

# **UNIVERSITI PUTRA MALAYSIA**

EFFECTS OF SUBSTITUTING RICE STRAW WITH DIFFERENT LEVELS OF Leucaena leucocephala (Lam.) DE WIT AND Manihot esculenta CRANTZ LEAVES ON RUMEN FERMENTATION CHARACTERISTICS AND MICROBIAL POPULATION IN GOATS

NUR LIYANA AKMAL BINTI HARUN

FP 2015 50



# EFFECTS OF SUBSTITUTING RICE STRAW WITH DIFFERENT LEVELS OF Leucaena leucocephala (Lam.) DE WIT AND Manihot esculenta CRANTZ LEAVES ON RUMEN FERMENTATION CHARACTERISTICS AND MICROBIAL POPULATION IN GOATS



By

NUR LIYANA AKMAL BINTI HARUN

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

December 2015

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

# EFFECTS OF SUBSTITUTING RICE STRAW WITH DIFFERENT LEVELS OF Leucaena leucocephala (Lam.) DE WIT AND Manihot esculenta CRANTZ LEAVES ON RUMEN FERMENTATION CHARACTERISTICS AND MICROBIAL POPULATION IN GOATS

By

## NUR LIYANA AKMAL BINTI HARUN

### December 2015

## Chairman Faculty

Anjas Asmara @ Ab. Hadi b. Samsudin, PhD Agriculture

The leaves of L. leucocephala and M. esculenta are among good source of protein and its use in animal feed would further increase the nutritive value of poor quality feed. Generally, the presence of the anti-nutritive factors contained in both of the forages limit their usefulness as animal feed, but not in ruminant. Ruminant have a dynamic and complex rumen ecology that may have the ability to degrade the anti-nutritive factors contained in the L. leucocephala and M. esculenta leaves. Most of the previous studies are focused on the effect of L. leucocephala and M. esculenta leaves on performances and health of the animal. Therefore, current study was conducted to evaluate the effect of substitution rice straw with different level of L. leucocephala and M. esculenta leaves on rumen fermentation characteristics and rumen microbial population in goats. Seven treatment groups; T1: concentrates (C)/rice straw (RS) (40:60) (Control); T2: C/RS/leucaena leaves (40:45:15); T3:C/RS/leucaena leaves (40:30:30); T4: C/RS/leucaena leaves (40:15:45); T5:C/RS/cassava leaves (40:45:15); T6:C/RS/cassava leaves (40:30:30) and T7: C/RS/cassava leaves (40:15:45) were used in this experiment. An in-vitro study was carried out to evaluate the effect of substitution of rice straw with different level of L. leucocephala and M. esculenta leaves diet on rumen gas production and fermentation characteristics. In the *in-vitro* study, the production of propionate and IVDMD were significantly affected with the treatment diets (P<0.05). Propionate production was significantly increased meanwhile the percentages of IVDMD was decreased significantly with T2 showed the highest value in the propionate production and the lowest value of IVDMD. Similar observation were found among treatment diets in rumen pH, rumen ammonia, acetate, butyrate, total VFA production and total gas production of the *in-vitro* study. In the *in*vivo study, 21 local Boer goats were randomly divided into seven diets and were placed in metabolic cages individually for a period of 7 days as an adjustment period followed by an administration diet period for 10 days. Urine samples were taken daily for 5 days period for urinary purine derivatives analysis and rumen samples were taken at day 10 for rumen fermentation characteristics and rumen microbial population quantification analysis. Result shown that propionate (P<0.05), butyrate (P<0.01), A to P ratio (P<0.05), total VFA production (P<0.01) were affected significantly among the treatment diets. The highest value of acetate, A to P ratio and total VFA were found in

T2 with the lowest value of propionate. Supplementation of L. leucocephala and M. esculenta leaves diets demonstrated significant increase (P<0.01) in the production of rumen ammonia. There were no differences shown in the rumen pH and production of acetate. Urinary allantoin (P<0.05), xanthine (P<0.05), total PD (P<0.05), microbial purine absorbed (P<0.01) and microbial N supply (P<0.01) were affected significantly with T2 and T6 showed the highest value among the treatment diets whereas production of uric acid, hypoxanthine showed similar observation. In the microbial population study, the supplementation of L. leucocephala or M. esculenta leaves in the diet had increased the population of total protozoa and R. flavefacien significantly (P<0.05) with T4 and T7 showed the highest value, respectively. Meanwhile, the populations of *R. albus*, and *F. succinogenes* were significantly reduced (P<0.05) with T3 and T7 showed the lowest value, respectively. In conclusion, T2 (25% of L. leucocephala leaves) and T6 (50% of M. esculenta leaves) supplementation diet are suitable diet for goat consumption due to improvement in the VFA production, urinary purine derivative productions, microbial purine absorbed and microbial N supply with moderate amount rumen microbial population.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

## KESAN PENGGANTIAN JERAMI PADI DENGAN BERLAINAN ARAS DAUN Leucaena Leucocephala (Lam.) DE WIT DAN Manihot esculenta CRANTZ PADA CIRI-CIRI FERMENTASI RUMEN DAN POPULASI MIKROB RUMEN PADA KAMBING

Oleh

### NUR LIYANA AKMAL BINTI HARUN

#### **Disember 2015**

### Pengerusi Fakulti

Anjas Asmara @ Ab. Hadi b. Samsudin, PhD Pertanian

Daun L. leucocephala dan M. esculenta adalah antara sumber protein yang baik dan penggunaannya dalam makanan haiwan akan meningkatkan lagi nilai nutrien makanan yang berkualiti rendah. Umumnya, kehadiran faktor anti-nutrien yang terkandung di dalam kedua-dua foraj menghadkan penggunaan mereka sebagai makanan ternakan, tetapi tidak dalam ruminan. Ruminan mempunyai rumen ekologi yang dinamik serta kompleks dan mungkin mempunyai keupayaan untuk mendegradasi faktor anti-nutrien yang terkandung di dalam daun L. leucocephala dan M. esculenta. Kebanyakan kajian sebelum ini memberi tumpuan kepada kesan daun L. leucocephala dan M. esculenta pada prestasi dan kesihatan haiwan. Oleh itu, kajian semasa telah dijalankan untuk menilai penggantian jerami padi dengan perbezaan aras daun L. leucocephala dan M. esculenta pada ciri-ciri fermentasi rumen dan rumen populasi mikrob dalam kambing. Tujuh kumpulan rawatan; T1:konsentrat (K)/jerami padi (JP) (40:60) (Kontrol); T2:K/JP/daun leucaena (40:45:15); T3:K/JP/daun leucaena (40:30:30); T4:K/JP/daun leucaena (40:15:45); T5:K/JP/daun leucaena (40:45:15); T6:K/JP/daun leucaena (40:30:30) dan T7:K/JP/daun leucaena (40:15:45) telah digunakan di dalam ekperimen ini. Satu kajian in-vitro telah dijalankan untuk menilai kesan penggantian jerami padi dengan perbezaan aras daun L. leucocephala dan M. esculenta di dalam diet pada pengeluaran gas rumen dan ciri-ciri fermentasi. Total pengeluaran gas yang sama diperhatikan apabila perbandingan dibuat diantara diet dengan diet kontrol. Dalam kajian in-vitro, produksi asid propionic dan ketercernaan bahan kering in-vitro telah terjejas secara signifikan dengan diet rawatan (P<0.05). Produksi propionik telah meningkat secara signifikan sementara peratus ketercernaan bahan kering in-vitro telah berkurangan secara signifikan dengan T2 menunjukkan nilai tertinggi di dalam produksi asid propionic dan nilai terendah di dalam ketercernaan bahan kering *in-vitro*. Pemerhatian yang serupa telah didapati di antara diet rawatan pada pH rumen, rumen ammonia, asid asetik, asid butyric, total asid lemak meruap dan total produksi gas meruap dalam kajian in-vitro. Dalam kajian in-vitro, 21 kambing Boer tempatan telah dibahagi secara rawak kepada 7 diet dan telah diletakkan di dalam kandang metabolic secara individual selama 7 hari sebagai tempoh adaptasi diikuti dengan tempoh pentadbiran rawatan selama 10 hari. Sampel air kencing diambil setiap hari selama 5 hari untuk analisis derivatif purin dalam urin dan sampel rumen telah diambil pada hari

iii

10 untuk analisis ciri-ciri rumen fermentasi dan kuantifikasi populasi rumen mikrob. Keputusan menunjukkan bahawa asid propionik (P < 0.05), asid butyrik (P < 0.01), nisbah A kepada P (P<0.05), total asid lemak meruap (P<0.01) telah terkesan secara signifikan di antara rawatan diet. Nilai tertinggi asid asetik, nisbah A kepada P dan total asid meruap telah dijumpai pada T2 dengan asid propionic pada nilai yang terendah. Penggantian jerami padi dengan daun L. leucocephala and M. esculenta diet telah menunjukkan peningkatan yang signifikan (P<0.01) di dalam produksi rumen ammonia. Terdapat tiada perbezaan yang ditunjukkan di dalam pH rumen dan produksi asid asetik. Alantoin urin (P<0.05), xantina (P<0.05), jumlah derivatif purin (P<0.05), penyerapan purin mikrob (P<0.01) dan bekalan N mikrob (P<0.01) telah terjejas secara signifikan dengan T2 dan T6 menunjukkan nilai tertinggi di antara rawatan diet dimana produksi asid urik, hipoxantina menunjukkan pemerhatian yang sama. Di dalam kajian populasi mikrob, penggantian jerami padi dengan daun L. leucocephala dan M. esculenta di dalam diet telah meningkatkan populasi jumlah protozoa serta R. flavefacien dengan ketara (P<0.05) dengan T4 dan T7 masing-masing menunjukkan nilai tertinggi. Sementara itu, populasi R. Albus dan F. succinogenes telah dikurangkan dengan ketara (P<0.05) dengan T3 dan T7 masing-masing menunjukkan nilai terendah. Kesimpulannya, diet T2 (25% daun L. leucocephala diet) dan T6 (50% daun M. esculenta diet) adalah sesuai untuk pengunaan kambing disebabkan penambahbaikan di dalam produksi asid meruap, produksi derivative purin, penyerapan purin mikrob dan bekalan N mikrob dengan populasi rumen mikrob yang sederhana.

## ACKNOWLEDGEMENTS

Here, I would like to express my deepest gratitude to my supervisor, Dr. Anjas Asmara @ Ab. Hadi b. Samsudin, whose selfless time and care were sometimes all that kept me going. Thank you for your guidance, patience, knowledge, encouragement and support in completing my project and thesis. Extended gratitude goes to my co-supervisor, Prof. Razak b. Alimon, who had shared with me his knowledge as well as ideas in executing my experiment.

