



UNIVERSITI PUTRA MALAYSIA

***MODULATION OF RECEPTOR FOR ADVANCED GLYCATION END
PRODUCTS SIGNAL TRANSDUCTION PATHWAY AS THERAPEUTIC
OPTION FOR MALARIA THERAPY***

CHUAH YAW KUANG

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By

CHUAH YAW KUANG

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in
Fulfillment of the Requirements for the Degree of Master of Science

April 2015

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of
the requirement for the degree of Master of Science

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April 2015

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Receptor for advanced glycation endproducts (RAGE), an important receptor in the regulation of innate immune response, has been associated with many inflammatory related diseases such as septicaemia, rheumatoid arthritis, and arteriosclerosis. Malaria is also considered as an inflammatory disease involving excessive inflammatory response towards parasite invasion and severe systemic inflammation occurred during the infection has been closely linked to morbidity and mortality of the disease. However, RAGE involvement during malaria infection has yet to be revealed. In this study, the role and involvement of RAGE during malaria infection was investigated and the effects of modulating RAGE on the course of the infection, the release of major inflammatory cytokines and the histopathological consequences in major affected organs during malaria were evaluated. *Plasmodium berghei* (*P. berghei*) ANKA infection in male ICR mice was used as a model for malaria infection. The mice were inoculated intraperitoneally with 2×10^7 parasite-infected red blood cells (PRBCs) whereas the control mice received an equivalent dilution of normal RBCs. The plasma levels of RAGE in malarial mice were measured by ELISA. Results showed that RAGE was upregulated during malaria especially at the late critical phase of infection and there is a positive correlation between RAGE concentration and parasitaemia development suggesting that RAGE could be one of the important factors in mediating the severity of the infection.

Modulation of RAGE expression was carried out by treatment of malarial mice with recombinant mouse RAGE Fc chimera (rmRAGE/Fc Chimera) or mouse RAGE polyclonal antibody (mRAGE/pAb) intravenously. Both treatments did not affect the parasitaemia development during malaria infection. Blocking RAGE signaling pathway during the infection period significantly result in an elevation in the plasma levels of interleukin (IL)-4 and IL-17A, a further increase in IL-10 and IL-2 plasma levels, and reduced secretion of interferon (IFN)- γ in the plasma. But no effect on the release of tumor necrosis factor (TNF)- α and IL-6 was observed. Histopathological examination was performed on five major organs affected during malaria including liver, spleen,

brain, kidney, and lung. The results showed that modulation of RAGE expression improve the histopathological conditions of malaria to some degree. Both treatment groups showed an overall better outcome in histopathological conditions of all five organs despite the lack of effect on the course of the parasitaemia. In conclusion, the findings from this study showed that RAGE is involved during immune response towards malaria infection and blocking of RAGE may prove beneficial by reducing tissue injury to a lesser degree. Hence, this suggests the potential of RAGE as an immunotherapeutic target in malaria, in which the host may benefit from its inhibition.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

**MODULASI RECEPTOR FOR ADVANCED GLYCATION END PRODUCTS
SEBAGAI SASARAN TERAPEUTIK UNTUK TERAPI MALARIA**

Oleh

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Receptor for Advanced glycation endproducts (RAGE), suatu reseptor penting dalam pengawalaturan gerakbalas imun semulajadi, telah dikaitkan dengan banyak penyakit berkaitan inflamasi seperti septisemia, artritis reumatoïd dan arteriosklerosis. Malaria juga dianggap sebagai suatu penyakit inflamasi melibatkan gerakbalas inflamasi yang berlebihan terhadap pencerobohan parasit dan inflamasi sistemik tenat yang berlaku semasa jangkitan telah dikaitkan secara rapat dengan morbiditi dan mortaliti penyakit. Walau bagaimanapun, penglibatan RAGE semasa jangkitan malaria belum lagi dirungkaikan. Dalam kajian ini, peranan dan penglibatan RAGE semasa jangkitan malaria diselidiki dan kesan-kesan modulasi RAGE ke atas keadaan jangkitan, pembebasan sitokin inflamasi utama dan kesan histopatologi dalam organ-organ utama yang terkesan semasa jangkitan dinilai. Jangkitan *Plasmodium berghei* (*P. berghei*) ANKA dalam mencit ICR jantan telah digunakan sebagai model bagi jangkitan malaria. Mencit diinokulasi secara intraperitoneum dengan 2×10^7 sel-sel darah terjangkit parasit, manakala mencit kawalan menerima pencairan setara sel-sel darah normal. Tahap plasma RAGE dalam mencit malaria diukur menggunakan ELISA. Keputusan menunjukkan bahawa RAGE meningkat dalam mencit malaria pada fasa kritikal akhir jangkitan dan terdapat korelasi positif antara kepekatan RAGE dan perkembangan parasitaemia, yang mencadangkan RAGE mungkin salah satu faktor penting dalam memperantarakan jangkitan yang tenat.

Modulasi ekspresi RAGE dijalankan dengan merawat mencit malaria dengan RAGE Fc kimera mencit rekombinan (rmRAGE/Fc Chimera) atau antibodi poliklonal mencit (mRAGE/pAb) secara intravena. Kedua-dua rawatan tidak memberikan kesan ke atas perkembangan parasitaemia semasa jangkitan malaria. Merencat RAGE semasa jangkitan menyebabkan peningkatan secara signifikan interleukin-4 dan IL-17A pada tahap plasma, meningkatkan lagi tahap plasma IL-10 dan IL-2, dan mengurangkan pembebasan IFN- γ dalam plasma. Tetapi tiada kesan ke atas TNF- α dan IL-6 diperhatikan. Pemeriksaan histopathologi telah dijalankan ke atas lima organ utama yang terkesan semasa jangkitan malaria termasuk hati, limpa, otak, ginjal dan paru-paru. Keputusan menunjukkan modulasi ekspresi RAGE mampu memperbaiki keadaan

histopatologi malaria. Kedua-kedua kumpulan rawatan menunjukkan hasil keseluruhan yang lebih baik ke atas keadaan histopatologi kelima-lima organs walaupun tiada kesan ke atas parasitemia. Kesimpulannya, dapatan dari kajian ini menunjukkan bahawa RAGE terlibat semasa gerakbalas imun terhadap jangkitan malaria dan merencat RAGE mungkin berfaedah dengan mengurangkan kecederaan tisu ke tahap lebih rendah. Jadi, ini mencadangkan potensi RAGE sebagai sasaran imunoterapeutik dalam malaria, di mana hos mungkin mendapat faedah dari perencatannya.

