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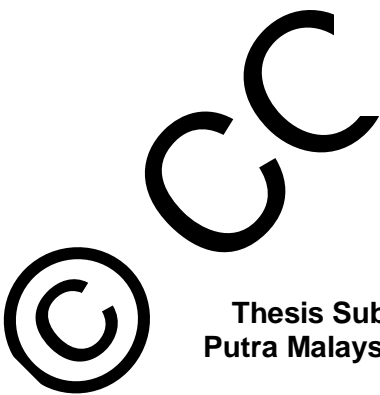
***CLONING AND CHARACTERIZATION OF A NOVEL TRANSCRIPT
ENCODING A RNA-BINDING PROTEIN FROM OIL PALM
(*Elaeis guineensis* Jacq.)***

YEAP WAN CHIN

FBSB 2015 7



**CLONING AND CHARACTERIZATION OF A NOVEL TRANSCRIPT
ENCODING A RNA-BINDING PROTEIN FROM OIL PALM (*Elaeis
guineensis* Jacq.)**



**Thesis Submitted to the School of Graduate Studies, Universiti
Putra Malaysia, in Fulfilment of the Requirements for the Degree of
Doctor of Philosophy**

May 2015

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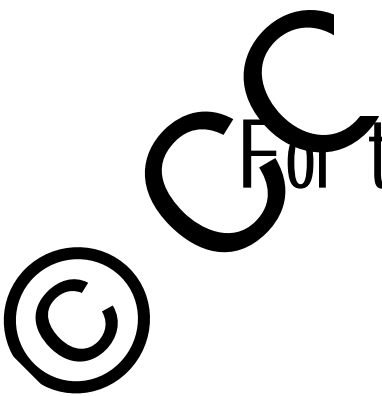
Specially Dedicated

to my



Father
&
Mother

For their Love and Supports



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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

**CLONING AND CHARACTERIZATION OF A NOVEL TRANSCRIPT
ENCODING A RNA-BINDING PROTEIN FROM OIL PALM (*Elaeis guineensis*
Jacq.)**

By

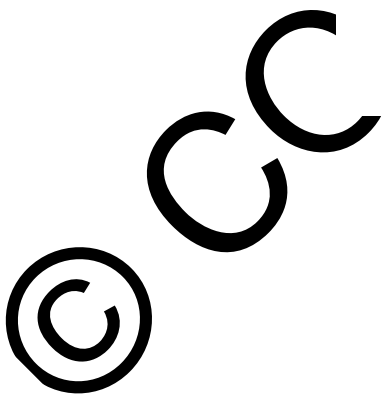
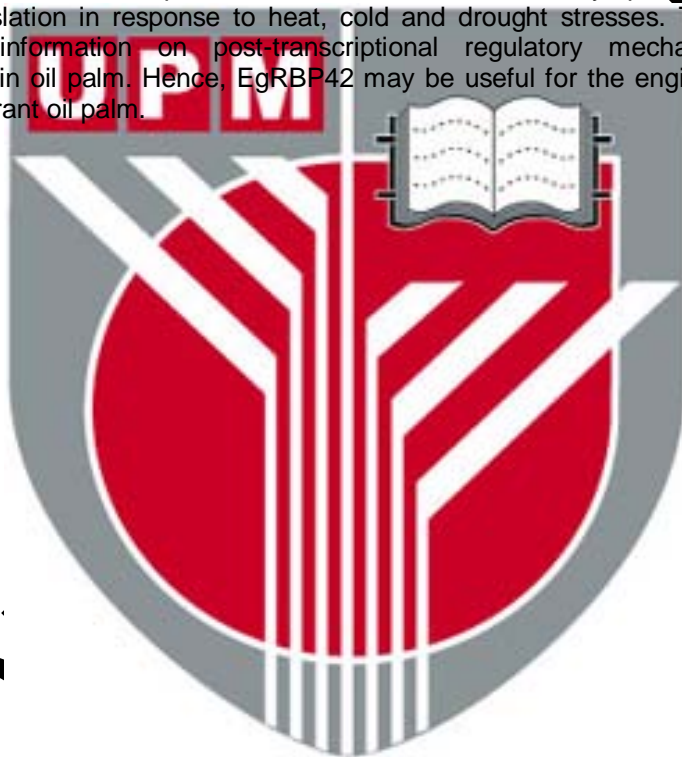
YEAP WAN CHIN



**Chairman: Associate Professor Ho Chai Ling, PhD
Faculty: Biotechnology and Biomolecular Sciences**

RNA-binding proteins (RBPs) have been implicated as regulatory proteins involved in the post-transcriptional processes of gene expression in plants that influence floral development, circadian rhythms, hormone signaling, plant growth, abiotic stress response and tolerance. RBPs have received much attention in *Arabidopsis*, tobacco and rice. Oil palm (*Elaeis guineensis* Jacq.) is the most efficient oil-yielding crop in the world. Environmental stresses have a major impact on oil palm production mainly plant growth, physiology and oil yield. However, the biological functions of RBPs in post-transcription regulation of gene expression in response to stresses are still poorly understood in oil palm. This study aimed to understand the regulation of target transcripts by a RBP from oil palm at post-transcriptional level, the regulatory factors associated with the target RBP in the ribonucleoprotein complex and the involvement of RBP in post-transcriptional RNA mechanism in response to environmental stimuli in oil palm. In this study, a gene designated as *EgRBP42*, encoding a plant heterogeneous nuclear ribonucleoprotein-like RBP was isolated from oil palm. *EgRBP42* was identified from an expressed sequence tag (EL684239) from the oil palm female inflorescence. *EgRBP42* protein consists of two N-terminal RNA recognition motifs and a glycine-rich domain at the C-terminus. The upstream region of *EgRBP42* has multiple light-responsive, stress-responsive and flower development related regulatory elements. Real-time RT-PCR analysis showed that *EgRBP42* was expressed in all oil palm tissues tested, including leaf, shoot apical meristem, root, female inflorescence, male inflorescence and mesocarp with the lowest transcript level in the roots. *EgRBP42* protein interacted with transcripts associated with stress responses, transcription and translation. Validation of consensus sequence of interactive transcripts binding to *EgRBP42* indicated that *EgRBP42* binds to the AG-rich region on its interactive transcripts. Three variants of *EgRBP42* DNA binding regions were detected from

oil palm leaf tissue. The accumulation of *EgRBP42*, its interacting transcripts D Q G L W V D Q W R and transcripts were up-regulated (> 2 fold change) by abiotic stresses, including salinity, drought, submergence, cold and heat stresses in leaf discs (short-term stress treatment for 30 min to 28 hr) and leaves from oil palm seedlings (long-term stress treatment for 7 days). Co-immunoprecipitation and yeast II hybrid interaction studies showed that *EgRBP42* protein interacted with various regulatory factors involved in transcription, nucleocytoplasmic transport, mRNA degradation and translation. The protein accumulation of *EgRBP42* was up-regulated (> 2-fold change) by heat, cold, drought and salinity in oil palm seedlings exposed to long-term stress treatments for 7 days. Collectively, the data suggested that *EgRBP42* is responsive to various abiotic stresses. It is potentially a nucleocytoplasmic transporter of stress-responsive mRNAs from nucleus to cytoplasm for their rapid translation in response to heat, cold and drought stresses. This study provided information on post-transcriptional regulatory mechanisms of *EgRBP42* in oil palm. Hence, *EgRBP42* may be useful for the engineering of stress tolerant oil palm.

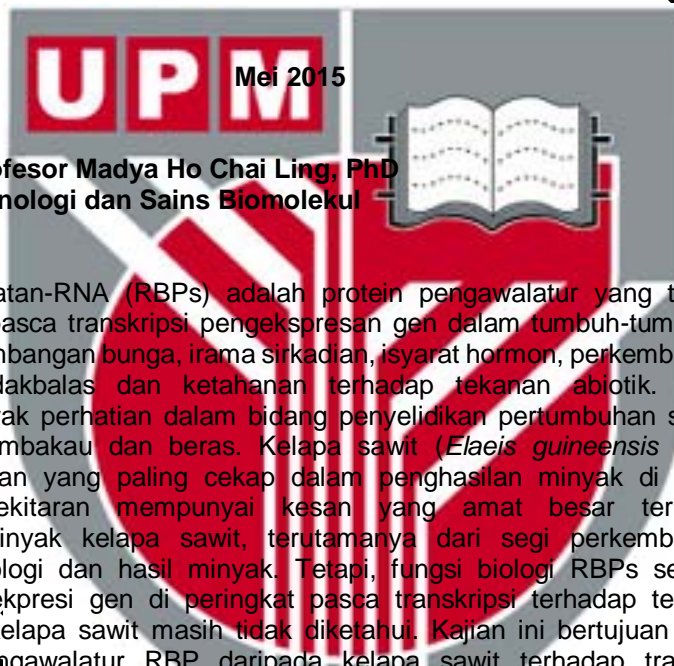


Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PENGLONAN DAN PENCIRIAN TRANSKRIP NOVEL YANG
MENGEKODKAN PROTEIN PENGIKATAN-RNA DARIPADA KELAPA SAWIT
(*Elaeis guineensis* Jacq.)**

