

UNIVERSITI PUTRA MALAYSIA

PRODUCTION, EXPRESSION AND CHARACTERIZATION OF A HEAT-STABLE ORGANIC SOLVENT TOLERANT LIPASE FROM BACILLUS SP, STRAIN 42

MOHAMRD ABDALLAH ELTAWEEL.

FBSB 2005 11



PRODUCTION, EXPRESSION AND CHARACTERIZATION OF A HEAT-STABLE ORGANIC SOLVENT TOLERANT LIPASE FROM BACILLUS SP. STRAIN 42

By

MOHAMED ABDALLAH ELTAWEEL

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfillment of the Requirements for the Degree of Doctor of Philosophy

October 2005



DEDICATION

I dedicate this humble effort, the fruit of my thoughts and study,
to the great and helpful wife Hawa Safar,
dear sons Abdallah, Abdalrahman, Abdalraouf
and sweet daughter Halima
who have inspired me to higher ideals of life.



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

PRODUCTION, EXPRESSION AND CHARACTERIZATION OF A HEAT-STABLE ORGANIC SOLVENT TOLERANT LIPASE FROM BACILLUS SP. STRAIN 42

By

MOHAMED ABDALLAH ELTAWEEL

October 2005

Chairman: Associate Professor Raja Noor Zaliha Raja Abd Rahman, PhD

Faculty: Biotechnology and Biomolecular Sciences

Ninety two bacterial strains were isolated from oil palm effluent from Bangi, Selangor; Kluang, Johor; Alor Gajah hot spring (up to 54 °C) Melaka and Slim River hot spring (up to 91°C) Perak. An enrichment culture technique was used to isolate bacteria utilizing olive oil as a substrate. Cultures were incubated at 60°C to select for the thermophilic bacteria. Eight isolates showed lipolytic activity on tributyrin and triolein agar plates. In order to screen for highest lipase producer, six production media were used. Isolate 42 was observed to produce the highest level (0.059 U/ml) after 72h. Its crude lipase retained its full activity when preincubated at 70°C for 30 min. It also showed high stability in several organic solvents (25% v/v). Furthermore, its activity was enhanced in benzene, hexane and hexadecane while, completely inhibited by butanol. Isolate 42 was



identified as *Bacillus* sp. Strain 42 using 16S rDNA. The nucleotide sequence deposited at GenBank under accession number AY 763118.

Further optimization studies were done in order to determine the best lipase production condition. Inoculum size of 3% proved to be the best for lipase production, with an optimum temperature of 50°C when, grown under shaking condition of 150 rpm. A combination of tryptone and yeast extract was the best nitrogen source. Lipase production was stimulated by olive oil.

The lipase gene was amplified by polymerase chain reaction using consensus primers based on multiple aligned sequences of thermophilic genes from other thermophilic *Bacillus* species. Nucleotide sequence comparison shared high homology with the thermostable genes in *Geobacillus* sp., *Bacillus* stearothermophilus and *Bacillus* thermoleovorans. Nucleotide sequence deposited at GenBank under accession number AY 787835. The amplified gene was successfully cloned using a pQE-30 UA expression vector and induced by IPTG at the optimum concentration of 0.75 mM.

The recombinant lipase was facilitated by the fusion of 6-histidine and this allowed a one-step purification of the lipase enzyme using Ni-NTA affinity chromatography. The histidine-tagged lipase was purified 6-fold with a yield of 21.7%. Purified lipase migrated as a single band with a molecular mass of ~43 KDa on SDS-PAGE.



