

UNIVERSITI PUTRA MALAYSIA

YIELD AND NUTRITIVE QUALITY OF FOUR NAPIER (Pennisetum purpureum Schumach.) CULTIVARS HARVESTED AT DIFFERENT AGES AS FRESH AND ENSILED FODDER

MOHAMAD ZAIHAN BIN ZAILAN

FP 2016 38



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By

MOHAMAD ZAIHAN BIN ZAILAN

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

May 2016

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

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May 2016

Chair: Associate Professor Dr. Halimatun Yaakub, PhD

Faculty: Agriculture

Studies were conducted to evaluate the yield and nutritive quality of four Napier (*Pennisetum purpureum*) cultivars namely Common, Silver, Red and Dwarf Napier harvested at 4, 6 and 8 weeks age as fresh and ensiled fodder. Common, Silver and Red Napier were classified as tall cultivars while Dwarf Napier is a short cultivar. The harvesting ages selected were within of the range of optimum cutting age for Napier grass.

Study 1 was conducted to determine the dry matter yield and leaf to stem ratio of fresh Napier cultivars at 4, 6 and 8 weeks old. The dry matter yield of Common Napier reached a peak of 6 tonnes ha^{-1} cut⁻¹ at 6-week old. The dry matter yield of Red Napier gradually increased and peaked 6 tonnes ha^{-1} cut⁻¹ at 8 weeks old. Silver Napier yielded a similar dry matter production as Dwarf Napier as well as the lowest yield throughout the study. The leaf to stem ratio of Napier cultivars declined significantly from 3.24 at 4 weeks to 1.94 at 6 weeks. Dwarf Napier had the highest leaf to stem ratio (3.93) among the cultivars.

The nutritional composition and digestibility of fresh Napier cultivars at 4, 6 and 8 weeks were evaluated in Study 2. Dwarf Napier had the highest nutritive quality among the cultivars throughout the harvesting ages (12 to 20% CP). Overall, tall cultivars have higher NDF, ADF and ADL content than Dwarf Napier. Interestingly, the crude protein content of Red Napier (11%) remained unchanged throughout the harvesting ages. In terms of digestibility, Dwarf Napier and 6-week old Red Napier were classified as high quality feed (> 70% IVDMD and > 65% IVOMD).

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Study 3 was conducted to evaluate and compare the nutritional composition and digestibility of fresh and ensiled cultivars at 6 and 8 weeks harvesting age. The crude protein of Common Napier increased significantly after ensiling process from 8 to 9%. In contrast, a significant loss in crude protein content was observed in ensiled Silver Napier (9% CP) compared to the fresh forage (10% CP). A significant loss in cell wall constituent of Napier grass was observed in ensiled cultivars regardless of the harvesting ages. All cultivars have similar IVDMD, and the IVDMD declined from 68 to 60% after ensilation. Nevertheless the improvement in IVOMD from 52 to 58% in silage might derive from the bacteria population.

In conclusion, Common Napier is recommended to be harvested at 6 weeks age to obtain highest dry matter yield as well as minimizing loss of nutritive value. Red Napier could be harvested at 6 and 8 weeks age since there were no change in crude protein content. Dwarf Napier had superior quality and could be harvested at 6 weeks since the crude protein was able to fulfill the requirement for growing and lactating animals. The dry matter yield of Dwarf Napier could be maximized by harvesting at 8 weeks age. Silver Napier had similar dry matter yield and no advantage in nutritive quality compared to Dwarf Napier. Nevertheless, Silver Napier is suggested to be harvested at 6 weeks since significant loss in nutritive value was observed especially crude protein at 8 weeks.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

HASIL DAN KUALITI NUTRITIF EMPAT NAPIER (Pennisetum purpureum Schumach.) KULTIVAR DITUAI PADA UMUR YANG BERBEZA SEBAGAI FODER SEGAR DAN PERAM

Oleh

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Satu kajian telah dijalankan untuk menilai hasil dan kualiti nutritif empat Napier (*Pennisetum purpureum*) kultivar dinamakan sebagai Napier Umum, Perak, Merah, Kerdil yang dituai pada minggu ke-4, 6 dan 8 sebagai foder segar dan silaj. Napier Umum, Perak dan Merah diklasifikasikan sebagai kultivar tinggi manakala rumput Kerdil adalah kultivar rendah. Peringkat penuaian telah dipilih dalam lingkungan umur tuaian optima bagi rumput Napier.

Kajian 1 telah dijalankan untuk menentukan hasil berat kering dan nisbah daun kepada batang bagi Napier kultivar pada minggu tuaian ke-4, 6 dan 8. Hasil berat kering bagi Napier Umum mencecah kemuncak, 6 tan ha⁻¹ potong⁻¹ pada minggu ke-6. Hasil berat kering Napier Merah meningkat secara berperingkat dan mencanak naik kepada 6 tan ha⁻¹ potong⁻¹ pada umur minggu ke-8. Napier Perak menghasilkan berat kering yang sama seperti Napier Kerdil sekaligus merupakan hasil yang terendah sepanjang kajian dijalankan. Nisbah daun kepada batang bagi kultivar Napier menurun secara signifikan dari 3.24 pada minggu ke-4 kepada 1.94 pada minggu ke-6. Napier Kerdil mempunyai nisbah daun kepada pada tertinggi (3.93) diantara kultivar.

Komposisi nutrisi dan kecernaan bagi kultivar Napier segar pada minggu ke-4, 6 dan 8 telah dinilai dalam kajian 2. Napier Kerdil mempunyai kualiti nutritif yang tertinggi diantara kultivar sepanjang umur tuaian (12 ke 20% CP). Secara keseluruhan, Napier tinggi menpunyai kandungan NDF, ADF dan ADL yang lebih tinggi berbanding Napier Kerdil. Yang menariknya, protin kasar bagi Napier Merah (11% CP) tidak berubah sepanjang umur tuaian,. Dalam istilah kecernaan, Napier Kerdil dan Napier Merah pada umur 6 minggu diklasifikasikan sebagai makanan kualiti tinggi (> 70% IVDMD and >65% IVOMD).

