



**UNIVERSITI PUTRA MALAYSIA**

***AGRO-MORPHOLOGICAL, PHYSIOLOGICAL AND BIOCHEMICAL  
CHARACTERISTICS OF RICE SUBJECTED TO CYCLIC WATER STRESS  
AND POTASSIUM FERTILIZERS***

**NURUL AMALINA BT MOHD ZAIN**

**ITA 2015 1**



**AGRO-MORPHOLOGICAL, PHYSIOLOGICAL AND BIOCHEMICAL  
CHARACTERISTICS OF RICE SUBJECTED TO CYCLIC WATER STRESS AND  
POTASSIUM FERTILIZERS**

By

**NURUL AMALINA BT MOHD ZAIN**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in  
Fulfillment of the Requirements for the Degree of Doctor of Philosophy**

**August 2015**

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## DEDICATION

*For my beloved father, Mohd Zain and my dearest mother, Doyah, thank you for all hardship to guide me from child until now, from primary until my PhD. To My grandfather, Abdul Wahab and my late grandmother, Rokiah, who raised me since child, they are my idol and my backbone in my life. And for my beloved husband, Mohd Hafiz bin Ibrahim, thank you for always be with me, believed in me, guiding me along the journey. Without you there would be no excuses for me to stand still and work hard to achieve my dreams. And to my daughter, Nurul Alisha Fathia, you are my precious diamond and always my forever supporter. My heartfelt gratitude for all love, encouragement and support through the years of my quest for knowledge. May this achievement shall be our stepping stone towards living our dreams and ambitions.....*

*The vegetation of a good land comes forth (easily) by the Permission of its Lord; and that which is bad, brings forth nothing but (a little) with difficulty. Thus do We explain variously the Ayât (proofs, evidences, verses, lessons, signs, revelations, etc.) for a people who give thanks”.*

*“[Al-A'râf 7 : 58]*

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirements for the degree of Doctor of Philosophy

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By

**Nurul Amalina Mohd Zain**

**August 2015**

**Chairman : Professor Mohd Razi Ismail, PhD**

**Institute : Institute Tropical Agriculture**

Water deficit is a major problem in rice production due to increase scarcity of water resource. To solve this problem, the present study was conducted to determine how the use of cyclic water stress and potassium fertilizer could be used to alleviate water stress in rice. In the first experiment, a series of cyclic water stress by days [CW5, CW10, CW15, CW20, CW25 and CW30 including Control flooded (CF), control saturated (CS)] were used to investigate the effect of cyclic water stress on yield, growth, physiological and biochemical response of rice. It was found that higher duration of cyclic water stress (CW15, CW20, CW25 and CW30) had reduced grain yield, total biomass, filled spikelet, 1000 grains weight, total panicle hill<sup>-1</sup>, plant height and total tillers hill<sup>-1</sup> and increase oxidative stress and tiller mortality. Grain yield was the highest both in CF and CS treatments and followed by CW5 and CW10 with a 14.3% difference. The application of CW10 was found to have higher yield and water use efficiency under reduction of water. In the second experiment, three rates of potassium [80 kg K<sub>2</sub>O/ha (control), 120 kg K<sub>2</sub>O/ha and 160 kg K<sub>2</sub>O/ha] and three levels of cyclic water stress (CW5, CW10 and CW15) was used to characterize the role of potassium in alleviating water stress. It was observed that rice yield, harvest index, leaf gas exchange, total chlorophyll content and relative water content was influenced by interaction effects between cyclic water stress and potassium rates. The 1000 grains weight, total biomass and water productivity was influenced by potassium rates. It was found that cyclic water stress 10 days (CW10) with potassium fertilization at 120 kg K<sub>2</sub>O/ha was the best practices in achieving higher yield with less water, maximum efficiency of photosystem II ( $F_v/F_m$ ), 1000 grains weight, total biomass production as well as uptake of major nutrient elements (N, P, K, Ca, Mg and Fe) in rice. In the third experiment, four levels of potassium rates [Control, 80 kg K<sub>2</sub>O/ha, 120 kg K<sub>2</sub>O/ha and 160 kg K<sub>2</sub>O/ha] and two types potassium sources (KCl and K<sub>2</sub>SO<sub>4</sub>) were used to

investigate the influence of potassium fertilization in minimizing the effect of cyclic water stress in rice production. It was observed that panicle dry weight  $\text{hill}^{-1}$ , root dry weight, rice yield, Catalase activity (CAT), proline, malondialdehyde (MDA) and harvest index was influenced by potassium rates. The leaves numbers, total tillers and 1000 grains weight were influenced by potassium types. Interaction effects (potassium rate x potassium types) was observed in shoot dry weight, leaf area, total spikelet  $\text{panicle}^{-1}$ , net assimilation rate, transpiration rate and water use efficiency. From the study, the application either KCl or  $\text{K}_2\text{SO}_4$  at 120 kg  $\text{K}_2\text{O}/\text{ha}$  was efficient in minimizing yield reduction under water stress. In the fourth experiment, five treatments including (1) standard local grower's practice (control, 80CF = 80 kg  $\text{K}_2\text{O}/\text{ha}$  + control flooded); (2) 120CW15 = 120 kg  $\text{K}_2\text{O}/\text{ha}$  + cyclic water stress 15 days; (3) 120DS15V = 120 kg  $\text{K}_2\text{O}/\text{ha}$  + drought stress for 15 days during the vegetative stage; (4) 120DS25V = 120 kg  $\text{K}_2\text{O}/\text{ha}$  + drought stress for 25 days and (5) 120DS15R = 120 kg  $\text{K}_2\text{O}/\text{ha}$  + drought stress for 15 days during the reproductive stage, were evaluated to assess the effects of different water stress combined with potassium fertilization on growth, yield, leaf gas exchanges and biochemical changes in rice. It was found that rice under 120CW15 treatment showed tolerance to drought stress by having high water use efficiency, peroxidase (POX), catalase (CAT), proline, maximum efficiency of photosystem II ( $F_V/F_M$ ) and lower minimal fluorescence ( $F_0$ ), compared to other treatments. Based on the result, the sub experiment was conducted to identify *LEA* gene relation with water stress and potassium input on rice. Three treatments were evaluated i.e. Control (control flooded + 80 kg  $\text{K}_2\text{O}/\text{ha}$ ), Water stress 25 days + 80 kg  $\text{K}_2\text{O}/\text{ha}$  and Water stress 25 days + 120  $\text{K}_2\text{O}/\text{ha}$  to detect the expression of Late Embryogenesis Abundant (*LEA*) genes under water stress conditions. The result showed that imposition of 25 day water stress with 120 kg  $\text{K}_2\text{O}/\text{ha}$  (WSK) was proven to reduce *LEA* gene expression and achieve high plant growth and yield. Meanwhile, the 25 day water stress with 80 kg  $\text{K}_2\text{O}/\text{ha}$  (WS) was shown to have the highest induction of the *LEA* gene and lower rice yield and plant growth. From this project, it can be concluded that application of cyclic water stress and potassium fertilizer was able to mitigate water stress and maximize water use efficiency in rice.

Abstrak tesis yang dikemukakan kepada Senat Univerisiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

**AGRO-MORFOLOGI, FISIOLOGI DAN CIRI-CIRI BIOKIMIA PADI  
BERDASARKAN KEPADA TEGASAN AIR BERKITAR DAN BAJA KALIUM**

Oleh

**NURUL AMALINA BT MOHD ZAIN**

**Ogos 2015**

**Pengerusi : Profesor Mohd Razi Ismail, PhD**

**Institusi : Institut Pertanian Tropika**

Defisit air adalah masalah utama bagi produksi padi kerana sumber air semakin sukar didapati. Untuk menyelesaikan masalah ini, kajian ini telah dijalankan untuk menentukan bagaimana penggunaan yaring penggunaan tegasan air berkitar dan pbaja kalium pula boleh digunakan untuk mengurangkan kesan tegasan air pada padi. Dalam eksperimen pertama, beberapa siri tegasan air berkitar berdasarkan hari [CW5, CW10, CW15, CW20, CW25 dan CW30 termasuk kawalan banjir (CF), kawalan tepu (CS)] telah dilaksanakan untuk mengkaji kesan tegasan air berkitar pada hasil, pertumbuhan, tindak balas fisiologi dan biokimia. Kajian mendapati, lebih lama tempoh tegasan air berkitar (CW15, CW20, CW25, CW30) telah mengurangkan hasil bijirin, jumlah biomass, bijirin padi berisi, berat 1000 bijirin padi, jumlah tangkai padi per rumpun, ketidaksurutan tangkai padi, ketinggian pokok, jumlah tangkai padi per rumpun dan meningkatkan tekanan oksidatif. Hasil bijirin adalah paling tinggi dalam kedua-dua CF dan CS dan diikuti oleh CW5 and CW10 dengan perbezaan 14.3%. Penggunaan CW10 didapati dapat memberikan hasil dan kecekapan penggunaan air (WUE) tinggi dalam kekurangan air. Dalam eksperimen kedua, tiga kadar kalium [ 80 kg K<sub>2</sub>O/ha, 120 kg K<sub>2</sub>O/ha dan 160 kg K<sub>2</sub>O/ha] dan tiga tegasan air berkitar (CW5, CW10 dan CW15) telah digunakan untuk mengkaji kesan kalium dalam mngurangkan kesan tegasan air. Pemerhatian menunjukkan hasil bijirin, indeks tuaian, pertukaran gas daun, jumlah kandungan klorofil dan kandungan air relatif telah dipengaruhi oleh interaksi antara tegasan air berkitar dan kadar kalium. Berat 1000 bijirin, jumlah biomass, produksi air telah dipengaruhi oleh kadar kalium. Penemuan menunjukkan tegasan air berkitar 10 hari dengan pembajaan kalium pada 120 kg K<sub>2</sub>O/ha (CW10) adalah praktis terbaik dalam mencapai hasil tuaian lebih tinggi hasil dengan pengurangan air, tinggi kecekapan maksimum Fotosistem II (Fv/Fm), berat 1000 bijirin, jumlah pengeluaran biomass dan juga pengambilan nutrien utama (N, P,K, Ca, Mg and Fe) dalam padi. Dalam eksperimen ketiga, empat tahap kadar kalium [Kawalan, 80 kg K<sub>2</sub>O/ha, 120 kg



$K_2O/ha$  dan  $160\text{ kg }K_2O/ha$  dan dua jenis kalium ( $KCl$  dan  $K_2SO_4$ ) telah digunakan pada padi untuk mengkaji pengaruh pembajaan kalium dalam mengurangkan kesan tegasan air berkitar dalam pengeluaran padi. Diperhatikan bahawa berat kering tangkai per rumpun, berat kering akar, hasil bijirin, aktiviti katalase (CAT), prolin, maliondialdehida (MDA) dan indeks tuaian telah dipengaruhi oleh kadar kalium. Bilangan dedaun, jumlah tiller padi dan berat 1000 bijirin telah dipengaruhi oleh jenis kalium. Kesan interaksi (kadar kalium x jenis kalium) diperhatikan pada berat kering bahagian pokok selain akar, luas daun, jumlah spikelet padi tangkai<sup>-1</sup>, kadar asimilasi bersih, kadar transpirasi dan kecekapan penggunaan air. Daripada kajian, aplikasi sama ada  $KCl$  atau  $K_2SO_4$  pada  $120\text{ kg }K_2O/ha$  adalah efisien dalam mengurangkan pengurangan hasil ketika defisit air. Dalam eksperimen keempat, lima rawatan termasuk (1) standard praktis penanam tempatan (kawalan,  $80CF = 80\text{ kg }K_2O/ha +$  kawalan banjir); (2)  $120CW15 = 120\text{ kg }K_2O/ha +$  tegasan air kitaran 15 hari; (3)  $120DS15V = 120\text{ kg }K_2O/ha +$  tegasan kemarau untuk 15 hari ketika peringkat vegetatif; (4)  $120DS25V = 120\text{ kg }K_2O/ha +$  tegasan kemarau untuk 25 hari dalam peringkat vegetatif dan (5)  $120DS15R = 120\text{ kg }K_2O/ha +$  tegasan kemarau untuk 15 hari dalam peringkat reproduktif, telah dikaji dalam eksperimen ini untuk menilai kesan-kesan kepelbagaian tegasan air berkombinasi dengan sistem pembajaan kalium pada pertumbuhan, hasil, pertukaran gas daun, pertukaran biokimia dalam pertumbuhan padi. Didapati bahawa padi di bawah rawatan  $120CW15$  mempunyai toleransi kepada tegasan kemarau melalui peningkatan kecekapan penggunaan air, peroksidase (POX), katalase (CAT), prolin, kecekapan maksimum fotosistem II ( $F_v/F_m$ ) dan fluorescence minimum yang lebih rendah ( $F_0$ ) berbanding dengan rawatan lain. Berdasarkan keputusan eksperimen keempat, sub eksperimen telah dijalankan untuk mengenalpasti kaitan gen *LEA* dengan tegasan air dan input kalium pada padi. Tiga rawatan telah dijalankan i.e. kawalan (kawalan banjir +  $80\text{ kg }K_2O/ha$ ), tegasan air 25 hari +  $80\text{ kg }K_2O/ha$  dan tegasan air 25 hari +  $120\text{ kg }K_2O/ha$  untuk mengesan ekspresi gen Late Embryogenesis Abundant (*LEA*) dalam keadaan tegasan air. Perlaksanaan ketegasan air 25 hari dengan  $120\text{ kg }K_2O/ha$  terbukti mengurangkan ekspresi gen *LEA* dan mencapai pertumbuhan pokok dan hasil yang tinggi. Sementara, ketegasan air 25 hari dengan  $80\text{ kg }K_2O/ha$  telah menunjukkan paling tinggi induksi gen *LEA* dan hasil padi dan pertumbuhan pokok yang rendah. Daripada projek ini, dapat disimpulkan bahawa penggunaan tegasan air berkitar dan baja kalium pada padi boleh mengurangkan tegasan air dan memaksimumkan kecekapan penggunaan air di dalam padi.