The purified lipase showed high activity at 70°C with its optimum at pH 8.0. The enzyme was stable over a broad range of pH from 6.0 to 10.0. It also showed high stability with half-lives of 315 min at 60°C , 120 min at 65°C , and 45 min at 70°C . Preincubation enzyme activity was stimulated with Na⁺, K⁺ and Ca²⁺. While, Zn^{2+} , Mn²⁺ and Fe ²⁺ at high concentration (10 mM) were greatly inhibitory. Protease inhibitors Bestatin and pepstatin stimulated the lipase activity while, phenylmethylsulfonyl fluoride (PMSF) completely inhibited the lipase activity. Tween 80 (0.1%) enhanced the lipase activity while higher concentration (1%) dramatically decreased the lipase activity. The activity of preincubated enzyme in heptanol (log P 2.4) and octanol (log P 2.9) was slightly enhanced while, remains very stable with other organic solvents tested. Solvents such as ethylbenzene (log P 3.1) and dodecane (log P 6.6) reduced the lipase activity up to 35% and 38%, respectively. The highest specificity was observed towards tricaprylin (C₈), followed by tricaprin (C₁₀). Its hydrolyzed all the natural oils tested, with highest hydrolysis rate on olive oil.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia Sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

PENGHASILAN, PENGEKSPRESIAN DAN PENCIRIAN LIPASE TAHAN PELARUT ORGANIK DAN TERMOSTABIL DARI BACILLUS SP. STRAIN 42

Oleh

MOHAMED ABDALLAH ELTAWEEL

Oktober 2005

Pengerusi: Profesor Madya Raja Noor Zaliha Raja Abd Rahman, PhD

Fakulti: Bioteknologi Dan Seins Biomolekul

Sebanyak 92 strain bakteria telah dipencilkan daripada sisa kumbahan sawit yang di perolehi dari Bangi, Selangor; dan Kluang Johor, serta kolam air panas di Alor Gajah, Melaka (suhu 54°C) dan di Slim River, Perak (91°C). Minyak zaitun sebagai substrat telah digunakan untuk memencilkan bakteria melalui kaedah pengkayaan kultur. Pengeraman pada suhu 60°C digunakan untuk menggalakkan pertumbuhan bakteria yang rintang suhu atau termofilik. Sebanyak lapan isolat telah menunjukkan aktiviti lipolitik tertinggi di atas plat agar tributirin and triolein. Di dalam media cecair, penghasilan tertinggi didapati pada strain 42. Bagi meningkatkan penghasilan lipase, sebanyak enam jenis media digunakan. Media M3 didapati menghasillkan lipase tertinggi, iaitu 0.059 U/mL pada 72 jam pengeraman, dengan kadar goncangan 150 rpm, pada suhu 60°C.



Kajian dilanjutkan bagi menentukan penghasilan terbaik enzim lipase. Inokulum bersaiz 3% terbukti menghasilkan enzim lipase tertinggi dengan suhu optimum pada 50°C. Penghasilan lipase paling tinggi adalah pada kadar goncangan 150 rpm per min. Penghasilan lipase adalah dirangsang oleh penambahan minyak zaitun sebagai substrat. Lipase mentah menunjukkan kestabilan yang tinggi sehingga mencapai suhu 70°C.

Ekstrak enzim mentah diuji terhadap beberapa pelarut organik berkepekatan 25% v/v, selama 30 min untuk menentukan kestabilannya. Peningkatan aktiviti berlaku di dalam pelarut benzena, heksana dan heksadekana, tetapi ia juga stabil di dalam pelarut toluena, xylina, dekanol, isooktana dan tetradekana. Aktiviti enzim menurun sebanyak 34.5% di dalam pelarut propanol dan 63.6% di dalam perlarut propilasetat berbanding kawalan dan direncatkan sepenuhnya oleh butanol. Gen lipase dari *Bacillus* sp. strain 42 telah digandakan melalui tindakbalas rantaian polimerase (PCR) menggunakan primer konsensus berdasarkan padanan jujukan berganda gen termofilik daripada spesis *Bacillus*. Perbandingan jujukan nukleotida menunjukkan gen lipase strain 42 mempunyai homologi yang tinggi dengan gen termostabil dari *Geobacillus* sp, *Bacillus stearothermophilus* dan *Bacillus thermoleovorans*. Gen yang digandakan ini telah berjaya diklon ke dalam vector pQE-30 UA dan telah diekspreskan dengan kehadiran IPTG pada kepekatan optimum 0.75 mM.

