by a Natural Product. *The Open Microbiology Journal* 5:96-106
- Hamendra, S. P. and Anand, K. (2007): Antidiabetic potential of *Citrus sinensis* and *Punicagranatum* peel extracts in alloxan treated male mice. *Bio Factors* 31: 17-24
- Han, F., Walker, R. D., Janes, M. E., Prinyawiwatukul, W. G. B. (2007). Antimicrobial susceptibilities of *Vibrio parahaemolyticus* and *Vibrio vulnificus* isolates from Louisiana Gulf and retail raw oysters. *Applied and Environmental Microbiology* 73 (21): 7096-7098
- Han, S. T. (1998). Medicinal Plants in the South Pacific, World Health Organization, (WHO), Regional Publications West Pacific Series, No. 19, Manila
- Hara-Kudo, Y., Nishina, T., Nakagawa, H., Konuma, H., Hasegawa, J., Kumagai, S. (2001). Improved method for detection of *Vibrio parahaemolyticus* in seafood. *Applied and Environmental Microbiology* 67 (12): 5819-5823
- Harvey, J., Keenan, K. P. and Gilmour, A. (2007). Assessing biofilm formation by *Listeria monocytogenes* strains. *Food Microbiology* 24: 380-392
- Hlady, W. G., and Klontz, K. C. (1996). The epidemiology of *Vibrio* infections in Florida (1981-1993). *Journal of Infectious Diseases* 173: 1176-1183
- Honda, T. and Iida, T. (1993). The pathogenicity of *Vibrio parahaemolyticus* and the role of the thermostable direct haemolysin and related haemolysin. *Review in Medical Microbiology* 4: 106-113
- Hood, S. K. and Zottola, E. A. (1995). Biofilms in food processing. *Food Control* 6: 9-18
- Hosseini, H., Cheraghali, A. M., Yalfani, R., and Razavilar, V. (2004). Incidence of *Vibrio* spp. in shrimp caught off the South Coast of Iran. *Food Control* 15: 187-190

- Hwang, J. K., Shim, J.S and Chung, J.Y. (2004). Anticariogenic activity of some tropical medicinal plants against *Streptococcus mutans*. *Fitoterapia* 75:596–598
- International Commission on Microbiological Specification for Food (ICMSF). (1995). Roberts TA, Baird-Parker A C and Tompkin RB (eds.). Microorganisms in foods 5. Blackie Academic and Professional, London
- Ito, C., Itoigawa, M., Miyamoto, Y., Onoda, S., Rao, K.S., Mukainaka, T., Tokuda, H., Nishino, H. and Furukawa, H. (2003). Polyprenylated benzophenones from *Garcinia assugu* and their potential cancer chemopreventive activities. *Journal of Natural Products* 66(2): 206–209
- Jay, J. M. (2000). Modern Food Microbiology. 6<sup>th</sup> ed., APAC Publisher, Singapore: 425-568
- Jiang, X. and Chai, T.J. (1996). Survival of *Vibrio parahaemolyticus* at Low Temperatures under Starvation Conditions and Subsequent Resuscitation of Viable, Nonculturable Cells. *Applied Environment Microbiology* 62(4):1300
- Jin, Y., Samaranayake, L. P., Samaranayake, Y. and Yip, H. K. (2004). Biofilm formation of *Candida albicans* is variably affected by saliva and dietary sugars. *Archives Oral Biology* 49: 789–798
- Joshi, D. D. (2012). Herbal Drugs and Fingerprints: Evidence Based Herbal Drugs. Springer India
- Kaletka, E.F. (2013). Disinfection in poultry medicine - aims and means. *Veterinary Journal* 48 (2) 112-115
- Kawasaki S, Horikoshi, N., Okada, Y. (2005). Multiplex PCR for simultaneous detection of *Salmonella spp.*, *Listeria monocytogenes*, and *Escherichia coli* O157:H7 in meat samples. *Journal Food Protection* 68 : 551–556
- Kaysner, C., DePaola, A.J. (2004): U.S. Food and Drug Administration: Bacteriological Analytical Manual: Methods for specific pathogens: Chapter 9: *Vibrio*. Accessed on 24 May 2007. Available at <http://www.fda.gov/Food/ScienceResearch/LaboratoryMethods/BacteriologicalAnalyticalManualBAM/ucm070830.htm>.
- Kim, Y. B., Okuda, J., Matsumoto, C., Takahashi, Nakabayashi, Y., Kumagai, K., (1999). Molecular Identification of *Vibrio parahaemolyticus* strains at the direct hemolysin (TDH) and TDH-related hemolysin of species level by PCR targeted to the *toxR* gene. *Journal of Clinical Microbiology* 37 (4): 1173-1177

