



UNIVERSITI PUTRA MALAYSIA

***PARTIAL PURIFICATION AND CHARACTERIZATION OF MOLYBDENUM
REDUCING ENZYME FROM *Escherichia coli* STRAIN K12***

MOHD HARIS BIN SULAIMAN

FBSB 2015 10



**PARTIAL PURIFICATION AND CHARACTERIZATION OF MOLYBDENUM
REDUCING ENZYME FROM *Escherichia coli* STRAIN K12**

By

MOHD HARIS BIN SULAIMAN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfilment of the Requirements for the Degree of Master of Sciences**

March 2015

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

PARTIAL PURIFICATION AND CHARACTERIZATION OF MOLYBDENUM REDUCING ENZYME FROM *Escherichia coli* STRAIN K12

By

MOHD HARIS BIN SULAIMAN

March 2015

Chairman: Associate Prof. Mohd Yunus Abdul Shukor, PhD
Faculty: Biotechnology and Biomolecular Sciences

Molybdenum is becoming a threat to environment due to its toxic effect in high concentration due to various industrial use. Therefore improper management of this heavy metal waste will cause it to remain in the ecosystem. To date, the best way to treat heavy metals is via bioremediation. *Escherichia coli* (*E. coli*) have been reported to have the capability to reduce the heavy metal molybdenum (molybdate) to molybdenum blue. Thus, this study was conducted to determine the optimum environmental and nutrient conditions of *Escherichia coli* strain K12; to partially purify molybdenum reducing enzyme; and to characterize the molybdenum reducing enzyme. *E. coli* are Gram-negative bacteria that belong to the γ -proteobacteria. They exist in a straight rod shaped cells and about 2 μm long and 0.5 μm wide, which can grow and divide rapidly by binary fission. In this experiment, *E. coli* was used to reduce molybdate Mo^{6+} forming a Mo-blue with the aid of Molybdenum reductase (Mo-reducing enzyme) which give non-toxic effect to the environment. Bacteria were cultured in a low phosphate media, pH 7.5 at temperature 35°C and incubated for 2 days. The optimization studies were carried out to optimize the production of molybdenum blue. The combination of 1% (w/v) glucose, 0.4% (w/v) ammonium chloride, 0.2% (w/v) yeast extract, and in ratio of 5mM phosphate and 80mM molybdate at pH 7.5 gave the optimum production of Molybdenum blue. Based on the maximum absorption peak at 865nm, this wavelength was used for the measurement of molybdenum blue produced in subsequent experiments. The effect of heavy metals on molybdenum blue production were studied. Thirteen metal ions and heavy metals were screened on *E. coli*. Mercury (Hg), Argentum (Ag), Copper (Cu) and Chromium (Cr) totally inhibited molybdate reductase enzyme while Zink (Zn), Nickel (Ni), Cobalt (Co), Arsenic (As), Lead (Pb) Aluminium (Al), Cadmium (Cd), and Magnesium (Mg) decreased the molybdenum reduction activity. Partial purification and characterization were conducted on molybdenum reducing enzyme with anion exchange chromatography using GE-Healthcare Mono-QTM column and gel filtration chromatography using Agilent ZorbaxTM (GF-250) column. Two bands were visualized on the gel filtration fraction at 95.64 and 84.42 kDa using SDS polyacrylamide-gel electrophoresis (SDS-PAGE). In enzyme kinetic studies, characterization of enzyme and stability of enzyme is being studied. NADH serves as the substrate for electron donor and 12-Molybdophosphate act as the substrate. The K_m and V_{max} for NADH were 2.156 mM and 15.015 units/ mg enzyme respectively. While the values for 12-MP were 3.549 mM and 54.348 units/ mg enzyme respectively. The characterization of Mo-reducing

enzyme studies were carried out at optimum pH of 6.0 using phosphate buffer at 35 °C. For enzyme temperature stability, enzyme are stable at range of 5°C - 40 °C for a period of 24 hours. Based on the result obtained, *Escherichia coli* strain K12 was proven to be effective for reduction of molybdate forming molybdenum blue.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

PENULENAN SEPARA DAN PENCIRIAN ENZIM PENURUNAN MOLIBDENUM OLEH BAKTERIA *Escherichia coli* STRAIN K12

Oleh

MOHD HARIS BIN SULAIMAN

Mac 2015

Pengerusi: Profesor Madya Mohd Yunus Abdul Shukor, PhD
Fakulti : Bioteknologi dan Sains Biomolekul

Molibdenum di dalam kepekatan yang tinggi telah menjadi satu ancaman kepada persekitaran disebabkan oleh kesan toksiknya serta penggunaan meluas di dalam industri. Oleh itu, salah pengurusan bagi sisa logam berat akan menyebabkannya kekal di dalam ekosistem persekitaran. Setakat ini cara yang terbaik bagi merawat sisa logam berat adalah melalui bioremediasi. *Escherichia coli* (*E.coli*) telah dilaporkan mempunyai keupayaan untuk menurunkan logam berat molibdenum (molibdat) kepada molibdenum biru. *E.coli* merupakan bakteria gram-negatif dan tergolong dalam kumpulan γ -proteobacteria yang wujud dalam sel-sel berbentuk rod kira-kira 2 μm panjang dan 0.5 μm lebar yang mempunyai pertumbuhan yang cepat dan menjalani belahan dedua. Dalam eksperimen ini, *E.coli* digunakan bagi mengkaji penurunan molibdat Mo^{6+} membentuk molibdenum biru melalui enzim penurun Molibdenum. Pengoptimuman telah dikaji untuk menentukan kadar optimum penghasilan molibdenum biru. Bakteria dikultur dalam media fosfat berkepekatan rendah pada pH 7.5 dan suhu 35 °C selama 2 hari. Kombinasi 1% (w/v) kepekatan glukosa, 0.4% (w/v) ammonium klorida, 0.2% (w/v) kepekatan yis dan nisbah 5mM fosfat dan 80mM molibdat pada pH 7.5 memberikan hasil optimum molibdenum biru. Molibdenum biru diukur berpanduan gelombang pada 865nm dalam keseluruhan eksperimen. Tiga belas ion logam dan logam berat telah diuji kepada *E.coli*. Merkuri (Hg), Argentum (Ag), Kuprum (Cu), dan Kromium (Cr) merencat sepenuhnya enzim penurun molibdat manakala Zink (Zn), Nikel (Ni), Kobalt (Co), Arsenik (As), Plumbum (Pb), Aluminium (Al), Kadmium (Cd), dan Magnesium (Mg) merendahkan aktiviti enzim penurun molibdenum. Penulenan separa dan pencirian telah dijalankan kepada enzim penurun molibdenum dengan kaedah kromatografi penukaran anion menggunakan kolum GE-Healthcare Mono QTM dan kromatografi penurasan gel fraksi menggunakan kolum Agilent ZorbaxTM (GF-250). Dua jalur dapat dilihat pada penurasan gel pada fraksi 95.64 dan 84.42 kDa menggunakan SDS elektroforesis-gel poliakrilamida. Dalam kajian kinetik enzim, NADH berfungsi sebagai substrat untuk menderma elektron dan 12-MP bertindak sebagai substrat untuk menerima elektron dimana K_m dan V_{max} untuk NADH adalah 2.156mM dan 15.015 mM unit / mg enzim masing-masing. Bagi 12-MP pula nilai K_m dan V_{max} adalah 3.549 mM dan 54.348 mM unit / mg enzim masing-masing. Pencirian bagi enzim penurun molibdenum telah dijalankan dan pH optimum adalah pada pH 6.0 didalam larutan penimbal fosfat pada suhu 35 ° C. Bagi ujian kestabilan suhu, enzim

