



UNIVERSITI PUTRA MALAYSIA

***IMPACT OF WATER STRESS AND CO₂ ENRICHMENT ON GROWTH
AND FIBER DEVELOPMENT OF KENAF (*Hibiscus cannabinus* L.)***

AMIR MAHDI KHALATBARI

FP 2016 43



**IMPACT OF WATER STRESS AND CO₂ ENRICHMENT ON GROWTH AND
FIBER DEVELOPMENT OF KENAF (*Hibiscus cannabinus* L.)**

By

AMIR MAHDI KHALATBARI

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfillment of the Requirements for the Degree of Doctor of Philosophy**

July 2016

© COPYRIGHT UPM



All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



© COPYRIGHT UPM



DEDICATION

Dedicated with Love to

My Kind Father, Mohsen Khalatbari

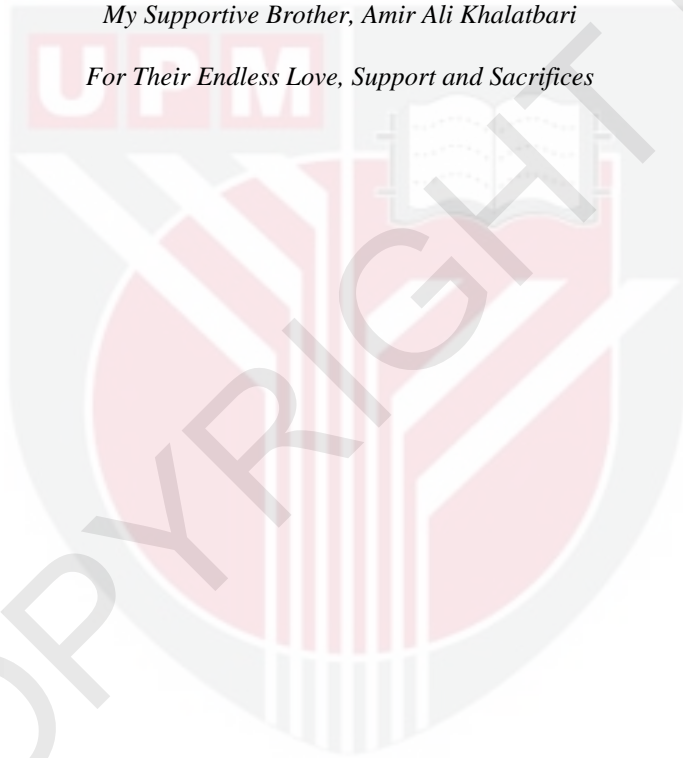
and

My Beloved Mother, Azam Porsheikhani

and

My Supportive Brother, Amir Ali Khalatbari

For Their Endless Love, Support and Sacrifices



© COPYRIGHT UPM



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Doctor of Philosophy

IMPACT OF WATER STRESS AND CO₂ ENRICHMENT ON GROWTH AND FIBER DEVELOPMENT OF KENAF (*Hibiscus cannabinus* L.)

By

AMIR MAHDI KHALATBARI

July 2015

Chairman: Associate Professor Hawa ZE Jaafar, PhD
Faculty: Agriculture

In this study, the effect of some environmental factors (different water treatments and CO₂ concentration) on morphological, physiological and histochemical characteristics of some kenaf varieties namely Fuhong (FH991, FH992 and FH952), V36, Kohn-Kaen60 (KK60) and TK were considered. Kenaf (*Hibiscus cannabinus* L.), a fast growing C3 plant native of tropical Africa, is being investigated as a new source of bioenergy as well as an industrial crop and has high potential to be used in Malaysia. Kenaf is considered as a great source of cellulose fiber for a wide range of paper products. Information on lignocellulose properties (lignin and cellulose) of these kenaf varieties as affected by environmental factors such as water stress, time of water stress imposition and CO₂ enrichment are still scarce and such data would be useful to provide information on fiber products containing high levels of lignocellulose attributes. A Randomized Complete Block Design (RCBD) experiment was conducted to determine performance and growth rate of varieties namely Fuhong (FH991, FH992 and FH952), V36, Kohn-Kaen60 (KK60) and TK during the seedling establishment. Seedling establishment is one of plant's growth phases in which some of the factors are precisely influential in final stage of growth and development for kenaf which is known as fiber product. The results showed that different varieties had significant effects on growth parameters such as shoot height and stem diameter which are important for producing higher fibre content and yield. Varieties FH991 and V36 obtained higher values of plant height (74.2 cm and 72 cm respectively) and stem diameter (5.53 mm and 5.50 mm) whereas variety KK60 recorded the lowest value for root parameters such as root average diameter value which was 0.62 mm. These three varieties were selected for further studies for their fiber yield and quality evaluation. A split plot experiment was arranged to determine effect of three different water treatments (100% ER; well watered, 50% ER; high water stress and 25%

ER; severe water stress) on morphological, physiological, fiber yield and lignocellulose quality of varieties FH991, V36 and KK60. The highest plant height belonged to variety FH991 at 100% of ER and followed by variety V36 (242.67 cm and 230.66 cm respectively) whereas variety KK60 with water treatment of 25% of ER obtained the lowest average height of 190.67 cm at the end of experimental period. The highest net photosynthesis rate belonged to variety FH991 at 100% of ER with the value of $23.57 \mu\text{mol m}^{-2} \text{s}^{-1}$ at the end of experimental period. The lowest net photosynthesis rate $10.87 \mu\text{mol m}^{-2} \text{s}^{-1}$ was recorded by variety V36 at 25% of ER. The highest fiber length for bast was recorded by varieties FH991 and V36 (2.59 mm and 2.57 mm respectively) at 100% ER. The highest fiber length of core belonged to varieties V36 and FH991 with 100% ER recording the value of 0.95 mm and 0.93 mm respectively. The lowest fiber length for bast belonged to variety V36 at 25% ER with the value of 1.32 mm. The lowest fiber length for core was recorded by variety KK60 with the value of 0.48 mm. The optimum value of bast fiber yield was obtained by variety V36 with 100% ER water treatment ($13.3 \text{ g plant}^{-1}$) followed by variety FH991 ($12.96 \text{ g plant}^{-1}$) and KK60 ($11.77 \text{ g plant}^{-1}$) respectively. The lowest value for bast fiber yield was recorded by variety V36 at 25% ER ($4.98 \text{ g plant}^{-1}$). The highest core fiber yield of $21.72 \text{ g plant}^{-1}$ belonged to variety FH991 with 100% ER water treatment which was followed by variety V36 with value of $21.32 \text{ g plant}^{-1}$. Variety V36 at 25% ER attained the lowest value of $8.44 \text{ g plant}^{-1}$. The evidence from this study elucidated that the fiber quality of for all three varieties decreased by increasing the severity of water stress from 100% ER to 25% ER. For the third experiment a split plot experiment was arranged to determine effect of three different times of water stress imposition at different growth stages namely daily watering (100% ER; well watered), water stress imposition 1 month after seedling establishment completion (1MAS) and finally water stress imposition at flowering stage (AFS) on morphological and histochemical attributes of varieties FH991, V36 and KK60. The highest value of net photosynthesis rate belonged to variety FH991 ($23.33 \mu\text{mol m}^{-2} \text{s}^{-1}$) and it was followed by varieties KK60 with value of $23.27 \mu\text{mol m}^{-2} \text{s}^{-1}$ and V36 at $21.63 \mu\text{mol m}^{-2} \text{s}^{-1}$. The lowest net photosynthesis rate of $11.37 \mu\text{mol m}^{-2} \text{s}^{-1}$ belonged to variety KK60 subjected to water treatment of 1MAS. Considering the impact of different water treatments on fiber dimensions of these varieties, the highest bast and core fiber length of 2.59 mm and 0.91 mm was recorded when all varieties were under control water treatment (no stress). The lowest bast and core fiber length belonged to water treatment of 1MAS with value of 1.62 mm and 0.64 mm respectively. The highest bast holocellulose, α -cellulose and lignin belonged to varieties under control treatment (85.22%, 56.42% and 13.75% respectively). Core lignocellulose attributes attained highest percentages under control water treatment in which holocellulose recorded 83.17%, α -cellulose 46.85% and lignin 20.08%. The lowest bast and core lignocelluloses belonged to varieties under water stress treatment of 1MAS. Bast holocellulose of 80.67%, α -cellulose of 56.42 and lignin of 13.75% were recorded when plants were under water stress of 1MAS. Plants under water stress treatment of 1MAS had lowest core holocellulose of 78.63%, α -cellulose of 41.97% and lignin of 15.24. As an interesting result water stress imposition at flowering stage increased bast fiber length and core lumen diameter for selected varieties that could describe possible positive effect

of water stress imposition in this stage of growth on kenaf fiber dimension. For the last experiment varieties FH991, V36 and KK60 were exposed to 400 and 800 $\mu\text{mol mol}^{-1}$ CO_2 in a split plot experiment. Results of the analysis of variance showed that different CO_2 concentrations (400 $\mu\text{mol mol}^{-1}$ and 800 $\mu\text{mol mol}^{-1}$) had significant impact on morphological, physiological and histochemical properties of all three varieties for all traits measured; indicating enriched CO_2 level (800 $\mu\text{mol mol}^{-1}$) had a huge impact on measured traits. The highest bast fiber length was recorded by 800 $\mu\text{mol mol}^{-1}$ CO_2 level with value of 3.10 mm whereas CO_2 level of 400 $\mu\text{mol mol}^{-1}$ recorded bast fiber length of 2.68 mm. For core fiber attributes, the highest core fiber length belonged to elevated CO_2 level (800 $\mu\text{mol mol}^{-1}$) with value of 0.98 mm whereas ambient CO_2 level (400 $\mu\text{mol mol}^{-1}$) recorded core fiber length of 0.92 mm. The highest bast holocellulose, α -cellulose and lignin belonged to varieties under enriched CO_2 level of 800 $\mu\text{mol mol}^{-1}$ (87.34%, 57.85% and 14.32% respectively). Core lignocellulose attributes attained highest percentages under elevated CO_2 level (800 $\mu\text{mol mol}^{-1}$) in which holocellulose recorded 84.24%, α -cellulose 47.52% and lignin 21.60%. The most obvious finding to emerge from this study is that CO_2 enriched kenaf plants exhibited the ability to synthesize higher fiber yield and lignocellulose properties which were not detected from kenaf grown under ambient CO_2 concentration. These results indicate that the fiber yield and histochemical attributes of these kenaf varieties can be enhanced by controlled environment production and CO_2 enrichment in top soil.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah.

**PENGARUH TEKANAN AIR DAN PERKAYAAN CO₂ TERHADAP
PERTUMBUHAN DAN PEMBENTUKAN FIBER KENAF
(*Hibiscus cannabinus* L.)**

Oleh

AMIR MAHDI KHALATBARI

Julai 2015

**Pengerusi: Prof. Madya Hawa ZE Jaafar, PhD
Fakulti: Pertanian**

Kenaf (*Hibiscus cannabinus* L.), tumbuhan C3 yang cepat membesar berasal dari Afrika tropika, sedang disiasat sebagai sumber baru bioteknologi dan juga sebagai tanaman industri berpotensi tinggi untuk digunakan di Malaysia. Kenaf dipertimbangkan sebagai sumber hebat fiber selulosa untuk produk kertas berskala besar. Satu kajian telah dilakukan untuk menilai dan membandingkan morfologi, fisiologi dan ciri-ciri histokimia enam variety kenaf terhadap perbezaan rawatan air dan tahap CO₂ di Serdang, Selangor, Malaysia. Objektif kajian ini secara keseluruhan adalah untuk menentukan keadaan optimum dari segi tekanan air dan kepekatan CO₂ untuk hasil dan kualiti fiber. Secara umum, semua variety kenaf adalah lebih baik apabila disiram dengan secukupnya berbanding dengan kenaf yang diletakkan di bawah rawatan tekanan air berdasarkan kaedah penggantian evapotranspirasi atau perbezaan masa tekanan air dilaksanakan. Varieti FH991 dan V36 didapati menghasilkan panjang pucuk paling tinggi dan diameter batang yang merupakan ciri paling penting untuk penghasilan hasil. Nilai tertinggi untuk kadar bersih fotosintesis direkodkan oleh variety FH991 dan V36 berikutan rawatan air kawalan (tiada tekanan) yang mana disebabkan konduksi stomata yang tinggi. Pengurangan adalah lebih parah dalam kes potensi air daun, membawa kepada stomata yang hampir tertutup, pengurangan pengambilan CO₂ dan menyebabkan aktiviti fotosintesis yang terhad untuk semua variety di bawah rawatan tekanan air. FH991 dan V36 mencapai hasil kulit fiber, hasil pusat fiber, hasil keseluruhan fiber dan nilai lignoselulosa tertinggi yang mana lebih signifikan dari tumbuhan dibawah tekanan air. Keputusan analisis variant menunjukkan perbezaan kepekatan CO₂ (400 $\mu\text{mol mol}^{-1}$ dan 800 $\mu\text{mol mol}^{-1}$) mempunyai kesan yang signifikan terhadap morfologi, fisiologi dan sifat histokimia kesemua tiga variety untuk semua ciri yang diukur; menunjukkan kekayaan tahap CO₂ (800 $\mu\text{mol mol}^{-1}$) mempunyai kesan yang besar terhadap ciri-ciri yang diukur. Tumbuhan yang terdedah