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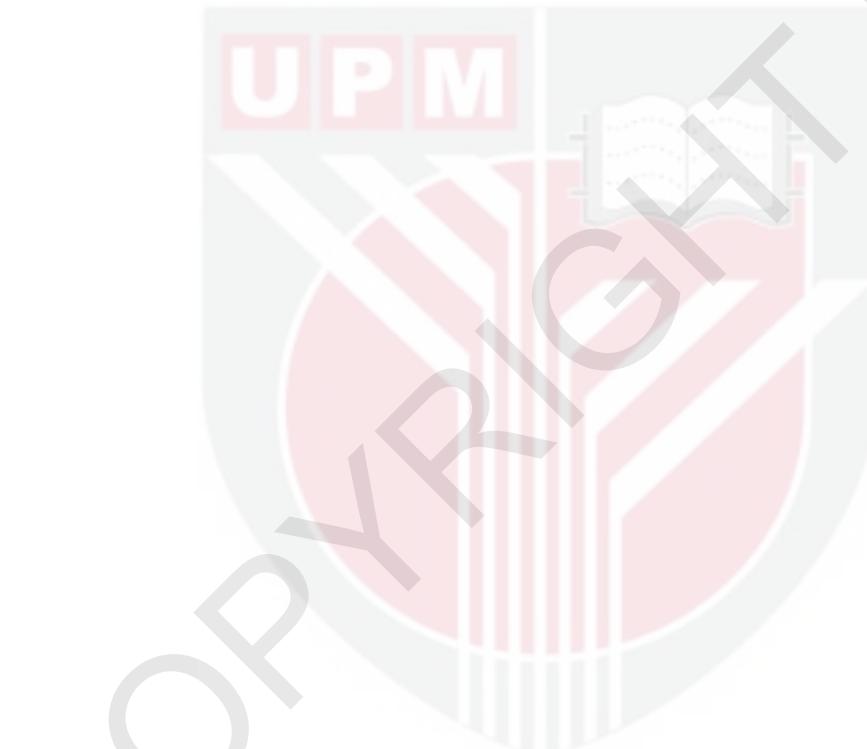
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LIST OF ABBREVIATIONS

ADCI	antibody-dependent cellular inhibition
ALI	acute lung injury
ANOVA	one-way analysis of variance
APCs	antigen-presenting cells
ARDS	acute respiratory distress syndrome
B cells	B lymphocyte cells
CBA	cytometric bead array
cRAGE	cleaved RAGE
CTL	cytotoxic T cells
DCs	dendritic cells
DNA	deoxyribonucleic acid
Na ₂ HPO ₄	disodium hydrogen phosphate anhydrous
ELISA	enzyme-linked immunosorbent assay
esRAGE	endogenous secretory RAGE
<i>et al.</i>	and others
fl-RAGE	full length, membrane-bound form RAGE
GPI	glycosylphosphatidylinositol
GM-CSF	granulocytes macrophage-colony stimulating factor
Ig	immunoglobulin
ICAM-1	intercellular adhesion molecule-1
IFN-γ	interferon-gamma
IL-	interleukin
i.p.	intraperitoneal
i.v.	intravenous

JAK	janus kinase
μg	microgram
μL	microliter
μm	micrometer
mM	milimolar
min	minute
mRAGE/pAb	mouse RAGE polyclonal antibody
ng	nanogram
nm	nanometer
NaN_3	sodium azide
NaCl	sodium chloride
NK cells	natural killer cells
NMD pathway	nonsense-mediated decay pathway
NO	nitric oxide
iNOS	nitric oxide synthase
NF- κB	nuclear factor kappa B
n	number of observation
PfEMP-1	<i>P.falciparum</i> -encoded erythrocyte membrane protein-1
PRBCs	parasite-infected red blood cells
PBS	phosphate buffer saline
pg	pictogram
<i>P.</i>	<i>Plasmodium</i>
KCl	potassium chloride
KH_2PO_4	potassium dihydrogen phosphate anhydrous
PGE_2	prostaglandin E ₂
RAGE ^{-/-} mice	homozygous RAGE deficient mice

RBCs	red blood cells
rmRAGE/Fc Chimera	recombinant mouse RAGE Fc chimera
rpm	revolution per minute
sRAGE	soluble RAGE
STAT	signal transducer and activator of transcription
s.e.m.	standard error of the mean
Th1	T-helper type 1
Th2	T-helper type 2
T _h cells	T helper cells
TLR	toll-like receptor
TGF-β	tumor growth factor-beta
TNF-α	tumor necrosis factor-alpha
T _{regs}	Regulatory T cells
VCAM-1	vascular cell adhesion molecule-1
w/v	weight per volume

CHAPTER 1

INTRODUCTION

1.1 Background

Although being investigated for over hundreds years, malaria still remains a tough challenge to mankind, creating an enormous social, economic, and health burden. According to World Malaria Report 2012, malaria is reported as being endemic in over 104 countries and territories, spanning all continents of the world except Antarctica and Australia, with 99 of these countries had on-going malaria transmission. Despite the extensive efforts in controlling and eradicating malaria since 1955, half of the world population or approximately 3.3 billion people remain at risk of this parasitic infection. In 2010, there were an estimated 219 million cases of malaria, causing 660 000 deaths (WHO, 2012). Every year, malaria imposes huge financial costs on afflicted persons as well as the governments of the endemic countries, putting an immense economic burden on those countries (WHO, 2012; Roll back malaria, 2010).