adalah stabil pada julat suhu 5 °C - 40 °C selama tempoh 24 jam. Berdasarkan keputusan yang diperolehi, *Eschericia coli* strain K12 telah terbukti berkesan untuk penurunan molibdat membentuk molybdenum biru.



ACKNOWLEDGEMENTS

بِسْمِ اللَّهِ الرَّحْمَنِ الرَّحِيمِ

In the name of Allah, the most Gracious, and Most Merciful.

I would like to express my special appreciation and thanks to my supervisor Assoc. Prof. Dr. Mohd Yunus Abdul Shukor and not to forget my Co-supervisor, Prof. Mohd Ariff Syed, both of you have been a tremendous mentor for me and not to forget all lecturers from Biochemistry Department. Thank you for the support and guide for all this while until the completion of this thesis.

My appreciation also goes to all the people that help me on each milestone to complete this research, Universiti Putra Malaysia, colleagues, lab partner; Mohd Ezuan Khayat, Mohd Afif Aziz, Mohd Badrin, Baskaran, Siti Aqlima, and all others lab mates for the tears and joy during the lab session. They give me emotional and spiritual strength to success in the Msc. Research.

My special thanks and gratitude also goes to all my parents and wife for the support and long lasting motivation for my success and the trust they put during my up and down moments. I always believe that Allah will always be with the patience and in every difficulty, lies the opportunity.

Thank you.

Mohd Haris Bin Sulaiman

I certify that a Thesis Examination Committee has met on 16 March 2015 to conduct the final examination of Mohd Haris Bin Sulaiman on his thesis entitled “Partial Purification and Characterization of Molybdenum-reducing Enzyme from *Escherichia coli* Strain K12” in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The committee recommends that the student be awarded the Masters of Science.

Members of the Examination Committee are as follows:

Syahida Binti Ahmad, PhD

Lecturer

Faculty of Biotechnology & Biomolecular Sciences

Universiti Putra Malaysia

(Chairman)

Son Radu, PhD

Professor

Faculty of Food Science and Technology

Universiti Putra Malaysia

(Internal examiner)

Phang Lai Yee, PhD

Lecturer

Faculty of Biotechnology & Biomolecular Sciences

Universiti Putra Malaysia

(Internal examiner)

Mohd Azmuddin Abdullah, PhD

Associate Professor

Universiti Teknologi PETRONAS

Malaysia

(External Examiner)

ZULKARNAIN ZAINAL, PhD

Professor and Deputy Dean

School of Graduate Studies

Universiti Putra Malaysia

Date : 5 November 2015

This thesis was submitted to the Senate of Universiti Putra Malaysia has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Mohd Yunus Bin Abd Shukor, PhD

Associates Professor

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Chairman)

Mohd Arif Bin Syed, PhD

Professor

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date :

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of the thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No. : Mohd Haris Bin Sulaiman (GS25162)

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____
Name of
Chairman of
of Supervisory
Committee: Mohd Yunus Bin Abd Shukor

Signature: _____
Name of
Member of
Supervisory
Committee: Mohd Arif Bin Syed

TABLE OF CONTENTS

| | Page |
|--|----------|
| ABSTRACT | i |
| ABSTRAK | iii |
| ACKNOWLEDGEMENTS | v |
| APPROVAL | vi |
| DECLARATION | viii |
| LIST OF TABLES | xv |
| LIST OF FIGURES | xvi |
| LIST OF ABBREVIATIONS | xix |
| CHAPTER | |
| 1 INTRODUCTION | 1 |
| 2 LITERATURE REVIEW | 3 |
| 2.1 Molybdenum. | 3 |
| 2.1.1 History on Molybdenum | 3 |
| 2.1.2 The chemistry of Molybdenum | 3 |
| 2.1.3 Molybdenum-containing enzyme in biological systems | 4 |
| 2.1.3.1 Mechanism action of molybdenum-containing enzyme in biological systems | 5 |
| 2.1.4 Molybdenum in industry | 6 |
| 2.1.4.1 Molybdenum source | 6 |
| 2.1.4.2 Molybdenum applications | 7 |
| 2.1.5 The molybdate ion | 7 |
| 2.1.6 Molybdenum in biochemistry | 7 |
| 2.1.6.1 Molybdenum in air, soil and water | 8 |
| 2.1.6.2 Molybdenum in plant and soil microbial symbiotic interaction. | 9 |
| 2.1.7 Molybdenum toxicity | 9 |
| 2.1.7.1 Molybdenum toxicokinetic | 9 |
| 2.1.7.2 Toxicity in animals | 10 |
| 2.1.7.3 Toxicity in human | 11 |
| 2.1.8 Molybdenum pollution | 12 |
| 2.1.9 Comparison of several bacteria in molybdenum reduction | 12 |
| 2.1.10 Enzymatic and microbial action on Molybdenum | 13 |
| 2.1.10.1 Mo-blue | 13 |
| 2.1.10.2 Mo-reducing enzyme | 14 |
| 2.1.10.3 Mo-reducing enzyme purification | 16 |
| 2.1.10.4 Molybdenum-blue quantification from molybdate reduction | 16 |
| 2.2 Bioremediation | 17 |
| 2.2.1 Advantages of bioremediation | 18 |
| 2.2.2 Bioremediation of heavy metals | 18 |