My sincerest thanks to the lecturer and staff of Animal Science Department for the assistance and providing facilities needed in completing my laboratory analyses. I would also want to express my deepest thanks to Dr. Faseleh Jahromi of Animal Production Laboratory in Tropical Agricultural Institute, who has shared his knowledge and facilitate in the microbial quantification analysis by using real-time PCR. Special thanks on Agro-Biotechnology Institute for providing me the instruments needed in completing my quantification assessment. Not to forget, I am indebted to my colleagues who help me through thick and thin. It is indeed a fun learning environment which I would missed, greatly.

For my beloved parents, Dr. Harun b. Ongah and Ruhaidah bt. Yahya, I want to thank both of you from the bottom of my heart for your patience and supports in my decision to further my study. I am grateful enough to have both of you as my parents. My sincerest thanks to my husband, Muhammad Afiq Khalit for your support towards the completion of my thesis. Lastly, I offer my regards and blessings to all of those who supported me in any respect during the completion of the project. I certify that a Thesis Examination Committee has met on 28 December 2015 to conduct the final examination of Nur Liyana Akmal bt Harun on her thesis entitled "Effects of Substituting Rice Straw with Different Levels of *Leucaena leucocephala* (Lam.) de Wit and *Manihot esculenta* Crantz Leaves on Rumen Fermentation Characteristics and Microbial Population in Goats" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

# **Dahlan bin Ismail, PhD** Professor Faculty of Agriculture

Universiti Putra Malaysia (Chairman)

# Liang Juan Boo, PhD

Associate Professor Institute of Tropical Agriculture Universiti Putra Malaysia (Internal Examiner)

### Wan Khatijah binti Embong, PhD Professor

University of Malaya Malaysia (External Examiner)



**ZULKARNAIN ZAINAL, PhD** Professor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date: 24 March 2016

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Anjas Asmara @ Ab. Hadi Bin Samsudin, PhD Senior Lecturer Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Abdul Razak Bin Alimon, PhD Professor Faculty of Agriculture Universiti Putra Malaysia (Member)

# BUJANG KIM HUAT, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

## **Declaration by graduate student**

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature:

Date:

Name and Matric No.: Nur Liyana Akmal binti Harun ,GS35571

# **Declaration by Members of Supervisory Committee**

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in Rule 41 in Rules 2003 (Revision 2012-2013) were adhered to.

Name of Chairman of Supervisory	Signature:	
	Name of	
Supervisory	Chairman of	
Supervisory	Supervisory	
Committee: Dr.Anjas Asmara @ Ab. Hadi Bin Samsudin	Committee:	Dr.Anjas Asmara @ Ab. Hadi Bin Samsudin
Signature:	Signature:	

Name of Member of Supervisory Committee:

Professor Dr. Abdul Razak Bin Alimon

# TABLE OF CONTENTS

			Page
	TRACT <i>TRAK</i>	,	i iii
		EDGEMENTS	III V
	ROVAL		vi
DEC	LARAT	TION	viii
	Г ОГ ТА		xiii
LIST	r of fi	GURES	xiv
CHA	PTER		
1	INT	RODUCTION	1
_			
2		ERATURE REVIEW	4
	2.1 2.2	Goat nutritions in the tropics	4 4
	2.2	Leguminous forages as animal feed 2.2.1 Leucaena leucocephala	4 5
		2.2.1 Leucaena reucocepnaia 2.2.2 Manihot esculenta	5
	2.3	Nutritional quality of <i>L. leucocephala</i> and <i>M. esculenta</i>	6
	2.4	Presence of anti-nutritive factor in <i>L. leucocephala</i> and <i>M.</i>	6
		esculenta	
		2.4.1 Mimosine	7
		2.4.2 Hydrogen cyanide	8
	2.5	2.4.3 Tannin	9
	2.5	Rumen fermentation characteristics as affected by	10
	2.6	<i>L. leucocephala</i> and <i>M. esculenta</i> supplementations Urinary purine derivatives and its relation to microbial N	12
	2.0	supply	12
	2.7	Microbial populations in the ruminant	14
	2.8	Conclusion	16
3	GEN	NERAL MATERIAL AND METHODS	17
	3.1	Experimental diets	17
	3.2	Proximate analyses	18
		3.2.1 Dry matter determination	18
		3.2.2 Organic matter determination	19 10
		<ul><li>3.2.3 Crude protein determination</li><li>3.2.4 Crude Fat determination</li></ul>	19 19
		3.2.5 Neutral detergent fibre determination	19 20
		3.2.6 Acid detergent fibre determination	20 20
		3.2.7 Acid detergent lignin determination	20
4	DIF ANI	E EFFECT OF SUBSTITUTING RICE STRAW WITH FERENT LEVELS OF <i>L. leucocephala</i> (Lam.) DE WI D <i>M. esculenta</i> CRANTZ LEAVES DIET ON <i>IN-VITRO</i> MEN FERMENTATION CHARACTERISTICS O	Г Э

X

C

GOATS

4.1	Introdu	uction	22
4.2	Materi	als and method	23
	4.2.1	Animal, sample and rumen liquor sampling	23
	4.2.2	Buffer and minerals solutions preparations	23
	4.2.3	Preparation of media	23
	4.2.4	Preparation of the incubation syringes	23
	4.2.5	Incubation procedure	24
	4.2.6	Gas production determinations	24
	4.2.7	In-vitro dry matter digestibility (IVDMD)	25
	4.2.8	pH determination	25
	4.2.9	Rumen ammonia determination	25
	4.2.10	Volatile fatty acid determination	26
	4.2.11		26
4.3	Result	S	26
	4.3.1	Rumen pH	26
	4.3.2	Rumen ammonia nitrogen	26
	4.3.3	Volatile fatty acid	26
	4.3.4	Total <i>In</i> -vitro gas production	27
	4.3.5	In-vitro dry matter digestibility	27
4.4	Discus	sions	29
	4.4.1	Rumen pH	29
	4.4.2	Rumen ammonia nitrogen	29
	4.4.3	Volatile fatty acid	30
	4.4.4	Total gas production and <i>in-vitro</i> dry matter digestibility	30
4.5	Conclu		31
		CT OF SUBSTITUTING RICE STRAW WITH	32
		T LEVELS OF L. leucocephala (Lam.) DE WIT	
		ulenta CRANTZ LEAVES DIET ON RUMEN	
		ATION CHARACTERISTICS AND URINARY	
		ERIVATIVES OF GOATS	22
5.1			32
5.2		al and methods	32
	5.2.1	Animal, management and facilities	32
	5.2.2		32
	5.2.3	Sampling technique and procedures	32
	5.2.4	Rumen pH determination	33
	5.2.5	Ammonia-N determination	33
	5.2.6	VFA determination	33
	5.2.7	Urinary purine derivatives determination	33
	5.2.8	Microbial purine absorption and microbial N supply determination	34
5.3	Statisti	ical analyses	34
5.4			34
	5.4.1	Rumen pH	34
	5.4.2	Rumen ammonia-N	35
	5.4.3	Rumen volatile fatty acid	35
	5.4.4	Urinary purine derivatives determinations	37
5.5			39
	5.5.1	Rumen pH	39

		5.5.2 Rumen ammonia-N	39
		5.5.3 Volatile fatty acid	40
		5.5.4 Urinary purine derivatives	41
	5.6	Conclusion	43
6	EFF	ECT OF SUBSTITUTING RICE STRAW WITH	44
	DIF	FERENT LEVEL OF <i>L. leucocephala</i> (Lam.) DE WIT and	
		sculenta CRANTZ LEAVES DIETS ON RUMEN	
	MIC	CROBIAL POPULATION OF GOATS	
	6.1	Introductions	44
	6.2	Materials and method	45
		6.2.1 Preparation of rumen sample	45
		6.2.2 Deoxyribonucleic acid (DNA) extractions	45
		6.2.3 Standard curve preparation	45
		6.2.4 Real time PCR assay	46
		6.2.5 Statistical analysis	46
	6.3	Results	48
		6.3.1 Rumen microbial population	48
	6.4	Discussions	50
	6.5	Conclusion	51
7	GEN	NERAL DISCUSSION	52
8	COI	NCLUSION	55
REF	ERENC	ES	56
APP	ENDIC	ES	77
BIO	DATA (	DF STUDENT	81
LIST	OF PU	BLICATIONS	82

xii

 $[\mathbf{G}]$ 

# LIST OF TABLES

Table		Page
2.1	Chemical compositions of L. leucocephala and M. esculenta foliage	6
3.1	Treatments diet	17
3.2	Nutrient composition (% of dry matter) of the diet components	17
3.3	Nutrient composition (% of dry matter) of the treatment diets	18
4.1	<i>In-vitro</i> gas production, dry matter digestibility and fermentation profile of the treatment diets containing different level of <i>L. leucocephala</i> and <i>M. esculenta</i> leaves	28
5.1	The effects of substitution rice straw with different level of <i>L. leucocephala</i> and <i>M. esculenta</i> leaves on rumen fermentation profile of goats	36
5.2	The effects of substitution rice straw with different level of <i>L. leucocephala</i> and <i>M. esculenta</i> leaves on urinary purine derivatives, microbial purine absorption and microbial N supply	38
6.1	Primers used for real-time PCR assay to target total bacteria, total protozoa, methanogens, <i>R. albus, R. flavefaciens</i> and <i>F. succinogenes</i> .	47
6.2	The effects of substitution rice straw with different level of L. leucocephala and M. esculenta leaves on rumen microbial population in goats (log10 copy no. per ml DNA extract)	49

# LIST OF FIGURES

Figure		Page
2.1	Mimosine transformation into dihydroxypyridine by rumen microorganism	7
2.2	The mechanism of hydrogen cyanide formation	9
2.3	Tannin classifications	10
2.4	Purine nucleotides degradation and purine derivatives formation	13
4.1	In-vitro gas production technique	24

6

#### **CHAPTER 1**

## **INTRODUCTION**

In Malaysia, small ruminant subsectors plays inconsiderable role in the livestock industry. Statistics shows that Malaysia self-sufficiency level for beef and mutton were increased in 4.7% over the past five years (DVS, 2014) with average 0.68 kg per capita consumption in Peninsular Malaysia, alone. Currently, market niches for small ruminant product are developing as they did not only cater for religious purposes but also from middle-east restaurant. However, the increasing awareness in Malaysian population regarding healthy lifestyle with high income level makes the never ending demand.

A lot of measures have been taken by the government and non-government agencies to further improve the industry especially the small scale farmers. These include campaign, policy and allocations aiming at increasing the domestic supply in the industry. Nevertheless, the key limiting factors that discourage the small scale farmer to further expand their livestock project was feeding cost. In the times of need, they would rely on concentrates to feed the animals especially during rainy season and thus increasing the cost of feed. Low nutritional value of locally found grass and rice straw were the other options of feeding the animals.