In Malaysia, the national malaria eradication program has been a success in recent decades, steadily reduces the incidence of malaria from 59208 cases (29.7 per 10,000 populations) in 1995 to 6650 cases (2.4 per 10,000 populations) in 2010 (Lokman, 2011). The reported malaria death cases also remain steady within 20-40 cases annually for the last decade (Western Pacific Region WHO, 2012). The majority of malaria incidences in Malaysia are reported in both Sabah and Sarawak of Malaysian Borneo, accounted for 38% and 33% respectively of all reported cases (Ministry of Health Malaysia, 2011). Noteworthy, most of the malaria cases are confined to rural and semi-rural areas no matter in Peninsular Malaysia or Malaysian Borneo (Rundi, 2011) and largely concentrated among immigrant workers (legal/illegal), workers in land schemes, and hinterland aborigines who are mostly socio-economically disadvantaged (Ministry of Health Malaysia, 2011).

The appearance of first drug resistant case to one of the most common antimalarial drug, chloroquine, along the Thai-Combodian border in late 1950s, has indicated the start of a new chapter in the history of combating malaria. Since then, more and more cases reporting the resistance of the malaria parasites to anti-malarial chemotherapy were detected worldwide. To date, resistance *in vivo* has been observed in almost all currently used antimalarial drugs, including chloroquine, quinine, sulphadoxine-pyrimethamine, and mefloquine (Farooq & Mahajan, 2004). To make the situation worse, not only drug resistance of malaria parasites is widespread, the vector *Anopheles* mosquitoes themselves also have developed resistance to insecticide used for malaria control (WHO, 2012). Since no vaccine is yet fully available and new antimalarial agents will be facing resistance problem eventually, the need to develop a new therapeutic option for malaria therapy by targeting the immune system is great indeed.

Excessive inflammatory response to the parasite invasion is the disastrous endpoint of an overstimulated immune reaction, which in turn lead to malaria susceptibility, severe immunopathological conditions, septic shock and multi organ failure due to end organ damage (Plebanski & Hill, 2000). Although not much is known for the mechanisms in the pathogenesis of severe malaria, considerable evidences have revealed that high levels of pro-inflammatory cytokines such as interferon-gamma (IFN- γ), tumor necrosis factor-alpha (TNF- α), and interleukin-1 (IL-1) are correlated with severity of malaria and hyperinflammation is implicated in the development of severe malaria (Lyke *et al.*, 2004; Artavanis-Tsakonas, Tongren & Riley, 2003). These findings suggest an idea that using immunomodulatory approach to reduce the overproduction pro-inflammatory cytokines and limiting the hyperactivated immune response may be beneficial in reducing morbidity and mortality due to severe malaria. In this case, receptor for advanced glycation end products (RAGE) has potential to be an attractive immunomodulatory target because RAGE signaling and its downstream pathways has been identified to be essential in perpetuation and amplification of inflammatory reactions (Bierhaus *et al.*, 2005) as well as in the production of various pro-inflammatory cytokine, including TNF- α , IFN- γ , and IL-6 (Lotze & Tracey, 2005; Treutiger *et al.*, 2003).

The receptor of advanced glycation endproducts (RAGE) is a newly identified multi-ligand receptors and a member of the immunoglobulin superfamily. It is involved in the signal transduction from pathogen substrates to cell activation during the onset and perpetuation of inflammation. The binding of RAGE ligands, including advanced glycation endproducts (AGEs) and high mobility group box protein 1 (HMGB1), to their receptor has been found to initiate a series of intracellular signal transduction pathways that leads to a sustained inflammatory reaction (Lander *et al.* 1997; Wautier *et al.* 2001; Ishihara *et al.* 2003; Huang *et al.* 2001) as well as amplify the cytokine cascade during systemic inflammation (Andersson *et al.* 2000).

Upregulation of RAGE occurred in the blood vessels, neurons and transformed epithelial during many inflammatory-related pathologic conditions such as septicaemia, rheumatoid arthritis, inflammatory kidney disease, arteriosclerosis, and inflammatory bowel disease (Bierhaus *et al.* 2005). The potential of RAGE as therapeutic target in disease conditions has been demonstrated in several studies. Blocking of RAGE signal transduction pathway for example can increase survival in experimental sepsis (Wang *et al.* 1999; Yang *et al.* 2004), reduce the signs of lung damage in acute inflammation during lung injury (Abraham *et al.* 2000) and increase survival after massive liver resection (Cataldegirmen *et al.* 2005). The most interesting finding was that RAGE knockout mice were protected from lethal septic shock as compared with the wild-type controls (Liliensiek *et al.* 2004).

Most data from the previous studies suggest that RAGE perpetuates and amplifies inflammatory reactions and targeting this receptor might help curbing the hyperinflammatory responses that occur in many inflammation-associated conditions. Since malaria is also considered as an inflammatory disease involving excessive inflammatory response towards parasite invasion and severe systemic inflammation has been closely linked to morbidity and mortality of the disease, it is necessary to investigate whether modulation of RAGE signaling pathway would produce any

beneficial outcomes during malaria infection. If modulation of RAGE signaling pathways can produce impact on the pathological conditions seen during malaria infection then targeting RAGE would be beneficial and it can represent a promising new therapeutic option for malaria therapy. This can at least reduce the morbidity and mortality associated with malaria infection and may be a breakthrough in the effort of treating the disease.

1.2 Hypotheses

In this study, it is hypothesized that RAGE is involved in malaria infection and modulating the RAGE signaling pathway would give a positive impact on the pathophysiology of the disease.