- Koburger, J. A., Lazarus, C. R. (1974). Isolation of *Vibrio parahaemolyticus* from salt springs in Florida. *Applied Microbiology* 27: 435-436
- Kraa, E. (1995). Surveillance and epidemiology of foodborne illness in NSW, Australia. *Food Australia* 47: 418-423
- Krämer, M., Bongaerts, J., Bovenberg, R., Kremer, S., Müller, U.; Orf, S., Wubbolts, M., Raeven, L. (2003). Metabolic engineering for microbial production of shikimic acid. *Metabolic Engineering* 5 (4): 277-283
- Krumperman, P. H. (1983). Multiple antibiotic resistance indexing of *Escherichia coli* to identify high-risk sources of fecal contamination of foods. *Applied and Environmental Microbiology* 46: 165-170
- Krupesha, S., Shankar, K. M., Sathyanarayana, M. L., Sahoo, A. K., Patil, R., Narayanaswamy, HD. and Rao, S. (2010). Evaluation of immune response and resistance to diseases in tiger shrimp, *Penaeus monodon* fed with biofilm of *Vibrio alginolyticus*. *Fish & Shellfish Immunology* 29: 724-732
- Kudva, I. T., Jelacic, S., Tarr, P. I., Youderian, P., and Hovde, C. J. (1999). Biocontrol of *Escherichia coli* O157 with O157-specific bacteriophages. *Applied and Environmental Microbiology* 65, 3767-3773
- Laksamana, S. P., Srinivasa, R.T.C.H.H., Srinivasa, L., Prasad, A.K.C. (1997). *In vitro* antibacterial activity of culinary spices Aniseed, Star Anise and Cin.namon against bacterial pathogens of fish. *International Journal of Pharmacy and Pharmaceutical Sciences* Vol 4(3) : 221-228.
- Lapornik B, Prosek M, Wondra, A. G. (2005). Comparison of extracts prepared from plant by-products using different solvents and extraction time. *Journal of Food Engineering* 71: 214-222.
- Le, X. T., Munekage, Y. and Kato, S. (2005). Antibiotic resistance in bacteria from shrimp farming in mangrove areas. *Science of the Total Environment*, vol.349, pp.95-105.
- Lee, H. C., and Pogliano, K. (2007). *Vibrio parahaemolyticus*: Microbe Wiki. Accessed on 4 January 2010. Available at [http://microbewiki.kenyon.edu/index.php/Vibrio\\_parahaemolyticus](http://microbewiki.kenyon.edu/index.php/Vibrio_parahaemolyticus).
- Lee, J. (2009). Laboratory studies of growth conditions of *Vibrio parahaemolyticus* in Pacific Oyster (*Crassostrea gigas*) with international considerations to shellfish-associated illnesses in South Korea. Bachelor Thesis. Oregon State University.