Penulenan enzim lipase rekombinan dipermudahkan dengan kehadiran 6histidina pada vektor, ini membolehkan penulenan satu langkah dengan



menggunakan Ni-NTA kromatografi afiniti. Lipase pembawa histidina telah ditulenkan sebanyak 5.65 kali dengan hasilan 21.7%. Lipase rekombinan tulen bergerak sebagai satu jalur dengan jisim molekular ~43 KDa pada SDS-PAGE. Lipase tulen menunjukkan aktiviti tertinggi pada suhu 70°C dengan pH optimum 8.0. Enzim adalah stabil pada julat pH dari 6.0 ke 10.0. Ia juga menunjukkan kestabilan tertinggi dengah tempoh separuh hayat 315 min pada 60°C, 120 min pada 65°C dan 45 min pada 70°C.

Lipase tulen menghidrolisiskan kesemua minyak semulajadi yang diuji dengan kadar hidrolisis tertinggi terhadak minyak zaitun. Spesifisiti substrat tertinggi adalah terhadap trikaprilin (C₈) diikuti oleh trikaprin (C₁₀). Aktiviti lipase meningkat dengan penambahan ion logam seperti Ca⁺ dan Na⁺. Tween 80 pada kepekatan 0.1% meningkatkan aktiviti enzim tetapi aktiviti menurun pada 1%. Bestatin dan pepstatin juga meningkatkan sedikit aktiviti enzim tetapi EDTA tidak meningkatkan sebarang kesan. Sebaliknya aktiviti enzim direncatkan oleh fenilmetilsulfonilfluorida iaitu perencat protease serine.



ACKNOWLEDGEMENTS

All praise is to the Almighty Allah, the Merciful and the Beneficent. Had it not been due to his will and favour, the completion of this study would not have possible.

I avail myself of this opportunity to record my sincerest thanks and appreciation to Associate Professor Dr. Raja Noor Zaliha Raja Abd Rahman, chairman of my supervisory committee, for her dedicated efforts, support, invaluable advice, intellectual guidance and encouragement in the conduct of my research and in the preparation of this thesis.

Grateful thanks are also due to my supervisory committee members, Professor Dr. Abu Bakar Salleh and Professor Dr. Mahiran Basri, for their constructive comment, advice and help throughout my studies and in the preparation of this final manuscript.

I am exceedingly grateful to the Libyan Higher Learning Popular Committee for their financial support. Thanks are also extended to all staff member in the department of Biotechnology and Biomolecular Sciences, lab mates and friends who have helped me in one way or another.

Finally, I also take this opportunity to express my deep gratitude to my affectionate Mother, brothers, sisters, helpful wife Hawa Safar, dear sons



Abdallah, Abdalrahman, Abdalraouf and sweet daughter Halima. I thank them for all their love, patience, support and encouragement throughout my study in UPM and whole life. Without their understanding and sacrifices this project would have been nigh impossible.





TABLE OF CONTENTS

					Page
AB AB AP DE LIS	PROVECLAR STOF STOF	CT K VLEDGEM	i e		II III VI IX XII XIII XVI XVII XXI
CHA	APTER				
1	INTRO	DUCTIO	N		1
	Object	tive of this	study		7
2	LITER	ATURE R	EVIEW		8
	2.1 2.2 2.3	2.2.1 2.2.2 2.2.3	of Lipases Plant Lipases Mammalian Lipases Microbial Lipases able Lipase		8 10 10 11 11
	2.4	Organic S	Solvent Tolerant Lipas		16
	2.5 2.6 2.7	Selection Detection		polytic Microorganisms	19 21 24
	2.8 2.9	Effect of I 2.9.1	n of Microbial Lipase Nutritional Factors on Carbon Sources		27 28 28
	2.10	2.10.1	Nitrogen Sources Substrates Minerals Physical Factors on L Temperature	ipase Production	30 32 33 34 34
		2.10.2 2.10.3	pH Cultivation Period		35 35