In improving nutritional value of animal feed, locally found legumes forage shows high potential to be used in the animal feed ration. Leguminous forage such as *Leucaena leucocephala* and *Manihot esculenta* are widely known and available abundantly in tropical country and traditionally has been used as livestock feed due to its high nutritional values. The usage of these forages in the animal feed shows auspicious result in growth rate, dry matter (DM) digestibility (Adejumo and Ademosun, 1991), DM intake (Srivastava and Sharma, 1997) for leucaena and DM intake, nutrient digestibility and weight gain (Hue *et al.*, 2010; Phengvichith and Ledin, 2007; Chunjula *et al.*, 2004) for *M. esculenta* regardless of feeding forms and methods.

Most of the plants have a mechanism in which to defense themselves against predators. Aside from physical forms like having thorns, there is secondary compound in aiding a defense mechanism from predators like insects. These secondary compounds like mimosine in *L. leucocephala* and hydrogen cyanide in *M. esculenta* affects the nutritive value of forages and animal consumed them (Aganga and Tshwenyane, 2003; Oni *et al.*, 2010). These forages are naturally high in crude protein (CP) where the CP content are 22.5 and 24 g/kg DM, respectively (Speedy and Pugliese, 1992). The consumption of these high quality forages with low quality straw may enhance their diet regimes quality despite the presence of anti-nutritive factors which may limits the usage of the forages as well as decrease their nutritional value as sole feed. Thus, the consumption of high protein forages may increase the microbial protein synthesis as more N is available for the microbes to utilize for their growth.

Foregut fermenter like ruminant was classified by their digestion fermentation site, pregastric fermentation chamber. In this chamber, a vast population of microorganism coexisted together with the host animal where they are involved in symbiotic relationship in the feed digestion processes. With the help of microbial population in the rumen, carbon dioxide, hydrogen, methane gas, and short-chain fatty acid such as acetate, propionate and butyrate were produced (Stevens and Hume, 1998). These short-chain fatty acids are the sourced of energy used by the host animal (Godoy-Vitorino *et al.*, 2012). Most of the studies were focused on rumen fermentation product in relation to diets, species, breed, age, sex and physiological status of the animal (Sokerya *et al.*, 2010; Oni *et al.*, 2010; Kang *et al.*, 2012) regardless of rumen microorganism involved.

Rumen microbial population is a stable and dynamic ecosystem where it change considerably through a set of constrain regardless of natural environment or feed-associated constraint such as the presence of anti-nutritive compounds (Kamra, 2005). In the rumen fed with plant contains secondary compound, some microbes may thrive and some may limits their growth due to its anti-microbial activity. Extensive research has been done on the response of anti-nutritive compound regardless of its form towards animal growth performance, nutrient digestibility and fermentation product (Yami *et al.*, 2000; Oduguwa *et al.*, 2013; Tan *et al.*, 2011). However, the respond of feeding plant-containing anti-nutritive compound on the rumen microorganism were still unclear.

Utilisations of rice straw in the tropical region are not uncommon practices among farmers. The availability throughout the year and can be easily incorporated in the animal feed makes this crop residue an easy option compared with green forages. Fermentation rice straw in rumen would result in higher methane production due to an increase in retention time in the gastrointestinal tract as rice straw is highly fibrous and low in digestibility. Hence, the combination use of rice straw as an inexpensive roughage source with *L. leucocephala* and *M. esculenta* foliage as N source may activates microbial fermentation in straw which leads to the final result being improve rumen fermentation condition as well as rumen microbial population without increasing N losses to the environment. The combination of rice straw and high quality forage, *L.leucocephala* and *M. esculenta* offers on farm practicality with high quality of diets being offered to the animals. However, an optimum level of combination must be explored so that the diet can be fed to the animal without deteriorating the rumen function of the animal and environment.

Therefore, this study were conducted to test the hypothesis that such substitution of rice straw with certain level of *L. leucocephala* and *M. eculenta* leaves diet may improve rumen fermentation condition and microbial population as such that would increase in the microbial N supply. Hence, the objectives of this study were;

- 1. to investigate the *in-vitro* fermentation characteristics on different level of *L*. *leucocephala* and *M. esculenta* leaves diet,
- 2. to investigates the effect of substituting rice straw with different level of *L*. *leucocephala* and *M. esculenta* on *in-vivo* rumen fermentation characteristics and urinary purine derivatives of the goats,
- 3. to quantify the rumen microbial community of goats as affected by substitution of rice straw with different level of *L. leucocephala* and *M. esculenta* leaves diets using real-time PCR.



#### REFERENCES

- Abecia, L., Fondevila, M., Rodríguez-Romero, N., Martinez, G., and Yáñez-Ruiz, D.R. (2013). Comparative study of fermentation and methanogen community structure in the digestive tract of goats and rabbits. *Journal of Animal Physiology And Animal Nutrition*. 97(s1): 80-88.
- Adegbola, T.A. and Okonkwo, J.C. (2002). Nutrient intake, digestibility and growth rate of rabbits fed varying levels of cassava leaf meal. *Nigerian Journal of Animal Production*. 29(1): 21-26.
- Adejumo, J.O. and Ademosun, A.A. (1991). Utilization of leucaena as supplement for growing dwarf sheep and goats. *Journal of Small Ruminant Research*. 4: 75-82.
- Adesope, A.I. and Ojo, V.O.A. (2011). The effect of Leucaena leaf supplementation to maize residues on village goat performance. *Journal of Animal and Plant Sciences*. 10(2): 1276-1282.
- Aganga, A.A. and Tshwenyane, S.O. (2003). Feeding Values and Anti-Nutritive Factors of Forage Tree Legumes. *Pakistan Journal of Nutrition*. 2: 170–177.
- Al-Dehneh, A., Pierzynowski, S.G., Smuts, M., Sahlu, T., and Fernandez, J.M. (1994). Blood metabolite and regulatory hormone concentrations and response to metabolic challenges during the infusion of mimosine and 2, 3 dihydroxypyridine in alpine goats. *Journal of Animal Science*. 72(2): 415-420.
- Aletor, V.A. and Adeogun, O.A. (1995). Nutrients and anti- nutrient components of some tropical leafy vegetables. *Food Chemistry Journal*. 53(4): 375-379.
- Allison, M.J., Hammond, A.C. and Jones, R.J. (1990). Detection of ruminal bacteria that degrade toxic dihydroxypyridine compounds produced from mimosine. *Journal of Applied and Environmental Microbiology*. 56: 590–594.
- Alvarez, F.J., Wilson, A., and Preston, T.R. (1977). *Leucaena leucocephala* as a combined source of protein and roughage for steers fattened on molasses/urea. *Journal of Tropical Animal Health and Production*. 2: 288-291.
- Ammar, H. (2002). Compositión química, digestibilidad y cinética de fermentacion ruminal *in-vitro* de arbustos. Ph.D. thesis, Universidad de León, Spain.
- AOAC. (1990). *Official methods of analysis* (15th ed.). Arlington, VA: Association of Official Analytical Chemists.
- Ash, A.J. (1990). The effect of supplementation with leaves from legumes trees *Sesbania grandiflora Abizia chinensis* and *Gliricidia sepium* on the intake and digestibility of guinea grass hay by goats. *Journal of Animal Feed Science* and *Technology*. 28: 225–232.

- Ash, R.W. (1959). Inhibition and excitation of reticulo-rumen contractions following the introduction of acids into the rumen and abomasum. *Journal* of Physiology. 147: 58.
- Atawodi, S.E., Mari, D., Atawodi, J.C. and Yahaya, Y. (2008). Assessment of Leucaena leucocephala leaves as feed supplement in laying hens. African Journal of Biotechnology. 7: 317-321.
- Aung, A. Meulen, U., Gessler, F. and Böhnel, H. (2011). Isolation of Mimosine Degrading Bacteria from Rumen Juice and Mass Production by Göttingen Bioreactor Technology. *Journal of Agricultural Science and Technology*. 1: 764-772.
- Bach, A., Calsamiglia, S., and Stern, M.D. (2005). Nitrogen metabolism in the rumen. *Journal of Dairy Science*. 88: E9-E21.
- Balcells, J., Guada, J.A., Castrillo, C. and Gasa, J. (1993). Rumen digestion and urinary excretion of purine derivatives in response to urea supplementation of sodium-treated straw fed to sheep. *British Journal of Nutrition*. 69: 721-732.
- Balogun, R.O., Jones R.J. and Holmes, J.H.G. (1998). Digestibility of some tropical browse species varying in tannin content. *Journal of Animal Feed Science and Technology*. 76: 77-88.
- Bannink, A. (2007). Modelling volatile fatty acid dynamics and rumen function in lactating cows. PhD Thesis, Wageningen University., the Netherlands. pp. 262.
- Barry, T.N. and Manley, T.R. (1984). The role of condensed tannins in the nutritional value of *Lotus peduculatus* for sheep 2. Quantitative digestion of carbohydrates and protein. *British Journal of Nutrition*. 51: 493.
- Belanche, A., Abecia, L., Holtrop, G., Guada, J.A., Castrillo, C., de la Fuente, G., and Balcells, J. (2011). Study of the effect of presence or absence of protozoa on rumen fermentation and microbial protein contribution to the chyme. *Journal* of Animal Science. 89(12): 4163-4174.
- Belanche, A., de la Fuente, G., and Newbold, C.J. (2014). Study of methanogen communities associated with different rumen protozoal populations. *FEMS Microbiology Ecology*. 90(3): 663-677.
- Belenguer, A., Yanez, D., Balcells, J., Ozdemir Baber, N.H. and Gonzalez Ronquillo, M.N. (2002). Urinary excretion of purine derivatives and prediction of rumen microbial outflow in goats. *Livestock Production Science Journal*. 77: 127– 135.
- Bergman, E.N. (1990). Energy contributions of volatile fatty-acids from the gastrointestinal tract in various species. *Physiological Review*. 70: 567-590.