- Lesley, M. B., Son, R., Bahaman, A. R., Raha, A. R., Suhaimi, N., Michael, W. C. V. L., Gwendelynn, B. T. and Nishibuchi, M. (2005). Detection of *Vibrio parahaemolyticus* in cockle (*Anadara granosa*) by PCR. *Federation of European Microbiological Societies (FEMS) Microbiology Letters* 252: 85-88
- Lelieveld, H. L, Mostert, M. A, Holah, J. and White, B. (2003). Hygiene in Food Processing, Cambridge, Woodhead Publishing Limited.
- Lindow, S.E. and Brandl, M.T. ( 2003). Microbiology of the phyllosphere. *Applied and Environmental Microbiology* 69: 1875-1883
- Lorian, V. (2005). Antibiotics in Laboratory Medicine, 5th ed., *Lippincott Williams and Wilkins, London*
- López-Malo, A., Alzamora, S.M., and Argáiz, A. (1997). Effect of natural vanillin on germination and radial growth of moulds in fruit-based agar systems. *Food Microbiology* 12: 213-219
- Magnus, P., Jaakkola, J.J., Skrondal, A., Alexander, J., Becher, G., Krogh, T., (1999). Water chlorination and birth defects. *Epidemiology* 10(5):513-517
- Mahmud, Z. H., Kassu, A., Mohammad, A., Yamato, M., Bhuiyan, N. A., Nair, G. B., and Ota, F. (2005). Isolation and molecular characterization of toxigenic *Vibrio parahaemolyticus* from the Kii Channel, Japan. *Microbiological Research*. Available online at [www.elsevier.de/micres](http://www.elsevier.de/micres)
- Mallavarapu, G. R., Ramesh, S., Syamasunday, K.V., Chandrasekhara, R.S. (1999). Composition of Indian curry leaf oil. *Journal Essential oil Research* 11: 176-178
- Mandrell, R. E. and Wachtel, M. R. (1999). Novel detection techniques for human pathogens that contaminate poultry. *Current Opinion Biotechnology, Review*. 10(3):273-278
- Mao, Z. J., Yu, L., You, Z.Q., Wei, Y.W. and Liu, Y. (2007). Cloning, expression and immunogenicity analysis of five outer membrane proteins of *Vibrio parahaemolyticus* zj2003. *Fish Shellfish Immunology* 23, 567-575
- Maruti J. Dhanavade, Chidamber B. Jalkute, Jai S. Ghosh and Kailash D. Sonawane Study Antimicrobial Activity of Lemon (*Citrus lemon L.*) Peel Extract. *British Journal of Pharmacology and Toxicology* 2(3): 119-122
- Mata, A. I., Blanco, M. M., Dominguez, L., Fernandez-Garayzabal, J., Gibello, I. (2004). Development of a PCR assay for *Streptococcus iniae* based on the



lactate oxidase (lctO) gene with potential diagnostic value. *Veterinar Microbiology* 101(2): 109-116

Matsumoto, C., Okuda, J., Ishibashi, M., Iwanaga, M., Garg, P., Ramamurthy, T., Wong, H., Depaola, A., Kim, Y. B. (2000). Pandemic spread of an O3:K6 clone of *Vibrio parahaemolyticus* and emergence of related strains evidenced by arbitrarily primed and toxRS sequence analyses. *Journal Clinical Microbiology* 38: 578–585

McCarter, L. (1999). The multiple identities of *Vibrio parahaemolyticus*, *Journal of Microbiology Biotechnology* 1: 51-57

McCarter, L. L. (2006). Motility and Chemotaxis. In *The Biology of Vibrios*, ed. F.L. Thompson, B. Austin, and J. Swings, pp. 115-132. Washington D. C.: ASM Press

McCarthy S.A., (1992). Attachment of *Listeria monocytogenes* to chitin and resistance to biocides. *Food Technology* 46, 84–88.

McKeon, D.M., Calabrese, J.P. and Bissonette, G. K. (1995) Antibiotic resistant gram-negative bacteria in rural groundwater supplies. *Water Research* 29: 1902–1908

McLaughlin, J. B., DePaola, A., Bopp, C. A., Martinek, K. A., Napolilli, N., Allison, C, Murray, S. L., Thompson, E. C., Bird, M. M., Middaugh, J.P. (2005). Outbreak of *Vibrio parahaemolyticus* gastroenteritis associated with Alaskan oysters. *New England Journal of Medicine* 353 (14): 1463-1470

Mereghetti, L., Quentin, R., Marquet-Van Der Mee, N., Audurier, A. (2000). Low sensitivity of *Listeria monocytogenes* to quaternary ammonium compounds. *Applied Environmental Microbiology* 66 5083–5086

Merrill, A. Biel (2010). Biofilm Infections. Photodynamic Therapy of Bacterial and Fungal FEMS Microbiology Letters 232