1.3 Objectives

The general objective of this research is to study and determine the possible roles and involvement of RAGE during malaria infection. The specific objectives of this study are listed as follows:

- 1) To investigate the involvement of RAGE during malaria infection by determining its expression at systemic level.
- 2) To modulate the expression of RAGE *in vivo* by means of neutralizing antibody against RAGE and chimera binding protein as an antagonist to RAGE ligands.
- 3) To evaluate the effects of RAGE pathway modulation on the pathological changes seen during malaria infection, whether blocking of RAGE pathway would improve the pathological conditions associated with disease.
- 4) To evaluate the modulatory effects of RAGE on the pattern of major cytokines release during the infection. This includes the pro-inflammatory cytokines IL-2, IL-6, IL-17A, TNF- α and IFN- γ , and the anti-inflammatory cytokines IL-10 and IL-4.

REFERENCES

- Abraham, E., Arcaroli, J., Carmody, A., Wang, H., & Tracey, K. J. (2000). Cutting edge: HMG-1 as a mediator of acute lung inflammation. *The Journal of Immunology*, 165(6), 2950-2954.
- Aitken, E. H., Negri, E. M., Barboza, R., Lima, M. R., Álvarez, J. M., Marinho, C. R., ... & Epiphanio, S. (2014). Ultrastructure of the lung in a murine model of malaria-associated acute lung injury/acute respiratory distress syndrome. *Malaria Journal*, 13(1), 230.
- Akdis, C. A., & Blaser, K. (1999). IL-10-induced anergy in peripheral T cell and reactivation by microenvironmental cytokines: two key steps in specific immunotherapy. *The FASEB Journal*, 13(6), 603-609.
- Akirav, E. M., Preston-Hurlburt, P., Garyu, J., Henegariu, O., Clynes, R., Schmidt, A. M., & Herold, K. C. (2012). RAGE expression in human T cells: a link between environmental factors and adaptive immune responses. *PLoS One*, 7(4), e34698.
- Akirav, E. M., Henegariu, O., Preston-Hurlburt, P., Schmidt, A. M., Clynes, R., & Herold, K. C. (2014). The receptor for advanced glycation end products (RAGE) affects T cell differentiation in OVA induced asthma. *PloS One*, 9(4), e95678.
- Amante, F. H., Haque, A., Stanley, A. C., de Labastida Rivera, F., Randall, L. M., Wilson, Y. A., ... & Engwerda, C. R. (2010). Immune-mediated mechanisms of parasite tissue sequestration during experimental cerebral malaria. *The Journal of Immunology*, 185(6), 3632-3642.
- Amodu, O. K., Adeyemo, A. A., Olumese, P. E., & Gbadegesin, R. A. (1998). Intraleucocytic malaria pigment and clinical severity of malaria in children. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 92(1), 54-56.
- Anand, A. C., & Puri, P. (2005). Jaundice in malaria. *Journal of Gastroenterology and Hepatology*, 20(9), 1322-1332.
- Andersson, U., Wang, H., Palmlad, K., Aveberger, A. C., Bloom, O., Erlandsson-Harris, H., ... & Tracey, K. J. (2000). High mobility group 1 protein (HMG-1) stimulates proinflammatory cytokine synthesis in human monocytes. *The Journal of Experimental Medicine*, 192(4), 565-570.
- Andrade, B. B., Reis-Filho, A., Souza-Neto, S. M., Clarêncio, J., Camargo, L. M., Barral, A., & Barral-Netto, M. (2010). Severe *Plasmodium vivax* malaria exhibits marked inflammatory imbalance. *Malaria Journal*, 9(13), 10-1186.
- Angulo, I., & Fresno, M. (2002). Cytokines in the pathogenesis of and protection against malaria. *Clinical and Diagnostic Laboratory Immunology*, 9(6), 1145-1152.

- Anisman, H., & Merali, Z. (1999). Anhedonic and anxiogenic effects of cytokine exposure. *Advances in Experimental Medicine and Biology*, 461, 199–233.
- Anstey, N. M., Jacups, S. P., Cain, T., Pearson, T., Ziesing, P. J., Fisher, D. A., ... & Maguire, G. P. (2002). Pulmonary manifestations of uncomplicated falciparum and vivax malaria: cough, small airways obstruction, impaired gas transfer, and increased pulmonary phagocytic activity. *Journal of Infectious Diseases*, 185(9), 1326-1334.
- Antinori, S., Galimberti, L., Milazzo, L., & Corbellino, M. (2013). Plasmodium knowlesi: the emerging zoonotic malaria parasite. *Acta Tropica*, 125(2), 191-201.
- Arancio, O., Zhang, H. P., Chen, X., Lin, C., Trinchese, F., Puzzo, D., ... & Du Yan, S. S. (2004). RAGE potentiates A β -induced perturbation of neuronal function in transgenic mice. *The EMBO Journal*, 23(20), 4096-4105.
- Artavanis-Tsakonas, K., & Riley, E. M. (2002). Innate immune response to malaria: Rapid induction of IFN-gamma from human NK cells by live *Plasmodium falciparum* infected erythrocytes. *Journal of Immunology*, 169(6):2956-2963.
- Artavanis-Tsakonas, K., Tongren, J. E., & Riley, E. M. (2003). The war between the malaria parasite and the immune system: immunity, immunoregulation and immunopathology. *Clinical & Experimental Immunology*, 133(2), 145-152.
- Autino, B., Corbett, Y., Castelli, F., & Taramelli, D. (2012). Pathogenesis of malaria in tissues and blood. *Mediterranean journal of Hematology and Infectious Diseases*, 4(1), e2012061.
- Babikir, H. E. H. (2010). Cerebral malaria in children: A review of pathophysiology, clinical manifestations and management. *Journal of Paediatrics and Child Health*, 10, 14-23.
- Baheti, R., Laddha, P., & Gehlot, R. S. (2003). Liver involvement in falciparum malária-A histo-pathological analysis. *Journal Indian Academy of Clinical Medicine*, 4(1), 34-38.
- Bailey, J. W., Williams, J., Bain, B. J., Parker-Williams, J., & Chiodini, P. (2007). General Hematology Task Force. *Guideline for laboratory diagnosis of malaria*. London: British Committee for Standards in Haematology. Retrieved from <http://guideline.gov/content.aspx?id=11997>
- Barber, B. E., William, T., Dhararaj, P., Anderios, F., Grigg, M. J., Yeo, T. W., & Anstey, N. M. (2012). Epidemiology of Plasmodium knowlesi malaria in north-east Sabah, Malaysia: family clusters and wide age distribution. *Malaria Journal*, 11(401), 10-1186.
- Barsoum, R. S. (1998). Malarial nephropathies. *Nephrology Dialysis Transplantation*, 13(6), 1588-1597.