Kajian 3 dijalankan bagi menilai dan membandingkan komposisi nutrisi dan kecernaan bagi kultivar segar dan peram pada tuaian minggu ke-6 dan 8. Protin kasar bagi Napier Umum meningkat ketara selepas proses pemeraman daripada 8 ke 9%. Sebaliknya, kehilangan signifikan bagi kandungan protin kasar dalam Napier Perak peram (9% CP) berbanding jenis segar (10% CP). Kehilangan ketara bagi kandungan dinding sel dalam rumput Napier diperhatikan dalam kultivar peram tanpa mengira

umur tuaian. Kesemua kultivar mempunyai IVDMD yang sama, dan IVDMD menurun daripada 68% ke 60% selepas diperam. Akan tetapi, peningkatan dalam IVOMD dari 52 ke 58% dalam silaj kemungkinan berasal daripada populasi bakteria.

Sebagai kesimpulan, Napier Umum, disarankan untuk dituai pada umur 6 minggu untuk memiliki hasil berat kering yang tertinggi sekaligus meminimakan kehilangan nilai nutritif. Napier Merah boleh dituai pada umur 6 dan 8 minggu memandangkan tiada perubahan dalan kandungan protin kasar. Napier Kerdil mempunyai kualiti atasan dan boleh dituai pada minggu ke-6 memandangkan protin kasar dapat memenuhi keperluan bagi haiwan yang meningkat dewasa dan haiwan dalam laktasi. Hasil berat kering bagi Napier Kerdil boleh dimaksimakan dengan tuaian pada minggu ke-8. Napier Perak mempunyai hasil berat kering yang sama dan tiada kelebihan dalam kualiti nutritif dibandingkan dengan Napier Kerdil. Namun begitu, Napier Perak disarankan untuk dituai pada minggu ke-6 memandangkan kehilangan signifikan dalam nilai nutritif diperhatikan terutamanya protin kasar pada minggu ke-8.

ACKNOWLEDGEMENTS

I am using this opportunity to express my sincere gratitude to Assoc. Prof. Dr. Halimatun Yaakub and Dr. Shokri Jusoh for their patience, motivation and immense knowledge Grateful thanks are also due for their guidance and support that helped me to overcome many crisis situations during the study.

I would like to express my sincere appreciation to Prof. Dr. Abdul Razak Alimon and Prof. Dr. Jothi Malar Panandam for their aspiring guidance and invaluably constructive criticism with inspiring suggestion.

Thanks are also due to the contribution of staffs of Field 2 especially Mr. Mohd Faizal Yeop Baharudin and Mr. Hidayat Ali Aman Ali. I also wish to convey my thanks to Mr. Saparin Denim, Mr. Khairul Anwar Bahari and Mrs Rohaida Abd Rashid, staffs of the Nutrition Laboratory, Ms. Nurul Syuhada Adnan, staff of the Pasture Laboratory, Department of Animal Science, Faculty of Agriculture for their assistance in conducting the experiment.

Above all, I would like to acknowledge tremendous sacrifices that my father, Zailan Ismail and mother, Saerah Baharom made to ensure that I had excellent education Thank you both for giving me strength to reach the stars and chase my dreams.

Finally and most importantly, I would like to thank to my wife Ezzah Mahmudah Salim. Her support, encouragement, patience and unwavering love were undeniable.

I certify that a Thesis Examination Committee has met on 3 May 2016 to conduct the final examination of Mohamad Zaihan Bin Zailan on his thesis entitled "Yield and nutritive quality of four Napier (*Pennisetum purpureum* Schumach.) cultivars harvested at different ages as fresh and ensiled fodder" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

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LIST OF ABBREVIATIONS

| ADF | Acid detergent fiber |
|--------|---------------------------------------|
| ADL | Acid detergent lignin |
| ATP | Adenosine triphosphate |
| AA | Amino acid |
| ANF | Anti-nutritive factor |
| СНО | Carbohydrate |
| CO_2 | Carbon dioxide |
| CT | Condensed tannin |
| СР | Crude protein |
| °C | Degree Celsius |
| DE | Digestible energy |
| DMY | Dry matter yield |
| G3P | Glyceryldehyde-3-phosphate |
| g | Gram |
| GE | Gross energy |
| ha | Hectare |
| HCl | Hydrochloric acid |
| IVDMD | In vitro dry matter digestibility |
| IVOMD | In vitro organic matter digestibility |
| kg | Kilogram |
| LAB | Lactic acid bacteria |
| LSR | Leaves to stem ratio |
| MJ | Mega joule |
| ME | Metabolisable energy |
| CH4 | Methane gas |
| mM | Milli Molarity |
| ml | Milliliter |
| mm | Millimeter |
| NDF | Neutral detergent fiber |
| Ν | Nitrogen |
| OM | Organic matter |
| OAA | Oxaloacetate |

| h^{-1} | per hour |
|-----------|--------------------------------------|
| % | Percentage |
| RUBISCO | Ribulose-1,5-biphosphate carboxylase |
| RUP | Rumen undegradable protein |
| NaOH | Sodium hydroxide |
| H_2SO_4 | Sulphuric acid |
| WSC | Water soluble carbohydrate |

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CHAPTER 1

INTRODUCTION

The ruminant industry in Malaysia is faced with the problem of high cost of production. The scarcity of feed resources has made farmers dependent on imported commercial concentrate feed. Farmers need to find alternative sources of feed to substitute the heavy use of expensive commercial concentrates. Ironically, Malaysia can grow a wide range of forages that could reduce the use of concentrate feed and lower the feed cost.

Feed cost represents major single cost item for livestock production as it accounts for more than 70% of the total cost of production. Farmers are inclined to choose cheap and accessible feed source to maximize their profitability. The considerations of nutritional composition of forages and requirement of the animal itself are crucial in formulating the least cost ration. With this knowledge, the utilization of available sources of feedstuffs such as forages could be optimised. The self-sufficiency of livestock products especially dairy products are extremely low compared to other tropical countries. There should be collaborations among farmers and researchers in searching for suitable adaptive strategies such as pasture-based production to increase the productivity intended for consumption.