Miller, K., O'Neill, A. J. and Chopra, I. (2002). Response of *Escherichia coli* hypermutators to selection pressure with antimicrobial agents from different classes. *Journal Antimicrobiology Chemotherapy* 49:925-934

Mills-Robertson, F., Addy, M. E., Mensah, P., and Crupper, S. S. (2002). Molecular characterization of antibiotic resistance in clinical *Salmonella typhi* isolated in Ghana. *FEMS microbiology Letters* 215: 249-253

Minakshi, De, Amit Krishna, De, Parimal Sen and Arun, B. (2002). Antimicrobial Properties of Star Anise (*Illicium verum* Hook f). *Phototherapy research* 16, 94–95

- Miwatani, T. and Takeda, Y. (1976). *Vibrio parahaemolyticus* - a causative bacterium of food poisoning . *National Library of Australia* 6 :110-136
- Mizunoe, Y., Wai, S. N., Ishikawa, T., Takade, A., and Yoshida, S. (2000). Resuscitation of viable but nonculturable cells of *Vibrio parahaemolyticus* induced at low temperature under starvation. *FEMS Microbiology Letters* 186: 115-120
- Molina-Aja, A., Garcia-Gasca, A., Abreu-Grobois, A., Bolan-Mejia, C., Roque, A., and Gomez-Gil, B. (2002). Plasmid profiling and antibiotic resistance of *Vibrio* strains isolated from cultured penaeid shrimp. *FEMS Microbiology Letters* 213: 7-12
- Monfort, P., Piclet, G. and Plusquellec, A. (2000). *Listeria innocua* and *Salmonella panama* in estuarine water and seawater: a comparative study. *Water Residue* 34: 983-989
- Nakamura, C.V., Ishida, K., Faccin, L.C. Filho, B.P. Cortez, D. A., Rozental, S., de Souza, W. and Ueda-Nakamura, T. (2004). In vitro activity of essential oil from *Ocimum gratissimum* against four *Candida* species. *Research Microbiology* 155: 579-586
- National Committee for Clinical Laboratory Standards (NCCLS)(2001). Performance standards for antimicrobial susceptibility testing: eleventh informational supplement. Wayne (PA) (NCCLS document no. M-100-S11)
- National Environment agency (2013). Sustaining our environment. National Environment Agency Annual Report 2012/13
- Negi, P. S., A.S. Chauhan, G.A. Sadia, Y.S. Rohinishree and R.S. Ramteke (2005). Antioxidant and antibacterial activities of various seabuckthorn (*Hippophae rhamnoides* L.) seed extracts. *Food Chemistry* 92: 119-124
- Nishibuchi, M., Kaper, J. B. (1995). Thermostable direct haemolysin gene of *V. parahaemolyticus*: a virulence gene acquired by a marine bacterium. *Infection and Immunity* 63: 2093-2099
- Noube, N. S., Afolayan, A. J., and Okoh, A. I. (2008). Assessment techniques of antimicrobial properties of natural compounds of plant origin: current methods and future trends. *African Journal of Biotechnology* 7 (12): 1797-1806
- Ngwoke, K. G., Odimegwu, D. C., Esimone, C. O. (2011). Antimicrobial natural products. In: Mendez-Vilas A, editor. Science against microbial pathogens:

communicating current research and technology advances. Badajoz, Spain: FORMATEX pp. 1011.

Odugbemi, T. O. (2006). Outlines and pictures of medicinal plants from Nigeria, University of Lagos Press, Lagos, Nigeria, pp. 91

Oliver, J. D. and Kaper, J. B. (1997). *Vibrio* species. In: Doyle, M, Beuchat, L, R, Montville, T.J., (Eds), *Food Microbiology: Fundamental and Frontiers*, ASM Press, Washington D. C., pp. 228-264

Oliver, J. D., Hite, F., McDougald, D., Barrett, T., Glover, L. A., Prosser, J. I. (1995). Entry into and resuscitation from, the viable nonculturable state by *Vibrio vulnificus* in an estuarine environment. *Applied and Environmental Microbiology* 61: 2624-2630.

