



**UNIVERSITI PUTRA MALAYSIA**

***AGRO-MORPHOLOGICAL, GLUCOMANNAN CONTENT AND  
MOLECULAR CHARACTERIZATION OF MALAYSIAN  
Amorphophallus spp. BLUME GERMPLOSM COLLECTION***

***SURISA PHORNVILLAY***

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By

**SURISA PHORNVILLAY**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in  
Fulfilment of the Requirements for the Degree of  
Master of Science**

**September 2015**

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**I dedicate this dissertation works in memory of beloved my mother and grandmother, who I deeply miss.**

**A special dedication to my father, sisters and other family members for always be here, by my side through many ups and downs till the completion of my master study**

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the Degree of Master of Science

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**September 2015**

**Chairman : Siti Hajar Ahmad, PhD**  
**Faculty : Agriculture**

*Amorphophallus* has attracted much attention as it contains glucomannan and also possess other medicinal properties. Prior to the collection of propagating materials and cultivation, identification and diversity information of the *Amorphophallus* species are essential as different species perform differently under cultivation. However, limited research has been conducted on morphological and genetic variations of *Amorphophallus* spp. in Peninsular Malaysia. Therefore, the present study was conducted with objectives, namely collection and establishment of the *Amorphophallus* spp. in Peninsular Malaysia, morphological and genetic variations, and glucomannan (GM) content of the established *Amorphophallus* spp. accessions. The current observation from the collection of *Amorphophallus* spp. from six locations found that *Amorphophallus* spp. were abundantly spotted in disturbed areas of secondary forest. They were found thriving well in damp, moist and shady areas, mostly near a riverbank. The accessions were established, and their life cycle was observed. *Amorphophallus* had three distinct growth phases, namely, vegetative, generative and dormancy. The plant had a vegetative growing phase with the leaf development for about 5 months and then underwent a dormancy period for about 2 to 3 months. The plant resumed its growth in the next season or followed by generative life cycle, whereby the inflorescence was produced. Sixty accessions of *Amorphophallus* spp., with 10 accessions representing six locations, were used to assess morphological vegetative characters and molecular variations. The morphological variation data were also supplemented with the assessment of 20 *Amorphophallus* inflorescence accessions. There were variations in morphological characteristics among accessions of *Amorphophallus* spp. based on corm size, corm shape, cormel number per corm, petiole nature and distinct inflorescence characters, spathe colour, appendix colour and appendix shape and pattern. Genetic variations of the plants were assessed using nine screened microsatellite primers. Principle coordinates and cluster analysis results for morphological and molecular characters clearly separated Kota Bahru, Kelantan (KKB) accessions from the other 50 accessions, thus grouping the accessions into two major groups. The KKB population accounted for 22.22% genetic variations while the remaining five populations varied 77.78 to 100% genetically. Nonetheless, high inbreeding coefficients (0.21 to 0.88) were detected within each of the population. For the determination of glucomannan (GM) content, the 3,5-dinitrosalicylic acid

colorimetric assay was employed. The Hulu Langat, Selangor accession SHL24 was found to contain the highest GM content at 8.29%. The lowest GM content was from Perak, Taiping; PT37 at only 2.55%. The results also showed that there was a significant interaction effect between the populations and the corm size of GM content. Based on the results, PUK population of 500 g had the highest GM content. In conclusions, both the morphology and molecular analysis results suggest that KKB accessions are identified as *A. paeoniifolius* while other accession are *A. prainii* particularly based on distinct inflorescence characteristics. This study provides valuable insight and a better understanding of the habitat, growth, phenotypic and genotypic characteristics and GM content of the available *Amorphophallus* spp. accessions in Peninsular Malaysia. These findings could be used as the basic information for breeding, improvement and conservation program of this plant in Malaysia.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

**AGRO-MORFOLOGI, KANDUNGAN GLUKOMANAN DAN PENCIRIAN MOLEKUL BAGI KOLEKSI GERMPLASMA *Amorphophallus* spp. BLUME MALAYSIA**

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*Amorphophallus* telah menarik banyak perhatian kerana kandungan glukomanan serta memiliki ciri perubatan. Pengenalpastian dan pengumpulan maklumat tentang kepelbagaian spesies *Amorphophallus* adalah penting kerana spesies yang berlainan menunjukkan prestasi yang berbeza semasa penanaman dan diikuti dengan pengumpulan bahan-bahan tanaman dan penanaman. Namun begitu, penyelidikan yang dijalankan terhadap variasi morfologi dan genetik tumbuhan *Amorphophallus* spp. di Semenanjung Malaysia masih terhad. Oleh itu, kajian ini dijalankan dengan menggunakan dua objektif iaitu pengumpulan dan pengukuhan bagi spesies *Amorphophallus* di Semenanjung Malaysia, variasi morfologi dan genetic, serta kandungan glukomanan (GM) daripada aksesori *Amorphophallus* spp. yang dikumpulkan. Hasil pemerhatian daripada pengumpulan *Amorphophallus* spp. daripada enam lokasi mendapati bahawa *Amorphophallus* spp. boleh ditemui dengan banyak di kawasan hutan sekunder yang telah diterokai. Ia didapati tumbuh dengan subur di kawasan lembap dan teduh yang kebanyakannya adalah berhampiran dengan tebing sungai. Aksesori *Amorphophallus* spp. telah diperolehi dan kitaran hidup tumbuhan ini diperhatikan. *Amorphophallus* mempunyai tiga fasa pertumbuhan yang berbeza iaitu, vegetatif, generatif dan dorman. Tumbuhan tersebut mempunyai fasa vegetatif dengan pertumbuhan daun selama kira-kira lima bulan dan kemudian menjalani satu tempoh dorman selama lebih kurang 2 hingga 3 bulan. Pertumbuhan diteruskan semula pada musim berikutnya atau diikuti dengan kitaran hidup generatif yang menghasilkan bunga akan dihasilkan. Sebanyak 60 aksesori *Amorphophallus* spp. dengan 10 aksesori mewakili enam lokasi telah digunakan untuk penilaian morfologi aksara vegetatif dan variasi molekul. Penambahan terhadap data bagi variasi morfologi dilakukan dengan penilaian terhadap 20 aksesori bunga *Amorphophallus*. Terdapat kepelbagaian pada ciri-ciri morfologi di antara aksesori *Amorphophallus* spp. berdasarkan saiz dan bentuk umbisi, bilangan anak umbisi per umbisi, sifat petiol dan ciri-ciri jambak bunga yang jelas dengan warna *spathe*, dan umbaian (appendix) serta bentuk dan corak umbaian (appendix). Variasi genetik tumbuh-tumbuhan dinilai menggunakan sembilan primer mikrosatelit. Prinsip koordinasi dan analisis berkelompok bagi pencirian morfologi dan molekul telah memisahkan aksesori Kota Bahru, Kelantan (KKB) daripada 50 aksesori yang lain dengan jelas dan membentuk dua kumpulan yang besar. Populasi KKB