| | | |
|-----------|--|-----------|
| 3 | MATERIALS AND METHODS | 20 |
| 3.1 | Equipments, Chemicals, Buffer and Chemical Solutions | 20 |
| 3.1.2 | Microorganism and culture medium | 20 |
| 3.2 | Methodology of Experiment | 20 |
| 3.2.1 | Scanning Spectra of <i>E.coli</i> strain-K12's | |
| | Molybdenum Blue | 20 |
| 3.2.2 | Growth optimization of <i>E.coli</i> | 20 |
| 3.2.3 | Bacterial growth optimization over time | 21 |
| 3.2.4 | Optimization of Carbon Sources | 21 |
| 3.2.5 | Optimization of Nitrogen Sources | 21 |
| 3.2.6 | Optimization of pH | 22 |
| 3.2.7 | Optimization of Molybdate (Mo O_4^{2-}) and Phosphate | 22 |
| 3.2.8 | Effect of Phosphate and Molybdate ratio on Molybdate reduction | 23 |
| 3.2.9 | Optimization of Temperature | 24 |
| 3.2.10 | Effect of Heavy Metals and Metal Ions on Molybdate reduction by <i>Escherichia coli</i> strain K12 | 25 |
| 3.2.11 | Scale Up of <i>E.coli</i> strain K12 Growth and Enzyme Purification | 25 |
| 3.2.11.1 | Growth of Bacteria | 25 |
| 3.2.11.2 | Small Scale Growth | 25 |
| 3.2.11.3 | Large Scale Growth | 25 |
| 3.2.11.4 | High Speed Centrifugation | 26 |
| 3.2.11.5 | Sonication and Ultracentrifugation | 26 |
| 3.2.11.6 | Partial Purification by pre-packed Mono-Q TM Strong Anion Exchanger | 26 |
| 3.2.11.7 | Column Chromatography Preparation for Mono-Q TM Strong Anion Exchanger | 26 |
| 3.2.11.8 | Sample Preparation and Application | 27 |
| 3.2.11.9 | Partial Purification using Agilent Zorbax TM (G-250) Gel Filtration | 27 |
| 3.2.11.10 | Column Chromatography Preparation for Agilent Zorbax TM (G-250) Gel Filtration | 27 |
| 3.2.11.11 | Sample Preparation and Application | 28 |
| 3.2.11.12 | SDS Polyacrylamide Gel Electrophoresis of Protein | 28 |
| 3.2.11.13 | SDS-Polyacrylamide Gels Preparation | 28 |
| 3.2.11.14 | Preparation of Samples and Running the Gel | 29 |
| 3.2.11.15 | Gel Staining | 30 |
| 3.2.12 | Enzymatic Studies on the Reduction of Molybdenum | 30 |
| 3.2.12.1 | Molybdenum-reducing Enzyme Assay | 30 |
| 3.2.12.2 | Protein Concentration Determination | 31 |
| 3.2.12.3 | Molybdenum-reducing Enzyme Kinetic Studies | 31 |
| 3.2.12.4 | K_m and V_{max} NADH as the Substrate Electron Donor | 31 |
| 3.2.12.5 | K_m and V_{max} LPPM as the Substrate Electron Donor | 32 |

| | | |
|----------|--|-----------|
| 3.2.12.6 | Effect of Temperature on Molybdenum-Reducing Enzyme Activity | 32 |
| 3.2.12.7 | Effect of pH on Molybdenum-Reducing Enzyme Activity | 32 |
| 3.2.12.8 | Molybdenum-reducing Enzyme Temperature Stability | 32 |
| 4 | RESULTS AND DISCUSSION | 34 |
| 4.1 | Optimization of Molybdate reduction by <i>Escherichia coli</i> Strain K-12 | 34 |
| 4.1.1 | Molybdenum Blue (Mo-Blue) Spectroscopic Scanning | 34 |
| 4.1.2 | Optimization of <i>E. coli</i> strain K-12 Molybdate reduction over Time | 35 |
| 4.1.3 | Optimization of Temperature | 37 |
| 4.1.4 | Optimization of Carbon Sources | 38 |
| 4.1.5 | Optimization of Glucose Concentration | 39 |
| 4.1.6 | Optimization of Nitrogen Sources | 40 |
| 4.1.7 | Optimization of Ammonium Sulphate Concentrations | 42 |
| 4.1.8 | Optimization of pH | 43 |
| 4.1.9 | Optimization of MoO_4^{2-} and Phosphate | 45 |
| 4.1.9.1 | Effect of Phosphate and Molybdate Ratio on Molybdate Reduction | 45 |
| 4.2 | The Effect of Heavy Metals and Metal Ions on Molybdate Reduction by <i>Escherichia coli</i> strain K12 | 48 |
| 4.3 | Partial Purification of Mo-Reducing Enzyme | 50 |
| 4.4 | Purification Analysis of Molybdenum Reducing Enzyme | 52 |
| 4.5 | SDS Polyacrylamide Gel Electrophoresis | 55 |
| 4.6 | Enzymatic Studies on Reduction of Molybdenum by <i>Escherichia coli</i> strain K12 | 56 |
| 4.6.1 | Kinetic Studies by Mo-Reducing Enzyme | 56 |
| 4.6.1.1 | Kinetic Studies Using NADH as the Substrate Electron Donor | 56 |
| 4.6.1.2 | Kinetic Studies Using 12-PM as the Substrate Electron Acceptor | 58 |
| 4.6.2 | Effect of Different Temperatures on Mo-reducing Enzyme Activity | 60 |
| 4.6.3 | Effect of pH on Mo-reducing Enzyme Activity | 61 |
| 4.7 | Determination of Mo-reducing Enzyme Temperature Stability | 62 |
| 5 | CONCLUSIONS | 66 |
| | REFERENCES | 67 |
| | APPENDICES | 74 |
| | BIODATA OF STUDENT | 79 |