- Bhatnagar, R., Kataria, M. and Verna, S.V.S. (1996). Effect of dietary Leucaena leaf meal on the performance and egg characteristics in white leghorn hens. *Journal of Animal Science*. 66: 1291-1294.
- Bhatta, R., Saravanan, M., Baruah, L., Sampath, K.T. and Prasad, C.S. (2013). Effect of plant secondary compounds on in vitro methane, ammonia production and ruminal protozoa population. *Journal of Applied Microbiology*. 115(2): 455-465.
- Blummel, M. and Becker, K. (1997). The degradability characteristics of fifty-four roughages and roughage neutral detergent fibers as described by *in-vitro* gas production and their relationship to voluntary feed intake. *British Journal of Nutrition*. 77: 757-768.
- Bonsi, M.L.K., Osuji, P.O., Tuah, A.K., and Umunna, N.N. (1995). Intake, digestibility, nitrogen balance and certain rumen characteristics of Ethiopian Menz sheep fed teff straw supplemented with cotton seed cake, dry sesbania, dry leucaena or fresh leucaena. *Journal of Agroforestry Systems*. 31(3): 243-256.
- Brewbaker, J.L. and Hylin, J.W. (1965). Variation in mimosine content among leucaena species and related mimosaceae. *Crop Science Journal*. 5: 348–349.
- Broderick, G.A. and Balthrop, J.E., (1979). Chemical inhibition of amino acid deamination by ruminal microbes *in vitro*. *Journal of Animal Science*. 49: 1101-1111.
- Broderick, G.A. (2003). Effects of varying dietary protein and energy levels on the production of lactating dairy cows. *Journal of Dairy Science*. 86: 1370–1381.
- Broderick, G.A. and Muck, R.E. (2009). Effect of alfalfa silage storage structure and rumen protected methionine on production in lactating dairy cows. *Journal of Dairy Science*. 92: 1281–1289.

Buitrago, A.(1990). La yuca em la alimentación animal. CIAT.

- Calsamiglia, S., Cardozo, P.W., Ferret, A., and Bach, A. (2008). Changes in rumen microbial fermentation are due to a combined effect of type of diet and pH. *Journal of Animal Science*. 86(3): 702-711.
- Canbolat, O., Ozkan, C.O., and Kamalak, A. (2007). Effects of NaOH treatment on condensed tannin contents and gas production kinetics of tree leaves. *Journal of Animal Feed Science And Technology*. 138(2): 189-194.
- Carvalho, J.L.H. de (1990). A mandioca: raiz e parte aérea na alimentação animal. Brasília: EMBRAPA, CPAC.
- Chanchay, N. and Poosaran, N. (2009). The reduction of mimosine and tannin contents in leaves of *Leucaena leucocephala*. *Asian Journal of Food and Agro-Industry*. 2(Special Issue): S137-S144.

- Chandrasekharaiah, M., Thulasi, A., Suresh, K. P., and Sampath, K. T. (2011). Rumen degradable nitrogen requirements for optimum microbial protein synthesis and nutrient utilization in sheep fed on finger millet straw (*Eleucine coracana*) based diet. Animal Feed Science and Technology, 163(2): 130-135.
- Chanjula, P., Wanapat, M., Wachirapakorn, C. and Rowlinson, P. (2004). Effect of level of cassava hay and urea-treated rice straw on rumen ecology and digestibility in swamp buffaloes. *Asian-Australasian Journal of Animal Science*. 17: 663-669.
- Chen, X.B., Hovell, F.D.DeB., Ørskov, E.R. and Brown, D.S. (1990). Excretion of purine derivatives by ruminants: effect of exogenous nucleic acid supply on purine derivatives excretion in sheep. *British Journal of Nutrition*. 63: 131-142.
- Chen, X.B. and Gomez, M.J. (1995). Estimation of microbial protein supply to sheep and cattle based on urinary excretion of purine derivatives - An overview of the technical details. Occasional Publication 1992, International Feed Resources Unit, Rowette Research Institute, Aberdeen, UK.
- Chen, X.B., Samaraweera, L., Kyle, D.J., Ørskov, E.R. and Abeygunawardene, H. (1996). Urinary excretion of purine derivatives and tissue xanthine oxidase (EC 1.2. 3.2) activity in buffaloes (*Bubalis bubalis*) with special reference to differences between buffaloes and Bos taurus cattle. *British Journal of Nutrition*. 75(03): 397-407.
- Cheng, K.J., Forsberg, C.W., Minato, H. and Costerton, J.W. (1991) Microbial ecology and physiology of feed degradation within the rumen. In T. Tsuda, Y. Sasaki, and R. Kawashima. *Physiological Aspects of Digestion and Metabolism in Ruminants.* (pp. 595–624). Toronto: Academic Press.
- Cieslak, A., Szumacher-Strabel, M., Stochmal, A. and Oleszek, W. (2013). Plant components with specific activities against rumen methanogens. *Animal : Journal Of Animal Bioscience*. 7(s2): 253-265.
- D'Mello, J.P.E. and Taplin, D.E. (1978). *Leucaena leucocephala* in poultry diets for the tropics. World Review of Animal Production. 24: 41-47.
- D'Mello, J.P.E., Acamovic, T. and Walker, A.G. (1987). Evaluation of leuceana leaf meal for broiler growth and pigmentation. *Journal* of *Tropical Agriculture* (Trinidad). 64: 33-35.
- D'Mello, J.P.F. (1992). Chemical constraints to the use of tropical legumes in animal nutrition. *Journal of Animal Feed Science and Technology*. 38: 237–261.
- De Boever, J. L., Iantcheva, N., Cottyn, B. G., De Campeneere, S., Fiems, L. O., and Boucqué, C. V. (1998). Microbial protein synthesis in growing-finishing bulls estimated from the urinary excretion of purine derivatives. *Animal Feed Science and Technology*, 75(2): 93-109.

- Demeyer, D.I. and Van Nevel, C.J. (1975). Methanogenesis, an integrated part of carbohydrate fermentation, and its control. In I. W. Mcdonald and A. C. I. Warner. *Digestion and Metabolism in the Ruminant*. (pp. 366- 382). New South Wales, Australia: The University of New England Publishing Unit.
- Denman, S.E., and McSweeney, C.S. (2006). Development of a real-time PCR assay for monitoring anaerobic fungal and cellulolytic bacterial populations within the rumen. FEMS Microbiology Ecology. 58(3): 572-582.
- Department of Veterinary Services. (2014). Livestock statistics. Malaysia : Self-Sufficiency In Livestock Products (%), 2004-2013. Retrieved from http://www.dvs.gov.my/documents/10157/69177f95-9ec4-49d38d27342b4d1 8bbe. [Accessed on December 12, 2014].
- Dipu, M.T., George, S.K., Singh, P., Verma, A.K., and Mehra, U.R. (2006). Measurement of microbial protein supply in Murrah buffaloes (Bubalus bubalis) using urinary purine derivatives excretion and PDC index. Asian Australasian *Journal of Animal Sciences*. 19(3): 347.
- Do, H.Q., Son, V.V., Hang, B.P.T., Tri, V.C. and Preston, T.R. (2003). Effect of supplementation of ammoniated rice straw with cassava leaves or grass on intake, digestibility and N retention by goats. In: Proceedings of National Workshop-Seminar on Sustainable Livestock Production on Local Feed Resources, March, 25–27. Hue, Vietnam.
- Duncan, A.J. and Young, S.A. (2002). Can goats learn about foods through conditioned food aversions and preferences when multiple food options are simultaneously available? *Journal of Animal Science*. 80: 2091–2098.
- Dung, N.T., Mui, N.T. and Ledin, I. (2005). Effect of replacing a commercial concentrate with cassava hay (*Manihot esculenta Crantz*) on the performance of growing goats. *Journal of Animal Feed Science and Technology*. 119: 271– 281.
- Dziba, I.E., Scogings, P.F., Gordon, I.J. and Raats, J.G. (2003a). Effects of season and breed on browse species intake rates and diet selection by goats in the False Thornveld of the Eastern Cape, South Africa. *Journal of Small Ruminant Research*. 47: 17–30.
- Dziba, I.E., Scogings, P.F., Gordon, I.J. and Raats, J.G. (2003b). The feeding height preferences of two goat breeds fed *Grewia occidentalis L. (Tiliacecae)* in the Eastern Cape, South Africa. *Journal of Small Ruminant Research.* 47: 31–38.
- Eggum, B.O. (1970). The protein quality of cassava leaves. British Journal of Nutrition. 24(03): 761-768.
- Erfle, J.D., Boila, R.J., Teather, R.M., Mahadevan, S. and Sauer, F.D. (1982). Effect of pH on fermentation characteristics and protein degradation by rumen microorganisms in vitro. *Journal of Dairy Science*. 65(8): 1457-1464.

- Estrada-Liévano, J. M., Sandoval-Castro, C. A., Ramirez Avilés, L., and Capetillo-Leal, C. M. (2009). *In-vitro* fermentation efficiency of mixtures of *Cynodon nlemfuensis, Leucaena leucocephala* and two energy sources (maize or sugar cane molasses). *Tropical and Subtropical Agroecosystems*, 10(3): 497-503.
- Fedele, V., Claps, S., Rubino, R., Calandrelli, M. and Pilla, A.M. (2002). Effect of freechoice and traditional feeding systems on goat feeding behavior and intake. *Livestock Production Science Journal*. 74: 19–31.
- Filípek, J. and Dvořák, R. (2009). Determination of the volatile fatty acid content in the rumen liquid: comparison of gas chromatography and capillary isotachophoresis. *Journal Acta Veterinaria Brno*. 78: 627-633.
- Firkins, J.L., Yu, Z., and Morrison, M. (2007). Ruminal Nitrogen Metabolism: Perspectives for Integration of Microbiology and Nutrition for Dairy 1, 2. Journal of Dairy Science. 90: E1-E16.
- Fisher, D.S., Mayland, H.F. and Burns, J.C. (1999). Variation in ruminants' preference for tall fescue hays cut either at sundown or at sunup. *Journal of Animal Science*. 77: 762–768.
- Forsberg, C.W., Cheng, K.J. and White, B.A. (1997). Polysaccharide degradation in the rumen and large intestine. In *Gastrointestinal microbiology*. pp. 319-379. Springer US.
- Francis, G., Kerem, Z., Makkar, H.P., and Becker, K. (2002). The biological action of saponins in animal systems: a review. *British Journal of Nutrition*. 88(06): 587-605.
- Franzolin, R. and Dehority, B.A. (1996). Effect of prolonged high-concentrate feeding on ruminal protozoa concentrations. *Journal of Animal Science*. 74(11): 2803-2809.
- Franzolin, R., and Alves, T.C. (2010). The Ruminal Physiology in Buffalo Compared with Cattle. *Revista Veterinaria*. 21(1).
- Frutos, P., Hervas, G., Giráldez, F.J. and Mantecón, A.R. (2004). Review. Tannins and ruminant nutrition. *Spanish Journal of Agricultural Research*. 2(2): 191-202.
- Fujihara, T., Iwakuni, M., Kyle, D.J., Ørskov E.R.(1999). The effect of feeding level and frequency on the microbial protein synthesis in the rumen of sheep. *Protein Metabolism and Nutrition (Book of abstracts of the VIII<sup>th</sup> International Symposium on Protein Metabolism and Nutrition)*. p.54.
- Garcia, G. W., Ferguson, T. U., Neckles, F. A., and Archibald, K. A. E. (1996). The nutritive value and forage productivity of *Leucaena leucocephala*. *Animal Feed Science and Technology*, 60(1):29-41.
- Geihauser, T., Linhart, N., Neidl, A. and Reimann, A. (2012). Factors associated with ruminal pH at herd level. *Journal of Dairy Science*. 95(8): 4556-4567.