kepada perkayaan tahap CO₂ 800 μmol mol⁻¹ juga didapati merekodkan tinggi pokok dan biomas keseluruhan tertinggi. Di kalangan varieti, FH991 mempunyai ketinggian pokok, luas daun keseluruhan dan biomas keseluruhan paling tinggi yang mana memberikan hasil fiber keseluruhan tertinggi. Ciri fisiologi terutamanya kadar fotosintesis dan kadar transpirasi meningkat apabila tumbuhan didedahkan kepada perkayaan tahap CO₂ terutamanya FH991 dan ini kerana konduksi stomata yang tinggi yang dilakukan oleh tumbuhan dibawah kepekatan CO₂ yang tinggi iaitu 800 μmol mol⁻¹. Varieti berbeza tidak mempunyai kesan signifikan terhadap ciri fisiologi dan tiada kesan interaksi yang signifikan antara varieti dan tahap perkayaan CO₂ terhadap ciri fisiologi. Apabila tumbuhan terlibat dengan perkayaan tahap CO₂ 800 μmol mol⁻¹, varieti FH991 dan V36 mempunyai hasil kulit dan pusat fiber yang tertinggi dan menunjukkan nilai panjang fiber dan ciri lignoselulosa yang lebih tinggi terutamanya holoselulosa dan α-selulosa yang mana penting untuk kualiti fiber yang tinggi sebagai pertimbangan untuk pengeluaran kertas. Mengambil kira keseluruhannya, kajian ini memberikan maklumat bernilai untuk pembentukan fiber, ciri-ciri histokimia varieti kenaf tempatan yang mana penting untuk usaha pembangunan di masa hadapan.

ACKNOWLEDGEMENTS

I would like to express my sincere thanks to Associate Professor Dr. Hawa ZE Jaafar, chairman of my supervisory committee, for her dedicated efforts, support, invaluable advice and intellectual guidance during the accomplishment of this research work. I would also like to thank my supervisory committee members, Associate Professor Dr. Hazandy Abdul Hamid and Dr. Mohd. Ridzwan Abd Halim for their guidance, assistance, encouragements and constructive comments throughout the period of this study. I greatly appreciate all the help and support provided by the supervisory committee during my study in Malaysia.

Special thanks go to Universiti Putra Malaysia (UPM) for their contribution for this project number of 5523867 FRGS (Fundamental Research Grant Scheme).

I am very grateful to Mr. Rodhi Ahmed and all staff/officers of TPU and Ladang 2, Universiti Putra Malaysia for their help during my field work. I am also grateful to the laboratory technicians of the Department of Crop Science, Universiti Putra Malaysia.

My sincere thanks and appreciations also go to my friends and fellow students especially Dr. Ali Ghasemzadeh and Dr. Ehsan Karimi for their help, support and encouragements during the period of my study.

My deepest gratitude goes to my father Mohsen, my mother Azam and my brother Ali for their help and continuous moral support throughout my study.

I certify that a Thesis Examination Committee has met on 8 July 2015 to conduct the final examination of Amir Mahdi Khalatbari on his thesis entitled “Impact of Water Stress and CO₂ Enrichment on Growth and Fiber Development of Kenaf (*Hibiscus cannabinus* L.)” in accordance with the Universities and University College Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A)106] 15 March 1998. The committee recommends that the student be awarded the degree of Doctor of Philosophy.

Members of the thesis Examination Committee were as follows:

Uma Rani a/p Sinniah, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Mohd Rafi bin Yousp, PhD

Professor
Institute of Tropical Agriculture
Universiti Putra Malaysia
(Internal Examiner)

Izham bin Ahmad, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Internal Examiner)

Hakoomat Ali, PhD

Professor
Department of Agronomy
Faculty of Agriculture Science and Technology
Bahauddin Zakariya University Multan, Pakistan
(External Examiner)

ZULKARNAIN ZAINAL, PhD

Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Hawa Zee Jaafar, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Hazandy Abdul Hamid, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

Mohd Ridzwan A. Halim, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: AMIR MAHDI KHALATBARI / GS26937

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____
Name of Chairman of
Supervisory
Committee: Associate Professor Dr. Hawa Zee Jaafar

Signature: _____
Name of Member of
Supervisory
Committee: Associate Professor Dr. Hazandy Abdul Hamid

Signature: _____
Name of Member of
Supervisory
Committee: Associate Professor Dr. Mohd Ridzwan A. Halim

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iv
ACKNOWLEDGEMENTS	vi
APPROVAL	vii
DECLARATION	ix
LIST OF TABLES	xiv
LIST OF FIGURES	xvii
LIST OF ABBREVIATIONS	xix

CHAPTER

1 INTRODUCTION	1
1.1 Justification	2
1.2 Hypothesis	3
1.3 General Objectives	3
1.4 Specific Objectives	3
2 LITERATURE REVIEW	4
2.1 Origin of Kenaf	4
2.2 Taxonomy and Botany of Kenaf	4
2.3 Environmental conditions for kenaf growth	5
2.4 Importance of kenaf to be cultivated	5
2.4.1 Kenaf production in Malaysia	5
2.4.2 Uses of kenaf	9
2.5 Agronomy of Kenaf	9
2.6 Kenaf Fiber Morphology and Quality	10
2.7 Water Stress and Its Effects on Plants	11
2.7.1 Morphological Effects	12
2.7.2 Physiological Effects	13
2.8 Carbon Dioxide and Plant Responses	16
2.9 Carbon Dioxide Enrichment	17
2.10 Plants Adaptation to Increases in CO ₂ Levels	19
2.10.1 Effects on Biomass and Growth	19
2.10.2 Leaf Gas Exchange Effects under Elevated CO ₂	20
2.10.3 Maximal Photochemical Efficiency (Fv/Fm) under CO ₂ Enrichment	21
2.10.4 Carbohydrates Increases under Elevated CO ₂	21
3 THE EVALUATION OF SEEDLING ESTABLISHMENT OF SIX DIFFERENT VARIETIES OF YOUNG KENAF (<i>Hibiscus cannabinus</i> L.) UNDER GLASSHOUSE CONDITION	23
3.1 Introduction	23
3.2 Materials and Methods	24
3.2.1 Experimental Site	24
3.2.2 Plant Material	24
3.2.3 Plant Maintenance, Irrigation System and Watering Regulation	24
3.2.4 Soil Sampling and Analysis	24

3.2.5	Growth Measurements	25
3.2.6	Root System Measurements	25
3.2.7	Experimental Design and treatments	26
3.2.8	Data Collection	26
3.2.9	Data Analysis	27
3.3	Results and discussion	27
3.3.1	Growth Attributes	27
3.3.2	Root System Attributes	30
3.3.3	Correlation between Shoot and Root	31
3.4	Conclusion	32
4	THE EFFECT OF DIFFERENT WATER TREATMENTS ON GROWTH, GAS EXCHANGE RATE, WATER RELATIONS, PHYSICAL AND HISTOCHEMICAL FIBER PROPERTIES OF THREE VARIETIES OF KENAF (<i>Hibiscus cannabinus</i> L.)	33
4.1	Introduction	34
4.2	Materials and Methods	34
4.2.1	Experimental Site	34
4.2.2	Plant Material	34
4.2.3	Water Treatments Application	35
4.2.4	Growth Parameters	35
4.2.5	Gas Exchange Parameters	35
4.2.6	Water Relationship Attributes	34
4.2.7	Fiber Dimensions and Their Values	36
4.2.8	Chemical Analysis	36
4.2.9	Fiber Quality Data Collection	37
4.2.10	Experimental Design and Statistical Analysis	39
4.3	Results and Discussion	39
4.3.1	Growth and Biomass Allocation	40
4.3.2	Gas Exchange Measurement	42
4.3.3	Water Relationship Measurement	42
4.3.4	Correlation among Growth Parameters, Gas Exchange Attributes and Water Relation	43
4.3.5	Fiber Dimension	45
4.3.6	Fiber Values	46
4.3.7	Fiber Yield	47
4.3.8	Correlation among Bast and Core Fiber Parameters	51
4.3.9	Chemical Properties	51
4.4	Conclusion	54
5	THE EFFECT OF DIFFERENT TIMES OF WATER STRESS IMPOSITION ON MORPHOLOGICAL, PHYSIOLOCAL TRAITS AND FIBER DIMENSION OF THREE VARIETIES OF KENAF (<i>Hibiscus cannabinus</i> L.)	55
5.1	Introduction	55
5.2	Materials and Methods	56
5.2.1	Experimental Site	56
5.2.2	Plant Material	56
5.2.3	Water Treatment Application	56
5.2.4	Growth Parameters, Gas Exchange Parameters and Water Relationship Attributes	57

5.2.5	Chlorophyll Content	57
5.2.6	Chlorophyll Fluorescence	57
5.2.7	Fiber Dimension, Lignocellulose Properties and Chemical Analysis	57
5.2.8	Experimental Design and Statistical Analysis	57
5.3	Results and Discussion	58
5.3.1	Growth and Biomass Allocation	58
5.3.2	Gas Exchange Measurement	59
5.3.3	Water Relationship Measurement	62
5.3.4	Total Chlorophyll Content and chlorophyll Fluorescence (Fv/Fm)	63
5.3.5	Correlation among Growth Parameters, Gas Exchange Attributes and Water Relation	64
5.3.6	Fiber Dimension	64
5.3.7	Fiber Values	65
5.3.8	Chemical Properties	66
5.3.9	Fiber Yield	69
5.4	Conclusion	72
6	GROWTH, GAS EXCHANE TRAITS, FIBER DIMENSION AND LIGNOCELLULOSE PROPERTIES OF THREE VARIETIES OF KENAF (<i>Hibiscus cannabinus</i> L.) UNDER DIFFERENT CO₂ LEVELS	73
6.1	Introduction	73
6.2	Materials and Methods	74
6.2.1	Experimental Location, Plant Materials and Treatments	74
6.2.2	Growth House Microclimate and CO ₂ Enrichment Treatment	75
6.2.3	Growth Parameters, Gas Exchange Parameters, Fiber dimension and lignocellulose properties	76
6.2.4	Experimental Design and Statistical Analysis	76
6.3	Results and Discussion	76
6.3.1	Growth and Biomass Allocation	78
6.3.2	Gas Exchange Measurement	78
6.3.3	Fiber Dimension	79
6.3.4	Fiber Values	82
6.3.5	Chemical Properties	83
6.3.6	Fiber Yield	84
6.4	Conclusion	85
7	SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCHES	86
	REFERENCES	89
	APPENDICES	108
	BIODATA OF STUDENT	121
	LIST OF PUBLICATIONS	122

LIST OF TABLES

Table	Page
2.1 Kenaf production in different countries (FAO, 2012).	8
2.2 Fiber dimensions of bast and core of kenaf.	11
2.3 Influence of water stress on Carambola vegetative growth (Ismail et al., 1994)	12
2.4 Properties of carbon dioxide	18
2.5 Photosynthetic characteristics of three major plant groups	21
3.1 Six varieties of Kenaf used in the studies with source and country of origin.	25
3.2 The characteristics of top-clay soil (Serdang series) used in the experiment.	26
3.3 Means comparison of cumulative growth rate (CGR) and root system parameters of six Kenaf (<i>Hibiscus cannabinus</i> L.) varieties; FH991, FH992, FH952, V36, TK and KK60. Means followed by ($P \leq 0.05$) differ significantly by Duncan at 4 weeks of age. N=3.	31
3.4 Pearson's correlation coefficients between growth and root system attributes of six varieties of kenaf (FH991, FH992, FH952, V36, TK and KK60) at 4 weeks of age.	31
4.1 Plant height, Stem diameter, Total leaf area and total dry biomass of three different kenaf varieties subjected to three different water treatments at three months of age. N=3.	41
4.2 Net photosynthesis rate, water potential, stomata conductance and leaf relative water content of three different kenaf varieties subjected to three different water treatments at three months of age. N=3.	44
4.3 Pearson's correlation coefficients among the mean shoot height, stem diameter, total biomass, stomata conductance and net photosynthesis rate of three varieties of kenaf (FH991, V36 and KK60) under three different water treatments (25%, 50% and 100% of evapotranspiration replacement) at three months of age.	45
4.4 Interaction effect of different water treatments (25%, 50% and 100% ER) and three different kenaf varieties: Fuhong 991 (FH991), V36 and Kohn-Kaen 60 (KK60) on fibre length, fibre diameter, fibre lumen diameter and cell wall thickness of bast and core at the end of experimental period (at three months of age). N=3.	48
4.5 Interaction effect of different water treatments (25%, 50% and 100% ER) and three different kenaf varieties including: Fuhong 991 (FH991), V36 and Kohn-Kaen 60 (KK60) on Slender ratio, Suppleness coefficient and Runkle ratio of bast and core at the end of experimental period (at three months of age). N=3.	49