- Barsoum, R. S. (2000). Malarial acute renal failure. *Journal of the American Society of Nephrology*, 11(11), 2147-2154.
- Basa-Dalay, V., Limpaiboon, R., Looareesuwan, S., Kitayaporn, D., Karbwang, J., Tantraporn, W., ... & Charoenlarp, P. (1991). The relationship between splenomegaly and severity of falciparum malaria. *Journal of Infectious Diseases and Antimicrobial Agents*, 8(3), 153-160.
- Basir, R. (1998). *Tumour Necrosis Factor- α and Nitric Oxide in rodent malaria*. (Unpublished doctoral dissertation). University of Manchester, UK.
- Basir, R., Hasballah, K., Jabbarzare, M., Gam, L. H., Abdul Majid, A. M. S., Yam, M. F., ... & Abdullah, W. O. (2012). Modulation of interleukin-18 release produced positive outcomes on parasitaemia development and cytokines production during malaria in mice. *Tropical Biomedicine*, 29(3), 405-421.
- Basir, R., Rahiman, S. F., Hasballah, K., Chong, W. C., Talib, H., Yam, M. F., ... & Ahmad, Z. (2012). *Plasmodium berghei* ANKA infection in ICR mice as a model of cerebral malaria. *Iranian Journal of Parasitology*, 7(4), 62-74.
- Basta, G., Lazzerini, G., Massaro, M., Simoncini, T., Tanganeli, P., Fu, C., ... & De Caterina, R. (2002). Advanced glycation end products activate endothelium through signal-transduction receptor RAGE a mechanism for amplification of inflammatory responses. *Circulation*, 105(7), 816-822.
- Bellone, G., & Trinchieri, G. (1994). Dual stimulatory and inhibitory effect of NK cell stimulatory factor/IL-12 on human hematopoiesis. *The Journal of Immunology*, 153(3), 930-937.
- Berendt, A. R., Simmons, D. L., Tansey, J., Newbold, C. I., & Marsh, K. (1989). Intercellular adhesion molecule-1 is an endothelial cell adhesion receptor for *Plasmodium falciparum*. *Nature*, 341, 57-59.
- Berendt, A. R., Turner, G. D. H., & Newbold, C. I. (1994). Cerebral malaria: the sequestration hypothesis. *Parasitology Today*, 10(10), 412-414.
- Berretta, F., St-Pierre, J., Piccirillo, C. A., & Stevenson, M. M. (2011). IL-2 contributes to maintaining a balance between CD4+ Foxp3+ regulatory T cells and effector CD4+ T cells required for immune control of blood-stage malaria infection. *The Journal of Immunology*, 186(8), 4862-4871.
- Biamonte, M. A., Wanner, J., & Le Roch, K. G. (2013). Recent advances in malaria drug discovery. *Bioorganic & Medicinal Chemistry Letters*, 23(10), 2829-2843.
- Bianchi, M. E., & Manfredi, A. A. (2007). High-mobility group box 1 (HMGB1) protein at the crossroads between innate and adaptive immunity. *Immunological Reviews*, 220(1), 35-46.

- Biemba, G., Gordeuk, V. R., Thuma, P., & Weiss, G. (2000). Markers of inflammation in children with severe malarial anaemia. *Tropical Medicine & International Health*, 5(4), 256-262.
- Bierhaus, A., Schiekofer, S., Schwaninger, M., Andrassy, M., Humpert, P. M., Chen, J., ... Nawroth, P. P. (2001). Diabetes-associated sustained activation of the transcription factor nuclear factor-kappaB. *Diabetes*, 50(12), 2792-2808.
- Bierhaus, A., Haslbeck, K. M., Humpert, P. M., Liliensiek, B., Dehmer, T., Morcos, M., ... & Nawroth, P. P. (2004). Loss of pain perception in diabetes is dependent on a receptor of the immunoglobulin superfamily. *Journal of Clinical Investigation*, 114(12), 1741-1751.
- Bierhaus, A., Humpert, P. M., Morcos, M., Wendt, T., Chavakis, T., Arnold, B., ... & Nawroth, P. P. (2005). Understanding RAGE, the receptor for advanced glycation end products. *Journal of Molecular Medicine*, 83(11), 876-886.
- Bierhaus, A., Stern, D. M., & Nawroth, P. P. (2006). RAGE in inflammation: A new therapeutic target? *Current Opinion In Investigational Drugs*, 7(11), 985-991.
- Biswas, S., Karmarkar, M. G., & Sharma, Y. D. (2001). Antibodies detected against *Plasmodium falciparum* hemozoin with inhibitory properties to cytokine production. *FEMS Microbiology Letters*, 194(2), 175-179.
- Bolad, A., & Berzins, K. (2000). Antigenic diversity of *Plasmodium falciparum* and antibody-mediated parasite neutralization. *Scandinavian Journal of Immunology*, 52(3), 233-239.
- Bombini, G., Canetti, C., Rocha, F. A., & Cunha, F. Q. (2004). Tumour necrosis factor- α mediates neutrophil migration to the knee synovial cavity during immune inflammation. *European Journal of Pharmacology*, 496(1), 197-204.
- Bompart, F., Kiechel, J. R., Sebbag, R., & Pecoul, B. (2011). Innovative public-private partnerships to maximize the delivery of anti-malarial medicines: lessons learned from the ASAQ Winthrop experience. *Malaria Journal*, 10(143), 1475-287.
- Bonaldi, T., Talamo, F., Scaffidi, P., Ferrera, D., Porto, A., Bachi, A., ... Bianchi, M. (2003). Monocytic cells hyperacetylate chromatin protein HMGB1 to redirect it towards secretion. *The EMBO Journal*, 22(20), 5551-5560.
- Bopp, C., Bierhaus, A., Hofer, S., Bouchon, A., Nawroth, P. P., Martin, E., & Weigand, M. A. (2008). Bench-to-bedside review: The inflammation-perpetuating pattern-recognition receptor RAGE as a therapeutic target in sepsis. *Critical Care*, 12(1), 201-208.
- Bouharoun-Tayoun, H., Attanath, P., Sabchareon, A., Chongsuphajaisiddhi, T., & Druilhe, P. (1990). Antibodies that protect humans against *Plasmodium falciparum* blood stages do not on their own inhibit parasite growth and invasion in vitro, but act in cooperation with monocytes. *The Journal of Experimental Medicine*, 172(6), 1633-1641.