LIST OF TABLES

| Tables | Page |
|--|------|
| 1 Techniques of <i>in situ</i> bioremediation | 17 |
| 2 Volumes of disodium phosphate solution for preparing different concentration of molybdate on molybdate reduction | 24 |
| 3 Solution for preparing 12% resolving gels for Tris-glycine SDS-polyacrylamide Gel electrophoresis | 29 |
| 4 Solution for preparing 5% stacking gels for Tris-glycine SDS-polyacrylamide Gel Electrophoresis | 29 |
| 5 Mo-reducing enzyme putification table | 54 |
| 6 Overall results of Mo-reducing bacteria (<i>Escherichia coli</i> strain K12) Mo-reducing bacteria optimization and enzymatic studies of Mo-reducing enzyme from <i>Escherichia coli</i> strain K12 | 65 |

LIST OF FIGURES

| Figure | Page |
|--|------|
| 1 Active site structure of the three families of mononuclear molybdenum containing enzymes (a). Xanthine Oxidase family (b) DMSO reductase family (c) Sulfite Oxidase family | 4 |
| 2 General mechanism for oxidative (a) and reductive reaction (b) catalysed by molybdenum containing enzyme (Brondino <i>et al.</i> , 2006) | 5 |
| 3 World molybdenum reserves (19,000,000 metric tonnes) as reported by U.S geological survey, mineral commodity summaries, January 2005 | 6 |
| 4 Structure of molybdenum blue which is formed by 12 tetrahedral MoO_4^{2-} and one phosphate (PO_4^{3-}) ion | 13 |
| 5 A schematic presentation of the mechanism of molybdenum reduction to Mo-blue by EC 48 (Ghani <i>et al.</i> , 1993) | 14 |
| 6 Newly suggested schematic presentation of the mechanism of molybdate reduction to Mo-blue by EC 48 (modified from Shukor <i>et al.</i> , 2000) | 15 |
| 7 Scanning spectrum of Molybdenum blue from <i>Escherichia coli</i> strain K12 after 24 hours incubation period | 35 |
| 8 Molybdate reduction curve of <i>E.coli</i> strain K-12 over period of time | 36 |
| 9 The effect of temperature on the molybdate reduction by <i>E.coli</i> strain K-12 | 38 |
| 10 The effect of carbon sources on the molybdate reduction by <i>E.coli</i> strain K-12 | 39 |
| 11 The effect of glucose concentration on the molybdate reduction by <i>E.coli</i> strain K-12 | 40 |
| 12 The effect of nitrogen source on the molybdate reduction by <i>E.coli</i> strain K-12 | 42 |

| | | |
|----|--|----|
| 13 | The effect of ammonium chloride concentration on the molybdate reduction by <i>E.coli</i> strain K-12 | 43 |
| 14 | Effect of pH on the molybdate reduction by <i>E.coli</i> Strain K-12 | 44 |
| 15 | The effect of molybdate concentration on the molybdate reduction by <i>E.coli</i> strain K-12 | 46 |
| 16 | The effect of phosphate concentration on the molybdate reduction by <i>Escherichia coli</i> strain K12 | 47 |
| 17 | The effect of phosphate and molybdate concentration on the molybdate reduction by <i>E.coli</i> strain K-12 | 48 |
| 18 | Effect of twelve heavy metal (1ppm) on molybdate reduction by <i>E.coli</i> strain K-12 | 49 |
| 19 | Relative percentage activity (%) of twelve heavy metal (1ppm) on molybdate reduction by <i>E.coli</i> strain K-12 | 50 |
| 20 | Elution profile of Mo-reducing enzyme using Mono-Q TM anion-exchanger | 51 |
| 21 | Elution profile of Mo-reducing enzyme using Agilent Zorbax TM (GF-250) gel filtration column | 52 |
| 22 | The electrophoresis analysis of the crude and partially purified molybdenum reducing enzyme | 56 |
| 23 | Michealis-Menten plot of amount of Molybdenum-blue formed (Units/mg enzyme/min) versus the electron donor substrate (NADH) | 57 |
| 24 | Lineweaver-Burke plot of reciprocal amount of molybdenum blue form (Units/mg enzyme) versus reciprocal electron donor substrates, NADH (mM) | 58 |
| 25 | Michealis-Menten plot of amount of Molybdenum-blue formed (Units/mg enzyme/min) versus the electron acceptor substrate (12-Phosphomolybdate) | 59 |

| | | |
|----|--|----|
| 26 | Lineweaver-Burke plot of reciprocal amount of molybdenum blue form (nmole/min/mg protein) versus reciprocal electron acceptor substrates, 12-Phosphomolybdate (mM) | 60 |
| 27 | The Effect of Temperature on Mo-reducing Enzyme Activity | 61 |
| 28 | Effects of different pH and buffers on Mo-reducing enzyme activity | 62 |
| 29 | Effect of prolonged pre-incubation temperatures on Mo-reducing enzyme | 64 |

