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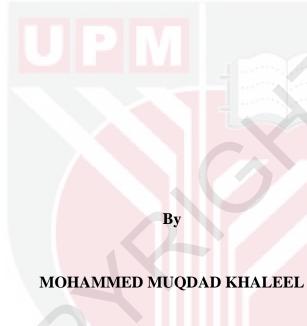
ESTABLISHMENT OF HAEMORRHAGIC SEPTICAEMIA IN MICE INOCULATED WITH WATER CONTAMINATED WITH PASTEURELLA MULTOCIDA TYPE B:2

MOHAMMED MUQDAD KHALEEL

FPV 2014 16



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Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfillment of the Requirements for the Degree of Master of Veterinary Science

March, 2014

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in Fulfillment of the requirement for the degree of Master of Veterinary Science

ESTABLISHMENT OF HAEMORRHAGIC SEPTICAEMIA IN MICE USING WATER CONTAMINATED WITH *PASTEURELLA MULTOCIDA TYPE B: 2*

By

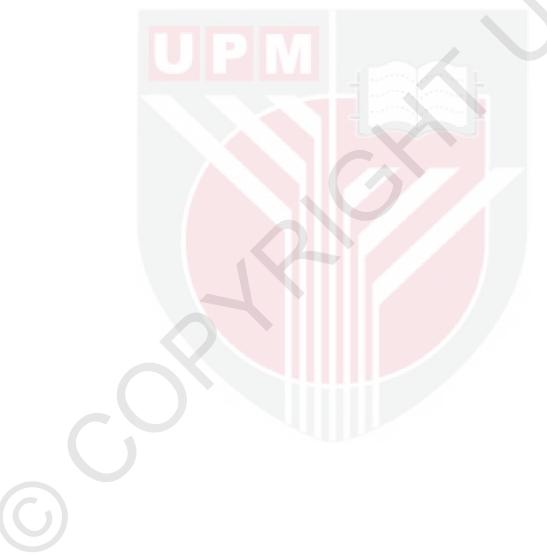
MOHAMMED MUQDAD KHALEEL

March, 2014

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Haemorrhagic septicaemia (HS) is an infectious disease of cattle and buffalo inflicted by serotypes B: 2 and E: 2 of Pasteurella multocida in Asian and African countries respectively and characterised by an acute, highly fatal septicaemia with high morbidity and mortality. Therefore, the present study aims at evaluating the presence of P. multocida type B: 2 in various organs using Polymerase chain reaction (PCR), and host cell response in mice infected with *Pasteurella multocida* type B: 2 in contaminated river water with mice carcasses kept for 24, 48 and 72 hours. This study postulated that the outbreak of HS among buffaloes and cattle could be due to the consumption of river water contaminated with infected HS carcasses and the aerosol routes could perhaps be a readily available route for effective vaccine administration and heightened immunity in animals considering the progressive responses of APPs through this route. Sixty five healthy BALC male mice of eight to ten weeks old were used in this study. The wild-type P. multocida B: 2 used in this study were obtained from stock culture. The river water was obtained from Hulu Langat and was cultured to confirm that it was free from P. multocida type B: 2, Fifteen mice were initially inoculated with 1.0 mL of P. multocida type B: 2 intraperitoneally. Five infected mice carcasses were placed in each tank for 24, 48 and 72 hours and 1ml of the contaminated river water containing Pasteurella multocida type B: 2 were inoculated via the intraperitoneally and aerosol routes while, 0.4 ml of Pasteurella multocida type B: 2 was inoculated orally into five mice in each group and after 48 hours the mice were euthanized by cervical dislocation. Blood samples were collected directly from the heart into plain tubes from the moribund animals to obtain serum for the analysis of serum amyloid A (SAA) and haptoglobin. The fourth group is the control group comprised of five mice and was inoculated with 1.0 mL of sterile Phosphate Buffered Saline (PBS) pH7. Post mortem was conducted and the brain, kidney, heart, spleen, lungs and liver were sampled for histopathology. All the organs were cultured on the blood agar and incubated at 37°C for 24 hours. PCR was performed on the organs from the mice. The concentration of SAA increased significantly (p<0.05) in the mice that were infected with the contaminated river water for 72 hours followed intraperitoneal group compared to the control, oral and aerosol group. There were also significant increase (p<0.05) in the concentrations of Hp in the group of mice that were infected with contaminated river water for 24 hours intraperitoneally relative to the control, oral and the aerosol groups. The PCR results revealed the presence of P.

multocida from the brain, kidney, heart, spleen, lung and liver in the group of mice from the intraperitoneal, oral and aerosol groups. The river water kept for 24 hours was positive for *P. multocida* in the intraperitoneal, oral and the aerosol groups. The river water kept for 48 and 72 hours were positive for *P. multocida* in the intraperitoneal and oral groups. The cellular changes in the vital organs include thrombosis, inflammatory cells, hemorrhage, degeneration and necrosis. In the brain, heart, kidney, liver, lung and spleen, the degeneration and necrosis was significantly high (p < 0.05) compared to the other cellular changes in the mice carcasses infected with *P. multocida* in contaminated river water kept for 72 hours. In conclusion, mice model could be used to enhance the understanding of the progression of the disease and control of the natural disease through the various routes of the disease transmission and contaminated river water infected with HS carcasses could be a potential source of infection if the carcasses are not removed from the river water immediately.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains Veterinar

