

UNIVERSITI PUTRA MALAYSIA

IN VITRO ANTICANCER PROPERTIES OF LINAMARIN CONTROLLED RELEASE FROM BIODEGRADABLE POLY-LACTIC CO-GLYCOLIC ACID NANOPARTICLE

WEDAD ASHOUR AL FOURJANI.

FK 2005 12



IN VITRO ANTICANCER PROPERTIES OF LINAMARIN CONTROLLED RELEASE FROM BIODEGRADABLE POLY-LACTIC CO-GLYCOLIC ACID NANOPARTICLE

By

WEDAD ASHOUR AL FOURJANI

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia in Fulfilment of the Requirement for the Degree of Master of Science

November 2005



DEDICATIONS

To my husband and my son Abdo



Abstract of thesis presented to the Senate of University Putra Malaysia in fulfilment of the requirement for the degree of Master of Master of Science.

IN VITRO ANTICANCER PROPERTIES OF LINAMARIN CONTROLLED RELEASE FROM BIODEGRADABLE POLY-LACTIC CO-GLYCOLIC ACID NANOPARTICLE

By

WEDAD ASHOUR ALFOURJANI

November 2005

Chairman: Norhafizah Abdullah, PhD

Faculty : Engineering

There are many interests in finding new chemotherapeutic agents for cancer. The current work involved screening of linamarin as the therapeutic agent on different cancer cells, as no such study has been performed previously. Improved bioavailability and delivery of the linamarin to the targeted tumour cells can be engineered by proper selection of its carrier. There are many advantages of choosing biodegradable nanoparticles as a drug carrier. These include an improved bioavailability and efficacy of the drug. It also offers a controlled release mechanism in which the activity of the drug can be prolonged at the affected sites. Besides, the biodegradability character of the carrier means these particles are easily dissolved in the system without exerting any side effects to the body. The



present study investigated fabrication of linamarin encapsulation into biodegradable nanoparticles to kill cancer cells.

The present study was initiated with an investigation of the toxic effect of linamarin on cancer cells and their cell cycles. The *in vitro* study on the effect of linamarin was performed on two tumour cell lines, HeLa (cervical tumour cell line) and CAOV3 (ovarian tumour cell line). The cytotoxicity of linamarin was determined by the MTT assay. Both cell lines showed significant cell death when exposed to linamarin with the IC50 values well within the efficacious limit (IC50 of 30 mg/ml and 58 mg/ml for HeLa and CAOV3 cell lines, respectively, when exposed to pure linamarin). This result indicated that linamarin has the potential as a for drug candidate for cancer treatment. The subsequent cell cycle analysis performed by flow cytometry to determine the arrested point of linamarin within the cell cycle. Results showed significant effect of linamarin on the G1 phase. However, no significant effect was observed on the S and G2-M stage of the cell cycle stage after treatment with the linamarin for 24 hours.

The second part of the study was on fabrication of biodegradable linamarin loaded nanoparticles. Poly (lactic-co-glycolic acid) (PLGA) was chosen as the polymeric material of the nanoparticles. The water-in-oil-in-water emulsification process was the method of choice for the encapsulation of linamarin inside polymeric particles. The linamarin nanoparticles based on two different mole fraction of PLGA copolymer (50/50 and 85/50 of lactic acid/glycolic acid, respectively) were successfully fabricated using water-in- oil-in-water double emulsion extraction/evaporation technique. The SEM



analysis on the morphologies of the nanoparticles showed the particles are spherical in shape with porous surface structure and well within nano-scale in size.