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I certify that a Thesis Examination Committee has met on 21 August 2015 to conduct the final examination of Nurul Amalina binti Mohd Zain on her Doctor of Philosophy thesis entitled “AGRO-MORPHOLOGICAL, PHYSIOLOGICAL AND BIOCHEMICAL CHARACTERISTICS OF RICE SUBJECTED TO CYCLIC WATER STRESS AND POTASSIUM FERTILIZERS”, in accordance with the Universities and University Colleges Act 1971 and the Constitution of the University Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the candidate be awarded the Doctor of Philosophy.

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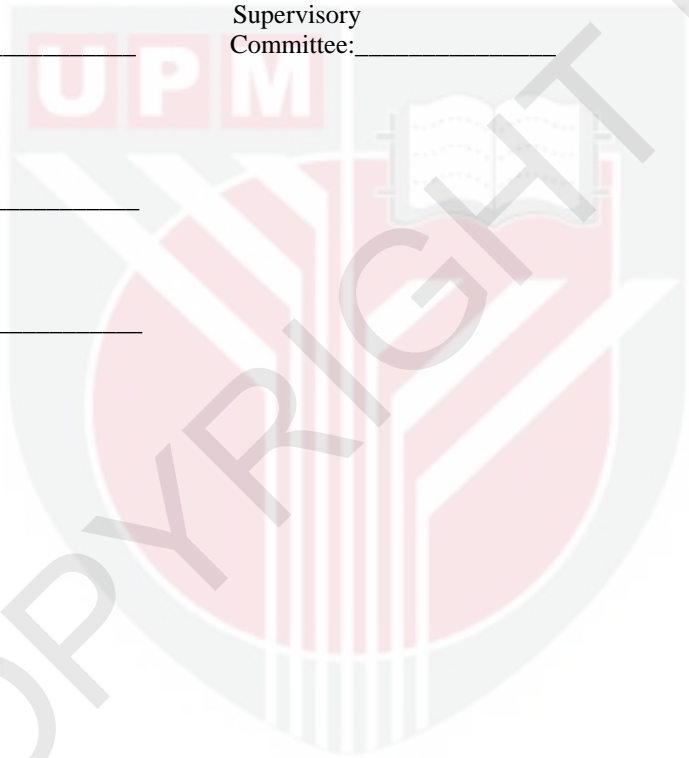
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## LIST OF ABBREVIATIONS AND SYMBOLS

%	percent
*	significant at 0.05 probability level
**	significant at 0.01 probability level
1000gw	1000 grains weight
<i>18sRNA</i>	18S ribosomal RNA
$\mu\text{g g}^{-1}$	micro gram per gram
$\mu\text{mol m}^{-2}\text{s}^{-1}$	micro mole per meter square per second
$\mu\text{mol m}^{-1}$	micro mole carbon dioxide per mole air
$\mu\text{A}$	micro ampere
$\mu\text{L}$	micro litre
$^{\circ}\text{C}$	degree-celcius
L	litre
3PGA	3-Phosphoglicolate Acid
A	net photosynthesis
AA	ascorbic acid
ABA	abscisic acid
AtHK1	Arabidopsis Histidine Kinase
$A_{\text{max}}$	maximum net photosynthesis/net assimilation rate
ANOVA	analysis of variance
AOX	alternative oxidase
APX	ascorbate peroxidases
ATP	adenosine triphosphate
AVP	average vapor pressure
CF	Control flooded
cm	centimeter
$\text{cm}^2$	centimeter square
C.V	coefficient variation
$\text{C}_3$	carbon 3 species
$\text{C}_4$	carbon 4 species
Ca	calcium
CAT	catalase
CAM	crassulacean acid metabolism
cDNA	Complementary Deoxyribonucleic Acid
Chl a	Chlorophyll a
Chl b	Chlorophyll b
$\text{C}_i$	intercellular carbon dioxide concentration
$\text{CO}_2$	carbon dioxide
C/N	carbon to nitrogen ratio
DAS	day after sowing
DAT	day after transplanting
DREB	dehydration-responsive transcription factors
DMRT	Duncan Multiple Range Test
$\text{dS m}^{-1}$	electrical conductivity (EC) formula
DW	dry weight
E	transpiration rate
e.g	example

FAO	Food and Agricultural Organization of the United Nations
Fe	Iron
$F_M$	maximal fluorescence
$F_O$	minimal fluorescence
FS	filled spikelet
$F_V$	variable fluorescence
$F_V/F_O$	indication of maximum quantum yield of photochemical and non-photochemical processes in photosystem II and correlates with leaf photosynthetic capacity
$F_V/F_M$	maximum quantum efficiency of PSII system
FW	fresh weight
g	gram
GA3	gibberellin
<i>GAPDH</i>	Glyceraldehyde 3-phosphate dehydrogenase
GR	glutathione reductase
gs	stomata conductance
GLM	General Linear Model
Gly Bet	glycinebetaine
h	hour
ha	hectare
$H_2O$	water
$H_2O_2$	hydrogen peroxide
$H_2SO_4$	sulphuric acid
HCl	hydrochloric acid
HI	harvest index
Hsps	heat-shock proteins
IAA	indole-3-acetic acid
i.e	that is
IRRI	International Rice Research Institute
WUE	water use efficiency
K	potassium
$K^+$	Ion potassium
KCl	potassium chloride
$K_2SO_4$	potassium sulphate
kg	kilogram
kg N/ha	kilogram nitrogen per hectare
kg $P_2O_5$ /ha	kilogram phosphorus pentoxide per hectare
kg $K_2O$ /ha	kilogram potassium oxide per hectare
Kj	kilo joule
$km^3$	cubic kilometre
kPa	kilo pascal
LAR	leaf area ratio
LEA	Late Embryogenesis-Abundant
LSD	least significant difference
$LSD_{0.05}$	least significant difference at 5 % level
lbs	pound
m	metre
$m^3$	cubic metre (volume)

MDA	Malondialdehyde
MDAR	Monodehydroascorbate Reductase
Mg	magnesium
mg/g	mili gram per gram
mg/kg	mili gram per kilo gram
mm	milimeter
mmol/m <sup>2</sup> /s	mili mole per meter square per second
MOA	Ministry of Agriculture
mol m <sup>-2</sup> s <sup>-1</sup>	mole per meter square per second
MOP	Muriate of Potash
m s <sup>-1</sup>	meter per second
mRNA	messenger ribonucleic acid
N	Nitrogen
nm	nano meter
n.s.	not significant
NADPH	nicotinamide adenine dinucleotide phosphate
NaOH	sodium hydroxide
NaNO <sub>3</sub>	sodium nitrate
NAR	net assimilation rate
NCED3	9-cis-epoxycarotenoid dioxygenase
NTC	No Template Control
O <sub>2</sub>	oxygen
PAL	Phenyl alanine ammonia lyase
pH	power of hydrogen
Phe	Phenylalanine
p	probability
P	Phosphorus
PDW	panicle dry weight
Pi	inorganic phosphorus
P <sub>N</sub>	Net photosynthesis
PNUE	photosynthesis nitrogen use efficiency
pKa	acidity
PAR	photosynthetically active radiation
Pc	photosynthesis carboxylation
PSII	photosystem ii
PEPCase	pep carboxylase
POD	peroxidases
qRT-PCR	Quantitative Reverse-Transcriptase Polymerase Chain Reaction
RCBD	Randomized Complete Block Design
RDW	root dry weight
RGR	relative growth rate
RH	relative humidity
ROS	reactive oxygen species
rpm	rotations per minute
R <sub>d</sub>	dark respiration rate
Rubisco	ribulose biphosphate carboxylase/oxygenase
RuBP	ribulose biphosphate
RWC	relative water content

$r^2$	coefficient of determination or r square
s	second
SAS	Statistical Analysis System
SDW	shoot dry weight
SEM	standard error of difference between means
SLA	specific leaf area
SOD	superoxide dismutase
S/R	shoot to root ratio
SVP	saturation vapor pressure
SPS	sucrose phosphate synthase
SPSS	Statistical Product and Service Solutions
t	time
TB	total biomass
TBA	thiobarbituric acid
TCC	total chlorophyll content
TNC	total non structural carbohydrate
tonnes/ha	metric tonnes per hectare or 1,000 kilogram per hectare
TSP	Triple Super Phosphate
TSS	total soluble sugar
TW	turgid weight
UV	ultra violet ray
VPD	vapor pressure deficit
Var.	variety
$V_{cmax}$	RuBP carboxylation efficiency of PSII
WP	water productivity
WUE	water use efficiency

## CHAPTER 1

### INTRODUCTION

#### 1.1 Overview

Rice (*Oryza sativa* L.) is a major staple crop for about 75% of the world population. It is a good source of energy that contains nutrients, vitamins and minerals. Rice is also a major export crop for several Asian countries. In Malaysia, it is cultivated in eight granary areas along Peninsular Malaysia with an area of approximately 389, 544 ha (Jabatan Pertanian, 2012). In Sabah and Sarawak, hill rice is the major rice found suitable for that environment. Rice comes under the Gramineae Family and there are only two species generally used as food for humans, i.e. *Oryza sativa* and *Oryza glaberrima* (IRRI, 2012). It is a unique crop that can resist and tolerate submergence conditions.

Crop cultivation in Malaysia uses large quantities of water. It is estimated that total water withdrawal for agriculture was 4.520 km<sup>3</sup> (34%), while 3.902 km<sup>3</sup> (30%) was for use by municipalities and 4.788 km<sup>3</sup> (36%) was used by industries (Frenken, 2012). Double cropping of rice cultivation started on the 1960s and by early 1970 created a high demand for irrigation development and efficient water management. Moreover, current water scarcity due to the rapidly increasing world population, progressive global warming, competition of water consumption from domestic and urban sectors and the lack of fresh water sources has resulted in a dextrous problem for crop cultivation. From the irrigation management perspective, drought has become a scary and dextrous environmental factor limiting rice productivity and farmers income (Chai *et al.*, 2006; Yang and Zhang, 2006; Sarvestani *et al.*, 2008).

#### 1.2 Problem Statement /Significance of The Study

Water stress occurs when water uptake by plants decreases over water transpiration and in severe conditions, it could stop physical plant growth and disrupt chemical balance in plants. As a further impact, water stress increases yield reduction and retards crop growth. For tuber crops like potato and radish, the depletion of vegetative storage organs occurs, while in cereal crops aborted tillers, unfilled spikelets and damaged grains frequently occur, and in fruit crops the size and content of the fruits are reduced due to water stress. In facing water scarcity risks, knowledge and technology needs to be rapidly developed to mediate the damaging effects of water scarcity, not only in the urban sector, but also in the agriculture and domestic sectors. In agriculture, research is important to determine new ways to decrease water stress impact on agriculture, in relation to changes in the physiology, breeding, chemistry or molecular approaches (Frenken, 2012).

Therefore, it is important to find the means to reduce water usage and increase the yield of rice. One possible way is to study rice water usage by applying cyclic water stress to the plants. Izanloo *et al.* (2008) have shown that wheat cultivars that are adapted to cyclic water stress gained highest grain number per spike and reduced aborted tillers. The usage of potassium fertilization can minimize the water stress effects on rice. Potassium is a pre-requisite for normal functioning of all plant biochemical and physiological systems. It has been shown to reduce the effects of water stress in many plants (Bajehbaj *et al.*, 2009). Previous studies had reported that potassium was able to mediate adverse effects of water stress on mung bean (Fooladivanda *et al.*, 2014), canola (Rose *et al.*, 2008), and beans (Nasri and Khalatbary, 2011).

No information is available on the water requirements of Malaysian rice varieties and the effect of fertilization with potassium to minimize the effects of water stress on rice productivity. Research on the use of potassium fertilization in Malaysia rice is also scarce. Hence, it is pertinent to establish the water requirements of Malaysian rice by using cyclic water stress technique and the effects of potassium fertilization to minimize the water stress effects on rice yields. This information would be beneficial for efficient use of water by Malaysian rice farmers.