- Giner-Chávez, B.I. (1996). Condensed tannins in tropical forages. Doctoral Thesis. Cornell University. Ithaca, NY, USA.
- Gleadow, R.M. and Woodrow, I.E. (2002). Mini-Review: constraints on effectiveness of cyanogenic glycosides in herbivore defense. *Journal of Chemical Ecology*. 28(7): 1301-1313.
- Goncalves, A.L., Lana, R.P., Rodrigues, M.T., Viera, R.A.M., Queiroz, A.C. and Henrique, D.S. (2001). Nictemeral pattern of ruminal pH and feeding behavior of dairy goats fed diets with different roughage to concentrate ratio. *Revista Brasileira de Zootecnia*, 30: 1886–1892.
- Gupta, H.K., and Atreja, P.P. (1998). Influence of gradual adaptation of cattle to *Leucaena leucocephala* leaf meal on biodegradation of mimosine and 3-hydroxy-4 (1H)-pyridone (3, 4 DHP) in rumen, their levels in blood, fate and influence of absorbed DHP on thyroid hormones and liver enzymes. *Journal of Animal Feed Science and Technology*. 74(1): 29-43.
- Guyader, J., Eugène, M., Nozière, P., Morgavi, D.P., Doreau, M. and Martin, C. (2014). Influence of rumen protozoa on methane emissions in ruminants: A meta-analysis approach. *Animal Journal*. 8: 1816-1825.
- Hagerman, A.E., Robbins, C.T., Weerasuriya, Y., Wilson, T.C., and McArthur, C. (1992). Tannin chemistry in relation to digestion. *Journal of Range Management*. 57-62.
- Hang, D.T. and Preston, T.R. (2005). The effects of simple processing methods of cassava leaves on HCN content and intake by growing pigs. *Livestock Research for Rural Development Journal*. 17(99).
- Harborne, J.B. (1989). Biosynthesis and function of anti-nutritional factors in plants. *Aspects of Applied Biology Journal*. 19: 21–28.
- Hariadi, B.T. and Santoso, B. (2010). Evaluation of tropical plants containing tannin on *in-vitro* methanogenesis and fermentation parameters using rumen fluid. *Journal of the Science of Food and Agriculture*. 90: 456–461.
- Hegarty, M.P., Schinkel, P.G. and Court, R.D. (1964). Reaction of sheep to the consumption of *Leucaena glauca* (Benth) and its toxic principle mimosine. *Australian Journal of Agricultural Research*. 15: 153-167.
- Hegarty, M.P., Court, R.D., Christie, G.S. and Lee, C.P. (1976). Mimosine in *Leucaena leucocephala* is metabolised to a goitrogen in ruminants. *Australian Veterinary Journal*. 52: 490.
- Hegarty, M.P. (1977) in "Leucaena: Promising forage and tree crop for the tropics". Editor: F.R. Ruskin, pp. 31. National Academy of Sciences, Washington, D.C.

- Hernandez, P., Salema, A.Z.M, López, S., Sun, X.Z., Rojo, R., Camachoe, L.M., Elghandour, M.M.Y., and Gonzalez-Ronquillo, M. (2014). Influence of *Salix babylonica* and *Leucaena leucocephala* leaf extracts on ruminal fermentation characteristics, urinary purine derivative excretion and microbial protein synthesis of lambs. *Journal of Livestock Science*. 163: 80–84.
- Hervas, G., Frutos, P., Giraldez, F.J., Mantecon, A.R. and Pino, M.C.A.D. (2003). Effect of different doses of quebracho tannins extract on rumen fermentation in ewes. *Journal of Animal Feed Science and Technology*. 109: 65–78.
- Hoover, W.H. (1986). Chemical factors involved in ruminal fiber digestion. *Journal of Dairy Science*. 69: 2755.
- Hoover, W.H. and Stokes, S.R. (1991). Balancing carbohydrates and proteins for optimum rumen microbial yield. *Journal of Dairy Science*. 74: 3630.
- Hue, K.T., Van, D.T.T. and Ledin, I. (2008). Effect of supplementing urea treated rice straw and molasses with different forage species on the performance of lambs. *Journal of Small Ruminant Research* 78: 134–143.
- Hung, L.V., Wanapat, M. and Cherdthong, A. (2013). Effects of leucaena leaf pellet on bacterial diversity and microbial protein synthesis in swamp buffalo fed on rice straw. *Livestock Science Journal*. 151(2): 188-197.
- Hutjens, M.F. (2003). Hoard's Dairyman Feeding Guide, Second Edition. W.D. Hoards & Sons Company; Fort Atkinson, WI.
- Ichinohe, T., and Fujihara, T. (2008). Adaptive changes in microbial synthesis and nitrogen balance with progressing dietary feeding periods in sheep fed diets differing in their ruminal degradation synchronicity between nitrogen and organic matter. *Animal Science Journal*, 79(3): 322-331.
- Isah, O.A., Oguntuyo, S.A., Dawodu, R.O., Diya, O.O., Afolabi, M.O., and Omoniyi, L.A. (2013). Feed utilization, rumen parameters, and microbial profile of goats fed different tropical browse plants with *Pennisetum purpureum* as basal diet. *Pacific Journal of Science and Technology*. 14(1): 397-405.
- Jetana, T., Thongruay, S., Uswang, S. and Hengtrakulsin, R. (2012). A comparative study on mimosine, 3, 4-dihydroxy pyridone (3, 4-DHP) and 2, 3-dihydroxy pyridone (2, 3-DHP), purine derivatives (PD) excretion in the urine, thyroid hormone and blood metabolites profiles of Thai swamp buffalo (Bubalus bubalis) and Murrah buffalo (Bubalus bubalis). *Tropical Animal Health and Production*. 44(4): 887-897.
- Jones, R.J. (1979). The value of *Leucaena leucocephala* as a feed for ruminants in the tropics. *World Animal Review*. 31(1): 3-2.

- Jones, G. A., McAllister, T. A., Muir, A. D., and Cheng, K. J. (1994). Effects of sainfoin (*Onobrychis viciifolia Scop.*) condensed tannins on growth and proteolysis by four strains of ruminal bacteria. *Applied and Environmental Microbiology*, 60(4): 1374-1378.
- Joomjantha, S. and Wanapat, M. (2007). Effect of intercropping of cassava cultivation on biomass yield, and chemical compositions. Tropical Feed Resources Research and Development Center, Department of Animal Science, Faculty of Agriculture, Khon Kaen University: Khon Kaen, Thailand.
- Jurgens, M.H. (1997). Animal feeding and Nutrition. 8 edition. Kendall/Hunt publishing company. Dubuque, Iowa, USA.
- Kang, S., Wanapat, M., Pakdee, P., Pilajun, R. and Cherdthong, A. (2012). Effects of energy level and Leucaena leucocephala leaf meal as a protein source on rumen fermentation efficiency and digestibility in swamp buffalo. *Journal of Animal Feed Science and Technology*. 174(3-4): 131-139.
- Karachi, M. (1998). Variation in the nutritional value of leaf and stem fractions in nineteen Leucaena lines. *Journal of Animal Feed Science and Technology*. 70: 305.
- Karda, I.W. (2007). Effects of leucaena (*L. leucocephala* cv *Tarramba*) leucaena with urea or leucaena with sucrose supplementation on intake and rhodes grass (*Chloris gayana* cv *callide*) hay fed to sheep. Ejournal Universitas Udayana. 10(2): 13.
- Kayouli, C., Demeyer, D.I., Van Nevel, C.J. and Dendooven, R. (1984). Effect of defaunation on straw digestion in sacco and on particle retention in the rumen. *Animal Feed Science and Technology*. 10(2): 165-172.
- Khang, D.N. and Wiktorsson, H. (2004). Effects of fresh cassava tops on rumen environment parameters, thyroid gland hormones and liver enzymes of local yellow cattle fed urea-treated fresh rice straw. *Journal of* Tropical *Animal Health and Production*. 36: 751–762.
- Khampa, S. and Wanapat, M. (2006). Influences of energy sources and levels supplementation on ruminal fermentation and microbial protein synthesis in dairy steers. *Pakistan Journal of Nutrition*. 5(4): 294-300.
- Khampa, S., and Wanapat, M. (2006). Supplementation Levels of Concentrate Containing High Levels of Cassava Chip on Rumen Ecology and Microbial Protein Synthesis in Cattle. *Pakistan Journal of Nutrition*, 5(6): 501-506.
- Khanbabaee, K., and van Ree, T. (2001). Tannins: classification and definition. *Natural Product Reports*. 18(6): 641-649.
- Khy, Y., Wanapat, M., Haitook, T. and Cherdthong, A. (2012). Effect of *Leucaena leucocephala* pellet (LLP) supplementation on rumen fermentation efficiency and digestibility of nutrients in swamp buffalo. *Journal of Animal and Plant Sciences*. 22(3): 564-569.

- Klieve, A.V., Ouwerkerk, D., Tuner, A. and Roberton, R. (2002). The production and storage of a fermenter-grown bacterial culture containing *Synergistes jonesii* for protecting cattle against mimosine and 3-hydroxy-4(1H)-pyridone toxicity from feeding on *Leucaena leucocephala*. *Australian Journal of Agricultural Research*. 53: 1-5.
- Koakhunthod, S., Wanapat, M., Wachirapakorn, C., Nontaso, N., Rowlinson, P. and Sornsungnern, N. In *Effect of cassava hay in high-quality feed block supplementation on milk production in lactating dairy cows*. Proceeding of International Workshop on Current Research and Development of Cassava as Animal Feeds. T. R. Preston, B. Ogle and M. Wanapat Eds. 2001.
- Koike, S. and Kobayashi, Y. (2001). Development and use of competitive PCR assays for the rumen cellulolytic bacteria: *Fibrobacter succinogenes, Ruminococcus albus* and *Ruminococcus flavefaciens. FEMS Microbiology Letters.* 204(2): 361-366.
- Kowalik, B., Skomiał, J., Pająk, J.J., Taciak, M., Majewska, M. and Bełżecki, G. (2012). Population of ciliates, rumen fermentation indicators and biochemical parameters of blood serum in heifers fed diets supplemented with yeast (*Saccharomyces cerevisiae*) preparation. *Animal Science Papers and Reports*. 30(4): 329-338.
- Krause, D.O., Denman, S.E., Mackie, R.I., Morrison, M., Rae, A.L., Attwood, G. T. and McSweeney, C.S. (2003). Opportunities to improve fiber degradation in the rumen: microbiology, ecology, and genomics. *FEMS Microbiology Reviews*. 27(5): 663-693.
- Krause, M.K. and Oetzel, G.R. (2005). Inducing subacute ruminal acidosis in dairy. *Journal of Dairy Science*. 88: 3633-3639.
- Kudo, H.K., Cheng, J., Majak, W., Hall, J.W. and Costerton, J.W., (1984). Degradation of mimosine in the rumen fluid from cattle and sheep in Canada. *Canadian Journal of Animal Science*. 64: 937–942.
- Kumar, R. (2003). Anti-nutritive factors, the potential risks of toxicity and methods to alleviate them. http://www.fao.org/docrep/003/T0632E/T0632E10.htm.
- Lana, R.P., Russell, J.B. and Van Amburgh, M.E.(1998). The role of pH in regulating ruminal methane and ammonia production. *Journal of Animal Science*.76: 2190-2196.
- Lane, D.J. (1991). 16S/23S rRNA sequencing. Nucleic Acid Techniques in Bacterial Systematics. 125-175.
- Laurent, F. and Vignon, B. (1979). Variations in urinary excretion of total nitrogen urea and allantoin in sheep and he-goats [influence of level and source of nitrogen in diet]. *Bulletin de l'Ecole Nationale Superieure d'Agronomie et des Industries Alimentaires*. 21: 115–124.

