



**UNIVERSITI PUTRA MALAYSIA**

***DEVELOPMENT OF MINIATURIZED TAPERED FIBER OPTICS DIAMINE  
OXIDASE-BASED SENSOR FOR HISTAMINE DETECTION***

***HAMZA BOKO USMAN***

**FBSB 2015 9**



**DEVELOPMENT OF MINIATURIZED TAPERED FIBER OPTICS DIAMINE  
OXIDASE-BASED SENSOR FOR HISTAMINE DETECTION**

By

**HAMZA BOKO USMAN**

**Thesis submitted to the School of Graduate Studies, Universiti Putra Malaysia, in  
fulfillment of the Requirements for the Degree of Master of Science**

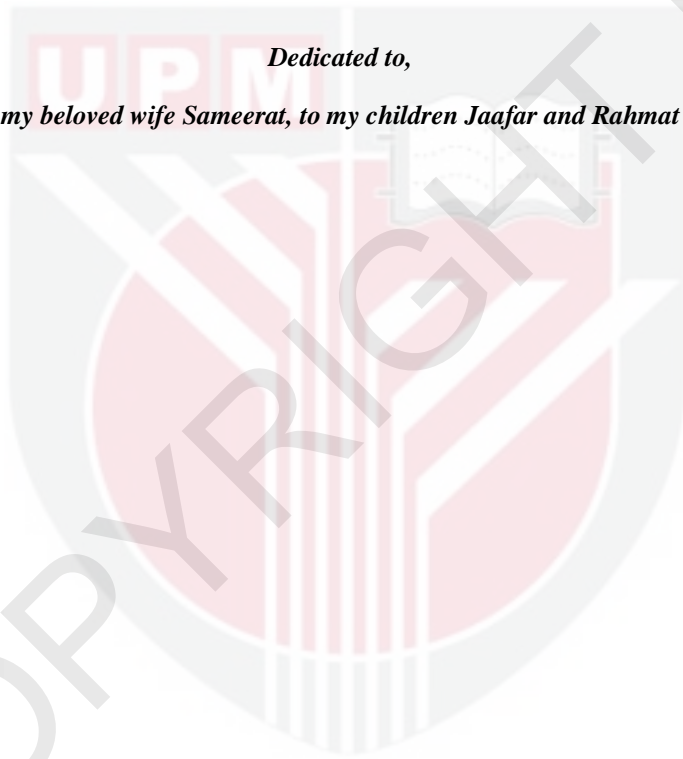
**May 2015**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



*Dedicated to,  
my beloved wife Sameerat, to my children Jaafar and Rahmat*



© COPYRIGHT UPM

© COPYRIGHT UPM



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

**DEVELOPMENT OF MINIATURIZED TAPERED FIBER OPTICS DIAMINE OXIDASE-BASED SENSOR FOR HISTAMINE DETECTION**

**By**

**USMAN BOKO HAMZA**

**May, 2015**

**Chairman: Abu Bakar Salleh, PhD**

**Faculty: Biotechnology and Biomolecular Sciences**

Estimating the level of histamine in fish and fish products is very important because of their implication in fish poisoning in human; hence, ascertaining histamine levels in the afore-mentioned serves as a chemical index for spoilage. Factors such as expensive instrumentation, time consumption, size and weight are some of the problems with conventional analytical methods of detection. Tapered fiber optics-enzyme based sensors are prospective candidates towards resolving these quagmires due to their portability and simplicity vis-à-vis currently available detectors. Taking into cognizance immobilization as the backbone of any biosensor, this work, reports a technique to immobilize an ordered multilayer of diamine oxidase (DAO) by means of chemical cross-linking on the biconical tapered fiber surface step-wisely alternating between chitosan, glutaraldehyde (GA) and the enzyme. The optimum parameters for the fabricated biosensor included 160 mg/ml DAO, 0.5% chitosan, 2.5% GA, pH 7.0, and tapered fiber surface of waist diameter 12  $\mu\text{m}$  and length 20 mm. A spectrophotometric signal resulted from horseradish peroxidase catalyzed reduction of  $\text{H}_2\text{O}_2$ , a secondary product of the oxidative deamination of histamine monitored at 450 nm in 0.1 M phosphate buffer (pH 7.0 and room temperature). Atomic force microscopy (AFM), scanning electron microscope (SEM) and spectrophotometric technique confirmed the functionality of the biosensor. The biosensor showed a response and recovery time of 14 sec, a linear response range up to 1.5 mM, a good sensitivity of  $0.64 \text{ mM}^{-1}$  with detection and quantification limits towards histamine of 0.086 mM (15.8 ppm) and 0.204 mM (37.7 ppm) and a linear response range of 0-1.5 mM. The sensor showed an excellent anti-interferents property towards the common interferents' agents of <5%, with good recovery performance towards varying concentration of histamine ranging from 95.6 to 103.6% (RSD <5%). It showed operational stability to up to 40 repeated analyses without significant loss of sensitivity. The developed miniaturized biosensor has a good potential for use in quantitative measurement of histamine in seafood.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