Onayade, D. A., and Adebajo, A. C. (2000). Composition of the leaf volatile oil of *Murrayakoenigii* growing in Nigeria. *Journal Herbs, Spices Medicine Plants* 7(4): 59-66

O'Toole, G. H. B., Kaplan, R., Kolter, H. (2000). Biofilm formation as microbial development. *Annula Review Microbiology* 54:49-79.

Pandey, R. R, Dubey, R. C, Saini, S. (2010). Phytochemical and antimicrobial studies on essential oils of some aromatic plants. *African Journal of Biotechnology* 9(28):4364-68

Pankey, G. A., Ashcraft, D. S. (2010). In vitro antibacterial activity of tigecycline against resistant Gram-negative *bacilli* and enterococci by time-kill assay. *Diagnostic Microbiology Infection Disease* 64: 300-304

Palaniswamy, U. R. (2007). Asian horticultural crops and human dietetics. *HortTechnology* 11:504-509

Pavithra, G. (2014). Effect of Spices on Bacteria -A Short Review. *Journal of Pharmacy Science* Vol. 6(8), 268-270

Perret, C., Tabin, R., Marcoz, J. P., Lor, J., Cheseaux, J. J., (2011). Apparent life-threatening event in infants: think about star anise intoxication! *Archives de Pédiatrie* 18(7): 750-75

Phillip, J. (1981). Curryleaf and its issues. *World crops*, 33:125-127

Piera Serra, G. (2003). Estudio de biofilms: Formación y consecuencia.

Prudent, D., F. Perineau, J.M. Bessiere, G.M. Michel and J.C. Baccou (2005). Analysis of the essential oil of wild oregano from Martinique (*Coleus*



*aromaticus* Benth.)—evaluation of its bacteriostatic and fungistatic properties. *Journal Essential Oil Research* 7: 165– 173

- Pruthi, J.S. (1976). Spices and condiments. New Delhi: National Book Trust, India
- Rajkowski, K. T. (2009). Biofilms in fish processing. In P. M. Fratamico, B. A. Annous, and N. W. Gunther (Eds.), Biofilms in the food and beverage industries . Woodhead Publishing pp. 499 - 516
- Raman, M, (2001). Malaysia: Each prawn produced represents a teardrop. *World Rainforest Movement Bulletin* 51: 1-2
- Rasmussen, T. B., Skindersoe, M. E., Bjarnsholt, T., Phipps, R. K., Christensen, K. B., Jensen, P. O., et al. (2005). Identity and effects of quorum-sensing inhibitors produced by *Penicillium* species. *Microbiology* 151, 1325–1340
- Reeves, M. W., Evins, G. M., Heiba, A. A., Plikaytis, B. D., Farmer, J. J. (1989). Clonal nature of *Salmonellatyphi* and its genetic relatedness to other *Salmonellae* as shown by multilocus enzyme electrophoresis, and proposal of *Salmonella bongori*. *Journal Clinical Microbiology* 27: 313-320
- Reddy, V., Urooj, A. and Kumar, A. (2005). Evaluation of antioxidant activity of some plant extracts and their application in biscuits. *Food Chemical* 90: 317-321
- Reisch, J., Goj, O., Wickramasinghe, A. and Herath, H.M.T.B. (1992). Carbazole alkaloids from seeds of *Murrayakoenigii*. *Phytochemistry* 31: 2877-2879
- Reichelt, J. L., Baumann, P., Baumann, L. (1976). Study of genetic relationships among marine species of the genera *Beneckea* and *Photobacterium* by means of in vitro DNA/DNA hybridization. *Archives of Microbiology* 110: 101-120
- Ronner, A. B., and Wong, A. C. (1993). Biofilm development and sanitizer inactivation of *Listeria monocytogenes* and *Salmonella typhimurium* on stainless steel and Buna-n rubber.. *Journal Food Protection* 56:750-758
- Roth, L.S. (2006). Mosby's Handbook of Herbs and Natural Supplements. 3rd Ed. Mosby, StLouis
- Rukayadi, Y., Lee, K., Han, S., Yong, D., Hwang, J. K. (2009). In Vitro Activities of Panduratin A against Clinical *Staphylococcus* Strains. *Antimicrobial Agents Chemotherapy* 53: 4529—4532
- Rukayadi, Y., Han, S., Yong, D. and Hwang, J.K. (2010 ). In Vitro Antibacterial Activity of Panduratin A against Enterococci Clinical Isolates. *Biology Pharmacy Bulletin* 3(9): 1489-1493