### **1.3 Objectives of the Study**

1. To characterize plant growth, physiology and gas exchange responses of rice under different cyclic water stress.
2. To investigate the usage of potassium rates and cyclic water rates to remedy the effect of cyclic water stress on growth, physiology and biochemistry of rice.
3. To determine biochemical regulation of primary metabolites and antioxidative enzymes of rice subjected to cyclic water stress under different potassium rates and sources.
4. To assess the effects of different cyclic water stress combined with potassium fertilization regimes on growth, yield, leaf gas exchanges and biochemical changes in rice grown in pots compared with standard local rice growing.
5. To characterize gene activities during recovery by potassium remediation.

It was hypothesized that cyclic water stress can increase plant resistance to water stress, leaf gas exchange and enhance root growth. This promotes flowering and leaf senescence. It was also hypothesized that addition of potassium to rice can increase plant water status and osmotic adjustments under cyclic water stress.

## REFERENCES

- Abd-Alla, M.H. and Wahab, A.M.A. 1995. Response of nitrogen fixation, nodule activities, and growth to potassium supply in water-stressed broad bean. *Journal of Plant Nutrition* 18:1391-1402.
- Abdel Wahab, A.M. and Abd-Alla, M.H. 1995. The role of potassium fertilizer in nodulation and nitrogen fixation of faba bean (*Vicia faba* L.) plants under drought stress. *Biology and Fertility of Soils* 20:147-150.
- Aebi, H. 1983. Catalase. In *Methods of Enzymatic Analysis*, 3rd ed.; Bergmeyer, H., Verlag, C., Weinheim, A., Eds.; Verlag Chemie: Weinheim, Germany, 273-277.
- Ahmed, A.L., Sultan, M.S., Badawi, M.A. and Abd El-Rahman, A.A. 2010. Performance of some rice cultivars as affected by irrigation and potassium fertilizer under saline soil condition: I. yield and yield components. *Crop and Environment* 1(1):18-21.
- Ahmed, I.M., Dai, H.; Zheng, W., Cao, F., Zhang, G., Sun, D. and Wu, F. 2013. Genotypic differences in physiological characteristics in the tolerance to drought and salinity combined stress between Tibetan wild and cultivated barley. *Plant Physiology Biochemistry* 63:49-60.
- Ahmed, K.M., Bhattacharya, P., Hasan, M.A., Akhter, S.H., Alam, S.M.M., Bhuyian, M.A.H., Imam, M. B., Khan, A. A., and Sracek, O. 2004. Arsenic enrichment in groundwater of the alluvial aquifers in Bangladesh: An overview. *Application Geochemistry* 19:181-200.
- Akram, M.S., Ashraf, M., Shahbaz, M. and Akram, N.A. 2009. Growth and photosynthesis of salt-stressed sunflower (*Helianthus annuus*) plants as affected by foliar-applied different potassium salts. *Journal Plant Nutrient and Soil Science* 172:884-893.
- Alam, R., Iqbal, A., Khan, I., Ali, I., Munir, I., Tahir, M., Jan, N. and Swati, Z.A. 2011. Carbon and nitrogen stoichiometry in *Brassica napus* L. seedlings after supplementation with  $Ca^{2+}$  and  $K^{+}$  under irrigated and drought stress conditions. *African Journal of Biotechnology* 10:18418-18424.
- Ali, M., Jensen, C.R., Mogensen, V.O., Andersen, M.N. and Henson, I.E. 1999. Root signalling and osmotic adjustment during intermittent soil drying sustain grain yield of field grown wheat. *Field Crops Research* 62:35-52.
- Andersen, M.N., Jensen, C.R. and Losch, R. 1992. The interaction effects of potassium and drought in field-grown barley. I. Yield, water-use efficiency and growth. *Soil Plant Science* 42:34-44.



- Aown, M., Raza, S., Saleem, M.F., Anjum, S.A., Khaliq, T. and Wahid, M.A. 2012. Foliar application of potassium under water deficit conditions, Improved the growth and yield of wheat (*Triticum aestivum* L.). *The Journal of Animal and Plant Sciences* 22(2):431–437.
- Apel K. and Hirt H. 2004. Reactive oxygen species: Metabolism, oxidative stress and signal transduction. *Annual Review of Plant Biology* 55:373–399.
- Aref, F. and Rad, H.E. 2012. Physiological characterization of rice under salinity stress during vegetative and reproductive stages. *Indian Journal of Science and Technology* 5:2578–2586.
- Asch, F., Dingkuhn, M., Sow, A. and Audebert, A. 2004. Drought-induced changes in rooting patterns and assimilate partitioning between root and shoot in upland rice. *Field Crops Research* 93: 223–236.
- Ashraf, M.A., Ahmad, M.S.A., Ashref, M., Al-Qurainy, F. and Ashraf, M.Y. 2011. Alleviation of waterlogging stress in upland cotton (*Gossypium hirsutum* L.) by exogenous application of potassium in soil and as a foliar spray. *Crop and Pasture Science* 62: 25-38.
- Asch, F., Dingkuhn, M., Wittstock, C. and Doerffling, K. 1998. Sodium and potassium uptake of rice panicles as affected by salinity and season in relation to yield and yield components. *Plant and Soil* 207:133–145.
- Assmann, S.M. and Shimazaki, K. 1999. The multisensory guard cell, stomatal responses to blue light and abscisic acid. *Plant Physiology* 119: 809–815.
- Atkin, O.K. and Macherel, D. 2009. The crucial role of plant mitochondria in orchestrating drought tolerance. *Annual Botany* 103:581–597.
- Auge, R.M. and Moore, J.L. 2002. Stomatal response to non-hydraulic root to shoot communication of partial soil drying in relation to foliar dehydration tolerance. *Environmental Experiment Botany* 47:217–229.
- Baek, M.H., Byung, Y.C., Kim, J.H., Seung, G.W., Kim, J.S. and Lee, I.J. 2006. Gamma radiation and hormone treatment as tools to reduce salt stress in rice (*Oryza sativa* L.). *Journal of Plant Biology* 49:257–260.
- Bague, M.A., Karim, M.A., Hamid, A. and Tetsushi, H. 2006. Effect of fertilizer potassium on growth, yield and nutrient uptake of wheat under water stress condition. *South Pacific Studies* 1:25-37.
- Bajehbaj, A.A., Qasimov, N. and Yarnia M. 2009. Effects of drought stress and potassium on some of the physiological and morphological traits of sunflower (*Helianthus annuus* L.) cultivars. *Journal of Food, Agriculture and Environment* 7:448–451.
- Bajji, M., Kinet, J.M. and Lutts, S. 2002. The use of the electrolyte leakage method for assessing cell membrane stability as a water stress tolerance test in durum wheat. *Plant Growth Regulation* 36:61–70.

- Bajji, M, Lutts, S and Kinet, J.M. 2001. Water deficit effects on solute contribution to osmotic adjustment as a function of leaf ageing in three durum wheat (*Triticum durum* Desf.) cultivars performing differently in arid conditions. *Plant Science* 60:669–681.
- Baker, N.R. and Rosenqvist, E. 2004. Applications of chlorophyll fluorescence can improve crop production strategies: An examination of future possibilities. *Journal of Experimental Botany* 55:1607–1621.
- Bandurska, H. 2004. Free proline accumulation in leaves of cultivated plant species under water deficit conditions. *Acta Agrobotany* 57:57–67.
- Banoc, D.M., Yamauchi, A., Kamoshita, A., Wade, L.J. and Pardales, J.R. 2000. Genotypic variations in response of lateral root development to fluctuating soil moisture in rice. *Plant Production Science* 3: 335–343.
- Basavaraja, H., Mahajanashetti, S.B., Udagatti and Naveen, C. 2007. Economic Analysis of Post-harvest Losses in Food Grains in India: A Case Study of Karnataka. *Agricultural Economics Research Review* 20:117–126.
- Bates, L.S., Waldren, R.P, and Teare, I.D. 1973. Rapid determination of free proline for water stress studies. *Plant Soil* 39:205–211.
- Battista, J.R., Park, M.J. and McLemore, A.E. 2001. Inactivation of two homologues of proteins presumes to involved in the desiccation tolerance of plants sensitizes *Deinococcus radiodurans* RI to dessication. *Cryobiology* 43: 133–139.
- Baumgartl, T. and Kock, B. 2004. Modeling volume change and mechanical properties with hydraulic models. *Soil Science Society of America Journal* 68:57–65.
- Beadle, C.L. 1998. Growth analysis. In: Hall DO, Scurlock JMO, Bolhar N, Leegood RC, Long SP (eds) Photosynthesis and production in a changing environment. Chapman & Hall, New York.
- Benlloch-Gonzalez, M., Arquero, O., Fournier, J.M., Barranco, D. and Benlloch, M. 2008. K<sup>+</sup> starvation inhibits water-stress-induced stomatal closure. *Plant Physiology* 165:623–630.
- Benlloch-Gonzalez, M., Romera, J., Cristescu, S., Harren, F., Fournier, J.M. and Benlloch, M. 2010. K<sup>+</sup> starvation inhibits water-stress-induced stomatal closure via ethylene synthesis in sunflower plants. *Experimental Botany* 61:1139–1145.
- Beyrouthy, C.A. Grigg, B.C., Norman, R.J. and Wells, B.R. 1994. Nutrient uptake by rice in response to water management. *Journal Plant Nutrition* 17:39–55.
- Blackman, S.A., Obendorf, R.L. and Leopold, A.C. 1995. Desiccation tolerance in developing soybean seeds: The role of stress proteins. *Plant Physiology* 93:630–638.

- Bokor, M., Csizmok, V., Kovacs, D., Banki, P., Friedrich, P., Tompa, P. and Tompa, K. 2005. NMR relaxation studies on the hydrate layer of intrinsically unstructured proteins. *Biophysical Journal* 88:2030–2037.
- Bouman, B.A.M. and Tuong, T.P. 2001. Field water management to save water and increase its productivity in irrigated rice. *Agriculture Water Management* 49:11–30.
- Brag, H. 1972. The influence of potassium on the transpiration rate and stomatal opening in *Triticum aestivum* and *Pisum sativum*. *Plant Physiology* 26:250–255.
- Brar, D. S. and Khush, G. S. 2003. Utilization of wild species of genus *Oryza* in rice improvement. In *Monograph on Genus Oryza*; Nanda, J.S. and Sharma, S.D., Eds., 283–309.
- Bray, E.A. 1993. Molecular response to water deficit. *Plant Physiology* 103:1035–1040.
- Bustin, S.A. 2002. Quantification of mRNA using real-time reverse transcription PCR RT-PCR: trends and problems. *Journal of Molecular Endocrinology* 29: 23–29.
- Cabangon, R.J., Tuong T.P. and Abdullah N.B. 2002. Comparing water input and water productivity of transplanted and direct-seeded rice production systems. *Agriculture Water Management* 57: 11–31.
- Cakmak I. 2000. Possible roles of zinc in protecting plant cells from damage by reactive oxygen species. *New Phytology* 146:185–205.
- Cakmak I. 2005. Potassium alleviates detrimental effects of abiotic stresses in plants. *Journal of Plant Nutrition* 268: 121–130.
- Cassman, K.G. and Harwood, R.R. 1995. The nature of agricultural systems: food security and environmental balance. *Food Policy*, 20: 439–454.
- Cassman, K.G., Kerby, T.A., Roberts, B.A., Bryant D.C. and Higashi S.L. 1990. Potassium nutrition effects on lint yield and fiber quality of acala cotton. *Crop Sciences* 30:672–677.
- Centritto, M., Lauteri, M., Monteverdi, M.C. and Serraj, R. 2009. Leaf gas exchange, carbon isotope discrimination, and grain yield in contrasting rice genotypes subjected to water deficits during the reproductive stage. *Journal of Experimental Botany* 60:2325–2339.
- Chai, Y., Wang, W., Zhu, Z., Zhang, Z., Lang, Y and Zhu, Q. 2006. Effect of water stress during grain filling period on rice grain yield and its quality under different nitrogen levels. *National Center for Biotechnology Information, U.S. National Library of Medicine, National Institute of Health* 17:1201–1206.