4.6	Pearson's correlation coefficients among the mean bast fibre yield, bast diameter, bast lumen diameter, bast cell wall thickness, core fibre yield, core diameter, core lumen diameter and core cell wall thickness of three varieties of kenaf (FH991, V36 and KK60) under three different water treatments (25%, 50% and 100% of ER) (At three months of age).	51
4.7	Bast and core lignocellulose properties (holocellulose, α -cellulose and lignin) of three different kenaf varieties (Fuhong 991(FH991), V36 and Kohn-Kaen 60(KK60) (at three months of age). N=3.	53
4.8	Bast and core lignocellulose properties (holocellulose, α -cellulose and lignin) of kenaf varieties (Fuhong 991(FH991), V36 and Kohn-Kaen 60(KK60) affected by different water treatments of 25%, 50% and 100% ER (at three months of age). N=3.	53
4.9	Interaction effect of different water treatments (25%, 50% and 100% ER) and three different kenaf varieties including: Fuhong 991 (FH991), V36 and Kohn-Kaen 60 (KK60) on α -cellulose and lignin of core fiber (at three months of age). N=3.	54
5.1	Plant height, Stem diameter, Total leaf area and total dry biomass of three different kenaf varieties subjected to three different water treatments at three months of age. N=3.	60
5.2	Means of photosynthesis rate, water potential, stomata conductance and transpiration rate of three different kenaf varieties subjected to three different water treatments at three months of age. N=3.	61
5.3	Means of maximum quantum efficiency, total chlorophyll content and Intrinsic water use efficiency of three different kenaf varieties subjected to three different water treatments at three months of age. N=3.	63
5.4	Pearson's correlation coefficients among the mean shoot height stem diameter, total biomass, total leaf area, water potential, stomata conductance and net photosynthesis rate of three varieties of kenaf (FH991, V36 and KK60) under three different water treatments at three months of age.	64
5.5	Means of fiber dimension of bast and core of different Kenaf varieties: Fuhong 991 (FH991), V36 and Kohn-Kaen 60 (KK60) at three months of age. N=9.	67
5.6	Impact of different times of water stress imposition on fiber dimension of BAST and CORE of three varieties of Kenaf Fuhong 991 (FH991), V36 and Kohn-Kaen 60 (KK60) at three months of age. N=9.	67
5.7	Bast and core fiber values of three varieties of Kenaf (Fuhong 991 (FH991), V36 and Kohn-Kaen 60 (KK60)) under three water treatments (Control, water stress imposition after 2 months of	68

growth (1MAS) and water stress imposition at flowering stage initiation (AFS) at three months of age. N=9.

- 5.8 Effect of three different water treatments on Holocellulose, α -cellulose and Lignin of three varieties of Kenaf including: Fuhong 991 (FH991), V36 and kohn-kaen 60 (KK60) at three months of age. N=9. 69
- 6.1 Plant height, stem diameter, total leaf area and total dry biomass of three different kenaf varieties subjected to two different CO₂ levels at three months of age. N=9. 77
- 6.2 Plant height, Stem diameter, Total leaf area and total dry biomass of three different kenaf varieties namely: FH991, V36 and KK60 at the end of experiment period subjected to two different CO₂ levels at three months of age. N=6. 77
- 6.3 Net photosynthesis rate, stomata conductance, transpiration rate and Water Use Efficiency (WUE) of three varieties of kenaf namely: FH991, V36 and KK60 subjected to two different CO₂ levels (400 and 800 $\mu\text{mol mol}^{-1}$) at three months of age. N=9. 78
- 6.4 Bast and core fiber dimensions of three different kenaf varieties (FH991, V36 and KK60) subjected to two different CO₂ levels (400 and 800 $\mu\text{mol mol}^{-1}$) at three months of age. N=9. 81
- 6.5 Bast and core fiber length and diameter of three different kenaf varieties (FH991, V36 and KK60) subjected to two different CO₂ levels (400 and 800 $\mu\text{mol mol}^{-1}$). N=6. 81
- 6.6 Bast and core fiber dimensions of three different kenaf varieties (FH991, V36 and KK60) subjected to two different CO₂ levels (400 and 800 $\mu\text{mol mol}^{-1}$) at three months of age. N=3. 83
- 6.7 Bast and core lignocellulose of three different kenaf varieties (FH991, V36 and KK60) subjected to two different CO₂ levels (400 and 800 $\mu\text{mol mol}^{-1}$) at three months of age. N=9. 84
- 6.8 Bast, core and total fiber yield of three different kenaf varieties (FH991, V36 and KK60) subjected to two different CO₂ levels (400 and 800 $\mu\text{mol mol}^{-1}$) at three months of age. N=3. 85

LIST OF FIGURES

Figure	Page
3.1 Means comparison of shoot height of six Kenaf (<i>Hibiscus cannabinus</i> L.) varieties; FH991, FH992, FH952, V36, TK and KK60. Means followed by ($P \leq 0.05$) differ significantly by Duncan at 4 weeks of age. N=3.	28
3.2 Means comparison of stem diameter of six Kenaf (<i>Hibiscus cannabinus</i> L.) varieties; FH991, FH992, FH952, V36, TK and KK60. Means followed by ($P \leq 0.05$) differ significantly by Duncan at 4 weeks of age. N=3.	29
3.3 Means comparison of leaf area meter of six Kenaf (<i>Hibiscus cannabinus</i> L.) varieties; FH991, FH992, FH952, V36, TK and KK60. Means followed by ($P \leq 0.05$) differ significantly by Duncan at 4 weeks of age. N=3.	29
3.4 Means comparison of leaf number of six Kenaf (<i>Hibiscus cannabinus</i> L.) varieties; FH991, FH992, FH952, V36, TK and KK60. Means followed by ($P \leq 0.05$) differ significantly by Duncan at 4 weeks of age. N=3.	30
4.1 Instrument of water potential measurements Scholander pressure chamber (Model 615, Plant Water Status Console, Santa Barbara, CA).	36
4.2 Taken pictures in this study indicating method of measuring bast fiber length and (a) core fiber length (b) taken from high resolution microscope (Magnification = 40x).	38
4.3 Taken pictures in this study indicating method of measuring bast fiber diameter, lumen diameter and cell wall thickness (a) core fiber diameter, lumen diameter and cell wall thickness (b) taken from high resolution microscope (Magnification = 40 x).	39
4.4 Bast fiber yield of Kenaf as affected by interaction of different water levels (25%, 50% and 100% of evapotranspiration replacement (ER) and Three different varieties (Fuhong 991(FH991), V36 and Kohn-Kaen 60(KK60)) (at three months of age). Bars represent standard error of differences between means (SEM). N=3.	50
4.5 Core fiber yield of Kenaf as affected by interaction of different water levels (25%, 50% and 100% of ER) and three different varieties (Fuhong 991(FH991), V36 and Kohn-Kaen 60(KK60)) (at three months of age). Bars represent standard error of differences between means (SEM). N=3.	50
5.1 Bast holocellulose of three different kenaf varieties (Fuhong 991(FH991), V36 and Kohn-Kaen 60 (KK60)). Means with different letters are significantly different ($P \leq 0.05$) at three months of age. N=9.	68

- 5.2 Bast, core and total fiber yield of three different kenaf varieties (Fuhong 991(FH991), V36 and Kohn-Kaen 60(KK60). Means with different letters are significantly different ($P \leq 0.05$) at three months of age. N=9. 70
- 5.3 Bast, core and total fiber yield of three different kenaf varieties (Fuhong 991(FH991), V36 and Kohn-Kaen 60(KK60) affected by different water stress imposition of 1 month after seedling establishment (1MAS), at flowering stage (AFS) and control at three months of age. Bars represent standard error of differences between means (SEM). N=9. 71
- 6.1 Experimental layout using nine mini structures as the main plot of, with 400 = 400 $\mu\text{mol mol}^{-1}$ and 800 = 800 $\mu\text{mol mol}^{-1}$. The subplot consists of three varieties namely Fuhong (FH991), V36 and Kohn-Kaen60 (KK60). Each row represented as a block. 75

LIST OF ABBREVIATIONS

°C	Degree Celsius
°N	North
°S	South
AFS	At Flowering Stage
ANOVA	Analysis of variance
B.C.	Before Christ
CEC	Cation exchange capacity
CGR	Cumulative Growth Rate
Chl	Chlorophyll
Chl F	Chlorophyll fluorescence
cm	Centimetre
CO ₂	Carbon Dioxide
C.V	Coefficient of variation
d.f	Degree of freedom
DNMRT	Duncan New Multiple Range Test
dNTP	2'-Deoxynucleoside 5'- triphosphate
ER	Evapotranspiration Replacement
EDTA	Ethylenediaminetetraacetic acid
FC	Field Capacity
FAO	Food and Agriculture Organization of the United Nations
Fv/fm	Fluorescence variable/ fluorescence maximum
g	Gram
ha	Hectare
hr	Hour
K	Potassium
kPa	Kilo Pascal
m ²	Square meter
M ³	Cubic meter
LRWC	Leaf Relative Water Content
LSD	Least significant difference
LWP	Leaf Water Potential
MAS	Month After Seedling
MPa	Mega Pascal
MS _e	Mean square of error
µg	Microgram
µl	Microlitre
µm	Micrometer
µM	Micromolar
mm	Millimeter
mmol	Millimol
µmol	Micromol
N	Nitrogen
NS	Not Significant
P	Phosphorus
pH	Potential of Hydrogen
ppm	Parts per million
R	Correlation coefficient
RCBD	Randomized complete block design
RWC	Relative Water Content

S	Second(s)
SAS	Statistical Analysis System
S.E.	Standard Error
S.O. V	Source of variance
SPAD	Special Products Analysis Division
St.Dev.	Standard deviation
TBE	Tris-borate/EDTA
UV	Ultraviolet
V	Volt
v/v	Volume per volume
WP	Water Potential
WUE	Water Use Efficiency



CHAPTER 1

INTRODUCTION

Kenaf (*Hibiscus cannabinus* L., Malvaceae) is a warm-season annual crop, with a C₃–photosynthetic pathway, achieving high biomass yields. Although kenaf is a C₃ crop and uses solar radiation, CO₂, water and nitrogen more efficiently than C₄ crops (Danalatos et al., 2005). In the early 1970s, kenaf was introduced to Malaysia and then identified as a potential alternative and cheaper source of fibrous material (Azizi et al., 2010). It has enormous potential to be Malaysia's next industrial crop since it can be used in different types of products with commercial value (Abdul Khalil et al., 2010). The National Kenaf Research and Development Program has been established for research and future development of kenaf-based industry, whereas the government has considered and allocated RM 12 million under the 9th Malaysia Plan (2006-2010) for this purpose (Edeerozey et al., 2007). Kenaf is a demanding crop and is less sensitive to weather and environmental conditions compared to tobacco as kenaf is known to be a tolerant plant to water stress. This has encouraged farmers to replace tobacco with kenaf (Azizi et al., 2010).