Oleh

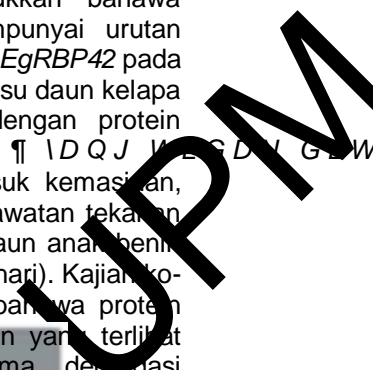
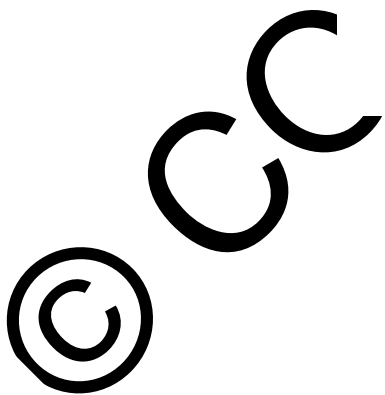
YEAP WAN CHIN



**Pengerusi: Profesor Madya Ho Chai Ling, PhD
Fakulti: Bioteknologi dan Sains Biomolekul**

Protein pengikatan-RNA (RBPs) adalah protein pengawalatur yang terlibat dalam proses pasca transkripsi pengekspresan gen dalam tumbuh-tumbuhan dari segi perkembangan bunga, irama sirkadian, isyarat hormon, perkembangan tumbuhan, tindakbalas dan ketahanan terhadap tekanan abiotik. RBPs mendapat banyak perhatian dalam bidang penyelidikan pertumbuhan seperti *Arabidopsis*, tembakau dan beras. Kelapa sawit (*Elaeis guineensis* Jacq.) adalah tumbuhan yang paling cekap dalam penghasilan minyak di dunia. Tekanan persekitaran mempunyai kesan yang amat besar terhadap penghasilan minyak kelapa sawit, terutamanya dari segi perkembangan tumbuhan, fisiologi dan hasil minyak. Tetapi, fungsi biologi RBPs sebagai pengawalatur ekspresi gen di peringkat pasca transkripsi terhadap tekanan abiotik pokok kelapa sawit masih tidak diketahui. Kajian ini bertujuan untuk memahami pengawalatur RBP daripada kelapa sawit terhadap transkrip sasaran di peringkat pasca transkripsi, faktor-faktor pengawalatur yang berkaitan dengan RBP dalam kompleks ribonucleoprotein dan juga penglibatan RBP dalam mekanisme RNA di peringkat pasca transkripsi dan tindakbalasnya terhadap rangsangan alam sekitar di dalam kelapa sawit. Dalam kajian ini, gen yang dinamakan sebagai *EgRBP42*, mengekodkan ribonucleoprotein nuklear heterogen telah dipencilkan dan diklonkan daripada kelapa sawit. *EgRBP42* didapati daripada tag jujukan terekspres (EL684239) bunga betina kelapa sawit. *EgRBP42* mengandungi dua motif pengenalan RNA di pangkalan-N dan satu domain kaya dengan glisin di pangkalan-C. Rantau penganjur *EgRBP42* mengandungi pelbagai elemen pengawalaturan yang berkaitan dengan tindakbalas terhadap cahaya, tindakbalas terhadap tekanan dan perkembangan bunga. Analisis RT-PCR masa nyata menunjukkan bahawa *EgRBP42* digekspres dalam semua tisu-tisu kelapa sawit yang diuji, termasuk daun, pucuk meristem apikal, akar, bunga betina, bunga jantan dan mesokarp dengan aras transkrip yang paling rendah di dalam akar. Protein *EgRBP42* berinteraksi

dengan transkrip yang berkaitan dengan proses tindakbalas terhadap tekanan, transkripsi dan terjemahan protein. Pengesahan jujukan konsensus pada transkrip berinteraktif kepada protein EgRBP42 menunjukkan bahawa EgRBP42 terikat kepada transkrip berinteraktif yang mempunyai urutan nukleotida yang kaya dengan adenine dan guanine. Tiga varian *EgRBP42* pada ORNDVL ¶ \DQJ anak dan anak benih kelapa pada tisu daun kelapa sawit. Ekspresi *EgRBP42*, transkripsi yang berinteraksi dengan protein EgRBP42 dan transkripsi varian *EgRBP42* GLORNDVL ¶ \DQJ ¶ \SD ¶ \G ¶ W ditingkatkan (> 2 kali ganda) oleh tekanan abiotik, termasuk kemasinan, kemarau, banjir, kesejukan dan kepanasan di cakera daun (rawatan tekanan jangka masa pendek selama 30 min sehingga 28 jam) dan daun anak benih kelapa sawit (rawatan tekanan jangka masa panjang selama 7 hari). Kajian ko-pengendapan imun dan interaksi yis II hibrid menunjukkan bahawa protein EgRBP42 berinteraksi dengan pelbagai faktor pengawalaturan yang terlibat dalam tindakan transkripsi, pengangkutan nukleus-sitoplasma, degradasi mRNA dan terjemahan mRNA. Ekspresi protein EgRBP42 telah ditingkatkan (> 2 kali ganda) dalam anak benih kelapa sawit yang terdedah kepada kepanasan, kesejukan, kemarau dan kemasinan dalam rawatan tekanan jangka masa panjang selama 7 hari. Secara keseluruhan, data yang diperolehi mencadangkan bahawa protein EgRBP42 bertindakbalas terhadap pelbagai tekanan abiotik dan ia berpotensi sebagai pengangkut nukleus-sitoplasma kepada mRNA yang responsif terhadap tekanan alam sekitar supaya penerjemahan cepat akan dijalankan sebagai tindakbalas terhadap kepanasan, kesejukan dan kemarau. Kajian ini memberi maklumat berguna mengenai mekanisme pasca transkripsi yang dikawalatur oleh EgRBP42 di dalam kelapa sawit. Oleh itu, EgRBP42 mungkin memainkan peranan yang penting dalam kejuruteraan genetik kelapa sawit untuk meningkatkan tahap ketahanan terhadap tekanan persekitaran.



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I certify that a Thesis Examination Committee has met on 21 May 2015 to conduct the final examination of Yeap Wan Chin on her thesis *H Q W L Wong* ³ and Characterization of a Novel Transcript Encoding a RNA-Binding Protein from Oil Palm (*Elaeis guineensis* Jacq.) in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

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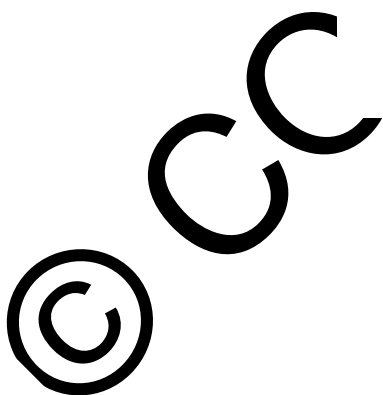
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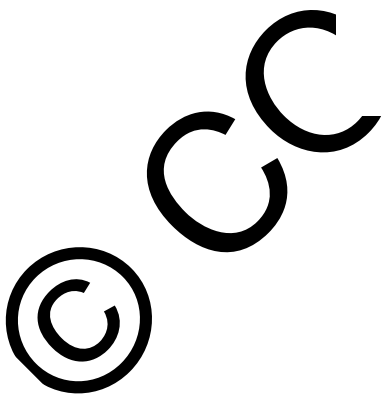
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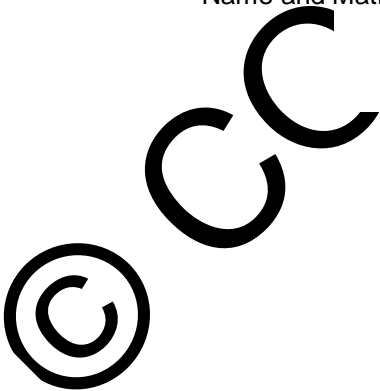
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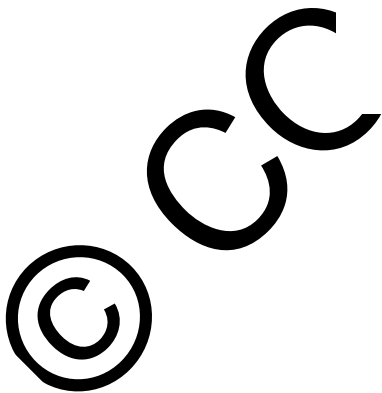
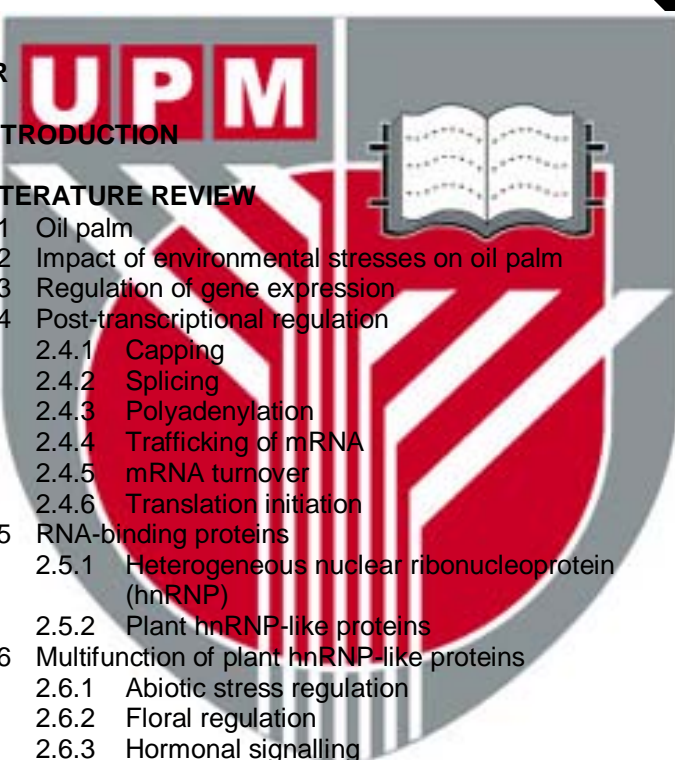