PENGHASILAN HAWAR BERDARAH DI DALAM MENCIT DENGAN MENGGUNAKAN AIR TERCEMAR DENGAN *Pasteurella multocida* JENIS B: 2

Oleh

MOHAMMED MUQDAD KHALEEL

Mac, 2014

Pengerusi: Faez Firdaus Jesse Abdullah, Ph.D. Fakulti: PerubatanVeterinar

Hawar berdarah merupakan penyakit berjangkit bagi lembu dan kerbau disebabkan oleh bakteria Pasteurella multocida jenis serotaip B:2 dan E:2 dinegara Asia dan Afrika masing-masing dan penyakit ini mempunyai ciri-ciri akut, septisemia berbahaya, dengan morbiditi dan kematian yang tinggi. Oleh itu, kajian ini bertujuan untuk melihat kehadiran baketria P. multocida B: 2 dalam pelbagai organ dengan menggunakan aplikasi reaksi rantai polymerase (PCR), dan tindak balas sel tuan rumah dalam mencit yang dijangkiti dengan bakteria Pasteurella multocida B: 2 dalam air sungai yang tercemar dengan bangkai mencit disimpan selama 24, 48 dan 72 jam. Kajian ini merumuskan bahawa wabak hawar berdarah dikalangan kerbau dan lembu mungkin disebabkan oleh penggunaan air sungai tercemar dengan jangkitan bangkai hawar berdarah dan laluan aerosol boleh menjadi salah satu laluan sedia ada untuk pentadbiran vaksin yang berkesan dan imuniti yang memuncak pada haiwan. Enam puluh lima ekor mencit jantan BALC yang sihat berumur antara lapan hingga sepuluh minggu digunakan dalam kajian ini. Bakteria jenis liar *P. multocida* B: 2 yang digunakan dalam kajian ini diperolehi daripada stok kultur. Air sungai yang digunakan dalam kajian ini diperolehi dari Hulu Langat dan dikultur untuk mengesahkan bahawa ia adalah bebas daripada bakteria P. multocida jenis B: 2, Lima belas ekor mencit pada mulanya disuntik dengan 1.0 mL P. multocida Jenis B: 2 melalui intraperitoneum. Lima bangkai mencit yang dijangkiti dengan telah hawar berdarah diletakkan di dalam setiap tangki air sungai selama 24, 48 dan 72 jam and1ml air sungai yang dicemari dengan Pasteurella multocida jenis B: 2 telah disuntik melalui intraperitoneum dan laluan aerosol manakala, 0.4 ml air sungai tercemar telah disuntik secara lisan kepada lima ekor mencit dalam setiap kumpulan, dan selepas 48 jam kesemua mencit telah ditakai dengan cara terkehel leher. Sampel darah yang diambil secara langsung daripada jantang ke dalam tiub kosong daripada mencit yang menunjukkan tanda hampir menemui ajal bagi mendapatkan serum untuk analisis serum amiloid A (SAA) dan haptoglobin. Kumpulan keempat adalah kumpulan kawalan terdiri daripada lima ekor mencit yang telah disuntik dengan 1.0 mL steril larutan penampan fosfat (PBS) pH7. Bedah siasat telah dijalankan dan organ seperti otak, buah pinggang, jantung, limpa, paru-paru dan hati telah disampel untuk tujuan kajian histopatologi . Semua organ-organ telah dikultur pada agar darah dan dieram pada 37°C selama 24 jam. PCR telah dijalankan ke atas organ-organ tersebut . Kepekatan SAA meningkat dengan ketara (p < 0.05) pada mencit yang dijangkiti dengan air sungai yang tercemar selama 72 jam diikuti kumpulan 'intraperitoneal' berbanding dengan kawalan, kumpulan lisan dan 'aerosol'. Terdapat juga peningkatan yang signifikan (p < 0.05) dalam kepekatan Hp

dalam kumpulan mencit yang dijangkiti dengan air sungai yang tercemar selama 24 jam 'intraperitoneally' berbanding dengan kawalan, lisan dan kumpulan aerosol. Keputusan PCR mendedahkan kehadiran P. multocida pada organ seperti, buah pinggang, jantung, limpa, paru-paru dan hati pada mencit dari kumpulan intraperitoneal, lisan dan aerosol. Air sungai yang disimpan selama 24 jam adalah positif bagi P. multocida bagi kumpulan 'intraperitoneal', lisan dan aerosol. Air sungai disimpan selama 48 dan 72 jam adalah positif bagi P. multocida bagi kumpulan 'intraperitoneal' dan lisan. Perubahan struktur sel dalam organ-organ adalah seperti trombosis, sel-sel radang, pendarahan, degenerasi dan nekrosis . Bagi organ otak, jantung , buah pinggang , hati, paru-paru dan limpa, kemerosotan dan nekrosis adalah ketara tinggi (p < 0.05) berbanding dengan perubahan sel lain dalam kumpulan yang bangkai mencit dijangkiti P. multocida dalam air sungai yang tercemar disimpan selama 72 jam. Kesimpulannya, model mencit boleh digunakan untuk meningkatkan pemahaman perkembangan penyakit dan kawalan penyakit semula jadi melalui pelbagai laluan jangkitan penyakit dan air sungai yang tercemar dijangkiti bangkai hawar berdarah boleh menjadi salah satu sumber yang berpotensi jangkitan wabak penyakit hawar berdarah jika bangkai tersebut tidak dikeluarkan dari air sungai serta-merta.

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I certify that a Thesis Examination Committee has met on 15th March, 2014 to conduct the final examination of Mohammed Muqdad Khaleel on his thesis entitled "establishment of haemorrhagic septicaemia in mice inoculated with water contaminated with *Pasteurella multocida* type B:2" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U. (A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Veterinary Science.