**PEMBANGUNAN DIAMINE OXIDASE FIBER OPTIK BERSAIZ KECIL  
TIRUS BERASASKAN SENSOR UNTUK PENGESANAN HISTAMINE**

Oleh

**USMAN BOKO HAMZA**

**Mei 2015**

**Pengerusi: Abu Bakar Salleh, PhD**

**Fakulti: Bioteknologi dan Sains Biomolekul**

Penganggaran tahap histamin dalam ikan dan produk ikan adalah sangat penting kerana implikasinya terhadap keracunan ikan pada manusia; oleh itu, penentuan tahap histamin adalah penting untuk mengetahui tahap kerosakan indeks kimia. Antara masalah bagi analisis pengesanan konvensional adalah punca masalah disebabkan oleh faktor seperti peralatan mahal, penggunaan masa, saiz dan berat badan. Maka, pengesanan yang berasaskan gentian optik tirus-enzim telah digunakan untuk menyelesaikan masalah tersebut oleh kerana kebolehan pengesanan yang mudah alih dan sederhana untuk digunakan pada masa kini. Menyedari ia sebagai tulang belakang kepada mana-mana biosensor, projek ini melaporkan teknik untuk memegunkan satu lapisan tersusun dengan mengarahkan diamine oxidase (DAO) dengan cara kimia silang pada permukaan gentian tirus yang dwikon secara selingan antara kitosan, glutaraldehid (GA) dan enzim. Parameter optimum untuk biosensor yang direka adalah dengan menggunakan 160 mg/ml DAO, 0.5% kitosan, 2.5% GA, pH 7.0, dan permukaan gentian tirus yang mempunyai diameter 12  $\mu\text{m}$  dan panjang 20 mm. Isyarat spektrofotometri dihasilkan oleh horseradish peroxidase yang menjadi pemangkin pengurangan  $\text{H}_2\text{O}_2$ , produk sekunder oleh pengoksidaan deaminasi histamin yang telah dipantau pada 450 nm dalam 0.1 M larutan penimbal fosfat (pH 7.0 dan suhu bilik). Daya mikroskop atom (AFM), mikroskop elektron imbasan (SEM) dan teknik spektrofotometri telah mengesahkan keupayaan biosensor itu. Hasil keputusan biosensor telah menunjukkan tindak balas masa selama 14 saat, dan peningkatan linear sehingga 1.5 mM, kepekaan yang baik iaitu  $0.64 \text{ mM}^{-1}$  dengan pengesanan dan had kuantifikasi histamin sebanyak 0.086 mM (15.8 ppm) dan 0.204 mM (37.7 ppm) dan tindak balas linear sebanyak 0-1.5 mM. Sensor ini telah menunjukkan kejayaan anti-interferen terhadap ejen yang biasa iaitu <5%, dengan peningkatan prestasi terhadap perbezaan kepekatan histamine di antara 95.6 sehingga 103.6% (RSD <5%). Ini menunjukkan kestabilan operasi meningkat dalam 40 kali analisis tanpa kehilangan tahap sensitiviti dengan ketara. Pembangunan biosensor mini ini telah menunjukkan potensi yang baik untuk digunakan dalam penentuan kuantitatif sebatian histamine makanan lain.

## ACKNOWLEDGEMENTS

In the name of Allaah, the most gracious, the most merciful, all praises and glorifications are due to him.

The author would like to thank my always-good-to-go Advisor, Professor Dato' Dr Abu Bakar Salleh for his wonderful insight and mentorship throughout my graduate studies. In addition, I acknowledge my Co-Supervisors Dr Amir Syahir Hamzah, Dr Mohammad Hafiz Abubakar for their tireless effort and to all members of the EMTECH Biosensor Research Group.

Lastly, I am grateful to all members of my lovely family for their supports, and prayers. Special thanks to my beloved wife for being with me throughout the period of my studies in Malaysia.

I certify that a Thesis Examination Committee has met on 26<sup>th</sup> May 2015 to conduct the final examination of (student's name) on his (her) thesis entitled "development of miniaturized tapered fiber optics diamine oxidase-based sensor for histamine detection" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science Degree in Biochemistry.

Members of the Thesis Examination Committee were as follows:

**Adam Leow Thean Chor, PhD**

Faculty of Biotechnology and Biomolecular Sciences  
Universiti Putra Malaysia  
(Chairman)

**Haslina Ahmad, PhD**

Faculty of Sciences  
Universiti Putra Malaysia  
(Internal Examiner)

**Azila Binti Abd Aziz, PhD**

Associate Professor  
Faculty of Chemical Engineering  
Universiti Teknologi Malaysia  
Malaysia  
(External Examiner)

---

**Zulkarnain Zainal, PhD**

Professor and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

**Abu Bakar Salleh, PhD**

Professor

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Chairman)

**Amir Syahir Amir Hamzah, PhD**

Senior Lecturer

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Member)

**BUJANG KIM HUAT, PhD**

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date:

## Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No.: \_\_\_\_\_

## Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: \_\_\_\_\_  
Name of Chairman of  
Supervisory  
Committee: \_\_\_\_\_