- Lebas, F., Bannelier, C., Adoukonou, J., and Djago, A.Y. (2012). Chemical Composition Of Some Raw Materials Available For Rabbit Feeding In Benin. World Rabbit Congress. 10: 581 – 584.
- Lee, H. and Lee, I. (2002). A study on the dry matter yield and nutritive values of wild Korean lespedeza (*Lespedeza stipulacea Maxim.*). *Asian–Australasian Journal* of Animal Science. 15: 396–400.
- Lettat, A. and Benchaar, C. (2013). Diet-induced alterations in total and metabolically active microbes within the rumen of dairy cows. *PloS one*. 8(4): e60978.
- Limcango-Lopez, P.D. (1997). The use of shrubs and tree fodders by non-ruminants. *In: Cassava as Animal Feed*. 61-75.
- Liu, Q., Wang, C., Huang, Y.X., Dong, K.H., Yang, W.Z., Zhang, S.L. and Wang, H. (2009). Effects of isovalerate on ruminal fermentation, urinary excretion of purine derivatives and digestibility in steers. *Journal of Animal Physiology* and Animal Nutrition. 93(6): 716-725.
- Lohan, O. P., Lall, D., Makkar, H. P. S., and Negi, S. S. (1981). Inhibition of rumen urease activity by tannins in oak leaves [India]. *Indian Journal of Animal Sciences* (India).
- Ly, V.L,. and Chinh, B.V. (1996). Animal production in sustainable agricultural systems, Agricultural Publishing House, Ha Noi, 88-91.
- Madruga, M.S. and Camara, F.S. (2000). The chemical composition of "Multimistura" as a food supplement. *Food Chemistry*. 68(1): 41-44.
- Majak, W. and Cheng, K.J. (1984). Cyanogenesis in bovine rumen fluid and pure cultures of rumen bacteria. *Journal of Animal Science*. 59: 784- 790.
- Majak, W. and Cheng, K.J. (1987). Hydrolysis of the cyanogenic glycosides amygdalin, prunasin and linamarin by ruminal microorganisms. *Canadian Journal of Animal Science*. 67: 1133-1137.
- Majak, W., McDiarmid, R.E., Hall, J.W. and Cheng, K.J. (1990). Factors that determine rates of cyanogenesis in bovine ruminal fluid in vitro. *Journal of Animal Science*. 68(6): 1648-55.
- Makkar, H.P.S., Singh, B. and Dawra, R.K. (1988). Effect of tannin rich leaves of Oak (Quercus incana) on various microbial enzyme activities of the bovine rumen. *British Journal of Nutrition*. 6: 287–296.
- Makkar, H.P.S. and Becker, K. (1996). Effect of pH, temperature and time on inactivation of tannin and possible implications in detannification studies. *Journal of Agricultural and Food Chemistry*. 44: 1291-1295.
- Males, J.R. and Purser, D.B. (1970).Relationship Between Rumen Ammonia Levels and the Microbial Population and Volatile Fatty Acid Proportions in Faunated and Defaunated Sheep. *Journal of Applied Microbiology*. 19(3): 485.

- Mangan, J.L. (1988). Nutritional effects of tannins in animal feeds. *Nutrition Abstracts and Reviews*. 1:209-231.
- Marie-Magdeleine, C., Mahieu, M., Philibert, L., Despois, P. and Archimède, H. (2010). Effect of cassava (*Manihot esculenta*) foliage on nutrition, parasite infection and growth of lambs. *Journal of Small Ruminant Research*. 93: 10 18.
- Martin, S.A. and Akin, D.E. (1998). Effect of phenolic monomers on the growth and  $\beta$ -glucosidase activity of *Bacteroides ruminicola* and on Carboxymethyl cellulase,  $\beta$ -glucosidase,  $\beta$ -xylanase activities of *Bacteroides succinogens*. *Applied and Environmental Microbiology*. 54: 3600–3604.
- Matsumoto, M., Kobayashi, T. and Itabashi, H. (1991). Effects of the absence of rumen ciliate protozoa on urinary allantoin excretion in goats. *Journal of Animal Science and Technology*. 62: 939–946.
- McAllister, T.A., Okine, E.K., Mathison, G.W. and Cheng, K.J., (1996). Dietary environmental and microbiological aspects of methane production in ruminants. *Canada Journal of Animal Science*. 76: 231-243.
- McDonald, P., Edwards, R.A. and Greenhalgh, J.F.D., (1988). Animal Nutrition, 4<sup>th</sup> ed. Longman Scientific and Technical, John Wiley & Sons, Inc., New York, p.42– 157.
- McSweeney, C.S., Palmer, B., Bunch, R. and Krause, D.O. (1999). Isolation and characterization of proteolytic runnial bacteria from sheep and goats fed the tannin-containing shrub legume *Calliandra calothyrsus. Journal of Applied Environment Microbiology*. 65: 3075–3083.
- McSweeney, C.S., Palmer, B., Krause, D.O. and Brooker, J.D. (1999). Rumen microbial ecology and physiology in sheep and goats fed a tannin-containing diet. In *ACIAR Proceedings* (pp. 140-145). ACIAR.
- McSweeney, C.S., Denman, E.S. and Mackie, R.I. (2005). Rumen Bacteria. In H.P.S. Makkar and C.S. McSweeney. *Methods in Gut Microbial Ecology for Ruminants* (edited). pp 23-37.
- McLeod, M.N. (1974). Plant tannins, their role in forage quality. *Nutrition Abstracts and Reviews*. 44: 804–815.
- Meale, S.J., Chaves, A.V., Baah, J. and McAllister, T.A. (2012). Methane Production of Different Forages in *In-vitro* Ruminal Fermentation. *Asian-Australasian Journal* of *Animal Science*. 25(1): 86-91.
- Mendoza, R.C. (1975). Herbage crude protein and digestible dry matter yield of Ipil-Ipil (*Leucaena latisilique* Cv. Peru). Hedge Rows-Animal scientific Convention of the Philippines Society of Animal Science.

- Meulen, U., Pucher, F., Szyszka, M., and El-Harith, E.A. (1984). Effects of administration of Leucaena meal on growth performance of, and mimosine accumulation in, growing chicks. *Journal* of *European Poultry Science*. 48: 41-44.
- Michalet-Doreau, B., Fernandez, I., Peyron, C., Millet, L. and Fonty, G. (2001). Fibrolytic activities and cellulolytic bacterial community structure in the solid and liquid phases of rumen contents. *Reproduction Nutrition Development*. 41(2): 187-194.
- Min, B.R., Barry, T.N., Attwood, G.T. and McNabb, W.C. (2003). The effect of condensed tannins on the nutrition and health of ruminants fed fresh temperate forages: a review. *Journal of Animal Feed Science and Technology*. 106: 3– 19.
- Miron, J., Ben-Ghedalia, D. and Morrison, M. (2001). Invited Review: Adhesion mechanisms of rumen cellulolytic bacteria. *Journal of Dairy Science*. 84: 1294-1309.
- Mole, S. and Waterman, P.G. (1987). Tannic acid and proteolytic enzymes: enzyme inhibition or substrate deprivation?. *Journal of Phytochemistry*. 26: 99-102.
- Morris, C.D. and Du Toit, L.P. (1998). The performance of Boer goats browsing Leucaena leucocephala in KwaZulu-Natal, South Africa. Tropical Grasslands. 32: 188-194.
- Mosoni, P., Martin, C., Forano, E. and Morgavi, D.P. (2011). Long-term defaunation increases the abundance of cellulolytic ruminococci and methanogens but does not affect the bacterial and methanogen diversity in the rumen of sheep. *Journal of Animal Science*. 89(3): 783-791.
- Moss, A. R., Jouany, J. P., and Newbold, J. (2000). Methane production by ruminants: its contribution to global warming. In *Annales de zootechnie*, 49(3):231-253). EDP Sciences.
- Mtenga, L.A. and Shoo, R.A. (1990). Growth rate, feed intake and feed utilization of small East African goats supplemented with *Leucaena leucocephala*. Journal of Small Ruminant Research. 3: 9-18.
- Mueller-Harvey, I. and McAllan, A.B. (1992). Tannins. Their biochemistry and nutritional properties. In I.M. Morrison. *Advances in Plant Cell Biochemistry and Biotechnology*. (pp. 151-217). JAI Press Ltd.
- Mupangwa, J. F., Ngongoni, N. T., Topps, J. H., Acamovic, T., Hamudikuwanda, H., and Ndlovu, L. R. (2000). Dry matter intake, apparent digestibility and excretion of purine derivatives in sheep fed tropical legume hay. *Small Ruminant Research*, 36(3): 261-268