A preliminary investigation on *in vitro* drug (linamarin) release was also carried out. The *in vitro* drug (linamarin) release was characterised by an initial burst and incomplete dissolution of the drug. When decreasing the polymer/drug ratio, the release appeared more controlled and prolonged up to 8hr. It can be concluded that nanoparticles prepared by water-in-oil-in-water emulsification followed by solvent evaporation is a good potential for a controlled released-drug carriers for linamarin.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

SIFAT-SIFAT ANTIKANSER LINAMARIN SECARA IN-VITRO DAN PELEPASAN TERKAWALNYA DARIPADA NANOZARAH ASID PLGA BOLEH BIOROSOT

Oleh

WEDAD ASHOUR AL FOURJANI

November 2005

Pengerusi: Norhafizah Abdullah, PhD

Fakulti : Kejuruteraan

Terdapat banyak minat dalam penemuan agen kemoterapi yang baru untuk penyakit kanser. Kajian ini melibatkan penyaringan linamarin sebagai agen terapi untuk pelbagai sel-sel kanser memandangkan tiada kajian yang sama dijalankan terdahulu. Peningkatan bioavailabiliti dan penghantaran linamarin ke sel-sel tumor yang ingin ditujui boleh dijuruterakan dengan pemilihan pembawa yang sesuai. Terdapat banyak kebaikan dalam memilih nanopartikel yang boleh dibiodegradasikan sebagai pembawa ubat. Ini termasuk peningkatan bioavailabiliti dan keberkesanan ubat. Ia juga membolehkan pengawalan terhadap mekanisma pelepasan di mana aktiviti ubat tersebut boleh diperpanjangkan di kawasan yang terjangkit. Disamping itu, sifat pembawa yang boleh dibiodegradasikan juga bermakna partikel-partikel tersebut mudah larut dalam sistem tanpa membawa sebarang kesan sampingan kepada badan. Kajian ini bertujuan mengaji fabrikasi linamarin yang dikapsulkan dalam nanopartikel yang boleh dibiodegradasikan untuk tujuan pembunuhan sel-sel kanser.



Kajian ini dimulakan dengan menyiasat kesan ketoksikan linamarin terhadap sel-sel kanser serta kitaran selnya. Pengajian kesan linamarin di luar tubuh badan dilakukan ke atas dua jujukan sel tumor, iaitu HeLa (jujukan sel tumor servik) dan CAOV3 (jujukan sel tumor ovari). Sitotoksiksiti linamarin ditentukan dengan asei MTT. Kedua-dua jenis jujukan sel menunjukkan kematian sel yang nyata apabila didedahkan kepada linamarin pada nilai IC₅₀ yang berada di dalam julat keberkesanan. (Nilai IC50 untuk HeLa adalah 30 mg/ml dan 58 mg/ml untuk sel CAOV3 apabila kedua-dua sel ini didedahkan kepada linamarin yang tulen. Keputusan ini menunjukkan bahawa linamarin mempunyai potensi sebagai calon ubat dalam rawatan kanser. Kitaran sel yang nyata pada fasa G1 kitaran sel. Ini bermakna terdapatnya nombor sel yang nyata yang telah disekat pada fasa G1. Walaubagaimanapun, tiada kesan yang nyata yang diperhatikan pada fasa S dan G2-M kitaran sel selepas dirawatkan dengan linamarin selama 24 jam.

Bahagian kedua kajian ini adalah mengenai fabrikasi nanopartikel dengan muatan linamarin yang boleh dibiodegradasikan. Poli (laktik – ko – asid glikolik) (PLGA) dipilih sebagai bahan polimerik nanopartikel. Air-dalam-minyak-dalam air adalah proses pengemulsian yang dipilih untuk mengkapsulkan linamarin ke dalam partikel polimerik. Nanopartikel linamarin yang berasaskan dua pecahan mol kopolimer PGLA yang berlainan (50/50 dan 85/15 masing-masing untuk pecahan mol asid laktik kepada asid glikolik) telah berjaya dihasilkan dengan teknik pengekstrakan/ penyejatan dua kali ganda pengemulsian air-dalam-minyak-dalam-air. Analisis morfologi nanopartikel



dengan SEM menunjukkan bahawa partikel-partikel yang dihasilkan adalah dalam bentuk sfera dengan struktur permukaan yang berliang dan saiz yang berada dalam skala nano. Kajian pada peringkat awal tentang pelepasan ubat (linamarin) di luar tubuh badan juga dijalankan. Pelepasan ubat (linamarin) di luar tubuh badan bercirikan peletusan pada permulaan dan keterlarutan ubat yang tidak lengkap. Apabila nisbah polimer kepada ubat dikurangkan, pelepasan ubat didapati lebih terkawal dan berlanjutan sehingga 8 jam. Kesimpulannya, nanopartikel yang disediakan dengan pengemulsian air-dalam-minyakdalam-air dan diikuti dengan penyejatan pelarut merupakan satu potensi yang baik untuk pelepasan pembawa ubat linamarin yang terkawal.