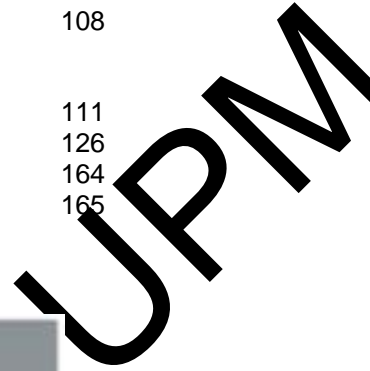
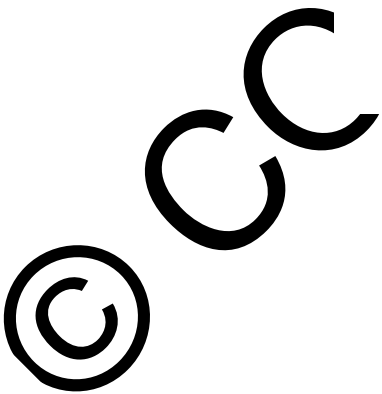
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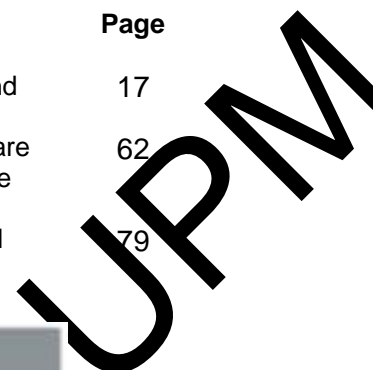
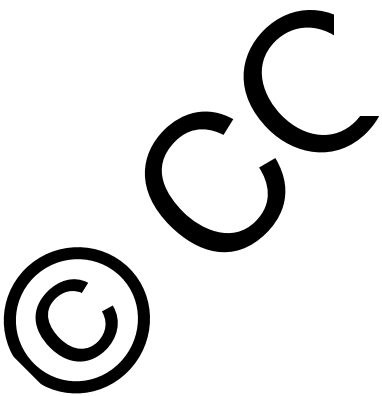
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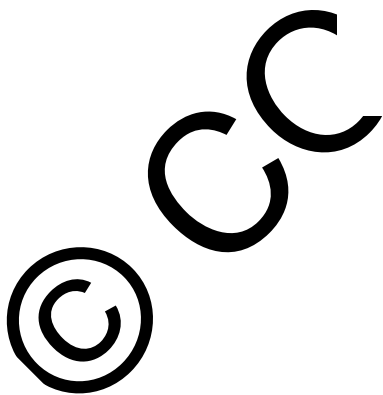
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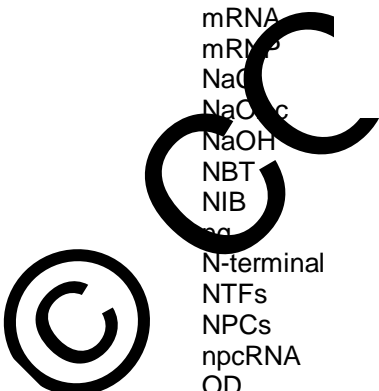
LIST OF ABBREVIATIONS

.	alpha
	beta
	gamma
/	delta
%	percentage
°C	degree Centigrade
AAPK	ABA-activated protein kinase
AbA	Aureobasidin A
ABA	Abscisic acid
AD	activation domain
Ade	adenine
AG	AGAMOUS
AKIP1	AAPK-interacting protein 1
ARE	adenylate/uridylate-rich element
ATP	adenosine triphosphate
AtTRN1	transportin 1
BCIP	5-bromo-4-chloro-3-indolyl-phosphate
bp	base pair
BSA	bovine serum albumin
CaCl ₂	calcium chloride
CBC	cap-binding complex
C-terminal	carboxyl terminal
cDNA	complementary DNA
cm	centimetre
CPL1	C-terminal domain phosphatase-like 1
cpRNP	chloroplastid ribonucleoproteins
CPSF	cleavage and polyadenylation specific factor
DNA	deoxyribonucleic acid
DNase I	deoxyribonuclease
dNTP	deoxynucleotides
ds-cDNA	double stranded cDNA
DEPC	diethyl pyrocarbonate
DMSO	dimethylsulphonyl oxide
DTT	dithiothreitol
EtBr	ethidium bromide
EDTA	ethylenediaminetetraacetic acid
EGTA	ethylene glycol bis-(2-aminoethyle ether)
ELRBP42	<i>Elaeis guineensis</i> RNA-binding protein 42 kDa
eIF	translation initiation factors
EJC	exon junction complex
EMSA	electrophoretic mobility shift assay
FCA	flowering control locus A
FLC	FLOWERING LOCUS C
FLK	flowering locus K
FPA	flowering time control protein A
FY	flowering locus Y
GA	giberrellin acid

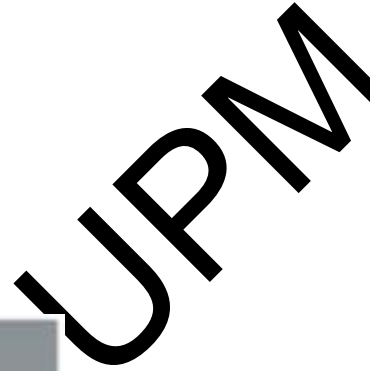
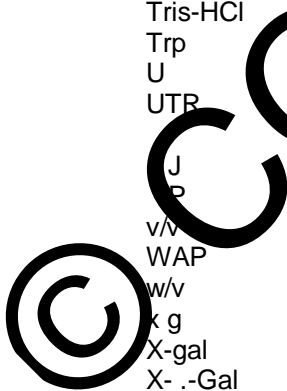
g	gram
GR-RBP	glycine-rich RNA-binding protein
GST-His6	glutathione-S transferase-hexa histadine
HCl	hydrochloric acid
His	histidine
HEPES	N-2-hydroxyethylpiperazine-N' -2-ethanesulfonic acid
hr	hours
hnRNA	heterogeneous nuclear ribonucleic acid
hnRNP	heterogeneous nuclear ribonucleoprotein
IPTG	isopropyl- β -D-thioga1actoside
k	kilo
kb	kilo base pair
KCl	potassium chloride
kDa	kilot Dalton
KH	K homology
KOAc	potassium acetate
L	liter
Leu	leucine
LHP1	LIKE HETEROCHROMATIN PROTEIN1
LIF2	LHP1 Interacting Factor 2
LiCl	lithium chloride
M	molar
M9	M9 nucleocytoplasmic export signal
MBE	Musashi binding element
mg	milligram
MgCl ₂	magnesium chloride
MgSO ₄	magnesium sulphate
min	minutes
miRNA	microribonucleic acid
mL	millilitre
/	microlitre
mm	millimeter
mM	millimolar
MOPS	3-(N-morpholino) propane-sulphonic acid
mRNA	messenger ribonucleic acid
mRNP	messenger ribonucleoprotein
NaCl	sodium chloride
NaOAc	sodium acetate
NaOH	sodium hydroxide
NBT	nitro blue tetrazolium
NIB	nuclear isolation buffer
ng	nanogram
N-terminal	amino terminal
NTFs	nuclear transport factors
NPCs	nuclear pore complexes
npcRNA	non-protein coding ribonucleic acid
OD	optical density
ORF	open reading frame
P bodies	processing bodies
PABP	polyadenylation-binding protein



UPM



PAGE	polyacrylamide gel electrophoresis
PABPII	polyadenylation binding protein II
PAP	polyadenylation polymerase
PBS	phosphate buffer saline
PCR	polymerase chain reaction
pI	isoelectric point
PMSF	phenylmethyl-sufonyl fluoride
poly (A)	polyadenylation
pre-mRNA	precursor messenger ribonucleic acid
PTB	polypyrimidine-tract binding
PVDF	polyvinylidene difluoride
PVPP	polypolyvinylpyrrolidone
qRT-PCR	quantitative real-time RT-PCR
RBD	RNA-binding domain
RBP	RNA-binding protein
RGG	arginine-glycine-glycine
RNA	ribonucleic acid
rRNA	ribosomal ribonucleic acid
RNAPII	RNA polymerase II
RNase	ribonuclease
RNP	ribonucleoprotein
rpm	revolutions per minute
RRM	RNA recognition motif
RT	reverse transcriptase
sec	second
SD	synthetic dropout
SDS	sodium dodecyl sulphate
snRNP	small ribonucleoprotein particles
SR	serine/arginine rich
TAE	tris acetate EDTA
TCA	trichloro-acetic acid
TE	tris-EDTA
TEMED	,N,N,N',N'-tetramethylethylenediamine
Tris	tris[hydroxymethyl]aminomethane
Tris-HCl	tris[hydroxymethyl]aminomethane hydrochloride
Trp	tryptophan
U	unit
UTR	untranslated region
µl	microliter
µg	microgram
µm	micrometer
v/v	volume per volume
WAP	weeks after pollination
w/v	weight per volume
x g	relative centrifugal force
X-gal	5-bromo-4-chloro-3-indolyl-β-D-galactopyronoside
X-α-Gal	5-bromo-4-chloro-3-indolyl α-D-galactopyranoside



CHAPTER 1

INTRODUCTION

Gene expression during growth and development is governed by both transcription and post-transcription regulation of mRNAs. Transcriptional regulation affects the expression of genes. However, the discordance between the mRNA and protein levels in eukaryotes is mainly due to post-transcriptional processing and regulation. This regulation can be achieved either directly by RNA binding proteins (RBPs) or indirectly via modulation of other regulatory factors in eukaryotes (Lorkovic, 2009). Gene encoding RBPs with RNA recognition motifs (RRMs) have received more attention in plant research recently. These RBPs are emerging as multifunctional cellular regulatory proteins involved in RNA metabolism including the regulation of transcriptional processes such as RNA synthesis, pre-mRNA splicing, capping, polyadenylation, exporting RNA from nucleus, pre-rRNA complex formation, mRNA stability and degradation. Besides that, RBPs participate in all aspects of translational processes whereby they regulate translation of functional mRNAs and storage of non-translated mRNAs. Some RBPs are also involved in chromosome structuring such as telomere maintenance that is important for chromosome stability and integrity (Chen and Varani, 2005; Glisovic *et al.*, 2008).