- Ngwa, A.T., Nsahlai, I.V. and Iji, P.A. (2003). Effect of feeding legume pods or alfalfa in combination with poor quality grass straw on microbial enzyme activity and production of VFA in the rumen of South African Merino sheep. *Small Ruminant Research*. 48(2): 83-94.
- Nolan, J.V. and Leng, R.A. (1972). Dynamic aspects of ammonia and urea metabolism in sheep. *British Journal of Nutrition*. 27: 177-183.
- NRC. (2001). Nutrient requirements of dairy cattle.7th rev. ed. 381.
- National Research Council (US). Committee on Nutrient Requirements of Small Ruminants. (2007). Nutrient requirements of small ruminants: sheep, goats, cervids, and new world camelids.
- Odo, B.I., Omeje, F.U. and Okwor, J.N. (2001). Forage species availability, food preference and grazing behavior of goats in south eastern Nigeria. *Journal of Small Ruminant Research*. 42: 163–168.
- Oliva, M.L.V., Souza-Pinto, J.C., Batista, I.F., Araujo, M.S., Silveira, V.F., Auerswald, E.A. and Sampaio, C.A. (2000). *Leucaena leucocephala* serine proteinase inhibitor: primary structure and action on blood coagulation, kinin release and rat paw edema. *Biochimica et Biophysica Acta (BBA)-Protein Structure and Molecular Enzymology*. 1477(1): 64-74.
- Oppenheim, E.W., Nasrallah, I.M., Mastri, M.G. and Stover, P.J. (2000). Mimosine is a cell-specific antagonist of folate metabolism. *Journal of Biological Chemistry*. 275: 19268–19274.
- Orden, E.A., Abdulrazak, S.A., Cruz, E.M., Orden, M.E.M., Ichinohe, T., and Fujihara, T. (2000). Leucaena leucocephala and Gliricidia sepium supplementation in sheep fed with ammonia treated rice straw: effects on intake, digestibility, microbial protein yield and live-weight changes. Asian-Australasian Journal of Animal Sciences. 13(12): 1659-1666.
- Osakwe, I.I. and Steingass, H. (2006). Ruminal fermentation and nutrient digestion in West African Dwarf (WAD) sheep fed *Leucaena leucocephala* supplemental diets. *Journal of Agroforestry Systems*. 67: 129–133.
- Otukoya, F.K. and Babayemi, O.J. (2008). Supplementation of *Leucaena leucocephala* hay as protein enrichment for cassava peels in West African dwarf goats. *Journal of Food Agriculture And Environment*. 6(2): 247.
- Paengkoum, P., and Paengkoum, S. (2010). Effects of supplementing rice straw with Leucaena (*Leucaena leucocephala*) and Madras thorn (*Pithecellobium dulce*) foliages on digestibility, microbial N supply and nitrogen balance of growing goats. *Journal of Animal Physiology And Animal Nutrition*. 94(5): e59-e65.
- Pantastico, J.B. and Baldia, J.P. (1980). Ipil-ipil leaf meal as supprement feeding of *Tilapia mossambica*. In fin fish nutrition and fish feed technology. Halver, J.E. and Tiews, K. (Editors), Heenemann, Berlin, 1: 587-593.

- Parsons, T.R., Maita, Y. and Lalli, C.M. (1984). A manual of Chemical and Biological Metlwds for Seawater Analysis. pp. 173. Pergamon Press, New York.
- Phengvichith, V. and Ledin, I. (2007). Effects of supplementing gamba grass (Andropogon gayanus) with cassava (Manihot esculenta Crantz) hay and cassava root chips on feed intake, digestibility and growth in goats. Asian-Australasian Journal of Animal Sciences. 20(5): 725-732.
- Phesatcha, K., Wanapat, M. and Mcsweeney, C.(2013). Effect of Dried Leucaena Leaf Supplementation on Rumen Ecology, Nutrient Digestibility and Urinary Excretion of 2, 3-Dihydroxy Pyridone (2, 3-DHP) and 3, 4-Dihydroxy Pyridone (3, 4-DHP) in Swamp Buffaloes. Buffalo Bulletin Vol.32 (Special Issue 2): 975-979.
- Phuc, B.H.N., Ogle, B. and Lindberg, J.E. (2000). Effect of replacing soybean protein with cassava leaf protein in cassava root meals based diets for growing pigs on digestibility and N retention. *Journal of Animal Feed Science and Technology*. 83(3-4): 223-235.
- Pinho, E.Z.de, Costa, C., Arrigoni, M.D.B., Silveira, A.C., Padovani, C.R. and Pinho, S. Z. de. (2004). Fermentation and nutritive value of silage and hay made from the aerial part of cassava (*Manihot esculenta Agricola*. 61(4): 364-370.
- Pintus, A. (2000). Effect of polyethylene glycol on browsing behaviour and performance of late lactating goats. *Cahiers Options Mediterraneennes*. 52: 147–150.
- Pratchett, D., Jones, R.J. and Syrch, F.X. (1991). Use of DHP-degrading rumen bacteria to overcome toxicity in cattle grains irrigated leucaena pasture. *Tropical Grasslands Journal*. 25: 268–274.
- Preston T.R. (1995). Biological and chemical analytical methods. In: T.R. Preston (Editor). *Tropical Animal Feeding: a Manual for Research Workers*. FAO, Rome.
- Promkot, C., Wanapat, M., Wachirapakorn, C. and Navanukraw, C. (2007). Influence of sulfur on fresh cassava foliage and cassava hay incubated in rumen fluid of beef cattle. *Asian-Australasian Journal of Animal Sciences* 20(9): 1424-1432.
- Promkot, C. and Wanapat, M. (2009). Effect of elemental sulfur supplementation on rumen environment parameters and utilization efficiency of fresh cassava foliage and cassava hay in dairy cattle. *Asian-Australasian Journal of Animal Sciences*. 22(10): 1366-1376.
- Raghavendra, B., Shinde, A.K., Sankhyan, S.K. and Verma, D.L. (2002). Nutrition of range goats in a shrubland of western India. *Asian Australasian Journal* of *Animal Sciences*. 15: 1719–1724.

- Rahman, M.M., Salleh, M.A.M., Sultana, N., Kim, M.J. and Ra, C.S. (2013). Estimation of total volatile fatty acid (VFA) from total organic carbons (TOCs) assessment through in vitro fermentation of livestock feeds. *African Journal of Microbiology Research*. 7(15): 1378-1384.
- Ravindran, V. (1993). Cassava leaves as animal feed: potential and limitations. *Science of Food and Agriculture Journal*. 61: 145–150.
- Ravindran, G. and Ravindran, V. (1988). Changes in the nutritional composition of cassava (Manihot esculenta Crantz) leaves during maturity. *Food Chemistry Journal*. 27(4): 299-309.
- Ravindran, V., Kornegay, E. T., and Rajaguru, A. S. B. (1987). Influence of processing methods and storage time on the cyanide potential of cassava leaf meal. *Animal Feed Science and Technology*, 17(4): 227-234.
- Reed, J.D., Soller, H. and Wood, A. (1990). Fodder tree and straw diets for sheep intake, growth, digestibility and the effect of phenolics on nitrogen utilization. *Journal of Animal Feed Science and Technology*. 30: 39-50.
- Reed, J.D. (1995). Nutritional toxicology of tannins and related polyphenols in forage legumes. *Journal of Animal Science*. 73: 1516.
- Reis, P.J., Tunks, D.A. and Chapman, R.E. (1975). Effects of mimosine, a potential chemical deflecting agent, on wool growth and the skim of sheep. *Australian Journal of Biological Science*. 28: 69-84.
- Reis, P.J., Puchala, R., Sahlu, T. and Goetsch, A.L. (1999). Effects of mimosine and 2,3-dihydroxypyridine on fibre shedding in Angora goats. *Journal of Animal Science*. 77: 1224–1229.
- Rira, M., Marie-Magdeleine, C., Archimède, H., Morgavi, D.P. and Doreau, M. (2013). Effect of condensed tannins on methane emission and ruminal microbial populations. In *Energy and protein metabolism and nutrition in sustainable animal production* (pp. 501-502). Wageningen Academic Publishers.
- Rira, M., Morgavi, D.P., Archimède, H., Marie-Magdeleine, C., Popova, M., Bousseboua, H., and Doreau, M. (2015). Potential of tannin-rich plants for modulating ruminal microbes and ruminal fermentation in sheep. *Journal of Animal Science*. 93(1): 334-347.
- Roza, E., Suardi, M.S., Nurdin, E., and Aritonang, S.N. (2013). Digestibility Test of Cassava Leaves in Feed Supplement on Buffaloes by *In-vitro*. *Pakistan Journal of Nutrition*. 12(5): 505-509.
- Rosa, G.D., Moio, L., Napolitano, F., Grasso, F., Gubitosi, L. and Bordi, A. (2002). Influence of flavor on goat feeding preferences. *Journal of Chemical Ecology*. 28: 269–281.

- Rodríguez, R., Mota, M., Castrillo, C. and Fondevila, M., (2010). *In-vitro* rumen fermentation of the tropical grass *Pennisetum purpureum* and mixtures with browse legumes: effects of tannin contents. *Journal of Animal Physiology and Animal Nutrition*. 94: 696-705.
- Rogers, D.J. and Milner, M. (1963). Amino acid profile of manioc leaf protein in relation to nutritive value. *Economic Botany Journal*. 17(3): 211-216.
- Ruskin F.R. (1977). Leucaena: Promising forage and tree crop for the tropics. 1st Edn. National Academy Press and National Research Council, Washington, DC, USA.
- Sahlu, T., Fernandez, J.M., Lu, C.D. and Manning, R. (1992). Dietary protein level and ruminal degradability for mohair production in Angora goats. *Journal of Animal Science*. 70: 1526-1533.
- Sallam, S.M.A., Bueno, I.C.S., Godoy, P.B., Nozella, E.F., Vitti, D.M.S.S. and Abdalla, A.L. (2010). Ruminal fermentation of tannins bioactivity of some browses using a semi-automated gas production technique. *Journal of Tropical Subtropical Agroecosystem*. 12: 1-10.
- Sandiago, C.B., Aldaba, M.B., Laron, M.A. and Reyes, O.S. (1988). Reproductive performance and growth of Nile tilapia, *Oreochromis niloticus* broodstock fed diets containing *Leucaena leucocephala* leaf meal. Aquaculture. 70(1-2): 53-62.
- Satyanarayana Reddy, P.V.V., Ramachandra Reddy, R. and Sudba Reddy, K.(1987). Utilisation of Subabul (*Leucaena leucocephala*) leaf meal in male chick diets. *Veterinary Journal.* 64: 1078-1079.
- Schofield, P., Mbugua, D.M. and Pell, A.N. (2001). Analysis of condensed tannins: a review. *Journal of Animal Feed Science and Technology*. 91: 21-40.
- Schöner, F.J. (1981). Schätzung des energetischen Futterwertes von Milchleistungsfuttern (Ergänzungsfutter fur Milchkühe) unter besonderer Berücksichtigung des Hohenheimer Futterwerttestes. Dissertation, Bonn University, Bonn, Germany.
- Semenye, P.P. (1990). Toxicity response of goats fed on *Leucaena leucocephala* forage only. *Small Ruminant Research*. 3(6): 617-620.
- Seng, S. and Preston, T.R. (2003). Effect of grass or cassava foliage on growth and nematode parasite infestation in goats fed low or high protein diets in confinement. Journal of *Livestock Research for Rural Development*. 15(8): 1– 15.
- Shaheen, H.M. (2001). The effect of feed and water deprivation on ingestive behavior and blood constituents in camels: comparison with sheep and goats. *Journal of Camel Practice and Research.* 8: 153–162.