LIST OF ABBREVIATIONS

| | |
|---|-----------------------------------|
| % | Percent |
| (NH ₄) ₂ SO ₄ | Ammonium sulphate |
| < | Less than |
| > | More than |
| °C | Degree celsius |
| μl | Microlitre |
| μM | Micromolar |
| 12-MP | Twelve-molybdophosphate |
| Ag | Argentum |
| As | Arsenic |
| ATP | Adenosine triphosphate |
| Cd | Cadmium |
| cm | Centimeter |
| Co | Cobalt |
| Cr | Chromium |
| Cu | Copper |
| DEAE | Diethylaminoethylamine |
| dH ₂ O | Distilled water |
| DNA | Deoxyribonucleic acid |
| DTT | Dithiothreitol |
| EDTA | Ethylene diamine tetraacetic acid |
| <i>et al</i> | And friends |
| Fe | Ferum |

| | |
|----------------------|--|
| Glc | Glucose |
| g | Gravity (relative centrifugal force) |
| HCl | Hydrochloric acid |
| Hg | Mecury |
| HPLC | High performance liquid chromatography |
| HPM | High phosphate media |
| hr | Hour |
| K | Kelvin |
| kb | Kilobase |
| kDa | Kilodalton |
| Kg | Kilogram |
| K _m | Michealis-Menten constant |
| L | Litre |
| LPM | Low phosphate media |
| m | Meter |
| M | Molar |
| mA | Miliampere |
| mAu | Mili Absorbance unit |
| mg | Miligram |
| MgSO ₄ | Magnesium Sulphate |
| min | Minutes |
| mM | Milimolar |
| Mo | Molybdenum |
| Mo-blue | Molybdenum blue |
| Mo-reducing bacteria | Molybdenum reducing bacteria |

| | |
|---|--|
| Mo-reducing enzyme | Molybdenum reducing enzyme |
| MT | Milestones |
| MW | Molecular weight |
| $\text{Na}_2\text{HPO}_4 \cdot 2\text{H}_2\text{O}$ | Disodium-hydrogen phosphate |
| $\text{Na}_2\text{MoO}_4 \cdot 2\text{H}_2\text{O}$ | Disodium molybdate |
| NaCl | Sodium chloride |
| NAD^+ | Nicotinamide adenine dinucleotide oxidized form |
| NADH | Nicotinamide adenine dinucleotide reduced form |
| Ni | Nickel |
| nm | Nanometer |
| OD | Optical Density |
| PAGE | Polyacrylamide gel electrophoresis |
| Pb | Plumbum |
| pH | $-\log$ concentration of H^+ ion (<i>Puissance hydrogene</i>) |
| PMSF | Phenylmethylsulfonylfluoride |
| PO_4^{3-} | Phosphate |
| RNA | Ribonucleic acid |
| rpm | Revolution per minute |
| SDS | Sodium dodecyl sulphate |
| Sn | Stannum |
| T50-7.5-buffer | 50 mM Tris-HCl at pH 7 |
| TEMED | N,N,N',N'-tetramethyl-ethylenediamine |
| UV | Ultraviolet |
| v/v | Volume/ volume |
| V_{max} | Maximum velocity |

w/v

Weight/ volume

XOD

Xanthine oxidase

Zn

Zink



CHAPTER 1

INTRODUCTION

Water pollution due to heavy metals is a very important issue as it reduces the viable water resources by creating a negative feedback loop involving increasing economic pressure and decreasing quality of supply. Water covers nearly 70% of our planet, yet the majority of it is salt water. We have very little reserve of fresh water yet great portion of our reserved water have been badly polluted or contaminated.

Heavy metal is one of the major concerns that contributed to pollution of our water ways and has become a growing threat to the environment and humanity. Heavy metals such as mercury, lead and arsenic are widely recognized as highly toxic and dangerous to organisms. Major sources of heavy metal pollution today come from the combustion of leaded gasoline, mining and processing, steel, iron, cement and fertilizer production, nuclear and other industrial effluents and sludge, dumping and land filling of industries wastes, biocide and preservatives including organometallic compounds.

Molybdenum has been widely used in industrial applications which cause vast contamination of molybdenum in the environment. Many reports on molybdenum pollution due to molybdenum mining activity such as at Tokyo Bay and Black Sea in 1991, Red Sea in 1996 and Tyrol in 2000 (Davis, 1991; Slifer 1996; Neuhauserer *et al.*, 2000). Exposure to higher concentration of molybdenum affected the reproduction and caused mortality in humans and animals.

Heavy metal is different from organic pollutants because it cannot be detoxified by degradation and remains in the ecosystem (Shukor *et al.*, 2000). The best way to remove heavy metals is via bioremediation. Bioremediation is a process which involves the transformation/detoxification of pollutants using microorganisms and plants. Bioremediation cleans up the environment effectively and cheaper than any other methods.

In the early 90's, Ghani and his team (Ghani *et al.*, 1993) isolated a heterotrophic bacterium, *Enterobacter cloacae* strain 48 (EC 48) from Chengkau, Malaysia which was able to reduce molybdate (molybdenum 6+) to molybdenum blue (molybdenum 5+) with NADH as an electron donor. This study brought the Molybdenum-blue research to University Putra Malaysia. There were many other studies which have indicated that molybdenum can be reduced to molybdenum blue using several bacteria, such as *Serratia Marcescens* strain DRY6 (Shukor *et al.*, 2008a), *Serratia sp.* strain Dr. Y8 (Shukor *et al.*, 2009a), *Pseudomonas sp* strain DRY2 (Shukor *et al.*, 2009b), *Enterobacter sp.* strain Dr. Y13 (Shukor *et al.*, 2009c), *Serratia marcescens* strain Dr. Y9 (Shukor *et al.*, 2009d), *Serratia sp.* strain DRY5 (Shukor *et al.*, 2009e) and *Acinetobacter calcoaceticus* strain Dr. Y12 (Shukor *et al.*, 2010).

The earliest work on the Mo-reducing bacterium was on *E. coli* (Capaldi and Proskauer, 1896) followed by the work of Campbell *et al.*, (1985).

In order to present a better comparison on the efficiency and characteristics of *E. coli* more specifically *E. coli* strain K12, characterisation of this bacterium on low phosphate media as used by all of the more recent isolates is warranted.

E.coli strain K12 was chosen in this experiment as it is easy to handle, complete gene sequence, has the ability to grow under both aerobic and anaerobic condition and the outcome for post-bioremediation are less harmful as compared to bacteria studied previously.

The importance of purifying and characterizing the molybdenum-reducing enzyme allows the elucidation of the mechanism of molybdenum-reducing activity. Purification of molybdenum-reducing enzyme could identify the protein take role in molybdenum-reducing activity and determine the protein structure thus contribute in protein genetic engineering process. Comparison can be made with other purified molybdenum reducing enzymes from different sources in terms of homologous sequences and also for the best enzyme characteristics such as broad substrates capacity for the source of reducing power, strong affinity for heteropolymolybdates and heat stability for the purpose of bioremediation. This information contributes in future molybdate reduction and bioremediation studies.