Functional studies and RNA sequencing of plant RBPs clearly showed that a family of RBPs that are defined as heterogeneous nuclear ribonucleoprotein (hnRNP)-like proteins are also expressed in higher plants and serve specific plant functions (Lambermon *et al.*, 2000; Lorkovic *et al.*, 2000). In plants, these hnRNP-like proteins have been reported to influence floral induction and development, circadian rhythms, hormone signaling, differentiation, chloroplast regulation, phloem translocation, stress response and stress tolerance (Nakamura *et al.*, 1999; Li *et al.*, 2002; Macknight *et al.*, 2002; Quesada *et al.*, 2003; Simpson *et al.*, 2003; Razem *et al.*, 2006; Staiger *et al.*, 2003; Ham *et al.*, 2009; Tillich *et al.*, 2009). Post-transcriptional gene regulation plays a crucial role in the response of plants towards stress stimulus. Regulatory factors such as small RNAs and RBPs have emerged as regulators of RNA mechanism in all aspects of post-transcriptional gene regulation in plant stress responses, tolerance and adaptation. RBPs respond to stress signals through the regulation of downstream stress-related genes expression and enhance tolerance of plants towards various abiotic adversities (Lee *et al.*, 2009).

Oil palm (*Elaeis guineensis* Jacq.) is the most efficient oil-yielding crop in the world. Oil palm produces 32% of global oils and fats and utilizes the least global land area for cultivation (5.5%) compared to other oilseed crops (Oil world 2013). One hectare of oil palm plantation produces up to ten times more oil (average oil yield of ~4 tonnes of oil per hectare) than other oilseed crops (Oil world 2013). Palm oil is the most consumed oil among 17 major oils and fats traded globally

and the global consumption for palm oil was 52.1 million tonnes in 2012 (Oil world 2013). The palm oil industry is one of the key economic drivers of the agricultural sector in Malaysia. Malaysia is the second largest palm oil producer utilizing 5.1 million hectares of land for oil palm cultivation, and accounted for 10% or 18.8 million tonnes of global vegetable oils and fats output in 2012 (Oil world 2013).

Environmental stresses have a major impact on oil palm production mainly plant growth, physiology and oil yield. Environmental stresses such as flood, drought, cold and heat stress have been reported to affect abortion of inflorescence, determination of inflorescence ratio and bunch ripening in matured oil palm (Corley and Donough, 1995; Gawankar *et al.*, 2003; Cha-Um *et al.*, 2010). Intermittent water stress (rainfed) reduces 91% of oil palm fresh fruit bunch production resulting in 88.46% reduction in the yield of fresh fruit bunches (Gawankar *et al.*, 2003). Under intermittent water stress, oil palm female inflorescence production is reduced by 86% while leaf production is reduced by 30% in the early growth phase and 12.5% in the later growth phase (Gawankar *et al.*, 2003).

The biological functions of RBPs in post-transcription regulation of gene expression in response to stresses are still poorly understood in oil palm. Hence, an in-depth functional analysis of RBPs mediated regulation of target RNA metabolisms will increase the understanding of the roles of RBPs in oil palm stress responses and adaptation that are vital for the engineering of stress-tolerant plant. The aim of this research was to understand the functional role of oil palm RBP and the network of gene regulation operating at the post-transcriptional level in response to environmental stimuli in oil palm. Through the study of messenger ribonucleoproteins (mRNPs) and the constituents of RNP complexes, networks of gene regulation and the underlying mechanism operating at the post-transcriptional level in response to environmental stimuli in oil palm can be elucidated.

The specific objectives of this study were:

1. To clone and isolate full length cDNA, variants, promoter and the gene encoding putative RBP (*EgRBP42*) from *Elaeis guineensis* Jacq.
2. To identify transcripts interacting with the *EgRBP42* protein and their consensus binding site.
3. To identify the regulatory factors associated with *EgRBP42* in the ribonucleoprotein complex and the functional roles of *EgRBP42* in post-transcriptional regulation.
4. To profile the protein accumulation and transcript abundance of *EgRBP42* and its variants under various abiotic stress conditions in oil palm.

REFERENCES

- Baurle, I., Smith, L., Baulcombe, D. C., & Dean, C. (2007). Widespread role for the flowering-time regulators FCA and FPA in RNA-mediated chromatin silencing. *Science*, 318(5847), 109-112.
- Blazquez, M. A., Green, R., Nilsson, O., Sussman, M. R., & Weigel, D. (1998). Gibberellins promote flowering of *Arabidopsis* by activating the L^{EAFY} promoter. *Plant Cell*, 10(5), 791-800.
- Bomszyk, K., Denisenko, O., & Ostrowski, J. (2004). hnRNP K: one protein multiple processes. *Bioessays*, 26(6), 629-638.
- Bove, J., Kim, C. Y., Gibson, C. A., & Assmann, S. M. (2008). Characterization of wound-responsive RNA-binding proteins and their splice variants in *Arabidopsis*. *Plant Molecular Biology*, 67(1-2), 71-88.
- Cartegni, L., Maconi, M., Morandi, E., Cobiainchi, F., Riva, S., & Biamonti, G. (1996). hnRNP A1 selectively interacts through its Gly-rich domain with different RNA-binding proteins. *Journal of Molecular Biology*, 259(3), 337-348.
- Chabot, B., Blanchette, M., Lapierre, I., & La Branche, H. (1997). An intron element modulating 5' splice site selection in the hnRNP A1 pre-mRNA interacts with hnRNP A1. *Molecular and Cellular Biology*, 17(4), 1776-1786.
- Charlesworth, A., Wilczynska, A., Thampi, P., Cox, L. L., & MacNicol, A. M. (2006). Musashi regulates the temporal order of mRNA translation during *Xenopus oocyte* maturation. *The EMBO Journal*, 25(12), 2792-2801.
- Chattopadhyay, S., Puente, P., Deng, X.-W., & Wei, N. (1998). Combinatorial interaction of light-responsive elements plays a critical role in determining the response characteristics of light-regulated promoters in *Arabidopsis*. *Plant Journal*, 15(1), 69-77.
- Chatum, S., Takabe, T., & Kirdmanee, C. (2013). Osmotic potential, photosynthetic abilities and growth characters of oil palm (*Elaeis guineensis* Jacq.) seedlings in responses to polyethylene glycol-induced water deficit. *African Journal of Biotechnology*, 9(39), 6509-6516.
- Chen, Y., & Varani, G. (2005). Protein families and RNA recognition. *FEBS Journal*, 272(9), 2088-2097.
- Cheng, S., & Gallie, D. R. (2007). eIF4G, eIFiso4G, and eIF4B bind the poly (A)-binding protein through overlapping sites within the RNA recognition motif domains. *Journal of Biological Chemistry*, 282(35), 25247-25258.
- Cheng, Y., & Chen, X. (2004). Posttranscriptional control of plant development. *Current Opinion in Plant Biology*, 7(1), 20-25.

Cheng, Y., Kato, N., Wang, W., Li, J., & Chen, X. (2003). Two RNA Binding Proteins, HEN4 and HUA1, Act in the Processing of *AGAMOUS* Pre-mRNA in *Arabidopsis thaliana*. *Developmental Cell*, 4(1), 53-66.

Cheong, Y. H., Chang, H.-S., Gupta, R., Wang, X., Zhu, T., & Luan, S. (2002). Transcriptional profiling reveals novel interactions between wounding, pathogen, abiotic stress, and hormonal responses in *Arabidopsis*. *Plant Physiology*, 129(2), 661-677.

Chien, C. T., Bartel, P. L., Sternglanz, R., & Fields, S. (1991). The two-hybrid system: a method to identify and clone genes for proteins that interact with a protein of interest. *Proceedings of the National Academy of Sciences of the United States of America*, 88(21), 9578-9582.

Chinnusamy, V., Gong, Z., & Zhu, J. K. (2008). *Nuclear RNA export and its importance in abiotic stress responses of plants: Nuclear pre-mRNA Processing in Plants* (pp. 235-255): Springer-Verlag Berlin-Heidelberg, Germany.

Colgan, D. F., & Manley, J. L. (1997). Mechanism and regulation of mRNA polyadenylation. *Genes & Development*, 11(21), 2755-2766.

Conesa, A., Götz, S., García-Gómez, J. M., Terol, J., Talón, M., & Robles, M. (2005). Blast2GO: a universal tool for annotation, visualization and analysis in functional genomics research. *Bioinformatics*, 21(18), 3674-3676.

Corley, R., Ng, M., & Donough, C. (1995). Effects of defoliation on sex differentiation in oil palm clones. *Experimental Agriculture*, 31(2), 177-190.

Corley, R.H.V. and Tinker, P.B. (2003). *The oil palm*; 4th edition; Blackwell Science Ltd.: Oxford UK.

