



UNIVERSITI PUTRA MALAYSIA

***EVALUATION OF NATURAL IMMUNOSTIMULANTS FOR GROWTH
PROMOTION AND PROTECTION AGAINST *Aeromonas hydrophila*
IN JUVENILE RED HYBRID TILAPIA (*Oreochromis* sp.)***

NUR HIDAYAHANUM HAMID

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By

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**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfillment of the Partial Requirements for the Degree of Master of Science**

May 2014

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DEDICATION

With appreciation and respect, this thesis is dedicated to:

My beloved parent, brothers and sisters.

then calculated. Daily mortality was recorded for a week. From the study, 11 isolates of bacteria were successfully identified by using morphological, biochemical (conventional and commercial kit) and physiological tests. The bacteria isolates were characterized as Gram-negative, motile, catalase positive, oxidase positive, possessed straight rod cell; approximately up to 3 μm in length which appeared in ukpingu"cpf" rcktu0"Vjg"kuqncvgu"ujqygf" -haemolysis on horse blood agar and were able to grow on selective agar (Rimler-Shott agar). All bacteria isolates were found to be sensitive towards streptomycin, kanamycin, chloramphenicol and gentamicin but showed resistant to amoxicillin. In a pathogenicity test run by intraperitoneal injection, the infection caused marked clinical sign of abnormalities such as exophthalmia, lethargy, enlargement of kidney, spleen and liver. Histopathological examinations showed marked congestion and haemorrhages in the spleen, liver and kidney tissues. The median lethal dose (LD_{50}) at 96 hours of the isolate for juvenile Red hybrid tilapia (*Oreochromis* sp.) was 6.3×10^6 cfu/ml. Administration of LPS cpf" -glucan by intraperitoneal injection, long-term bath exposure and oral feeding significantly enhanced the RPS. The survivability was higher in fish treated with both compounds, which was significantly high (more than 50%) when compared to control group (less than 50%). The results indicated that oral feeding had the highest survival among the treatment groups followed by i.p injection and bath due to the long duration of feeding exposure. In haematological assay, there were no significant difference between control and treated groups. However, the fish cf o kpkuvgtgf" ykvj "NRU"cpf" -glucan showed significant increased in total leucocytes count ($P < 0.05$) and also significantly ($P < 0.05$) improve the growth performance of tested fish compared with the control group. In histopathological examinations, different types of MMC (melano-macrophage centre) were observed in spleen tissues of control and treated groups. The present study revealed that the presence of numerous hemosiderin granules (brownish yellow) was higher in the spleen of control group compared to treated groups. The MMC in the treated group was more enriched in melanin pigment which is dark pigment-containing cells (macrophage). The findings in this study indicates that *A. hydrophila* infection has become an important health issue in tilapia farms. This study also demonstrates that NRU"cpf" -glucan administration through injection, long-term bath exposures and oral feeding effectively stimulates the non-specific cellular as well as growth performances and offers protection against *A. hydrophila* infection in juvenile Red hybrid tilapia (*Oreochromis* sp.). Data from biochemical tests, haematological and histological studies provide valuable and previously unknown information associates with *A. hydrophila* in cultured juvenile Red hybrid tilapia (*Oreochromis* sp.). In addition, this study also reveals that NRU"cpf" -glucan could be used as an alternative for prophylaxis against *A. hydrophila* infection and the important of long term exposure of immunostimulants to obtain maximum protection against bacterial infection.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**PENILAIAN KE ATAS IMMUNOSTIMULASI SEMULAJADI DALAM
MENGALAKKAN TUMBESARAN DAN PERLINDUNGAN MENENTANG
Aeromonas hydrophila PADA REGA IKAN TILAPIA MERAH HIBRID
(*Oreochromis* sp.)**

Oleh

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Mei 2014

Pengerusi : Prof. Madya Hassan Hj. Mohd Daud, PhD

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Aeromonas hydrophila terkenal dengan meluas sebagai spesies bakteria yang biasa dalam habitat air tawar dan kadangkala dikenali sebagai patogen ikan. Di Malaysia, penggunaan immunostimulasi terutamanya dalam industry akuakultur kurang dikaji kerana kurangnya promosi dan pengetahuan terutamanya para penternak ikan. Oleh itu, kajian ini dilakukan untuk mengenalpasti *A. hydrophila* daripada ikan air tawar yang dijangkiti penyakit. Bakteria tersebut juga digunakan untuk ujian sensitivity antimikrobial. Dua jenis immunostimulasi (Lipopolisakarida fcp" -glukan) digunakan untuk menilai keberkesanan bagi memeriksa perubahan dalam hematologi, sel tak-spesifik, daya rintang penyakit dan penilaian tumbesaran rega ikan tilapia merah hibrid (*Oreochromis* sp.). Dalam kajian ini, isolate *A. hydrophila* melalui perwarna Gram, Identiti isolate *A. hydrophila* dipastikan dengan API 20E[®] dengan kombinasi ujian oksidasi dan katalase. Kemudian bacteria tersebut diinokulasi atas agar darah kuda dan agar Rimler-Shotts. Ujian sensitiviti antimikrobial dijalankan menggunakan kaedah Kirby-Bauer. Selepas itu, kepatogenan *A. hydrophila* diungkap sebagai median dosis maut (LD₅₀) dan ikan nazak diteruskan dengan ujian histopatologi. Daripada kajian ini, keberkesanan LPS fcp" -glukan turut dikaji pada rega tilapia merah hibrid (*Oreochromis* sp.) menentang patogen bakteria, *A. hydrophila*. Sekumpulan ikan dibahagi kepada tujuh kumpulan yang berbeza termasuk kawalan dimana setiap kumpulan mengandungi 15 ekor ikan. Setiap kumpulan memiliki dua replika. Mgrgmcvcp"NRU" fcp" -glukan yang berbeza digunakan untuk suntikan (i.p.), mandian dalam jangka masa panjang dan pemakanan secara oral iaitu 50 µg/ikan, 50µg/L dan 25 mg/kg, setiap satu. Untuk suntikan (i.p.) dan ocpfkcpcp"NRU" fcp" -glukan diberi pada hari pertama, ke-tujuh dan ke-empat belas. Sementara, kumpulan ikan yang diberi makanan secara oral diberi makan pellet mq ogtukcnc" {cp i" fkv o dcj"NRU" cvcw" -glukan setiap hari sehingga 40 hari. Pada hari ke-tujuh, sampel darah ikan diambil daripada setiap kumpulan untuk ujian hematologi dan imunologi. Ikan kawalan dan kajian dijangkitkan dengan *A. hydrophila* berkepekatan LD₅₀ pada hari ke-16 (suntikan intraperitoneal dan

mandian) dan hari ke-41 (oral). Peratus tahan hidup relatif (RPS) dan kesan modulasi imun lipopolisakarida fcp" -glukan dikaji pada rega hybrid ikan tilapia merah (*Oreochromis* sp.) menentang bacteria patogen, *A. hydrophila*. Mortaliti direkodkan setiap hari selama seminggu dan RPS dikira. Daripada kajian ini, sebelas *A. hydrophila* isolasi bacteria telah Berjaya dipencilkan melalui ujian morfologi, ujian biokimia (konvensional dan kit komersial) dan ujian fisiologi. Bacteria yang diisolasi dikenalpasti sebagai Gram-negatif, motil, positif katalase, positif oksidase, terdiri daripada sel morfologi rod lurus lebih kurang 3 µm panjang, terdapat dalam bentuk tunggal dan pasangan. Bacteria tersebut ogpwlwmmcp" -hemolisis di atas agar darah kuda dan boleh tumbuh di atas agar selektif (agar Rimler-Shott). Kesemua isolate bacteria adalah peka terhadap streptomisin, kanamisin, kloramfenikol dan gentamisin, tetapi rintang terhadap amoxicillin. Dalam ujian patogenisiti secara suntikan intraperitoneal, jangkitan menyebabkan tanda-tanda klinikal abnormal seperti exoftalmia, letargi, pembesaran ginjal, limpa dan hati. Dalam ujian histopatologi, terdapatnya congesti dan hemoraj di dalam tisu ginjal, limpa dan hati. Median dosis maut, LD₅₀ pada 96 jam oleh pencilan bacteria (*A. hydrophila*) bagi rega tilapia merah hybrid (*Oreochromis* sp.) secara suntikan intraperitoneal adalah 6.3x10⁶ cfu/ml. RPS bertambah secara signifikan dengan rgodgtkcp" NRU" fcp" -glukan secara suntikan intraperitoneal, mandian dan pemberian secara oral. Keupayaan tahan hidup adalah tinggi padaikan yang diberi rawatan kedua-dua sebatian LPS dan -glukan (lebih daripada 50%) berbanding dengan kawalan (kurang daripada 50%). Hasil kajian menunjukkan pemberian secara oral mempunyai tahan hidup yang tertinggi berbanding kumpulan rawatan yang lain kerana pemberian makanan dalam jangkamasa yang panjang. Bagi ujian hematologi, nilainya adalah tidak signifikan untuk kmcp" {cpi" fkdgtk" NRU" fcp" -glukan dengan kumpulan kawalan. Jumlah leukosit keseluruhan serta pelaksanaan pertumbuhan juga bertambah secara signifikan untuk kmcp" {cpi" fkdgtk" NRU" fcp" -glukan. Dalam kajian histopatologi, MMC (pusat melano-makrofaj) yang berlainan jenis dapat dilihat di dalam tisu limpa. Kajian ini mendedahkan banyaknya kehadiran hemosiderin (coklat keperangan) di dalam tisu limpa bagi kumpulan kawalan berbanding kumpulan yang dirawat. MMC dalam kumpulan yang dirawat lebih kaya dengan pigmen melanin dimana ia adalah pigmen gelap mengandungi sel (makrofaj). Kajian ini mendapati dcjcycc" NRU" fcp" -glukan yang diberi secara suntikan, mandian dan pemakanan secara oral merangsang dengan efektif sel tak-spesifik serta pelaksanaan pertumbuhan dan member perlindungan terhadap jangkitan *A. hydrophila* pada rega tilapia merah hybrid (*Oreochromis* sp.). Hasil daripada ujian biokimia, hematologi dan histology member maklumat yang bernilai dan tidak diketahui sebelum ini mengenai *A. hydrophila* pada rega tilapia merah hybrid (*Oreochromis* sp.). Tambahan lagi, kajian ini turut mendedahkan bahawa LPS fcp" -glukan boleh digunakan sebagai profilaksis alternative menentang infeksi *A. hydrophila* dan pentingnya pendedahan jangka masa panjang immunostimulasi untuk mendapat perlindungan maksimum terhadap jangkitan bacteria.