- Bouharoun-Tayoun, H., Oeuvray, C., Lunel, F., & Druilhe, P. (1995). Mechanisms underlying the monocyte-mediated antibody-dependent killing of *Plasmodium falciparum* asexual blood stages. *The Journal of Experimental Medicine*, 182(2), 409-418.
- Brett, J., Schmidt, A. M., Yan, S. D., Zou, Y. S., Weidman, E., Pinsky, D., ... Stern, D. (1993). Survey of the distribution of a newly characterized receptor for advanced glycation end-products in tissues. *American Journal of Pathology*, 143(6), 1699-1712.
- Brian de Souza, J., Hafalla, J. C., Riley, E. M., & Couper, K. N. (2010). Cerebral malaria: why experimental murine models are required to understand the pathogenesis of disease. *Parasitology*, 137(05), 755-772.
- Bruneel, F., Gachot, B., Wolff, M., Regnier, B., Danis, M., & Vachon, F. (2001). Resurgence of blackwater fever in long-term European expatriates in Africa: report of 21 cases and review. *Clinical Infectious Diseases*, 32(8), 1113-1140.
- Bucciarelli, L. G., Wendt, T., Qu, W., Lu, Y., Lalla, E., Rong, L. L., ... & Schmidt, A. M. (2002). RAGE blockade stabilizes established atherosclerosis in diabetic apolipoprotein E-null mice. *Circulation*, 106(22), 2827-2835.
- Bueno, L. L., Morais, C. G., Lacerda, M. V., Fujiwara, R. T., & Braga, É. M. (2012). Interleukin-17 producing T helper cells are increased during natural *Plasmodium vivax* infection. *Acta Tropica*, 123(1), 53-57.
- Buffet, P. A., Safeukui, I., Milon, G., Mercereau-Puijalon, O., & David, P. H. (2009). Retention of erythrocytes in the spleen: a double-edged process in human malaria. *Current Opinion in Hematology*, 16(3), 157-164.
- Cabantous, S., Pouliougou, B., Oumar, A. A., Traore, A., Barry, A., Vitte, J., ... & Dessein, A. J. (2009). Genetic evidence for the aggravation of *Plasmodium falciparum* malaria by interleukin 4. *Journal of Infectious Diseases*, 200(10), 1530-1539.
- Carter, R., & Walliker, D. (1975). New observations on the malaria parasites of rodents of the Central African Republic-*Plasmodium vinckeii* petteri subsp. nov. and *Plasmodium chabaudi* Landau, 1965. *Annals of Tropical Medicine and Parasitology*, 69(2), 187-196.
- Carter, R., & Graves, P. M. (1988). Gametocytes. In W. H. Wernsdorfer and I. McGregor (Eds.), *Malaria: principles and practice of malariology*. (Vol. 1, pp. 233-305). Edinburgh: Churchill Livingstone.
- Carvalho, L. H., Sano, G. I., Hafalla, J. C., Morrot, A., de Lafaille, M. A. C., & Zavala, F. (2002). IL-4-secreting CD4+ T cells are crucial to the development of CD8+ T-cell responses against malaria liver stages. *Nature Medicine*, 8(2), 166-170.
- Carvalho, L. J., Ferreira-da-Cruz, M. F., Daniel-Ribeiro, C. T., Pelajo-Machado, M., & Lenzi, H. L. (2007). Germinal center architecture disturbance during