Cowling, V. (2010). Regulation of mRNA cap methylation. *Biochemistry Journal*, 425, 295-302.

Crofts, A. J., Crofts, N., Whitelegge, J. P., & Okita, T. W. (2010). Isolation and identification of cytoskeleton-associated *prolamine* mRNA binding proteins from developing rice seeds. *Planta*, 231(6), 1261-1276.

Day, D. J., & Tuite, M. F. (1998). Post-transcriptional gene regulatory mechanisms in eukaryotes: an overview. *Journal of Endocrinology*, 157, 361-371.

Degenhardt, J., & Tobin, E. M. (1996). A DNA binding activity for one of two closely defined phytochrome regulatory elements in an *Lhcb* promoter is more abundant in etiolated than in green plants. *Plant Cell*, 8(1), 31-41.

De Lucia, F., & Dean, C. (2011). Long non-coding RNAs and chromatin regulation. *Current Opinion in Plant Biology*, 14(2), 168-173.

- Ding, Y. -F., & Zhu, C. (2009). The role of microRNAs in copper and cadmium homeostasis. *Biochemical and Biophysical Research Communications*, 386(1), 6-10.
- dit Frey, N. F., Muller, P., Jammes, F., Kizis, D., Leung, J., Perrot-Rechenmann, C., & Bianchi, M. W. (2010). The RNA binding protein Tudor-SN is essential for stress tolerance and stabilizes levels of stress-responsive mRNAs encoding secreted proteins in *Arabidopsis*. *Plant Cell*, 22(5), 1575-1590.
- Doroshenk, K. A., Crofts, A. J., Morris, R. T., Wyrick, J. J., & Okita, T. W. (2009). Proteomic analysis of cytoskeleton-associated RNA binding proteins in developing rice seed. *Journal of Proteome Research*, 8(10), 4641-4653.
- Dreyfuss, G., Matunis, M. J., Pinol-Roma, S., & Burd, C. (1993). hnRNP proteins and the biogenesis of mRNA. *Annual Review of Biochemistry*, 62(1), 289-321.
- Egea-Cortines, M., Saedler, H., & Sommer, H. (1999). Ternary complex formation between the MADS-box proteins SQUAMOSA, DEFICIENS and GLOBOSA is involved in the control of floral architecture in *Antirrhinum majus*. *The EMBO Journal*, 18(19), 5370-5379.
- Fujisaki, K., & Ishikawa, M. (2008). Identification of an *Arabidopsis thaliana* protein that binds to tomato mosaic virus genomic RNA and inhibits its multiplication. *Virology*, 380(2), 402-411.
- Garneau, N. L., Wilusz, J., & Wilusz, C. J. (2007). The highways and byways of mRNA decay. *Nature Reviews Molecular Cell Biology*, 8(2), 113-126.
- Gawankar, M., Devmore, J., Jamadagni, B., Sagvekar, V., & Khan, H. (2003). Effect of water stress on growth and yield of Tenera oil palm. *Journal of Applied Horticulture*, 5(1), 39-40.
- Gebauer, F., & Hentze, M. W. (2004). Molecular mechanisms of translational control. *Nature Reviews Molecular Cell Biology*, 5(10), 827-835.
- Giamartino, D. C. D., Nishida, K., & Manley, J. L. (2011). Mechanisms and consequences of alternative polyadenylation. *Molecular Cell*, 43, 853-866.
- Glisic, T., Bachorik, J. L., Yong, J., & Dreyfuss, G. (2008). RNA-binding proteins and post-transcriptional gene regulation. *FEBS Letters*, 582(14), 1977-1986.
- Gong, Z., Dong, C. -H., Lee, H., Zhu, J., Xiong, L., Gong, D., Stevenson, B., & Zhu, J. -K. (2005). A DEAD box RNA helicase is essential for mRNA export and important for development and stress responses in *Arabidopsis*. *Plant Cell*, 17(1), 256-267.

Gowik, U., Burscheidt, J., Akyildiz, M., Schlue, U., Koczor, M., Streubel, M., & Westhoff, P. (2004). *cis*-Regulatory elements for mesophyll-specific gene expression in the C4 plant *Flaveria trinervia*, the promoter of the C4 *phosphoenolpyruvate carboxylase* gene. *Plant Cell*, 16(5), 1077-1090.

Grewal, S. I. S & Jia, S. (2007). Heterochromatin revisited. *Nature Reviews Genetics*, 8(1), 35-46.

Gutierrez, R. A., Macintosh, G. C., & Green, P. J. (1999). Current perspectives on mRNA stability in plants: multiple levels and mechanisms of control. *Trends in Plant Science*, 4(11), 429-438.

Hall, T. A. (1999). *BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT*. Paper presented at the Nucleic acids symposium series.

Ham, B.-K., Brandom, J. L., Xoconostle-Cázares, B., Ringgold, V., Lough, T. J., & Lucas, W. J. (2009). A polypyrimidine tract binding protein, pumpkin RBP50, forms the basis of a phloem-mobile ribonucleoprotein complex. *Plant Cell*, 21(1), 197-215.

Han, J., Xiong, J., Wang, D., & Fu, X. -D. (2011). Pre-mRNA splicing: where and when in the nucleus. *Trends in Cell Biology*, 21(6), 336-343.

Hanano, S., Sugita, M., & Sugiura, M. (1996). Isolation of a novel RNA-binding protein and its association with a large ribonucleoprotein particle present in the nucleoplasm of tobacco cells. *Plant Molecular Biology*, 31(1), 57-68.

He, Y., & Amasino, R. M. (2005). Role of chromatin modification in flowering-time control. *Trends in Plant Science*, 10(1), 30-35.

Hellemans, J., Mortier, G., De Paepe, A., Speleman, F., & Vandesompele, J. (2007). qBase relative quantification framework and software for management and automated analysis of real-time quantitative PCR data. *Genome Biology*, 8(2), R19.

Hirose, T., & Sugiura, M. (2001). Involvement of a site specific trans-acting factor and a common RNA-binding protein in the editing of chloroplast mRNAs: development of a chloroplast in vitro RNA editing system. *The EMBO Journal*, 20(5), 1144-1152.

Hirano, Y., & Ohkuma, Y. (2007). Phosphorylation of the C-terminal domain of RNA polymerase II plays central roles in the integrated events of eucaryotic gene expression. *Journal of Biochemistry*, 141(5), 601-608.

Ho, C. L., Kwan, Y. Y., Choi, M. C., Tee, S. S., Ng, W. H., Lim, K. A., Lee, Y. P., Ooi, S. E., Lee, W. W., Tee, J. M., & Tan, S. H., (2007). Analysis and functional annotation of expressed sequence tags (ESTs) from multiple tissues of oil palm (*Elaeis guineensis* Jacq.). *BMC Genomics*, 8(1), 381.

Hutchison, S., LeBel, C., Blanchette, M., & Chabot, B. (2002). Distinct sets of adjacent heterogeneous nuclear ribonucleoprotein (hnRNP) A1/A2 binding VLWHV FRQWURO •VSOLFH VLWH VHOHFWLRQ LQ WKH KQ513 \$51\$SUHFUXVRU *Journal of Biological Chemistry*, 277(33), 29745-29752.

Isshiki, M., Tsumoto, A., & Shimamoto, K. (2006). The serine/arginine-rich protein family in rice plays important roles in constitutive and alternative splicing of pre-mRNA. *Plant Cell*, 18(1), 146-158.

Jain, M., Khurana, P., Tyagi, A. K., & Khurana, J. P. (2008). Genome-wide analysis of intronless genes in rice and *Arabidopsis*. *Functional & Integrative Genomics*, 8(1), 69-78.

Jedamzik, B., & Eckmann, C. R. (2009). Analysis of RNA-protein complexes by RNA coimmunoprecipitation and RT-PCR analysis from *Caenorhabditis elegans*. *Cold Spring Harbor Protocols*, 2009(10), pdb-prot5300.

Jiang, J., Wang, B., Shen, Y., Wang, H., Feng, Q., & Shi, H. (2013). The *Arabidopsis* RNA Binding Protein with K Homology Motifs, SHINY1, Interacts with the C-terminal Domain Phosphatase-like 1 (CPL1) to Repress Stress-Inducible Gene Expression. *PLoS Genetics*, 9(7), e1003625.

Jung, J.-H., Seo, P. J., Ahn, J. H., & Park, C. -M. (2012). *Arabidopsis* RNA-binding protein FCA regulates microRNA172 processing in thermosensory flowering. *Journal of Biological Chemistry*, 287(19), 16007-16016.

Kaneko, M., Inukai, Y., Ueguchi-Tanaka, M., Itoh, H., Izawa, T., Kobayashi, Y., Hattori, T., Miyao, A., Hirochika, H., Ashikari, M., & Matsuoka, M. (2004). Loss-of-function mutations of the rice *GAMYB* JHQH LPSDLU amylose expression in aleurone and flower development. *Plant Cell*, 16(1), 33-44.

Kawahara, H., Imai, T., Imataka, H., Tsujimoto, M., Matsumoto, K., & Okano, H. (2008). Neural RNA-binding protein Musashi1 inhibits translation initiation by competing with eIF4G for PABP. *The Journal of Cell Biology*, 181(4), 639-653.

Kedarsha, N. L., Gupta, M., Li, W., Miller, I., & Anderson, P. (1999). RNA-binding proteins TIA-1 and TIAR link the phosphorylation of eIF-4G to the formation of mammalian stress granules. *The Journal of Cell Biology*, 147(7), 1431-1442.