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This thesis was submitted to the Senate of Universiti Putra Malaysia as fulfillment of the requirements for the degree of Master of Science. The members of the supervisory committee were as follows:

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LIST OF ABBREVIATIONS

cfu	Colony-forming unit
DO	Dissolved oxygen
dpi	Days post inoculation
FAO	Food and Agriculture Organization of the United Nations
H&E	Hematoxylin and eosin
i.p	Intraperitoneal
LD ₅₀	Median lethal dose
MMC	Melano-macrophage centres
Ppt	Parts per thousand
SCP	single cell proteins
TSA	Trypticase soy agar
TSB	Trypticase soy broth
w/v	Weight per volume
w/w	Weight per Weight

CHAPTER 1

INTRODUCTION

1.1 Fish Diseases and Health Management in Malaysia

Aquaculture is one of important industries all over the world. From its beginning in the 1920s, aquaculture in Malaysia has developed quickly and become an important activity nowadays (FAO, 2011). Various kinds of marine and freshwater fish have been cultured and the production of cultured fish increases every year. In Malaysia, fishes are usually cultured in enclosed system such as ponds, net cages and the latest one is in recirculating aquaculture system (RAS). Nowadays, the main aim of commercial aquaculture is to increase production by intensification; which are by increasing stocking density, increased seed production, and feeding with good quality of feed. However, disease in intensive aquaculture is said to be the great importance in Malaysia due to economic loss observed in recent years. Consequently, the use of chemicals and antibiotics had become common practices in fish farm.

Due to the intensive aquaculture practices, fish in cultured systems are susceptible to numerous types of bacterial, viral and parasitic diseases. The presence and development of fish disease is the result of the interaction between pathogen, host and environment. In the cultured system, poor handling and overcrowding always tend to give unfavourably affect to the fish health. These conditions may lead to the production of poor physiological environment and increase susceptibility of the cultured fish to infection disease (Sakai, 1998). Among the three major causes of disease, most frequent related to freshwater fish is bacterial infections. Hasty and uncontrolled growth of pathogens and indiscriminate use of antibiotics to prevent the emerging of pathogen have resulted in the emergence of several resistant pathogens in aquaculture. Currently, these two factors are the most fundamental concerns for both public health workers and farmers.

Even though the infectious diseases which caused by pathogenic species of bacteria in freshwater fish have been described in the majority of the existing taxonomic groups, however only a relatively small number are responsible of important economic losses in cultured fish worldwide. *Aeromonas* spp., *Pseudomonas* spp., *Streptococcus* spp. and *Flexibacter columnari* are some of the regularly bacteria isolated from fish and become primary pathogenic agents which frequently reducing the production of cultured freshwater fish. Among this pathogen of bacterial origin, motile aeromonads play an important role in freshwater systems.

1.2 Aeromoniasis Study in Malaysia

Aeromonas hydrophila and other motile aeromonads are among the most common bacteria in freshwater habitats throughout the world. These bacteria frequently cause diseases among cultured and wild fishes. Because of this, the bacteria species is said to be commonly isolated from diseased fish. *Aeromonas hydrophila* is a primary

(Esteve *et al.*, 1993), secondary (Joice *et al.*, 2002) and opportunistic pathogen (Dooley and Trust, 1988) of a wide variety of aquatic and domestic animals, including humans.

In Malaysia, only few report of disease outbreaks that was caused by *Aeromonas hydrophila* in aquaculture system especially in food fish have been documented. Infections due to *Aeromonas hydrophila* are common and pose a threat especially to the development of the aquaculture sector. Taufik and Wong (1990) showed that *Aeromonas hydrophila* was the major pathogenic bacterium in catfishes from paddy field Kedah and Perak, West Malaysia. In addition, Najiah *et al.* (2008) reported the presence of *Aeromonas hydrophila* not only in the food fish but also in freshwater imported ornamental fish (*Xiphophorus maculatus*, *Barbus pentazona hexazona*, *Symphysodon* spp., *Colisa lalia*, *Gymnocorymbus ternetzi*, *Poecilia reticulata*, *Pangasius sutchi*, and *Osphronemus goramy*) in retail pet shop in Kuala Terengganu.

1.3 Current Study on Fish Health Management

Various chemicals and antibiotics have been used to treat bacterial infections in cultured fish. It provides a useful means of control to the infections. However, there are many problems associated with the development of drug-resistant bacteria and the high cost of treatment. At present, preventive and management measures are fundamental concern to overcome such outbreak of diseases. Immunostimulants are considered as a helpful and effective means for enhancing immune status of cultured fish. There are several approaches to disease prevention that previously have been successfully used in other animal industries such as vaccines and immunostimulants. Both approaches also have been effectively used in some aquaculture industries and should be considered as a part of fish health management options.

Vaccination is a useful prophylaxis for infectious diseases and it is already commercially available for bacterial infections such as vibriosis, redmouth disease and furunculosis and also for viral infection such as IPN (Sakai, 1998). In the salmon industry, it has been used for about 30 years and known to be one of the major reasons that salmon production has been successful. The use of vaccine also dramatically decreased the use of antibiotics (Somerset *et al.*, 2005). For example, in Norway (1987), before the extensive use of vaccines, almost 50,000 kg of antibiotics were used. In 1997, when vaccines had become more routine practices, antibiotic usage had decreased to less than 1000-2000 kg (Somerset *et al.*, 2005). For the time being, even though vaccination may be the most effective method of controlling fish diseases but not all diseases can be treated with vaccine. According to Sakai (1998), the vaccine development for *Renibacterium salmoninarum*, is still not far from being successful. Therefore, it is impossible for us to rely only on vaccination in order to control the fish diseases.

Nowadays the main problem in the fish rearing during larval and on-growing stages is microbial pathogens. Therefore, it is important for us to develop methods for establishing microbial control at all stages of the cultivation progress. One

possibility is immunostimulation, which includes methods of enhancing the capacities of the specific and non-specific immune systems. It is valuable for the control of fish diseases and may also be useful in fish culture. Immunostimulants increase resistance to infectious disease, not only by enhancing specific immune response but also by enhancing non-specific defense mechanisms (Sakai, 1998). There are many experiments on non-specific immunostimulation of fish that suggest the method has considerable potential for reducing losses in aquaculture, both during larval and on-growing stages.

1.4 Statement of Problem and Significance of Study

Vaccination using immunostimulant is very effective and acts as a potential approach in order to increase the immunocompetency and disease resistance of fish. Immunostimulants are said to be safer than antibiotic and chemotherapeutic as their range of efficacy are wider and broader compared to common vaccine (Sakai, 1998). However, in Malaysia, the use of immunostimulant especially in aquaculture industry has not been studied widely due to the lack of promotion and education especially to fish farmers. Moreover, there is a relatively few of the end product that show guarantee in a research context become available for use by the fish farmer. For aquaculture industry in Malaysia, it will lead to high return and income since this industry is expanding throughout the region. Therefore, the aim of this study is to evaluate the status and potential of immunostimulation as an element in the strategy for solving microbial problems in promoting fish health against pathogen. The immunostimulants that were used in this study were lipopolysaccharide (LPS) and β -glucan. The assay will be done on juvenile Red hybrid tilapia (*Oreochromis* sp.), but the results are also applicable to other species of fish and organisms relevant to aquaculture.

1.5 Hypothesis of Study

Natural immunostimulants which are lipopolysaccharide (LPS) and β -glucan might be used to promote health and protection against *Aeromonas hydrophila* in juvenile Red hybrid tilapia (*Oreochromis* sp.).

1.6 Objectives of the study

The objectives of this study were;

1. to isolate and identify *Aeromonas hydrophila* from freshwater fish.
2. to perform antimicrobial sensitivity test on *Aeromonas hydrophila* isolates.
3. to investigate whether the application of immunostimulants (β -glucan and LPS) can be used to improve fish health, survival against *Aeromonas hydrophila* infection as well as growth performance.