ACKNOWLEDGEMENTS

I wish to express my profound gratitude to Dr. Norhafizah who thoroughly supervised this work with great interest and enthusiasm. The timely support, comments and evaluation allowed me to complete the research project on schedule that played a huge part in making it possible for me to pursue the dream of obtaining a master degree. Special thanks to Associate Professor Dr. Rozita Rosli, for providing the assistance in the cytotoxicity experiments. In this regard, I owe my most sincere thankfulness to my other members of my dissertation committee respectively Assoc. Prof. Dr. Robiah Yunus and Dr. Nashiru Billa, for sharing their knowledge and wisdom with me. Not to forget Dr. luky Sunny who supervised me in the first and second semester.

My parents: Mr. and Mrs. M. Al Fourjani for their prayers love. I thank my sisters; Aisha, basma, and my entire family in Libya for their unceasing mails.

At last, but definitely not the least, I would like to give my special thanks to a very special person in my life-my husband, Kadri Lyeaas and my son Abdo. I am most grateful to god for the precious gift. Kadri who have been a solid support and continuous source of encouragement. He is not only very understanding and supportive to my studies, but also shows me what life is really about besides books, research and internet. More importantly, he helps me how to face difficulties and cherish life. I am thankful that I have him in my life.

Thank you!!

TABLE OF CONTENTS

DEDICHERAL	
DEDICATION	ii
ABSTRACT	iii
ABSTRAK	vi
ACKNOWLEDGEMENT	ix
APPROVAL	х
DECLARATION	xii
LIST OF TABLES	xvi
LIST OF FIGURES	xvii
LIST OF ABBREVIATIONS	XX

CHAPTER

1	INTRODUCTION	1
	1.1 Introduction	1
	1.2 Problem Statement	3
	1.3 Objectives and Strategies of The Thesis	5
2	LITERATURE REVIEW	7
	2.1. Linamarin	7
	2.1.1. Linamarin as the Toxic Compound in Cassava	7
	2.1.2. Potential Application of Linamarin	8
	2.2. Cell cycle	11
	2.3 Introduction to control release	13
	2.4 Polymer system in controlled release	15
	2.4.1 Polymeric matrices	16
	2.4.1.1 Water soluble polymer	16
	2.4.1.2 Biodegradable polymer	17
	2.4.1.3 Non biodegradable polymer	18
	2.4.2 Drug Released Mechanism in Polymeric System	19
	2.5 Controlled drug delivery based on biodegradable polymer	23
	2.5.1 Physical and chemical properties of biodegradable	24
	Polymers	
	2.5.2 Degradation and erosion of biodegradable system	27
	2.5.3 Modeling of biodegradable system	31
	2.5.4. Nanoparticles	33
	2.5.5. Primary method for nanoparticles preparation	34
	2.5.5.1. Emulsion-Solvent Evaporation/Extraction Method	35