		2.10.4	Shaking Rate	36
	2.11	Purification	on of Lipase	37
	2.12	Properties	s of Purified Microbial Lipases	39
	2.13	Substrate	Specificity	41
		2.13.1	Positional Specificity	41
		2.13.2	Fatty Acid Specificity	42
		2.13.3	Partial Glycerides Specificity	44
	2.14	Polymera	se Chain Reaction	44
	2.15	Primer De	esign	45
	2.16	Optimizat	ion of PCR	48
	2.17	Selection	of Vector	49
	2.18	Selection	of Host	51
		_	nd Expression	53
		Cloning b		54
	2.21	Direct Sel	lection Strategy of Lipase Gene	57
			on of Lipase Gene	58
	2.23	Application	on of Lipases	61
3	MATE	ERIALS AN	ID METHODS	64
	- 1			
	3.1	Materials		64
	3.2	Methods		71
		3.2.1	Preparation of Media and Solutions	71
		3.2.2	Bacterial Sources	80
		3.2.3	Enrichment Culture Technique	80
		3.2.4	Isolation of Bacteria	80
		3.2.5	Screening of Lipase Producing Microorganisms	81
		3.2.6	Slant Agar Stock Culture	81
		3.2.7	Glycerol Stock Culture	82
		3.2.8	Preparation of Inoculum	82
		3.2.9	Assay of Lipase Activity	82
		3.2.10	Effect of Different Liquid Media on Lipase Production	02
		2 2 44	Effect of Temporature on Crude Engume Stability	83 84
		3.2.11 3.2.12	Effect of Temperature on Crude Enzyme Stability	04
		3.2.12	Effect of Organic Solvents on Crude Enzyme Stability	84
		3.2.13	Bacterial Identification	86
		3.2.14	Growth Curve and Lipase Production of <i>Bacillus</i> sp.	OC.
		0.Z. I T	Isolate 42	88
		3.2.15	Growth Optimization Study for Maximum Enzyme	
		0.2.10	Production	88
		3.2.16	Physical Factors Affecting the Growth and Lipase	
		3.2.,0	Production by <i>Bacillus</i> sp. Isolate 42	89
		3.2.17	Nutritional Factors Affecting the Growth and Lipase	-
		J.—	Production by <i>Bacillus</i> sp. Isolate 42.	91
		3.2.18	Cloning and Sequencing of Thermostable Organic	



			Solvent Tolerant Lipase Gene	95
		3.2.19	Amplification of Thermostable Organic Solvent Tolerant	
			Lipase Gene by PCR	97
		3.2.20	PCR Amplification of the Lipase Gene from Genomic	
			DNA	97
		3.2.21	Purification of the Amplified PCR Product	99
		3.2.22	Preparation of Competent E. coli	100
		3.2.23	Cloning PCR Product using pQE-30 UA Vector	100
		3.2.24	Transformation	101
		3.2.25	Plasmid Isolation	101
		3.2.26	Analysis of Positive Colonies	102
		3.2.27	Stock Culture	103
		3.2.28	Expression of Recombinant Thermostable Organic	100
		3.2.20	Solvent Tolerant Lipase Gene	103
		3.2.29	Preparation of Culture Supernatant and Cell Extract	104
		3.2.29	SDS-PAGE Analysis of Bacteria Protein	105
			Optimum IPTG Concentration for Expression	106
		3.2.31		107
		3.2.32	Assay of Recombinant Lipase Activity	101
		3.2.33	Purification of Thermostable Organic Solvent Tolerant	107
		0.004	Recombinant Lipase	107
		3.2.34	Protein Determination	109
		3.2.35	Characterization of Purified Thermostable Organic	400
			Solvent Tolerant Recombinant Lipase	109
	DEOL	II TO AND	PICOLICCION	117
4	RESU	JL15 AND	DISCUSSION	117
	4.4	11-4:	and Care aning of Thermonhilia Linguitic Pactoria	117
	4.1		and Screening of Thermophilic Lipolytic Bacteria	118
	4.2		f Different Liquid Media on Lipase Production	125
	4.3		Temperature on Crude Enzyme Stability	126
	4.4		Organic Solvents on the Crude Lipase Activity	
	4.5		Identification	130
		4.5.1	16S rDNA Identification and Phylogenetic Tree	400
			Analysis	130
	4.6	Growth C	Curve and Lipase Production by Bacillus sp. Isolate 42	400
				138
	4.7	Effects o	of Physical Factors on Growth and Lipase Production	
				141
		4.7.1	Temperature	141
		4.7.2	Agitation	143
		4.7.3	pH	146
		4.7.4	İnoculum Size	148
		4.7.5	Medium Volume	150
	4.8		Nutritional Factors on growth and Lipase Production	
				151
		4.8.1	Carbon Sources	151
		4.8.2	Inorganic Nitrogen Sources	156
		4.8.3	Organic Nitrogen Sources	156
				1