- Chaitanya, K., Sundar, D., Masilamani, S. and Ramachandra, R.A. 2002. Variation in heat stress induced antioxidant enzyme activities among three mulberry cultivars. *Plant Growth Regulators* 36:175–180.
- Cha-um, S. and Kirdmanee, C. 2008. Effect of osmotic stress on proline accumulation, photosynthetic abilities and growth of sugarcane plantlets (*Saccharum officinarum* L.). *Pakistan Journal of Botany* 40:2541–2552.
- Chaves, M.M., Maroco, J.P and Pereira, J.S. 2003. Understanding plant responses to drought from genes to the whole plant. *Functional Plant Biology* 30:239–264.
- Chaves, M.M., Flexas, J. and Pinheiro, C. 2009. Photosynthesis under drought and salt stress: regulation mechanisms from whole plant to cell. *Annual Botany* 103:551–560.
- Chaves, M.M., Pereira, J.S., Maroco, J., Rodrigues, M.L., Ricardo, C.P.P., Osorio, M.L., Carvalho, I., Faria, T. and Pinheiro, C. 2002. How plants cope with water stress in the field. Photosynthesis and growth. *Annals of Botany* 89:907–916.
- Chen, J.Q., Meng, Q.P., Zhang, Y., Xia, M. and Wang, X.P. 2008. Over-expression of OsDREB genes lead to enhanced drought tolerance in rice. *Biotechnology Letters* 30:2191–2198.
- Chen, J.X., Xuan, J.X., Du, C.L. and Xie, J.C. 1997. Effect of potassium nutrition of rice on rhizosphere redox status. *Plant Soil* 188:131–137.
- Chen, J.X. and Chen, J.X. 1997. Effect of K nutrition on rice root growth and nutrient uptake. *Acta Pedology Sinica* 34:182–188.
- Chimenti, C.A., Pearson, J. and Hall, A.J. 2002 Osmotic adjustment and yield maintenance under drought in sun-flower. *Field Crops Research* 75:235–246.
- Chutipaijit, S., Cha-Um, S. and Sompornpailin, K. 2012. An evaluation of water deficit tolerance screening in pigmented indica rice genotypes. *Pakistan Journal of Botany* 4:65–72.
- Chutipaijit, S., Cha-Um, S. and Sompornpailin, K. 2009. Differential accumulations of proline and flavonoids in indica rice varieties against salinity. *Pakistan Journal of Botany* 41:2497–2506.
- Chutipaijit, S., Suriyan, C.U. and Sompornpailin, K. 2010. Proline accumulation and physiological responses of indica rice genotypes differing in tolerance to salt and drought stresses. *Philippine Agriculture Science* 93:165–169.
- Clerkx, E.J.M., El-Lithy, M.E., Vierling, E., Ruys, G.J., Blankestijn-DeVries, H., Groot, S.P.C., Vreugdenhil, D. and Koornneef, M. 2004. Analysis of natural allelic variation of *Arabidopsis* seed germination and seed longevity traits between the accessions *Landsberg erecta* and *Shakdara* using a new recombinant inbred line population. *Plant Physiology* 135:432–443.

- Close, T.J. and Lammers, P.J. 1993. An osmotic stress protein of cyanobacteria is immunologically related to plant dehydrins, *Plant Physiology* 101:773–779.
- Colmer, T.D., Gibberd, M.R., Wiengweera, A. and Tinh, T.K. 1998. The barrier to radial oxygen loss from roots of rice (*Oryza sativa* L.) is induced by growth in stagnant solution. *Journal of Experimental Botany* 49:1431–1436.
- Cruz de Carvalho, M.H. 2008. Drought stress and reactive oxygen species: Production, scavenging and signaling. *Plant Signal Behavior* 3:156–165.
- Cuellar, T., Pascaud, F., Verdeil, J.L., Torregrosa, L., Adam-Blondon, A.F., Thibaud, J.B., Sentenac, H. and Gaillard, I. 2010. A grapevine Shaker inward K<sup>+</sup> channel activated by the calcineurin B-like calcium sensor 1-protein kinase CIPK23 network is expressed in grape berries under drought stress conditions. *Plant* 61:58–69.
- DaCosta, M. and Huang, B. 2006. Osmotic adjustment associated with variation in bent grass tolerance to drought stress. *Journal of America Society of Horticulture Sciences* 131:338–344.
- Dai, X., Xu, Y., Ma, Q., Xu, W., Wang, T., Xue, Y. and Chong, K. 2007. Overexpression of an R1R2R3 MYB gene, OsMYB3R–2, increases tolerance to freezing, drought and salt stress in transgenic Arabidopsis. *Plant Physiology* 143:1739–1751.
- Dalal, V.K. and Tripathy, B.C. 2012. Modulation of chlorophyll biosynthesis by water stress in rice seedlings during chloroplast biogenesis. *Plant Cell Environment* 35(9):1685–703.
- Dardick, C., Chen, J., Richter, T., Ouyang, S. and Ronald, P. 2007. The rice kinase database. A phylogenomic database for the rice kinome. *Plant Physiology* 143:579–586.
- Dat, J., Vandenabeele, S., Vranova, E., van Montagu, M., Inze, D. and van Breusegem, F. 2000. Dual action of the active oxygen species during plant stress responses. *Cellular and Molecular Life Sciences* 57:779–795.
- Degenkolbe, T., Do, P.T., Zuther, E., Repsilber, D., Walther, D., Hinch, D.K. and Kohl, K.I. 2009. Expression profiling of rice cultivars differing in their tolerance to long-term drought stress. *Plant Molecular Biology* 69:133–153.
- Deka, M. and Baruah, K.K. 2000. Comparable studies of rainfed upland winter rice (*Oryza sativa*) cultivars for drought tolerance. *Indian Journal of Agricultural Science* 70:135–139.
- Demmig-Adams, B. and Adams, W.W. 2006. Photoprotection in an ecological context: the remarkable complexity of thermal energy dissipation. *New Phytology* 171:11–21.



- Denby, K. and Gehring, C. 2005. Engineering drought and salinity tolerance in plants: lessons from genome-wide expression profiling in Arabidopsis. *Trends in Biotechnology* 23:547–552.
- Dingkuhn, M., Jones, M.P., Johnson D.E. and Sow, A. 1998. Growth and yield potential of *Oryza sativa* and *O. glaberrima* upland rice cultivars and their interspecific progenies. *Field Crops Research* 57:57–69.
- Dingkuhn, M., Cruz, R.T., O'Toole, J.C., Turner, N.C. and Doerffling, K. 1991. Responses of seven diverse rice cultivars to water deficits. III. Accumulation of abscisic acid and proline in relation to leaf water-potential and osmotic adjustment. *Field Crops Research* 27:103–117.
- Dong, B., Molden, D., Loeve, R., Li, Y.H., Chen, C.D., and Wang, J.Z. 2004. Farm level practices and water productivity in Zanghe Irrigation System. *Paddy Water Environment* 2:217–226.
- Duan, J. and Cai, W. 2012. OsLEA3-2, an abiotic stress induced gene of rice plays a key role in salt and drought tolerance. *Plos One* 7:1–11.
- Dure, L. 1993. A repeating II-mer amino acid motif and plant desiccation. *Plant Journal* 3:363–369.
- Dure, L., Crouch, M., Harada, J., Ho, T., Mundy, J., Quatrano, R., Thomas, T. and Sung, Z.R. 1989. Common amino acid sequence domains among the LEA proteins of higher plant. *Plant Molecular Biology* 12:475–486.
- Egilla, J.N., Davies, F.T. and Boutton, T.W. 2005. Drought stress influences leaf water content, photosynthesis, and water-use efficiency of *Hibiscus rosasinensis* at three potassium concentrations. *Photosynthetica* 43:135–140.
- Egilla, J.N., Davies, F.T. and Drew, M.C. 2001. Effect of potassium on drought resistance of *Hibiscus rosasinensis* cv. *Leprechaun*: Plant growth, leaf macro- and micronutrient content and root longevity. *Plant Soil* 229:213–224.
- Ekanayake, I.J., De Datta, S.K. and Steponkus, P.L. 1989. Spikelet sterility and flowering response of rice to water stress at anthesis. *Annual Botany* 63:257–264.
- FAO, 2012. FAO Statistical Yearbook. Available at <http://www.fao.org/statistics/yearbook>
- FAO, 2008. Sustainable rice production for food security Proceedings of the 20<sup>th</sup> Session of the International Rice Commission Bangkok, Thailand, 23–26 July 2007.
- FAO, 2006. FAOSTAT Agriculture Data [online]. In: [www.fao.org](http://www.fao.org). Available at <http://faostat.fao.org>, Rome, Italy: Food and Agriculture Organization (FAO).
- Farooq, M., Aziz, T., Chemma Z.A., Hussian, M. and Khaliq, A. 2008. Activation of antioxidant system by KCl improves the chilling tolerance in hybrid maize. *Agronomy Crop Sciences* 194:438–448.

- Farooq, M., Wahid, A., Basra, S.M.A. and Islam, U. 2009. Improving water relations and gas exchange with brassinosteroids in rice under drought stress. *Journal of Agronomy Crop Science* 195: 262–269.
- Farooq, M., Wahid, A., Kobayashi, N., Fujita, D. and Basra, S.M.A. 2009 Plant drought stress: Effects, mechanisms and management. *Agronomy for Sustainability Development* 29:185–212.
- Fayyaz, P., Etemadi, E., Julaiee-Manesh, N. and Zolfaghari R. 2013. Sodium and potassium allocation under drought stress in Atlas mastic tree (*Pistacia atlantica* subsp. Mutica). *Journal of Forest* 6: 90–94.
- Fernández-Escobar, R., García, T. and Benlloch, B. 1994. Estado nutritivo de las plantaciones de olivar en la provincia, *Granada ITEA*.90:39–49.
- Flexas, J., Bota, J., Loreto, F., Cornic, G. and Sharkey, T.D. 2004. Diffusive and metabolic limitations to photosynthesis under drought and salinity in C3 plants. *Plant Biology* 6:269–279.
- Florez-Sarasa, I.D., Bouma, T.J., Medrano, H., Azcon-Bieto, J. and Ribas-Carbo, M. 2007 Contribution of the cytochrome and alternative pathways to growth respiration and maintenance respiration in *Arabidopsis thaliana*. *Physiology Plantarum* 129:143–151.
- Fooladivanda, Z., Hassanzadehdelouei, M. and Zarifinia, N. 2014. Effects of water stress and potassium on Quantity traits of two varieties of Mung Bean (*Vigna radiata* L.). *Cercetări Agronomice În Moldova* 1:157–158.
- Ford, C.W. and Wilson, J.R. 1981. Changes in levels of solutes during osmotic adjustment to water stress in leaves of four tropical pasture species. *Australian Journal Plant Physiology* 8:77–91.
- Foyer, C.H., Vanacker, H., Gomez, L.D. and Harbinson, J. 2002. Regulation of photosynthesis and antioxidant metabolism in maize leaves at optimal and chilling temperatures. *Review of Plant Physiology and Biochemistry* 40:659–668.
- Foyer, C.H., Decourviers, P. and Kenerk, K.J. 1994. Protection against oxygen radicals: An important defense mechanism studied in transgenic plants. *Plant Cell Environment* 17: 507–523.
- Foyer, C.H. and Paul, M.J. 2001. Source-sink relationship. *Encyclopedia Of Life Sciences*. Nature Publishing Group / [www.els.net](http://www.els.net)
- Frenken K. 2012. Irrigation In Southern And Eastern Asia In Figures AQUASTAT Survey – 2011. FAO Water Report. Food And Agriculture Organization Of The United Nations, Rome, 311–322.



- Fu, G.F., Song, J., Li, Y.R., Yue, M.K., Xiong, J. and Tao, L.X. 2010. Alterations of panicle antioxidant metabolism and carbohydrate content and pistil water potential involved in spikelet sterility in rice under water-deficit stress. *Rice Science* 17:303–310.
- Fu, X.Q., He, H.H., Wen, P., Luo, X.D. and Xie, J.K. 2012. Drought resistance evaluation system for backcross lines of Dongxiang common wild rice (*Oryza rufipogon* Griff.). *China Journal Applied Ecology* 23:1277–1285.
- Fukai, S. and Cooper, M. 1995. Development of drought-resistant cultivars using physio-morphological traits in rice. *Field Crops Research* 40:67–86.
- Gachon, C., Mingam, A. and Charrier, B. 2004. Real-time PCR: what relevance to plant studies? *Journal of Experimental Botany* 55:1445–1454.
- Gajdanowicz, P., Michard, E., Sandmann, M., Rocha, M. and Dreyer, I. 2011. Potassium gradients serve as a mobile energy source in plant vascular tissues. *Proceedings of the National Academy of Sciences* 108:864–869.
- Gal, T.Z., Glazer, I and Koltai, H. 2003. Differential gene expression during desiccation stress in the insect-killing nematodes *Steinernema feltiae* IS-6. *Journal Parasitology* 89:761–766.
- Galmez, J., Pou, A., Alsina, M.M., Tomas, M., Medrano H. and Flexas, J. 2007. Aquaporin expression in response to different water stress intensities and recovery in Richter-110 (*Vitis* sp.): Relationship with ecophysiological status. *Planta* 226:671–681.
- Gauthami, P., Subrahmanyam, D., Padma, V., Raghuvver Rao, P. and Voleti, S.R. 2013. Influence of simulated post-anthesis water stress on stem dry matter remobilization, yield and its components in rice. *Indian Journal of Plant Physiology* 18:177–182.
- Ge, T.D., Sun, N.B., Bai, L.P., Tong, C.L. and Sui, F.G. 2012. Effects of drought stress on phosphorus and potassium uptake dynamics in summer maize (*Zea mays*) throughout the growth cycle. *Acta Physiologiae Plantarum* 34:2179–2186.
- Gibson, S.I. 2005. Control of plant development and gene expression by sugar signaling. *Current Opinion in Plant Biology* 8:93–102.
- Gong, P., Zhang, J. and Li, H. 2010. Transcriptional profiles of drought-responsive genes in modulating transcription signal transduction, and biochemical pathways into tomato. *Experimental Botany* 61:3563–3575.
- Gopalan, C., Rama Sastri, B.V. and Balasubramanian, S. 2007. Nutritive Value of Indian Foods, published by National Institute of Nutrition (NIN), ICMR.