Emission of carbon dioxide (CO₂) has become a major concern in many developed and developing countries in order to deal with global warming issues. Therefore, choosing proper strategies to reduce the CO₂ concentration would be achieved by executing carbon capture method. Kenaf as a green source is a great CO₂ sequester which could be great choice to attain this target. Kenaf is an environmental-friendly industrial organic material which Kyoto Protocol has recognized it as a plant with the ability to combat global warming issues. It was reported by Lam et al., (2003) that growing global warming and its environmental concern have led to increasing interest in kenaf as a source of cellulosic fiber for its high CO₂ fixation ability. The use of kenaf as an alternative raw material to wood will protect some forest resources from further deforestation and results in environmental stabilities. Carbon dioxide enrichment entails increasing carbon dioxide concentration to 2 to 4 times higher than the normal atmospheric levels, to between 800-1500 $\mu\text{mol m}^{-2} \text{s}^{-1}$, in an enclosed space (Woodward, 2002). Based on this scientific definition two CO₂ concentration of 400 $\mu\text{mol m}^{-2} \text{s}^{-1}$ as an ambient level and 800 $\mu\text{mol m}^{-2} \text{s}^{-1}$ were introduced in this study to kenaf varieties to evaluate the effect of CO₂ on fiber yield and quality. CO₂ enrichment has been shown to enhance growth rate and plant yields (Tisserat *et al.*, 1997; Wang *et al.*, 2003; Onoda *et al.*, 2007) and the increase of CO₂ concentration in the atmosphere is also well documented (Woodward, 2002). Reported positive effects of enriched CO₂ levels on plant growth encouraged optimism among scientists for future agricultural production (Jaafar, 2006).

As an abiotic stress, water stress is the one of the most serious limitations in agriculture thus; water tolerance performed by any plant is really immensely important (Shao et al., 2008). This type of tolerance involves the changes and differences in biochemistry, physiological responses like leaf gas exchange of plant and plant water relation (Guerfel et al., 2009). Sufficient amount of water should be supplied for plants in order to maintain their cells in good conditions at the early stage of development in order to produce new tissues and cells progressively (Neumann, 2008).

1.1 Justification

The Malaysian pulp and paper industry identified kenaf fiber as a stable source of non-wood materials and a suitable alternative for wood fiber (Roda and Rathi, 2006). However, introduction and determination of specific varieties from local and foreign sources with fiber production is still in its early stages in Malaysia. In Malaysia Development of kenaf industry has both weak and strong noticeable aspects. First of all, Malaysia as a tropical country has the suitable climate in which kenaf cultivation can be very successful. The government and Research Departments of Malaysia are strongly interested in the cultivation of this plant due to prospective market of fiber production in the region. However, lack of enough cultivation knowledge is main setback in developing kenaf industry in Malaysia.

No articles have reported data on the physiology and histochemical properties of kenaf to be grown in top soil (SERDANG series) in Malaysia. Kenaf was reported to produce higher stem production on fertile soils as oppose on BRIS soil. It was concluded after a number of trials of Kenaf cultivation on BRIS that Kenaf is adapted to a wide range of soil types, but performs best on the fine to medium textured clay or loamy, well drained, fertile soils like Serdang series. The Serdang Series as a well-drained soil to over 100 cm depth can be considered. It has good permeability. A large variety of crops can be grown on these soils. These include crops such as oil palm, rubber, fruit trees etc. (Panton, 1954). So it seems valuable to determine the performance of this plant in this type of soil under controlled condition especially for conducting CO₂ enrichment experiment and different water treatments.

The production of fiber may be much affected by the environmental factors which can dwindle as climate scenario changes in future. In order to bring proper insight, examination and determination of physiological and histochemical aspects of Malaysian kenaf cultivars under condition of changing climate (such as different water treatments and CO₂ levels) seem valuable. Crop cultivation modern high technology in controlled environmental system (CES) structures such as glasshouses, greenhouse and rain shelters have many advantages (Jaafar, 2006). The findings were, however, not conclusive and need further research to establish the environmental conditions that would enhance growth and carbon assimilation for increased yield and sustainable production of quality fiber for kenaf under controlled environment system. One of the major driving forces of photosynthesis is CO₂. The greenhouse industry has taken advantage by manipulating this factor for the benefit of crops grown in controlled conditions for extended periods. In order to have proper control and to observe the effect of specific water treatments and different CO₂ concentrations on kenaf growth, fiber yield and quality, experiments had to be done in glasshouse to discard all other undesirable environmental factors.

One of the means to increase different aspects of production is to speed up the growth process. This could be plausible by using carbon dioxide (CO₂) with different levels from seedling stage on. Increase in carbon gain is justified by augment of total plant dry biomass considering enhancement of photosynthesis with extra supplies of carbon dioxide. Despite the enhancement of photosynthesis, growth and production would be

facilitated with reduction of transpiration that results in higher water use efficiency (Hsiao and Jackson, 1999). Plant growth and photosynthesis stimulation have been demonstrated when level of CO₂ is higher than ambient (Coleman and sage, 2001). Despite its great potential for multiple uses (paper, construction materials, and energy), data on assimilation rate as affected by environmental factors such as water stress, time of water stress imposition and CO₂ enrichment effect on kenaf are scarce. Knowledge on kenaf carbon balance, as a response of assimilation and respiration rate to environmental conditions such as CO₂ concentration, is fundamental for understanding and assessing kenaf growth and productivity.

1.2 Hypothesis

As little study has been done to demonstrate that kenaf may continue growth under water stress condition and time of stress imposition, the present study is about understanding the impact of water stress and CO₂ enrichment on growth and fiber development of kenaf. There are possibilities of positive effect of CO₂ enrichment and specific water treatment (as a trigger) on fiber quality of kenaf especially before flowering time. The reduction in transpiration rate in plants under water deficit may also be attributed to morphological changes such as increased cell wall thickness and cell wall lignifications which could bring higher fiber quality (Netondo, 1999).

1.3 General Objective

To determine performance of specific varieties from local and foreign sources with fiber production under some influential environmental factors such as water stress and CO₂ enrichment.

1.4 Specific Objectives

- 1) To evaluate the performance of kenaf varieties on top soil
- 2) To determine the best water treatment and the most effective time of stress imposition for higher fiber yield and quality.
- 3) To examine impact of CO₂ enrichment on kenaf yield, especially on bast and core fiber biophysical properties.
- 4) To identify lignocellulose properties of each selected varieties under water treatments, the time of stress imposition and CO₂ enrichment.
- 5) To determine the most potential productive varieties under CO₂ enrichment.

REFERENCES

- Abbate, P. E., Dardanellib, J. L., Cantarero, M. G., Maturano, M., Melchiorid, R. J. M. and Sueroa, E. E. (2004). Climatic and water availability effects on water use efficiency in wheat. *Crop Science* 44: 474-483.
- Abdul-Hamid, H., Yusoff, M., Ab-Shukor, N., Zainal, B., & Musa, M. (2009). Effects of different fertilizer application level on growth and physiology of *Hibiscus cannabinus* L.(kenaf) planted on BRIS soil. *Journal of Agricultural Science* 1(1): 121-127.
- Abdul-Hamid, M. R. (2008). Kenaf Ganti Tembakau. *Berita Harian*.
- Abdul Khalil, H., Yusra, A., Bhat, A., & Jawaid, M. (2010). Cell wall ultrastructure, anatomy, lignin distribution, and chemical composition of Malaysian cultivated kenaf fiber. *Industrial Crops and Products* 31(1): 113-121.
- Ackerson, R. C. and Kreig, R. D. (1977). Stomatal and non-stomatal regulation of water use in cotton, corn and sorghum. *Plant Physiology* 60: 850-853.
- Ackerson, R.C., Krieg, D.R., Miller, T.D. and Zartman, R.E. (1977). Water relations of field grown cotton and sorghum: Temporal and diurnal changes in leaf water, osmotic, and turgor potentials. *Crop Science* 17:76-80.
- Acreche, M., Gray, L., Collavino, N. and Mariotti, J. (2005). Effect of row spacing and lineal sowing density of kenaf (*Hibiscus cannabinus* L.) yield components in the north-west of Argentina. *Spanish Journal of Agricultural Research* 3(1): 123-129.
- Aini, N, A.S., Hamzah, M.B., Hazandy, A.H., Ghizan, S. and Fadzhel, M.M.N. (2009). Growth and phenology of kenaf (*Hibiscus cannabinus* L.) varieties. *Pertanika Journal of Tropical Agriculture Science*. 32:29-33.
- Akin, D.E., Kimball, B.A., Mauney, J.R., LaMorte, R.L., Hendrey, G.R., Lewin, K., Nagy, J. and Gates, R.N. (1994). Influence of enhanced CO₂ concentration and irrigation on sudangrass digestibility. *Agriculture, Forestry and Meteorology* 70: 279–287.
- Allen, L.H., Valle, R.R. Jr., Mishoe, J.W. and Jones, J.W. (1994). Soybean leaf gas-exchange responses to carbon dioxide and water stress. *Agronomy Journal* 86: 625–636.
- Allen, D.J. and Ort, D.R. (2001) Impacts of chilling temperatures on photosynthesis in warmclimate plants. *Trends in Plant Science* 6: 36-42.
- Alexopoulou, E., Christou, M., Mardikis, M. and Chatziathanassiou, A.(2000). Growth and yields of kenaf varieties in central Crece. *Industrial Crop and Products* 11: 163-172.
- Alexopoulou, E., Cosentino, S.L., Danalatos, N., Venturi, G. and Fernando, A.L. (2005). *Biokenaf: A european chain of the kenaf*. Proceedings of the 14th European Biomass network for the biomass production Conference.

- Aminah, A., Wong, C.C. and Hasim, I. (2004). *Kenaf fiber production as affected by plant population and plant age on bris soil*, p. 9-16. In: Third technical review meeting on the national Kenaf Research project. MARDI.
- Amthor, J.S. and Loomis, R.S. (1996). *Integrating knowledge of crop responses to elevated CO₂ and temperature with mechanistic simulation models: Model components and research needs*, p. 317–346. In: Koch, G.W. and Mooney, H.A. (eds.). Carbon dioxide and terrestrial ecosystems. Academic Press, San Diego, CA.
- Amthor, J.S. (2001). Effects of atmospheric CO₂ concentration on wheat yield: review of results from experiments using various approaches to control CO₂ concentration. *Field Crops Research* 73: 1–34.
- Angelini, L., Macchia, M., Ceccarini, L., & Bonari, E. (1998). Screening of kenaf (*Hibiscus cannabinus* L.) genotypes for low temperature requirements during germination and evaluation of feasibility of seed production in Italy. *Field Crops Research* 59(1): 73-79.
- Anjum, F., Yaseen, M., Rasul, E., Wahid, A. and Anjum, S. (2003). Water stress in barley (*Hordeum vulgare* L.). I. Effect on chemical composition and chlorophyll contents. *Pakistan Journal of Agriculture Science* 40: 45-49.
- Anonymous. (2008a). <http://gaken.com/densidadaceite/condicionpalm.pdf>. Retrieved in 29 September 2008.
- Anonymous.(2008b).<http://www.andhranews.net/Technology/2008/September/23-current-cutbacks-65522.asp>. Retrieved on 6 October 2008.
- Arcelia, M.A.A. and Meribel, L.D.S. (1999). *Fundamentals of plant physiology*. In Plant Physiology society of Philipines: Pasig City, Philipines.
- Arndt, A. K., Clifford, S., Wanek, W., Jones, H. G. and Popp, M. (2001). Physiological and morphological adaptations of the fruit tree *Ziziphus rotundifolia* in response to progressive drought. *Tree Physiology* 21: 1-11.
- Arp, W.J. (1991). Effects of source sink relations on photosynthetic acclimation to elevated CO₂. *Plant Cell Environment* 14: 869 – 875.
- Asch, F., Dingkuhn, M., Sow, A. and Audebert, A. (2005). Drought-induced changes in rooting patterns and assimilate partitioning between root and shoot in upland rice. *Field Crops Research* 93: 223-236.
- Asfaliza, R., Mansur, P., Ghawas, M.M. and Wong, C.C. (2001). *Assessment of kenaf germplasm accessions for their adaptability to the Malaysian environment*. p. 13-24. In: First technical review meeting on the national Kenaf Research project. Malaysian Agriculture Research Institute (MARDI), Serdang Malaysia.
- Ashori, A. (2006). Pulp and paper from kenaf bast fibre. *Fibers Polymer* 7:26-29.
- Association of Official Seed Analysts (AOSA), (1993). Rules for Testing Seeds. *Journal of Seed Technology* 16 (3).