		4.8.4	Formulation of Nitrogen Source for Optimal Production	
				160
		4.8.5	Metal lons	164
		4.8.6	Substrates	166
	4.9	4.8.7	Tweens Sequencing and Expression of the Organic Solvent	169
	4.9	.	Lipase gene	171
		4.9.1	Genomic DNA Extraction	171
	4.10	-	tion of Thermostable Organic Solvent Tolerant Lipase	
			Polymerase Chain Reaction (PCR)	173
	4.11	Gene Ana		180
		4.11.1	Nucleotide Sequence and Deduced Amino Acid	
			Sequence of <i>Bacillus</i> sp. Isolate 42	180
	4.12		of Organic Solvent Tolerant Lipase Gene using pQE-30	400
	4.40	UA vecto		186
	4.13 4.14	•	of Positive Colonies	188
	4.14	Gene	on of Thermophillic Organic Solvent Tolerant Lipase	195
	4.16		GE Analysis of Bacterial Proteins	197
	4.15		IPTG Concentration	197
	4.17		on of Th <mark>ermostable</mark> Organic Solvent Tolerant Lipase	
				200
			Affinity Chromatography	200
	4.18		rization of Purified Organic Solvent Tolerant Lipase	203
		4.18.1	Determination of Molecular Weight	203
		4.18.2	Effect of pH on Lipase Activity	205
		4.18.3 4.18.4	Effect of pH on Lipase Stability	208 210
		4.18.5	Effect of Temperature on Lipase Activity Effect of Temperature on Lipase Stability	212
		4.18.6	Effect of Metal Ions on Lipase Activity	214
		4.18.7	Effect of Surfactants	217
		4.18.8	Effect of Inhibitors	219
		4.18.9	Substrate Specificity	222
		4.18.10		
			Activity	224
			Effect of Organic Solvents on Lipase Activity	226
		4.18.12	Positional Specificity	230
5	CONC	CLUSION		233
	REFE	RENCES		236
	APPE	NDICES		258
		ATA OF T	HE ALITHOD	000
	RIOD	AIAUFI	HE AUTHOR	266



LIST OF TABLES

Table	•	Page
1	The log P Value of Common Solvents	23
2	Properties of Purified Lipases from some Bacterial Sources	40
3	Preparation of Free Fatty Acid Solutions	75
4	Bovine Serum Albumin Solutions	78
5	Thermophillic Lipase Producer from <i>Bacillus</i> sp Precursors Nuculeotide Sequences Extracted from the NCBI Database.	98
6	Oligonucleotide Used as Primers for Specific Amplification of Genes Encoding for Organic Tolerant Lipase Gene Fragments	98
7	Screening the Activity of Lipolytic Thermophilic Bacteria Isolated from Different Local Regions in Malaysia	119
8	Effect of Different Liquid Media on the Isolates Lipase Production	120
9	Effect of Different Organic Solvents on Strain 42 Crude Lipase Activity	129
10	Summary of the Purification Recombinant 6 x His-tagged Lipase Produced with <i>E. coli</i> M15	202
11	Effect of Organic Solvents on Lipase Stability	227