menyumbang kepada 22.22% variasi genetik dan lima populasi dengan 77.78%-100% adalah variasi selebihnya. Namun begitu, koefisien pembiakbakaan yang tinggi (0.21 hingga 0.88) dikesan bagi setiap populasi. Penentuan kandungan glukomanan (GM) daripada spesies *Amorphophallus* menggunakan asai kolorimetrik asid 3,5-dinitrosalisilik. Aksesori Hulu Langat, Selangor, SHL24 didapati mengandungi kandungan GM yang paling tinggi pada 8.29%. Kandungan GM yang paling rendah adalah daripada PT37 dengan hanya 2.55%. Keputusan juga menunjukkan bahawa terdapat kesan interaksi di antara aksesori dan saiz umbi dengan kandungan GM. Kesimpulannya, berdasarkan kedua-dua analisis morfologi dan molekul mencadangkan bahawa aksesori KKB dikenal pasti sebagai *A. paeoniifolius* manakala aksesori lain adalah *A. prainii*, terutamanya berdasarkan ciri bunga yang berbeza. Kajian ini memberikan pandangan yang berharga dan pemahaman yang mendalam tentang habitat, pertumbuhan, ciri-ciri fenotip dan genotip serta kandungan GM di dalam aksesori *Amorphophallus* spp. di Semenanjung Malaysia. Penemuan ini boleh digunakan sebagai asas untuk pembiak-bakaan dan program pemuliharaan tumbuhan ini di Malaysia.



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This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

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## LIST OF ABBREVIATIONS

GM	Glucomannan
NS	North-south
EW	East-west
DAP	Days after planting
M	Molar
WHO	World Health Organization
NAA	$\alpha$ -naphthaleneacetic acid
MS	Murashige and Skoog medium
DNA	Deoxyribonucleic acids
PCR	Polymerase chain reaction
RAPD	Random amplified polymorphic DNA
AFLP	Amplified fragment length polymorphisms
ISSR	Inter simple sequence repeat
SSR	Simple sequence repeat
EST	Expressed sequence tag
PCA	Principal component analysis
PCoA	Principal coordinate analysis
PC	Principal component
UPGMA	Unweighted pair group method based on arithmetic averages
UPM	Universiti Putra Malaysia
CRD	Completely Randomized Design
ANOVA	Analysis of variance
LSD	Least significant different
PUK	Ulu Kenas, Perak

PT	Taiping, Perak
KKB	Kota Bahru, Kelantan
KKP	Kubur Panjang, Kedah
PBJ	Bukit Jambul, Penang
SHL	Hulu Langat, Selangor
MgCl <sub>2</sub>	Magnesium chloride
dNTP	2'-deoxynucleoside 5'-triphosphate
TBE	Tris-borate-EDTA
I	Shanon's information index
H <sub>o</sub>	Observed heterozygosity
H <sub>e</sub>	Expected heterozygosity
PIC	Polymorphic information content
CF	Crude flour
NTSYS-pc	Numerical taxonomy multivariate analysis system
PAST	Paleontological statistics
EDTA	Ethylene diamine tetraacetic acid
g	Gram
μl	Microliter
μM	Micromolar
min	Minute
s	Second
°C	Degree celsius
w/v	Weight per volume
ng	Nanogram
kg	Kilogram



mM	Milimolar
ml	Milliliter
h	Hour
nm	Nanometer
syn	Synonym
bp	Base pair
cm	Centimeters
mg	Milligram
$T_a$	Annealing temperatures
$F_{is}$	Fixation index
$F_{ST}$	Coefficient of gene differentiation
$N_m$	Estimate of gene flow
$n_a$	Average number of alleles
$n_e$	Number of effective alleles
6-BAP	6-benzylaminopurine
2,4-D	2, 4-dicholophenoxy acetic acid
3,5-DNS	3,5-dinitrosalicylic

## CHAPTER 1

### GENERAL INTRODUCTION

*Amorphophallus* spp. is a perennial plant belonging to the Araceae family, consisting of approximately 200 species. They have been found distributed across the tropical and subtropical regions ranging from West Africa to Polynesia (Hettterscheid and Ittenbach, 1996; Sedayu et al., 2010). The *Amorphophallus* spp. has an underground storage organ known as the corm. The corm produced enormous solitary leaves consisting of trunk-like petiole, often with mottled and dissected umbrella shaped leaf lamina (Hettterscheid and Ittenbach, 1996; Hejnowicz and Barthlott, 2005). In Peninsular Malaysia, *A. paeoniifolius* and *A. prainii* are reported to be in abundance, followed by *A. muelleri* and *A. elegans* (Mansor et al., 2012; Shahbudin, 2012).

Plants of the *Amorphophallus* have been long used as folk medicine, food source, animal fodder, and ornamental and religious ceremony (Long, 1998; Bown, 2000; Follet and Douglas, 2002). *A. prainii*, *A. aphyllus*, *A. commutatus*, *A. paeoniifolius* and *A. sylvaticus*, for instance, are used to treat snake bite, as well as for arrow poison and an analgesic (Quattrocchi, 2012). *A. paeoniifolius* is grown commercially in India. The corm, young shoot and flower are eaten, either as boiled or baked vegetables, and used in Ayurvedic medicine. The ash of the corm is prescribed to treat piles, haemorrhoids, gout, asthma, bronchitis and stomach indigestion while the petiole juice is used to cure diarrhoea (Khare, 2004). The corm extract possesses antitumour, antioxidant and cytotoxic properties and has synergistic depressant effect when used with diazepam (Madhurima et al., 2012). In China, *A. konjac* is used traditionally for detoxification, as well as an antitumor and to treat asthma; cough and skin disorder (Bown, 2000; Niwa et al., 2000).

However, recent interest is towards the flour content extracted from the corm tissues of some of *Amorphophallus* spp., such as *A. konjac*, *A. albus* and *A. muelleri*. The corm contains high glucomannan (GM) content which is a water-soluble polysaccharide, fermentable dietary fibre or hemicelluloses (Keithley and Swanson, 2005). Traditionally, the flour has been used to prepare noodles, tofu and snacks (Long, 1998; Chua et al., 2010). As the flour has no calorific content, therefore, the purified form is potential to be used in health food products such as supplement, diet cookies, noodles and rice. Many studies have reported that GM is an effective treatment of obesity, diabetes, hypertension, cardiovascular disease and high cholesterol problems (Liu et al., 1998; Kraemer et al., 2007). Consumption of GM based food products increased high-density lipoprotein cholesterol and total cholesterol ratio (Keithley and Swanson, 2005; Kraemer et al., 2007).

Development of GM based-food products is significant, especially for developed and developing countries, including Malaysia, whereby the above diseases are listed as the primary cause of major health problem and death. According to the World Health Organization (WHO), approximately 3.4 million adults die each year due to diabetes,

ischaemic heart disease, cancer and obesity (WHO, 2014). The diseases are caused by the consumption of high calorific content of foods. Thus, consuming GM food-based products, with little calories, could help to overcome the problem.