- Ates, S., Ni, Y.H., Akgul, M. and Tozluoglu, A. (2008). Characterization and evaluation of *Paulownia elongata* as a raw material for paper production. *African Journal of Biotechnology* 7: 4153-4158.
- Azizi, A. M., Harun, J., Tahir, P., Resalati, H., Ibrahim, R., Fallah Shamsi, S., & Mohammmed, A. (2010). A review of literatures related of using kenaf for pulp production (beating, fractionation, and recycled fiber). *Modern Applied Science* 4(9): 21-29.
- Baker, J.T., Allen, L.H., Boote, K.J. Jr., Jones, P. and Jones, J.W. (1990). Rice photosynthesis and evapotranspiration in subambient, ambient, and superambient carbon dioxide concentrations. *Agronomy Journal* 82:834–840.
- Baker, J.T. and Allen, L.H. (1994). Assessment of the impact of rising carbon dioxide and other potential climate changes on vegetation. *Environmental Pollution* 83: 223 – 235.
- Banuelos, G.S. Bryla, D.R. and Cook, C.G. (2002). Vegetative production of kenaf and canola under irrigation in central California. *Industrial Crops Products* 15, 237–245.
- Balogun, M. O., Raji, J. A. and Akande, S. R. (2008). Morphological characterization of 51 kenaf (*Hibiscus cannabinus* L.) accessions in Nigeria. *Revista UDO Agrícola* 8(1): 23-28.
- Begg, J.E. and Turner, N.C. (1976). Crop water deficits. *Advanced Agronomy* 28:161-217.
- Bettarini, I., Vacari, F.P. and Miglietta, F. (1998). Elevated CO₂ concentration and stomatal density: observations from 17 plant species growing in a CO₂ spring in central Italy. *Global Change Biology* 4: 17–22.
- Booker, F.L. (2000). Influence of carbon dioxide enrichment, ozone and nitrogen fertilization on cotton (*Gossypium hirsutum* L.) leaf and root composition. *Plant Cell Environment* 23: 573–583.
- Bordovsky, D.G., Jordan, W.R., Hiler, E.A. and Howell, T.A. (1974). Choice of irrigation timing indicator for narrow row cotton. *Agronomy Journal* 66:88-91.
- Bota, J., Flexas, J. and Medrano, H. (2004). Is photosynthesis limited by decreased Rubisco activity and RuBP content under progressive water stress? *New Phytologist* 162: 671-681.
- Bowes, G. (1993). Facing the inevitable: Plants and increasing atmospheric CO₂. *Annu. Rev. Plant Physiol. Plant Molecular Biology* 44:309–332.
- Boyer, J.S. (1985). *Water transport*. Annual Review of Plant Physiology.36: 473-516.
- Bremner, J.M. (1960). Determination of nitrogen in soil by the Kjeldahl method. *Journal of Agriculture Science*. 55: 11-31.
- Brevoort, P. (1998). The blooming United State botanical market: A new overview. *Herbalgram*. 44: 33–46.

- Brown, K and Higginbotham, K.O. (1986). Effects of carbon dioxide enrichment and nitrogen supply on growth of boreal tree seedlings. *Tree physiology* 2: 223 – 232.
- Buwalda, J. G. and Smith, G. S. (1992). Acquisition and utilization of carbon, mineral nutrients and water by the kiwi fruit vines. *Horticulture Review* 13: 307-347.
- Carberry, P.S. and Muchow, R.C. (1992). A simulation model for kenaf assisting fibre industry planning in northern Australia. II. Leaf area Development. *Australian Journal of Agriculture Research* 43:1515–1526.
- Carter, E.B., Theodorou, M.K. and Morris, P. (1999). Responses of *Lotus corniculatus* to environmental change. 2. Effect of elevated CO₂, temperature and drought on tissue digestion in relation to condensed tannin and carbohydrate accumulation. *Journal of Science Food and Agriculture* 79:1431–1440.
- Cattivelli, L., Rizza, F., Bedeck, F. W., Mazzucotelli, E., Mastrangelo, A. M., Francia, E., Mare, C., Tondelli, A. and Stanca, A. M. (2008). Drought tolerance improvement in crop plants: An integrative view from breeding to genomics. *Field Crops Research* 105: 1-14.
- Ceulemans, R. and Mousseau, M. (1994). Effects of elevated atmospheric CO₂ on woody plants. *New Phytologist* 127:425–446.
- Ceulemans, R., Janssens, I.A. and Jach, M.E. (1999). Effects of CO₂ enrichment on trees and forests; lessons to be learned in view of future ecosystem studies. *Annals of Botany* 84: 577–590.
- Chapin, F.S., Bloom, III, A.J. Field, C.B. and Waring, R.H. (1987). Plant responses to multiple environmental factors. *Bioscience* 37:49–55.
- Charles, L.(2002). *Trends in New Crops and New Use*. ASHS Press, Alexandria, VA.
- Cheng, Z., Lu, B., Sameshima, K., Fu, D. and Chen, J. (2004). Identification and genetic relationships of kenaf (*Hibiscus cannabinus* L.) germplasm revealed by AFLP analysis. *Genetic Resources and Crop Evolution* 51(4): 393-401.
- Chia, C., Zakaria, S., Ahamd, S., Abdullah, M. and Jani, S. (2006). Preparation of magnetic paper from kenaf: lumen loading and *in situ* synthesis method. *American Journal of Applied Sciences* 3(3): 1750-1754.
- Coetzee, R. (2004). *Characterization of Kenaf Cultivars in South Africa*. Unpublished doctoral dissertation, University of the Free State, South Africa.
- Coleman, J.R. and Sage, R.F. (2001). Effects of low atmospheric carbon dioxide on plants: more than a thing of the past. *Trends in Plant Science* 6: 18 -24.
- Cornic, G. and Massacci, A. (1996). *Leaf photosynthesis under drought stress* in: Baker, N. R., (Ed.). *Photosynthesis and the Environment*. Kluwer Academic Publishers. The Netherlands.
- Cosentino, S.L. and Copani, V. (2003). Leaf photosynthesis in kenaf (*Hibiscus cannabinus* L.) in response to water stress. *Agroindustria* 2: 137-145.

- Cosentino, S.L., Riggi, E. and D'Agosta, G.(2004). *Leaf photosynthesis in kenaf (Hibiscus cannabinus L.) in response to water stress*. Proceedings of the 2nd World Biomass Conference, Roma, Italy, pp. 374-376.
- Costa, L. D., Vedove, G. D., Gianquinto, G., Giovanardi, R. and Peressoti, A. (1997). Yield, water use efficiency and nitrogen uptake in potato: influence of drought stress. *Potato Research* 40: 19-34.
- Creelman, R. A., Mason, R. J., Benson, J. S., Boyer and Mullet, J. E. (1990). Water deficit and abscisid acid (ABA) cause differential inhibition of shoot versus root growth in soybean seedlings. *Plant Physiologist* 92: 423-430.
- Cundiff, J.S. (1979). Tobacco seed emergence related to differences in terminal velocity. *Tobacco Science* 23:49-51.
- Cure, J.D. and Accock, B. (1986). Crop responses to carbon dioxide doubling: A literature review survey. *Agriculture Forest Metereology* 38: 127 – 145.
- Danalatos, N.G. and Archontoulis, S.V.(2004). *Potential growth and biomass productivity of kenaf under central Greek conditions: II. The influence of variety, sowing time and plant density*. Proceedings of the 2nd World Biomass Conference, , Roma, Italy, pp. 319-322.
- Danalatos, N.G. and Archontoulis, S.V.(2005). *Sowing time and plant density effects on growth and biomass productivity of two kenaf varieties in central Greece*. Proceedings of the International Conference on Industrial Crops and Rural Development, Murcia, Spain, pp. 889-901.
- Daud, M.D.M. (2006) . *Development of kenaf production in Malaysia*. p. 119-130. In: Fourth technical review meeting on the National Kenaf Research project. MARDI.
- Davidson, E. A., Verchot, L. V., Cattanio, J. H., Ackerman, I. L. and Carvalho, H. M. (2000). Effects of soil water content on soil respiration in forests and cattle pastures of eastern Amazonian, *Biogeochemistry* 48: 53-69.
- Davis, T.D. and Potter, J.R. (1983). High CO₂ applied to cuttings:effects on rooting and subsequent growth in ornamental species. *HortScience* 18: 194-196.
- Dehghani-Firouzabadi, M. R., Shakhes, J. and Namakiyan, R. (2008). *The effects of harvesting times on fiber morphological properties and kenaf yield*. The 1st Iranian Conference on supplying Raw materials and Development of Wood and Paper Industries, Gorgan University of Natural Resources and Agriculture.
- Dempsey, J.M.(1975). *Fiber Crops*. Rose Printing Company, Tallahassee, FL.
- Drake, B.G. and Leadley, P.W. (1989). Canopy photosynthesis of C₃ and C₄ communities exposed to long term elevated carbon dioxide treatment. *Plant Cell and Environment* 14: 853 – 860.
- Edeerozey, A., Akil, Hk., Azhar, A., and Ariffin, M. (2007). Chemical modification of kenaf fibers. *Materials Letters* 61(10): 2023-2025.

- Eamus, D. (1991). The interaction between rising carbon dioxide and temperature with water use efficiency. *Plant, Cell and Environment* 14: 843 - 852.
- Encyclopedia Britannica. (2006). Carbon dioxide [Electronic version]. Retrieved 23 February 2006.
- Enoch, H.Z. and Honour, H.J. (1993). *Significance of increasing carbon dioxide for plant growth and survival, and interaction with air pollution*. In *Interacting Stresses on Plant in a Changing Climate*, ed. Jackson, M.B. and Black, C.R., pp. 51 -75. Berlin: Springer Verlag.
- Evans, J.R. (1999). Leaf anatomy enables more equal access to light and CO₂ between chloroplasts. *New Phytologist* 143: 93–104.
- FAO. (2012). *FAO Production Year Book* 3. New York: FAO.
- Farooq, M., Wahid, A., Kobayashi, N., Fujita, D., Basra, S. M. A. (2009). Plant drought stress: effects, mechanisms and management. *Agronomy Sustainable Development*. 1-28.
- Farquahar, G.D., and Von, C. (1982). *modelling of photosynthetic response to environmental condition* . In. *Encyclopedia of plant physiology* vol 12B. Edited by, Lange, O.L., Nobel, P.S., Osmond, C.B., Ziegler, H. Springer-verlag, Heidelberg. Pp 549 – 587.
- Fernandez-Conde, M. E. (1998). *Effects of drought on growth and photosynthetic capacity of cotton (Gossypium hirsutum L.)*. Paper Presented at the Internet World Congress. Dec, 7-16. Hamilton. Ontario, Canada.
- Fischer, R.A. and Turner, N.C. (1978). Plant Productivity in the Arid and Semiarid Zones. *Annual Review of Plant Physiology* 29: 277-317.
- Franck, R. (2005). *Bast and Other Plant Fibres*. Cambridge: Woodhead Publishing.
- Francois, L.E., Donovan, T.J. and Maas, E.V. (1992). Yield, vegetative growth, and fibre length of kenaf grown on saline soil. *Agronomy Journal* 84: 592–598.
- Frehner, M., Lu'scher, A., Hebeisen, T., Zanetti, S., Schubiger, F. and Scalet, M. (1997). Effects of elevated partial pressure of carbon dioxide and season of the year on forage quality and cyanide concentration of *Trifolium repens* L. from a FACE experiment. *Acta Oecology* 18: 297–304.
- Fritschi, F.B., Boote, K.J., Sollenberger, L.E. and Allen Jr., L.H. (1999). Carbon dioxide and temperature effects on forage establishment: tissue composition and nutritive value. *Global Change Biology* 5: 743–753.
- Fu, J. and Huang, B. (2001). Involvement of antioxidants and lipid peroxidation in the adaptation of two cool-season grasses to localized drought stress. *Environment Experimental Botany* 45: 105-114.