Forages are the best feed resources for ruminants as forages do not compete with agricultural production of concentrate feed for human consumption. Forages can be digested and utilised by ruminant as a source of nutrient. There is no doubt that ruminant digestive tract is capable in providing a favourable environment as a host for symbiotic microbes (bacteria, protozoa and fungi) to hydrolyze cellulose, hemicellulose and other substances that are resistant to enzymes secreted by the host animal.

Guinea (*Megathyrsus maximus*) and Napier are the common cultivated grass species in Malaysia. Guinea grass yielded from 9 to 12 tonnes dry matter yield ha⁻¹ cut⁻¹ with cumulative mean of more than 20 tonnes ha⁻¹ yr⁻¹ (Ahmed *et al.*, 2012; Munyasi *et al.*, 2015). Napier is the most popular forage species due to high dry matter production, high nutritive value and it can be easily established through stem propagation. Besides, the Napier grass tends to produce high dry matter yield up to 70 tonnes ha⁻¹ yr⁻¹ (Wijitphan *et al.*, 2009). The broad range in yield and nutritional composition is influenced by the morphology and management of Napier cultivars.

Generally, the determination of limiting factors is prerequisite in evaluating the yield and quality of grasses. The cultivar selection, cutting management (cutting frequency, interval and height) application of fertilizer (rate and type of fertilizer), soil condition, and environmental factors are among the crucial factors (Jusoh *et al.*, 2014; Lounglawan, *et al.*, 2014). Napier grass was first introduced to Malaysia in the 1920's and there were many cultivars introduced in Malaysia since 1950's known as Common Napier, Red Napier, Taiwan Napier, Dwarf Napier, Dwarf "Mott", Australian Dwarf, Indian Napier, Uganda Napier and King grass (Halim *et al.*, 2013; Jusoh, 2005). However, very few comparative studies on Napier cultivars had been done.

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The terminology of "cultivar" and "variety" bring a different meaning and these two terms often abused by farmers. According to Haynes (2009), varieties often occur in nature and most varieties are true to type, meanwhile cultivars is a combination of "cultivated" and "variety" which are not necessarily true type and it was selected and cultivated by humans. Nevertheless, it is possible for a plant to have both variety and cultivars. The documentation of Napier cultivars are scanty and cannot be relied upon for choosing the best cultivars. A number of Napier grass cultivars have been in circulation, often with more than one name (Struwig, 2007).

Napier grass grows best in high-rainfall areas up to 1500 mm rainfall yr⁻¹ but it does not tolerate flooding (FAO). The estimated area prone to flood disaster is 9% of total area in Malaysia (D/iya *et al.*, 2014). In spite of flood, tropical countries are more vulnerable to drought compared to temperate countries and therefore, the conservation of feed is crucial to preserve the quality and supply adequate feed to livestock. Silage was found to be more suitable than hay making process because of high relative humidity, more than 90% which will easily spoil the hay.

Statement of problem

Many cultivars of Napier grass (*Pennisetum purpureum*) have been planted in Malaysia as discussed previously. Common Napier, Red Napier, Taiwan Napier, Indian Napier, Uganda Napier, King grass, Zanzibar Napier and Kobe Napier are classified as tall cultivars whereas Dwarf Napier, Dwarf "Mott", Australian Dwarf are short cultivars. Generally, tall cultivars with a high yielding grass are normally grown in several areas under a cut-and-carry system in Malaysia. Common Napier is among the highest yielding crops and has a better nutritive value as compared to Uganda Napier (Halim *et al.*, 2013). Red Napier was high in metabolisable energy and this crucial parameter reflected the actual level of energy available for absorption (Haryani *et al.*, 2012). Dwarf Napier has high leaf to stem ratio and this associated with good forage quality. However, Silver Napier was recently introduced without any documentation regarding the performance of this cultivar. There is a need to have comparative evaluation of these Napier cultivars so that definite recommendations can be made in the choice and management of the respective cultivars. Four cultivars (Common, Red, Dwarf and Silver Napier) were selected for this study.

Objectives

The general aim of the proposed project is to investigate the yield and nutritive value of four Napier (*Pennisetum purpureum*) cultivars harvested at different ages as fresh and ensiled fodder.

The specific objectives of the projects are as below:

- 1) To determine the dry matter yield and proportion of leaves to stem fraction of different Napier cultivars harvested at 4, 6 and 8 weeks age
- 2) To evaluate the nutritional composition and digestibility of different Napier cultivars harvested at 4, 6 and 8 weeks age.
- To evaluate and compare the nutritional composition and digestibility of fresh and ensiled Napier cultivars harvested at 6 and 8 weeks age