- Guan, L.M., Zhao, J., and Scandalios, J.G. 2000. Cis-elements and trans-factors that regulate expression of the maize Cat1 antioxidant gene in response to ABA and osmotic stress: H<sub>2</sub>O<sub>2</sub> is the likely intermediary signaling molecule for the response. *Plant Journal* 22:87–95.
- Hajiboland, R. and Beiramzadeh, N. 2008. Growth, gas exchange and function of antioxidant defense system in two contrasting rice genotypes under Zn and Fe deficiency and hypoxia. *Acta Biologica Szegediensis* 52:283–294.
- Hara, Y. 2013. Suppressive effect of sulfate on establishment of rice seedlings in submerged soil may be due to sulfide generation around the seeds. *Plant Production Science* 16:50–60.
- Hassan, Z. and Arshad, M. 2008. Evaluating factors affecting cotton tolerance to potassium deficiency stress using path analysis. *International Journal Agriculture Biology* 10:511–516.
- Herhog, V. and Fahimi H. 1973. Determination of the activity of peroxidase. *Analytical Biochemistry* 55:554 – 562.
- Holthusen, D., Peth, S. and Horn, R. 2010. Impact of potassium concentration and matric potential on soil stability derived from rheological parameters. *Soil and Tillage Research* 111:75–85.
- Horn, R. and Peth, S. 2011. Mechanics of unsaturated soils for agricultural applications. In *Handbook of Soil Sciences*; Huang, P.M., Li, Y., Sumner, M.E., Eds., 2nd ed; CRC Press (Chapter 3).
- Hsaio, T.C. and Xu, L.K. 2000. Sensitivity of growth of roots versus leaves to water stress: biophysical analysis and relation to water transport. *Experimental Botany* 51:1596–1616.
- Hu, L., Wang Z., and Huang, B. 2013. Effects of cytokinin and potassium on stomatal and photosynthetic recovery of kentucky bluegrass from drought stress. *Crop Science* 53:221–231.
- Hundertmark, M. and Hinch, D.K. 2008. Late embryogenesis abundant (LEA) proteins and their encoding genes in *Arabidopsis thaliana*. *BMC Genomics* 9:118–139.
- Hunt, R. 1990. Basic growth analysis. London. Unwin Hyman.
- Ibrahim M.H., Jaafar H.Z.E. and Karimi E. 2012. Ghasemzadeh A., Primary, secondary metabolites, photosynthetic capacity and antioxidant activity of the Malaysian Herb Kacip Fatimah (*Labisia pumila* Benth) exposed to potassium fertilization under greenhouse conditions. *International Journal of Molecular Science* 13:15321–1534.

- Ibrahim, M.H. and Jaafar, H.Z.E. 2011. Photosynthetic capacity, photochemical efficiency and chlorophyll content of three varieties of *Labisia pumila* Benth. exposed to open field and greenhouse growing conditions. *Acta Physiologicae Plantarum* 33:2179–2185.
- Ibrahim, M.H. and Jaafar, H.Z.E. 2012. Reduced photoinhibition under low irradiance enhanced kacip fatimah (*Labisia pumila* Benth) secondary metabolites, phenyl alanine lyase and antioxidant activity. *International Journal of Molecular Science* 13:5290–5306.
- Ingram J. And Bartels D. 1996. The molecular basis of dehydration tolerance in plants. *Annual Review of Plant Physiology and Plant Molecular Biology* 47:377–403.
- International Rice Research Institute (IRRI), 2002. Growth and Morphology of the Rice Plant. [http://www.knowledgebank.irri.org/pu\\_growthMorph.htm](http://www.knowledgebank.irri.org/pu_growthMorph.htm) accessed in 27 February 2013.
- IRRI, International Rice Research Institute 2012. <http://irri.org/about-rice/rice-facts/rice-basics>, <http://ricestat.irri.org:8080/wrs/> accessed in 17 March 2013.
- Iskandar, H.M, Simpson, R.S., Casu, R.E., Bonnett, G.D., Maclean, D.J. and Manners, J.M. 2004. Comparison of reference genes for quantitative real-time polymerase chain reaction analysis of gene expression in sugarcane. *Plant Molecular Biology Report* 22:325–337.
- Islam, MR, Xue, X, Mao, S, Ren, C, Eneji, A.E and Hu, Y. 2011. Effects of water-saving superabsorbent polymer on antioxidant enzyme activities and lipid peroxidation in oat (*Avena sativa* L.) under drought stress. *Journal Science Food Agriculture* 91:680–686.
- Itoh, R. and Kumara, A. 1987. Acclimation of soybean plants to water deficit. V. Contribution of potassium and sugar to osmotic concentration in leaves. *Japan Journal of Crop Science* 56: 678–684.
- Izanloo, A., Anthony G.C., Longridge, P., Tester, M. and Schnurbusch, T., 2008. Different mechanism of adaptation to cyclic water stress in two South Australian bread wheat cultivars. *Journal of Experimental Botany* 59:3324–3346.
- Jabatan Pertanian, 2012. Perangkaan Padi Malaysia 2011, Ed; Jabatan Pertanian Semenanjung Malaysia, pp. 41.
- Jaleel, C.A., Manivannan, P., Lakshmanan, G.M., Gomathinayagam, M. and Panneerselvam, R. 2008. Alterations in morphological parameters and photosynthetic pigment responses of *Catharanthus roseus* under soil water deficits. *Colloids Surfaces. Bulletin Biointerfaces* 61:298–303.
- Ji, X., Shiran, B. and Wan, J. 2010. Importance of pre-anthesis anther sink strength for maintenance of grain number during reproductive stage water stress in wheat. *Plant, Cell and Environment* 33:926–942.

- Jiang, M. and Zhang, J. 2001. Effect of abscisic acid on active oxygen species, antioxidative defence system and oxidative damage. *Plant Cell Physiology* 42:1265–1273.
- Jiang, M. and Zhang, J. 2002. Water stress-induced abscisic acid accumulation triggers the increased generation of reactive oxygen species and up-regulates the activities of antioxidant enzymes in maize leaves. *Journal of Experimental Botany* 53:2401–2410.
- Jin, S.H., Huang, J.Q., Li, X.Q., Zheng, B.S., Wu, J.S., Wang, Z.J., Liu, G.H. and Chen, M. 2011. Effects of potassium supply on limitations of photosynthesis by mesophyll diffusion conductance in *Carya cathayensis*. *Tree Physiology* 31:1142–1151.
- Jones, H. 2004. What is water use efficiency? In: *Water Use Efficiency in Plant Biology*; Bacon, M.A., Ed.; Blackwell: Oxford, UK, pp. 27–41.
- Jones, M.M., Turner, N.C. and Osmond, C.B. 1981. Mechanisms of drought resistance. In: *The Physiology and Biochemistry of Drought Resistance in Plants*; Paleg, L.G., Aspinall, D., Eds.; Academic Press: New York, NY, USA, pp. 15–35.
- Jones, R.G. and Pollard, A. 1983. Proteins Enzymes and Inorganic Ions. In *Inorganic Plant Nutrition*; Lauchli, A., Bileski, R.L., Eds.; Springer: New York, NY, USA, 528–562.
- Juenger, T.E., Sen, S., Bray, E., Stahl E., Wayne, T., McKay, J. and Richards, J.H. 2010 Exploring genetic and expression differences between physiologically extreme ecotypes: comparative genomic hybridization and gene expression studies of Kas-1 and Tsu-1 accessions of *Arabidopsis thaliana*. *Plant Cell Environment* 33:1268–1284.
- Kalai, T., Khamassi, K., Teixeira, da Silva J.A., Gouia, H. and Bettaieb, Ben-Kaab L. 2014. Cadmium and copper stress affect seedling growth and enzymatic activities in germinating barley seeds. *Archives of Agronomy and Soil Science* 60:25–37.
- Kanai, S., Moghaieb, R.E., El-Shemy, H.A., Panigrahi, R., Mohapatra, P.K., Ito, J., Nguyen, N.T., Saneoka, H. and Fujita, K. 2011. Potassium deficiency affects water status and photosynthetic rate of the vegetative sink in green house tomato prior to its effects on source activity. *Plant Sciences* 180:368–374.
- Kandil, A.A., Sultan, M.S., Badawi, M.A., Abd El-Rahman, A.A. and Zayed, B.A. 2010. Performance of rice cultivars as affected by irrigation and potassium fertilizer under saline soil conditions: I. Yield and yield Components Crop. *Journal of Environment* 1:18–21.
- Kant, S. and Kafkafi, U. 2002. Potassium and Abiotic Stresses in Plants. In *Potassium for Sustainable Crop Production*; Pasricha N.S., Bansal S.K., Eds.; Potash Institute of India: Gurgaon, India, 233–251.

- Kato, M. and Shimizu, S. 1985. Chlorophyll metabolism in higher plant. VI. Involvement of peroxidase in chlorophyll degradation. *Plant Cell Physiology* 26:1291–1301.
- Kaya, C., Tuna, A.L., Ashraf, M., Altunlu, H. 2007. Improved salt tolerance of melon (*Cucumis melo* L.) by the addition of proline and potassium nitrate. *Environmental Experimental Botany* 60:397–403.
- Kehoe, D.M. and Gutu, A. 2006. Responding to color: The regulation of complementary chromatic adaptation. *Annual Review of Plant Biology* 57:127–150.
- Khadr, M.S., Negm, A.Y., Khalil, F.A., Antoun, L.W. 2004. Effect of potassium chloride in comparison with potassium sulfate on sugar cane production and some soil chemical properties under Egyptian conditions. In IPI regional workshop on Potassium and Fertigation development in West Asia and North Africa; Rabat, Morocco, 24-28 November, 2004. Pp 1–8.
- Khan, A.W., Mann, R.A. and Saleem, M. 2012. Comparative rice yield and economic advantage of foliar  $KNO_3$  over soil applied  $K_2SO_4$ . *Pakistan Journal of Agriculture Science* 49 481–484.
- Khush, G.S. 2005. What it will take to Feed 5.0 Billion Rice consumers in 2030. *Plant Molecular Biology* 59:1–6.
- Kim, B.R., Nam, H.Y., Kim, S.U., Kim, S.I. and Chang, Y.J. 2003. Normalization of reverse transcription quantitative PCR with housekeeping genes in rice. *Biotechnology Letters* 25:1869–1872.
- Kirnak, H., Kaya, C., Tas, I. and Higgs, D. 2001. The influence of water deficit on vegetative growth, physiology, fruit yield and quality in eggplants. *Plant Physiology* 27: 34–46.
- Kirkby, E.A., LeBot, J., Adamowicz, S. and Römheld, V. 2009. Nitrogen in Physiology. In *An Agronomic Perspective and Implications for the Use of Different Nitrogen Forms*; International Fertiliser Society; Cambridge, York, UK, 653:1–48.
- Kobata, T. and Takami, S. 1986. Changes in respiration, dry-matter production and its partition in rice (*Oryza sativa* L.) in response to water deficits during the grain-filling period. *Botany Magazine Tokyo* 99:379–393.
- Kramer, P. J. 1983. Plant and soil water relationships (a modern synthesis). McGraw-Hill Publishing, New York, USA.
- Lambers, H., Robinson, S. A. and Ribas-Carbo, M. 2005. Regulation of respiration in vivo. In *Plant Respiration: From Cell to Ecosystem. Advances in Photosynthesis and Respiration Series*; Lambers H., and Ribas-Carbo M., Eds; Springer, The Netherlands, 18:1–15.