Thus the objectives of this study are:

- To determine the optimum environmental and nutrient conditions of *Escherichia coli* strain K12 on Low Phosphate Media (LPM),
- To partially purify Mo-reducing enzyme, and
- To characterize the Molybdenum reducing enzyme.

REFERENCES

- Abumrad, N. N., Schneider, A. J., Steel, D., and Rogers, L. S. 1981. Amino acid intolerance during prolonged total parenteral nutrition reversed by molybdate therapy. *American journal of clinical nutrition* **34**: 2551-2559.
- Acinas, S. G., Rodríguez-Valera, F. and Pedros-Alio, C. 1997. Spatial and temporal variation in marine bacterioplankton diversity as shown by RFLP fingerprinting of PCR amplified 16S rDNA. *FEMS Microbiology and Ecology* **24**: 7–40.
- Anke, M., Groppel, B., Krause, U., Arnhold, W., and Langer, M. 1991. Trace element intake (zinc, manganese, copper, molybdenum, iodine and nickel) of humans in Thuringia and Brandenburg of the Fed Rep of Germany. *Journal of trace elements and electrolytes in health and disease* **5(2)**: 69-74.
- Ariff, A.B., M. Rosfarizan, B. Ghani, T. Sugio and M.I.A. Karim 1997: Mo-reducing Enzyme in *Enterobacter cloacae* strain 48. *World Journal of Microbiology and Biotechnology* **13**, 643–647.
- Astwood, A. C. and Wais, A. C. 1998. Psychrotrophic bacteria isolated from a constantly warm tropical environment. *Current Microbiology* **36(3)**: 148-151.
- Barceloux, D.G. and Barceloux, D. 1999. Molybdenum, *Journal of Toxicology: Clinical Toxicology* **37(2)**: 231-237.
- Boopathy, R. 2000. "Factors Limiting Bioremediation Technologies." *Bioresource Technology* **74(1)**: 63–67.
- Booth, I.R. 1985. Regulation of Cytoplasmic pH in Bacteria. *Microbiology Reviews* **49(4)**: 356-378
- Bradford, M. 1976. A rapid and sensitive method for the quantification of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry* **72**: 248-252.
- Braithwaite, E. R. 1981: Molybdate ion. In: Thompson, R. (Ed.) *Specialty Inorganic Chemicals*. The Royal Society of Chemistry. London. pp 350-351.
- Brondino, C. D., Rivas, M. G., Romão, M. J., Moura, J. J., and Moura, I. 2006. Structural and electron paramagnetic resonance (EPR) studies of mononuclear molybdenum enzymes from sulfate-reducing bacteria. *Accounts of chemical research* **39 (10)**: 788-796.
- Burris, R. H and Roberts, G. P. 1993. Biological nitrogen fixation. *Annual Review of Nutrition* **13**:317–335.

- Campbell, M. A., Campbell, A. D. and Villaret, D. B. 1985. Molybdate reduction by *Escherichia coli* K-12 and its *Chl* mutants. *Proceedings of the National Academy of Science of the United States of America* **82**: 227-231.
- Chappell, W. R., Meglen, R. E. Moure-Eraso, R. *et al.*, 1979. Human health effects of molybdenum in drinking water. EDA-600A-79-006. US EDA, Cincinnati, Ohio.
- Clark, N. A., Teschke, K., Rideout, K., and Copes, R. 2007. Trace element levels in adults from the west coast of Canada and associations with age, gender, diet, activities and levels of other trace elements. *Chemosphere* **70**: 155-164.
- Clesceri, L.S., Greenberg, A.E. and Trussel, R.R. (eds.), 1989: Standard Methods for the Examination of Wastewater, 17th ed. American Public Health Association, Port City Press, Baltimore, Maryland, pp. (4-166)–(4-178).
- Cotton, F. A. and Wilkinson, G. 1980. Advanced inorganic chemistry: A comprehensive. Wiley-Interscience. New York. pp 453-455.
- Counotte, G. H. M., Prins, R. A. 1979. Calculation of K_m and V_{max} from substrate concentration versus time plot. *Applied and Environmental Microbiology* **38(4)**: 758-760.
- Davis, G. K. 1991. Molybdenum. In: Merian, E. (Ed). *Metals and their compounds in the environment, occurrence, analysis and biological relevance*. VCH Weinheim. New York. pp 1089–1100.
- Dawson, R. M. C, Elliott, D.C., Elliott, W.H. and Jones, K.M. 1969. Data for biochemical research. Clarendon Press. Oxford. pp. 484-485
- Deutscher, M. P. 1990. Setting up a laboratory. In: Deutscher, M. P. (ed). *Methods in enzymology: Guide to protein purification*. Volume 182. Academic Press, San Diego. pp19-24
- Dick, A. T. 1956. Molybdenum in animal nutrition. *Soil Science* **81**: 229-236.
- Donald, G. B. and Donald, B. 1999. Molybdenum. *Clinical Toxicology* **37(2)**: 231-237.
- Drancourt, M., Bollet, C., Carlouz, A., Martelin, R., Gayral, J. P., Raoult, D. 2000. 16S ribosomal DNA sequence analysis of a large collection of environmental and clinical unidentifiable bacterial isolates. *Journal of Clinical Microbiology* **38 (10)**: 3623-3630.
- Dworkin, M. and Falkow, S. 2006. The Prokaryotes: Proteobacteria: gamma subclass. 3rd ed. Springer, New York, United State.
- Elverum, -J, S.M, Shivik, J.A. and Clark, L. 2001. Importance of Bacterial Decomposition and Carrion Substrate To Foraging Brown Treesnakes. *Journal of Chemical Ecology* **27 (7)**: 1315-1331.