- Plasmodium berghei* ANKA infection in CBA mice. *Malaria Journal*, 6(1), 59.
- Cataldegirmen, G., Zeng, S., Feirt, N., Ippagunta, N., Dun, H., Qu, W., ... & Emond, J. C. (2005). Rage limits regeneration after massive liver injury by coordinated suppression of TNF- α and NF- κ B. *The Journal of Experimental Medicine*, 201(3), 473-484.
- Chaiyaroj, S. C., Rutta, A. S., Muenthaisong, K., Watkins, P., Ubol, M. N., & Looareesuwan, S. (2004). Reduced levels of transforming growth factor- β 1, interleukin-12 and increased migration inhibitory factor are associated with severe malaria. *Acta Tropica*, 89(3), 319-327.
- Chakravorty, S. J., & Craig, A. (2005). The role of ICAM-1 in *Plasmodium falciparum* cytoadherence. *European Journal of Cell Biology*, 84(1), 15-27.
- Chang, Y. J., Holtzman, M. J., & Chen, C. C. (2002). Interferon- γ -induced epithelial ICAM-1 expression and monocyte adhesion involvement of protein kinase C-dependent C-Src tyrosine kinase activation pathway. *Journal of Biological Chemistry*, 277(9), 7118-7126.
- Charoenpan, P., Indraprasit, S., Kiatboonsri, S., Suvachittanont, O., & Tanomsup, S. (1990). Pulmonary edema in severe falciparum malaria. Hemodynamic study and clinicophysiological correlation. *Chest Journal*, 97(5), 1190-1197.
- Chavakis, T., Bierhaus, A., Al-Fakhri, N., Schneider, D., Witte, S., Linn, T., ... Nawroth, P. P. (2003). The pattern recognition receptor (RAGE) is a counterreceptor for leukocyte integrins: A novel pathway for inflammatory cell recruitment. *Journal of Experimental Medicine*, 198(10), 1507-1515.
- Chen, Q., Schlichtherle, M., & Wahlgren, M. (2000). Molecular aspects of severe malaria. *Clinical Microbiology Reviews*, 13(3), 439-450.
- Chen, Y., Yan, S. S., Colgan, J., Zhang, H. P., Luban, J., Schmidt, A. M., ... & Herold, K. C. (2004). Blockade of late stages of autoimmune diabetes by inhibition of the receptor for advanced glycation end products. *The Journal of Immunology*, 173(2), 1399-1405.
- Chen, Y., Akirav, E. M., Chen, W., Henegariu, O., Moser, B., Desai, D., ... Herold, K. C. (2008). RAGE ligation affects T cell activation and controls T cell differentiation. *The Journal of Immunology*, 181(6), 4272-4278.
- Cheng, C., Tsuneyama, K., Kominami, R., Shinohara, H., Sakurai, S., Yonekura, H., ... Yamamoto, Y. (2005). Expression profiling of endogenous secretory receptor for advanced glycation end products in human organs. *Modern Pathology*, 18(10), 1385-1396.
- Chotivanich, K., Udomsangpetch, R., McGready, R., Proux, S., Newton, P., Pukrittayakamee, S., ... & White, N. J. (2002). Central role of the spleen in malaria parasite clearance. *Journal of Infectious Diseases*, 185(10), 1538-1541.

- Clark, I. A., Cowden, W. B., Butcher, G. A., & Hunt, N. H. (1987). Possible roles of tumor necrosis factor in the pathology of malaria. *The American Journal of Pathology*, 129(1), 192-199.
- Clark, I. A., Rockett, K. A., & Cowden, W. B. (1992). Possible central role of nitric oxide in conditions clinically similar to cerebral malaria. *The Lancet*, 340(8824), 894-896.
- Clark, I. A., & Rockett, K. A. (1994). The cytokine theory of human cerebral malaria. *Parasitology Today*, 10(10), 410-412.
- Clark, I. A., & Cowden, W. B. (2003). The pathophysiology of falciparum malaria. *Pharmacology & Therapeutics*, 99(2), 221-260.
- Clark, I. A., Budd, A. C., Alleva, L. M., & Cowden, W. B. (2006). Human malarial disease: a consequence of inflammatory cytokine release. *Malaria Journal*, 5(1), 85.
- Clark, I. A., Alleva, L. M., Budd, A. C., & Cowden, W. B. (2008). Understanding the role of inflammatory cytokines in malaria and related diseases. *Travel Medicine and Infectious Disease*, 6(1), 67-81.
- Clark, I. A., & Alleva, L. M. (2009). Is human malarial coma caused, or merely deepened, by sequestration?. *Trends in Parasitology*, 25(7), 314-318.
- Claser, C., Malleret, B., Gun, S. Y., Wong, A. Y. W., Chang, Z. W., Teo, P., ... & Rénia, L. (2011). CD8+ T cells and IFN- γ mediate the time-dependent accumulation of infected red blood cells in deep organs during experimental cerebral malaria. *PLoS One*, 6(4), e18720.
- Clynes, R., Moser, B., Yan, S. F., Ramasamy, R., Herold, K., & Schmidt, A. M. (2007). Receptor for AGE (RAGE): Weaving tangled webs within the inflammatory response. *Current Molecular Medicine*, 7(8), 743-751.
- Coban, C., Ishii, K. J., Kawai, T., Hemmi, H., Sato, S., Uematsu, S., ... & Akira, S. (2005). Toll-like receptor 9 mediates innate immune activation by the malaria pigment hemozoin. *The Journal of Experimental Medicine*, 201(1), 19-25.
- Collins, W. E., & Jeffery, G. M. (2005). Plasmodium ovale: parasite and disease. *Clinical Microbiology Reviews*, 18(3), 570-581.
- Collins, W. E., & Jeffery, G. M. (2007). Plasmodium malariae: parasite and disease. *Clinical Microbiology Reviews*, 20(4), 579-592.
- Collison, K. S., Parhar, R. S., Saleh, S. S., Meyer, B. F., Kwaasi, A. A., Hammami, M. M., ... Al-Mohanna, F. A. (2002). RAGE-mediated neutrophil dysfunction is evoked by advanced glycation end products (AGEs). *Journal of Leukocyte Biology*, 71(3), 433-444.