- Goford. (1994). *The evaluation of kenaf as an oil absorbent*. In Fuller, M. J. (ed) A summary of kenaf production and product development research 1989-1993. Miss. Agri. And Forestry Exp. Sta., Mississippi State, MS Bulletin 1011: 25.
- Gonzalez, M., Ribascabo, M.A. and Siedow, J.N. (1996). Direct inhibition of plant mitochondrial respiration by elevated carbon dioxide. *Plant physiology* 112: 1349 – 1355.
- Guerfel, Baccouri, M.O., Boujnah. D., Chaibi, W. and Zarrouk, M. (2009). Impacts of water stress on gas exchange, water relations, chlorophyll content and leaf structure in the two main Tunisian olive (*Olea europaea* L.) cultivars. *Scientific Horticulture* 119(3): 257-263.
- Hamid, A., Yusoff, M., A. Shukor, N., Zainal, B., & Musa, M. (2009). Effects of different fertilizer application level on growth and physiology of *Hibiscus cannabinus* L. planted on BRIS soil. *Journal of Agriculture Science* 1(1): 121-131.
- Hamilton, J.G., Thomas, R.B. and Delucia, E.H. (2001). Direct and indirect effects of elevated carbon dioxide on leaf respiration in a forest ecosystem. *Plant Cell and environment* 24: 975 – 982.
- Haniff, M.H. (2006). Gas exchange of excised oil palm (*Elaeis guineensis*) fronds. *Asian Journal of Plant Sciences* 5(1): 9 – 13.
- Hawa, Z.E. (2005). *Carbon dioxide enrichment production technology for controlled environment system in the lowland tropics*. In International symposium on greenhouses, environmental controls and in-house mechanization for crop production in the tropics and sub-tropics: 15 -17 June 2004.
- Henson, I.E., Jensen, C.R. and Turner, N.C. (1989). Leaf gas exchange and water relation of Lupins and Wheat. 1. Shoot responses to soil water deficits. *Australian Journal of Plant Physiology* 16: 401–413.
- Hicklenton, P.R. (1988). *Carbon dioxide enrichment in the greenhouse: principle and practices*. Portland , Timber Press: USA.
- Higginbotham, K.O., Mayo, J.M., Lhironnelle, S.L. and Krystofiak, D.K. (1985). Physiological ecology of lodgepine (*pinus contorta*) in an enriched CO₂ environment. *Canada Journal forest Research* 15: 417- 421.
- Hiler, E.A. and Clark, R.N. (1971). Stress day index to characterize effects of water stress on crop yields. *Trans ASAE* 14:757-761.
- Hiler, E.A., Van Bavel, C.H.M., Hossain, M.M. and Jordan, W.R. (1972). Sensitivity of southern peas to plant water deficits at three growth stages. *Agronomy Journal* 64:60-64.
- Hiroimi, H., Ninomiya, I., Koike, T. and Ogino, K. (1999). Stomatal regulation of canopy trees in a tropical rain forest. *Japanease Journal of Ecology* 49: 68-76.

- Hng, P.S., Khor, B.N., Tadashi, N., Aini, A.S.N. and Paridah, M.T. (2009). Anatomical Structures and Fiber Morphology of New Kenaf Varieties. *Asian Journal of science research*. 3:161-166.
- Hollowell, J. (1997). *Nutritional and Yield Evaluation of Kenaf as a Potential High Quality Forage for the Southeastern United States*. Unpublished doctoral dissertation, Mississippi State University, Mississippi.
- Horn, R. and Setterholm, V. (1990). Fiber morphology and new crops. In J. Janick & J. Simon. *Advances in New Crops*. pp. 270-275. Portland: Timber Press.
- Hosomi, K. (2000). *Utilization of dried kenaf-leaves to the meals*. Proceedings of the 2000 international kenaf symposium, Hiroshima, Japan. 13-14, pp.171-176.
- Hossain, M.D., Hanafi, M.M., Jol, H. and Hazandy, A.H. (2011). Growth, yield and fiber morphology of kenaf (*Hibiscus cannabinus* L.) grown on sandy bris soil as influenced by different levels of carbon. *Asian Journal of Botrony* 10(50): 10087-10094.
- Hsiao, T. C. (1973). Plant responses to stress. *Annual Review of Plant Physiology* 24: 519-570.
- Hsiao, T.C. and Jackson, R.B. (1999). *Interactive effects of water stress and elevated carbon dioxide on growth, photosynthesis and water use efficiency*. In *Carbon dioxide and environmental stress*, ed by Yiqi, L. and Mooney, H.A. pp 3 - 26. Academic press.
- Huang, B. R. and Fu, J. (2000). Photosynthesis, respiration and carbon allocation of two cool-season perennial grasses in response to surface soil drying. *Plant Soil* 227: 17-26.
- Ibrahim, M.H., Jaafar, H.Z.E., Haniff, M.H., Raffi, M.Y. (2010). Changes in growth and photosynthetic patterns of oil palm seedling exposed to short term CO₂ enrichment in a closed top chamber. *Acta Physiology Plant* 32: 305–313.
- Idem, N.U.A. and Adeoti, A.A. (1988). *Kenaf: A potential pulp source for Nigeria*. A paper presented at the 2nd Annual Conference of the Botanical Association of Nigeria, 10-14th April, 1988 at ABU Zaria.
- Idris, A.B.G.K., Chin, F.Y. and Wong, C.C. (2001). *Preliminary studies on kenaf sseed production at sintok farm, kedah. Proceeding*. p. 6-10 .In: First technical review meeting on the National Kenaf Research project. MARDI.
- Idso, S.B., Kimball, B.A., Anderson, M.G. and Maugney, J.R. (1987). Effects of atmospheric CO₂ enrichment on plant growth : The interactive role of air temperature. *Agriculture Ecosystem Environment* 20: 1 -10.
- Idso, K.E. and Idso, S.B. (1994). Plant responses to atmospheric CO₂ enrichment in the face of environmental constraints: a review of the past 10 years' research. *Agricultural and Forest Meteorology* 69: 153–203.

- Iker, A., Pilar, P., Libia, H., Juan, J.I., Zita, G., Rafael, M.C. and Manuel, S.D. (2005). The response of nodulated alfalfa to water supply, temperature and elevated carbon dioxide: photosynthesis downregulation. *Physiologica Plantarum* 123: 348 – 358.
- IPCC (2011). *Intergovernmental Panel of Climatic Change WGII*, fourth assessment report,. Available from <http://www.ipcc.ch/publications> and data/publications and data reports.htm
- Ismail, M. R. and Awang, M. (1992). Growth and physiological changes in of *Averrhoa carambola* as influenced by water availability. *PERTANIKA* 15 (1): 1-7.
- Ismail, M. R., Burrage, S. W., Tarmizi, H., Aziz, M. A. (1994). Growth, plant water relations, photosynthesis rate and accumulation of proline in young carambola plants in relation to water stress. *Scientia Horticulture* 60: 101-114.
- Jackson, R.B., Sala, O.E., Field, C.B. and Mooney H.A. (1994). Carbon dioxide alters water use, carbon gain, and yield of dominant species in a natural grassland. *Oecologia* 98: 257 – 262.
- Jaafar, H.Z.E. (1995). *Impact of Environmental Stress on Reproductive Development in Sweet pepper (Capsicum anuum)*. Ph.D. Thesis, University of Nottingham, Nottingham, UK.
- Jaafar, H.Z.E. (2006). Carbon dioxide enrichment technology for improved productivity under controlled environment systems in tropics. *Acta Horticulture 2*: 353–363.
- Jeffrey, Q.C and Whendee, L.S. (2006). *Some aspect of ecophysiological and biogeochemical responses of tropical forest to atmospheric changes*. Philos. Trans. R. Soc. Lond. B. 2004; 359:549–555.
- Jensen, C.R. (1989). *Plant water relations approaches and measurements*. in: Persson, R., Wredin, A., (Eds.). Vattminsbehov och Narings-tillforsel. Foredrag presenterade vid NJF-seminarium nr. 151 Landskrona, 1–3. 45–63.
- Jones, M.D., Puentes, C. and Suarze, R.(1955). Isolation of kenaf for seed increase. *Agronomy Journal*. 47: 256-257.
- Jones, P., Allen, L.H., Jones, J.W. Jr., Boote, K.J. and Campbell, W.J. (1984). Soybean canopy growth, photosynthesis, and transpiration responses to whole-season carbon dioxide enrichment. *Agronomy Journal* 76:633–637.
- Jones, P., Allen, L.H., Jones, J.W. Jr. and Valle, R. (1995). Photosynthesis and transpiration responses of soybean canopies to short- and long-term CO₂ treatments. *Agronomy Journal* 77:119–126.
- Jones, R.J. and Mansfield, T.A. (1970). Increases in the diffusion resistances of leaves in a carbon dioxide-enriched atmosphere. *J. Expt. Bot.* 21: 951–958.
- Kaldor, A., Karlgren, C., & Verwest, H. (1990). Kenaf: a fast growing fiber source for papermaking. *TAPPI Journal (USA)*. 73(11): 205-208.

- Kalina, J., Kajanek, M., Spunda, V and Marek, M.V. (1997). *Changes of the primary photosynthetic reactions of Norway spruce under elevated carbon dioxide*. In Impacts of global change on Tree Physiology and Forest Ecosystem, ed. Mohrem, G.M.J, Kramer, K and Sabate, S. pp 59 – 69. Kluwer Academic Publication: Dordrecht.
- Kim, J. Y., Mahe, A., Brangeon, J. and Prioul, J. L. (2000). A maize vacuolar invertase, IVR2 is induced by water stress. Organ/tissue specificity and diurnal modulation of expression. *Plant Physiology* 124: 71-84.
- Kimball, B.A. (1983). Carbon dioxide and agricultural yield: An assemblage and analysis of 430 prior observations. *Agronomy Journal* 75:779-788.
- Kimball, B.A. and Idso, S.B. (1983). Increasing atmospheric CO₂: Effects on crop yield, water use, and climate. *Agriculture Water Management* 7:55–72.
- Kimball, B.A. and Mauney, J.R. (1993). Response of cotton to varying CO₂, irrigation, and nitrogen: yield and growth. *Agronomy Journal* 85: 706–712
- Kimball, B.A., Kobayashi, K. and Bindi, M. (2002). Responses of agricultural crops to free-air CO₂ enrichment. *Advanced Agronomy* 77: 293–368.
- Klapwijk, D. and De Lint, P.J.A.L. (1974). Fresh weight and flowering of tomato plants as influenced by container type and watering condition. *Acta Horticulture* 39:237–247.
- Komor, E. (2000). Source physiology and assimilate transport: the interaction of sucrose metabolism, starch storage and phloem export in source leaves and the effects on sugar status in phloem. *Australian Journal of Plant Physiology* 27: 497-505.
- Kramer, P.J. and Kozlowski, T.T. (1979). *Physiology of woody plants*. Academic press: New York.
- Kramer, P.J. (1981). Carbon dioxide concentration, photosynthesis and dry matter production. *Bioscience* 31: 1 – 33.
- Kramer, P.J. (1983). *Water Relations of Plants*. Academic Press, New York.
- Kristova, V., Kriska, M., Canova, R., Hejdova, E., Kobzova, D. and Dobrocky, P. (1993). Endothelial changes following repeated effect of vasoconstrictive substances in vitro. *Acta Physiology Hungarian* 81: 363-370.
- Krupa, S.V. and Kickert, P.N. (1989). The greenhouse effects: impact of ultraviolet-B-(UV-B) radiation, carbon dioxide (CO₂) and ozone (O₃) on vegetation. *Environmental Pollution* 61: 263 – 393.
- Kubota, F. and Hamid, A. (1992). Comparative analysis of dry matter production and photosynthesis between mungbean (*Vigna radiata* L. Wlczek) and blackgram (*Vigna mungo* L. Hepper) grown in different light intensities. *J. Fac. Agric. Kyushu Univ.* 37: 71-80.
- Kulger (1996). *Kenaf commercialization*. 1986-1995. In Janick, J. (ed) Progress in new crops. ASHS Press, Arlington, VA. 129-132.