- Lampayan, R.M., Bouman, B.A.M., DeDios, J.L., Lactaen A.T., Espiritu, A.J., Norte, T.M., Quilang, E.J.P., Tabbal, D.F., Llorca, L.P., Soriano, J.B., Corpuz, A.A. and Malasa, R.B. 2005. Transfer of water saving technologies in rice production in the Philippines. In *Transitions in Agriculture for Enhancing Water Productivity*; Thiyagarajan, T.M., Hengsdijk, H., and Bindraban, P., Eds., Proceedings of the International Symposium on Transitions in Agriculture for Enhancing Water Productivity, September 23–25, 2003, Kilikulam, Tamil Nadu Agricultural University, Tamil Nadu, India, 111–132.
- Lapinski, J. and Tunnacliffe, A. 2003. Anhydrobiosis without trehalose in bdelloid rotifers, *FEBS Letters* 553:387–390.
- Lasnitzki, A. 1964. Potassium and carbohydrate metabolism. *Nature* 146: 99–100.
- Lawlor, D.W. and Cornic, G. 2002. Photosynthetic carbon assimilation and associated metabolism in relation to water deficits in higher plants. *Plant, Cell and Environment* 25:275–294.
- Lawlor, D.W. and Tezara, W. 2009. Causes of decreased photosynthetic rate and metabolic capacity in water-deficient leaf cells: a critical evaluation of mechanisms and integration of processes. *Annual Botany* 103:561–579.
- Lee, S.C., Lee, M.Y., Kim, S.J., Jun, S.H., An, G. and Kim, S.R. 2005. Characterization of abiotic stress-inducible dehydrin gene OsDhn1 in rice (*Oryza sativa* L.). *Molecules and Cells* 19:212–218.
- Levitt, J. 1980. Stress terminology. In: *Adaptation of Plants to Water and High Temperature Stress*; Turner, N.C.; Kramer, P.J., Eds.; Wiley: New York, NY, USA, pp. 473–439.
- Liang, Y., Hu, F., Yang, M. and Yu, J. 2013. Antioxidative defenses and water deficit-induced oxidative damage in rice (*Oryza sativa* L.) growing on non-flooded paddy soils with ground mulching. *Plant and Soil* 257: 407–416
- Lilley, J.M. and Fukai, S. 1994. Effect of timing and severity of water deficit on four diverse rice cultivars. II. Physiological responses to soil water deficit. *Field Crops Research* 37:215–223.
- Lima, A.L.S., Damatta, P.M., Pinheiro, H.A., Totola, M.R. and Loureiro, M.E. 2002. Photochemical responses and oxidative stress in two clones of *Coffea canephora* under water deficit conditions. *Environmental and Experimental Botany* 47:239 – 247.
- Lindhauer, M.G. 1995. Influence of K nutrition and drought on water relations and growth of sunflower (*Helianthus annuus* L.). *Plant Nutrition and Soil Sciences* 148:654–669.
- Lindhauer, M.G. 1985. Influence of K nutrition and drought and water stressed sunflower plants differing in K nutrition. *Journal of Plant Nutrition* 10:1965–1973.

- Liu, S.-H., Chen, G.X., Yin, J.J. and Lu, C.G. 2011. Response of the flag leaves of a super-hybrid rice variety to drought stress during grain filling period. *Journal of Agronomy. Crop Science* 197:322–328.
- Liu, S.P., Zheng, L.Q., Xue, Y.H., Zhang, Q.A., Wang, L. and Shou, H.X. 2010. Over expression of OsVP1 and OsNHX1 increases tolerance to drought and salinity in rice. *Journal Plant Biology* 53:444–452.
- Livak, K.J. and Schmittgen, T.D. 2001. Analysis of Relative Gene Expression Data Using Real-Time Quantitative PCR and the  $2^{-DDCT}$  Method. [[PubMed](#)] *Methods* 25:402–408.
- Ludlow, M.M. 1989. Strategies of response to water stress. In *Structural and Functional Responses to Environmental Stresses*. Edited by Kreeb KH, Hinckley HRTM. SPB Academic 269–281.25.
- Manunta, P., Grant, R.F. and Feng, Y. 2002. Changes in mass and energy transfer between the canopy and the atmosphere model development and testing with a free-air CO<sub>2</sub> enrichment (FACE) experiment. *International Journal of Biometeorology* 46(1):9–21.
- Maqsood, M., Shehzad ,M.A., Wahid, A. and Butt, A.A. 2013. Improving drought tolerance in maize (*Zea mays*) with potassium application in furrow irrigation Systems. *International Journal Of Agriculture & Biology*,15(6): 1193–1198.
- MARDI,2015.[http://agromedia.mardi.gov.my/magritech/tech\\_detail\\_fdcrop.php?id=34](http://agromedia.mardi.gov.my/magritech/tech_detail_fdcrop.php?id=34)
- Markgraf, W. and Horn, R., 2006. Rheological-stiffness analysis of K<sup>+</sup> treated and CaCO<sub>3</sub>-rich soils. *Plant Nutrition and Soil Science* 169:411–419.
- Marschner, H. and Cakmak, I. 1989. High light intensity enhances chlorosis and necrosis in leaves of zinc, potassium, and magnesium deficient bean (*Phaseolus vulgaris*) plants. *Plant Physiology* 134:308–315.
- Marschner P. 2012. Marschner's Mineral Nutrition of Higher Plants, 3rd ed.; Academic Press: London, UK, 178–189.
- Marschner, H., 1995. Mineral Nutrition of Higher Plants.2<sup>nd</sup> Ed. Academic Press, San Diego, California, USA.
- McDonald, A.J.S. and Davies, W.J. 1996. Keeping in touch: responses of the whole plant to deficits in water and nitrogen supply. *Advances in Botanical Research* 22:229–300.
- McKay, J.K., Richards, J.H., Nemali, K.S., Sen, S., Mitchell-Olds, T., Boles, S., Stahl, E.A., Wayne T. and Juenger, T.E. 2008. Genetics of drought adaptation in *Arabidopsis thaliana* II. QTL analysis of a new mapping population, Kas-1 Tsu-1. *Evolution* 62:3014–3026.

- Mensha, J.K., Obadoni, B.O., Eroutor, P.G., Onome, I.F. 2006. Simulated flooding and drought effects on germination, growth and yield parameters of Sesame (*Sesamum indicum* L.). *African Journal Biotechnology* 5:1249–1253.
- Mittler, R. 2006 Abiotic stress, the field and environment and stress combination. *Trends in Plant Science* 11:15–19.
- Mohammad Khani, N. and Heidari, R. 2008. Drought-induced accumulation of soluble sugars and proline in two maize varieties. *World Applied Science Journal*, 13:448–453.
- Mohd Zain N.A., Ismail M.R., Mahmood M., Puteh A. and Ibrahim M.H. 2014. Alleviation of water stress effects on MR220 rice by application of periodical water stress and potassium fertilization, *Molecules* 19:1795–1819.
- Mohiti, M., Ardalan, M.M., Mohammadi, T.A. and ShokriVahed, H. 2011. The efficiency of potassium fertilization methods on the growth of rice (*Oryza sativa* L.) under salinity stress. *African Journal of Biotechnology*. 10:15946–15952.
- Mohamed, A. A., Saleh, M. I and Adel, D. A. 2014. Effect of potassium soil and foliar spray fertilization on yield, fruit quality and nutrient uptake of ‘Seweda’ date palm grown in sandy loam soil. *Journal of Food, Agriculture and Environment* 12 (1): 305-311.
- Molden, D., Murray-Rust, H., Sakthivadivel, R., and Makin, I. 2003. A Water-Productivity Framework for Understanding and Action. *Water Productivity in Agriculture: Limits and Opportunities for Improvement*, Kijne J.W., Barker R. and Molden D. (eds), Comprehensive Assessment of Water Management in Agriculture Series, No 1. International Water Management Institute, Sri Lanka
- Molina, J., Sikora, M., Garud, N., Flowers, J. M., Rubinstein, S., Reynolds, A., Huang, P., Jackson, S., Schaal, B. A., Bustamante, C. D., Boyko, A. R. and Purugganan, M. D. 2011. Molecular evidence for a single evolutionary origin of domesticated rice. *Proceedings of the National Academy of Sciences* 108(20):8351.
- Moons, A. De Keyser, A. and Van Mantagou, M A.1997. Group of 3 LEA cDNA of rice, responsive to abscisic acid, but not to jasmonic acid, shows variety specific differences in salt stress response, *Gene* 191:197–204.
- Moore, J.P., Le, N.T., Brandt, W.F., Driouich, A. and Farrant, J.M. 2009. Towards a systems-based understanding of plant desiccation tolerance. *Trends Plant Sciences* 14:110–117.
- Moore, J.P., Vre-Gibouin, M., Farrant, J.M. and Driouich, A. 2008. Adaptations of higher plant cell walls to water loss: drought vs desiccation. *Physiology Plantarum* 134:237–245.



- Moumeni, A., Satoh, K., Kondoh, H., Asano, T., Hosaka, A., Venuprasad, R. Serraj, R. and Kikuchi, S. 2011. Comparative analysis of root transcriptome profiles of two pairs of drought-tolerant and susceptible rice near-isogenic lines under different drought stress. *BMC Plant Biology* 11:174–180.
- Murata, T. and Akazawa, T. 1968. Stimulative effect of potassium ion on starch synthetase of different plant origins. *Plant Cell Physiology* 10:457–460.
- Najafabadi, M.S. 2012. Improving rice (*Oryza sativa* L.) drought tolerance by suppressing a NF-YA. *Iran Journal of Biotechnology* 10:40–48.
- Nakamura, K., Ohto, M., Yoshida, N. and Nakamura, K. 1991. Sucrose induced accumulation of amylase occurs concomitant with the accumulation of starch and sporamin in leaf petiole cuttings of sweet potato. *Plant Physiology* 96:902–909.
- Nandwal, A.S., Hooda, A. and Datta, D., 1998. Effect of substrate moisture and potassium on water relations and C, N and K distribution in *Vignaradiata*. *Biologia Planatarium* 41:149–153.
- Naser, Z., Aynehband, A. and Lak S. 2012. Evaluation of physiological traits changes in drought stress, the application of potassium and their impact on the yield of mung bean cultivars and promising lines. *Advances Environment Biology* 6:23–241.
- Nasri, M. and Khalatbary, M., 2011. Effect of nitrogen fertilizer, potassium and zinc on quantitative and qualitative characteristics of green bean genotypes. *Journal of Crop Ecophysiology* 3:82-93.
- Nawaz, F., Ahmad, R., Waraich, E.A., Naeem, M.S. and Shabbir, R.N. 2012. Nutrient uptake, physiological responses, and yield attributes of wheat (*Triticum aestivum* L.) exposed to early and late drought stress. *Journal of Plant Nutrition*, 35: 961-974.
- Nguyen, G.N., Hailstones, D.L., Wilkes, M. and Sutton B.G. 2010. Role of carbohydrate metabolism in drought-induced male sterility in rice anthers. *Journal of Agronomy and Crop Science* 196: 346–357.
- Nilsen, E.T. and Orcutt, D.M. 1996. *Physiology of Plants under Stress, Abiotic Factors*. 2nd Ed., John Wiley and Sons Inc, New York, 689.
- Oddo, E., Inzerillo, S., La Bella, F., Grisafi, F., Salleo, S. and Nardini, A. 2011. Short-term effects of potassium fertilization on the hydraulic conductance of *Laurus nobilis* L. *Tree Physiology* 31:131–138.
- Oerke, E.C. and Dehne, H.W. 2004. Safeguarding production-losses in major crops and the role of crop protection. *Crop Protection* 23:275–285.
- Oka, H.I. 1988. *Origin of cultivated rice*. Elsevier, Amsterdam.

- Okuda, T., Matsuda, Y., Yamanaka, A. and Sagisaka, S. 1991. An abrupt increase in the level of hydrogen peroxide in leaves of winter wheat is caused by cold treatment. *Plant Physiology* 97:1265–1267.
- Ouvrard, O., Cellier, F. and Ferrare, K. 1995. Differential expression of water stress-regulated genes in drought tolerant or sensitive sunflower genotypes. In *Proceedings of the International Conference of Integrated Studies on Drought Tolerance of Higher Plants*, International Drought, vol.95.
- Pantuwan, G., Fukai, S., Cooper, M., Rajatasereekul, S. and O'Toole, J.C. 2000. Field screening for drought resistance. Increased lowland rice production in the Mekong Region. Proceeding International Workshop, 30 October-2 November, 2000, pp: 69–77. Vientiane, Laos.
- Pantuwan, G. 2001. Yield responses in rice (*Oryza sativa* L.) genotypes to water deficit in rain fed lowlands. Ph.D. Thesis. University of Queensland, Australia.
- Patel, A.L. and Singh, J. 1998. Nutrient uptake and distribution in aerial parts of wheat under water stress at different growth stages. *Annals of Biological Research* 3:5-8.
- Pattanagul, W. 2011. Exogenous abscisic acid enhances sugar accumulation in rice (*Oryza sativa* L.) under drought stress. *Asian Journal Plant Science* 10:212–219.
- Pedersen, O., Rich, S.M. and Colmer, T.D. 2009. Surviving floods: Leaf gas films improve O<sub>2</sub> and CO<sub>2</sub> exchange, root aeration, and growth of completely submerged rice. *Plant Journal* 58:147–156.
- Pel, Z.M., Murata, Y. and Benning, G. 2000. Calcium channels activated by hydrogen peroxide mediate abscisic acid signaling in guard cells. *Nature* 406:731–734.
- Peleg, Z. and Blumwald, E. 2011. Hormone balance and abiotic stress tolerance in crop plants. *Current Opinion in Plant Biology* 14:290–295.
- Peng, Y.H., Zhu, Y.F., Mao, Y.Q., Wang, S.W., Su, W.A., Tang, Z.C. 2004. Alkali grass resists salt stress through high [K<sup>+</sup>] and an endodermis barrier to Na<sup>+</sup>. *Journal of Experimental Botany* 55:939–949.
- Percival, G.C., Fraser, G.A. and Oxenham, G. 2003. Foliar salt tolerance of *Acer* genotypes using chlorophyll fluorescence. *Journal Arboriculture* 29:61–65.
- Preek, A., Sopory, S.K., Bahnert, H.J and Govindjee. 2010. Abiotic stress adaptation in plants. *Physiological, molecular and genomic foundation*. Springer, The Netherland, pp. 7.
- Pervez, H., Makhdum, M.I. and Ashraf, M. 2006. The interactive effects of potassium nutrition on the uptake of other nutrients in cotton (*Gossypium hirsutum* L.) under an arid environment. *Chemical Society of Pakistan* 28:256–265.