Signature: \_\_\_\_\_  
Name of Member of  
Supervisory  
Committee: \_\_\_\_\_

## TABLE OF CONTENTS

	Page
<b>ABSTRACT</b>	<b>i</b>
<b>ABSTRAK</b>	<b>ii</b>
<b>ACKNOWLEDGEMENTS</b>	<b>iii</b>
<b>APPROVAL</b>	<b>iv</b>
<b>DECLARATION</b>	<b>vi</b>
<b>LIST OF TABLES</b>	<b>x</b>
<b>LISTS OF FIGURES</b>	<b>xi</b>
<b>LIST OF ABBREVIATIONS</b>	<b>xiii</b>
<b>CHAPTER</b>	
<b>1 INTRODUCTION</b>	<b>1</b>
1.1 Rationale and Motivation	1
1.2 Objectives	3
<b>2 LITERATURE REVIEW</b>	<b>4</b>
2.1 Histamine Poisoning	4
2.2 Diamine Oxidase (DAO)	5
2.3 Biosensors	5
2.4 Bioreceptor (Biorecognition Element)	6
2.5 Enzyme Immobilization	6
2.5.1 Advantages of Enzymes Immobilization	7
2.5.2 Immobilization Techniques/Methods	7
2.5.3 Polymer for Enzyme Immobilization	8
2.5.4 Chitin/Chitosan as Polymer for Enzyme Immobilization	9
2.6 Biosensor Classification Based on Transducer Type	10
2.7 Fiber Optics Study	10
2.7.1 Chemical Composition of Typical Optical Fiber	11
2.7.2 Detection Principle in Evanescent Sensor	11
2.7.3 Tapered Fiber Geometries	12
2.8 Biogenic Amine Biosensors	14
<b>3 METHODOLOGY</b>	<b>19</b>
3.1 Chemicals and Reagent	19
3.2 Instruments	20
3.3 Preparation of Standard Solutions	24
3.3.1 Preparation of 0.1 M sodium phosphate buffer	24
3.3.2 Preparation of 1% acetic acid	24
3.3.3 Preparation of 2.5% glutaraldehyde (GA)	24
3.3.4 Preparation of 2% chitosan film	24
3.3.5 Preparation of standard DAO solution	24
3.3.6 Preparation of 75 mM histamine (substrate)	24
3.3.7 Preparation of assay solution	24
3.4 Preparation of Bioreceptor	25
3.4.1 Diamine oxidase immobilization on chitosan	

	(Cross-linking Method)	25
3.4.2	Effect of chitosan concentration on immobilized DAO	25
3.4.3	Effect of enzyme (DAO) loading	25
3.4.4	Effect of pH on DAO immobilization	26
3.5	Transducer Fabrication, Characterization and Optimization	27
3.5.1	Fiber Tapering	27
3.5.2	Tapered Fiber Characterization	27
3.5.3	Taper Cleaning/Activation	27
3.6	Biosensor Integration	27
3.6.1	Enzyme Immobilization on Tapered Fiber	27
3.6.2	Experimental	28
3.6.3	Histamine Detection Experiments	29
3.7	Biosensor Validation	29
3.7.1	Repeatability and Reproducibility	29
3.7.2	Response Time	30
3.7.3	Sensitivity	30
3.7.4	Limit of Detection (LOD) and Limit of Quantification (LOQ)	30
3.7.5	Interference Study	30
3.7.6	Recovery Characteristic of the Histamine Biosensor	31
3.7.7	Operational Stability of the Biosensor	31
<b>4</b>	<b>RESULTS AND DISCUSSION</b>	<b>32</b>
4.1	Bioreceptor Preparation	32
4.1.1	Immobilization of Diamine Oxidase (Cross-linking Method)	32
4.1.2	The effect of Chitosan Concentration on the Activity of Immobilized DAO	32
4.1.3	The effect of DAO Loading	33
4.1.4	Effect of pH on DAO Immobilization	34
4.2	Transducer Preparation	35
4.2.1	Taper Characterization and Optimization	35
4.3	Biosensor Integration	41
4.3.1	Enzyme Multilayer Assembly	41
4.3.2	Histamine Detection	44
4.4	Biosensor Validation	47
4.4.1	Sensitivity	47
4.4.2	Limit of Detection and Limit of Quantification	47
4.4.3	Repeatability and Reproducibility	47
4.4.4	Response Time	48
4.4.5	Interference Studies	49
4.4.6	Recovery Performance of Histamine Biosensor	50
4.4.7	Operational Stability of Biosensor	51
<b>5</b>	<b>CONCLUSION AND RECOMMENDATION</b>	<b>52</b>
	<b>REFERENCES</b>	<b>53</b>
	<b>BIODATA OF STUDENT</b>	<b>59</b>

## LIST OF TABLES

<b>Table</b>	<b>Page</b>
2.1. Biogenic amines (BAs) and there precursors	4
2.2. Types of transducers and their detection principle	10
2.3. Summary of basic characteristics of some biogenic amine biosensors fabricated over the years.	15
4.1. Immobilization yield of diamine oxidase (DAO) in 0.5 % chitosan.	34
4.2. Recovery performance of histamine biosensor.	50
4.3. Short time operational stability of the biosensor	51