Purified konjac flour (>~90.0% GM) has the potential to be developed as a pharmaceutical excipient to control drug delivery system due to its unique gelling and rheological properties (Zhang et al., 2014). As a natural biopolymer, GM is also used to make the capsule for medicine, whereby it can replace gelatin as edible coating for postharvest produce and to restructure foods (Parry, 2010). As a gelatin replacer, GM has gained much attention in the Muslim world because gelatin is derived from the animal source, considered as non-halal. GM seems to be the most promising source of a plant-based medicinal capsule for the halal industry. Therefore, due to its benefits and market demand, Japan, China and Indonesia regard these plants as a high-value crop and are commercially planting *A. konjac*, *A. albus* and *A. muelleri*. China is the world largest producer and exporter of GM flour with 60% of total global production followed by Japan at 28% (Liu, 2004).

Despite the economic importance and benefits of *Amorphophallus* plants, there is still lack of detail information available for the Malaysian *Amorphophallus* spp. in Peninsular Malaysia. To date, only a few studies have been conducted on *Amorphophallus* spp. cultivation and glucomannan content, much less on genetic diversity of this plant in Malaysia. Therefore, in order to promote breeding program, such fundamental knowledge of morphological and genetic variation is crucial as it underlies the crop improvement and selection. Moreover, *Amorphophallus* plant is easy to grow under plant shade, and the climate in Malaysia provides suitable conditions for the plant to grow. It is apparent that *Amorphophallus* spp. has the potential to be cultivated as a new crop.

Therefore, the objectives of this study were to (i) collect and establish the accessions of *Amorphophallus* spp. from selected locations in Peninsular Malaysia, (ii) determine phenotypic and genotypic characteristics, as well as the GM content of the accessions.

## REFERENCES

- Abraham, Z., Latha, M., Asha, K. I., Varghese, C., Lakhminarayan, S. and Pareek, S. K. (2006). *Minimal descriptors of agri-horticultural crops Part V: spices, tubers and plantations crops*. Thrissur: National Bureau of Plant Genetic Resources, Regional station.
- Afifi, A., May, S. and Clark, V. A. (2011). *Practical Multivariate Analysis, Fifth Edition*: Taylor & Francis.
- Agarwal, M., Shrivastava, N. and Padh, H. (2008). Advances in molecular marker techniques and their application in plant sciences. *Plant Cell Reports*, 27, 617-631.
- Ajibade, S. R., Weeden, N. F. and Chite., S. M. (2000). Inter-simple sequence repeat analysis of genetic relationships in the genus *Vigna*. *Euphytica*, 111, 47-55.
- Al-Saghir, M. G., Malkawi, H. I. and El-Oqlah, A. (2009). Morphological Diversity in *Hordeum spontaneum* C. Koch of Northern Jordan (Ajloun Area). *Middle-East Journal of Scientific Research*, 4(1), 24-27.
- An, N. T., Thien, D. T., Dong, N. T., Dung, P. L. and Dub, N. V. (2010). Characterization of glucomannan from some *Amorphophallus* species in Vietnam. *Carbohydrate Polymers*, 80, 308-311.
- Anil, S. R., Siril, E. A. and Beevy, S. S. (2011). Morphological variability in 17 wild elephant foot yam (*Amorphophallus paeoniifolius*) collections from southwest India. *Genetics Resources and Crop Evolution*, 58, 1263-1274.
- Anil, S. R., Sirila, E. A. and Beevy, S. S. (2014). Diversity analysis in *Amorphophallus* using isozyme markers. *International Journal of Vegetable Science*, 20, 305-321.
- Anonymous. (2012). A big problem, Health, *The Star Online*. Retrieved from <http://www.thestar.com.my/Lifestyle/Health/2012/03/25/A-Big-problem/>
- Armstrong, W. P. (2002). Botany 115 Terminology. Flower Terminology Part 2 Retrieved 20 August 2012, from <http://waynesword.palomar.edu/termfl2.htm>
- Banerjee, B. (1992). Botanical classification of tea. In K. C. Willson and M. N. Clifford (Eds.), *Tea: Cultivation to consumption* (pp. 53-86). London, UK: Chapman & Hall.
- Bown, D. (2000). *Aroids, Plants of the Arum Family* (second ed.). Oregon, U.S.A: Timber Press.
- Boys, J., Cherry, M. and Dayanandan, S. (2005). Microsatellite analysis reveals genetically distinct populations of red pine (*Pinus resinosa*, Pinaceae). *American Journal of Botany*, 92(5), 833-841.

- Brown, D. M., Zeef, L. A. H., Ellis, J. and Turner, S. R. (2005). Identification of novel genes in *Arabidopsis* involved in secondary cell wall formation using expression profiling and reverse genetics. *Plant Cell*, 17, 2281-2295.
- Burkil, I. H. (1996). *A dictionary of the economic products of the Malay Peninsula*. Kuala Lumpur, Malaysia: Governments of Malaysia and Singapore by the Ministry of Agriculture and Co-operatives.
- Chae, S. S. and Warde, W. D. (2006). Effect of using principal coordinate and principal components on retrieval of clusters. *Computational Statistics and Data Analysis*, 50, 1407-1417.
- Chen, H. L., Fan, Y. H., Chen, M. E. and Chan, Y. (2005). Unhydrolyzed and hydrolyzed konjac glucomannan modulated cecal and fecal microflora in Balb/c mice. *Nutrition*, 22, 36-42.
- Chen, H. L., Cheng, H. C., Liu, Y. J., Liu, S. Y. and Wu, W. T. (2006). Konjac acts as a natural laxative by decreasing stool bulk and improving colonic ecology in healthy adults. *Nutrition*, 22, 1112-1119.
- Chen, H. L., Cheng, H. C., Wu, W. T., Liu, Y. J. and Liu, S. Y. (2008). Supplementation of konjac glucomannan into a low-fibre Chinese diet promoted bowel movement and improved colonic ecology in constipated adults: a placebo-controlled, diet-controlled trial. *Journal American College Nutrition*, 27, 102-108.
- Chua, M., Baldwin, T. C., Hocking, J. T. and Chan, K. (2010). Traditional uses and potential health benefits of *Amorphophallus konjac* K. Koch ex N.E.Br. *Journal of Ethnopharmacology*, 128, 268-278.
- Chua, M., Chan, K., Hocking, T. J., Williams, P. A., Perry, C. J. and Baldwin, T. C. (2012). Methodologies for the extraction and analysis of konjac glucomannan from corms of *Amorphophallus konjac* K. Koch. *Carbohydrate Polymers*, 87, 2202-2210.
- Cuesta, G., Suarez, N., Bessio, M. I., Ferreira, F. and Massalsi, H. (2003). Quantitative determination of pneumococcal capsular polysaccharide serotype 14 using a modification of phenol-sulphuric acid method. *Journal of Microbiological Methods*(52), 69-73.
- Dave, V., Sheath, M., McCarthy, S. P., Ratto, J. A. and Kaplan, D. L. (1998). Crystalline rheological and thermal properties of konjac glucomannan. *Polymer*, 39, 1139-1148.
- Dey, Y. N. and Gosh, A. K. (2010). Evaluation of anthelmintic activity of the methanolic extract of *Amorphophallus paeoniifolius* tuber. *International Journal Pharmaceutical Science and Research*, 1(11), 117-121.
- Dhua, R. S., Ghosh, S. R., Biswas, J., Mitra, S. K. and Sen, H. (1988). Effect on some chemicals on sprouting, growth and corm yield of *Amorphophallus campanulatus*. *Journal Root Crops*, 14, 47-49.