REFERENCES

- Abdel-Tawwab, M., Khattab, Y.A.E., Ahmad, M.H. and Shalaby, A.M.E. 2006. Compensatory growth, feed utilization, whole-body composition and hematological changes in starved juvenile Nile tilapia, *Oreochromis niloticus* (L.). *Journal of Applied Aquaculture*, 18: 17–36.
- Abulhamd A. 2009. Characterization of *Aeromonas hydrophila* Isolated from Aquatic Environments Using Phenotypic and Genotyping Methods. *Resolution of Journal Agriculture and Biological Science*, 5(9):923-931.
- Adams, C.A., Bundy, A., Thompson, K., and Horne, M.T. 1998. Molecular characterization of plasmid mediated oxytetracycline resistance in *Aeromonas salmonicida*. *Applied Environmental Microbiology*, 64: 41 94-4201.
- Anbarasu, K., and Chandran, M.R. 1998. Evaluation of protective immunity in catfish, *Mystus gulin* against *Aeromonas hydrophila* infection using crude lipopolysaccharide vaccine. In: Proceeding of the 10th International Congress of Immunology. New Delhi, India, pp. 1509-1513.
- Anbarasu, K., Thangakrishnan, K., Aruna, B.V., and Chandran, M.R. 1998. Assessment of immune response in freshwater catfish *Mystus vittatus* to different bacterins of *Aeromonas hydrophila*. *Indian Journal of Experimental Biology*, 36: 990- 995.
- Anbarasu, K., and Chandran, M.R. 2000. Effect of ascorbic acid on the immune response of the catfish, *Mystus gulin* (Hamilton) to different bacterins of *Aeromonas hydrophila*. *Fish and Shellfish Immunology*, 11: 347-355.
- Anderson, D.P. 1974. *Diseases of Fishes*. In Book 4: Fish Immunology. TFH Publications, Neptune City, New Jersey, pp. 239.
- Anderson, D.P. 1992. Immunostimulants, adjuvants, and vaccine carriers in fish: applications to aquaculture. *Annual Review of Fish Disease*, 2: 281–307.
- Anderson, D.P., Siwicki, A.K., and Rumsey, G.L. 1996. *Injection or immersion delivery of selected immunostimulants to trout demonstrate enhancement of nonspecific defense mechanisms and protective immunity*. In Shariff, M., Subasinghe, R.P., Arthur, J.R. (Eds.), *Diseases in Asian Aquaculture*. Fish Health Section, Asian Fisheries Society, Manila, Philippines, pp. 413–426.
- Anon, 2012, <http://www.thefishsite.com/articles/58/>. (Accessed on 1st October 2010).
- Afifi, S.H., Al-Thobiati, S., and Hazaa, M.S. 2000. Bacteriological and histopathological studies on *Aeromonas hydrophila* infection of Nile tilapia (*Oreochromis niloticus*) from fish farms in Saudi Arabia. *Assiut Veterinary Medical Journal*, 84: 195-205
- Agius, C. and Roberts, R.J. 2003. Melano-macrophage centers and their role in fish pathology. *Journal of Fish Diseases*, 26: 499-509.

- Ai, Q., Mai K., Zhang L., Tan B., Zhang W., Xu W., and Li H. 2007. Effects of dietary β -1,3 glucan on innate immune response of large yellow croaker, *Pseudosciaena crocea*. *Fish Shellfish Immunology*, 22: 394–402.
- Anju, P., Naik, M. and Dubey S.K. 2010. Hemolysin, Protease, and EPS Producing Pathogenic *Aeromonas hydrophila* Strain An4 Shows Antibacterial Activity against Marine Bacterial Fish Pathogens. *Journal of Marine Biology*, 2010: 1-9.
- Alagappan, K.M., Deivasigamani, B., Kumaran, S. and Sakthivel, M. 2009. Histopathological alterations in estuarine catfish (*Arius maculatus*; Thunberg, 1792) due to *Aeromonas hydrophila*. *Infection World Journal Fish Marine Science*, 1: 185–189.
- Alderman, D.J. and Michel, C. 1992. *Chemotherapy in aquaculture today*. In Michel, C., Alderman, D.J. (Editors.), *Chemotherapy in Aquaculture: From Theory to Reality*. Office International des Epizootics, Paris, pp. 3-24.
- Al-Harbi, A.H. and Austin, B. 1992. Immune response of turbot, *Scophthal mus maximus* (L.), to lipopolysaccharide from a pathogenic Cytophagea-like bacterium. *Journal of Fish Disease*, 15: 449–452.
- Amend, D.F., and R.W. Eshenour. 1980. Development and use of commercial fish vaccines. *Salmonid*. Volume III, No. 6: S-12.
- Andrzej, K., Siwicki, Zdzisaw Z., Sylwia T., Agata K., Krzysztof K. and Edward G. 2003. Selected hematological and biochemical parameters of pikeperch *sander lucioperca* (L.) from intensive culture. *Archives of Polish Fisheries*, 11: 117-22.
- Angka, S.L. 1990. The pathology of the walking catfish, *Clarias batrachus* (L) infected intraperitoneally with *Aeromonas hydrophila*. *Asian Fisheries Science*, 3(3):343-351.
- Ansary, A., Haneef, R.M., Torres, J.L., and Yadav, M. 1992. Plasmids and antibiotic resistance in *Aeromonas hydrophila* isolated in Malaysia from healthy and diseased fish. *Journal of Fish Disease*, 15(3): 191-196.
- Aoki, T. 1992. *Chemotherapy and drug resistance in fish farms in Japan*. In: *Diseases in Asian Aquaculture*, M. Shariff., R.P. Subasinghe and J.R. Arthur (Editors), Fish Health section, Asian Fisheries Society, Manila, Philippines, Volume 1, pp. 519-529.
- Araujo, V.S., Pagliares, V.A., Queiroz, M.L.P. and Freitas-Almeida, A.C. 2000. Occurrence of *Staphylococcus* and enteropathogens in soft cheese commercialized in the city of Rio de Janeiro, Brazil. *Journal of Applied Microbiology*, 92: 1172- 77.
- Austin B., and Austin, D.A. 1987. *Bacterial fish pathogens*. In: E. Horwood (Ed), *Disease in Farmed and Wild Fish*, Chichester, pp. 191-197.

- Austin, B., and Adams, C. 1996. *Fish pathogens*. In: Austin, B., Altweig, M., Gosling, P.J., Joseph, S.W. (Eds.), *The Genus Aeromonas*. Willey, Singapore, pp. 197-229.
- Azeli, A. Potensi besar ternakan tilapia. KOSMO: 19th July 2007, p. 9.
- Baba, T., Imamura, J. and Izawa, K. 1988. Immune protection in carp, *Cyprinus carpio* L., after immunization with *Aeromonas hydrophila* crude lipopolysaccharide. *Journal of Fish Diseases*, 11: 237–244.
- Baba, T., Imamura, J., Izawa, K. and Ikeda, K. 1999. Cell-mediated protection in carp, *Cyprinus carpio* L. against *Aeromonas hydrophila*. *Journal of Fish Disease*, 11: 171-178.
- Balm, P.H.M., Lieshout, E., Lokate, J., and Wendelaar Bonga, S.E. 1995. Bacterial lipopolysaccharide (LPS) and interleukin 1 (IL-1) exert multiple physiological effects in the tilapia *Oreochromis mossambicus* (Teleostei). *Journal of Comparative Physiology B: Biochemical, Systemic and Environmental Physiology*, 165: 85-92.
- Bauer, A.W., Kirby, W.M.M., Sherris, J.C. and Turck, M. 1966. Antibiotic susceptibility testing by a standardized single disk method. *American Journal of Clinical Pathology*, 45: 493 – 496.
- Baulny, M.O.D., Quentel, C., Fournier, V., Lamour, F. and Gouvello, R.L. 1996. Effect of long-term oral administration of β glucan as an immunostimulant or an adjuvant on some non-specific parameters of the immune response of turbot *Scophthal musmaximus*. *Disease of Aquatic Organism*, 26:139-147.
- Beveridge, M.C.M., and McAndrew, B.J. 1998. *Tilapias: Their biology and exploitation*. London: Chapman and Hall.
- Black, J.G. 1999. *Microbiology- Principles and Exploration*. 4th edition London: Prentice Hall.
- Black, J. G. 2008. *Microbiology: Principles and Explorations*. Jefferson City: John Wiley and Sons.
- Bols, C.N., John Brubacher, L., Rosemarie Ganassin, C. and Lucila Lee, E.J., 2001. Ecotoxicology and innate immunity in fish. *Development & Comparative Immunology*, 1: 853–873.
- Bjorlund, H., Bondestam, J, and Bylund G. 1990. Residues of Oxytetracycline in wild fish and sediments from fish farms. *Aquaculture*, 86: 359-367.
- Boulanger, Y., Lallier, R., and Cousineau, G. 1977. Isolation of enterotoxigenic *Aeromonas* from fish. *Canada Journal of Microbiology*, 23: 1161-1164.