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TABLE OF CONTENT

			Page
ABS'	TRACT		ii
ABS'	ГКАК		iv
ACK	NOWLEDGEMENTS		V
APP	ROVAL		vii
DEC	LARATION		ix
LIST	COF TABLES		xii
LIST	COF FIGURES		xiv
LIST	COF ABBREVIATIONS		XV
	PTER		
1	INTRODUCTION		2
•			
2	LITERATURE REVIEW		3
	2.1 Haemorrhagic septicae	emia (HS)	3 3 5
	2.2 Pathogenesis		
	2.3 Diagnosis		5
	2.4 Acute Phase Proteins		6
	2.5 Pathology		7
3	MATERIALS AND MET	HODS	8
5	3.1 Mice		8
	3.2 River water		8
	3.3 Inoculum		8
		l design in mouse model	8
	3.3.2 Histopatholo		11
	3.3.3 Clinical signs		11
	3.3.4 Lesions scori		11
	3.4 Acute Phase Proteins (11
		on of Mouse Haptoglobin(HP)	12
		on of Mouse Serum Amyloid A (SAA)	12
	3.5 Sampling and culture	I OI MOUSE SEIUIII AIIIYIOIU A (SAA)	12
	3.5.1 DNA extract	ion	12
	3.5.2 PCR conditio		13
	3.5.3 Primer design		13
	3.5.4 Agarose gel		13
	3.5.5 Electrophore	-	13 14
	5.5.5 Electrophore	515	14
4	ACUTE PHASE PROTE	IN RESPONSES IN MICE INFECTED) 15
		CONTAMINATED BY PASTEURELLA	L
	MULTOCIDA TYPE B:2		
	4.1 Introduction		15
	4.2 Material and methods		15
	4.3 Results		15
	4.4 Discussion		19

5	POLYMERASE CHAIN REACTION DETECTION OF	20
	PASTEURELLA MULTOCIDA TYPE B:2 IN MICE INFECTED	
	WITH CONTAMINATED RIVER WATER	
	5.1 Introduction	20
	5.2 Material and methods	20
	5.3 Results	21
	5.4 Discussion and conclusion	28
6	PATHOLOGICAL CHANGES IN MICE INFECTED WITH RIVER WATER CONTAMINATED BY PASTEURELLA	29
	MULTOCIDA TYPE B:2	
	6.1 Introduction	29
	6.2 Materials and methods	29
	6.3 Results	30
	6.4 Discussion	40
7	GENERAL DISCUSSION AND CONCLUSION	44
	7.1 General discussion and conclusion	44
REI	FERENCES	49
BIO	DATA OF STUDENT	50
LIS	T OF PUBLICATIONS	51

 \bigcirc

LIST OF TABLES

Table

- 3.1 The microscopic lesions scored for the organs examined
- 5.1 Identification of P. multocida type B:2 in vital organs inoculated via different routes with river water contaminated with infected mice carcasses kept for 24 hours by using PCR
- 5.2 Identification of P. multocida type B:2 in vital organs inoculated via different routes with river water contaminated with infected mice carcasses kept for 48 hours by using PCR
- 5.3 Identification of P. multocida type B:2 in vital organs inoculated via different routes with river water contaminated with infected mice carcasses kept for 72 hours by using PCR.