- Douglas, J. A., Follett, J. M. and Waller, J. E. (2005). Research on konjac (*Amorphophallus konjac*) production in New Zealand. *Proceedings of the First International Symposium on Root and Tuber Crops*, p. 173-180.
- Du, Q., Wang, B., Wei, Z., Zhang, D. and Li, B. (2012). Genetic diversity and population structure of Chinese White Poplar (*Populus tomentosa*) revealed by SSR markers. *Journal of Heredity*, 103(6), 853-862.
- Eckert, C., Samis, K. and Dart, S. (2006). Reproductive assurance and the evolution of uniparental reproduction in flowering plants. In L. Harder and S. Barrett (Eds.), *Ecology and evolution of flowers* (pp. 183-200). Oxford, UK: Oxford University Press.
- Ehlers, B. K. and Olesen, J. M. (2004). Flower production in relation to individual plant age and leaf production among different patches of *Corydalis intermedia*. *Plant Ecology*, 174, 71-78.
- Esquinas-Alcázar, J. T. (2005). Protecting crop genetic diversity for food security: political, ethical and technical challenges. *Nature Reviews Genetics*, 6, 946-953.
- FDA. (2001). 4th October Import Alert. IA3315. FIARS.
- Fernandez, E. and Hafizuddin, A. (2012). Ministry battles obesity problem, *New Straits Times*. Retrieved from <http://www2.nst.com.my/nation/general/ministry-battles-obesity-problem-1.142433>
- FOA. (1997). The state of the world's plant genetic resources for food and agriculture. Retrieved from <http://www.fao.org/WAICENT/FAOINFO/AGRICULT/AGP/AGPS/Pgrfa/pdf/swrfull.pdf>
- Follet, J. M. and Douglas, J. A. (2002). Konjac production in Japan and potential for New Zealand. *Combined Proceedings of the International Plant Propagators' Society*, 52, 186-190.
- Fufa, H., Baenziger, P. S., Beecher, B. S., Dweikat, I. and Graybosch, R. A. (2005). Comparison of phenotypic and molecular marker-based classifications of hard red winter wheat cultivars. *Euphytica*, 145, 133-146.
- Ghebru, B., Schmidt, R. J. and Bennetzen, J. L. (2002). Genetic diversity of *Eritrean sorghum* landraces assessed with simple sequence repeat (SSR) markers. *Theoretical and Applied Genetics*, 105, 229-236.
- Gong, X. M., Yan, R. W., Xu, H. Y., Li, C., Guang, S. Y. and Fang, M. (2002). Study of the extraction methods and properties of konjac glucomannan. *Fine Chemicals*, 19, 486-488.
- Gower, J. C. (1966). Some distance properties of latent root and vector methods used in multivariate analysis. *Biometrika*, 53, 325-338.

- Gower, J. C. (1971). A general coefficient of similarity and some of its properties. *Biometrics*, 27(4), 857-871.
- Gupta, R. K. (2006). *Textbook of Systemic Botany*. New Delhi: CBS Publications.
- Hahn, S. K. (1995). Yams. In J. Smartt and N. W. Simmonds (Eds.), *Evolution of crop plants*. Harlow, UK: Longman.
- Hammer, O., Harper, D. A. T. and Ryan, P. D. (2001). PAST: Paleontological Statistics Software Package for Education and Data Analysis. *Palaeontologia Electronica*, 4(1), 1-9.
- Hanelt, P. (2001). *Mansfeld's encyclopedia of agricultural and horticultural crops*. Heidelberg, New York: Springer-Verlag, Berlin.
- Harijati, N. and Mastuti, R. (2014). Estimation of diverse porang (*Amorphophallus muelleri* Blume) age in forest are based on branching pattern of leaf petiolule. *Research Journal of Life Science*, 1, 20-26.
- Hejnowicz, Z. and Barthlott, W. (2005). Structural and mechanical peculiarities of the petioles of gaint leaves of *Amorphophallus* (Araceae). *American Journal of Botany*, 92(3), 391-403.
- Hemmat, M., Weeden, N. F., Manganaris, A. G. and Lawson, D. M. (1994). Molecular marker linkage map for apple. *Journal of Heredity*, 85, 4-11.
- Henderson, A. (2006). Traditional morphometrics in plant systematics and its role in palm systematics. *Botanical Journal of the Linnean Society*, 151, 103-111.
- Hetterscheid, W. L. A. and Ittenbach, S. (1996). Everything you always wanted to know about *Amorphophallus* but were afraid to stick your nose into. *Aroideana*, 19, 7-129.
- Hong, Y., Chen, X., Liang, X., Liu, H., Zhou, G., Li, S., Wen, S., Holbrook, C.C. and Guo, B. (2010). A SSR-based composite genetic linkage map for the cultivated peanut (*Arachis hypogaea* L.) genome. *BMC Plant Biology*, 10, 1-13.
- Hu, J., Gao, X., Liu, J., Xie, C. and Li, J. (2008). Plant regeneration from petiole callus of *Amorphophallus albus* and analysis of soma clonal variation of regenerated plant by RAPD and ISSR makers. *Botanical Studies*, 49, 189-197.
- Hu, J. B., Liu, J., Yan, H. B. and Xie, C. H. (2005). Histological observations of morphogenesis in petiole derived callus of *Amorphophallus riveri* Durieu in vitro. *Plant Cell Reports*, 24, 642-648.
- Ipor, I. B., Tawan, C. S. and Basrol, M. (2006). Growth pattern, biomass allocation and response of *Cryptocoryne ferruginea* Engler (Araceae) to shading and water depth. *Jurnal Biosains*, 17(2), 55-78.