- Pervez, H., Ashraf, M. and Makhdom, M.I. 2004. Influence of potassium nutrition on gas exchange characteristics and water relations in cotton (*Gossypium hirsutum* L.). *Photosynthetica* 42:251–255.
- Pier, P.A. and Berkowitz, G.A. 1987. Modulation of water stress effects on photosynthesis by altered leaf  $K^+$ . *Plant Physiology* 85:65–661.
- Plaxton, W. and Podesta, F. 2006. The functional organization and control of plant respiration. *Critical Review in Plant Sciences* 25:159–198.
- Postel, S. 1997. *Last Oasis: Facing Water Scarcity*. Norton and Company, New York, pp. 239.
- Pouchkina-Stantcheva, N.N., McGee, B.M., Boschetti, C., Tolleter, D., Chakrabortee, S., Popova, A.V., Meersman, F., Macherel, D., Hinch, D.K. and Tunnacliffe A. 2007. Functional divergence of former alleles in an ancient asexual invertebrate. *Science* 318: 268–271.
- Premachandra, G.S., Saneoka, H. and Ogata, S. 1991. Cell membrane stability and leaf water relations as affected by potassium nutrition of water-stressed maize. *Experimental Botany* 42:739–745.
- Premachandra, G.S., Saneoka, H., Fufita, K. and Ogata, S. 1992. Leaf water relations, osmotic adjustment, cell membrane competence, epicuticular wax load and growth as affected by increasing water deficits in sorghum. *Journal Experimental Botany* 43:1569–1576.
- Price, A.H. and Tomos, A.D. 1997. Genetic dissection of root growth in rice (*Oryza sativa* L.). II. Mapping quantitative trait loci using molecular markers. *Theoretical Applied Genetics* 95: 143–152.
- Pyingrope, S., Bhoomika, K. and Dubey, R.S. 2013. Reactive oxygen species, ascorbate-glutathione pool, and enzymes of their metabolism in drought-sensitive and tolerant indica rice (*Oryza sativa* L.) seedlings subjected to progressing levels of water deficit. *Protoplasma* 250:585–600.
- Quampah, A., Wang, R.M., Shamsil, I.H., Jilanil, G., Zhang, Q., Hua, S. and Xu, H. 2011. Improving Water Productivity by Potassium Application in Various Rice Genotypes. *International Journal of Agriculture Biology* 13:1560–1853.
- Rabello, A.R., Guimaraes, C. and Range, I. P.H.N. 2008. Identification of drought responsive genes in roots of upland rice (*Oryza sativa* L.). *BMC Genomics* 9:485–498.
- Raghavendra, A.S. and Padmasree, K. 2003. Beneficial interactions of mitochondrial metabolism with photosynthetic carbon assimilation. *Trends Plant Sciences* 8:546–553.

- Ramamoorthy, R., Jiang, S.-Y., Kumar, N. and Venkatesh, P.N. 2008. A comprehensive transcriptional profiling of the WRKY gene family in rice under various abiotic and phytohormone treatments. *Plant Cell Physiology* 49:865–879.
- Razak, A.A., Ismail, M.R., Karim, M.F., Megat, Wahab, P.E., Abdullah, S.N. and Kausar, H. 2013. Changes in leaf gas exchange, biochemical properties, growth and yield of chilli grown under soilless culture subjected to deficit fertigation. *Australian Journal of Crop Science* 7:1582–1589.
- Ren, Z.H., Zheng, Z.M., Chinnusamy, V., Zhu, J.H., Cui, X.P., Iida, K. and Zhu, J.K. 2010. RAS1, a quantitative trait locus for salt tolerance and ABA sensitivity in Arabidopsis. *Proceeding of the National Academy Sciences* 107:5669–5674.
- Ribas-Carbo, M., Taylor, N.L. and Gilesetal, L. 2005. Effects of water stress on respiration in soybean leaves. *Plant Physiology* 139:466–473.
- Rijsberman, F.R. 2006. Water scarcity: Fact or fiction? *Agriculture Water Management* 80:5–22.
- Rizhsky, L., Liang, H. and Mittler, R. 2002. The combined effect of drought stress and heat shock on gene expression in tobacco. *Plant Physiology* 130:1143–1151.
- Rodriguez, M.C.S., Edsgard, D., Hussain, S.S., Alquezar, D., Rasmussen, M., Gilbert, T., Nielsen, B.H., Bartels, D. and Mundy, J. 2010. Transcriptomes of the desiccation-tolerant resurrection plant *Craterostigma plantagineum*. *Plant Journal* 63:212–228.
- Roitsch, T. 1999. Source-sink regulation by sugar and stress. *Current Opinion in Plant Biology*, 2: 198–206.
- Romheld, V. and Kirkby, E.A. 2010. Research on potassium in agriculture: Needs and prospects. *Plant Soil* 335:155–180.
- Rong-hua, L., Pei-pol, G., Baumz, M., Grando, S. and Ceccarelli, S. 2006. Evaluation of chlorophyll content and fluorescence parameters as indicators of drought tolerance in barley. *Agriculture Science China* 5:751–757.
- Rose, T.J., Rengel, Z., Ma, Q. and Bowden, J.W. 2008. Post-flowering supply of P, but not K, is required for maximum canola seed yields. *Journal of Agronomy* 28:371–379.
- Rosenberg, N.J., Kimball, B.A., Martin, P. and Cooper, C.F. 1990. From climate and CO<sub>2</sub> enrichment to evapotranspiration. Climate Change and US Water Resources, P.E. Waggoner, Ed., pp. 151–175.
- Rouhier, N., Vieira Dos Santos C., Tarrago L., and Rey P. 2006. Plant methionine sulfoxide reductase A and B multigenic families. *Photosynthesis Research* 89:247–262.

- Ruan, J., Ma, L. and Shi, Y. 2013. Potassium management in tea plantations: Its uptake by field plants, status in soils, and efficacy on yields and quality of teas in China. *Journal of Plant Nutrition Soil Science* 176: 450-45.
- Sade, N., Alem, G., and Moshelion, M. 2012. Risk-taking plants. An isohydric behavior as a stress resistance trait. *Plant Signalling Behavior* 7:1-4.
- Salekdeh, G.H., Siopongco, H.J., Wade, L.J., Ghareyazie, B., and Bennett, J. 2002. A proteomic approach to analyzing drought- and salt-responsiveness in rice. *Field Crops Research* 76:199-219.
- Sanada, Y., Ueda, H., Kuribayashi, K., Andoh, T., Hayashi, F., Tamai, N. and Wada, K. 1995. Novel light-dark change of proline levels in halophyte (*Mesembryanthemum crystallinum* L.) and glycophytes (*Hordeum vulgare* L. and *Triticum aestivum* L.): Leaves and roots under salt stress. *Plant Cell Physiology* 36:65-970.
- Sangakkara, U.R., Frehner, M. and Nosberger, J. 2001. Influence of soil moisture and fertilizer potassium on the vegetative growth of mungbean (*Vigna radiate* L. Wilczek) and cowpea (*Vigna unguiculata* L. Walp). *Journal of Agronomy and Crop Science* 186:73-81.
- Sarkarung, S., Pantuwan, G., Pushpavesa, S. and Tanupan, P. 1997. Germplasm development for rainfed lowland ecosystems: breeding strategies for rice in drought-prone environments. Proceeding International Workshop, Ubon Ratchathani, Thailand. pp: 43-49.
- Sarvestani, Z.T., Pirdashti, H., Sanavy, S.A. and Balouchi, H. 2008. Study of water stress effects in different growth stages on yield and yield components of different rice (*Oryza sativa* L.) cultivars. *Pakistan Journal of Biological Science* 11:1303-1309.
- Savary, S., Horgan, F., Willocquet, L. and Heong, K.L. 2012. A review of principles for sustainable pest management in rice. *Crop Protection* 32:54
- Sengupta, A., Berkowitz, G.A. and Pier, P.A. 1989. Maintenance of photosynthesis at low leaf water potential in wheat. *Plant Physiology* 89:1358-1365.
- Sengupta, S. and Majumder, A.L. 2009. Insight into the salt tolerance factors of wild halophytic rice, *Porteresia coarctata*: A physiological and proteomic approach. *Planta* 229: 911-929.
- Shabala, S.N. and Lew, R.R. 2002. Turgor regulation in osmotically stressed *Arabidopsis* epidermal root cells. Direct support for the role of inorganic ion uptake as revealed by concurrent flux and cell turgor measurements. *Plant Physiology* 129:290-299.
- Shao, H.B., Chu, L.Y., Lu, Z.H., and Kang, C. M. 2008. Primary antioxidant free radical scavenging and redox signaling pathways in higher plant cells. *International Journal of Biological Science* 4: 8-14.



- Sharma, R.C. 1995. Tiller mortality and its relationship to grain yield in spring wheat. *Field Crops Research*, 41(1): 55–60.
- Sharma, K.D., Nandwal, A.S. and Kuhad, M.S. 1996. Potassium effects on CO<sub>2</sub> exchange, NRA and yield of clusterbean cultivars under water stress. *Journal of Potassium Research* 12: 412–423.
- Shih, M., Lin, S., Hsieh, J., Tsou, C., Chow, T., Lin, T and Hsing, Y. 2004. Gene cloning and characterization of a soybean (*Glycine max* L.) LEA protein, GmPM16. *Plant Molecular Biology* 56:689–703.
- Shin, R. and Schachtman, D.P. 2004. Hydrogen peroxide mediates plant root cell response to nutrient deprivation. *Proceeding of National Academy Science USA* 101:8827–8832.
- Shukla, N., Awasthi, R.P., Rawat, L. and Kumar, J. 2012. Biochemical and physiological responses of rice (*Oryza sativa* L.) as influenced by *Trichoderma harzianum* under drought stress. *Plant Physiology Biochemistry* 54:78–88.
- Siddiq, M.N. and Viraktamath, H.I. 2001. Rice. In *Breeding Field Crops*; Chopra, V.L. Ed.: Oxford and IBH Publishing Co. Pvt. Ltd., New Delhi, India, pp. 1–85.
- Siddiqui, M.H., Al-Whaibi, M.H., Sakran, A.M., Basalah, M.O. and Ali, H.M. 2012. Effect of calcium and potassium on antioxidant system of *Vicia faba* L. under cadmium stress. *International Journal of Molecular Sciences* 13: 6604–661.
- Siedow, J.N. and Umbach, A.L. 2000 The mitochondrial cyanide-resistant oxidase: structural conservation amid regulatory diversity. *Biochimica et Biophysica Acta* 1459: 432–439.
- Sikuku, P.A., Netondo, G.W., Onyango, J.C. and Mysyimi, D.M. 2010. Chlorophyll fluorescence, protein and chlorophyll content of three rainfed rice varieties under varying irrigation regimes. *ARPJ Journal of Agriculture and Biology Science* 5:19–25.
- Singh, V.K., Dwivedi, B.S., Buresh, R.J., Jat, M.L., Majumdar, K., Gangwar, B., Govil, V., and Singh, S.K. 2013. Potassium fertilization in rice-wheat system across northern India: Crop performance and soil nutrients. *Agronomy Journal* 105: 471–481.
- Siringam, K., Juntawong, N., Cha-um, S. and Kirdmanee, C. 2013. Exogenous application of potassium nitrate to alleviate salt stress in rice seedlings. *Journal of Plant Nutrition* 36: 607–616.
- Sritontip, C., Khaosumain, Y., Changjeraja, S., Changjeraja, R. 2008. Effects of light intensity and potassium chlorate on photosynthesis and flowering in 'Do' longan. *Acta Horticulturae* 787:285–288.