LIST OF FIGURES

Figur	re Pag	je
1	Hydrolytic and Synthetic Action of Lipase	9
2	Schematic Diagram of the PCR Process	46
3	Standard Curve for the Determination of Free Fatty Acid	76
4	Standard Curve for Protein Determination	79
5	Flow Chart for Lipase Assay Producer	85
6	Bacillus sp. Isolate 42 Streaked on Tributryin Plate with Control	122
7	Bacillus sp. Isolate 42 Streaked on Triolein Plate with Control	123
8	Bacillus sp. Isolate 42 Streaked on Rhodamine Plate with Control	124
9	Relative Stability of the Crude Lipase from Isolate 42	125
10	PCR Product of 16S rDNA Gene (1500 bp) from Isolate 42 Amplified using Universal Primers	131
11	16S rDNA Sequence of Bacillus sp. Strain 42	132
12	Comparison of 16S rDNA from <i>Bacillus</i> sp. BGSC W9A6 (AY 608903), <i>Bacillus</i> sp. BGSC W9A22 (AY 608987) and <i>Bacillus</i> sp Strain 42(AY 763118).	136
13	Rooted Phylogenetic Tree Showing the Relationships of Bacillus sp. Strain 42 to other Bacillus spp.	137
14	Time Profile of Lipase Production by Bacillus sp. Strain 42	140
15	Effect of Temperature on Growth and Lipase	142
16	Effect of Agitation on Bacterial population and Lipase production	145
17	Effect of pH on Bacterial Growth and Lipase Production	147
18	Effect of Inoculum Size on Bacterial Growth and Lipase	



	Production	149
19	Effect of Medium Volume on Bacterial Growth and Lipase Production	152
20	Effects of Carbon Sources on the Growth and Lipase Production by <i>Bacillus</i> sp. Strain 42	153
21	Effect of Inorganic Nitrogen Sources on Growth and Lipase Production by <i>Bacillus</i> sp. Isolate 42.	157
22	Effect of Organic Nitrogen Sources on Growth and Lipase Production by <i>Bacillus</i> sp. Strain 42.	159
23	Effects of Tryptone with some Other Nitrogen Sources on Bacterial Growth and Lipase Production by <i>Bacillus</i> sp. Strain	162
24	Effects of Different Concentrations of Tryptone and Yeast extract on Growth and Lipase Production by <i>Bacillus</i> sp. Isolate 42.	163
25	Effect of Metal lons (Ca ²⁺ , Mg ²⁺ and Fe ³⁺ , Individually and in Combination, on the Growth and Lipase Production by <i>Bacillus</i> sp. Strain 42.	165
26	Effect of Heavy Metal lons on the Growth and Lipase Production by <i>Bacillus</i> sp. Isolate 42.	167
27	Effects of Substrates on the Growth and Lipase Production by Bacillus sp. Strain 42.	168
28	Effect of Tween on the Growth and Lipase Production by Bacillus sp. Isolate 42	170
29	Genomic DNA Extraction from Strain 42	173
30	PCR Product of 308 bp Amplified by Using Hlip F1 and Hlip R1 Primers	175
31	PCR Product of 812 bp Amplified by Using Hlip F2 and Hlip R2 Primers	176
32	PCR Product of 771 bp Amplified by Using lip F12 and Hlip R 2 Primers	177
33	PCR Product of 790 bp Amplified by Using Hlip F2 and lip R8 Primers	178