Koh, J., & Buhtz, A. (2008). Long distance transport and movement of RNA through the phloem. *Journal of Experimental Botany*, 59(1), 85-92.

Kiesler, E., Miralles, F., Farrants, A. K. O., & Visa, N. (2003). The Hrp65 self-interaction is mediated by an evolutionarily conserved domain and is required for nuclear import of Hrp65 isoforms that lack a nuclear localization signal. *Journal of Cell Science*, 116(19), 3949-3956.

Kim, C. Y., Bove, J., & Assmann, S. M. (2008a). Overexpression of wound-responsive RNA-binding proteins induces leaf senescence and hypersensitive-like cell death. *New Phytologist*, 180(1), 57-70.

Kim, H. J., Lee, J. Y., Yoon, U. H., Lim, S. H., & Kim, Y. M. (2013). Effects of Reduced Prolamin on Seed Storage Protein Composition and the Nutritional Quality of Rice. *International Journal of Molecular Sciences*, 14(8), 17073-17084.

Kim, J. S., Jung, H. J., Lee, H. J., Kim, K., Goh, C. H., Woo, Y., Oh, S. H., Han, Y. S., & Kang, H. (2008b). Glycine-rich RNA-binding protein 7 affects abiotic stress responses by regulating stomata opening and closing in *Arabidopsis thaliana*. *The Plant Journal*, 55(3), 455-466.

Kim, J. Y., Kim, W. Y., Kwak, K. J., Oh, S. H., Han, Y. S., & Kang, H. (2010). Zinc finger-containing glycine-rich RNA-binding protein in *Oryza sativa* has an RNA chaperone activity under cold stress conditions. *Plant, Cell & Environment*, 33(5), 759-768.

Kim, J. Y., Park, S. J., Jang, B., Jung, C. H., Ahn, S. J., Goh, C. H., Cho, K., Han, O., & Kang, H. (2007a). Functional characterization of a glycine-rich RNA-binding protein 2 in *Arabidopsis thaliana* under abiotic stress conditions. *The Plant Journal*, 50(3), 439-451.

Kim, Y. O., Pan, S., Jung, C. H., & Kang, H. (2007). A zinc finger-containing glycine-rich RNA-binding protein, atRZ-1a, has a negative impact on seed germination and seedling growth of *Arabidopsis thaliana* under salt or drought stress conditions. *Plant and Cell Physiology*, 48(8), 1170-1181.

Kosturko, L. D., Maggipinto, M. J., Korza, G., Lee, J. W., Carson, J. H., & Barbarese, E. (2006). Heterogeneous nuclear ribonucleoprotein (hnRNP) E1 binds to hnRNP A2 and inhibits translation of A2 response element mRNAs. *Molecular Biology of the Cell*, 17(8), 3521-3533.

Kraut-Cohen, J., & Gerst, J. E. (2010). Review: Addressing mRNAs to the ER: cis sequences act up! *Trends in Biochemical Sciences*, 35, 459-469.

Krecic, A. M., & Swanson, M. S. (1999). hnRNP complexes: composition, structure, and function. *Current Opinion in Cell Biology*, 11(3), 363-371.

Kumar, S., Jiang, S., Jami, S.K., & Hill, R.D. (2011). Cloning and characterization of honey caryopsis FCA. *Physiologia Plantarum*, 143(1), 93-106.

Kupsch, C., Ruwe, H., Gusewski, S., Tillich, M., Small, I., & Schmitz-Linneweber, C. (2012). *Arabidopsis* chloroplast RNA binding proteins CP31A and CP29A associate with large transcript pools and confer cold stress tolerance by influencing multiple chloroplast RNA processing steps. *Plant Cell*, 24(10), 4266-4280.

Kwak, K. J., Park, S. J., Han, J. H., Kim, M. K., Oh, S. H., Han, Y. S., & Kang, H. (2011). Structural determinants crucial to the RNA chaperone activity of glycine-rich RNA-binding proteins 4 and 7 in *Arabidopsis thaliana* during the cold adaptation process. *Journal of Experimental Botany*, 62(11), 4003-4011.

Kwon, C., & Chung, I. K. (2004). Interaction of an *Arabidopsis* RNA-binding protein with plant single-stranded telomeric DNA modulates telomerase activity. *Journal of Biological Chemistry*, 279(13), 12812-12818.

Lambermon, M. H., Fu, Y., Kirk, D. A. W., Dupasquier, M., Filipowicz, W., & Simpson, G. G. (2000). UBP1, a multifunctional effector of pre-mRNA maturation in plants. *Molecular and Cellular Biology*, 22(12), 4346-4357.

Lambermon, M. H., Simpson, G. G., Wieczorek Kirk, D. A., Hemmings-Mieszczak, M., Klahre, U., & Filipowicz, W. (2000). UBP1, a novel hnRNP-like protein that functions at multiple steps of higher plant nuclear pre-mRNA maturation. *The EMBO Journal*, 19(7), 1638-1649.

Laporte, P., Merchan, F., Amor, B., Wirth, S., & Crespi, M. (2007). Riboregulators in plant development. *Biochemical Society Transactions*, 35(6), 1638-1642.

Latiff, A. 2000. The biology of the genus *Elaeis*. In Y. Basiron, B. S. Jalani and K. W. Chan Eds., *Advances in Oil Research Vol. 1* (pp. 19-38). Malaysia: Malaysian Palm Oil Board MPOB.

Latrasse, D., Germann, S., Houba-Hérin, N., Dubois, E., Bui-Prodhomme, D., Hourcade, D., Juul-Jensen, T., Le Roux, C., Majira, A., Simoncello, N., Granier F., Taconnat, L., Renou, J., & Gaudin, V. (2011). Control of flowering and cell fate by LIF2, an RNA binding partner of the polycomb complex component LHP1. *PLoS One*, 6(1), e16592.

Lee, J. H., Cho, Y. S., Yoon, H. S., Suh, M. C., Moon, J., Lee, I., Weigel, D., Jun, C. H., & Kim, J. K. (2005). Conservation and divergence of FCA function between *Arabidopsis* and rice. *Plant Molecular Biology*, 58(6), 823-838.

Lee, Y. W., Kim, K. P., Kim, B.-G., Hahn, J. S., & Hong, C. B. (2009). Flooding stress-induced glycine-rich RNA-binding protein from *Nicotiana tabacum*. *Molecules and Cells*, 27(1), 47-54.

Lee, Y. W., & Kim, W. T. (2010). Tobacco GTBP1, a homolog of human heterogeneous nuclear ribonucleoprotein, protects telomeres from aberrant homologous recombination. *Plant Cell*, 22(8), 2781-2795.

Lee, Y. W., & Kim, W. T. (2011). Roles of NtGTBP1 in telomere stability. *Plant Signaling & Behavior*, 6(4), 523-525.

Lee, Y. W., & Kim, W. T. (2013). Telomerase-Dependent 3' G-Strand Overhang Maintenance Facilitates GTBP1-Mediated Telomere Protection from Misplaced Homologous Recombination. *Plant Cell*, 25(4), 1329-1342.

Legros, S., Mialet-Serra, I., Caliman, J. P., Siregar, F., Clement-Vidal, A., Fabre, D., & Dingkuhn, M. (2009). Phenology, growth and physiological adjustments of oil palm (*Elaeis guineensis*) to sink limitation induced by fruit pruning. *Annals of Botany*, 104(6), 1183-1194.

Lei, H., Zhai, B., Yin, S., Gygi, S., & Reed, R. (2013). Evidence that a consensus element found in naturally intronless mRNAs promotes mRNA export. *Nucleic Acids Research*, 41(4), 2517-2525.

Li, J., Kinoshita, T., Pandey, S., Ng, C. K. Y., Gygi, S. P., Shimazaki, K., & Assmann, S. M. (2002). Modulation of an RNA-binding protein by abscisic acid-activated protein kinase. *Nature*, 418(6899), 793-797.

Li, P., Ham, B. -K., & Lucas, W. J. (2011). CmRBP50 protein phosphorylation is essential for assembly of a stable phloem-mobile high-affinity ribonucleoprotein complex. *Journal of Biological Chemistry*, 286(26), 23142-23149.

Li, W. F., Zhang, S. G., Han, S. Y., Wu, T., Zhang, J. H., & Qi, L. W. (2013). Regulation of *LaMYB33* by miR159 during maintenance of embryogenic potential and somatic embryo maturation in *Larix kaempferi* (Lamb.) Carr. *Plant Cell, Tissue and Organ Culture*, 113(1), 131-136.

Lim, M. H., Kim, J., Kim, Y. S., Chung, K. S., Seo, Y. H., Lee, I., Kim, J., Hong, C. B., Kim, H. J., & Park, C.M. (2004). A new *Arabidopsis* gene, *FLK*, encodes an RNA binding protein with K homology motifs and regulates flowering time via *FLOWERING LOCUS C*. *Plant Cell*, 16(3), 731-740.

Lindlöf, A., Bräutigam, M., Chawade, A., Olsson, O., & Olsson, B. (2009). *In silico* analysis of promoter regions from cold-induced genes in rice (*Oryza sativa* L.) and *Arabidopsis thaliana* reveals the importance of combinatorial control. *Bioinformatics*, 25(11), 1345-1348.

Lipman, D. J., Souvorov, A., Koonin, E. V., Panchenko, A. R., & Tatusova, T. A. (2002). The relationship of protein conservation and sequence length. *BMC Evolutionary Biology*, 2(1), 20.

/R/RYLü=-5ROHRISODQW51\$ -binding proteins in development, stress response and genome organization. *Trends in Plant Science*, 14(4), 229-236.