- Bravo, S., Dolz, H., Silva, M.T., Lagos, C., Millanao, A. and Urbina, M. 2005. Final Report. *Diagnosis on the use of pharmaceuticals and other chemicals in aquaculture*. Austral University of Chile. Faculty of Fishery and Oceanography, Aquaculture Institute. P.O. Box 1327. Port Montt, Chile. Project No. 2003-28.
- Bridle, A.R., Carter, C.G., Morrison, R.N., and Nowak, B.F. 2005. The effect of β -glucan administration on macrophage respiratory burst activity and Atlantic salmon *Salmo salar* L., challenged with amoebic gill disease—evidence of inherent resistance. *Journal of Fish Disease*, 28: 347–356.
- Bullock, G.L., Conroy, D.A. and Snieszko, S.F. 1971. *Bacterial disease of fish*. Book 2A. In: Snieszko, S.F., Axelrod, H.R. (Eds.), *Diseases of fishes*. T.F.W. Publications, Neptune, New Jersey.
- Buschmann, A. H., Riquelme, V. A., Hernandez Gonzalez, M. C., Varela, D., Jimenez, J. E., Henriquez, L.A., Vergara, P.A., Guíñez, R. and Filun, L. 2006. A review of the impacts of salmon farming on marine coastal ecosystems in the southeast Pacific. *Journal of Marine Science*, 63(7): 1269-1273.
- Cappucino, J.G. and Sherman, N. 2001. *Microbiology: A Laboratory Manual* 6th Edition. San Francisco: Benjamin Cummings.
- Chaudhury, A., Nath. G., Shukla, B.N. and Sanyal, S.C. 1996. Biochemical characterization, enteropathogenicity and antimicrobial resistance plasmids of clinical and environmental *Aeromonas* isolates. *Journal of Medical Microbiology*, 44: 434-437.
- Chen, C.Y., Wooster, G.A., and Bowser, P.R. 2004. Comparative blood chemistry and histopathology of tilapia infected with *Vibrio vulnificus* or *Streptococcus iniae* or exposed to carbon tetrachloride, gentamicin, or copper sulphate. *Aquaculture*, 239: 421-443.
- Choo, P.S. 1995. Withdrawal time for oxytetracycline in red tilapia cultured in freshwater. *Asian Fisheries Science*, 8: 169-176.
- Cipriano, R.C., Bullock, G.L. and Pyle, S.W. 1984. *Aeromonas hydrophila* and motile aeromonad septicemia of fish. U.S Fish and Wildfish Service, *Fish Disease Leaflet*, pp. 68.
- Cipriano, R.C. 2000. *Aeromonas hydrophila* and motile aeromonad septicemias of fish. United states fish and wildlife services, *Fish Disease Leaflet*, 68, pp:25. Revision of Fish Disease Leaflet 40 (1976), "Diseases of freshwaterfishes caused by bacteria of the genera *Aeromonas*, *Pseudomonas*, and *Vibrio*," by S. F. Snieszko and G. L. Bullock.
- Clem, L. W., Sizemore, R. C., Ellsaesser, C. F. and Miller, N. W. 1985. Monocytes as accessory cells in fish immune responses. *Development of Comparative Immunology*, 9: 803–809.

- Couillard, C.M. and Hodson, P.V. 1996. Pigmented macrophage aggregates: a toxic response in fish exposed to bleached-Kraft mill effluent? *Environmental Toxicology and Chemistry*, 15: 1844–1854.
- Cousoa, N., Castroa,R.,Magarin~osb B., Obachc,A. and Lamasa, J. 2003. Effect of oral administration of glucans on the resistance of gilthead seabream to pasteurellosis.*Aquaculture*, 219: 99–109.
- Dalmo, A., Arild, A. Kjerstad, Siri M. Arnesen, Peter S. Tobias and Bøggwald, J. 2000. Bath exposure of Atlantic halibut (*Hippoglossus hippoglossus* L.) yolk sac larvae to bacterial lipopolysaccharide (LPS): Absorption and distribution of the LPS and effect on fish survival. *Fish & Shellfish Immunology*, 10: 107–128.
- Dalmo, R.A. and Bøggwald, J. 1996. Distribution of intravenously and perorally administered *Aeromonas salmonicida* lipopolysaccharide in Atlantic salmon, *Salmo salar* L. *Fish and Shellfish Immunology*, 6: 427– 441.
- Dalmo, R.A. and Bøggwald, J. 2008. b-glucans as conductors of immune symphonies. *Fish Shellfish Immunology*, 25: 384–396.
- Deivasigamani, B. 2008. The immune response catfish, *Mystus gulio*. *Journal of Environmental Biology*, 29: 863–866.
- Defoirdt, T., Sorgeloos, P. and Bossier, P. 2011. Alternatives to antibiotics for the control of bacterial disease in aquaculture. *Current Opinion in Microbiology*, 14(3):251-8.
- Dooley, J.S., Trust and T.J. 1998. Surface protein composition of *Aeromonas hydrophila* strains virulent for fish: identification of a surface array protein. *Journal of Bacteriology*, 170: 499-506.
- Doukas, V., F. Athnassopoulou, E. Karagouni and Dotsika, E. 1998. *Aeromonas hydrophila* infection in cultured sea bass, *Dicentrarchus labrax* (L) and *Puntazzo puntazzo* (Cuvier) from the Aegean Sea. *Journal of Fish Disease*, 21: 317-320.
- Di Luzio, N.R. 1985. Update on the immunomodulating activities of glucans. *Springer Seminar in Immunopathology*, 8:387-400.
- Dixon, B.A., and Lssvoran, G. 1993. Antibacterial drug resistance in *Aeromonas* sp. isolated from domestic goldfish and koi from California. *Journal of World Aquaculture Society*, 24: 102-104.
- Duremdez, R., Al-Marzouk, A., Qasem, J.A., Al-Harbi, A., and Gharaball. H. 2004. Isolation of *S. agalactiae* from cultured silver pomfret, *Pampus argenteus*, in Kuwait. *Journal of Fish Diseases*, 27: 307-310.
- El-Boshy, M. E. El-Ashram, A. M., AbdelHamid, F. M. and Gadalla, H.A. 2010. Immunomodulatory effect of dietary *Saccharomyces cerevisiae*, -glucan

and laminaran in mercuric chloride treated Nile tilapia (*Oreochromis niloticus*) and experimentally infected with *Aeromonas hydrophila*. *Fish and Shellfish Immunology*, 28: 802-808.

- Egidius, E.C. and Anderson, K. 1979. Bath immunization - a practical and non-stressing method of vaccinating sea fanned rainbow trout, *Salmo gairdneri* (Richardson), against vibriosis. *Journal of Fish Disease*, 2: 405-410.
- Ellis A.E. and de Sousa M.A.B. 1974. Phylogeny of the lymphoid system. 1. A study of the fate of circulating lymphocytes in the plaice (*Pleuronectes platessa* L.). *European Journal of Immunology*, 4: 338-343.
- Erridge, C., Bennett-Guerrero, E., and Poxton, I.R. 2001. Structure and function of lipopolysaccharides. *Microbes and Infection*, 4(8): 837-851.
- Esteve, C., Biosca, E.G., and Amaro, C. 1993. Virulence of *Aeromonas hydrophila* and some other bacteria isolated from European eels *Anguilla anguilla* reared in fresh water. *Disease of aquatic organisms*, 16:15-20.
- Evans, J.J., Pasnik, D.J., Klesius, P.H., and Shoemaker, C.A. 2006. Identification and epidemiology of *Streptococcus iniae* and *Streptococcus agalactiae* in tilapia, *Oreochromis* spp. *International Symposium on Tilapia in Aquaculture*, Charles Town, WV, USA, American Tilapia Association, pp. 25-42.
- Evelyn, T.P.T. 1997. *A historical review of fish vaccinology*. In R. Gudding, A. Lillehaug, P.J. Midtlyng and F. Brown (Editors.) *Fish Vaccinology. Development of Biological Standards*, 90: 1-12.
- FAO, Food and Agriculture Organization of the United Nations corporate document repository, 1989. Selected aspects of warm water fish culture. <http://www.fao.org/docrep/field/003/T8389E/T8389E00.htm>. Accessed on 2nd December, 2011.
- FAO, Food and Agriculture Organization of the United Nations. 2003, National aquaculture sector overview: Colombia. Available at http://www.fao.org/fishery/countrysector/naso_colombia/en. (Accessed on 2nd December, 2011).
- FAO (Food and Agriculture Organization of the United Nations). 2004. *Fishstat plus*. <http://www.fao.org/fishery/statistics/software/fishstat/en>. (Accessed on 3rd August, 2011).
- Fitzsimmons, K. 2000. *Tilapia: The most important aquaculture species of the 21st century*. In K. Fitzsimmons and J. Carvalho Filho (Editors.), *Tilapia Aquaculture in the 21st Century* Fifth International Symposium on Tilapia Aquaculture Rio de Janeiro, Brazil, pp. 3-8.
- Fitzsimmons, K. 2003. Tilapia evolution. Growing industry moves from live fish to value-added products. *Global Aquaculture Advocate*, 6(6): 500-52.