- Sachse, K. and Frey, J. (2003). PCR Detection of Microbial Pathogens. Humana Press, Totowa, New Jersey 216: 3-80
- Sakazaki, R, Tamura, K., Kato, T. O., Yamai, S., Hobo, K. (1968). Studies on the enteropathogenic facultatively halophilic bacteria *Vibrio parahaemolyticus*: III. Enteropathogenicity. *Japanese. Journal of Medical Science and Biology* 21, 325–331
- Saumendu, D.R., Ratnali, B., Jashabir, R. (2012). Pharmacognostic, phytochemical, physicochemical property and antimicrobial activity studies of lemon peel oil. *Journal National Production Plant Resources* 2 (3):431-435
- Sandoe, J. A. T., Wysome, A. P., West, J., Heritage, M. H., Wilcox (2006). Measurement of ampicillin, vancomycin, linezolid and gentamicin activity against enterococcal biofilms. *Antimicrobial Chemotherapy* 57, 767—770
- Sarkar, B. L., Nair, G. B., Banerjee, A. K., Pal, S. C. (1985). Incidence and level of *Vibrio parahaemolyticus* in freshwater environs and in association with freshwater fishes in Calcutta. *Applied and Environmental Microbiology* 46: 132-136
- Scates, P., Moran, L., Madden, R. H. (2003). Effect of incubation temperature on isolation of *Campylobacter jejuni* genotypes from foodstuffs enriched in Preston broth. *Applied and Environmental Microbiology* 69: 4658-4661
- Schwach, T.S. and Zottola, E.A. (1982). Use of scanning electron microscopy to demonstrate microbial attachment to beef and beef contact surfaces. *Journal Food Science* 47: 1401–1405
- Settanni, L. and Corsetti, A. (2007). Application of bacteriocins in vegetable food biopreservation. *International Journal of Food Microbiology* 121; 123–138
- Sharma, M., and Anand, S. K. (2002). Characterization of constitutive microflora of biofilms in dairy processing lines. *Food Microbiology* 19, 627–636
- Shelef, L.A. (1980). Antimicrobial effects of spices. *Journal of Food Safety* 6: 29–44
- Shinoda, S., Honda, T., Takeda, Y., Miwatani, T. (1974). Antigenic difference between polar monotrichous and peritrichous flagella of *Vibrio parahaemolyticus*. *Journal of Bacteriology* 120: 923-928
- Shinoda, S., Kariyama, R., Ogawa, M., Takeda, Y., Miwatani, T. (1976). Flagellar antigens of various species of genus *Vibrio* and related genera. *International Journal of Systematic Bacteriology* 26: 97-101