Page

11

22

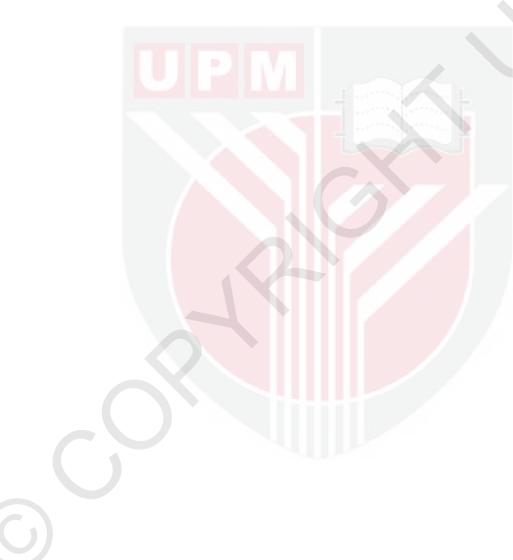
24

26

LIST OF FIGURES

Figure		Page
3.1	Flowchart of experimental design in mice model	10
3.2	picture of normal mice (left) and ruffled fur mice(right)	12
4.1	Responses of SAA in mice post inoculation with river water	
	contaminated with infected mice carcasses for 24, 48 and 72 hour	
	through the different routes of inoculation.	17
4.2	Responses of Hp in mice post inoculation with river water	
	contaminated with infected mice carcasses for 24, 48 and 72 hour	
	through the different routes of inoculation.	17
5.1	PCR identification of <i>P. multocida</i> type B: 2 in organs inoculated	
	through different routes with river water contaminated with infected	
	mice carcasses kept for 24 hours.	23
5.2	PCR identification of <i>P. multocida</i> type B: 2 in organs inoculated	
	through different routes with river water contaminated with infected	
	mice carcasses kept for 48 hours.	25
5.3	PCR identification of <i>P. multocida</i> type B: 2 in organs inoculated	
	through different routes with river water contaminated with infected	
	mice carcasses kept for 72 hours.	27
6.1	Section of the heart showing severe congestion, degeneration and	
	necrosis of cardiocytes, from mice inoculated intraperitoneally with	
	contaminated river water kept for 72 hours.	31
6.2	Section of the kidney showing severe congestion, tubular	
	degeneration and necrosis, from mice inoculated intraperitoneally	
	with contaminated river water kept for 72 hours.	31
6.3:	Section of the lung showing severe congestion, thickening of the	
	interalveolar septae with cellular infiltration, from mice inoculated	
	intraperitoneally with contaminated river water kept for 72 hours.	32
6.4	Section of the lung showing congestion and mild pulmonary oedema	
	from mice inoculated intraperitoneally with contaminated river water	
	kept for 72 hours.	32
6.5	Section of the spleen showing severe congestion and necrotic	
	follicles in the white pulp from mice inoculated intraperitoneally	
	with contaminated river water kept for 72 hours.	33
6.6	Section of the liver 72hrs showing congestion of the central vein,	
	degeneration and necrosis of hepatocytes from mice inoculated	
	intraperitoneally with contaminated river water kept for 72 hours.	33
6.7	Section of the liver showing congestion of the sinusoids and cellular	
	infiltration from mice inoculated intraperitoneally with contaminated	
	river water kept for 72 hours.	34
6.8	Section of the liver showing congestion in the sinusoids,	
	degeneration and necrosis of hepatocytes with leucocytic infiltration	
	from mice inoculated intraperitoneally with contaminated river water	24
6.0	kept for 72 hours.	34
6.9	Section of the brain showing capillary congestion, neuronal	
	degeneration and necrosis from mice inoculated intraperitoneally	25
	with contaminated river water kept for 72 hours.	35

- 6.10 Bar graph showing distribution of pathological lesions in organs of mice inoculated with water contaminated with infected mice carcass for 24 hours via aerosol, intraperitoneal and oral routes.
- 6.11 Bar graph showing distribution of pathological lesions in organs of mice inoculated with water contaminated with infected mice carcass for 48 hours via aerosol, intraperitoneal and oral routes.
- 6.12 Bar graph showing distribution of pathological lesions in organs of mice inoculated with water contaminated with infected mice carcass for 72 hours via aerosol, intraperitoneal and oral routes.



38

LIST OF ABBREVIATIONS

APP APR	Acute phase proteins Acute phase reaction
AGID	Agar gel immunodiffusion
BHI	Brain Heart Infusion
DNA	Deoxyribonucleic acid
ELISA	Enzyme Linked Immunosorbent Assay
FAO	Food and Agriculture Organization
Нр	Haptoglobin
HS	Haemorrhagic septicaemia
ICR	Institute of Cancer Research
IU	International unit
IHA	Indirect haemagglutination
MgCl ₂	Magnesium chloride
NC	necrotic cardiocytes
OIE	Office of International Epizootic
PBS	Phosphate Buffered Saline
PCR	Polymerase chain reaction
SAA	serum amyloid A
TSI	Triple Sugar Iron
VRI	Veterinary Research Institute
WHO	World Health Organization
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CHAPTER 1

INTRODUCTION

The production of ruminants in Malaysia is steadily changing from subsistence to intensive operations (Jesse et al., 2013a). The ruminants' population is predominantly made up of cattle, buffaloes, goats and sheep. The ruminant industry is the foremost in the production of food such as milk, meat and the by- products such as hide for the leather industry in Malaysia. The ruminant sector is vulnerable to the deadly and infectious disease outbreaks, which causes enormous losses to farmers and the country. The acute deadly disease is known as haemorrhagic septicaemia (HS) caused by a Gram negative bacterium called *Pasteurella multocida* type B: 2.

The pathogen is significant in animals for a protracted period and emerging as human pathogen (Jesse et al., 2013a) leading to a disease process termed Pasteurellosis. Notwithstanding, Pasteurellae have been indicated as the most dominant microflora of the upper respiratory tract in healthy animals (Abubakar et al., 2012; Jesse et al., 2013b). The organisms usually act as secondary invaders in animals with concomitant diseases or suffering from incapacitating stressful conditions (Abubakar and Zamri, 2011; Jesse et al., 2013c) and the infection frequently occurs via inhalation or ingestion of infected material (Shafarin et al., 2009).

In one study, research was conducted to investigate on the survival rate of *Pasteurella multocida* in water using different temperature where 18° C provides better survival rate then 2° C (Bredy and Botzler, 1989). Depending on the mode of storage and temperature where *Pasteurella multocida* was not isolated in river water and artificial sea water from the first to the 14^{th} day of inoculation (Thomson et al., 1992).

Animals experienced exogenous or endogenous challenge mount a strong response by activating the innate immune systems at the initial stage of acute infection before the onset of the acquired immune system (Eckersall, 2000). The acquired immune system ultimately leads to the development of specific cellular and humoral immune responses. During the preliminary challenge the continued existence of the host depends on the capability of the innate reactions to tussle the causes of disease (Eckersall, 2000).

The acute phase proteins (APPs) are proteins found in the blood and the circulating concentrations of APPs are correlated to the severity of the infection and hence the concentrations of APPs offers a ready means of assessing the presence and extent of the disease progression (Kent, 1992; Eckersall, 2000; Murata et al., 2004; Petersen et al., 2004; Ceron et al., 2005; Baumann and Gauldie, 1994; Tothova, et al., 2013).

However, there are substantial species dissimilarities in the serum concentrations of acute phase proteins (Eckersall and Bell, 2010). Six fold increases in Hp concentrations was observed in infected dairy cows and those with metabolic disease compared to animals with minor lesions (Hirvonen et al., 1999). Seven and forty fold increases in SAA and Hp were also observed in culled dairy cattle with acute lesions relative to healthy beef animals (Tourlomoussis et al., 2004). Isolation of *Pasteurella multocida* from calves with respiratory disease was associated with significant increased concentrations of acute phase proteins (Nikunen et al., 2007; Hodgson et al., 2005).