REFERENCES

- Ahmed, S. A., Halim, R. A. and Ramlan, M. F. (2012). Evaluation of the use of farmyard manure on a Guinea Grass (*Panicum maximum*) - Stylo (*Stylosanthes guianensis*) mixed pasture. Pertanika Journal of Tropical Agricultural Science, 35(1), 55–65.
- Allen, V. G., Batello, C., Berretta, E. J., Hodgson, J., Kothmann, M., Li, X., ... Sanderson, M. (2011). An international terminology for grazing lands and grazing animals. The Journal of the British Grassland Society, 66, 2–28.
- Ansah, T., Osafo, E. L. K. and Hansen, H. H. (2010). Herbage yield and chemical composition of four varieties of Napier (*Pennisetum purpureum*) grass harvested at three different days after planting. Agriculture and Biology Journal of North America, 1(5), 923–930.
- Ansah, T., Osafo, E. L. K. and Hansen, H. H. 2013: Variety, harvest date after planting and plant fraction of Napier grass influence *in vitro* gas production. Livestock Research for Rural Development. Volume 25, Article #78. Retrieved January 8, 2016, from http://www.lrrd.org/lrrd25/5/ansa25078.htm
- Archimède, H., Eugène, M., Marie Magdeleine, C., Boval, M., Martin, C., Morgavi, D. P.,Doreau, M. (2011). Comparison of methane production between C3 and C4 grasses and legumes. Animal Feed Science and Technology, 166-167, 59– 64.
- Bach, A., Calsamiglia, S. and Stern, M. D. (2005). Nitrogen metabolism in the rumen. Journal of Dairy Science, 88 Suppl 1(July 2004), E9–21.
- Bai, C., Zhang, R., Jiang, C., Yan, R., Han, J., Zhu, Y., and Zhang, Y. (2011). Characterization of carbohydrate fractions and fermentation quality in ensiled alfalfa treated with different additives. African Journal of Biotechnology, 10(48), 9958–9968.
- Balch, C. C. (1959). Structure of the ruminant stomach and the movement of its contents. Proceedings of the Nutritional Society, 18(2), 97–102.
- Ball, D., Collins, M., Lacefield, G., Martin, N., Mertens, D., Olson, K., ... Wolf, M. (2001). Understanding forage quality. American Farm Bureau Federation Publication, 1-01.
- Barton, F. E., Amos, H. E., Burdick, D. and Wilson, R. L. (1976). Relationship of chemical analysis to *in vitro* digestibility for selected tropical and temperate grasses. Journal of Animal Science, 43(2), 504–512.
- Beyero, N., Kapoor, V. and Tewatia, B. S. (2015). Effect of different roughage : concentrate ratio on milk yield and its fatty acid profile in dairy cows. Journal of Biology, Agriculture and Healthcare, 5(13), 176–186.

- Bhatti, M. B., Mohammad, D., Sartaj and Sultani, M. I. (1985). Effect of different inter- and intra-row spacings on forage yield and quality in elephant grass. Pakistan Journal of Agricultural Research. 6(2), 107-112.
- Blaxter, K. L., Wainman, F. W., Wilson, R. S. (1961). The regulation of food intake by sheep. *Animal Production* 3:51
- Blummel, B. Y. M. and Becker, K. (1997). The degradability characteristics of fiftyfour roughages and roughage neutral-detergent fibres as described by *in vitro* gas production and their relationship to voluntary feed intake. British Journal of Nutrition, 77, 757–768.
- Blummel, M. and Orskov, E. R. (1993). Comparison of *in vitro* gas production and nylon bag degradability of roughages in predicting feed intake in cattle. Animal Feed Science and Technology, 40, 109–119.
- Brock, F. M., Forsberg, C. W. and Buchanan-smith, J. G. (1982). Proteolytic Activity of Rumen Microorganisms and Effects of Proteinase Inhibitors. Applied and Environmental Mictobiology, 44(3), 561–569.
- Budiman, Soetrisno, R. D., Budhi, S. P. S. and Indrianto, A. (2012). Morphological characteristics, productivity and quality of three Napier grass (*Pennisetum purpureum* Schum) cultivars harvested at different age. Journal of the Indonesian Tropical Animal Agriculture, 37(4), 294–301.
- Buxton, D. R. (1996). Quality-related characteristics of forages as influenced by plant environment and agronomic factors. Animal Feed Science and Technology, 59, 37–49.
- Carro, M. D. and Miller, E. L. (1999). Effect of supplementing a fibre basal diet with different nitrogen forms on ruminal fermentation and microbial growth in an *in vitro* semi- continuous culture system (RUSITEC). British Journal of Nutrition, 82, 149–157.
- Catchpoole, V. R., and Hanzel, E. F. (1971). Silage and silage making from tropical herbage species. *Herbage Abstracts*. 41:213.
- Cherdthong, A. and Wanapat, M. (2013). Manipulation of *in vitro* ruminal fermentation and digestibility by dried rumen digesta. Livestock Science, 153(1-3), 94–100.
- Chesson, A. and C. W Forsberg. (1988). Polysaccharide degradation by rumen microorganisms. In: P.N. Hobson (Ed.) The rumen microbial ecosystem, p 251. Elsevier Science Publishers, London, England.
- Cone, J. W. and Gelder, A. H. V. (1999). Influence of protein fermentation on gas production profiles. Animal Feed Science and Technology, 76, 251–264.

- Cone, J. W., Van Gelder, A. H., Soliman, I. A., De Visser, H. and Van Vuuren, A. M. (1999). Different techniques to study rumen fermentation characteristics of maturing grass and grass silage. Journal of Dairy Science, 82(5), 957–966.
- Cruz Soto, R., Muhammed, S. A., Newbold, C. J., Stewart, C. S. and Wallace, R. J. (1994). Influence of peptides, amino acids and urea on microbial activity in the rumen of sheep receiving grass hay and on the growth of rumen bacteria *in vitro*. Animal Feed Science and Technology, 49(94), 151–161.
- D/iya, S. G., BarzaniGasim, M., Toriman, M. E. and Abdullahi, M. G. (2014). Floods in Malaysia: Historical reviews, causes, effects and mitigations approach. International Journal of Interdisciplinary Research and Innovations, 2(4), 59– 65.
- Dijkstra, J. A. N. and Tamminga, S. (1995). Simulation of the effects of diet on the contribution of rumen protozoa to degradation of fibre in the rumen. British Journal of Nutrition, 74, 617–634.
- Edwards, E. J. and Smith, S. A. (2010). Phylogenetic analyses reveal the shady history of C4 grasses. Proceedings of the National Academy of Sciences, 107(6), 2532–2537.
- Elfrink, S. J. W. H. O., Frank D., Jan, G. C., Sierk, S. F. (2000). Silage fermentation processes and their manipulation. In FAO *Electronic Conference Tropical Silage, Edition Netherland: Food Agriculture Organization.* 1-28
- Evitayani, Warly, L., Fariani, A., Ichinohe, T. and Fujihara, T. (2004). Study on nutritive value of tropical forages in North Sumatra , Indonesia. Asian-Australasian Journal of Animal Sciences, 17(11), 1518–1523.
- Fievez, V., Babayemi, O. J. and Demeyer, D. (2005). Estimation of direct and indirect gas production in syringes: A tool to estimate short chain fatty acid production that requires minimal laboratory facilities. Animal Feed Science and Technology, 123-124, 197–210.
- Filho, J. C. M. N. and Fondevila, M. (2000). *In vitro* microbial fermentation of tropical grasses at an advanced maturity stage. Animal Feed Science and Technology, 83, 145–157.
- Furbank, R. T. and Taylor, W. C. (1995). Regulation of photosynthesis in C3 and C4 plants: A molecular approach. The Plant Cell, 7(July), 797–807.
- Geren, H. and Kavut Y. T. (2015). Effect of different plan densities on the yield and some silage quality characteristics of giant king grass (*Pennisetum hybridum*) under Mediterranean climatic conditions. Turkish Journal of Field Crops. 20(1), 85-91.