34	PCR Product of 1251 bp Amplified by Using lip F12 and lip R12 Primers	179
35	Nucleotide and Deduced Amino Acid Sequences of Thermostable	182
36	The Putative Signal Peptide Cleavage Site	183
37	Comparison of the Amino Acids Sequence of Lipase Gene from <i>Bacillus</i> sp. Strain 42 with other Thermophilic Lipases Sequence Obtained from GenBank.	185
38	pQE-30 UA Vector	187
39	Clea <mark>ring Zone of <i>E. coli</i> M</mark> 15 Harboring Thermostable Organic Solvent Tolerant Lipase Gene on Tributyrin Plate	189
40	Experimental Procedure of Cloning the Thermostable Lipase Gene of Bacillus sp. Strain 42	190
41	Cloning and Sequence Strategy of the Cloned Fragments by pQE-30 UA vector	191
42	Intensive Zone of <i>E. coli</i> M 15 Harboring Thermostable Organic Solvent Tolerant Lipase Gene on Triolein Plate	192
43	Orange Fluorescence Halo of <i>E. coli</i> M 15 Harboring Thermostable Organic Solvent Tolerant Lipase Gene on Rhodamine Plate	193
44	Double Digestion of Recombinant pQE-30 UA Vector	194
45	Expression of Thermostable Organic Solvent Tolerant Lipase by pQE-30 UA Vector	196
46	SDS-PAGE of Bacterial Proteins Analysis	198
47	Effect of different IPTG Concentration at 3h Induction in <i>E. coli</i> Harboring <i>Bacillus</i> sp. Strain 42 Lipase Gene	199
48	Immobilization Metal Affinity Chromatography of Recombinant 6 x His-tagged Lipase	201
49	SDS-PAGE of the Purified sp. 42 Lipase	204
50	Estimation of the Molecular Weight of the Purified Lipase by Gel	



	Filtration Chromatography.	206
51	Effect of pH on Purified Lipase Activity	207
52	Effect of pH on Purified 42 Lipase Stability	209
53	Effect of Temperature on Purified Lipase Activity	211
54	Effect of Temperature on Purified Lipase Stability	213
55	Effect of Metal Ions on Purified Lipase Activity	215
56	Effect of Surfactants on Purified Lipase Activity	218
57	Effect of Inhibitors on Purified Lipase Activity	220
58	Chain Length Specificity of Purified Lipase	223
59	Specificity of the Purified Lipase Towards Natural Oils.	225
60	Thin-layer Chromatography of the Hydrolysis Products Obtained with Ttriolein as Substrate	231



LIST OF ABBREVIATIONS

POME Palm Oil Mill Effluent

cm Centimeter dH₂O Distilled Water

DNA Deoxyribonucleic Acid

dNTPs Deoxynucleotide Triphosphates EDTA Ethylenediaminetetraacetic Acid

mM Millimole g/L Gram per Liter

Gram g Nanogram ng Milligram Mg Microgram μg pmol Picomole Liter М Molar mL Milliliter Microliter ul % Percentage

rpm Rotation per Minute

Micromole

x g Gravity
UV Ultraviolet

umole

IPTG Isopropyl-β-D- Thiogalactopyranoside

X-gal 5-bromo-4chloro-3-indolyl-β-D galactopyranoside

h Hour Min Minute

PCR Polymerase Chain Reaction
ORF Open Reading Frame
GTE Glucose Tris-HCI-EDTA

PCI Phenol Chloroform Isoamylalcohol

SDS Sodium Dodecyl Sulphate

SDS-PAGE Sodium Dodecyl Sulphate Polyacrylamide Gel Electrophoresis

APS Amonium Persulphate
TCA Trichloroacetic Acid
TSB Tryptone Soy Broth

LB Luria-Bertani

BSA Bovine Serum Albumin

U/mL Unit per Milliliter
cfu Colony Form Units
v/v Volume per Volume
w/v Weight per Volume



psi Pound Persquare Inch

bp Base Pair

Ni-NTA Nickle-nitrilotriacetate acid

Kbp Kilo Base Pair

Da Dalton
KDa Kilo Dalton
mA Milliampere
sp Specie
spp Species
NCBI National Cei

NCBI National Center for Biotechnology Information