## LISTS OF FIGURES

<b>Figure</b>		<b>Page</b>
2.1.	Generalized Component of a simple biosensor	6
2.2.	Fabrication of a tapered tip	13
2.3.	Fabrication of biconical tapered fiber and its basic components (N = cladding diameter, a = up taper, b = waist length, c = up taper, and d = waist diameter).	14
3.1.	Generalized project overview.	19
3.2.	Halogen light source Type HL-2000	20
3.3.	USB4000 Vis-NIR spectrometer	21
3.4.	Fiber stripper	21
3.5.	Handy cleaver	22
3.6.	Programmable fusion splicer Type-37	22
3.7.	A complete set up of a glass-processing machine Vytran).	23
3.8.	A glass-processing machine (Vytran), showing the tapering part of the machine.	23
3.9.	Fabrication of diamine oxidase (DAO) sensor surfaces and experimental set up for histamine detection	28
3.10.	Schematic immobilization process through LbL assembly multilayer.	29
4.1.	Effect of Chitosan concentration on immobilized DAO membrane.	33
4.2.	Effect of membrane pH on the activity of immobilized DAO on histamine solution (0.1 M phosphate buffer pH 7.4)	35
4.3.	A taper profile for glass-processing machine (Vytran), showing the various input parameters.	36
4.4.	Transmission in taper with a length of 10 mm and 50 $\mu\text{m}$ waist diameter the wavelength of 400-800 nm.	37
4.5.	Transmission in taper with a length of 10 mm and 12 $\mu\text{m}$ waist diameter over the wavelength of 400-800 nm.	37
4.6.	Effect of variation in waist diameter on percentage transmission using water, glucose and histamine as analytes at 450 nm.	38
4.7.	Transmission change over the wavelength 400-800 nm because of change in taper length.	39
4.8.	SEM image of a normal fiber (untapered) with a total cladding diameter of 125 $\mu\text{m}$ (x150)	40
4.9.	SEM image of a tapered fiber with a waist diameter 20 $\mu\text{m}$	40
4.10.	SEM of tapered fiber with waist diameter 12 $\mu\text{m}$ before coating	41
4.11.	SEM image of a coated fiber with a waist diameter 12 $\mu\text{m}$ .	42
4.12.	AFM image of coated tapered fiber showing surface roughness in a fiber of 12 $\mu\text{m}$	43
4.13.	SEM image of the materials coated on the fiber.	43
4.14.	Vis- NIR spectra evolution of the biosensor showing the absorbance change towards different histamine concentration (H1 = 1 mM, H2 = 2 mM, H3 = 4 mM, H4 = 6 mM and H5 = 10 mM) in a taper with 20 $\mu\text{m}$ and 10 mm length.	44

4.15.	Vis-NIR spectra evolution of optimized biosensor showing the absorbance change towards different histamine concentration (H1=0.1, H2 =0.2, H3=0.4, H4 =0.6, H5=0.8, H6 = 1.0, and H7 =1.5 mM) in a taper with 12 $\mu$ m diameter and 20 mm length at 400-650 with distinct peak at 450 and 550nm.	45
4.16.	Response curve of the biosensor towards in the histamine concentration of 0- 4.5 mM.	46
4.17.	The linear dynamic range of the histamine biosensor.	46
4.18.	The reproducibility of the biosensor obtained when four different sensor where tested (RSD 2.5 %) with 0.6mM histamine	47
4.19.	The repeatability of the biosensor obtained from the analysis of 10 samples with 0.6 mM histamine (RSD 3.5%).	48
4.20.	A strip chart of response time (sec) of the biosensor towards different specific concentration of histamine.	48
4.21.	Response and recovery time (sec) taken from one absorbance peak in the strip chart.	49
4.22.	The strip showing the result of the anti-interference properties of the biosensor (H = histamine, A.A = ascorbic acid, and U.A = uric acid).	50

## LIST OF ABBREVIATIONS

AO	Amine oxidase
BA	Biogenic amine
Cad	Cadaverine
DAO	Diamine oxidase
DD	Degree of deacetylation
GA	Glutaraldehyde
GOD	Glucose oxidase
HMT	Histamine-N-methyl transferase
IUPAC	International union of Pure and applied chemistry
KDA	kilo Dalton
LbL	Layer by layer
LOD	Limit of detection
LOQ	limit of quantification
MMF	Multi-mode fiber
ODA	O-Dianisidine
PEGDGE	Poly(ethylene glycol) diglycidyl ether
PEM	Polyelectrolyte multilayer
POD	Peroxidase
PPM	Part per million
PSAO	Pea-Seedling amine oxidase
PUO	Putrescence oxidase
Put	Putrescine
SAM	Self-assembly multilayer
SPR	Surface Plasmon resonance
TFOB	Tapered Fiber Optic Biosensor