- Getachew, G., Blummel, M., Makkar, H. P. S. and Becker, K. (1998). *In vitro* gas measuring techniques for assessment of nutritional quality of feeds : a review. Animal Feed Science and Technology, 72, 261–281.
- Goto, I. and Minson, D. J. (1977). Prediction of the dry matter digestibility of tropical grasses using a pepsin-cellulase assay. Animal Feed Science and Technology, 2, 247–252.
- Grant, R. J. (1997). Interactions among forages and nonforage fiber sources. Journal of Dairy Science, 80(7), 1438–1446.
- Gwayumba, W., Christensen, D. A., McKinnon, J. J. and Yu, P. (2002). Dry matter intake, digestibility and milk yield by Friesian cows fed two Napier grass varieties. Asian-Australasian Journal of Animal Sciences, 15(4), 516-521.
- Halim, R. A., Shampazurini, S. and Idris, A. B. (2013). Yield and nutritive quality of nine Napier grass varieties in Malaysia. Malaysian Journal of Animal Science, 16(2), 37–44.
- Hamilton, D. F. and Johnson, C. R. (1978). Effects of organic matter and controlledrelease fertilizer in nutrient retention during intermittent-mist propagation. Scientia Horticulturae, 8, 155–162.
- Harmon, D. L. and Mcleod, K. R. (2001). Glucose uptake and regulation by intestinal tissues : Implications and whole-body energetics. Journal of Animal Science, 79(E. Suppl.), E59–E72.
- Haryani, H., Norfadzrin, F., Aswanimiyuni, A., Syed Hussein S. A., Abu Hassan, M. A., Azman, A. (2012). The growth performance and nutritive values of six types of Napier at different cutting age. In Proceedings of the 33rd Malaysian Society of Animal Production Conference. Langkawi, Malaysia. (Page: 102-103).
- Haynes C. (2009). "Cultivar versus variety". *Horticulture and Home Pest News*. Iowa State University. IC-499(2).
- Haynes, C. (2008). "Cultivar versus Variety". Horticulture and Home Pest News.IC-499 (2) – February 2008. Iowa state university. <u>http://www.ipm.iastate.edu/ipm/hortnews/2008/2-6/CultivarOrVariety.html</u>. Web. 19 Feb. 2016.
- Hersom, M. and Kunkle, W. E. (2014). Strategies for cost-effective supplementation of beef strategies to optimize cattle performance. SS-ANS-14, Gainesvill(UF/IFAS Extension), 1–7.
- Hetta, M., Cone, J. W., Bernes, G., Gustavsson, A.-M. and Martinsson, K. (2007). Voluntary intake of silages in dairy cows depending on chemical composition and *in vitro* gas production characteristics. Livestock Science, 106, 47–56.

- Hetta, M., Cone, J. W., Gustavsson, A.-M. and Martinsson, K. (2003). The effect of additives in silages of pure timothy and timothy mixed with red clover on chemical composition and *in vitro* rumen fermentation characteristics. Grass and Forage Science, 58(3), 249–257.
- Huhtanen, P., Brotz, P. G. and Satter, L. D. (1997). Omasal sampling technique for assessing fermentative digestion in the forestomach of dairy cows. Journal of Animal Science, 75, 1380–1392.
- Idris, A. B., A. R. Alimon and N. Abu Bakar. 2010. The growth performance and nutritive value of three varieties of Napier Grass fertilized at two levels of nitrogen. Proc 31st MSAP Ann. Con., Kota Bahru, Kelantan. Pp 101-102.
- Jenkins, T. C. (1993). Symposium: Advances in ruminant lipid metabolism. Journal of Dairy Science, 76(12), 3851–3863.
- Johnson, D. E. and Ward, G. M. (1996). Estimates of animal methane emissions. Environmental Monitoring and Assessment, 42, 133–141.
- Jusoh, S. (2005). Effects of sheep manure application on the production of Dwarf Napier grass (*Pennisetum Purpureum* cv. Mott), Master Thesis, Universiti Putra Malaysia.
- Jusoh, S., Alimon, A. R. and Kamiri, M. S. (2014). Agronomic properties, dry matter production and nutritive quality of Guinea Grass (*Megathrysus maximus*) harvested at different cutting intervals. Malaysian Journal of Animal Science, 17(2), 31–36.
- Kanjanapruthipong, J., Buatong, N. and Buaphan, S. (2001). Effects of roughage neutral detergent fiber on dairy performance under tropical conditions. Asian-Australasian Journal of Animal Sciences, 14(10), 1400–1404.
- Kariuki, J. N., Tamminga, S., Gachuiri, C. K., Gitau, G. K. and Muia, J. M. K. (2001). Intake and rumen degradation in cattle fed Napier grass (*Pennisetum purpureum*) supplemented with various levels of *Desmodium intortum* and *Ipomoea batatus* vines. South African Journal of Animal Science, 31(3), 149–157.
- Kering, M. K., Guretzky, J., Funderburg, E. and Mosali, J. (2011). Effect of nitrogen fertilizer rate and harvest season on forage yield, quality, and macronutrient concentrations in midland Bermuda grass. Communications in Soil Science and Plant Analysis, 42(16), 1958–1971.
- Khachatur, M. B. (2006). *In vitro* digestible organic matter and energy contents in wild growing forages of Armenia. Journal of Central European Agriculture, 7(3), 445–450.