- Fairhall, L. T., Dunn, R. C., Sharpless, N. E. and Pritchard, E. A. 1945. The toxicity of molybdenum. United States Public Health Service, Public Health Bulletin. pp293.
- Felsenstein, J. 1985. Confidence limits on phylogenies, an approach using the bootstrap. *Evolution* **39**: 783-791.
- Friberg, L., Lener, J. 1986. Molybdenum. In: *Handbook on the Toxicology of Metals. Vol II*. Friberg, L., Nordberg, G. F., Vouk, V. B., eds. Elsevier Science Publishing Co., Amsterdam. pp 446-461.
- Furr, A. K., Lawrence, A. W., Tong, S. S. C., *et al.* 1976. Multielement and chlorinated hydrocarbon analysis of municipal sewage sludges of American cities. *Environmental Science and Technology* **10**:68-687.
- Ghani, B., Takai, M., Hisham, N. Z., Kishimoto, N., Ismail, M. I. A., Tano T., and Sugio. T. 1993. Isolation and characterization of a Mo⁶⁺-reducing Bacterium. *Applied and Environmental Microbiology* **59**: 1176-1180
- Glenn, J. L., and Clane, E. L. 1956. Studies on metalloflavoprotein V. The action of silicomolybdate in the reduction of cytochrome c by aldehyde oxidase. *Biochimica et Biophysica Acta* **22**:111-115.
- Greenwood, N. N. 1984. Earnshaw A. Chemistry of the elements. Pergamon Press. Oxford. pp1167.
- Hille, R. 1996. The mononuclear molybdenum enzymes. *Chemical Review* **96**: 2757-2816
- Huisingh, J. and Matrone, G. 1972. Copper-molybdenum interactions with the sulfate-reducing system in rumen microorganisms. *Proceedings of the Society for Experimental Biology and Medicine* **139**:518-521.
- International Molybdenum Association, IMO, 1998.
- Isenberg, H. D. (ed). 1992. *Clinical Microbiology Procedures Handbook*. American Society for Microbiology, Washington, D.C.
- Jarrell, W. M., Page, A. L. and Elsewji, A. A. 1980. Molybdenum in the environment. *Residue Review* **7**:41-43.
- Johnsen, U., Selig, M., Xavier, K. B., Santos, H., and Schönheit, P. 2001. Different glycolytic pathways for glucose and fructose in the halophilic archaeon *Halococcus saccharolyticus*. *Archives of Microbiology* **175**: 52-61
- Johnson, J. L., Waud, W. R., Rajagopalan, K. V., *et al.* 1980. Inborn errors of molybdenum metabolism: Combined deficiencies of sulfite oxidase and xanthine dehydrogenase in a patient lacking the molybdenum cofactor. *Proceeding of the National Academy of Science* **77**: 3715-3719.

- Kazansky, L. P and Fedotov, M. A. 1980. Phosphorous-³¹ and oxygen-¹⁷ n.m.r. evidence of trapped electrons in reduced 18-molybdodiphosphate (v), $P_2Mo_{18}O_{62}^{28-}$. *Journal of the Chemical Society, Chemical Communications* **13**: 644–647.
- Killefer, D. H. and Linz, A. 1952. Molybdenum compounds; their chemistry and technology. Inter science Publisher. New York. pp50.
- Kim, J and Rees, D. C. 1992. Chrystallographic structure and functional implications of the nitrogenase molybdenum-iron protein from *Azotobacter Vinelandii*. *Nature* **360**:553–560.
- Koch, K.E., Nolte, K.D., Duke, E.R., McCarty, D.R. and Avigne, W.T. 1992. Sugar Levels Modulate Differential Expression of Maize Sucrose Synthase Genes. *American Society of Plant Physiologist* **4**: 59-69.
- Laemmli, U. K. 1970. Cleavage of structural proteins during the assembly of the head of bacteriophage T₄. *Nature* **227(5259)**: 680-685.
- Lee, J. D. 1977: *Concise inorganic chemistry*. Van Reinhold Company. New York. pp 325.
- Lener, J., Bibr, B. 1984. Effects of molybdenum on the organism (a review). *Journal of Hygiene, Epidemiology, Microbiology & Immunology* **4**: 405-419.
- Margush, T. and McMorris, F. R. 1981. Consensus n-trees. *Bulletin of Mathematical Biology* **43**: 239-244.
- Mason, J. 1986. Thiomolybdates: mediators of molybdenum toxicity and enzyme inhibitors. *Toxicology* **42**:99-109.
- Mitchell, P., Imgrund, H., Morhrbacher, H. and Outteridge, T. 2009. Moly Review. International Molybdenum Association. London. United Kingdom.
- Mizrahi, L. and Aчитuv, Y. 1989: Effect of heavy metals ions on enzyme activity in the Mediterranean mussel, *donax trunculus*. *Environmental Contemlnation and Toxicology* **42**; 854-859.
- Moura, J. J. G., Xavier, A.V., Bruschi, M., Le Gall, J., Hall, D. O. and Cammack, R. 1976. A molybdenum-containing iron-sulphur protein from *desulphovibrio gigas*. *Biochemical and Biophysical Research Communications* **72**: 782-789.
- Müller, A. and Serain, C. 2000. Soluble Molybdenum Blues. *Accounts of Chemical Research*, **33**; 2–10.
- Newton, D. E. 1992. Molybdenum. In: Baker, L.W.(Ed). Chemical elements from carbon to krypton. An imprint of the Gale Group. Detriot. pp 343-347
- Othman, a. R. et al. 2013. “Kinetics of Molybdenum Reduction to Molybdenum Blue by *Bacillus* Sp. Strain A.rzi.” *BioMed Research International* 2013: 9.