- Gado, M.S.M. 1998. Studies on the virulence of *Aeromonas hydrophila* in Nile Tilapia (*Oreochromis niloticus*). *Assiut Veterinary Medicine Journal*, 40: 190-200.
- Galeotti, M. 1998. Some aspects of the application of immunostimulants and a critical review of methods for their evaluation. *Journal of Applied Ichthyology*, 14: 189-199.
- Gannam, A.L., and Schrock, R.M. 2001. *Immunostimulants in fish diets*. In Lim, C., Webster, C.D. (Eds.), *Nutrition and Fish Health*. Haworth Press, Binghamton, New York, USA, pp. 35-266.
- Gbore, F.A., Oginni, O., Adewole, A.M., and Aladetan, J.O. 2006. The effect of transportation and handling stress on hematology and plasma biochemistry in fingerlings of *Clarias gariepinus* and *Tilapia zillii*. *World Journal of Agricultural Science*, 2(2): 208-212.
- González-Serrano, C.J., Santos, J.A., García-López, M.L., and Otero, A. 2002. Virulence markers in *Aeromonas hydrophila* and *Aeromonas veronii biovar sobria* isolates from freshwater fish and from a diarrhoea case. *Journal Applied Microbiology*, 93: 414-419.
- Guttvika, A., Bjørnar, P., Roy A.D., Sigrun, E., Vera, L., and Jarl, B. 2002. Oral administration of lipopolysaccharide to Atlantic salmon (*Salmo salar*) fry. Uptake, distribution, influence on growth and immune stimulation. *Aquaculture*, 214: 35-53.
- Hathaa, M., Vivekanandhan, A.A., Joicea, J. and Christola, G. 2005. Antibiotic resistance pattern of motile aeromonads from farm raised fresh water fish. *International Journal of Food Microbiology*, 98 131-134.
- Harikrisnan, R., Rani, M.N. and Balasundaram, C. 2003. Hematological and biochemical in common carp, *Cyprinus carpio* following herbal treatment for *Aeromonas hydrophila* infection. *Aquaculture*, 222: 41-50.
- Hettiarachchi, D. C. and Cheong, C. H. 1994. Some characteristics of *Aeromonas hydrophila* and *Vibrio* species isolated from bacterial disease outbreaks in ornamental fish culture in Sri Lanka. *Journal of National Science Council Sri Lanka*, 22: 361-369.
- Hibiya, T. 1982. *An Atlas of Fish Histology, Normal and Pathological Features*, pp. 64-65.
- Hoeger, B., Van Den Heuvel, M.R., Hitzfeld, B.C., and Dietrich, D.R. 2004. Effects of treated sewage effluent on immune function in rainbow trout (*Oncorhynchus mykiss*). *Aquatic Toxicology*, 70: 345-355.
- Huizinga, H.W., Esch, G.W., and Hazen, T.C. 1979. Histopathology of red-sore disease (*Aeromonas hydrophila*) in naturally and experimentally infected largemouth bass *Micropterus salmoides*. *Journal of Fish Biology*, 2: 263-277.

- Iliev, D.B., Roach, J.C., Mackenzie, S., Planas, J.V., and Goetz, F.W. 2005. Endotoxin recognition: In fish or not in fish? *FEBS Letters*, 579: 6519-6528.
- Islam, M.T., Mamnur Rashid, M. and Mostafa, K. 2008. Histopathological studies of experimentally infected shing, *Heteropneustes fossilis* with *Aeromonas hydrophila* bacteria. *Progressive Agriculture*, 19(1):89-96.
- Ismail, M.Z. 1992. Background paper on Marine and Freshwater fishes. In Malaysian National Conservation Strategy, pp. 7.
- Janda, J.M. 1991. Recent advances in the study of the taxonomy, pathogenicity, and infectious syndromes associated with the genus *Aeromonas*. *Journal of Clinical Microbiology Reviews*, 4: 397-410.
- Janda, J.M., and Duffey, P.S. 1988. Mesophilic *Aeromonas* in human diseases: Current taxonomy, laboratory identification and infectious disease spectrum. *Review of Infectious Disease*, 10: 980-997.
- Jeney, G., Galeotti, M., Volpatti, D., Jeney, Z. and Anderson, D.P. 1998. Prevention of stress in rainbow trout (*Oncorhynchus mykiss*) fed diets containing different doses of glucan. *Aquaculture*, 154: 1– 15.
- Jeney G., Yin, G., and Ardo', Z. L. 2009. The use of immunostimulating herbs in fish. An overview of research. *Fish Physiology and Biochemistry*, 35: 669–676.
- Joice, A., Shankar, K.M, and Mohan, C.V. 2002. Effect of bacterial biofilm in nursery on growth, survival and resistance to *Aeromonas hydrophila* of common carp, *Cyprinus carpio*. *Journal of Aquaculture in the Tropics*, 17:283-298.
- Kaleeswaran, B., Ilavenil, S., and Ravikumar, S. 2012. Changes in biochemical, histological and specific immune parameters in *Catlacatla* (Ham.) by *Cynodondactylon* (L.). *Journal of King Saud University Science*, 24: 139–152.
- Karunasagar, I., Rosalind, G.M., and Karunasagar, I., 1991. Immunological nresponse of the Indian major carps to *Aeromonas hydrophila* vaccine. *Journal of Fish Disease*, 14: 413-417.
- Ketover, B.P., Young, L.S. and Armstrong, D. 1973. Septicaemia due to *Aeromonas hydrophila*: lineal and immunologic aspects. *Journal of Infectious Disease*, 127: 284-290.
- Klesius, P.H., Shoemaker, C.A., and Evans, J.J. 2008. Streptococcus: A worldwide fish health problem. *8th International Symposium on Tilapia in Aquaculture* Cairo, Egypt, pp. 83-107.
- Kumaran, S., Deivasigamani, B., Alagappan, K.M. and Sakthivel, M. 2010. Infection and immunization trials of Asian seabass (*Latescalcarifer*) against fish pathogen *Vibrio anguillarum*. *Journal of Environmental Biology*, 31: 539-541.

- Lallier, R. and Higgins, R. 1988. Biochemical and toxigenic characteristics of *Aeromonas* spp. isolated from diseased mammals, moribund and healthy fish. *Veterinary Microbiology*, 18: 63-71.
- Lamers C.H.J. and De Haas M.J.H. 1985. Antigen localisation in the lymphoid organs of carp (*Cyprinus carpio*). *Cell and Tissue Research*, 242: 491-498.
- Lara-Flores, M., Olvera-Novoa, M.A., Guzman-Méndez, B.E. and López-Madrid, W. 2003. Use of the bacteria *Streptococcus faecium* and *Lactobacillus acidophilus*, and the yeast *Saccharomyces cerevisiae* as growth promoters in Nile tilapia (*Oreochromis niloticus*). *Aquaculture*, 216: 193-201.
- Lewis, D.H., and Plumb, J.A. 1979. Bacterial disease. In: Plumb, J.A. (Editors.), *Principal Diseases of Farm-raised Catfish*. Alabama Agricultural Experimental Station, Auburn University, Auburn, AL. pp. 15-24.
- Li, A., Yang, W., Hu, J., Wang, W., Cai, T. and Wang, J. 2006. Optimization by orthogonal array design and humoral immunity of the bivalent vaccine against *Aeromonas hydrophila* and *Vibrio fluvialis* infection in crucian carp (*Carassius auratus* L.). *Aquaculture Research*, 37: 813-820.
- Li, P. and Gatlin, D.M. 2005. Evaluation of the prebiotic GroBiotic®-A and brewers yeast as dietary supplements for subadult hybrid striped bass (*Morone chrysops* × *M. saxatilis*) challenged in situ with *Mycobacterium marinum*. *Aquaculture*, 248: 197-205.
- Luderitz, O., Westphal, O., Staub, A.M., and Nikaido, H. 1972. *Isolation and chemical and immunological characterization of bacterial lipopolysaccharides*. In: Weinbaum, G., Kadis, S., Ajl, S.J. (Editors). *Microbial Toxins*. New York: Academic press, pp. 145-233.
- Lunden, T., and Bylund, G. 2000. The influence of in vitro and in vivo exposure to antibiotics on mitogen- induced proliferation of lymphoid cells in rainbow trout (*Oncorhynchus mykiss*). *Fish and Shellfish Immunology*, 10: 395-404.
- MacFaddin, J. F. 2000. *Biochemical Tests for Identification of Medical Bacteria* (3rd. ed). USA: Philadelphia.
- Mackean, D.G., and Mackean, I. 2012. <http://www.biology-resources.com/all-fish.html>. (Accessed on 21st December, 2012).
- Michalin Leislation. Press, C., Dannevig, B.H. and Landsverk, T. 1994. Immune and enzyme histochemical phenotypes of lymphoid and nonlymphoid cells within the spleen and head kidney of Atlantic salmon (*Salmo salar* L.). *Fish and Shellfish Immunology*, 4: 79-93.
- Magnadottir, B. 2006. Innate immunity of fish (overview). *Fish and Shellfish Immunology*, 20: 137-151.