- Khandaker, Z. H. and Tareque, A. M. M. (1996). Studies on protein degradabilities of feedstuffs in Bangladesh. Asian-Australasian Journal of Animal Sciences, 9(6), 637–642.
- Koster, H. H., Meissner, H. H. and Coertze, R. J. (1992). Variation in the production and quality of Bana grass over the growing season using hand-clipped sample. South African Journal of Animal Science, 22(1), 31-34.
- Kurihara, M., Magner, T., Hunter, R. A. and Mccrabb, G. J. (1999). Methane production and energy partition of cattle in the tropics. British Journal of Nutrition, 81, 227–234.
- Lee, M., Hwang, S. and Chiou, P. W. (2000). Metabolizable energy of roughage in Taiwan. Small Ruminant Research, 36, 251–259.
- Liang, J. B. and Samiyah, M. N. (1988). Comparative intake , digestibility and utilization of guinea grass by buffaloes and cattle. MARDI Research Journal, 16(1), 43–47.
- Lopez, S., France, J., Gerrits, W. J. J., Dhanoa, M. S., Humphries, D. J. and Dijkstra, J. (2000). A generalized Michaelis-Menten equation for the analysis of growth. Journal of Animal Science, 78, 1816–1828.
- Lounglawan, P., Lounglawan, W. and Suksombat, W. (2014). Effect of cutting interval and cutting height on yield and chemical composition of King Napier grass (*Pennisetum purpureum x Pennisetum americanum*). APCBEE Procedia, 8, 27–31.
- Luchini, N. D., Broderick, G. A., Muck, R. E., Makoni, N. F. and Vetter, R. L. (1997). Effect of storage system and dry matter content on the composition of alfalfa silage. Journal of Dairy Science, 80(8), 1827–1832.
- Mannetje, L. T, (1992). Pennisetum purpureum Schumach.In: Mannetje, L.'t and Jones, R.M. (Editors). Plant Resources of South-East Asia No. 4: Forages. Pudoc, Wageningen, The Netherlands, pp. 191-192
- Manyawu, G. J., Chakoma, C., Sibanda, S., Mutisi, C., and Chakoma, I. C. (2003). The effect of harvesting interval on herbage yield and nutritive value of Napier grass and hybrid *Pennisetums*. Asian-Australasian Journal of Animal Sciences, 16(7), 996-1002.
- Mccrabb, G. J. and Hendricksen, R. E. (2000). Gross Energy Content of some Native Pasture Grasses in Tropical. Asian-Australasian Journal of Animal Sciences, 13(5), 124.
- McDonald, P. J., Henderson, A R., Heron, S. J. E (1991). *The Biochemistry of silage* (2nd Ed.) Mallow Chalcombe Publications, ISBN 0948617225

- Menke, K. H., Raab, L., Salewski, A., Steingass, H., Fritz, D., Schneider, W. (1979). The estimation of the digestibility and metabolizable energy content of ruminant feedingstuffs from the gas production when they are incubated with rumen liquor *in vitro*. Journal of Agricultural Science (Cambridge), 93:217-222
- Menke, K.H. and Steingass, H. (1988). Estimation of the energetic feed value obtained from chemical analysis and gas production using rumen fluid. Animal Research and Development. 28: 7-55.
- Mertens, D. R. (1997). Creating a system for meeting the fiber requirements of dairy cows. Journal of Dairy Science, 80(7), 1463–81.
- Miettinen, H. and Huhtanen, P. (1996). Effects of the ratio of ruminal propionate to butyrate on milk yield and blood metabolites in dairy cows. Journal of Dairy Science, 79(5), 851–861.
- Minson, D. J. (1990). Forage in Ruminant Nutrition. Academic Press, London, 483 pp.
- Mlay, P. S., Pereka, A., Phiri, E. C., Igusti, J., Hvelplund, T., Weisbjerg, M. R., ... Feed, J. M. (2006). Feed value of selected tropical grasses, legumes and concentrates. Veterinarski Arhiv, 76(1), 53–63.
- Mohammad, N., Butt, N. M. and Qamar, I. A. (1988). Effect of nitrogen fertilization and harvesting interval on the yield and nutritional value of Napier grass. Pakistan Journal of Agricultural Research, 9(4), 478–482.
- Mohd Najib, M. A. and Hassan A. W. (1985). Effects of farmyard manure and inorganic fertilizers on dry matter production of two grasses. MARDI Research Bulletin, 13(3), 323-332.
- Morrison, M., Mackie, R. I. and Kistner, A. (1990). 3-Phenylpropanoic acid improves the affinity of Ruminococcus albus for cellulose in continuous culture. Applied and Environmental Microbiology, 56(10), 3220–3222.
- Mouafi, F. E., Abdel-aziz, S. M. and Bashir, A. A. (2013). Nutritive value of ensiled Mangrove leaves by Lactobacillus plantarum I . fermentation characteristics and chemical composition. World Applied Sciences Journal, 28(4), 499–508.
- Mounika, B., Chellamuthu, V. and Sridevi, V. (2015). Plant spacing influence the relative productivity of Bajra Napier hybrid grasses. International Journal of Tropical Agriculture. 33(2), 875-878.
- Muck, R. E. (2010). Silage microbiology and its control through additives. Revista Brasileira de Zootecnia, 39, 183–191.
- Muck, R.E. (1989). Initial bacterial numbers on Lucerne prior to ensiling. Grass Forage Science. 44: 19-25.