/R/RUNRYLü=- %DUWD \$HQRPH DQDOVLV 51\$JHFRJQLWLRQ PRWLI (RRM) and K homology (KH) domain RNA-binding proteins from the flowering plant *Arabidopsis thaliana*. *Nucleic Acids Research*, 30(3), 623-635.

- ~~REMOVED~~ -Mieszczak, M., & Filipowicz, W. (2000). RBP45 and RBP47, two oligouridylylate-specific hnRNP-like proteins interacting with poly (A)+ RNA in nuclei of plant cells. *RNA*, 6(11), 1610-1624.
- Lohr, B., Streitner, C., Steffen, A., Lange, T., & Staiger, D. (2014). A glycine-rich RNA-binding protein affects gibberellin biosynthesis in *Arabidopsis*. *Molecular Biology Reports*, 41(1), 439-445.
- Lummer, M., Humpert, F., Steuwe, C., Caesar, K., Schuttpelz, M., Sauer, M., & Staiger, D. (2011). Reversible photoswitchable DRONPA-s monitor nucleocytoplasmic transport of an RNA-binding protein in transgenic plants. *Traffic*, 12(6), 693-702.
- Ma, A. S., Moran-Jones, K., Shan, J., Munro, T. P., Snee, M. J., Hoek, R. G., & Smith, R. (2002). Heterogeneous nuclear ribonucleoprotein A3, a novel RNA trafficking response element-binding protein. *Journal of Biological Chemistry*, 277(20), 18010-18020.
- Macknight, R., Duroux, M., Laurie, R., Dijkwel, P., Simpson, G., & Dean, C. (2002). Functional significance of the alternative transcript processing of the *Arabidopsis* floral promoter *FCA*. *Plant Cell*, 14(4), 877-888.
- MacNicol, M. C., Cragle, C. E., & MacNicol, A. M. (2011). Context-dependent regulation of Musashi-mediated mRNA translation and cell cycle regulation. *Cell Cycle*, 10(1), 39.
- Manzano, D., Marquardt, S., Jones, A. M., Bäurle, I., Liu, F., & Dean, C. (2009). Altered interactions within FY/AtCPSF complexes required for *Arabidopsis* FCA-mediated chromatin silencing. *Proceedings of the National Academy of Sciences of the United States of America*, 106(21), 8772-8777.
- Maxwell, D. P., Wang, Y., & McIntosh, L. (1999). The alternative oxidase lowers mitochondrial reactive oxygen production in plant cells. *Proceedings of the National Academy of Sciences of the United States of America*, 96(14), 8271-8276.
- Mayada, A., Munroe, S., Caceres, J., & Krainer, A. (1994). Function of conserved domains of hnRNP A1 and other hnRNP A/B proteins. *The EMBO Journal*, 13(22), 5483.
- Mazumder, B., Sampath, P., Seshadri, V., Maitra, R. K., DiCorleto, P. E., & Fox, P. L. (2003). Regulated release of L13a from the 60S ribosomal subunit as a mechanism of transcript-specific translational control. *Cell*, 115(2), 187-198.
- Mazumder, B., Seshadri, V., Imataka, H., Sonenberg, N., & Fox, P. L. (2001). Translational silencing of ceruloplasmin requires the essential elements of mRNA circularization: poly (A) tail, poly (A)-binding protein, and eukaryotic

translation initiation factor 4G. *Molecular and Cellular Biology*, 21(19), 6440-6449.

Mazzucotelli, E., Mastrangelo, A. M., Crosatti, C., Guerra, D., Stanca, A. M., & Cattivelli, L. (2008). Abiotic stress response in plants: When post-transcriptional and post-translational regulations control transcription. *Plant Science*, 174(4), 420-431.

McCarthy, D. J., & Smyth, G. K. (2009). Testing significance relative to a fold change threshold is a TREAT. *Bioinformatics*, 25, 765-71.

McKee, A. E., & Silver, P. A. (2007). Systems perspectives on mRNA processing. *Cell Research*, 17, 581-590.

Meier, I., & Somers, D. E. (2011). Regulation of nucleocytoplasmic trafficking in plants. *Current Opinion in Plant Biology*, 14(5), 538-546.

Minvielle-Sebastia, L., & Keller, W. (1999). mRNA polyadenylation and its coupling to other RNA processing reactions and to transcription. *Current Opinion in Cell Biology*, 11(3), 352-357.

Mitchell, P., & Tollervey, D. (2000). mRNA stability in eukaryotes. *Current Opinion in Genetics & Development*, 10(2), 193-198.

Mockler, T. C., Yu, X., Shalitin, D., Parikh, D., Michael, T. P., Liou, J., Huang, J., Smith, Z., Alonso, J. M., Ecker, J. R., Chory, J., & Lin, C. (2004). Regulation of flowering time in *Arabidopsis* by K homology domain proteins. *Proceedings of the National Academy of Sciences of the United States of America*, 101(34), 12759-12764.

Moran-Jones, K., Wayman, L., Kennedy, D. D., Reddel, R. R., Sara, S., Snee, M. J., & Smith, R. (2005). hnRNP A2, a potential ssDNA/RNA molecular adapter at the telomere. *Nucleic Acids Research*, 33(2), 486-496.

Morris, R. T., Dproshenk, K. A., Crofts, A. J., Lewis, N., Okita, T. W., & Wyrick, J. J. (2011). RiceRBP: A database of experimentally identified RNA-binding proteins in *Oryza sativa* L. *Plant Science*, 180(2), 204-211.

Manro, T. P., Magee, R. J., Kidd, G. J., Carson, J. H., Barbarese, E., Smith, L. M., & Smith, R. (1999). Mutational analysis of a heterogeneous nuclear ribonucleoprotein A2 response element for RNA trafficking. *Journal of Biological Chemistry*, 274(48), 34389-34395.

Murray, M., & Thompson, W. F. (1980). Rapid isolation of high molecular weight plant DNA. *Nucleic Acids Research*, 8(19), 4321-4326.

Nakaminami, K., Matsui, A., Shinozaki, K., & Seki, M. (2012). RNA regulation in plant abiotic stress responses. *Biochimica et Biophysica Acta*, 1819(2), 149-153.

Nakamura, T., Ohta, M., Sugiura, M., & Sugita, M. (1999). Chloroplast ribonucleoproteins are associated with both mRNAs and intron-containing precursor tRNAs. *FEBS Letters*, 460(3), 437-441.

Ogbourne, S., & Antalis, T. M. (1998). Transcriptional control and the role of silencers in transcriptional regulation in eukaryotes. *Biochemical Journal*, 331(1), 1-14.

Oil World (2013). *Oil World Annual 2013*. Hamburg, ISTA Mielke GmbH.

Ouyang, S., Zhu, W., Hamilton, J., Lin, H., Campbell, M., Childs, K., Thibaud, Nissen, F., Malek, R. L., Zheng, L., Orvis, J., Haas, B., Wortman, J., & Bell, C. R. (2007). The TIGR rice genome annotation resource: improvements and new features. *Nucleic Acids Research*, 35(suppl 1), D883-D887.

Örvar, B. L., & Ellis, B. E. (1997). Transgenic tobacco plants expressing antisense RNA for cytosolic ascorbate peroxidase show increased susceptibility to ozone injury. *The Plant Journal*, 11(6), 1297-1305.

Percipalle, P., & Visa, N. (2006). Molecular functions of nuclear actin in transcription. *The Journal of Cell Biology*, 172(7), 967-971.

Quesada, V., Dean, C., & Simpson, G. G. (2005). Regulated RNA processing in the control of *Arabidopsis* flowering. *International Journal of Developmental Biology*, 49(5/6), 773.

Razem, F. A., El-Kereamy, A., Abrams, S. R., & Hill, R. D. (2006). The RNA-binding protein FCA is an abscisic acid receptor. *Nature*, 439(7074), 290-294.

Reddy, A. S. (2004). Plant serine/arginine-rich proteins and their role in pre-mRNA splicing. *Trends in Plant Science*, 9(11), 541-547.

Riera, M., Redko, Y., & Leung, J. (2006). *Arabidopsis* RNA-binding protein UP12a relocates into nuclear speckles in response to abscisic acid. *FEBS Letters*, 580(17), 4160-4165.

Ripoll, J. J., Ferrándiz, C., Martínez-Laborda, A., & Vera, A. (2006). *PEPPER*, a novel K-homology domain gene, regulates vegetative and gynoecium development in *Arabidopsis*. *Developmental Biology*, 289(2), 346-359.

Ripoll, J. J., Rodríguez-Cazorla, E., González-Reig, S., Andújar, A., Alonso-Cantabrana, H., Perez-Amador, M. A., Carbonell, J., Martínez-Laborda, A., & Vera, A. (2009). Antagonistic interactions between *Arabidopsis* K-homology domain genes uncover *PEPPER* as a positive regulator of the central floral repressor *FLOWERING LOCUS C*. *Developmental Biology*, 333(2), 251-262.

Rühl, C., Stauffer, E., Kahles, A., Wagner, G., Drechsel, G., Rättsch, G., & Wachter, A. (2012). Polypyrimidine tract binding protein homologs from

Arabidopsis are key regulators of alternative splicing with implications in fundamental developmental processes. *Plant Cell*, 24(11), 4360-4375.

Sanan-Mishra, N., Kumar, V., Sopory, S. K., & Mukherjee, S. K. (2009). Cloning and validation of novel miRNA from basmati rice indicates cross talk between abiotic and biotic stresses. *Molecular Genetics and Genomics*, 282(5), 463-474.