Jesse et al. (2013c) successfully isolated Pasteurella multocida type B: 2 in mice following oral inoculation using polymerase chain reaction. The infection of mice with contaminated river water with P. multocida type B: 2 and subsequent alterations of SAA and Hp in a mice model has not been elaborated in previous studies. There is still inadequate information about the pathogenicity and epidemiology of HS through the intraperitoneal, oral and aerosol routes with contaminated river water with P. multocida type B:2 mice carcasses kept for 24, 48 and 72 hours. This study is a leap in the knowledge of HS transmission in a mice model. This study also postulated that the outbreak of HS among buffaloes and cattle could be due to the consumption of river water contaminated with infected HS carcasses and the aerosol routes could perhaps be a readily available route for effective vaccine administration and heightened immunity in animals considering the progressive responses of APPs through this route. Therefore, the present study aims at evaluating the presence of P. multocida type B: 2 in various organs using Polymerase chain reaction (PCR), and host cell response in mice infected with Pasteurella multocida type B: 2 in contaminated river water with mice carcasses kept for 24, 48 and 72 hours.

Therefore, the objectives of present study are as follows:

1. To detect the presence of *P. multocida* type B: 2 in various organs using Polymerase chain reaction (PCR) in mice through different routes of inoculation with river water contaminated with the bacteria.

2. To determine acute phase protein response associated with *Pasteurella multocida* type B: 2 infections in mice through different routes with river water contaminated with *Pasteurella multocida* type B: 2.

3. To determine the severity of cellular changes in various organs associated with *Pasteurella multocida* type B: 2 infections in mice through different routes with contaminated river water with *Pasteurella multocida* type B: 2.

Therefore the hypotheses of study were as outlined below:

1. Infection of mice with contaminated river water via different routes of inoculation will lead to the establishment of HS in mice and will be associated with clinical responses.

2. Contaminated river water with *P. multocida* type B: 2 inoculations through different routes can be detected in the various organs using Polymerase chain reaction (PCR).

3. Inoculation of mice through different routes with contaminated river water containing *Pasteurella multocida* type B: 2 will lead to changes in the concentrations of acute phase proteins.

4. There will be severe cellular changes in the various organs associated with the infection of *Pasteurella multocida* type B: 2 in mice inoculated through different routes with contaminated river water.

Therefore, the present study will provide a better comprehension of the characteristics of *Pasteurella multocida* type B; 2 infection and pathogenesis in the host post inoculation with contaminated river water.

REFERENCES

- Abdullah, F.F.J., M.M. Khaleel, L. Adamu, A.Y. Osman and A.W. Haron et al., (2013). Polymerase chain reaction detection of *Pasteurella multocida* type B: 2 in mice infected with contaminated river water. American Journal of Animal and Veterinary Sciences., 8:146-151.
- Abdullah, F.F.J., A.Y. Osman, L. Adamu, Z. Zakaria and R. Abdullah et al., (2013b). Acute phase protein profiles in calves following infection with whole cell, lipopolysaccharide and outer membrane protein extracted from *Pasteurella multocida* type B: 2.Asian Journal of Animal and Veterinary Advances .,8: 655-662.DOI:10.3923/ajava.2013.655.662.
- Abubakar, M.S. and M. Zamri-Saad, (2011). Clinicopathological changes in buffalo calves following oral exposure to B: 2. Basic Applied Pathology., 4: 130-135. DOI: 10.1111/j.1755-9294.2011.01113.x
- Allan, E.M., Gibbs, H.A., Wiseman, A. and Selman, I.E. (1985).Sequential lesions of experimental bovine pneumonic pasteurellosis. Veterinary Record. 117: 438-442.
- Alsemgeest, S.P.M., Jonker, F.H., Taverne, M.A.M., Kalsbeek, H.C., Wensing, T. and Gruys, E. (1995).Serum amyloid A and haptoglobin plasma concentrations in newborn calves. Theriogenology. 43:381-387.
- Alsemgeest, S.P.M., Kalsbeek, H.C., Wensing, T., Koeman, J.P., van Ederen, A.M. and Gruys, E.(1994). Concentrations of serum amyloid A and haptoglobin as parameters of inflammatory diseases in cattle. Veterinary Quarterly.16:21-23.
- Alsemgeest, S.P.M., Taverne, M.A.M., Boosman, R., Weyden, G.C. and Gruys, E. (1993). The acute phase protein serum amyloid A in plasma of cows and fetuses around parturition. American Journal of Veterinary Research.54:164-167.
- Ashraf, A. H., Tariq, S., Nadeem, S. S., & Manzoor, I. (2011). Characterization of *Pasteurella multocida* strains isolated from cattle and buffaloes in Karachi, Pakistan. Afr. J. Micrabiol. Res, 5, 4673-4677.
- Aziz, D.M. and Taha, M.B. (1997). Effect of dystocia on serum haptoglobin in Awassi ewes. Theriogenology.48: 559-562.
- Baumann, H and Gauldie, J. (1994). The acute phase response. Immunology today, 15(2), 74-80.
- Boyce, J. D., Chung, J. Y., & Adler, B. (2000). *Pasteurella multocida* capsule: composition, function and genetics. Journal of biotechnology, 83(1), 153-160.
- Boyce, J. D., & Adler, B. (2006). How does *Pasteurella multocida* respond to the host environment?. Current opinion in microbiology, 9(1), 117-122.
- Bredy, J. P and Botzler, R. G. (1989). The effects of six environmental variables on *Pasteurella multocida* populations in water. Journal of Wildlife Diseases, 25(2), 232-239.
- Brickell, S. K., Thomas, L. M., Long, K. A., Panaccio, M., &Widders, P. R. (1998). Development of a PCR test based on a gene region associated with the pathogenicity of *Pasteurella multocida* serotype B: 2, the causal agent of Haemorrhagic Septicaemia in Asia. Veterinary microbiology, 59(4), 295-307.
- Carrongeon, M. (1902). Bovine pasteurellosis in Malay Peninsular. The Veterinary Journal.55: 321-327.
- Carter, G. R., and M. C. L. De Alwis."Haemorrhagicseptcaemia." Pasturella and pasteurellosis/edited by C. Adlam, JM Rutter (1989).
- Carter, G.R.(1955). A haemagglutination test for the identification of serological types. American Journal of Veterinary Research., 16: 481-484.