2.5.5.2. Spontaneous Emulsification/ Solvent	Diffusion 37
2553 Salting Out /Emulsification Diffusio	n Mathad 20
2.5.5.4 Nano Precipitation Method	n Method 38
2.5.5.5. Production of Nanonarticles Using S	Superamitical 20
Eluid Technology	supercritical 39
2556 Polymerization Method	41
2.5.6 Factors Affecting Nanonarticles Production	41
2.5.7. Solvent Removal by Lyophilization (Freeze d	rving) 47
	.,
3 MATERIALS AND METHODS	48
3.1 Materials	48
3.1.1. Chemical and media	49
3.1.2. Equipments	49
3.2 Wethods	49
3.2.1 Cell culture	49
3.2.2 WIT I assay	49
3.2.2.1. Statistical Analysis for MTT Assay St	udy 50
3.2.3 Flow Cylometry	51
3.2.3.1. Flow Cylometry	51
2.2.2.2. Preparation of the cell for flow cytom	eter analysis 51
3.2.3.3. Statiling	52
3.2.5.4 Statistical analysis for flow cytometry	study 53
3.2.4 reparation of infamiliar noncoded nanoparticles	53
3.2.4.1 Poly vinyi alconol (PVA) solution	53
3.2.4.2 Single emulsion formulation	53
3.2.4.5 Double emulsion formulation	54
3.2.4.4 Nano-emulsincation step	. 54
3.2.5 Scanning Electron Microscope (S.E.M) analys	IS 55
3.2.6. Nanoparticles drug loading content and entrap	oment 55
2.2.7 In with draw release state	
5.2.7 In vitro drug release study	56
4 RESULTS AND DISCUSSION	57
4.1 Cytotoxicity Study of Lingmarin on Cancor Colla	57
4.2 The Effect of Drugs on Cell avaia by Flow Cytometer	· Charles 65
4.2.1 The Effect of Temovifen on the Call Crule of	/ Study 65
4.2.1. The Effect of Tamoxilen on the Cell Cycle of	Hela Cells 66
4.2.1.2. The effect on GI phase	67
4.2.1.2. The effect on S-phase	67
4.2.1.3. The effect of U-m line of the first	67
and CAOV3 cell lines	cle of HeLa 70
4.2.2.1. The effect on G1-phase	70
4.2.2.2. The effect on S-Phase	71



4.2.2.3. The effect on G2-M Phase	71
4.2.3 The Effect of Crude linamarin on The Cell	Cycle of 76
HeLa and CAOV3 cell lines	
4.2.3.1. The effect on G1-phase	76
4.2.3.2 The effect on S-Phase	77
4.2.3.3. The effect on G2-M Phase	77
4.3. Production of Linamarin loaded Biodegradable Nanop	articles 82
4.3.1. Scanning Electron Microscope (S.E.M) analys	sis 85
4.3.2. Entrapment efficiency of Linamarin in biodeg	radable 92
PLGA Nano- Particles	
4.3.3. Preliminary In vitro Drug Release Study	93
5 GENERAL CONCLUSION AND FUTURE WORK	96
5.1 Conclusion	96
5.2 Future development	97
5.2.1. Linamarin toxicity study on other cancer cell lines	97
5.2.2. Nanoparticle fabrication	97
5.2.3. Drug release study	98
REFERENCES	99

REFERENCES	99
APPENDICES	110
BIODATA OF THE AUTHOR	126



LIST OF TABLES

Table		Page
2.1.	Examples of Water-Soluble Polymers used as Drug Delivery Matrices	17
2.2	Examples of Biodegradable Polymers Used in Drug Delivery	18
2.3.	Examples of Non biodegradable Polymers Used in Drug Delivery	19
2.4	Characteristics of lactide/glycolide polyesters	27
2.5	Summary of methods used for preparation of polymeric nanoparticles.	43
2.6	Comparison of particles diameter for polymeric nanoparticles.	46
4.1	IC ₅₀ result of the MTT assay.	60
4.2	The effect of the polymer and the drug on the entrapment efficiency %.	92



LIST OF FIGURES

Figur	e.	Page
2.1	Structures of linamarin (I), lotaustralin (II) and acetone cyanohydrin (III).	9
2.2	Linamarin biosynthesis and breakdown pathway in cassava	10
2.3	Cell cycle diagram	12
2.4	Plasma concentration of drug as a function of time after administration.	14
2.5	Schematic of the drug delivery based on the different mechanism.	22
2.6	Structure of lactic/glycolic acid and poly lactic-co-glycol ides (PLGA)	24
2.7	The degradation of PLGA copolymer to form lactic and glycolic acid	25
2.8	Schematic of the surface erosion and bulk erosion.	30
2.9	Particle preparation methods via solvent evaporation method (single and double emulsion)	37
2.10	Schematic diagram of the supercritical anti solvent (SAS) method	40
2.11	Schematic representation for the production of poly (alkylcyanoacrylate) nanoparticles by anion polymerization	42
4.1	The effect of different drug on HeLa cell viability.	59
4.2	The effect of pure linamarin on HeLa cells	61
4.3	Effect of crude linamarin on HeLa cells	61.
4.4	Effect of tamoxifen on the HeLa cell as drug control.	62
4.5	Effect of tamoxifen on CAOV3 cells as a drug control	62
4.6	The effect of pure linamarin on CAOV3 cells	63
4.7.	The effect of crude linamarin on CAOV3 cells	63
4.8	The effect of pure linamarin with linamarase on HeLa cells	64