Sandberg, R., Neilson, J. R., Sarma, A., Sharp, P. A., & Burge, C. B. (2008). Proliferating cells express 515 ZLWKVKRUWHQHG KQWUDQVODWHG HLRQV and fewer microRNA target sites. *Science*, 320, 1643±1647.

Schomburg, F. M., Patton, D. A., Meinke, D. W., & Amasino, R. M. (2001). FPA, a gene involved in floral induction in *Arabidopsis*, encodes a protein containing RNA-recognition motifs. *Plant Cell*, 13(6), 1427-1436.

Schöning, J. C., Streitner, C., Meyer, I. M., Gao, Y., & Staiger, D. (2008). Reciprocal regulation of glycine-rich RNA-binding proteins via an interlocked feedback loop coupling alternative splicing to nonsense-mediated decay in *Arabidopsis*. *Nucleic Acids Research*, 36(22), 6977-6987.

Schöning, J. C., Streitner, C., Page, D. R., Hennig, S., Uchida, K., Wolf, E., Furuya, M., & Staiger, D. (2007). Auto-regulation of the circadian slave oscillator component AtGRP7 and regulation of its targets is impaired by a single RNA recognition motif point mutation. *The Plant Journal*, 52(6), 1119-1130.

Schuster, G., & Grissem, W. (1991). Chloroplast mRNA 3' end processing requires a nuclear-encoded RNA-binding protein. *The EMBO Journal*, 10(6), 1493.

Shah, S., Butler, N. M., Hannapel, D. J., & Rao, A. G. (2013). Mapping and Characterization of the Interaction Interface between Two Polypyrimidine-Tract Binding Proteins and a Nova-Type Protein of *Solanum tuberosum*. *PLoS One*, 8(5), e64783.

Simpson, G. G., Dijkwel, P. P., Quesada, V., Henderson, I., & Dean, C. (2003). FY Is an RNA 3' End-Processing Factor that Interacts with FCA to Control the *Arabidopsis* Floral Transition. *Cell*, 113(6), 777-787.

Sinclair, H., & Dreyfuss, G. (1995). A nuclear localization domain in the hnRNP A1 protein. *The Journal of Cell Biology*, 129(3), 551-560.

Smith, B., Williams, J., & Schulze-Kremer, S. (2003). *The ontology of the gene ontology*. Paper presented at the AMIA Annu Symp Proc.

Sorenson, R., & Bailey-Serres, J. (2014). Selective mRNA sequestration by OLIGOURIDYLATE-BINDING PROTIEN 1 contributes to translational

control during hypoxia in *Arabidopsis*, *Proceedings of the National Academy of Sciences of the United States of America*, 111, 2373±2378.

Staiger, D., & Green, R. (2011). RNA-based regulation in the plant circadian clock. *Trends in Plant Science*, 16(10), 517-523.

Staiger, D., Zecca, L., Kirk, D. A. W., Apel, K., & Eckstein, L. (2003). The circadian clock regulated RNA-binding protein AtGRP7 autoregulates its expression by influencing alternative splicing of its own pre-mRNA. *The Plant Journal*, 33(2), 361-371.

Stauffer, E., Westermann, A., Wagner, G., & Wachter, A. (2010). Polypyrimidine tract-binding protein homologues from *Arabidopsis* underlie regulatory circuits based on alternative splicing and downstream control. *The Plant Journal*, 64(2), 243-255.

Strambio-De-Castilla, C., Niepel, M., & Rout, M. P. (2010). The nuclear pore complex: bridging nuclear transport and gene regulation. *Nature Reviews Molecular Cell Biology*, 11(7), 490-501.

Streitner, C., Danisman, S., Wehrle, F., Schöning, J. C., Alfano, J. R., & Staiger, D. (2008). The small glycine-rich RNA binding protein AtGRP7 promotes floral transition in *Arabidopsis thaliana*. *The Plant Journal*, 56(2), 239-250.

Suárez-López, P., Wheatley, K., Robson, F., Onouchi, H., Valverde, F., & Coupland, G. (2001). CONSTANS mediates between the circadian clock and the control of flowering in *Arabidopsis*. *Nature*, 410(6832), 1116-1120.

Swindell, W. R., Huebner, M., & Weber, A. P. (2007). Transcriptional profiling of *Arabidopsis* heat shock proteins and transcription factors reveals extensive overlap between heat and non-heat stress response pathways. *BMC Genomics*, 8(1), 125.

Tamura, K., Dudley, J., Nei, M., & Kumar, S. (2007). MEGA4: molecular evolutionary genetics analysis (MEGA) software version 4.0. *Molecular Biology and Evolution*, 24(8), 1596-1599.

Terzigni, W. B., & Cashmore, A. R. (1995). Light-regulated transcription. *Annual Review of Plant Biology*, 46(1), 445-474.

Thompson, J. D., Higgins, D. G., & Gibson, T. J. (1994). CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Research*, 22(22), 4673-4680.

Tillich, M., Hardel, S. L., Kupsch, C., Armbruster, U., Delannoy, E., Gualberto, J. M., Lehwark, P., Leister, D., Small, I. D., & Schmitz-Linneweber, C. (2009). Chloroplast ribonucleoprotein CP31A is required for editing and stability of specific chloroplast mRNAs. *Proceedings of the National Academy of Sciences of the United States of America*, 106(14), 6002-6007.

Tourrière, H., Chebli, K., Zekri, L., Courselaud, B., Blanchard, J. M., Bertrand, E., & Tazi, J. (2003). The RasGAP-associated endoribonuclease G3BP assembles stress granules. *The Journal of Cell Biology*, 160(6), 823-831.

Tripet, B. P., Mason, K. E., Eilers, B. J., Burn, V. (3RZH003)LVFKHU0 & RSLp 9 6WUXFWXUDO DQG %LRFKHPLFDO QDOVLV RI *Hordeum vulgare* L. Hv GR-RBP1 Protein, a Glycine-Rich RNA-Binding Protein Involved in the Regulation of Barley Plant Development and Stress Response. *Biochemistry*, 53(50), 7945-7960.

Vandesompele, J., De Preter, K., Pattyn, F., Poppe, B., Van Roy, N., De Paepe, A., & Speleman, F. (2002). Accurate normalization of real-time quantitative RT-PCR data by geometric averaging of multiple internal control genes. *Genome Biology*, 3(7), research0034.

Wang, B. -B., & Brendel, V. (2004). The ASRG database: identification and survey of *Arabidopsis thaliana* genes involved in pre-mRNA splicing. *Genome Biology*, 5(12), R102.

Wang, S., & Okamoto, T. (2009). Involvement of polypyrimidine tract-binding protein (PTB)-related proteins in pollen germination in *Arabidopsis*. *Plant and Cell Physiology*, 50(2), 179-190.

Weber, C., Nover, L., & Fauth, M. (2008). Plant stress granules and mRNA processing bodies are distinct from heat stress granules. *The Plant Journal*, 56(4), 517-530.

Winicov, I. (2000). Alfin1 transcription factor overexpression enhances plant root growth under normal and saline conditions and improves salt tolerance in alfalfa. *Planta*, 210(3), 416-422.

Witzel, K., Weidner, A., Surabhi, G.-K., Börner, A., & Mock, H. -P. (2009). Salt stress-induced alterations in the root proteome of barley genotypes with contrasting response towards salinity. *Journal of Experimental Botany*, 60(14), 3545-3557.

Xiong, L., Lee, H., Ishitani, M., Tanaka, Y., Stevenson, B., Koiwa, H., Bressan, R. A., Ogasawara, P. M., & Zhu, J. -K. (2002). Repression of stress-responsive genes by FIERY2, a novel transcriptional regulator in *Arabidopsis*. *Proceedings of the National Academy of Sciences of the United States of America*, 99(16), 10899-10904.

Yang, D. H., Kwak, K. J., Kim, M. K., Park, S. J., Yang, K. Y., & Kang, H. (2014). Expression of *Arabidopsis* glycine-rich RNA-binding protein AtGRP2 or AtGRP7 improves grain yield of rice (*Oryza sativa*) under drought stress conditions. *Plant Science*, 214, 106-112.

Yeap, W. C., Loo, J. M., Wong, Y. C., & Kulaveerasingam, H. (2014). Evaluation of suitable reference genes for qRT-PCR gene expression normalization in

reproductive, vegetative tissues and during fruit development in oil palm. *Plant Cell, Tissue and Organ Culture*, 116(1), 55-66.

Yeap, W. C., Ooi, T. E. K., Namasivayam, P., Kulaveerasingam, H., & Ho, C. L. (2012). *EgRBP42* encoding an hnRNP-like RNA-binding protein from *Elaeis guineensis* Jacq. is responsive to abiotic stresses. *Plant Cell Reports*, 31(10), 1829-1843.

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Zhang, J., Zeng, R., Chen, J., Liu, X., & Liao, Q. (2008). Identification of conserved microRNAs and their targets from *Solanum lycopersicum* Mill. *Gene*, 423(1), 1-7.

Zhang, J., Zhao, Y., Xiao, H., Zheng, Y., & Yue, B. (2014). Genome-wide identification, evolution, and expression analysis of RNA-binding glycine-rich protein family in maize. *Journal of Integrative Plant Biology*, 56(10), 1020-1031.

Moore, C. (1999). Pta1, a component of yeast CF II, is required for both cleavage and poly (A) addition of mRNA precursor. *Molecular and Cellular Biology*, 19(11), 7733-7740.

Ziemienowicz, A., Haasen, D., Staiger, D., & Merkle, T. (2003). Arabidopsis transportin1 is the nuclear import receptor for the circadian clock-regulated RNA-binding protein AtGRP7. *Plant Molecular Biology*, 53(1-2), 201-212.