- Mukhtar, M., Ishii, Y., Tudsri, S., Idota, S. and Sonoda, T. (2003). Dry matter productivity and overwintering ability of the Dwarf and normal Napier grasses as affected by the planting density and cutting frequency. Plant Production Science. 6(1), 65-73.
- Mulligan, F. J., Quirke, J., Rath, M., Caffrey, P. J. and Mara, F. P. O. (2002). Intake, digestibility, milk production and kinetics of digestion and passage for diets based on maize or grass silage fed to late lactation dairy cows. Livestock Production Science, 74, 113–124.
- Munyasi, J. W., Auma, E. O., Ngode, L. and Muyekho, F. N. (2015). Evaluation of biomass yield of selected fodder species in relation to frequency of harvest alongside defoliation heights in Western Kenya. Journal of Agricultural Extension and Rural Development, 7(8), 257–262.
- Murphy M.R., Baldwin R.L. and Koon L.J. (1982) Estimation of stoichiometric parameters for rumen fermentation of roughage and concentrate diets. Journal of Animal Science. 55, 411-421.
- Na, R., Dong, H., Zhu, Z., Chen, Y. and Xin, H. (2013). Effects of forage type and dietary concentrate to forage ratio on methane emissions and rumen fermentation characteristics of dairy cows in China. American Society of Agriculture and Biological Engineers, 56(3), 1115–1122.
- National Research Council (1989). Nutrient Requirement of Dairy Cattle. 6th rev. ed. Natl. Acad. Sci., Washington, DC.
- Nocek, J. E. and Russell, J. B. (1988). Protein and Energy as an Integrated System. Relationship of Ruminal Protein and Carbohydrate Availability to Microbial Synthesis and Milk Production. Journal of Dairy Science, 71(8), 2070–2107.
- Noziere, P., Ortigues-Marty, I. and Sauvant, D. (2010). Carbohydrate quantitative digestion and absorption in ruminants: from feed starch and fibre to nutrients available for tissues. Animal, 4(7), 1057–1074.
- Nyambati, E. M., Muyekho, F. N., Onginjo, E. and Lusweti, C. M. (2010). Production, characterization and nutritional quality of Napier grass (*Pennisetum purpureum* (Schum.)) cultivars in Western Kenya. 4(12), 496-502.
- Obok, E. E., Ova, M. E. A. and Iwo, G. A. (2012). Forage potentials of interspecific hybrids between elephant grass selections and cultivated pearl millet genotypes of Nigerian origin. Journal of Plant Breeding and Crop Science, 4(9), 136–143.
- Ohmomo, S., Tanaka, O., KItamoto, H. K. and Cai, Y. (2002). Silage and microbial performance, old history but new problem. JARQ, Vol. 40, No. 2 (April 2002), pp. 59-71, ISSN

- Orskov, E. R., Hovell, F. D. and Mould, F. (1980). The use of the nylon bag technique for the evaluation of feedstuffs. Tropical Animal Production, 5(3), 195–213.
- Patil, D. B. and Joshi, A. B. (1962). Pusa Giant Napier. Indian Farming 18:7-8.
- Phiri, M. S., Ngongoni, N. T., Maasdorp, B. V, Titterton, M. and Mupangwa, J. F. (2007). Ensiling characteristics and feeding value of silage made from browse tree legume-maize mixtures. Tropical and Subtropical Agroecosystems, 7, 149– 156.
- Premaratne, S. and Premalal, G. G. C. (2006). Hybrid Napier (*Pennesitum purpureum* x *Pennisetum americanum*) var. CO-3 a resourceful fodder grass for dairy development in Sri Lanka. The Journal of Agricultural Sciences, 2(1), 22-33.
- Rahman, M. M. and Kawamura, O. (2011). Oxalate accumulation in forage plants : Some agronomic, climatic and genetic aspects. Asian-Australasian Journal of Animal Sciences, 24(3), 439–448.
- Rahman, M. M., Ishii, Y., Niimi, M. and Kawamura, O. (2009). Effect of clipping interval and nitrogen fertilisation on oxalate content in pot-grown napier grass (*Pennisetum purpureum*). Tropical Grassland, 43, 73–78.
- Ramadhan, A., Njunie, M. N. and Lewa, K. K. (2015). Effect of planting material and variety on productivity and survival of Napier grass (*Pennisetum purpureum* schumach) in the coastal lowlands of Kenya. East African Agriculture and Forestry Journal, 81(1), 40-45.
- Ridwan, R., Rusmana, I., Widyastuti, Y., Wiryawan, K. G., Prasetya, B., Sakamoto, M. and Ohkuma, M. (2015). Fermentation characteristics and microbial diversity of tropical grass-legumes silages. Asian-Australasian Journal of Animal Sciences, 28(4), 511-518.
- Robertson, L.J. and Waghorn, G.C. (2003). Dairy industry perspectives on methane emissions and production from cattle fed pasture or total mixed rations in New Zealand. Proceedings of the New Zealand Society of Animal Production, 62, 213 218.
- Rodrigues, L. R., Mott, G. O., Veiga, J. B. and Ocumpaugh, R. (1986). Tillering and mohphological characteristics of dwarf elephantgrass under grazing'. Pesquisa Agropecuaria Brasileira, 21(11), 1209–1218.
- Russell, J. B. and Wilson, D. B. (1996). Why are ruminal cellulolytic bacteria unable to digest cellulose at low pH? Journal of Dairy Science, 79(8), 1503–9.
- Russell, J. B., O'Connor, J. D., Fox, D. G., Van Soest, P. J. and Sniffen, C. J. (1992). A Net Carbohydrate and Protein System for Evaluating Cattle Diets: I. Ruminal Fermentation. Journal of Animal Science, 70, 3551–3561.