- Simoes, M., Simoes, L.C., Viera, M.J. (2010). A review of current and emergent biofilm control strategies. *LWT-Food Sciences and Technology* 43(4): 573-583
- Smack, D. P., Harrington, A.C., Dunn, C., Howard, R. S., Szkutnik, A. J., Krivda, S. J. Caldwell, J. B. and James, W. D. (1996). Infection and allergy incidence in ambulatory surgery patients using white petrolatum vs bacitracin ointment. A randomized controlled trial *JAMA : the journal of the American Medical Association* 276 (12): 972-7. PMID 8805732
- Sonia, S., Aparna, D.B., Gupta, G., Saksham, G. (2013). Antimicrobial activity of spices against isolated foodborne pathogens. *International Journal of Pharmacy and Pharmaceutical Sciences* 5 (1): 118-235
- Sorum, H., L., Abee, T. and Lund, T. M. (2002). Antibiotic resistance in food-related bacteria – a result of interfering with the global web of bacterial genetics. *International Journal of Food Microbiology* 78: 43-56
- Sommer, P., Martin-Rouas, C. and Mettler, E. (1999). Influence of the adherent population level on biofilm population, structure and resistance to chlorination. *Food Microbiology* 16: 503-515
- Spann, C. T, Taylor, S. C., Weinberg, J. M. (2004). "Topical antimicrobial agents in dermatology". *Disease-a-month* : DM 50 (7): 407-21
- Sujeewa, A. K. W., Norrakiah, A. S. and Laina, M. (2009). Prevalence of toxic genes of *Vibrio parahaemolyticus* in shrimps (*Penaeus monodon*) and culture environment. *International Food Research Journal* 16: 89-95
- Srivastava, N. (2005). Antibacterial Activity of Crude Ethanolic Extracts and Essential Oils of Spices Against *Salmonella* and other *Enterobacteria*. *KMITL Science Technology Journal* 5 (3) :6-12
- Staronline (2013). Accessed on 12 February 2014. <http://www.frim.gov.my/frim-conducts-course-on-plant-diseases-pests/?wppa-album=87&wppa-cover=0&wppa-occur=1&wppa-photo=311>
- Sutherland, I. W. (2001). The biofilm matrix – an immobilized but dynamic microbial environment. *Trends in Microbiology* 9, 222-227
- Su, Y. C. and Liu, C. A. (2007). *Vibrio parahaemolyticus*: a concern of seafood safety, *Food Microbiology* 24: 549-558
- Sybiya, V., Packiavathy, I. A., Agilandeswari, P., Syed Musthafa, K., Pandian, S. K., Ravi, A.V.(2012). Antibiofilm and quorum sensing inhibitory potential of

*Cuminumcyminum* and its secondary metabolite methyl eugenol against Gram negative bacterial pathogens. *Food Research International* 45: 85-92

- Szczuka, E. and Kaznowski, A. (2004). Typing of clinical and environmental *Aeromonas* sp. Strains by random amplified polymorphic DNA PCR, repetitive extragenic palindromic PCR, and enterobacterial repetitive intergenic consensus sequence PCR. *Journal of Clinical Microbiology* 42:220-228
- Tada, J., Ohashi, T., Nishimura, N., Shirasaki, Y., Ozaki, H., Fukushima, S., Takano, J., Nishibuchi, M. and Takeda, Y. (1992). Detection of thermostable direct hemolysin gene (trh) and thermostable direct hemolysin-related hemolysin gene (trh) of *Vibrio parahaemolyticus* by polymerase chain reaction. *Molecular and Cellular Probes* 6: 477-487
- Tendencia, E. A., and Pena, L. D. (2001). Antibiotic resistance of bacteria from shrimp ponds. *Aquaculture* 195: 193-204
- Thiem and Goslinska, (2001). Antimicrobial activity of *Rubuscha maemorus* leaves. *Fitoterapia* 75:93-95
- Thompson, F. L., Iida, T., Swings, J. (2004). Biodiversity of vibrios. *Microbiology and Molecular Biology Reviews* 38: 403-431
- Tomatake, H., Koga, T., Yamato, M., Kassu, A. and Ota, F. (2006). Antibacterial activity of citrus fruit juices against *Vibrio* species. *Journal Nutrition Science Vitaminology* 52(2):17-160
- Tu, T. H. Silvestre, F. Bernard, A. Douny, C. Phuong, T.N. Tao, T.C. Maghuin-Rogiste, G. and Kestemont, P. (2008). Oxidative stress response of black tiger shrimp (*Penaeus monodon*) to enrofloxacin and to culture system. *Aquaculture* 285, pp.244-248
- Tunung, R., Chai, L. C., Usha, M. R., Lee, H. Y., Fatimah, A. B., Farinazleen, A. B., Son, R. (2007). Characterization of *Salmonella enterica* isolated from street food and clinical samples in Malaysia. *ASEAN Food Journal* 14: 161-173
- Tsukamoto, K. K., Oyaizu, H., Nanba, K., Simidu, U. (1993). Phylogenetic relationship of marine bacteria, mainly members of the family Vibrionaceae, determined on the basis of 16S rRNA sequences. *International Journal System Bacteriology* 43:8-19
- Urakawa, H., Rivera, I. N. G. (2006). Aquatic Environment. In *The Biology of Vibrios*, ed. F.L. Thompson, B. Austin, and J. Swings, pp. 175-189. Washington D. C.: ASM Press