## CHAPTER 1

### INTRODUCTION

#### 1.1 Rationale and motivation

Histamine is a strong bioactive compound that can exert many responses within the body. These compounds apart from their biochemical significance to the biological system, they have been linked with fish poisoning (scombrototoxicosis, scombroid fish poisoning or histamine fish poisoning). An illness originally referred to as “scombroid poisoning”, due its association with fish, in the families’ Scombridae (such as mackerel and tuna) and Scomberesocidae (such as saury) although other non-scombroid species such as mahi-mahi, sardines, pilchards, anchovies, herring, marlin and bluefish also were implicated (Steiner, Meier, Duerkop, & Wolfbeis, 2010). Histamine poisoning is associated with higher level of histamine in fish (Keow et al., 2007). Histamine was observed to accumulate in tissues of fish and other seafood when spoilage commenced during storage because of inadequate refrigeration or preservation (Chemnitius & Bilitewski, 1996). Despite successes to unravel the recommended level as chemical indicator for safety or otherwise for the consumption of fish and it related products, with regard histamine toxicity, however, the big issue is how to unravel a simple and robust method to detect and determine these levels, with the global demand of fish and aquaculture products is constantly increasing. Current data for fish and aquaculture showed a total annual supply of 148, 154 and 158 million tons in 2010, 2011 and 2012, respectively, with increasing estimates for 2013 (FAO., 2014). Fish and fish products, cheese, meat and other variety of food had been widely documented to contain high level of biogenic amines preventing the consumption of spoiled food would be better than curing the disease.

In the past, several conventional technique such as derivatisation with fluorescent reagents, chromatographic methods of separation, electrophoresis, immunochemical methods, titration and lots more have been proposed as possible options (Tombelli & Mascini, 1998). However, these methods tends to disallow an easily and continuous monitoring process, because these instruments are costly, slow, needs well-trained personnel, require sample pre-treatment and also because of their physical characteristics such as size weight and fragility (Mello & Kubota, 2002).

The goal of biosensor research is to develop sensors capable of addressing these lapses, due to need for real time quantitative and/or qualitative measurement in the field and so far, it remains the best alternative to ease the routine analysis of fish and other seafood. Therefore, the design of a biosensor device capable of eliminating the lapses, and also maintaining the precision and accuracy of laboratory analysis is of paramount importance (Monk & Walt, 2004). Biosensor technology has attracted researchers due to their characteristic easy-to-use, accurate, fast, inexpensive, less sample pre-treatment, potential for miniaturization, simple and portable equipped platform for construction and high sensitivity and specificity in recognizing their analytes (Mello & Kubota, 2002).

Leland C Clark Jr is celebrated as the father of biosensor since he first coined the 'biosensor concept' in his published definitive paper on the oxygen electrode in 1956 (Chauhan, Vibhuti, & Singh, 2004). Clark's idea was realized when the first commercialized glucose biosensor was re-launch in 1975 (first launch 1973) by the Yellow Springs Instrument Company (Ohio). This is followed by a lactate analyzer (LA 640) device in 1976, by La Roche, Switzerland (Chaubey & Malhotra, 2002). Over the past decades, different devices evolved under the name 'biosensor', utilizing different platforms and biosensing receptors (biocatalytic, bioaffinity and hybrid receptors). Biosensor application has evolved from the area of laboratory testing to commercialization. They have gained wider application namely in the field of medical diagnostics, food safety, homeland security, industrial process control, battle field and environmental monitoring (Lee, 2008). In 2008, the global market for commercialized biosensors stands at about \$7 billion, with medical diagnostics (e.g., glucose biosensors and pregnancy test strips) being dominant and this is estimated to reach US\$12 billion by the year 2015 (Bahadır & Sezgentürk, 2015; Lee, 2008). Bayer, Roche Diagnostics, and LifeScan occupies the apex of competing mediated biosensors companies with a combined percentage sell of about 85% of the world market for biosensor.

There are many efforts geared towards developing a biosensor for biogenic amine detection, and with respect to the solid support. Few mentions amongst several of these works by these researchers are amperometry employing a screen printed electrode (SPE) (Chemnitiu & Bilitewski, 1996; Keow et al., 2012; Keow et al., 2007; Wimmerová & Macholán, 1999). Amperometry utilizing Pt electrode (Carelli, Centonze, Palermo, Quinto, & Rotunno, 2007; Carsol & Mascini, 1999; Karube, Satoh, Araki, Suzuki, & Yamada, 1980; Tombelli & Mascini, 1998), and utilizing a control pore-glass electrode (Yao, Satomura, & Wasa, 1992), and a capacitive sensor utilizing copper electrode (Wasoh et al., 2012). However, in any biotechnological research, the commercialization process is a stepwise gradual improvement from existing system until is made available to the final users in a portable and simplified form. Therefore, looking into the prospect of having a miniaturized diamine oxidase-based biosensor would be a step towards achieving this goal. The advantages offered by miniaturized biosensor include the requirement of small quantity of biorecognition element for biosensor development, portability, and thus a prospect for a miniaturized and disposable type of biosensor.

To the best of our knowledge, among the biogenic amine biosensors fabricated so far, no much effort geared towards fabricating a tapered fiber optic biosensor for DAO. Herein we propose the use of tapered fiber optics (taper) for this research work, due to its advantages over the use of other transduction techniques. These advantages include compactness, high sensitivity, affordability, fast response time, requirement for small volume of enzyme, lightweight and compatibility with optoelectronic devices (Baldini, Brenchi, Chiavaioli, Giannetti, & Trono, 2012).