- Steinberg, S.L., Miller J.C. and McFarland M.J. 1990. Dry matter partitioning and vegetative growth of young peach trees under water stress. *Australian Journal Plant Physiology* 17: 6–23.
- Stiller, V., Lafitte, H.R. and Sperry, J.S. 2003. Hydraulic properties of rice and the response of gas exchange to water stress. *Plant Physiology* 132:1698–1706.
- Su, D., Chen, N., Gao, T., Wang, C., Sheng, M. and Yang, C. 2012. Effects of Si<sup>+</sup>, K<sup>+</sup>, and Ca<sup>2+</sup> on antioxidant enzyme activities and osmolytes in *Halocnemum strobilaceum* under salt stress. *Advanced Materials Research* 356:2542–2550.
- Sudama, S., Tiwari, T.N., Srivastava, R.P., Singh G.P., and Singh S. 1998. Effect of potassium on stomatal behavior, yield and juice quality of sugarcane under moisture stress condition. *Indian Journal of Plant Physiology* 3:303–305.
- Szegletes, Z.S., Erdei, L., Tari, I. and Cseuz, L. 2000. Accumulation of osmoprotectants in wheat cultivars of different drought tolerance. *Cereal Research Communications* 28:403–410.
- Taira, M., Valtersson, U., Burkhardt, B. and Ludwig, R.A. 2004. *Arabidopsis thaliana* GLN2-encoded glutamine synthetase is dual targeted to leaf mitochondria and chloroplasts. *Plant Cell* 16:2048–2058.
- Taiz, L. and Zeiger, E. 2010. *Plant Physiology*, 5th edition. Sinauer Associates Inc. Publishers, Sunderland, Massachusetts, USA.
- Tatari M., Ghazvini R.F., Etemadi N., Ahadi A.M. and Mousavi A. 2012. Analysis of antioxidant enzymes activity, lipid peroxidation and proline content of *Agropyron desertorum* under drought stress. *South Western Journal of Horticulture, Biology and Environment* 3:9–24.
- Teixeira, J. and Pereira, S. 2007. High salinity and drought act on an organ-dependant manner on potato glutamine synthetase expression and accumulation. *Environmental Experiment Botany* 60:121–126.
- Tezera, W., Mitchell, V., Driscoll, S.P. and Lawlor, D.W. 2002. Effects of water deficit and its interaction with CO<sub>2</sub> supply on the biochemistry and physiology of photosynthesis in sunflower. *Journal of Experimental Botany*. 53:1781–1791.
- Thomashow, M.F. 1999. Plant cold acclimation: Freezing tolerance genes and regulatory mechanisms. *Annual Review Plant Physiology and Plant Molecular Biology* 50: 571–599.
- Tiwari, H.S., Agarwal, R.M. and Bhatt, R.K. 1998. Photosynthesis, stomatal resistance and related characters as influenced by potassium under normal water supply and water stress conditions in rice (*Oryza sativa* L.). *Indian Journal Plant Physiology* 3: 314–316.



- Tomemori, H., Hamamura, K. and Tanabe, K. 2002. Interactive effects of sodium and potassium on the growth and photosynthesis of spinach and komatsuna. *Plant Production Sciences* 5:281–285.
- Tompa, P., Banki, P., Bokor, M., Kamasa, P., Kovacs, D., Lasanda, G., and Tompa, K. 2006. Protein-water and protein-buffer interactions in the aqueous solution of an intrinsically unstructured plant dehydrin: NMR intensity and DSC aspects. *Biophysical Journal* 91:2243–2249.
- Tran, L.S.P. and Mochida, K. 2010. Functional genomics of soybean for improvement of productivity in adverse conditions. *Functional Integrative Genomics* 10:447–462.
- Tripathi, B.N., Bhatt, I. and Dietz, K.J. 2009. Peroxiredoxins: A less studied component of hydrogen peroxide detoxification in photosynthetic organism. *Protoplasma* 2:3–15.
- Tsonev, T., Velikova, V., Yildiz-Aktas, L., Gurel A. and Edreva, A. 2011. Effect of water deficit and potassium fertilization on photosynthetic activity in cotton plants. *Plant Biosystem* 145:841-847.
- Tuna, A.L., Kaya, C. and Ashraf, M. 2010. Potassium sulfate improves water deficit tolerance in melon plants grown under glasshouse conditions. *Nutrition* 33:1276–1286.
- Tunnacliffe, A., Lapinski, J. and McGee, B. 2005. A putative LEA protein, but no trehalose, is present in anhydrobiotic bdelloid rotifers. *Hydrobiologia* 546:315–321.
- Tuong, T.P., and Bouman, B.A.M. 2003. Rice production in water scarce environments. In *Water Productivity in Agriculture: Limits and Opportunities for Improvement* (Kijne, J.W., Barker, R. and Molden D., Eds.), CABI Publishing, Wallingford, UK. pp.53–67.
- Turner, N.C. 1981. Techniques and experimental approaches for the measurement of plant water stress. *Journal of Plant Soil* 58:339–366.
- Turner, N.C., O’Toole, J.C., Cruz, R.T., Yambao, E.B., Ahmad, S., Namuco, O.S., and Dingkuhn, M. 1986. Responses of seven diverse rice cultivars to water deficits: II. Osmotic adjustment, leaf elasticity, leaf extension, leaf death, stomatal conductance and photosynthesis. *Field Crops Research* 13:273–286.
- Turner, N.C., O’Toole, J.C. Cruz, R.T., Namuco, O.S. and Ahmad, S. 1986. Responses of seven diverse rice cultivars to water deficits. I. Stress development, canopy temperature, leaf rolling and growth. *Field Crops Research* 13:257–271.
- Umar, S. 2006. Alleviating adverse effects of water stress on yield of sorghum, mustard and groundnut by potassium application. *Pakistan Journal of Botany* 38:1373–1380.

- Valliyodan, B. and Nguyen, H.T. 2006. Understanding regulatory networks and engineering for enhanced drought tolerance in plants. *Current Opinion in Plant Biology* 9:189–195.
- Van Aken, O., Whelan, J. and Van Breusegem, F. 2010. Prohibitins: mitochondrial partners in development and stress response. *Trends in Plant Sciences* 15:275–282.
- Verbruggen, N. and Hermans, C. 2008. Proline accumulation in plants: A review. *Amino Acids* 35:753–759.
- Wagner, S., Cattle, S.R. and Scholten, T. 2007. Soil-aggregate formation as influenced by clay content and organic-matter amendment. *Plant Nutrition and Soil Science* 170:173–180.
- Wang, X.S., Zhu, H.B., Jin, G.L., Liu, H.L., Wu, W.R. and Zhu, J. 2007. Genome-scale identification and analysis of *LEA* genes in rice (*Oryza sativa* L.). *Plant Sciences* 172:414–420.
- Wang, Z.L. and Huang, B.R. 2004. Physiological recovery of kentucky bluegrass from simultaneous drought and heat stress. *Crop Sciences* 44:1729–1736.
- Werner, T., Nehnevajova, E., Keollmera, I., Novakb, O., Strnadb, M., Kreamerc, U. and Schmeullinga, T. 2010. Root-specific reduction of cytokinin causes enhanced root growth, drought tolerance, and leaf mineral enrichment in *Arabidopsis* and Tobacco. *Plant Cell* 22:3905–3920.
- Wise, M.J. 2003. Leaping to conclusions: a computational reanalysis of late embryogenesis abundant proteins and their possible role. *BMC Bioinformatics* 4:52–58.
- Wise, M.J. and Tunnacliffe, A. 2004. POPP the question: What do LEA proteins do? *Trends Plant Science* 9:13–17.
- Witham, F.A, Blaydes, D.F. and Devlin, R.M. 1986. Chlorophyll Absorption Spectrum and Quantitative Determination: Exercise in Plant Physiology. PWS Publishers, Boston. pp: 128–131.
- Wolf, B. 1982. A comprehensive system of leaf analysis and its use for diagnosing crop nutrient status. *Communication in Soil Sciences and Plant Analysis* 13:1035–1059.
- Wolkers, W.F., McCready, S., Brandt, W.F., Lindsey, G.G., and Hoekstra, F. 2001. Isolation and characterization of a D-7 LEA protein from pollen that stabilizes glasses in vitro, *Biochimica Biophysica Acta* 1544:196–206.
- Wopereis, M.C.S., Krop, V M.J., Maligaya, A.R., and Tuong, T.P. 1996. Drought-stress responses of two lowland rice cultivars to soil water status. *Field Crops Research* 46:21–39.

- Xiang, Y., Huang, Y. and Xiong, L. 2007. Characterization of stress-responsive CIPK genes in rice for stress tolerance improvement. *Plant Physiology* 144:1416–1428.
- Xiao, B., Huang, Y., Tang, N. and Xiong, L. 2007. Over expression of a *LEA* gene in rice improves drought resistance under the field conditions. *Theoretical and Applied Genetic* 115:35–46.
- Xiong, L., Wang, R.G., Mao, G. and Koczan, J.M. 2006. Identification of drought tolerance determinants by genetic analysis of root response to drought stress and abscisic acid. *Plant Physiology*. 142:1065–1074.
- Xiong, L., Schumaker, K.S and Zhu, J.K. 2002. Cell signalling during cold, drought, and salt stress. *Plant Cell* 14:165–183.
- Xu, D.Q., Huang, J., Guo, S.Q., Yang X., Bao, Y.M., Tang, H.J. and Zhang, H.S. 2008. Overexpression of a TFIIIA-type zinc finger protein gene ZFP252 enhances drought and salt tolerance in rice (*Oryza sativa* L.). *FEBS Letters* 582:1037–1043.
- Xu, Y.W., Zou, Y.T., Husaini, A.M., Zeng, J.W., Guan, L.L., Liu, Q. and Wu, W. 2011. Optimization of potassium for proper growth and physiological response of *Houttuynia cordata* Thunb. *Environment Experimental Botany* 71:292–295.
- Xue, G.P., McIntyre, C.L., Jenkins, C.L., Glassop, D., A.F., van Herwaarden and Shorter, R. 2008. Molecular dissection of variation in carbohydrate metabolism related to water-soluble carbohydrate accumulation in stems of wheat. *Plant Physiology* 146:441–454.
- Yadav, D.S., Goyal, A.K. and Vats, B.K. 1999. Effect of potassium in *Eleusine coracana* (L.) Gaertn. under moisture stress conditions. *Journal of Potassium Research* 15, 131–134.
- Yamada, M., Hidaka, T. and Fukamachi, H. 1996. Heat tolerance in leaves of tropical fruit crops as measured by chlorophyll fluorescence. *Scientia Horticulturae* 67:39–48.
- Yamakawa, H., Hirose, T., Kuroda, M. and Yamaguchi, T. 2007. Comprehensive expression profiling of rice grain filling-related genes under high temperature using DNA microarray. *Plant Physiology* 144:258–277.
- Yang, J. and Zhang, J. 2006. Grain filling under soil drying. *New Phytologist* 169:223–36.
- Yang, S., Vanderbeld B., Wan, J. and Huang, Y. 2010. Narrowing down the targets: towards successful genetic engineering of drought tolerant crops. *Molecular Plant* 3:469–490.
- Yooyongwech, S., Chaum, S. and Supaibulwatana K. 2013. Water relation and aquaporin genes (PIP1; 2 and PIP2;1) expression at the reproductive stage of

rice (*Oryza sativa* L. spp. indica) mutant subjected to water deficit stress. *Plant OMICS* 6:79–85.

- Yordanov, I., Velikova, V., and Tsone, v.T. 2000. Plant responses to drought, acclimation, and stress tolerance. *Photosynthetica* 38(2):171–186.
- Yu, X.Z., Zhang, F.Z. and Li, F. 2012. Phytotoxicity of thiocyanate to rice seedlings. *Bulletin of Environment. Contamination and Toxicology* 88:703–706.
- Zhang F., Tiyip T., Ding J.L., Taff, G.N. and He Q.S. 2009. The effects of the chemical components of soil salinity on electrical conductivity in the region of the delta oasis of Weigan and Kuqa Rivers, China. *Agricultural Science in China* 8:985–993.
- Zhang, S.W., Li, C.H., Cao, J., Zhang, Y.C., Zhang, S.Q., Xia, Y.F., Sun, D.Y. and Sun, Y. 2009. Altered architecture and enhanced drought tolerance in rice via the down-regulation of indole-3-Acetic Acid by TLD1/OsGH3.13 activation. *Plant Physiology* 151:1889–1901.
- Zhang, W. and Song, S.T. 1989. Irrigation model of water-saving high yield at lowland paddyfield. In “International Commission on Irrigation and Drainage, Seventh Afro-Asian Regional Conference.” October 15–25, 1989. Tokyo, Japan, Vol. I–C, 480–496.
- Zheng, J., Fu, J. and Gou, M. 2010. Genome wide transcriptome analysis of two maize inbred lines under drought stress. *Plant Molecular and Biology* 72:407–442.
- Zheng, X., Chen, B., Lu, G. and Han, B. 2009. Overexpression of a NAC transcription factor enhances rice drought and salt tolerance. *Biochemistry and Biophysics Research Communication* 379:985–989.
- Zhu J. K. 2002. Salt and drought stress signal transduction in plants. *Annual Review of Plant Biology* 53:247–273.
- Zhu, Q.H., Ramm, K., Shivakkumar, R., Dennis, E.S. and Upadhyaya, N.M. 2004. The anther indehiscence 1 gene encoding a single MYB domain protein is involved in anther development in rice. *Plant Physiology* 135:1514–1525.
- Zulkarnain, W.M., Razi, M.I., Ashrafuzzaman, N., Halimi, M.S. and Ismail, C.H. 2009. Rice growth and yield under rain shelter house as influenced by different water regimes. *International Journal of Agriculture and Biology* 11:556–560.