- Ipor, I. B., Tawan, C. S., Meekiong, K. and Simon, A. (2012). *Amorphophallus* diversity and conservation in Borneo and Malaysia. In G. G. Maiti and S. K. Mukherjee (Eds.), *Multidisciplinary approaches in angiosperm systematics* (Vol. 2, pp. 334-338). Kalyani, India: University of Kalyani.
- Ishurd, O., Kermagi, A., Elghazoun, M. and Kennedy, J. F. (2006). Structure of a glucomannan from *Lupinus varius* seed. *Carbohydrate polymer*, 65(4), 410-413.
- Jata, S. K., Sahoo, B. and Nedunchezian, M. (2009). Intercropping Elephant Foot Yam in Orchard Crops. *Orissa Review*, 82-84.
- Jentzsch, V. I. M., Bagshaw, A., Buschiazzo, E., Merkel, A. and Gemmell, J. E. (2008). *Evolution of microsatellite DNA*. Chichester, United Kingdom: John Wiley & Sons Ltd.
- Jones, C. J., Edwards, K. J., Castaglione, S., Winfield, M. O., Sala, F., Wiel, C. et al. (1997). Reproducibility testing of RAPD, AFLP, and SSR markers in plants by a network of European laboratories. *Molecular Breeding*, 3, 381-390.
- Joshi, S. P., Gupta, V. S., Aggarwal, R. K., Ranjekar, P. K. and Brar, D. S. (2000). Genetic diversity and phylogenetic relationship as revealed by intersimple sequence repeat (ISSR) polymorphism in the genus *Oryza*. *Theoretical and Applied Genetics*, 100, 1311-1320.
- Keithley, J. and Swanson, B. (2005). Glucomannan and obesity: a critical review. *Alternative Therapies in Health and Medicine*, 11, 30-34.
- Kenkel, N. C. (2006). On selecting an appropriate multivariate analysis. *Canadian Journal of Plant Science*, 86, 663-676.
- Khan, M. N. and Awasthi, L. P. (2006). Management of mosaic disease of *Amorphophallus campanulatus* through leaf extract of *Clerodendrum aculeatum*. *Indian Journal of Virology*, 17(1), 44-46.
- Khare, C. P. (2004). *Indian Herbal Remedies*. New York: Springer-Verlag Berlin Heidelberg.
- Khare, C. P. (2004). *Indian Herbal Remedies: Rational Western Therapy, Ayurvedic and Other Traditional Usage, Botany*. Germany: Springer-Verlag Berlin Heidelberg.
- Kite, G. C., Hettterscheid, W. L. A., Lewis, M. J., Boyce, P. C., Ollerton, J., Cocklin, E., Diaz, A. and Simmonds, M.J. (1998). Inflorescence odours and pollinators of *Arum* and *Amorphophallus* (Araceae). In S. J. Owens and P. J. Rudall (Eds.), *Reproductive Biology* (pp. 295-315). UK: Royal Botanic Gardens Kew.
- Kovach, W. L. (1999). *MVSP - a Multivariate Statistical package for WINDOWS, ver. 3.1*. Pentreath: Kovach Computing Services.



- Kraemer, W. J., Vingren, J. L., Silvestre, R., Spiering, B. A., Hatfield, D. L., Ho, J. Y., Fragala, M.S., Maresh, C.M. and Volek, J.S. (2007). Effect of adding exercise to a diet containing glucomannan. *Metabolism*, 56(8), 1149-1158.
- Kubik, C., Honig, J., Meyer, W. A. and Bonos, S. A. (2009). Genetic diversity of creeping bentgrass cultivars using ssr markers. *International Turfgrass Society Research Journal*, 11, 533-545.
- Kumar, P., Gupta, V. K., Misra, A. K., Modi, D. R. and Pandey, B. K. (2009). Potential of molecular markers in plant biotechnology. *Plant Omics Journal*, 2(4), 141-162.
- Kumara, G., Meenaa, B. L., Kara, R., Tiwaria, S. K., Gangopadhyaya, K. K., Bishta, I. S. and Mahajana, R. K. (2008). Morphological diversity in brinjal (*Solanum melongena* L.) germplasm accessions. *Plant Genetic Resources: Characterization and Utilization*, 6(3), 232-236.
- Kurniawan, A., Wibawa, I. P. A. H. and Adjie, B. (2011). Species diversity of *Amorphophallus* (Araceae) in Bali and Lombok with attention to genetic study in *A. paeoniifolius* (Dennst.) Nicolson. *Biodiversitas*, 12(1), 7-11.
- Lam, E., Kato, N. and Lawton, M. (2001). Programmed cell death, mitochondria and the plant hypersensitive response. *Nature*, 411, 848-853.
- Lebot, V. (2009). *Tropical root and tuber crops: cassava, sweet potato, yams and aroids*. UK: CABI.
- Legendre, P. and Legendre, L. F. J. (1998). *Numerical Ecology* (2 ed.). Amsterdam: Elsevier.
- Li, B., Xia, J., Wang, Y. and Xie, B. J. (2005). Grain-size effect on the structure and antiobesity activity of konjac flour. *Journal of Agricultural and Food Chemistry*, 53, 7404-7407.
- Lin, K. H., Lai, Y. C., Li, H. C., Lo, S. F., Chen, L. F. O. and Lo, H. F. (2009). Genetic variation and its relationship to root weight in the sweet potato as revealed by RAPD analysis. *Scientia Horticulturae*, 120, 2-7.
- Lindsay, H. (1973). A colorimetric estimation of reducing sugars in potatoes with 3,5-dinitrosalicylic acid. *Potato Research*, 16, 176-179.
- Liu, P. Y. (2004). *Konjac Science*. Beijing. 225 pp.: China Agriculture Press.
- Liu, P. Y., Lin, Z. S. and Guo, Z. X. (1998). Research and Utilization of *Amorphophallus* in China. *Acta Botanica Yunnanica*, 10, 48-61.
- Liu, P. Y., Zhang, S. L., Zhu, G. H., Chen, Y., Ouyang, H. X., Han, M., Wang, Z.F., Xiong, W. and Peng, H.Y. (2002). Professional standard for the classification, requirements and test methods of konjac flour; Technical Report NY/T 494 Retrieved from <http://www.konjacfoods.com/pdf/NY494-cn.pdf>

- Lobin, W., Neumann, M., Radscheit, M. and Barthlott, W. (2007). The cultivation of titan arum (*Amorphophallus titanum*) – a flagship species for botanic gardens. *The Journal of Botanic Garden Horticulture*, 5, 69-86.
- Long, C. L. (1998). Ethnobotany of *Amorphophallus* of China. *Acta Botanica Yunnanica (Suppl. 10)*, 89-92.
- Madhurima, P., Kuppast, I. J. and Mankani, K. L. (2012). A review on *Amorphophallus paeniifolius*. *International Journal of Advance Scientific Research and Technology*, 2(2), 99-111.
- Mansor, M., Boyce, P. C., Othman, A. S. and Sulaiman, B. (2012). *The Araceae of Peninsular Malaysia*. Penang, Malaysia. 146 pp.: Penerbit Universiti Sains Malaysia.
- Mayo, S. J., Bogner, J. and Boyce, P. C. (1997). *The genera of Araceae*. London: Royal Botanic Gardens Kew.
- Mckenzie, R. J., Ward, J. M., Lovis, J. D. and Breitwieser, I. (2004). Morphological evidence for natural intergeneric hybridization in the New Zealand *Gnaphalieae* (Compositae): *Anaphalioides bellidioides* × *Ewartia sinclairii*. *Botanical Journal of the Linnean Society*, 145, 59-75.
- McPherson, S. and Hetterschied, W. (2011). *Amorphophallus* in the wild and in cultivation. *The Plantsman*, 10(2), 91-97.
- Mekkerdchoo, O., Holford, P., Szrednicki, G., Prakitchaiwattana, C., Borompichaichartkul, C. and Wattananon, S. (2013). Genetic variation among *Amorphophallus* sp. from Northern Thailand and their glucomannan content. *Acta Horticulturae*, 989, 323-330.
- Millar, C. I. and Libby, W. J. (1989). Restoration: Disneyland or a native ecosystem? A question of genetics. *Fremontia*, 17, 3-10.
- Miller, G. L. (1959). Use of dinitrosalicylic acid reagent for determination of reducing sugar. *Analytical Chemistry*, 31, 426-428.
- Miura, K. and Watanabe, K. (1985). Effect of seed corm age and weight on the efficiency of the corm tuberization in konjac plants (*Amorphophallus konjac* K. Koch). *Japanese Journal of Crop Science*, 54, 1-7.
- Moreno, S., Martin, J. P. and Ortiz, J. M. (1998). Inter simple sequence repeats PCR for characterization of closely related grapevine germplasm. *Euphytica*, 101, 117-125.
- Morgante, M., Hanafey, M. and Powell, W. (2002). Microsatellites are preferentially associated with nonrepetitive DNA in plant genomes. *Nature Genetics*, 30, 194-200.