- Lam, T. B. T., Hori, K. and Iiyama, K. (2003). Structural characteristics of cell walls of kenaf (*Hibiscus cannabinus* L.) and fixation of carbon dioxide. *Journal of Wood Science*. 49.
- Lambers, H., Atkin, O. K. and Scheureater, I. (1996). *Respiratory patterns in roots in relation to their function*, in: Waisel Y. (Ed). *Plants Roots, The Hidden Half*. Marcel Dekker, New York.
- Lawlor, D.W. and Mitchell, R.A.C. (2000). *Crop ecosystem responses to climatic change: wheat*. In: Reddy, K.R. and Hodges, H.F. (Eds.), *Climate Change and Global Crop Productivity*. CAB International, Wallingford, pp. 57–80
- Leport, L., Turner, N. C., French, R. J., Barr, M. D., Dude, R. and Davies, S. L. (2006). Physiological responses of chickpea genotypes to terminal drought in a Mediterranean-type environment. *European Journal of Agronomy* 11: 279-291.
- Leuning, R. and Foster, L.J. (1991). Estimation of transpiration by single trees: Comparison of a ventilated chamber, leaf energy budgets and a combination equation. *Agriculture and Forest Metereology* 51: 63 – 68.
- Levitt, T. (1980). *Response of plant environmental stress to water, radiation, salt and other stresses*. *Physiol. Ecol.* 2nd Edn. Acad. Press Inc. Orlando, Florida USA. 365-488
- Li, P., Cheng, L., Peng, T. and Gao, H. (2009). CO₂ assimilation and chlorophyll fluorescence in green versus red *Brberis thunbergii* leaves measured with different quality irradiation. *Photosynthetica* 47:11-18.
- Lindhout, P. and Pet, G. (1990). Effects of CO₂ enrichment on young Plant growth of 96 genotypes of tomato (*Lycopersicon esculentum*). *Euphytica* 51(2): 191-196.
- Lippert, M., Steiner, K., Pfirman, T. and Payer, K. (1997). Assessing the impact of elevated ozone and carbon dioxide on gas exchange characteristics of differently supplied clonal Norway spruce trees during exposure and the following season. *Trees* 11: 306 – 316.
- Liu, A. M. (2003). *Making pulp and paper from kenaf*, Proceedings of the Natural Resource and Environment, Rome.
- Liu, H. S. and Li, F. M. (2005). Root respiration, photosynthesis and grain yield of two spring wheat in response to soil drying. *Plant Growth Regulators* 46: 233-240.
- Liu, Y. (2005). *Diallel and Stability Analysis of Kenaf (Hibiscus cannabinus L.) in South Africa*. Unpublished doctoral dissertation, University of the Free State, South Africa.
- Loats, K.V. and Rabbeck.J. (1999). Interactive effects of ozone and elevated carbon dioxide on the growth and physiology of black cherry, green ash and yellow poplar seedlings. *Environmental pollution* 106: 237 – 248.
- Loreto, F., Tricoli, D. and Di Marco, G. (1995). On the relationship between electron transport rate and photosynthesis in leaves of the C₄ plant Sorghum bicolor exposed

- to water stress, temperature changes and carbon metabolism inhibition. *Australian Journal of Plant Physiology* 22: 885-892.
- Ludlow, M. M. and Muchow, R. C. (1990). A critical evaluation of traits for improving crop yields in water-limited environments. *Advanced Agronomy* 43: 107-153.
- Luvaha, E., Netondo, G. W. and Ouma, G. (2008). Effect of water deficit on the physiological and morphological characteristics of mango (*Mangifera indica*) rootstock seedlings. *American Journal of Plant Physiology* 3(1): 1-15.
- Makino, A., Nakano, H. and Mae, T. (1994). Responses of ribulose-1,5-bisphosphate carboxylase, cytochrome f, and sucrose synthesis enzymes to leaf nitrogen in rice, and their relationships to photosynthesis. *Plant Physiology* 105: 1231-1238.
- Malaysian Industry-Government Group for High Technology (MIGHT). (2011). *Consultancy Services to Carry Out Business Strategy, Marketing and Implementation Plan for the Development of the Kenaf Industry in the East Coast Economic Region (ECER)*. East Coast Economic Region Development Corporation (ECERDC). Kuala Lumpur.
- Mambelli, S. and Grandi, S. (1995). Yield and quality of kenaf (*Hibiscus cannabinus* L.) stem as affected by harvest date and irrigation. *Industrial Crops Products* 4 (2): 97-104.
- Manikavelu, A., Nadarajan, N., Ganesh, S. K., Gnanamalar, R. P. and Babu, R. C. (2006). Drought tolerance in rice, morphological and molecular genetic consideration. *Plant Growth Regulators* 50: 121-138.
- Mansfield, T. J. and Atkinson, C. J. (1990). *Stomatal behavior in water stressed plants*, in: Alscher, R. G., Cumming, J. R. (Eds.). *Stress Responses in Plants: Adaptation and Acclimation Mechanisms*. Wiley-Liss. New York. 241-264.
- Mark, S.J. and Jackson, S.B. (2000). Growth responses of *Quercus petraea*, *Fraxinus excelsior* and *Pinus sylvestris* to elevated carbon dioxide, ozone and water supply. *New Phytologist* 146: 437-451.
- McCree, K. J. (1985). Whole plant carbon balance during osmotic adjustment to drought and salinity stress. *Australian Journal of Plant Physiology* 13: 33-43.
- McDonald, M.B. (2000). *Seed viability, germination, and vigor: Sorting out the terminology*. in: VanderVelde, J. (Ed.) *GrowerTalks on Plugs 3*. Ball Publ., Batavia, IL. 29-31
- McMillin, J.D., Wanger, M.R., Webber III, C.L., Mann, S.S., Nichols, J.D. and Jech, L. (1998). Potential for kenaf cultivation in south-central Arizona. *Industrial Crops Products* 9:73-77.
- Mehlich, A. (1953). Determination of *P, Ca, Mg, K, Na, and NH₄*. *Soil Test Division* (Mimeo). Raleigh, North Carolina, USA.
- Monakhova, O. F. and Chernyadev, I. I. (2002). Protective role of kartolin-4 in wheat plants exposed to soil drought. *Applied Biochemistry Microbiology* 38: 373-380.

- Monclus, R., Dreyer, E., Villar, M., Delmotte, F. M., Delay, D., Petit, J. M., Barbaroux, C., Thiec, D. L., Brechet, C. and Brignolas, F. (2006). Impact of drought on productivity and water use efficiency in 29 genotypes of *Populus deltoids* x *Populus nigra*. *New Phytologist* 169: 765-777.
- Morgan, P. W. (1990). *Effects of abiotic stresses on plant hormone systems*, in: Stress Responses in plants: adaptation and acclimation mechanisms. Wiley-Liss. Inc. 113-146.
- Morison, J.I.L. (1985). Sensitivity of stomata and water use efficiency to high CO₂. *Plant Cell environment* 14: 467 – 474.
- Morison, J.I.L. (1987). *Intercellular carbon dioxide concentration and stomatal responses to carbon dioxide*. In Stomatal Function, ed. Zeiger, E., Farquhar, G.D. and Cowan, I.R., pp. 229 - 251. Stanford, California: Stanford University Press.
- Morrison, W. H., Akin, D. E., Archibald, D. D., Dodd, R. B. and Raymer, P. L. (1999). Chemical and instrumental characterization of maturing knead core and bast. *Industrial Crops Products* 10: 21-34.
- Morison, J.I.L. (1993). Response of plant to carbon dioxide under water limited condition. *Vegetatio* 105: 193 – 209.
- Morocco, J. P., Pereira, J. S. and Chaves, M. M. (1997). Stomatal responses to leaf-to-air vapour pressure deficit in Sahelian species. *Australian Journal of Plant Physiology* 24: 381-387.
- Mortensen, L. M. (1987). Review: CO₂ enrichment in greenhouses. Crop responses. *Scientific Horticulture* 33:1–25.
- Mostofa, M. G., Islam, M. R., Alam, A. T. M., Ali, S. M. M. and Mollah, M. A. F. (2002). Genetic variability, heritability and correlation studies in kenaf (*Hibiscus cannabinus* L.). *Online Journal of Biological Sciences* 2(6): 422-424.
- Muchow, R.C. and Wood, I.M. (1980). Yield and growth responses of Kenaf (*Hibiscus cannabinus* L.) in a semi-arid tropical environment to irrigation regimes based on leaf water potential. *Irrigation Science* 1: 209-222.
- Nelson, P.V. (1985). *Greenhouse operation and management*. third edition. Prentice Hall: London.
- Netondo, G. W. (1999). *The use of physiological parameters in screening for salt tolerance in sorghum varieties grown in Kenya*. Ph.D. Thesis. Moi University, Kenya.
- Neumann, P. M. (2008). Coping mechanisms for crop plants in drought-prone environments. *Annals of botany* 101: 901-907.
- Nijs, I., Impens, I. and Behaeghe, T. (1988). Effects of long – term atmospheric carbon dioxide concentration on *Lolium Perrene* and *Trifolium repens* canopies in the course of terminal drought stress period, *Canadian Journal of Botany* 67: 2720 – 2725.

- Nieschlag, H.J., Nelson, G.H., Wolff, J.A. and Perdue, R.E. (1960). A search for new fiber crops. *Tappi* 43 (3): 193.
- N'kaa, F.A., Ogbonnaya, C.I. and Onyike, N.B. (2007). Effect of differential irrigation on physical and histochemical properties of kenaf (*Hibiscus cannabinus* L.) grown in the field in Eastern Nigeria. *Africani Journal of Agriculture Resrarch* Vol. 2 (6): 252-260.
- Norby, R.J. and O'neill, E.J. (1991). Leaf area compensation and nutrient interactions in carbon dioxide enriched seedlings of yellow-poplar. *New phytologist* 117: 515 – 528.
- Norby, R.J., Wullschleger, S.D. and Gunderson, C.A. (1996). *Tree responses to elevated CO₂ and the implications for forests*. In: Koch GW, Mooney HA, eds. Carbon dioxide and terrestrial ecosystems. San Diego, CA, USA: Academic Press, 1–21.
- Ogbonnaya, C.I., Nwalozie, M.C., Roy-Macauley, H. and Annerose, D.J.M. (1998). Growth and water relations of Kenaf (*Hibiscus cannabinus* L.) under water deficit on a sandy soil. *Industrial Crops Products* 8: 65–76.
- Okereke, O.O. (1962). *Studies on the fiber dimensions of some Nigerian timbers and raw materials*. Part 1: Research Report No. 16: Fed. Ministry of Commerce and Industry Lagos Nigeria.
- Ohyama, K, Kozai,T, Ishigami, Y. and Ochi, Y. (2000). *A carbon dioxide enrichment system for a greenhouse with a high ventilation rate*. Tottori University press: Japan.
- Omami EN (2005). *Salt tolerance of Amaranth as affected by seed priming*. Ph.D. Thesis Univ. of Pretoria etd.
- Onoda, Y., Hirose, T. and Hikosaka, H. (2007). Effect of elevated CO₂ levels on leaf starch, nitrogen and photosynthesis of plants growing at three natural CO₂ springs in Japan. *Ecological Research* 22: 475–484.
- Owensby, C.E., Cochran, R.M. and Auen, L.M. (1996). *Effects of elevated carbon dioxide on forage quality for ruminants*. In: Koerner, C. and Bazzaz, F. (Eds.), Carbon Dioxide, Populations and Communities. Physiological Ecology Series. Academic Press, pp. 363–371
- Ouma, J. P. (1988). *The effect of soil moisture deficit on photosynthesis and leaf water potential of field grown beans*. M. Sc. Thesis. University of Nairobi, Kenya.
- Panton, W. P. (1954). Soil Survey Report No. 1. Federal Experimental Station, Serdang. *Malaysian Agriculture Journal* 37:136-145.
- Parkhurst, J. (1986). *Internal leaf structure: a three dimensional perspective*. In: Givnish TJ, ed. On the economy of plant form and function. Cambridge, UK: Cambridge University Press, 215–250.
- Patané, C., D'Agosta, G.M., Mantineo, M. and Cosentino, S.L. (2007). *Radiation interception and use by kenaf (Hibiscus cannabinus L.) canopy under different water and nitrogen supply*. In: Proceeding of 15th European Biomass Conference, Berlin, Germany, pp. 791–794.

- Pate, J. and Joyner, J. (1958). The inheritance of a male sterility factor in kenaf, *Hibiscus cannabinus* L. *Agronomy Journal* 50: 402-403.
- Pearce, R.B., Carlson, G.E., Barnes, D.K., Hart, R.H. and Hanson, C.H. (1969). Specific leaf weight and photosynthesis in alfalfa. *Crop Science* 7-8 (9): 423-426.
- Peñuelas, J. and Matamala, R. (1990). Changes in N and S leaf content, stomatal density and specific leaf area of 14 plant species during the last three centuries of CO₂ increase. *Journal of Experimental Botany* 41: 1119–1124.
- Petri, R. (1952). Pulping studies with African tropical woods. *TAPPI* 35: 157-160.
- Peterson, R.B. and Havir, E.A. (2003). Contrasting modes of regulation of PSII light utilization with changing irradiance in normal and mutant leaves psbS of *Arabidopsis thaliana*. *Photosynthesis Research* 75: 57 -70.
- Phuong, M.T., Araki, T. and Kubota, F. (2005). Comparison of Growth Feature and Drought Tolerance between two High Productive Species, Kenaf (*Hibiscus cannabinus*, C3-plant) and Napiergrass (*Pennisetum purpureum*, C4-plant). *J. Fac. Agr., Kyushu Univ.* 50 (2): 521-532.
- Poole, I., Weyers, J.D.B., Lawson, T. and Raven, J.A. (1996). Variations in stomatal density and index: implications for palaeoclimatic reconstructions. *Plant, Cell & Environment* 19: 705–712.
- Poorter, H. Gifford, R.G., Kridelman, P.E. and Wong, S.E. (1992). A quantitative analysis of dark respiration and carbon content as factors in the growth response of plants to elevated CO₂. *Australian Journal Botany* 40: 501 – 513.
- Porter, M.A. and Grodzinski, B. (1985). CO₂ enrichment of protected crops. *Horticulture Research* 7:345–398.
- Prior, S.A., Torbert, H.A. Runion, G.B. and Rogers. H.H. (2003). Implications of elevated CO₂- induced changes in agroecosystem productivity. *Journal of Crop Products* 8:217–244.
- Radoglou, K.M. and Jarvis, P.G. (1990). Effects of CO₂ enrichment on four poplar clones. I. Growth and leaf anatomy. *Annals of Botany* 65: 617–626.
- Reddy, V.S., Safadi, F., Zielinski, R.E. and Reddy, A.S.N. (1999). Interaction of a kinesin-like protein with calmodulin isoforms from *Arabidopsis*. *Journal of Biology and Chemistry* 274: 31727–31733.
- Reddy, A. R., Chaitanya, K. V. and Vivekanandan, M. (2004). Drought-induced responses of photosynthesis and antioxidant metabolism in higher plants. *Journal of Plant Physiology* 161: 1189-1202.
- Roda, J. and Rathi, S. (2006). Feeding China's expanding demand for wood pulp: a diagnostic assessment of plantation development, fiber supply, and impacts on natural forests in China and South East Asia region: Vietnam report. *Jakarta, Indonesia: CIFOR.* 17: 979-924.