- Magnadottir, B. 2010. Immunological control of fish diseases. *Journal of Marine Biotechnology*, 12: 361–379.
- Matyar, F., Kaya, A. and Dinçer, S. 2007. Distribution and antibacterial drug resistance of *Aeromonas* spp. from fresh and brackish waters in Southern Turkey. *Annals of Microbiology*, 57(3): 443-447.
- Marina, M.P. Camargo Cláudia, and Martinez, B. R. 2007. Histopathology of gills, kidney and liver of a Neotropical fish caged in an urban stream. *Neotropical Ichthyology*, 5(3): 327-336.
- McNulty, S.T., Klesius, P.H., Shoemaker, C.A. and Evans, J.J. 2003. Hematological changes in Nile tilapia (*Oreochromis niloticus*) infected with *Streptococcus iniae* by inoculation. *Journal of World Aquaculture Society*, 34: 418-422.
- Miller, R.M., and Chapman, W.R. 1976. *Epistylis* sp. and *Aeromonas hydrophila* infections in fishes from North Carolina reservoirs. *Progressive Fish Culturist*, 38: 165-168.
- Mitchell, A.J and Plumb, J.A. 1980. Toxicity and efficacy of furazolidone on channel catfish infected experimentally with *Aeromonas hydrophila*. *Journal of Fish Diseases* 3: 93-100.
- Miyata, M., Aoki, T., Inglis, V., Yoshida, T. and Endo, M. 1995. RAPD Analysis of *Aeromonas salmonicida* and *Aeromonas hydrophila*. *Journal of Applied Bacteriology*, 79: 181-185.
- Miyazaki, T., and Kaige, N. 1985. A histopathological study on motile aeromonad disease of crucian carp. *Fish Pathology*, 21: 181-185
- Mohanty, S.N., Swain, S.K. and Tripathi, S.D. 1996. Rearing of catla (*Catla catla* Ham.) spawn on formulated diets. *Journal of Aquaculture Tropical*, 11: 253–258.
- Moral, C.H., Castillo, E.F.D., Fierro, P.L., Cortes, A.V., Castillo, J.A., Soriano, A.C., Salazar, M.S., Peralta, B.R., and Carrasco, G.N. 1998. Molecular characterisation of the *Aeromonas hydrophila* aroA gene and potential use of an auxotrophic and aroA Mutant as a live attenuated vaccine. *Infection Immunology*, 66: 1813-1825.
- Mostafa, K., Islam, M.T. and Mamnur Rashid, M. 2008. Experimental pathogenesis of *Aeromonas hydrophila* bacteria in stringing catfish *Heteropneustes fossilis*. *Bangladesh Journal of Fish Resolution*, 12(1): 27-33.
- Munro, P. B., and A. Birkbeck, T.H. 1995. Comparison of the growth and survival of larval turbot in the absence of culturable bacteria with those in the presence of *Vibrio anguillarum*, *Vibrio alginolyticus* or a marine *Aeromonas* sp. *Applied of Environmental Microbiology*, 61: 4425-4428.

- Murray, P. R. 2003. Manual of clinical microbiology. 7th edition., Washington: ASM Press.
- Najiah, M., Wee, W., H., Lee, Shaharom, F., and Wee, W. 2008. Surveillance of bacteria species in diseased freshwater ornamental fish from aquarium shop. *World Applied Sciences Journal* 3(6): 903-905.
- Nakanishi, Y. and Ototake, M. 1997. Antigen uptake and immune responses after immersion vaccination. In Gudding, R., Lillehaug, A., Midtlyng, P.J. and Brown, F. (eds.). *Fish Vaccinology. Development of Biological Standards Vol. 90*, Basel. p. 59-68.
- Nandlal, S., and Pickering, T. 2004. Tilapia fish farming in Pacific Island countries. *Volume 1. Tilapia Hatchery Operation*. Noumea, New Caledonia: Secretariat of the Pacific Community.
- NCCLS, 1999. National Committee for Clinical Laboratory Standards: Performance standards for antimicrobial susceptibility testing; 9th Informational Supplement, Vol 19, No. 1. NCCLS document M100-S9. Villanova: National Committee for Clinical Laboratory Standards.
- Newman, S.G. 1983. Bacterial vaccines for fish. *Annual Reviews of Fish Disease*, 3: 145- 185.
- Newman, S.G. 1993. *Aeromonas hydrophila*: a review with emphasis on its role in fish disease. In: Antigenes of fish pathogens, (Editors) by D.P. Anderson, M. Dorson and Ph. Dubourget). Collection Fondation Marcel Mérieux, Lyon, pp. 87-117.
- Nikl, L., Evelyn, T.P.T. and Albright, L.J. 1993. Trials with an orally and immersion administrated b-1,3glucan as an immunoprophylactic against *Aeromonas salmonicida* in juvenile chinook salmon *Oncorhynchus tshawytscha*. *Disease of Aquatic Organisms*, 17:191-196.
- Norimatsu, M., and Ono, T. 1999. Lipopolysaccharide-induced apoptosis in swine lymphocytes *in vivo*. *Infection and Immunity*, 63: 1122–1126.
- Nya, E.J. and Austin, B. 2010. Use of bacterial lipopolysaccharide (LPS) as an immunostimulant for the control of *Aeromonas hydrophila* infections in rainbow trout *Oncorhynchus mykiss* (Walbaum). *Journal of Applied Microbiology*, 108: 686–694.
- O'Donnell, G.B., Smith, P.R., Kilmartin, J.J. and Moran, A.P. 1994. Uptake and fate of orally administered bacterial lipopolysaccharide in brown trout (*Salmo trutta*). *Fish and Shellfish Immunology*, 4: 285– 299.
- Oliva-Teles, A. and Gonçalves, P. 2001. Partial replacement of fishmeal by brewers yeast *Saccharomyces cerevisiae* in diets for sea bass *Dicentrarchus labrax* juveniles. *Aquaculture*, 202: 269–278.

- Orozova, P., Chikova, V., Kolarova, V., Nenova, R., Konovska, M. and Najdenski, H. 2008. Antibiotic resistance of potentially pathogenic *Aeromonas* strains. *Trakia Journal of Science*, 6(1): 71–77.
- Ortuno, J., Cuesta, A., Rodríguez, A., Esteban, M.A., and Mesegure, J. 2002. Oral administration of yeast, *Saccharomyces cerevisiae*, enhances the cellular innate immune response of gilthead seabream (*Sparus aurata*, L.). *Veterinary of Immunology and Immunopathology*, 85: 41–50.
- Palumbo, S.A., Bencivengo, M.M., Corral, F.D., Williams, A.C. and Buchanan, R. L. 1989. Characterization of the *Aeromonas hydrophila* group isolated from retail foods of animal origin. *Journal of Clinical Microbiology*, 27: 854-859.
- Paniagua, C., Rivero, O., Anguita, J., and Naharro, G. 1990. Pathogenicity factors and virulence for rainbow trout (*Salmo gairdneri*) of motile *Aeromonas* spp. isolated from a river. *Journal of Clinical Microbiology*, 28: 350-355.
- Pepels, P.P.L.M., Wendelaar Bonga, S.E., and Balm, P.H.M. 2004. Bacterial lipopolysaccharide (LPS) modulates corticotropin-releasing hormone (CRH) content and release in the brain of juvenile and adult tilapia (*Oreochromis mossambicus*; Teleost). *The Journal of Experimental Biology*, 207: 4479-4488.
- Pérez-Casanova, J.C., Hamoutene, D., Volkoff, H., Mabrouk, G., Samuelson, S., and Burt, K. 2010. Effect of injection of lipopolysaccharides from *Aeromonas salmonicida* on some aspects of cod (*Gadus morhua*) immunity and appetite hormones. *Canada Technical Report of Fisheries Aquatic Sciences*, No 2878: 1-11.
- Plumb, J.A. and Shoemaker, C. 1995. Effect of temperature and salts concentration on latent *Edwardsiella ictaluri* infections in channel catfish. *Disease of Aquatic Organisms*, 21: 171-175.
- Popoff, M. 1984. *Genus III: Aeromonas*. In Kreig, N.R., and Holt, J.G. (Editors). *Bergey's Manual of Systematic Bacteriology*, Williams and Wilkins, Baltimore, London, pp. 545-7.
- Press, C.M., and Lillehaug, A. 1995. *Vaccination in European Salmonid aquaculture. A review of practices and prospects British*. In: Neter Jou, Thomson, J.C., Gardono, R., Newman, S.G., Kay, W.W., (Editors) 1991. Surface disorganized, attenuated mutants of *Aeromonas salmonicida* as furunculosis live vaccines. *Microbiology Pathology*, 11: 85-90.
- Press, C., Evensen O., Reitan, L.J. and Landsverk, T. (1996). Retention of furunculosis vaccine components in Atlantic salmon, *Salmo salar*, following different routes of administration. *Journal of Fish Diseases*, 19: 215–224.
- Press, C.M. 1998. *Immunology of fishes*. In: Handbook of vertebrate immunology (Eds.: P.P. Pastoret, P. Griegel, H. Bazin and A. Govaerts). Academic Press, San Diego, pp. 3-62.