## 1.2 Objectives

Therefore, the objectives of these studies are:

- i. To immobilize diamine oxidase on chitosan and cross-linked with bifunctional agent (GA).
- ii. To modify a multi-mode fiber (MMF) through tapering.
- iii. To fabricate a biosensor; through coating the enzyme multilayer, by alternating between chitosan, GA and DAO onto the tapered fiber.
- iv. To optimize and characterize the designed biosensor.



## REFERENCES

- Abdullah, J., Ahmad, M., Karuppiah, N., Heng, L. Y., & Sidek, H. (2006). Immobilization of tyrosinase in chitosan film for an optical detection of phenol. *Sensors and Actuators B: Chemical*, 114(2), 604-609.
- Abu Bakar, F. (2001). *Microbiological and biochemical changes of freshwater prawns (macrobrachium rosenbergii) during storage*. (Doctoral thesis).
- Ahmad, M., & Hench, L. L. (2005). Effect of taper geometries and launch angle on evanescent wave penetration depth in optical fibers. *Biosensors and Bioelectronics*, 20(7), 1312-1319.
- Azmi, N. E., Ahmad, M., Abdullah, J., Sidek, H., Heng, L. Y., & Karuppiah, N. (2009). Biosensor based on glutamate dehydrogenase immobilized in chitosan for the determination of ammonium in water samples. *Analytical Biochemistry*, 388(1), 28-32.
- Azofeifa, D. E., Arguedas, H. J., & Vargas, W. E. (2012). Optical properties of chitin and chitosan biopolymers with application to structural color analysis. *Optical Materials*, 35(2), 175-183.
- Bahadır, E. B., & Sezgintürk, M. K. (2015). Applications of commercial biosensors in clinical, food, environmental, and bioterror/bio warfare analyses. *Analytical Biochemistry*, 478(0), 107-120.
- Baldini, F., Brenchi, M., Chiavaioli, F., Giannetti, A., & Trono, C. (2012). Optical fibre gratings as a tools for chemical and biochemical sensing. *Anal Bioanal Chem*, 402(1), 109-116.
- Benner, R. A. (2012). Scombrotoxin. In K. A. Lampel, S. Al-Khaldi & S. M. Cahill (Eds.), *Food and drug administration. bad bug book, foodborne pathogenic microorganisms and natural toxins. second edition*. (2nd ed., pp. 207-210). U.S.: FDA.
- Carelli, D., Centonze, D., Palermo, C., Quinto, M., & Rotunno, T. (2007). An interference free amperometric biosensor for the detection of biogenic amines in food products. *Biosensors and Bioelectronics*, 23(5), 640-647.
- Carsol, M. -, & Mascini, M. (1999). Diamine oxidase and putrescine oxidase immobilized reactors in flow injection analysis: A comparison in substrate specificity. *Talanta*, 50(1), 141-148.
- Chaubey, A., & Malhotra, B. D. (2002). Mediated biosensors. *Biosensors and Bioelectronics*, 17(6-7), 441-456.

- Chauhan, S., Vibhuti, R., & Singh, H. B. (2004). Biosensors. *Resonance Journal of Science Education*, 9(12), 33-44.
- Chemnitiu, G. C., & Bilitewski, U. (1996). Development of screen-printed enzyme electrodes for the estimation of fish quality. *Sensors and Actuators B: Chemical*, 32(2), 107-113.
- Chemnitiu, G. C., Suzuki, M., Isobe, K., Kimura, J., Karube, I., & Schmid, R. D. (1992). Thin-film polyamine biosensor: Substrate specificity and application to fish freshness determination. *Analytica Chimica Acta*, 263(1-2), 93-100.
- Chen, Z., Kaplan, D. L., Gao, H., Kumar, J., Marx, K. A., & Tripathy, S. K. (1996). Molecular assembly of multilayer enzyme: Toward the development of a chemiluminescence-based fiber optic biosensor. *Materials Science and Engineering: C*, 4(3), 155-159.
- Corres, J. M., Matias, I. R., Bravo, J., & Arregui, F. J. (2008). Tapered optical fiber biosensor for the detection of anti-gliadin antibodies. *Sensors and Actuators B: Chemical*, 135(1), 166-171.
- Draisci, R., Volpe, G., Lucentini, L., Cecilia, A., Federico, R., & Palleschi, G. (1998). Determination of biogenic amines with an electrochemical biosensor and its application to salted anchovies. *Food Chemistry*, 62(2), 225-232.
- El-Sherif, M., Bansal, L., & Yuan, J. (2007). Fiber optic sensors for detection of toxic and biological threats. *Sensors*, 7, 3100-3118.
- Fan, X., White, I. M., Shopova, S. I., Zhu, H., Suter, D. J., & Sun, Y. (2008). Sensitive optical biosensors for unlabeled targets: A review. *Analytica Chimica Acta*, 620, 8-26.
- FAO. (Ed.). (2014). *The state of world fisheries and aquaculture; opportunities and challenges*. Rome: FAO.
- FAO., & WHO. (2012). *Joint FAO/WHO expert meeting on the public health risks of histamine and other biogenic amines from fish and fishery products*. (Meeting Report). Rome, Italy.: FAO/WHO.
- Fiorentino, D., Gallone, A., Fiocco, D., Palazzo, G., & Mallardi, A. (2010). Mushroom tyrosinase in polyelectrolyte multilayers as an optical biosensor for o-diphenols. *Biosensors and Bioelectronics*, 25(9), 2033-2037.
- Frébort, I., Skoupá, L., & Peč, P. (2000). Amine oxidase-based flow biosensor for the assessment of fish freshness. *Food Control*, 11(1), 13-18.
- Gifford, E. L. (2008). *Sensitivity control of optical fiber biosensors utilizing turnaround point long period gratings with self-assembled polymer coatings* (Doctoral dissertation).