- Muvaffak, A., Dogan, M. and Bilgin, C. C. (2001). A numerical taxonomic study of the genus *Acantholimon* Boiss. (Plumbaginaceae) in Ankara Province (Turkey). *Israel Journal of Plant Sciences*, 49, 298-300.
- Nashinari, K., Williams, P. A. and Philips, G. O. (1992). Review of the physico-chemical characteristics and properties of konjac mannan. *Food Hydrocolloids*, 6, 199-222.
- Nedunchezhiyan, M., Jata, S. K., Ray, R. C. and Misra, R. S. (2011). Management of mealy bug (*Rhizoecus amorphophallus*) in elephant foot yam (*Amorphophallus paenifolius*). *Experimental Agriculture*, 47(4), 717-728.
- Nei, M. (1973). Analysis of gene diversity in subdivided populations. *Proceedings of the National Academy of Science USA*, 70, 321-332.
- Nemati, Z., Zeinalabedini, M., Majidian, P., Jahromi, A. E. and Kiani, D. (2014). Phylogenetic relationships among Iranian and Spanish date palms (*Phoenix dactylifera* L.) revealed by microsatellite markers. *The Journal of Horticultural Science and Biotechnology*, 89(2), 114-120.
- Niwa, T., Etoh, H., Shimizu, A. and Shimizu, Y. (2000). Cis-N-(p-Coumaroyl) serotonin from from Konnyaku, *Amorphophallus konjac* K. Koch. *Bioscience, Biotechnology and Biochemistry*, 64, 2269-2271.
- Ogasawara, S., Yamazaki, H. and Nunomura, W. (1987). Electrophoresis on konjac mannan gel. *Seibutsu Butsuri Kagaku*, 31, 155-158.
- Okimasu, S. and Kishida, N. (1982). Hiroshima Joshi Daigaku. *Kaseigakubu Kiyo*, 13, 1-2.
- Palaniswami, M. S. (1999). Major pests of tropical tuber crops and their management. In R. R. Udpadhyay, K. G. Mukerjee and, O. P. Dubey (Eds.), *IPM system in Agriculture* (Vol. VI). New Delhi: Aditya Books Ltd.
- Palaniswami, M. S. and Peter, K. V. (2008). *Tuber and Root Crops*. New Delhi, India: New India Publishing Agency.
- Parry, J. M. (2010). Konjac glucomannan. In A. Imeson (Ed.), *Food Stabilisers, Thickeners and Gelling Agents* (pp. 198-217). United Kingdom: Blackwell Publishing Ltd.
- Paul, S., Wachira, F. N., Powel, W. and Waugh, R. (1997). Diversity and genetic differentiation among populations of Indian and Kenyan tea (*Camellia sinensis* (L.) O. Kuntze) revealed by AFLP markers. *Theoretical and Applied Genetics* 94, 255-263.
- Pavlista, A. D. (2004). Physiological aging of seed tubers. *16*(1), 1-4. Retrieved from [https://cropwatch.unl.edu/c/document\\_library/get\\_file?folderId=606641&name=DLFE-13914.pdf](https://cropwatch.unl.edu/c/document_library/get_file?folderId=606641&name=DLFE-13914.pdf)

- Pérombelon, M. C. M. and Kelman, A. (1980). Ecology of the soft rot *Erwinias*. *Annual Review of Phytopathology*, 18, 361-387.
- Phadung, T., Krisanapook, K. and Phavaphutanon, L. (2011). Paclobotrazol, water stress and nitrogen induced flowering in 'Khao Nam Phueng' pummelo. *Kasertsart Journal (Natural Science)*, 45, 189-200.
- Poerba, Y. S. and Martanti, D. (2008). Genetic variability of *Amorphophallus muelleri* Blume in Java based on Random Amplified Polymorphic DNA. *Biodiversitas*, 9(4), 245-249.
- Powell, W., Machray, G. C. and Provan, J. (1996a). Polymorphism revealed by simple sequence repeats. *Trend in Plant Science*, 1, 215-222.
- Powell, W., Morgante, M., Andre, C., Hanafey, M., Vogel, J., Tingey, S. and Rafalski, A. (1996b). The comparison of RFLP, RAPD, AFLP and SSR (microsatellite) markers for germplasm analysis. *Molecular Breeding*, 2, 225-238.
- Prychid, C. J., Jabaily, R. S. and Rudall, P. J. (2008). Cellular ultrastructure and crystal development in *Amorphophallus* (Araceae). *Annals of Botany*, 101(7), 983-995.
- Punekar, S. A. and Kumaran, K. P. N. (2010). Pollen morphology and pollination ecology of *Amorphophallus* species from North Western Ghats and Konkan region of India. *Flora*, 205, 326-336.
- Qian, W., Ge, S. and Hong, D. Y. (2001). Genetic variation within and among population of a wild rice *Oryza granulata* from China detected by RAPD and ISSR markers. *Theoretical and Applied Genetics*, 102, 440-449.
- Quattrocchi, U. (2012). *CRC World Dictionary of Medicinal and Poisonous Plants: Common Names, Scientific Names, Eponyms, Synonyms, and Etymology* (Vol. 5). Boca Raton, 3960 pp.: CRC Press.
- Raghu, A., Deepa, V. C. and Sundaran, K. (1999). A study on Soorana (*Amorphophallus paeniifolius*) The King of Tubers. In C. Balagopalan, T. V. R. Nayar, S. Sundaresan and K. R. Lakshmi (Eds.), *Tropical Tuber Crops in Food Security and Nutrition* (pp. 10-14). Calcutta: Oxford and IBH Publishing co. Pte. Ltd.
- Ranjbar, M., Esfahani, M. N., Esfahani, M. N. and Salehi, S. (2011). Phenology and morphological diversity of the main potato cultivars in Iran. *Journal of Ornamental and Horticultural Plants*, 2(3), 201-212.
- Rao, V. R. and Hodgkin, T. (2002). Genetic diversity and conservation and utilization of plant genetic resources. *Plant Cell, Tissue and Organ Culture*, 68, 1-19.
- Ravi, V., Ravindran, C. S. and Suja, G. (2009). Growth and productivity of elephant foot yam *Amorphophallus paeoniifolius* (Dennst. Nicolson) : An Overview. *Journal of Root Crops*, 35(2), 131-142.