- Quentel, C. and Vigneulle, M. 1997. Antigen uptake and immune responses after oral vaccination: In Gudding, R., Lillehaug, A., Midtlyng, P.J. and Brown, F. (Editors). *Fish Vaccinology. Development of Biological Standards, Volume 90*, pp. 69-78.
- Raa, J., Rorstad, G., Engstad, R., and Robertsen, B. 1992. *The use of immunostimulants to increase resistance of aquatic organisms to microbial infections*. In Shariff, I.M., Subasinghe, R.P., Arthur, J.R. (Editors.), *Diseases in Asian Aquaculture*. Fish Health Section, Asian Fisheries Society, Manila, Philippines, pp. 39-50.
- Randy White, M., and Swann, L. 1989. Diagnosis and Treatment of "Aeromonas hydrophila" Infection of Fish. A Guide to Approved Chemicals in Fish Production and Fishery Resource Management, University of Arkansas Cooperative Extension Service. *Fact Sheet, AS-461*.
- Rao, Y.V., Das, B.K., Jyotirmayee, P. and Chakrabarti, R. 2006. Effect of *Achyranthes aspera* on the immunity and survival of *Labeo rohita* infected with *Aeromonas hydrophila*. *Fish and Shellfish Immunology*, 20: 263-273.
- Rathore, G., Swaminathan, T. R., Abidi, R., Mahanta, P.C. and Kapoor, D. 2005. Isolation and characterization of motile aeromonads from aquatic environment. *Indian Journal of Fisheries*, 52(2): 241-248.
- Rehulka, J., and Adamec, V. 2004. Red blood cell induce for Rainbow trout (*Onchorhynchus mykiss*) reared in cage and raceway culture. *Acta Veterinaria Brno*, 73: 105-114.
- Rey, V.G., and Guerrero, G.A. 2007. Characterization of blood cells and hematological parameters in *Cichlasoma dimerus*. *Journal of Tissue Cell*, 39: 151-160.
- Rietschel, E.T., and Brade H. 1992. Bacterial endotoxins. *Scientific American*, 267(2): 54-61.
- Rijkers, G.T., R. Van Oosterom and Van Muiswinkel. 1981. The immune system of cyprinid Oxytetracycline and the regulation of humoral immunity in carp (*Cyprinus carpio*). *Veterinary Immunology and Immunopathology*, 2: 281-290.
- Robertsen, B. 1999. Modulation of the non-specific defence of fish by structurally conserved microbial polymers. *Fish Shellfish Immunology*, 9: 269-290.
- Roberts, R.J., Willoughby, L.G., and Chinabut, S. 1993. Mycotic aspects of epizootic ulcerative syndrome (EUS) of Asian fishes. *Journal of Fish Disease*, 16: 169-183.
- Roberts, R.J. 2001. *The bacteriology of teleosts*. In R.J., Roberts (Editors), *Fish Pathology*, W.B. Saunders, Philadelphia, pp. 315-321.

- Rohlenová, K., Morand, S., Hyršl, P., Tolarová, S., Flajšhans, M., and Šimková A. 2011. Are fish immune systems really affected by parasites? An immunoeological study of common carp (*Cyprinus carpio*). *Parasites and Vectors*, 4(120): 1756-4120.
- Rombout, J.H., Huttenhuis, H.B.T., Picchiatti, S., and Scapigliati, S. 2005. Phylogeny and ontogeny of fish leucocytes. *Fish and Shellfish Immunology*, 19: 441–455.
- Rukyani, A. 1994. *Status of epizootic ulcerative disease in Indonesia*. In: Roberts, R.J., Cambell, B., MacRae, I.H. (Editors.), Proceedings of the ODA regional seminar on epizootic ulcerative syndrome. Aquatic Animal Health Research Institute, Bangkok, pp. 25-27.
- Sabur, M.A. 2006. *Studies on the ecology of the pathogenic bacteria Aeromonas hydrophila in indigenous and exotic carps under polyculture condition*. Unpublished doctoral of philosophy dissertation. Department of Aquaculture, Bangladesh Agricultural University, Mymensingh, Bangladesh, pp.163.
- Saeed, M.O. and Plumb, A.J. 1986. Immune response of channel catfish to lipopolysaccharide and whole cell *Edwardsiella ictaluri* vaccines. *Disease Aquatic Organisms*, 2: 21–25.
- Sakai, M. 1998. Current research status of fish immunostimulants. *Aquaculture* 172: 63–92.
- Salati, F., Watanabe, K., Kawai, K. and Kusuda, R. 1989. Immune response of ayu against *Vibrio anguillarum* lipopolysaccharide. *Nippon Suisan Gakkaishi*, 55: 45–49.
- Santos, Y., Bandin, I., and Toranzo, A.E. 1996. Immunological analysis of extracellular products and cell surface components of motile *Aeromonas* isolated from fish. *Journal of Applied Bacteriology*, 81: 585-593.
- Scott, A.L. and Rogers, W.A. 1981. Hematological effects of prolonged sublethal hypoxia on channel catfish *Ictalurus punctatus* (Rafinesque). *Journal of Fish Biology*, 18: 591–601.
- Secombes, C. J., Manning M. J. and Ellis A. E. 1982. The effect of primary and secondary immunization on the lymphoid tissues of the carp, *Cyprinus carpio* L. *Journal of Experimental Zoology*, 220: 277-287.
- Secombes, C.J. 1996. *The nonspecific immune system: cellular defenses*. In Iwama, G., Nakanishi, T. (Editors.), *The Fish Immune System. Organism, Pathogen and Environment*. Academic Press, San Diego, CA, pp. 63–101.
- Selvaraj, V., Sampath, K. and Sekar, V. 2009. Administration of lipopolysaccharide increases specific and non-specific immune parameters and survival in carp (*Cyprinus carpio*) infected with *Aeromonas hydrophila*. *Aquaculture*, 286 176–183.

- Selvaraj, V., Sampath, K. and Sekar, V. 2005. Administration of yeast glucan enhances survival and some non-specific and specific immune parameters in carp (*Cyprinus carpio*) infected with *Aeromonas hydrophila*. *Fish and Shellfish Immunology*, 19: 293–306.
- Shariff, M., Gopinath, N., Chua, F.H.C., and Wong, Y.G. 1996. The use of chemicals in aquaculture in Malaysia and Singapore. In J.R., Arthur, C.R., Lavilla-Pittago and R.P., Subasinghe (Editors), *Proceeding of The Meeting on The Use of Chemicals in Aquaculture in Asia*, pp. 127-140.
- Shayo, S.D., Mwita, C.J., and Hosea, K. 2012. Ulcerative *Aeromonas* Infections in *Tilapia* (Cichlidae: Tilapiini) from Mtera Hydropower Dam, Tanzania. *Scientific Report*, 1:1.
- Shelton, W.L., and Popma, T.J. 2006. *Biology: Tilapia biology, culture and nutrition*. New York, NY, USA: Food Production Press.
- Sherwood, E.R., Williams, D.L., McName, R.B., Jones, E.L., Browder, I.W. and Di Luzio, N.R. 1987. Enhancement of interleukin-1 and interleukin-2 production by soluble glucan. *International Journal of Immunopharmacology*, 9:261-267.
- Shoemaker, C.A., and Klesius, P.H. 1997. *Streptococcal disease problem and control: A review*. Tilapia Aquaculture Ithaca, NY, USA. Northwest Regional Aquaculture Engineering Service.
- Shnyra, A., Luchi, M., and Morrison, D.C. 2000. *Preparation of endotoxin from pathogenic gram negative bacteria*. In: Evans, T.J. (Editors). *Methods in Molecular Medicine* (Vol. 36 Septic Shock Methods and Protocols). Totowa: Humana press, pp. 13-25.
- Shome, R., and Shome, R. 1999. A typical chronic form of *Aeromonas hydrophila* infection in Indian major carp, *Catla catla*, from Andaman. *Current Science*, 76:1188-1190.
- Shotts, E.B., Gaines, J.L., Jr., Martin, I., and Prestwood, A.K. 1972. *Aeromonas*-induced deaths among fish and reptiles in an eutrophic inland lake. *Journal of The American Veterinary Medicine Association*, 161: 603-607.
- Soltanian, S., Stuyven, E., Cox, E., Sorgeloos, P. and Bossier, P. 2009. Beta-glucans as immunostimulant in vertebrates and invertebrates. *Critical Reviews in Microbiology*, 35(2): 109–138.
- Somsiri, T., Chinabut, S., Phawichien, K., Soontornwit, S., and Prapaiwong, N. 1999. Impact of oxytetracycline on *Rana tigrina*. *Asian Fisheries Science*, 12: 361-369.
- Son, R., Rusul, G., Sahilah, A.M., Zainuri, A., Raha, A.R. and Salmah, I. 1997. Antibiotic sensitivity and plasmid profile of *Aeromonas hydrophila* isolates