- Roden, J.S. and Ball, M.C. (1996). The effect of elevated carbon dioxide on growth and photosynthesis of two eucalyptus species exposed to high temperature and water deficit. *Plant Physiology* 111: 909 – 919.
- Roderick, M.L, Berry, S.L. and Noble, I.R. (1999). The relationship between leaf composition and morphology at elevated CO₂ concentrations. *New Phytologist* 143: 63–72.
- Rogers, H.H. and Dahlman, R.C. (1993). Crop responses to CO₂ enrichment. *Vegetatio* 105:117–131.
- Rogers, H.H., Prior, S.A. Runion, G.B. and Mitchell, R.J. (1996). Root to shoot ratio of crops as influenced by CO₂. *Plant Soil* 187:229–248.
- Rydholm, S.A. (1967). *Pulping process*. New York, Wiley and Sons.(ed) Sellers, T., Miller, G.D. and Fuller, M.J. (1993). Kenaf core as a board raw material. *Forage Producys Journal* 43: 69-71.
- Saikia, C., Goswami, T., & Ali, F. (1997). Evaluation of pulp and paper making characteristics of certain fast growing plants. *Wood Science and Technology* 31(6): 467-475.
- Samarah, N. H. (2005). Effects of drought stress on growth and yield of barley. *Agronomy Sustainable Development* 25: 145-149.
- Samarakoon, A.B., Muller, W.J. and Gifford, R.M. (1995). Transpiration and leaf area under elevated carbon dioxide: Effects of soil water status and genotype in wheat. *Australian Journal of Plant Physiology* 22: 33 – 44.
- Sameshima, K.(2000). *Improvement of kenaf core oil absorption property by heat treatment at 200-500°C*. Proceedings of 3rd annual America kenaf society conference, Corpus Christi, TX, pp. 64-72.
- Sanders, G.E., Clark, A.G., and Colls, J.J. (1991). The influence of open top chambers on the growth and development of field bean. *New Phytologist* 117: 439 – 447.
- Saxe, H., Ellsworth, D.S. and Heath, J. (1998). Tree and forest functioning in an enriched CO₂ atmosphere. *New Phytologist* 139: 395–436.
- SAS (2007). *SAS/STAT User's Guide, Version 9.2*. SAS Institute Inc., Cary, NC, USA.
- Scott, S.W., Cook, C.G. and Taylor, C.S. (1999). *The effect of harvest timing on kenaf and sunn hemp*. p. 37–40 In: Proceedings of the Second Annual American Kenaf Society Meeting, 25–26 February 1999. San Antonio, TX, USA.
- Seth, R. S. and Page, D. H. (1988). Fiber properties and tearing resistance. *TAPPI Journal*. 71(2): 103-107.
- Setter, T. L., Flannigan, B. A. and Melkonian, J. (2001). Loss of kernel set due to water deficit and shade in maize: carbohydrate supplies, abscisic acid and cytokinins. *Crop Science* 41: 1530-1540.

- Shamshuddin, J. and Anda, M. (2008). Charge properties of soils in Malaysia dominated by kaolinite, gibbsite, goethite and hematite. *Bulletin of the Geological Society of Malaysia* 54: 27-31.
- Shao, H., Chu, L., Shao, M., Jaleel, C.A. and Mi. H. (2008). Higher plant antioxidants and redox signalling under environmental stresses. *Crop Resrearch. Biologies* 331: 433–441.
- Shaoyun, L., Chuanhao, C., Zhongcheng, W., Zhenfei, G. and Haihang, L. (2009). Physiological responses of somaclonal variants of triploid bermudagrass (*Cynodon transvaalensis* x *Cynodon dactylon*) to drought stress. *Plant Cell Rep.* 28:517-526.
- Shukor, N.A., Hamzah, M.B., Hazandy, A.H., Salleh, G. and Nasir, M.F. (2009). Growth and Phenology of Kenaf (*Hibiscus cannabinus* L.) Varieties. *Pertanika Journal of Tropical Agriculture Science* 32(1): 29 – 33.
- Siddique, M. R. B., Hamid, A. and Islam, M. S. (2001). Drought stress effects on water relations of wheat. *Botony Bulliten Academic Sinica.* 41: 35-39.
- Sionit, N., Rogers, H.H., Bingham, G.E. and Strain, B.R. (1984). Photosynthesis and stomatal conductance with CO₂-enrichment of container- and field-grown soybeans. *Agronomy Journal* 76:447–451.
- Slyvan, H.W. (2006). *Rising in carbon dioxide is good for plants.* www.purgit.com/co2ok.html. Accessed in 27 July 2006.
- Soussana, J.F. and Loiseau, P. (1997). Temperate grass swards and climatic changes, the role of plant–soil interactions in elevated CO₂. *Abstract Botany* 21: 223–234.
- Stegman, E.C., Schiele, L.H. and Bauer, A. (1976). Plant water stress criteria for irrigation scheduling. *Trans ASAE.* 19:850-855.
- Stephenson, R. A. and Callagher, E. C. (1990). Some aspects of water relations in macadamia. *Acta Horticulture* 75: 559-567.
- Stitt, M. (1991). Rising CO₂ levels and their potential significance for cotton flow in photosynthetic cell. *Plant Cell and Environment* 14: 741 – 762.
- Tahery, Y., Hazandy, A.H., Tahery, E., Deljoo, E. and Mogrilan, M. (2011). Comparative photosynthesis and transpiration of three varieties of *Hibiscus cannabinus* L. (Kenaf). *Australian Journal of Agriculture Research* 6(8): 2010-2014.
- Taiz, L. and Zeiger, E. (2006). *Plant physiology*, 4th Ed., Sinauer Associates Inc. Publishers, Massachusetts.
- Taylor, I. B. (1991). *Genetics of ABA synthesis*, in: Davies, W. J., Jones, H. G. (Eds.). *Abscisic acid: Physiology and Biochemistry.* Bios Scientific Publishers. Ltd. UK. 23-38.
- Taylor, J.A. and Llyold, J. (1992). Sources and sinks of atmospheric CO₂. *Australian Journal of Botany.* 40:401–418.

- Taylor, G., Ranasinghe, S, Bosac C., Gardner, S.D.L. and Ferris, R. (1994). Elevated CO₂ and plant growth: cellular mechanisms and responses of whole plants. *Journal of Experimental Botany* 45: 1761–1774.
- Tezera, W., Mitchell, V., Dviscoll, S. P. and Lawlor, D. W. (2002). Effects of water deficit and its interaction with CO₂ supply on the biochemistry and physiology of photosynthesis in sunflower. *Journal of Experimental Botany* 53: 1781-1791.
- Tischler, C. R., Polley, H. W. Johnson, H. B. and Pennington, R. E. (2000). Seedling response to elevated CO₂ in five epigeal species. *International Journal of Plant Sciences* 161:779–783.
- Tisserat, B., Herman, C., Silman, R. and Bothast, R.J. (1997). Using ultrahigh carbon dioxide levels enhances plantlet growth *in vitro*. *HortTechnology* 7: 282–289.
- Tissue, D.I. and Oechel, W.C.(1987). physiological responses of *Eriophorum vaginatum* to field elevated carbon dioxide and temperature in the *Alaskan tussock tundra*. *Ecology* 68: 401 – 410.
- Tolley, L.C. and Strain, B.R. (1984). Effects of CO₂ enrichment and water stress on growth of *Liquidambar styracifulia* and *pinus Taeda* seedlings. *Canadian Journal forest Research* 62: 2135 – 2139.
- Turner, N. C., Wright, G. C. and Siddique, K. H. M. (2001). Adaptation of grain legumes (pulses) to water-limited environments. *Advanced Agronomy* 71: 123-231.
- Vercruyse, J. (1984). *Semi-detailed soil survey of the gual periok zone, Kelantan*. Soil survey report-Belgain Aid Project. Department of Agriculture, Kuala Lumpur.
- Ververis, C., Georghiou, K., Christodoulakis, N., Santas, P., & Santas, R. (2004). Fiber dimensions, lignin and cellulose content of various plant materials and their suitability for paper production. *Industrial Crops and Products* 19(3): 245-254.
- Wahid, A. and Rasul, E. (2005). *Photosynthesis in leaf, stem, flower, and fruit*, in: Pessaraki M. (Ed.). *Handbook of Photosynthesis*. 2nd ed. CRC Press. Florida. 479-497.
- Wang, Y.S.H., Bunce, A.J. and Maas, L.J. (2003). Elevated carbon dioxide increases contents of antioxidant compounds in field grown strawberries. *Journal of Agricultural and Food Chemistry* 51: 4315–4320.
- Webber, C.L. III. (1993a). Yield components of five Kenaf cultivars. *Agronomic Journal* 85: 533–535.
- Webber, C.L. III. (1993b). Crude protein and yield components of six kenaf cultivars as affected by crop maturity. *Industrial Crop Products* 2: 27–31.
- Webber, C.L. III and Bledsoe, V.K. (2002). Plant maturity and kenaf yield components. *Industrial Crops Products* 16: 81-88.

- Wery, J., Silim, S. N., Knights, E. J., Malhotra, R. S., Cousin, R. (1994). Screening techniques and sources and tolerance to extremes of moisture and air temperature in cool season food legumes. *Euphytica*. 73: 73-83.
- Westgate, M. E. and Boyer, J. S. (1985). Osmotic adjustment and inhibition of leaf, root, stem and silk growth at low water potentials in maize. *Planta* 104: 540-554.
- Wikipedia Internet Encyclopedia.(2008).http://en.wikipedia.org/wiki/carbon_dioxide. Retrieved in 29 February 2008.
- Wise, L.E., Murphy, M. and D'Addieco, A.A. (1946). Chlorite holocellulose, its fractionation and bearing on summative wood analysis and studies on the hemicelluloses. *Paper Trade Journal* 122(2): 35-43.
- Wong, C.C., Nizam, A.M., Vijaysegaran, S., Mah, S.Y. and Ghawas, M. (2001). *Kenaf germplasm introductions and assessment of their adaptability in Malaysia for forage production*. p. 13-24. In: First technical review meeting on the National Kenaf Research project. MARDI.
- Woodward, F.I. and Bazzaz, F.A. (1988). The responses of stomatal density to CO₂ partial pressure. *Journal of Experimental Botany* 39: 1771–1781.
- Woodward, F.I. and Kelly, C.K. (1995). The influence of CO₂ concentration on stomatal density. *New Phytologist* 131: 1311–1327.
- Woodward, F.I. (2002). Potential impacts of global elevated carbon dioxide concentration on plants. *Current Opinion in Plant Biology* 5: 207–211.
- Zhang, T. (2003). *Improvement of kenaf yarn for apparel application*. Master thesis of Louisiana State University, US.
- Zhou, C., Ohtani, Y., Sameshima, K. and Zhen, M. (1997). Selection of kenaf (*Hibiscus cannabinus* L.) varieties for papermaking on arid hillside land in China. *Mok. Gakkais* 43(9): 770-777.
- Zhou, Y., Lam, H. M. and Zhang, J. (2007). Inhibition of photosynthesis and energy dissipation induced by water and high light stresses in rice. *Journal of Experimental Botany* 58: 1207-1217.