4.9.	The effect of pure linamarin with linamarase on CAOV3 cells	64
4.10	Histographs illustrating the 3 phases of cell cycles in HeLa and CAOV3, namely M1 (G1 phase), M2 (S phase) and M3 (G2-M phase).	66
4.11	Flow cytometery histographs of cell cycle analysis. HeLa cells were exposed to tamoxifen at different concentration (3,6 and 12 μ g/ml) for 24 hr.	68
4.12	Histogram showing HeLa cells treated with different concentration of tamoxifen for 24 hours.	69
4.13	Flow cytometry histograph of cell cycle analysis. HeLa cells exposed to pure linamarin for 24 hours.	72
4.14	Histographs showing flow cytometry of cell cycle analysis of CAOV3	
	cells	73
4.15	Histogram showing HeLa cell treated with pure linamarin for 24 hours	74
4.16	Histogram showing Caov-3cell treated with pure linamarin for 24 hours	75
4.17	Flow cytometry histographs of CAOV3 cell cycle analysis	78
4.18	Flow cytometry histographs of cell cycle analysis on HeLa cells exposed to crude linamarin for 24hr.	79
4.19	Histogram showing HeLa cells treated with crude linamarin for 24 hours	80
4.20	Histogram showing Caov-3cell treated with crude linamarin for 24 hours	81
4.21	SEM micrograph showing linamarin loaded PLGA nanoparticles	84
4.22	SEM micrographs of nanoparticles showing the shape and surface characteristic (a) PLGA 50/50 (b) PLGA 85/15	87
4.23	SEM micrograph of PLGA 50/50 nanoparticles loaded with 5mg linamarin.	88
4.24	SEM micrographs of PLGA 50/50 nanoparticles loaded with 10 mg linamarin.	89
4.25	SEM micrographs of PLGA 85/15 nanoparticles loaded with 5mg linamarin.	90



4.26	SEM micrograph of PLGA 85/15 nanoparticles loaded with 10 mg linamarin.	91
4.27	Release profiles of linamarin (5 mg) from different molar ratio of PLGA nanoparticles.	95
4.28	Release profiles of linamarin (10 mg) from different molar ratio of PLGA nanoparticles.	95



LIST OF ABBREVIATIONS

ACA	alkyl cyanocrylate
ANOVA	analysis of variance
CN ⁻	cyanide ion
CO ₂	carbon dioxide
DCM	dichloromethane
DMAB	didodecyl dimethyl ammonium bromides
DMSO	dimethylsulphoxide
DNA	dideoxyribonucleic acid
DSC	differential scanning calorimetry
FDA	Food and Drug Administration
GAS	gas anti solvent
HCN	hydrogen cyanide
HPLC	high performance liquid chromatography
IV	intravenous
MPS	mononuclear and phagocytic system
MTT	3-4, 5-dimethylthizol-2-yl)-2-5-diphenyl tetrazolium bromide solution
MW	molecular weight
OD	optical density
PACA	Poly alkyl cyanorylate
PBS	phosphate buffer saline



PDLLA	poly D, L lactic acid
PGA	poly glycolide
PI	propidium iodide
PLA	poly lactide
PLGA	poly lactic glycolic acid
PLLA	poly L-lactic acid
PVA	polyvinyl alcohol
RESS	rapid expansion of supercritical
RPMI media	Roswell Park Memorial Institutes media
RNA	ribonucleic acid
SAS	supercritical anti solvent
SEM	scanning electron microscope
W1/O	water-in-oil
W1/O/W2	water-in-oil- in-water