- Rencher, A. C. (2002). *Methods of multivariate analysis* (2nd ed.). Canada: John Wiley and Sons, Inc.
- Rohlf, F. J. (1994). *NTSYS-pc. Numerical taxonomy and multivariate analysis system, version 2.2*. New York, USA, 38 pp.: Exeter software.
- Rosado, T. B., Laviola, B. G., Faria, D. A., Pappas, M. R., Bhering, L. L., Quirino, B. and Grattapaglia, D. (2010). Molecular markers reveal limited genetic diversity in a large germplasm collection of the biofuel crop *Jatropha curcas* L. in Brazil. *Crop Science*, 50, 2372-2382.
- Santosa, E., Lian, C. L., Pisooksantivatana, Y. and Sugiyama, N. (2007). Isolation and characterization of polymorphic microsatellite markers in *Amorphophallus paeoniifolius* (Dennst.) Nicolson, Araceae. *Molecular Ecology Notes*, 7, 814-817.
- Santosa, E., Mine, Y., Nakata, M., Lian, C. and Sugiyama, N. (2010). Genetic diversity of cultivated elephant foot yam (*Amorphophallus paeoniifolius*) in Kuningan, West Java as revealed by microsatellite markers. *Journal of Applied Horticulture*, 12(2), 125-128.
- Santosa, E., Sugiyama, N., Hikosaka, S. and Takano, T. (2004). Classification of *Amorphophallus variabilis* in West Java, Indonesia based on morphological characteristics of inflorescences. *Japanese Journal of Tropical Agriculture*, 48, 25-34.
- Santosa, E., Sugiyama, N., Kawabata, S. and Hikosaka, S. (2012). Genetic variations of *Amorphophallus variabilis* Blume (Araceae) in Java using AFLP. *Journal of Agronomy Indonesia*, 40(1), 62-68.
- Santosa, E., Sugiyama, N., Nakata, M. and Lee, O. N. (2006). Growth and corm production of *Amorphophallus* at different shading levels in Indonesia. *Japanese Journal of Tropical Agriculture*, 50(2), 87-91.
- Sattler, R. and Rutishauser, R. (1997). The fundamental relevance of morphology and morphogenesis to plant research. *Annals of Botany* 80, 571-581.
- Seal, H. L. (1964). *Multivariate statistical analysis for biologists*. London, U.K.: Methuen.
- Sedayu, A., Eurlings, M. C. M., Gravendeel, B. and Hetterscheid, W. L. A. (2010). Morphological character evolution of *Amorphophallus* (Araceae) based on a combined phylogenetics analysis of *trnL* and *LEAFY* second intron sequences. *Botany Studies*, 51, 473-490.
- Semagn, K., Bjørnstad, Å. and Ndjiondjop, M. N. (2006). An overview of molecular marker methods for plants. *African Journal of Biotechnology*, 5(25), 2540-2568.

- Shahbudin, D. (2012). *Ecological studies and analysis of glucomannan content in selected Amorphophallus spp. of Peninsular Malaysia*. Master, Universiti Malaya, Kuala Lumpur, Malaysia.
- Shirasu, M., Fujioka, K., Kakishima, S., Nagai, S., Tomizawa, Y., Tsukaya, H., Murata, J., Manome, Y. and Touhara, K. (2010). Chemical identity of a rotting animal-like odor emitted from the inflorescence of the Titan Arum (*Amorphophallus titanum*). *Bioscience, Biotechnology and Biochemistry*, 74(12), 2550-2554.
- Simpson, M. G. (2006). Plant Morphology. In M. G. Simpson (Ed.), *Plant Systematics* (pp. 348-407). London: Elsevier Academic Press.
- Singh, R., Singh, P. P. and Singh, V. (2006). Integrated management of collar rot of *Amorphophallus paenifolius* Blume caused by *Sclerotium rolfsii* Saccardo. *Vegetable Science*, 33(1), 45-49.
- Singh, R., Yadav, R. S., Singh, V. and Singh, P. P. (2005). Integrated management of leaf blight of *Amorphophallus paenifolius* Blume. *Vegetable Science*, 32(2), 169-172.
- Smulders, M. J. M., Bredemeijer, G., Rus-Kortekass, W., Arens, P., and Vosman, B. (1997). Use of short microsatellite from database sequences to generate polymorphism among *Lycopersicon esculentum* cultivars and accessions of other *Lycopersicon* species. *Theoretical and Applied Genetics*, 97, 264-272.
- Sneath, P. H. A. and Sokal, R. R. (1973). *Numerical Taxonomy The Principal and Practice of Numerical Classification*. San Francisco, U.S.A: W.H. Freeman and Company.
- Souza, S. G. H., Carpentieri-Pípolo, V., Ruas, C. F. and Carvalho, V. P. (2008). Comparative analysis of genetic diversity among the maize inbred lines (*Zea mays* L.) obtained by studying genetic relationships in *Lactuca* spp. *Theoretical and Applied Genetics*, 93, 1202-1210.
- Su, M., Tsou, C. and Hsieh, C. (2007). Morphological Comparisons of Taiwan Native Wild Tea Plant (*Camellia sinensis* (L.) O. Kuntze forma *formosensis* Kitamura) and Two Closely Related Taxa Using Numerical Methods. *Taiwania*, 52(1), 70-83.
- Sugiyama, N., Santosa, E., Lee, O., Hikosaka, S. and Nakata, M. (2006). Classification of elephant foot yam (*Amorphophallus paeoniifolius*) cultivars in Java using AFLP markers. *Japanese Journal of Tropical Agriculture*, 50, 215-218.
- Sugiyama, N., Shimahara, H., Andoh, T., Takemoto, M. and Kamata, T. (1972). Molecular weights of konjac mannans of various sources. *Agricultural and Biological Chemistry*, 36, 1381-1387.
- Sun, G. L. and Solomon, B. (2003). Microsatellite variability and heterozygote deficiency in the arctic-alpine Alaskan wheatgrass (*Elymus alaskanus*) complex. *Genome*, 46, 729-737.