- United States Department of Food and Agriculture (USDA). (2005). Most Probable Number Procedure and Table. In Laboratory Guidebook. United States Department of Agriculture, Food Safety and inspection Service, Office of Public Health Science. pp. 1-8
- Valle, J., Re, D. S., Henry, N., Fontaine, T., Balestrino, D., Latour-Lambert, P. (2006). Broad-spectrum biofilm inhibition by a secreted bacterial polysaccharide. *Proceedings of the National Academy of Sciences USA*, 103, 12558–12563
- Vandamme, P., Pot, B., Gillis, M., de Vos, P., Kersters, K., Swings, J. (1996). Polyphasic taxonomy, a consensus approach to bacterial systematic. *Microbiological Reviews* 60: 407-438
- Wan, J., Wilcock, A. and Coventry, M.J. (1998) The effect of essential oils of basil on the growth of *Aeromonas hydrophila* and *Pseudomonas fluorescens*. *Journal of Applied Microbiology* 84, 152–158
- Wang, R., Huang, J., Zhang, W., Lin, G., Lian, J., Jiang, L., Lin, H., Wang, S., Wang, S. (1997). Detection and identification of *Vibrio parahaemolyticus* by multiplex PCR and DNA-DNA hybridization on a microarray. *Journal of Genetics and Genomic* 38(3):129-135
- Wekell, M. M., Manger, R., Kolburn, K., Adams, A. and Hill, W. (1994). Microbiological quality of Seafoods: viruses, bacteria and parasites. In Shahidi F., & Botta R. J (Eds). *Seafoods Chemistry, Processing Technology and Quality* pp : 220-232
- Welsh, J. and McClelland, M. (1990). Fingerprinting genome using PCR with arbitrary primers. *Nucleic Acid Research* 18: 7213-7218
- Williams, J. G. K., Kubelik, A. R., Livak, K. J., Rafalski, J. A., and Tingey, S. V. (1990). DNA polymorphisms applied by arbitrary primers useful as genetic markers. *Nucleic Acid Research* 18: 6531-6535
- Yearsley, G.K., Last, P.R. and Ward R.D. (eds) (1999). Australian seafood handbook: an identification guide to domestic species, CSIRO, Hobart. ISBN 0643 06194 0
- Yoshitsugu Nakaguchi (2013). MH Contamination by *Vibrio parahaemolyticus* and Its Virulent Strains in Seafood Marketed in Thailand, Vietnam, Malaysia, and Indonesia. *Tropical Medicine and Health* 41 No. 3, 2013, pp. 95-10
- Zhang, L., D. Pornpattananangkul, C. M. and Huang. C. M. (2010). Development of nanoparticles for antimicrobial drug delivery. *Current Medicinal Chemistry* 17(6): 585-594



Zink, D. L. (1997). The impact of consumer demands and trends on food processing. *Emerging Infectious Diseases*, 3(4), 467-469.  
<http://dx.doi.org/10.3201/eid0304.970408>

Zottola E.A. and Sasahara K.C. (1994): Microbial bio- films in the food-processing industry - should they be a concern. *International Journal of Food Microbiology*, 23: 125-148

Zulkifli, Y., Alitheen, N. B., Raha, A. R., Yeap, S. K., Marlina, Son, R. and Nishibuchi, M. (2009). Antibiotic resistance and plasmid profiling of *Vibrio parahaemolyticus* isolated from cockles in Padang, Indonesia. *International Journal of Food Research* 16: 53-58

Zulkifli, Y., Alitheen, N. B., Son, R., Yeap, S. K., Lesley, M. B. and Raha, A. R. (2009). Identification of *Vibrio parahaemolyticus* isolates by PCR targeted to the *toxR* gene and detection of virulence genes. *International Food Research Journal* 16: 289-296.