- Hamada, Y., Shinohara, Y., Yano, M., Yamamoto, M., Yoshio, M., Satake, K., . . . Usami, M. (2013). Effect of the menstrual cycle on serum diamine oxidase levels in healthy women. *Clinical Biochemistry*, 46(1–2), 99-102.
- Henao-Escobar, W., Domínguez-Renedo, O., Asunción Alonso-Lomillo, M., & Julia Arcos-Martínez, M. (2013). Simultaneous determination of cadaverine and putrescine using a disposable monoamine oxidase based biosensor. *Talanta*, 117(0), 405-411.
- Huang, Y., Mechref, Y., & Novotny, M. V. (1999). N-linked oligosaccharide structures in the diamine oxidase from porcine kidney. *Carbohydrate Research*, 323(1–4), 111-125.
- Kangde, Y., Junjie, L., & Fanglian, Yao and YuJi Yun (Eds.). (2012). *Chitosan-based hydrogels, functions applications*. Boca Raton FL: CRC Press.
- Karube, I., Satoh, I., Araki, Y., Suzuki, S., & Yamada, H. (1980). Monoamine oxidase electrode in freshness testing of meat. *Enzyme and Microbial Technology*, 2(2), 117-120.
- Keow, C. M., Bakar, F. A., Salleh, A. B., Heng, L. Y., Wagiran, R., & Siddiquee, S. (2012). Screen-printed histamine biosensors fabricated from the entrapment of diamine oxidase in a photocured poly(HEMA) film. *International Journal of Electrochemical Science*, 7(5), 4702-4715.
- Keow, C. M., Abu Bakar, F., Salleh, A. B., Heng, L. Y., Wagiran, R., & Bean, L. S. (2007). An amperometric biosensor for the rapid assessment of histamine level in tiger prawn (*penaeus monodon*) spoilage. *Food Chemistry*, 105(4), 1636-1641.
- Khijwania, S. K., & Gupta, B. D. (1999). Fiber optic evanescent field absorption sensor: Effect of fiber parameters and geometry of the probe. *Optical and Quantum Electronics*, 31(8), 625-636.
- Khoshnevisan, K., Bordbar, A., Zare, D., Davoodi, D., Noruzi, M., Barkhi, M., & Tabatabaei, M. (2011). Immobilization of cellulase enzyme on superparamagnetic nanoparticles and determination of its activity and stability. *Chemical Engineering Journal*, 171(2), 669-673.
- Kireyko, A. V., Veselova, I. A., & Shekhovtsova, T. N. (2006). Mechanisms of peroxidase oxidation of o -dianisidine, 3,3',5,5'-tetramethylbenzidine, and o -phenylenediamine in the presence of sodium dodecyl sulfate. *Russian Journal of Bioorganic Chemistry*, 32(1), 71-77.
- Krajewska, B. (2004). Application of chitin- and chitosan-based materials for enzyme immobilizations: A review. *Enzyme and Microbial Technology*, 35(2–3), 126-139.

- Lange, J., & Wittmann, C. (2002). Enzyme sensor array for the determination of biogenic amines in food samples. *Anal Bioanal Chem*, 372, 276-283.
- Lee, T. M. (2008). Over-the-counter biosensors: Past, present, and future. *Sensors*, 8, 5535-5559.
- Lehane, L., & Olley, J. (2000). Histamine fish poisoning revisited. *International Journal of Food Microbiology*, 58(1-2), 1-37.
- Leung, A., Rijal, K., Shankar, P. M., & Mutharasan, R. (2006). Effects of geometry on transmission and sensing potential of tapered fiber sensors. *Biosensors and Bioelectronics*, 21(12), 2202-2209.
- Leung, A., Shankar, P. M., & Mutharasan, R. (2007). A review of fiber-optic biosensors. *Sensors and Actuators B: Chemical*, 125(2), 688-703.
- Loughran, M. G., Hall, J. M., Turner, A. P. F., & Davidson, V. L. (1995). Amperometric detection of histamine at a quinoprotein dehydrogenase enzyme electrode. *Biosensors and Bioelectronics*, 10(6-7), 569-576.
- Male, K. B., Bouvrette, P., Luong, J. H. T., & Gibbs, B. F. (1996). Amperometric biosensor for total histamine, putrescine and cadaverine using diamine oxidase. *Journal of Food Science*, 61(5), 1012-1016.
- Mateo, C., Palomo, J. M., Fernandez-Lorente, G., Guisan, J. M., & Fernandez-Lafuente, R. (2007). Improvement of enzyme activity, stability and selectivity via immobilization techniques. *Enzyme and Microbial Technology*, 40(6), 1451-1463.
- Mei, Y., Ran, L., Ying, X., Yuan, Z., & Xin, S. (2007). A sequential injection analysis/chemiluminescent plant tissue-based biosensor system for the determination of diamine. *Biosensors and Bioelectronics*, 22(6), 871-876.
- Mello, L. D., & Kubota, L. T. (2002). Review of the use of biosensors as analytical tools in the food and drink industries. *Food Chemistry*, 77(2), 237-256.
- Miao, Y., & Tan, S. N. (2001). Amperometric hydrogen peroxide biosensor with silica sol-gel/chitosan film as immobilization matrix. *Analytica Chimica Acta*, 437(1), 87-93.
- Migneault, I., Dartiguenave, C., Bertrand, M. J., Karen, C., & Waldron, K. C. (2004). Glutaraldehyde: Behavior in aqueous solution, reaction with proteins, and application to enzyme crosslinking. *Biotechniques*, 37, 790-802.
- Miller, J. N., & Miller, J. C. (2010). *Statistics and chemometrics for analytical chemistry* (6th ed.). Edinburgh Gate, Harlow Essex CM20 2JE, England.: Pearson Education Limited.