- from cultured fish, *Telapia (Telapia mosambica)*. *Letters in Applied Microbiology*, 24: 479-482.
- Simin Rezania, Amirmozaffari, N., Tabarraei, B., Jeddi-Tehrani, M., Zarei, O., Alizadeh, O., Masjedian, F., and Zarnani, A.H. 2011. Avicenna Extraction, Purification and Characterization of Lipopolysaccharide from *Escherichia coli* and *Salmonella typhi*. *Journal of Medical Biotechnology*, 3(1): 3-9
- Siti-Zahrah, A., Misri, S., Padilah, B., Zulkaflī, R., Kua, B.C., Azila A. and Rimatulhana, R. 2004. Pre-disposing factors associated with outbreak of Streptococcal infection in floating cage-cultured red tilapia in reservoirs. Abstracts of the 7th Asian Fisheries Forum 04, The Triennial Meeting of The Asian Fisheries Society 30th Nov-4th Dec 2004, Penang, Malaysia, pp.129.
- Siwicki, A.K., Anderson, D.P., and Rumsey, G.L. 1994. Dietary intake of immunostimulants by rainbow trout affects non-specific immunity and protection against furunculosis. *Veterinary of Immunology and Immunopathology*, 41: 125-39.
- Snieszko, S.F., and Bullock, G.L. 1974. Diseases of freshwater fish caused by the genera *Aeromonas*, *Pseudomonas*, and *Vibrio*. *Fish Disease Leaflet No. 40*, U.S. Department of the Interior, pp. I-10.
- Sommerset, I., Bjørn, K., Eirik, B., and Petter, F. 2005. Review Vaccines for fish in aquaculture. *Expert Review of Vaccines*, 4(1): 89-101.
- Subramanian, S., Mackinnon, S.L., and Ross, N.W. 2007. A comparative study on innate immune parameters in the epidermal mucus of various fish species. *Comparative Biochemistry and Physiology Part B*, 148: 256–263.
- Subramanian, S., Ross, N.W., and Mackinnon, S.L. 2008. Comparison of antimicrobial activity in the epidermal mucus extracts of fish. *Comparative Biochemistry and Physiology Part B*, 150: 85–92.
- Suchanit, N., Kunihiko, F., Masato, E., Masashi, M., and Takayuki, K. 2010. Immunological effects of glucan and *Lactobacillus rhamnosus* GG, a probiotic bacterium, on Nile tilapia *Oreochromis niloticus* intestine with oral *Aeromonas* challenges. *Fish Science*, 76: 833–840
- Stevenson, R.M.W. 1998. Vaccination against *Aeromonas hydrophila*. In Ellis, A.E. (Editors.), *Fish vaccination*. Academic press, London, pp. 112-123.
- Swain, S.K., Rangacharyulu, P.V., Sarkar, S. and Das, K.M. 1996. Effect of a probiotic supplement on growth, nutrient utilization and carcass composition in mrigal fry. *Aquaculture*, 4: 29–35.
- Swain, P., Nayak, S.K., Nanda, P.K., and Dash, S. 2008. Biological effects of bacterial lipopolysaccharide (endotoxin) in fish: A review. *Fish and Shellfish Immunology*, 25: 191-201.

- Swam, L., and White. M.R. 1989. Aquaculture extension, Illinois landnanas sea grant program. Purdue University.
- Takase, Y., Shimizu, N., and Kubota, S. 1968. The absorption and distribution of a chemotherapeutic agent, P-7138 in fishes. *Bulletin of the Japanese Society Scientific Fisheries*, 34: 1118-1123.
- Taoka, Y., Maeda, H., Jo, J.Y., Jeon, M.J., Bai, S.C., Lee, W.J., Yuge, K. and Koshio, S. 2006. Growth, stress tolerance and non-specific immune response of Japanese flounder *Paralichthys olivaceus* to probiotics in a closed recirculating system. *Fisheries Science*, 72: 310–321.
- Taufik, P. and Wong, S.Y. 1990. The pathogenic of bacteria of paddy field catfishes (*Clarias batrachus* (L.) and *C. Macrocephalus Gunther*) from Kedah and Perak, west Malaysia. *Asian fisheries science*, 3: 361-368.
- Teh, S.J., Adams, S.M., and Hinton, D.E. 1997. Histopathological biomarkers in feral freshwater fish populations exposed to different types of contaminant stress. *Aquatic Toxicology*, 37: 51-70.
- Terry, C.H., Jenifer, L.C., and Stephen, A.S. 2000. Hematology and plasma chemistry reference intervals for cultured tilapia (*Oreochromis Hybrid*). *Veterinary of Clinical Pathology*, 29(1): 7-12.
- Thampuran, N. and Surendran, P. K. 1995. Incidence of motile aeromonads in marine environment, fishes and processed fishery products (Technological advancements in fisheries). In proceedings of the National Symposium on Technological Advancements in Fisheries and its Impact on Rural Development held at Cochin by School of Industrial Fisheries, Cochin University of Science and Technology during December 5 to 7, pp. 352-358.
- Thampuran, N., Surendran, P.K., Mukundan, M.K., and Gopakumar, K. 1995. Bacteriological studies on fish affected by epizootic ulcerative syndrome (EUS) in Kerala, India. *Asian Fisheries Science*, 8: 103-111.
- Thrope, J.E., and Roberts, R.J. 1972. An aeromonad epidemic in the brown trout (*Salmo tautta*). *Journal of Fish Biology*, 4: 441-451.
- Topic Popovic, N., Teskeredzic, E., Strunjak – Perovic, I. and Coz – Rakovac, R. 2000. *Aeromonas hydrophila* isolated from wild freshwater fish in Croatia. *Veterinary Research Communications*, 24: 371 – 377.
- Tovar, D., Zambonino-Infante, J.L., Cahu, C., Gatesoupe, F.J., Vázquez-Juárez, R. and Lésel, R. 2002. Effect of live yeast incorporation in compound diet on digestive enzyme activity in sea bass larvae. *Aquaculture*, 204: 113–123.
- Trust, T.J., Bull, M.L., Currie, B.R. and Buckley, J.T. 1974. Obligate anaerobic bacteria in the gastrointestinal microflora of the grass carp (*Ctenopharyngodon idella*), goldfish (*Carassius auratus*), and rainbow trout (*Salmo gairdneri*). *Journal of Fish Research Board Canada*, 36: 1174-1179.

- Tsujii T., and Seno, S. 1990. Melano-macrophage centers in the aglomerular kidney of the sea horse (teleosts): morphologic studies on its formation and possible function. *The Anatomical Record*, 226: 460–470.
- Uchiyama, T. 1982. Modulation of immune response by bacterial lipopolysaccharide LPS roles of macrophages and T cell in vitro adjuvant effects of LPS on antibody response to T cell dependent and T cell independent antigens. *Microbiology Immunology*, 26: 213–225.
- Van Muiswinkel, W.B., Lamers, C. H.J. and Rombout, J.H.W.M. 1991. Structural and functional aspects of the spleen in bony fish. *Research in Immunology*, 142: 362-366.
- Velji, M.I., Albright, L.J. and Evelyn, T.P. 1990. Protective immunity in juvenile Coho salmon *Oncorhynchus kisutch* following immunization with *Vibrio ordalii* lipopolysaccharide or from exposure to live *V. ordalii* cells. *Disease Aquatic Organisms*, 9: 25–29.
- Ventura, M.T., and Grizzle, J.M. 1988. Lesions associated with natural and experimental infections of *Aeromonas hydrophila* in channel catfish *Ictalurus punctatus* (Rafinesque). *Journal of Fish Disease*, 11: 397-407.
- Waché, Y., Auffray, F., Gatesoupe, F.J., Zambonino, J., Gayet, V., Labbé, L., and Quentel, C. (2006). Cross effects of the strain of dietary *Saccharomyces cerevisiae* and rearing conditions on the onset of intestinal microbiota and digestive enzymes in rainbow trout, *Onchorhynchus mykiss*, fry. *Aquaculture*, 258, 470–478.
- Wester, P.W. and Canton, J.H. 1991. The usefulness of histopathology in aquatic toxicity studies. *Comparative Biochemistry and Physiology (C)*, 100: 115-117.
- Wolf, M. 2004. Use and abuse of antibiotics. Time for its evaluation, beyond the human being. *Medical Journal of Chile*, 132: 909- 911.
- Wolke, R.E. 1992. Piscine macrophage aggregate: A review. *Annual Review of Fish Disease*, 2: 91–108.
- Whitman, K.A. and MacNair, N.G. 2004. *Finfish and shellfish bacteriology: Manual Techniques and Procedures*. USA: Iowa State Press.
- Whyte, S.K. 2007. The innate immune response of finfish: A review of current knowledge. *Fish and Shellfish Immunology*, 23: 1127–1151.
- Xu, D. and Rogers, W.A. 1995. Oxytetracycline residue in the muscle of Nile tilapia. *Asian Fisheries Science*, 8: 113-120.
- Yambot, A.V. and Inglis, V. 1994. *Aeromonas hydrophila* isolated from Nile tilapia (*Oreochromis niloticus*) with “eye disease”. In International Symposium on Aquatic Animal Health, University of California, School of Veterinary Medicine, Davis, CA. p. 103.

- Yambot, A.V. 1998. Isolation of *Aeromonas hydrophila* from *Oreochromis niloticus* during Fish Disease Outbreaks in the Philippines. *Asian Fisheries Science*, 10: 347-354.
- Yanong, R.P.E., and Francis-Floyd, R. 2002. Streptococcal infections of fish. Report from University of Florida. Series from the Department of Fisheries and Aquatic Sciences, Florida Cooperative Extension Service, Institute of Food and Agricultural Sciences, University of Florida.
- Yano, T., Matsuyama, H. and Mangindaan, R.E.P. 1991. Polysaccharide-induced protection of carp, *Cyprinus carpio* L. against bacterial infection. *Journal of Fish Disease*, 14: 577-582.
- Yardimci, B. and Aydin, Y. 2011. Pathological findings of experimental *Aeromonas hydrophila* infection in Nile tilapia (*Oreochromis niloticus*). *Ankara Üniversitesi Veteriner Fakültesi Dergisi*, 58: 47-54,
- Zuasti, A., Jara, J.R., Ferrer, C., and Solano, F. 1989. Occurrence of melanin granules and melano synthesis in the kidney of *Sparus auratus*. *Pigment Cell Research*, 2: 93-99.