- Prescott, L. M., Harley, J. P. and Klein, D. A. 2002. *Microbiology*. McGraw Hill. New York. p 1026.
- Rajagopalan, K. V. 1988. Molybdenum: an essential trace element in human nutrition. *Annual Review of Nutrition* **8**: 401-427.
- Rebelo, J. M., Dias, J. M., Huber, R., Moura, J. J., Romão, M. J. 2001. Structure refinement of the aldehyde oxidoreductase from *Desulfovibrio gigas* (MOP) at 1.28 Å. *Journal of Biological Inorganic Chemistry* **6**(8):791-800.
- Romão, M. J., Knäblein, J., Huber, R. and Moura, J. J. 1997. Structure and function of molybdopterin containing enzymes. *Progress in Biophysics and Molecular Biology* **68**: 121-144.
- Runnells, D. D., Kaback, D. S. and Thurman, E. M. 1976. Geochemistry and sampling of molybdenum in sediment, soils and plant in Colorado. In: Chappel, W. R., Peterson, K. K. (eds). *Molybdenum in the Environment*. Marcel and Dekker, Inc., New York.
- Schroeder H. A. 1970. A sensible look at air pollution by metals. *Archives of Environmental Health* **21**:798-806.
- Scopes, R. K. 1988. *Protein purification, principles and practice*. Springer-Verlag, New York. pp. 12
- Shineldecker, C. L. 1992. *Handbook of environmental contaminants: a guide to self assessment*. U.S.A., Lewis Publisher. pp76.
- Shukor, M. Y., Shamaan, N. A., Syed, M. A., Lee, C. H. and Karim, M. I. A. 2000. Characterization and quantification of molybdenum blue production in *Enterobacter cloacae* Strain 48 using 12-Molybdophosphate as the reference compound. *Asia Pacific Journal of Molecular Biology and Biotechnology* **8**(2): 167-172.
- Shukor, M. Y., Lee, C. H. Omar, I. Karim, M. I. A. Syed, M. A. and Shamaan, N. A. 2003. Isolation and characterization of a molybdenum-reducing enzyme in *Enterobacter cloacae* Strain 48. *Pertanika Journal of Science and Technology* **11**(2): 261-272.
- Shukor, Y., Adam, H. Ithnin, K., Yunus, I., Shamaan, N. A. and Syed, M. A. 2007. Molybdate Reduction to Molybdenum Blue in Microbe Proceeds via a Phosphomolybdate Intermediate. *Journal of Biological Science* **7**(8): 1448-1452.
- Shukor, M. Y., Habib, S. H. M., Rahman, M. F. A., Jirangon, H., Abdullah, M. P. A., Shamaan, N. A. and Syed, M. A. 2008a. Hexavalent molybdenum reduction to molybdenum blue by *S. marcescens* strain Dr.Y6. *Applied Biochemistry and Biotechnology* **149** (1): 33-43.

- Shukor, M. Y. A., Rahman, F. A., Shamaan, Lee, C. H., Karim, M. I. A. and Syed, M. A. 2008b. An Improved Enzyme Assay for Molybdenum-Reducing Activity in Bacteria. *Applied Biochemistry and Biotechnology* **144**: 293-300.
- Shukor, M.Y., Rahman, M.F., Suhaili, Z., Shamaan, N.A., Syed, M.A. 2009a. Bacterial reduction of hexavalent molybdenum to molybdenum blue. *World Journal Microbiology and Biotechnology* **25**: 1225-1234.
- Shukor, M. Y., Ahmad, S. A., Nadzir, M. M. M., Abdullah, M. P., Shamaan, N. A., and Syed, M. A. 2009b. Molybdate reduction by *Pseudomonas sp* strain DRY2. *Journal of Applied Microbiology* **108**: 2050-2058.
- Shukor, M. Y., Rahman, M.F., Suhaili, Z., Mustafa, S., Shamaan, N.A., and Syed, M.A. 2009c. Reduction of Mo (VI) by the bacterium *Serratia sp.* strain DRY5. *Journal of Environmental Biology* **30(1)**: 65-72.
- Shukor, M. Y., Rahman, M. F., Shamaan, N. A., and Syed, M. A. 2009d. Reduction of molybdate to molybdenum blue by *Enterobacter sp.* Strain Dr.Y13. *Journal of Basic Microbiology* **49**:1-12.
- Shukor, M. Y., Hamdan, M. H., Othaman, M. A., Shamaan, N. A., and Syed, M. A. 2009e. Mo (VI) Reduction to Molybdenum Blue by *Serratia marcescens* Strain Dr.Y9. *Polish Journal of Microbiology* **58(2)**: 141-147.
- Shukor, M. Y., Rahman, M. F., Suhaili, Z., Shamaan, N. A., and Syed, M. A. 2010. Hexavalent Molybdenum Reduction to Mo-blue by *Acinetobacter calcoaceticus*. *Folio Microbiologica* **55(2)**: 137-143.
- Sidgwick, N. V.1984. The chemical elements and their compounds. Clarendon Press. Oxford. pp 439-457
- Smyth, H. F. 1956. Hygienic standard for daily inhalation. *American Industrial Hygiene Association quarterly* **17**:129-185.
- Sugio, T., Tsujita, Y., Katagiri, T., Inagaki, K. and Tanao, T. 1988. Reduction of Mo⁶⁺ with elemental sulphur by *Thiobacillus ferrooxidans*. *Journal of Bacteriology* **170** (12): 5956-5959.
- Thompson, J. D., Higgins, D. G., Gibson, T. J., *et.al.* 1994. CLUSTAL W, improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Reserch* **22**: 4673-4680.
- Tsongas, T. A., Meglen, R. R., Walravens, P. A., and Chappell, W. R. 1980. Molybdenum in the diet: an estimate of average daily intake in the United States. *American Journal of Clinical Nutrition* **33**: 1103- 1107.
- Turnland, J. R., Keyes, W. R. and Peiffer, G. L. 1995. Molybdenum absorption, excretion, and retention studied with stable isotopes in young men at five intakes of dietary molybdenum. *American Journal Clinical of Nutrition* **62**:790-796.

Vyskocil, A. and Viau, C. 1999. Assessment of Molybdenum Toxicity in Humans, *Journal of Applied Toxicology* **19**: 185–192.

Underwood, E.J. (1977) "Trace Elements in Human and Animal Nutrition. 4th Edition. Academic Press, London, New York.

Wester, P. O. 1971. Trace element balances in two cases of pancreatic insufficiency. *Acta Medica Scandinavica* **190**:155-161.

Yoshida, M., Hattori, H., Ota, S., Yoshihara, K., Kodama, N., Yoshitake, Y., and Nishimuta, M. 2006. Molybdenum balance in healthy young Japanese women, *Journal of Trace Elements in Medicine and Biology* **20**: 245-252.