- Cerón, J. J., Eckersall, P. D and Martínez Subiela, S. 2005. Acute phase proteins in dogs and cats: current knowledge and future perspectives. Veterinary Clinical Pathology., 34(2), 85-99.
- Chandrasekaran, B. (1988). Generic tasks as building blocks for knowledge-based systems: The diagnosis and routine design example. Knowledge Eng. Review., 3(3), 183-210.
- Cole, D.J., Roussel, A.J. and Whitney, M.S.(1997). Interpreting a bovine CBC: Evaluating the leukon and acute-phase proteins. Veterinary Medicine.92: 470.
- Conner, J.G., Eckersall, P.D., Wiseman, A., Bain, R.K. and Douglas, T.A. (1989). Acute phase response in calves following infection with Pasteurella haemolytica, Ostertagiaostertagi and endotoxin administration. Research in Veterinary Science., 47:203-207.
- Cray, C., J. Zaias and N.H. Altman, (2009). Acute phase response in animals: A review. Comparative medicine., 59:517-526. PMID: 20034426.
- De Alwis, M. C. L. (1992). Haemorrhagic septicaemia—a general review. British Veterinary Journal, 148(2), 99-112.
- De Alwis, M. C. L. (1993). The epidemiology of haemorrhagic septicaemia in Sri Lanka.InAciar Proceedings (pp. 98-98).Australian centre for international agricultural research.
- De Alwis, M. C. L. (1995). Haemorrhagic septicaemia (*Pasteurella multocida* serotype B: 2 and E: 2 infection) in cattle and buffaloes. In Haemophilus, Actinobacillus, and Pasteurella (pp. 9-24).Springer US.
- Eckersall, P. D and Bell, R.(2010). Acute phase proteins: Biomarkers of infection and inflammation in veterinary medicine. The Veterinary Journal, 185(1), 23-27.
- Eckersall, P. D. (2000). Recent advances and future prospects for the use of acute phase proteins as markers of disease in animals. Revue de Médecine Vétérinaire, 151(7), 577-584.
- Eckersall, P.D. and R. Bell, (2010). Acute phase proteins: Biomarkers of infection and inflammation in veterinary medicine. Veterinary Journal., 185: 23-27. DOI:10.1016/j.tvjl.2010.04.009.
- Faez Firdaus Jesse Abdullah, Abdinasir Yusuf Osman, Lawan Adamu, Mohd Syamil Mohd Yusof, Abdul Rahman Omar, Abdul Aziz Saharee, Abd Wahid Haron, Rasedee Abdullah and Mohd Zamri-Saad, (2013). Polymerase Chain Reaction Detection of *Pasteurella multocida* Type B:2 in Mice Following Oral Inoculation. Asian Journal of Animal and Veterinary Advances, 8: 493-501.
- Faez Firdaus Jesse Abdullah, Abdinasir Yusuf Osman, Lawan Adamu, Mohd Syamil Mohd Yusof, Abdul Rahman Omar, Abdul Aziz Saharee, Abd Wahid Haron, Rasedee Abdullah and Mohd Zamri-Saad, (2013). Polymerase Chain Reaction Detection of *Pasteurella multocida* Type B:2 in Mice Following Oral Inoculation. Asian Journal of Animal and Veterinary Advances, 8: 493-501.

FAO-WHO-OIE (1994): Animal Health Yearbook, 1994.

- Food and Agricultural Organisation (FAO), (1991).Haemorrhagic septicaemia in Peninsular Malaysia. In Proceeding of the 4th International workshop on haemorrhagic septicaemia. Sri Lanka: FAO/APHCA Publication No. 1991/13.
- Gibbs, H.A., Allan, E.M., Wiseman, A. and Selman, I.E. (1984).Experimental production of bovine pneumonic pasteurellosis. Research in Veterinary Science., 37: 154-166.
- Godson, D.L., Campos, M., Attah-Poku, S.K., Redmond, M.J., Cordeiro, D.M., Sethi, M.S., Harland, R.J. and Babiuk, L.A.(1996). Serum haptoglobin as an indicator

of the acute phase response in bovine respiratory disease. Veterinary Immunology and Immunopathology.51: 277-292.