- Monk, D. J., & Walt, D. R. (2004). Optical fiber-based biosensors. *Anal Bioanal Chem*, 379, 931-945.
- Mureșan, L., Valera, R. R., Frébort, I., Popescu, C. L., Csöregi, E., & Nistor, M. (2008). Amine oxidase amperometric biosensor coupled to liquid chromatography for biogenic amines determination. *Microchim Acta*, 163, 219-225.
- Ng, L., Yuan, Y. J., & Zhao, H. (1998). Natural polymer-based sulfite biosensor. *Electroanalysis*, 10(16), 1119-1124.
- Pérez, S., Bartrolí, J., & Fàbregas, E. (2013). Amperometric biosensor for the determination of histamine in fish samples. *Food Chemistry*, 141(4), 4066-4072.
- Poli, B. M., Zampacavallo, G., Parisi, G., Poli, A., & Mascini, M. (2000). Biosensors applied to biochemical fish quality indicators in refrigerated and frozen sea bass reared in aerated or hyperoxic conditions. *Aquaculture International*, 8, 335-348.
- Ravi Kumar, M. N. V. (2000). A review of chitin and chitosan applications. *Reactive and Functional Polymers*, 46(1), 1-27.
- Rinaudo, M. (2006). Chitin and chitosan: Properties and applications. *Progress in Polymer Science*, 31(7), 603-632.
- Sai, V. V. R., Kundu, T., Deshmukh, C., Titus, S., Kumar, P., & Mukherji, S. (2010). Label-free fiber optic biosensor based on evanescent wave absorbance at 280 nm. *Sensors and Actuators B: Chemical*, 143(2), 724-730.
- Shalaby, A. R. (1996). Significance of biogenic amines to food safety and human health. *Sensors and Actuators B: Chemical*, 29(7), 675-690.
- Sharma, H., & Mutharasan, R. (2013). Review of biosensors for foodborne pathogens and toxins. *Sensors and Actuators B: Chemical*, 183(0), 535-549.
- Spagna, G., Andreani, F., Salatelli, E., Romagnoli, D., & Pifferi, P. G. (1998). Immobilization of  $\alpha$ -L-arabinofuranosidase on chitin and chitosan. *Process Biochemistry*, 33(1), 57-62.
- Steiner, M., Meier, R. J., Duerkop, A., & Wolfbeis, O. S. (2010). Chromogenic sensing of biogenic amines using a chameleon probe and the red-green-blue readout of digital camera images. *Analytical Chemistry*, 82(20), 8402-8405.
- Syms, R., & Cozens, J. (1992). *Optical guided waves and devices*. Maidenhead, Berkshire, SL6 2QL England: McGRAW - HILL Book Company Europe.
- Thévenot, D. R., Toth, K., Durst, R. A., & Wilson, G. S. (2001). Electrochemical biosensors: Recommended definitions and classification. *Biosensors and Bioelectronics*, 16(1-2), 121-131.

- Tombelli, S., & Mascini, M. (1998). Electrochemical biosensors for biogenic amines: A comparison between different approaches. *Analytica Chimica Acta*, 358(3), 277-284.
- Wasoh, H., Hengb, L. Y., Abu Bakar, F., Wagiran, R., Salleh, A. B., Yusof, N. A., . . . Abdul Rahmane, F. H. (2012). A simple capacitive biosensor device for histamine measurement. *Sensor Review*, 32(3), 245-250.
- Wimmerová, M., & Macholán, L. (1999). Sensitive amperometric biosensor for the determination of biogenic and synthetic amines using pea seedlings amine oxidase: A novel approach for enzyme immobilisation. *Biosensors and Bioelectronics*, 14(8-9), 695-702.
- Yao, T., Satomura, M., & Wasa, T. (1992). Amperometric flow-injection method for determination of biogenic diamines and hypoxanthine by combined use of immobilized enzyme reactors and a peroxidase electrode. *Analytica Chimica Acta*, 261(1-2), 161-165.
- Yu, J., Zhang, Y., & Liu, S. (2014). Enzymatic reactivity of glucose oxidase confined in nanochannels. *Biosensors and Bioelectronics*, 55(0), 307-312.