- Rymer, C., Williams, B. a., Brooks, A. E., Davies, D. R. and Givens, D. I. (2005). Inter-laboratory variation of *in vitro* cumulative gas production profiles of feeds using manual and automated methods. Animal Feed Science and Technology, 123-124, 225–241.
- Sandrin, C. Z., Domingos, M. and Figueiredo-Ribeiro, R. C. L. (2006). Partitioning of water soluble carbohydrates in vegetative tissues of Lolium multiflorum Lam . ssp . italicum cv . Lema. Brazilian Journal of Plant Physiology, 18(2), 299–305.
- Sarwatt, S. V., Urio, N. A.; Ekern, A. (1992). Evaluation of some tropical forage as silage. Improved Dairy Production from Cattle and Goats in Tanzania, NORAGRIC reports, 11, 14-24.
- Seymour, W. M., Campbell, D. R. and Johnson, Z. B. (2005). Relationships between rumen volatile fatty acid concentrations and milk production in dairy cows: a literature study. Animal Feed Science and Technology, 119, 155–169.
- Shafiq Zahid, M., Haqqani, A. M., Usman Mufti, M. and Safdar Shafeeq (2002). Optimization of N and P fertilizer for higher fodder yield and quality in Mottgrass under irrigation-cum rainfed conditions of Pakistan. Asian Journal of Plant Sciences, 1(6), 690-693.
- Soderlund, S. (1995). Effect of moisture level and fermentation components of ensiled feedstuffs on voluntary dry matter intake. In: Oklahama State University: Oklahama Agriculture Experiment Station. P-942, 264-271.
- Stern, M. D. and Hoover, W. H. (1979). Methods for determining and factors affecting ruman microbial protein synthesis: A review. Journal of Animal Science, 49(6), 1590–1603.
- Storm, E. and Orskov, E. R. (1983). The nutritive value of rumen micro-organisms in ruminants 1. Large-scale isolation and chemical composition of rumen microorganisms. British Journal of Nutrition, 50, 463–470.
- Struwig M. (2007). Fingerprinting *Pennisetum purpureum* varieties and cultivars using AFLP analyses. Thesis, Masters of Environmental Sciences, North West University.
- Sturm, C. D., Tiemann, T. T., Lascano, C. E., Kreuzer, M. and Hess, H. D. (2007). Nutrient composition and *in vitro* ruminal fermentation of tropical legume mixtures with contrasting tannin contents. Animal : An International Journal of Animal Bioscience, 138, 29–46.
- Sutton, J. D. (1985). Digestion and absorption of energy substrates in the lactating cow. Journal of Dairy Science, 68(12), 3376–3393.
- Tamminga, S. (1979). Protein degradation in the forestomachs of ruminants. Journal of Animal Science, 49(6), 1615–1629.

- Tessema, Z. and Baars, R. M. T. (2004). Chemical composition, *in vitro* dry matter digestibility and ruminal degradation of Napier grass (*Pennisetum purpureum* (L.) Schumach) mixed with different levels of Sesbania sesban (L.) Merr. Animal Feed Science and Technology, 117(1-2), 29–41.
- Torres, J., Rutherfurd, S. M., Muñoz, L. S., Peters, M. and Montoya, C. a. (2016). The impact of heating and soaking on the *in vitro* enzymatic hydrolysis of protein varies in different species of tropical legumes. Food Chemistry, 194, 377–82.
- Trujillo, W., Pitman, W. D., Chambliss, C. G. and Williams, K. (1996). Effect of height and frequency of cutting yield, quality and persistence of Desmanthus virgatus. Tropical Grassland, 30, 367–373.
- Tuah, A. K., Buadu, M. K., Fiagome, G. E. K. and Sackey, A. K. (1979). Studies on the nutritive value of giant star and guinea grass forages in the Ashanti forest belt of Ghana. Ghana Journal of Agricultural Science, (12), 103–111.
- Ukanwoko, A. I. and Igwe, N. C. (2012). Proximate composition of some grass and legume silages prepared in a humid tropical environment. International Research Journal of Agriculture Science and Soil Science, 2(2), 68–71.

Van Soest P J. (1982). Nutritional ecology of the ruminant. Cornell University Press, Ithaca, NY, USA.

- Van Soest, P. J. (1994). Nutritional ecology of ruminants. 2nd Edn, Cornell University Press, New York, ISBN-13: 978-0801427725.
- Walt, J. G. Van Der and Meyer, J. H. F. (1988). Protein digestion in ruminants. South African Society for Animal Science, 18(1), 30–41.
- Walton, P. D. (1984). Production and management of cultivated forages. Reston Publishing Company. Inc. Virginia. USA, pp 335.
- Wangchhuk, K., Rai, K., Nirola, H., Thukten, Dendup, C. and Mongar, D. (2015). Forage growth, yield and quality responses of Napier hybrid grass cultivars to three cutting intervals in the Himalayan foothills. Tropical grasslands, 3, 142-150.
- Warner, D., Hatew, B., Podesta, S. C., Klop, G., van Gastelen, S., van Laar, H., ... Bannink, A. (2015). Effects of nitrogen fertilisation rate and maturity of grass silage on methane emission by lactating dairy cows. Animal, 10(1), 34–43.
- Whittenbury, R., McDonald, P. and Bryan-Jones, D. G. (1967). A short review of some biochemical and bacteriological aspects of ensilage. Journal of the Science of Food and Agriculture, 18:441.

- Wijitphan, S., Lorwilai, P. and Arkaseang, C. (2009). Effect of cutting heights on productivity and quality of King Napier Grass (*Pennisetum purpureum* cv. King Grass) under irrigation. Pakistan Journal of Nutrition, 8(8), 1244–1250.
- Wolin, M. J. (1960). A theoretical rumen fermentation balance. Journal of Dairy Science, 43(10), 1452–1459.
- Yahaya, M. S., Goto, M., Yimiti, W., Smerjai, B. and Kawamoto, Y. (2004). Evaluation of fermentation quality of a tropical and temperate forage crops ensiled with additives of fermented juice of epiphytic lactic acid bacteria (FJLB). Asian-Australasian Journal of Animal Sciences, 17(7), 942–946.
- Yahaya, M. S., Kimura, A., Harai, J., Nguyen, H. V, Kawai, M., Takahashi, J. and Matsuoka, S. (2001). Effect of length of ensiling on silo degradation and digestibility of structural carbohydrates of lucerne and orchardgrass. Animal Feed Science and Technology, 92, 141–148.
- Zanine, A. D. M., Santos, E. M., Ricardo, J., Dórea, R., Dantas, A. D. S., Carvalho, T. and Pereira, O. G. (2010). Evaluation of elephant grass silage with the addition of cassava scrapings. Revista Brasileira de Zootecnia, 39(12), 2611–2616.
- Zewdu, T., Baars, R. M. T. and Yami, A. (2013). Effect of plant height at cutting and fertilizer on growth of Napier grass (*Pennisetum purpureum*). Tropical Science, 42, 57-61.