- Gomez-Laguna, J., F.J. Salguero, F.J. Pallares, M.F.D.Marco and I. Barranco et al. 2010.Acute phase response in porcine reproductive and respiratory syndrome virus infection. Comparative Immunology Microbiology Infectious Disese., 33: e51-e58. PMID:20004019.
- Gruys, E., Toussaint, M. J. M., Niewold, T. A., & Koopmans, S. J. (2005). Review: Acute phase reaction and acute phase proteins. Journal of Zhejiang University.Science.B, 6(11), 1045.
- Harper, M., Boyce, J. D., & Adler, B. (2006). *Pasteurella multocida* pathogenesis: 125 years after Pasteur. FEMS microbiology letters, 265(1), 1-10.
- Heddleston, K.L., Rhodes, K.R. and Reber, P.A. (1972). Fowl cholera. Gel diffusion precipitin test for serotyping *Pasteurella multocida* from anian species. Avian Disease.16: 925-936.
- Hiramune, T., & de Alwis, M. C. (1982).Haemorrhagic septicaemia carrier status of cattle and buffaloes in Sri Lanka. Tropical animal health and production, 14(2), 91-92.
- Hirvonen, J. and Pyorala, S. (1998). Acute-phase response in dairy cows with surgicallytreated abdominal disorders. Veterinary Journal.155:53-61.
- Hirvonen, J., Huszenicza, G., Kulcsar, M. and Pyörälä, S. (1999). Acute-phase response in dairy cows with acute postpartum metritis. Theriogenology, 51(6), 1071-1083.
- Hodgson, J. M., Burke, V and Puddey, I. B. (2005). Acute effects of tea on fasting and postprandial vascular function and blood pressure in humans. Journal of hypertension, 23(1), 47-54.
- Horadagoda, A., Eckersall, P.D., Hodgson, J.C., Gibbs, H.A. and Moon, G.M. (1994). Immediate responses in serum TNFα and acute phase protein concentrations to infection with Pasteurella haemolytica A1 in calves. Research in Veterinary Science. 57:129-132.
- Jesse, F.F.A., Y.O. Abdinasir, L. Adamu, M.Y. Syamil and A.R. Omar et al., (2013c). Polymerase Chain reaction detection of *Pasteurella multocida* type B: 2 in Mice Following Oral Inoculation. Asian Journal of Animal and Veterinary Advances. 8: 493-501. DOI: 10.3923/ajava. 2013.493.501.
- Jesse, F.F.A., L. Adamu, Y.O. Abdinasir, M.Z. Saad and Z. Zakaria et al., (2013b). Acute phase protein profiles and clinico-pathological changes in mice associated with the infection of *Pasteurella multocida* type B and the bacterial lipopolysaccharide and outer membrane protein immunogens. Journal of Animal and Veterinary Advances 12: 186-193.
- Jesse, F.F.A., L. Adamu, Y.O. Abdinasir, Z. Zakaria and R. Abdullah et al., (2013a). Clinico-pathological responses of calves associated with infection of Pasteurella multocida type В and the bacterial lipopolysaccharide and outer membrane protein immunogens. Int. Journal of Animal and Veterinary Advances 5: 190-198.
- Joseph, P.G. (1979). Haemorrhagic septicaemia in Peninsular Malaysia, Kajian Veterinar, 11: 65-79.
- Katch, N., Miyamoto, T., Nakagawa, H. and Watanabe, A. (1999) Detection of annexinI and IV and haptoglobin in bronchoalveolar lavage fluid from calves experimentally inoculated with Pasteurella haemolytica: American Journal of Veterinary Research.60: 1390-1395.
- Kent, J. (1992). Acute phase proteins: their use in veterinary diagnosis. British Veterinary Journal, 148(4), 279-282.

- Khaleel, M. M., Abdullah, F. F. J., Adamu, L., Osman, A. Y., Haron, A. W., Saad, M. Z., & Omar, A. R. (2013). Acute phase protein responses in mice infected with river water contaminated by *Pasteurella multocida* type B: 2. American Journal of Animal and Veterinary Sciences, 8(3), 159.
- Kharb, S., & Charan, S. (2013). Mouse model of haemorrhagic septicaemia: dissemination and multiplication of *Pasteurella multocida* B: 2 in vital organs after intranasal and subcutaneous challenge in mice. Veterinary research communications, 37(1), 59-63.
- Kumar, A.A., Shivachandra, S.B., Biswas, A., Singh, V.P., Singh, V.P., Srivastava, S.K., (2004). Prevalent serotypes of *Pasteurella multocida* isolated from different animal and avian species in India. Veterinary Research Communications 28 (8), 657–667.
- Khin, M. N., Zamri-Saad, M., & Noordin, M. M. (2010). Pathological Changes in the Lungs of Calves Following Intratracheal Exposure to *Pasteurella multocida* B: 2. Pertanika Journal of Tropical Agricultural Science, 33(1).
- Lane, E.P., Kock, N.D., Hill, F.W. and Mohan, K. (1992). An outbreak of haemorrhagic septicaemia (septicaemia pasteurellosis) in cattle in Zimbabwe. Tropical Animal Health and Production. 24(2): 97-102.
- Murata, H., Shimada, N and Yoshioka, M. (2004). Current research on acute phase proteins in veterinary diagnosis: an overview. The Veterinary Journal, 168(1), 28-40.
- Mustafa, A.A., Ghalib, H.W. and Shigidi, M.T. (1978). Carrier rate of *Pasteurella multocida* in a cattle herd associated with an outbreak of haemorrhagic septicaemia in the Sudan. British Veterinary Journal.134: 375-378.
- Namioka, S. (1978). *Pasteurella multocida*. Biochemical characteristic and serotype. In: Methods in Microbiology, 10. Academic Press, London, UK, 272-292.
- Nikunen, S., Härtel, H., Orro, T., Neuvonen, E., Tanskanen, R., Kivelä, S. L and Soveri, T. (2007). Association of bovine respiratory disease with clinical status and acute phase proteins in calves. Comparative immunology, microbiology and infectious diseases, 30(3), 143-151.
- Ozkanlar, Y., M.S. Aktas, O. Kaynar, S. Ozkanlar and E.Kireccl et al., (2012). Bovine respiratory disease in naturally infected calves: Clinical signs, blood gases and cytokine response. Revue Med. Vet., 163: 123-130.
- Perumalpillai, C. and Thambiayah, V.S. 1957. Outbreaks of haemorrhagic septicaemia in an epizootic form in Ceylon. Ceylon Veterinary Journal, 5, 24-28.
- Petersen, H. H., Nielsen, J. P and Heegaard, P. M. H. (2004). Application of acute phase protein measurements in veterinary clinical chemistry. Veterinary research, 35(2), 163-187.
- Pomorska-Mol, M., I. Markowska-Daniel, K. Kwit, K. Stepniewska and Z. Pejsak, (2013). C-reactive protein, haptoglobin, serum amyloid a and pig major acute phase protein response in pigs imultaneously infected with H1N1 swine influenza virus and *Pasteurella multocida*. BMC Veterinary Research., 9: 14-14.DOI: 10.1186/1746-6148-9-14.
- Priadi, A., & Natalia, L. (2000). Pathogenesis of haemorrhagic septicaemia (HS) in cattle and buffalo: clinical signs, pathological changes, re-isolation and detection of *Pasteurella multocida* using culture medium and polymerase chain reaction (PCR). JurnalIlmu Ternakdan Veteriner, 5(1), 65-71.
- Radostits, O. M. (2000). Veterinary medicine: a textbook of the diseases of cattle, sheep, pigs, goats and horses/cO. M. Radostits... [et al.