- Supapvanich, S., Prathan, P. and Tepsorn, R. (2012). Browning inhibition in fresh-cut rose apple fruit cv. Taaptimjaan using konjac glucomannan coating incorporated with pineapple fruit extract. *Postharvest Biology and Technology*, 73, 46-49.
- Takigami, S. (2000). Konjac mannan. In G. O. Phillips and P. A. Williams (Eds.), *Handbook of Hydrocolloids* (pp. 413-424). Florida: CRC Press.
- Tautz, D., Trick, M. and Dover, G. A. (1986). Cryptic simplicity in DNA is a major source of genetic variation. *Nature*, 322, 652-656.
- Tohme, J., Gonzalez, D. O., Beebe, S. and Duque, M. C. (1996). AFLP analysis of gene pool of a wild bean core collection. *Crop Science*, 36, 1375-1384.
- Udupa, S. M. and Baum, M. (2001). High mutation rate and mutational bias at (TAA)<sub>n</sub> microsatellite loci in chickpea (*Cicer arietinum* L.). *Molecular Genetics and Genomics* 265(6), 1097-1103.
- Upadhyaya, H. D., Gowda, C. L. L. and Reddy, V. G. (2007). Morphological diversity in finger millet germplasm introduced from Southern and Eastern Africa. *Journal of SAT Agricultural Research*, 3(1), 1-3.
- Upton, K. (1998). Leaf cuttings of *Amorphophallus titanum*. *Newsletter International Aroid Society*, 20(2), 6.
- Vlot, A. C., Klessig, D. F. and Park, S. W. (2008). Systemic acquired resistance: the elusive signal(s). *Current Opinion in Plant Biology*, 11, 436-442.
- Vos, P., Hogers, R., Bleeker, M., Reijans, M., Lee van de, T., Hornes, M., et al. (1995). AFLP: a new technique for DNA fingerprinting. *Nucleic Acids Research*, 23, 4407-4414.
- Vuksan, V., Sievenpiper, J. L., Xu, Z., Wong, E. Y. Y., Jenkins, A. L., Zdravkovic, U. B., Leiter, L.A., Josse, R.G. and Stavro, M.P. (2001). Konjac-mannan and American ginseng: emerging alternative therapies for type 2 diabetes mellitus. *Journal of the American College of Nutrition*, 20, 370-380.
- Wang, Z. F., Zhang, S. L. and Liu, P. Y. (2001). Study on the method of weighing mannose-hydrazone for determining content of glucomannan in konjac powder. *Food and Fermentation Industries*, 27, 53-56.
- Weinbing, C., Chunlin, W. and Jianfu, Z. (2001). Study on genetic diversity of RAPD markers in *Amorphophallus*. *Journal of Agriculture Biotechnology*, 60, 27-31.
- Weising, K., Nybom, H., Wolff, K. and Kahl, G. (2005). *DNA Fingerprinting in Plants: Principles, Methods and Applications*. Boca Raton, United States: CRC Press.
- WHO. (2014). Obesity and overweight. Retrieved 20 August, 2014, from <http://www.who.int/mediacentre/factsheets/fs311/en/>

- Williams, J. G. K., Kublelik, A. R., Livak, K. J., Rafalski, J. A. and Tingey, S. V. (1990). DNA polymorphism's amplified by arbitrary primers are useful as genetic markers. *Nucleic Acids Research*, 18, 6531-6535.
- WIPO. (1993). Clarified konjac glucomannan Retrieved 20 October 2012, from <http://www.sumobrain.com/patents/wipo/Clarifiedkonjacglucomannan/WO19930030471>.
- World Food Science. (2003). Part II: konjac-a program to help reduce poverty in northern Guangdong, China Retrieved 13 October 2012, from <http://www.worldfoodscience.org/cms/?pid=1003561>
- Wotton, A. J., Luker-Brown, M., Westcott, R. J. and Cheetam, P. S. J. (1993). The extraction of a glucomannan polysaccharide from konjac corms (Elephant Yam, *Amorphophallus riverii*). *Journal Science Food and Agriculture*, 61, 429-433.
- Wright, S. (1978). *Evolution and the Genetics of Populations: Variability within and among natural populations* (Vol. 4). Chicago: The University Chicago Press.
- Wu, C., Peng, S., Wen, C., Wang, X., Fan, L., Denga, R. and Pang, J. (2012). Structural characterization and properties of konjac glucomannan/curdlan blend films. *Carbohydrate Polymers*, 89, 497-503.
- Wu, J., Diao, Y., Gu, Y. and Hu, Z. (2010). Infection pathway of soft rot pathogens on *Amorphophallus konjac*. *African Journal of Microbiology Research*, 4(14), 1495-1499.
- Wu, Y. J., Meng, W. N., Chai, J. T. and Wang, J. F. (2002). Extraction of glucomannan from *Amorphophallus konjac*. *Science and Technology Food Industry*, 23, 41-43.
- Xiu, J. H., Ji, G. H., Wang, M., Yang, Y. L. and Li, C. Y. (2006). Molecular identification and genetic diversity in Konnyaku's soft rot bacteria. *Acta Microbiologica Sinica*, 46, 522-525.
- Yang, L., Yang, X., Petcavich, R. and Mao, L. (2001). Coating materials for preserving fresh produce. *United States Patent* 6, 23.
- Yee, M. C. F. (2011). *An investigation of the biology and chemistry of the chinese medicinal plant, Amorphophallus konjac*. Doctor of Philosophy Unpublished, University of Wolverhampton, United Kingdom.
- Yeh, F. C. and Boyle, T. J. B. (1997). Population genetic analysis of co-dominant and dominant markers and quantitative traits. *Belgian Journal of Botany*, 129, 157.
- Zanelle, C. M., Bruxel, M., Paggi, G. M., Goetze, M., Buttow, M. V., Cidade, F. W. and Bered, F. (2011). Genetic structure and phenotypic variation in wild populations of the medicinal tetraploid species *Bromelia antiacantha* (Bromeliaceae). *American Journal of Botany*, 98, 1511-1519.



- Zhang, C., Chen, J. and Yang, F. (2014). Konjac glucomannan, a promising polysaccharide for OCDDS. *Carbohydrate Polymer*, 104, 175-181.
- Zhao, H., Yu, J., You, F. M., Luo, M. and Peng, J. (2011). Transferability of microsatellite markers from *Brachypodium distachyon* to *Miscanthus sinensis*, a potential biomass crop. *Journal of Integrative Plant Biology*, 53(3), 231-245.
- Zhao, J., Zhang, D., Jianping, Z., Srzednicki, G., Borompichaichartkul, C. and Kanlayanarat, S. (2010a). Morphological and growth characteristics of *Amorphophallus muelleri* blume: a commercially important konjac species. *Acta Horticulturae*, 875, 501-508.
- Zhao, J., Zhang, D., Srzednicki, G., Kanlayanarat, S. and Borompichaichartkul, C. (2010b). Development of a low-cost two stage technique for production of low-sulphur purified konjac flour. *International Food Research Journal*, 17, 1113-1124.
- Zietkiewicz, E., Rafalski, A. and Labuda, D. (1994). Genome fingerprinting by simple sequence repeat (SSR)-anchored polymerase chain reaction amplification. *Genomics*, 20, 176-183.
- Zohary, D. (2004). Unconscious selection and the evolution of domesticated plants. *Economic Botany*, 58(1), 5-10.