- Shinkai, T. and Kobayashi, Y. (2007). Localization of ruminal cellulolytic bacteria on plant fibrous materials as determined by fluorescence in situ hybridization and real-time PCR. Applied and environmental microbiology. 73(5): 1646-1652.
- Shinozaki, K. (1959). Studies on experimental bloat in ruminants. 5. Effects of various volatile fatty acids introduced into the rumen on the rumen motility. *Tohoku Journal* of *Agricultural Research*. 9: 237.
- Singh, K.M., Tripathi, A.K., Pandya, P.R., Rank, D.N., Kothari, R.K. and Joshi, C.G. (2011). Dasytricha dominance in Surti buffalo rumen revealed by 18S rRNA sequences and real-time PCR assay. *Current microbiology*. 63(3): 281-288.
- Skillman, L.C., Toovey, A.F., Williams, A.J. and Wright, A.D.G. (2006). Development and validation of a real-time PCR method to quantify rumen protozoa and examination of variability between Entodinium populations in sheep offered a hay-based diet. *Applied and Environmental Microbiology*. 72(1): 200-206.
- Soltan, Y.A., Morsy, A.S., Sallam, S.M.A., Louvandini, H. and Abdalla, A.L. (2012). Comparative in vitro evaluation of forage legumes (prosopis, acacia, atriplex, and leucaena) on ruminal fermentation and methanogenesis. *Journal of Animal and Feed Sciences*. 575:54.
- Speedy, A., and Pugliese, P. L. (1992). Legume trees and other fodder trees as protein sources for livestock (No. 633.3/S742). FAO.
- Srivastava, S.N.L. and Sharma, K. (1998). Response of goats to pelleted diets containing different proportions of sun-dried *Leucaena leucocephala*. Journal of Small Ruminant Research. 28: 139–148.
- Sung, H.G., Kobayashi, Y., Chang, J., Ha, A., Hwang, I.H. and Ha, J.K. (2007). Low ruminal pH reduces dietary fiber digestion via reduced microbial attachment. *Asian-Australasian Journal of Animal Sciences*. 20(2): 200-207.
- Sutton, D.J., Morant, V.S., Bines, A.J., Napper, J.D. and Givens, I.D. (1993). Effect of altering the starch: fibre ratio in the concentrates on hay intake and milk production by Friesian cows. *Journal Agricultural Science*. 120: 379-390.
- Sylvester, J.T., Karnati, S.K., Yu, Z., Morrison, M. and Firkins, J.L. (2004). Development of an assay to quantify rumen ciliate protozoal biomass in cows using real-time PCR. *The Journal of Nutrition*. 134(12): 3378-3384.
- Sylvester, J.T., Karnati, S.K.R., Yu, Z., Newbold, C.J. and Firkins, J.L. (2005). Evaluation of a real-time PCR assay quantifying the ruminal pool size and duodenal flow of protozoal nitrogen. *Journal of Dairy Science*. 88(6): 2083-2095.
- Tan, H.Y., Sieo, C.C., Abdullah, N., Liang, J.B., Huang, X.D., and Ho, Y.W. (2011). Effects of condensed tannins from Leucaena on methane production, rumen fermentation and populations of methanogens and protozoa. *Journal of Animal Feed Science and Technology*. 169: 185-193.

- Tavendale, M. H., Meagher, L.P., Pacheco, D., Walker, N., Attwood, G.T. and Sivakumaran, S. (2005). Methane production from in vitro rumen incubations with *Lotus pedunculatus* and *Medicago sativa*, and effects of extractable condensed tannin fractions on methanogenesis. *Journal of Animal Feed Science and Technology*. 123:403-419.
- Ter Meulen, U., Struck, S., Schulke, E. and El-Harith, E.A. (1979). A review on the nutritive value and toxic aspects of *Leucaena leucocephala*. *Tropical Animal Production Journal*. 4: 113-26.
- Terry, R.A., Tilley, J.M.A. and Outen, G.E. (1969). Effect of pH on the cellulose digestion under *in-vitro* conditions. *Journal of the Science of Food and Agriculture*. 20: 317.
- Thang, C.M., Sanh, M.V. and Wiktorsson, H. (2004). Effects of supplementation of mixed cassava (*Manihot esculenta*) and legume (*Phaseolus calcaratus*) fodder on the rumen degradability and performance of growing cattle. *Asian-Australasian Journal of Animal Science*. 21(1): 66-74.
- Theodorou, M.K., Williams, B.A., Dhanoa, M.S., McAllan, A.B., France, J. (1994). A simple gas production method using a pressure transducer to determine the fermentation kinetics of ruminant feeds. *Journal of Animal Feed Science and Technology*. 48: 185-197.
- Thomas, D. and Addy, B.L. (1977). Stall-fed beef production in Malawi. *World Review* of Animal Production. 13: 23-30.
- Tiemann, T.T., Avila, P., Ramírez, G., Lascano, C.E., Kreuzer, M. and Hess, H.D. (2008). *In-vitro* ruminal fermentation of tanniniferous tropical plants: Plant-specific tannin effects and counteracting efficiency of PEG. *Journal of Animal Feed Science and Technology*. 146: 222–241.
- Van Soest P.J., Robertson J.B. and Lewis B.A. (1991). Methods for dietary fiber, neutral detergent fiber, and non-starch carbohydrates in relation to animal nutrition. *Journal of Dairy Science*. 74: 3583-3597.
- Van Soest, P.J. (1994). Nutritional ecology of the ruminant, 2<sup>nd</sup> ed. Cornell Univ Press. Ithaca, NY, USA. P.476.
- Van Son, V., Hang, B.P.T., Tri, V.C., and Preston, T.R. (2002). Effect of supplementation of ammoniated rice straw with cassava leaves or grass on intake, digestibility and N retention by goats. *Livestock Research for Rural Development*. 14(3).
- Vaithiyanathan, S., Sheikh, Q. and Kumar, R. (2005). Effect of transinoculation of goat rumen liquor on degradation and metabolism of mimosine in sheep fed with *Leucaena leucocephala* leaves. *Cellulose*. 4: 16-07.

- Vasta, V., Yáñez-Ruiz, D.R., Mele, M., Serra, A., Luciano, G., Lanza, M., and Priolo, A. (2010). Bacterial and protozoal communities and fatty acid profile in the rumen of sheep fed a diet containing added tannins. *Applied And Environmental Microbiology*. 76(8): 2549-2555.
- Verbic, J., Chen, X.B., MacLeod, N.A., and Ørskov, E.R. (1990). Excretion of purine derivatives by ruminants: effects of microbial nucleic acids infusion on purine derivative excretion by steers. *Journal of Agricultural Science (Cambridge)*. 114: 243-248.
- Vogt, G., Quinito, E.T. and Pasculal, F.P. (1986). *Leucaena leucocephala* leaves in formulated feed for *Penaeus monodon*: a concerte example of the application of histology in nutrition research. *Journal of Aquaculture*. 59: 209-234.
- Wanapat, M., Petlum, A. and Pimpa, O. (2000a). Supplementation of cassava hay to replace concentrate use in lactating Holstein-Friesian crossbreds. *Asian-Australasian Journal* of *Animal Science*. 13: 600-604.
- Wanapat, M. (2000). Rumen manipulation to increase the efficient use of local feed resources and productivity of ruminants in the tropics. *Asian-Australasian Journal* of *Animal Science*. 13: 59-67.
- Wanapat, M. and Pimpa, O. (1999). Effect of ruminal NH<sub>3</sub>-N levels on ruminal fermentation, purine derivatives, digestibility and rice straw intake in swamp buffaloes. *Asian-Australasian Journal* of *Animal Science*. 12: 904-907.
- Wanapat, M. (1999). Feeding of ruminants in the tropics based on local feed resources. Khon Kaen Publ. Comp. Ltd., Khon Kaen, Thailand. 236.
- Wang, Y., McAllister, T.A., Yanke, L.J. and Cheeke, P.R. (2000). Effect of steroidal saponin from Yucca schidigera extract on ruminal microbes. *Journal of Applied Microbiology*. 88(5): 887-896.
- Widiawati, Y., Winugroho, M., Teleni, E. and Thalib, A. (2007). Fermentation kinetics (*in-vitro*) of *Leucaena leucocephala*, *Gliricidia sepium* and *Calliandracallothyrsus* leaves (3) the pattern of gas production, organicmatterdegradation, pH,  $NH_3$  and VFA concentration; estimated  $CH_4$  and microbial biomass production. *Indonesian Journal of Animal and Veterinary Sciences*. 12(3).
- Wina, E., Muetzel, S. and Hoffman, E., Makkar, H.P.S. and Becker, K.(2005). Effect of secondary compounds in forages on rumen micro-organisms quantified by 16S and 18S rRNA. Applications of Gene-Based Technologies for ImprovingAnimal Production and Health in Developing Countries. 397-410.
- Wora-anu, S., Wanapat, M., Wachirapakorn, C. and Nontaso, N. In *Effect of different* tropical feed resources on rumen ecology of beef cattle, Proceedings 11th AAAP Animal Science Congress, Kuala Lumpur, 2005.

- Wora-anu, S., Wanapat, M., Wachirapakorn, C. and Nontaso, N. (2007). Effect of roughage sources on cellulolytic bacteria and rumen ecology of beef cattle. *Asian-Australasian Journal of Animal Science*. 20: 1705-1712.
- Yanez Ruiz, D.R., Moumen, A., Martin Garcia, A.I. and Molina Alcaide, E. (2004). Ruminal fermentation and degradation patterns, protozoa population, and urinary purine derivatives excretion in goats and wethers fed diets based on two-stage olive cake: Effect of PEG supply. *Journal of Animal Science*. 82(7): 2023-2032.
- Yang, C.J.M. and Varga, G.A. (1993) The effects of continuous ruminal dosing with dioctyl sodium sulphosuccinate on ruminal and metabolic characteristics of lactating cows. *British Journal of Nutrition*. 69: 397–408.
- Yulistiani, D., Jelan, Z. A., Liang, J. B., Yaakub, H., and Abdullah, N. (2015). Effects of supplementation of Mulberry (Morus alba) foliage and urea-rice bran as fermentable energy and protein sources in sheep fed urea-treated rice straw based diet. *Asian-Australasian Journal of Animal Sciences*, 28(4): 494.
- Zanu, H.K., Mustapha, M. and Addo Nartey, M. (2012). Response of broiler chickens to diets containing varying levels of Leucaena (*leucaena leucocephala*) leaf meal. *Journal of Animal and Feed Research*. 2(2): 108-112.
- Zhou, H., Li, M., Zi, X., Xu, T. and Hou, G. (2011). Nutritive value of several legumes in Hainan province in China. *Journal of Animal and Veterinary Advances*. 10(13): 1640-1648.
- Zucker, W.V. (1983). Tannins: does structure determine function? An ecological perspective. *American Naturalist Journal*. 121: 335-365.