- Rafidah O, Zamri-Saad M, Nasip E, Shahiruddin S, Saharee AA (2010). Analysis of haemorrhagic septicaemia outbreaks in cattle and buffalo in Malaysia. Online Journal of Veterinary Research. 14(2): 325-333.
- Ramdani, H. J. S., Johnson, R. B., & Spencer, T. L. (1991). Haemorrhagic septicaemia: correlation of vaccinal antibody responses in mice with protection against *Pasteurella multocida* strain M1404. Veterinary microbiology, 27(3), 309-326.
- Saharee, A. A., Salin, N. B., Rasedee, A., &Jainudeen, M. R. (1993).Haemorrhagic septicaemia carriers among cattle and buffalo in Malaysia. In ACIAR Proceedings (pp. 89-89). Australian centre for international agricultural research.
- Salonen, M., Hirvonen, J., Pyorala, S., Sankari, S. and Sandholm, M.(1996). Quantitative determination of bovine serum haptoglobin in experimentally induced Escherichia-coli mastitis. Research in Veterinary Science.60 :88-91.
- Schiefer, B., Ward, G.E. and Moffatt, R.E.(1978).Correlation of microbiological and histological findings in bovine pneumonia. Veterinary Pathology.15: 313-321.
- Shafarin, M. S., Zamri-Saad, M., Khairani, B. S and Saharee, A. A. (2009). Pathological Changes in the Respiratory Tract of Goats Infected by *Pasteurella multocida* B: 2. Journal of comparative pathology, 140(2), 194-197.
- Shah, N. H., Biewenga, J., Shah, N. H., &Graaf, F. K. (1996). Vacuolating cytotoxic activity of *Pasteurella multocida* causing haemorrhagic septicaemia in buffalo and cattle. FEMS microbiology letters, 143(1), 97-101.
- Singh, N. A. G. I. A. (1948). Nasal carriers in bovine pasteurellosis. Indian Journal of Veterinary Science and Animal Husbandry, 18, 261-278.
- Skinner, J. G. (2001). International standardization of acute phase proteins. Veterinary Clinical Pathology, 30(1), 2-7.
- Thomson, C. M., Chanter, N and Wathes, C. M. (1992). Survival of toxigenic *Pasteurella multocida* in aerosols and aqueous liquids. Applied and environmental microbiology, 58(3), 932-936.
- Tóthová, C., Nagy, O and Kováč, G. (2013). The Use of Acute Phase Proteins as Biomarkers of Diseases in Cattle and Swine.
- Tourlomoussis, P., Eckersall, P. D., Waterson, M. M and Buncic, S. (2004). A comparison of acute phase protein measurements and meat inspection findings in cattle. Foodbourne Pathogens & Disease, 1(4), 281-290.
- Wallace, W.R. (1929). Acute enzootic haemorrhagic septicaemia of the buffalo.Veterinary Record. 9:709-717.
- Wittum, T.E., Young, C.R., Stanker, L.H., Griffin, D.D., Perrino, L.J. and Littledike, E.T.(1996). Haptoglobin response to clinical respiratory tract disease in feedlot cattle. American Journal of Veterinary Research.57: 646-649.
- World Organization for Animal Health [OIE]. (2008). Manual of diagnostic tests and vaccines for terrestrial animals [online].Paris: OIE; 2008. Haemorrhagic septicemia. Available at: http://www.oie.int/eng/normes/mmanual/2008/pdf/2.04.12_HS.pdf.
- Yeo, B. K., & Mokhtar, I. (1993). Haemorrhagic Septicaemia in Sabah, Malaysia. Pasteurellosis in Production Animals. Patten BE, TL Spencer, RB Johnson, D. Hoffmann and L. Lehane (Eds.), ACIAR (Australian Centre for International Agricultural Research) Proceedings, 43, 112-115.