CHAPTER 1

INTRODUCTION

1.1 Introduction

The drug delivery system is a system which delivers or carries the drug to the infected sites. The system is characterised by its ability to incorporate drugs without damaging them, long *in vivo* stability, its tuneable release kinetics and targeting to specific organs and tissues. This tuneable release kinetics is a characteristic for a controlled drug delivery mechanism. The controlled drug delivery offers many advantages over conventional dosage forms, including improved efficacy, reduced toxicity, improved patient compliance, and cost effective therapeutic treatment. In particular, the controlled release mechanism is strongly required for unconventional drugs, such as proteins and oligopeptides.

In recent years, there has been significant effort to develop nanotechnology for drug delivery since it offers a suitable means for delivering small molecular weight drugs, as well as macromolecules such as protein, peptide or genes. Most of the works focus on formulation of therapeutic agents in biocompatible nano-composites such as nanoparticles, nanocapsules, micellar system, and conjugates. These systems are often polymeric based matrix and submicron in size.

These nanotechnology systems can be used to provide targeted delivery of drugs, to improve the oral bioavailability and to sustain drug effect in cancer tissues. They can



also be used to solubilize drugs for intravascular delivery and to improve the stability of therapeutic agents against enzymatic degradation. Much work in the past found that nanoparticulates drug carrier made of polymer appear to be more stable when in contact with biological fluids than other colloidal drug carriers (Kreuter *et al.*, 1988; Zambaux *et al.*, 1998). They also have been proposed as drug delivery systems for different routes of administration and for different types of active ingredients such as anticancer agents (Feng *et al.*, 2003and Fonseca *et al.*, 2002), anti-inflammatory compounds (Chacon *et al.*, 1999), oligonucleotides (Lambert *et al.*, 2001; Ulbrich *et al.*, 2004) and peptides (Lemoine and Preat, 1998).

Polymers can be used as a base matrix for nanoparticles. Polymeric nanoparticles generally vary in size from 10 to1000 nm. The fabrication of polymeric nanoparticles is via dissolvement, entrapment, encapsulation or attachment of the drug to a polymer matrix. The polymers used to make the nanoparticles for administration into the human body are significantly limited to a few types of polymers due to their biocompatibility and biodegradation although various polymers can be employed to make nanoparticles.

There has been intensive research in the development of nanoparticles of biodegradable polymers as an effective drug delivery system for medical practice, especially for chemotherapy and gene delivery. Progress in nanoparticles technology, material science of biodegradable polymers and cellular and molecular physiology and pathology have contributed to the advancements in chemotherapy and gene therapy of cancer and other



disease with polymeric nanoparticles been considered as promising carriers for the therapeutic agent.

Nanoparticulate delivery systems, based on poly (lactic-co-glycolic acid) (PLGA) polymers have been studied extensively for many years (Song.C.X, 1997). PLGA (lactic-co-glycolic acid) and its homo- or copolymers are the most widely used biodegradable polymers for fabricating nanoparticles. PLGA polymers have the advantage of being well characterized and have been commercially used as a microparticulate drug delivery systems. They are biocompatible, biodegradable and bio-resorbable.

1.2. Problem Statement

Chemotherapy is a complicated procedure in which many factors are involved in determining its success or failure. It carries a high risk due to drug toxicity and usually the more effective drugs tend to be more toxic. Problems related to drug side effects still exist even for successful chemotherapy, with patients not only have to tolerate the severe side effects but also sacrifice their quality of life. The effectiveness of chemotherapy depends on many factors, including the drug (s) used, the condition of the patient, the dosage and its form and schedule and others.

Most anticancer drugs are highly hydrophobic, and hence are not soluble in water and most pharmaceutical solvents. Adjuvants have to be used for the clinical administration of many anticancer drugs and this may cause serious side effects, some of which are life threatening. Development of effective carriers with little side effects for anticancer