- Cooper, A. L., Dascombe, M. J., Rothwell, N. J., & Vale, M. J. (1989). Effects of malaria on O₂ consumption and brown adipose tissue activity in mice. *Journal of Applied Physiology*, 67(3), 1020-1023.
- Corbett, C. E., Duarte, M. I., Lancellotti, C. L., Silva, M. A., & Andrade Junior, H. F. (1989). Cytoadherence in human *falciparum* malaria as a cause of respiratory distress. *The Journal of Tropical Medicine and Hygiene*, 92(2), 112-120.
- Cordeiro, R. S., Cunha, F. Q., Filho, J. A., Flores, C. A., Vasconcelos, H. N., & Martins, M. A. (1983). *Plasmodium berghei*: physiopathological changes during infections in mice. *Annals of Tropical Medicine and Parasitology*, 77(5), 455-465.
- Cordoliani, Y. S., Sarrazin, J. L., Felten, D., Caumes, E., Lévéque, C., & Fisch, A. (1998). MR of cerebral malaria. *American Journal of Neuroradiology*, 19(5), 871-874.
- Couper, K. N., Blount, D. G., Hafalla, J. C., van Rooijen, N., de Souza, J. B., & Riley, E. M. (2007). Macrophage-mediated but gamma interferon-independent innate immune responses control the primary wave of *Plasmodium yoelii* parasitemia. *Infection and Immunity*, 75(12), 5806-5818.
- Couper, K. N., Blount, D. G., & Riley, E. M. (2008). IL-10: the master regulator of immunity to infection. *The Journal of Immunology*, 180(9), 5771-5777.
- Couper, K. N., Blount, D. G., Wilson, M. S., Hafalla, J. C., Belkaid, Y., Kamanaka, M., ... & Riley, E. M. (2008). IL-10 from CD4+ CD25- Foxp3- CD127- adaptive regulatory T cells modulates parasite clearance and pathology during malaria infection. *PLoS Pathogens*, 4(2), e1000004.
- Cox, F. E. G. (1988). Major Animal Models in Malaria Research: Rodent. In W. H. Wernsdorfer and I. McGregor (Eds.), *Malaria Principles and Practice of Malaria* (Vol. 2, pp. 1503-1543). London: Churchill Livingstone.
- Cox, F. E. (2010). History of the discovery of the malaria parasites and their vectors. *Parasites & Vectors*, 3(1), 5.
- Cox-Singh, J., Davis, T. M., Lee, K. S., Shamsul, S. S., Matusop, A., Ratnam, S., ... & Singh, B. (2008). Plasmodium knowlesi malaria in humans is widely distributed and potentially life threatening. *Clinical Infectious Diseases*, 46(2), 165-171.
- Crutcher, J. M., Stevenson, M. M., Sedegah, M., & Hoffman, S. L. (1995). Interleukin-12 and malaria. *Research in Immunology*, 146(7), 552-559.
- Curfs, J. H., Van der Meer, J. W., Sauerwein, R. W., & Eling, W. M. (1990). Low dosages of interleukin 1 protect mice against lethal cerebral malaria. *The Journal of Experimental Medicine*, 172(5), 1287-1291.
- Curfs J. H., van der Meide, P. H., Billiau, A., Meuwissen, J. H., & Eling, W. M. (1993). *Plasmodium berghei*: recombinant interferon- γ and the development of

- parasitemia and cerebral lesions in malaria-infected mice. *Experimental Parasitology*, 77(2), 212-223.
- Daneshvar, C., Davis, T. M., Cox-Singh, J., Rafa'ee, M. Z., Zakaria, S. K., Divis, P. C., & Singh, B. (2009). Clinical and laboratory features of human Plasmodium knowlesi infection. *Clinical infectious diseases*, 49(6), 852-860.
- Das, B. S. (2008). Renal failure in malaria. *Journal of Vector Borne Diseases*, 45(2), 83.
- Dascombe, M. J., Huynh, T. T. T., & Owen, R. H. (2000). Behavioural thermoregulatory responses to malaria infection and interleukin-1 β in rats. *Journal of Thermal Biology*, 25(1), 11-15.
- Dattilo, B. M., Fritz, G., Leclerc, E., Kooi, C. W., Heizmann, C. W., & Chazin, W. J. (2007). The extracellular region of the receptor for advanced glycation end products is composed of two independent structural units. *Biochemistry*, 46(23), 6957-6970.
- Day, N. P., Hien, T. T., Schollaardt, T., Loc, P. P., Chuong, L. V., Chau, T. T., ... & Ho, M. (1999). The prognostic and pathophysiologic role of pro- and anti-inflammatory cytokines in severe malaria. *Journal of Infectious Diseases*, 180, 1288-1297.
- Day, K. P., & Fowkes, F. J. (2011). Quantifying malaria dynamics within the host. *Science*, 333(6045), 984-988.
- De Kossodo, S., & Grau, G. E. (1993). Profiles of cytokine production in relation with susceptibility to cerebral malaria. *The Journal of Immunology*, 151(9), 4811-4820.
- Del Portillo, H. A., Ferrer, M., Brugat, T., Martin-Jaular, L., Langhorne, J., & Lacerda, M. V. (2012). The role of the spleen in malaria. *Cellular microbiology*, 14(3), 343-355.
- Del Prete, G., Maggi, E., Parronchi, P., Chretien, I., Tiri, A., Macchia, D., ... & Romagnani, S. (1988). IL-4 is an essential factor for the IgE synthesis induced in vitro by human T cell clones and their supernatants. *The Journal of Immunology*, 140(12), 4193-4198.
- Delacollette, C., Taelman, H., & Wery, M. (1995). An etiologic study of hemoglobinuria and blackwater fever in the Kivu mountains, Zaire. *Annales-Societe Belge De Medecine Tropicale*, 75, 51-63.
- Dent, A. E., Bergmann-Leitner, E. S., Wilson, D. W., Tisch, D. J., Kimmel, R., Vulule, J., ... & Kazura, J. W. (2008). Antibody-mediated growth inhibition of *Plasmodium falciparum*: relationship to age and protection from parasitemia in Kenyan children and adults. *PLoS One*, 3(10), e3557.
- Depinay, N., Franetich, J. F., Grüner, A. C., Mauduit, M., Chavatte, J. M., Luty, A. J., ... & Réna, L. (2011). Inhibitory effect of TNF- α on malaria pre-

- erythrocytic stage development: influence of host hepatocyte/parasite combinations. *PLoS One*, 6(3), e17464.
- Deroost, K., Tyberghein, A., Lays, N., Noppen, S., Schwarzer, E., Vanstreels, E., ... & Van den Steen, P. E. (2013). Hemozoin induces lung inflammation and correlates with malaria-associated acute respiratory distress syndrome. *American Journal of Respiratory Cell and Molecular Biology*, 48(5), 589-600.
- de Waal Malefyt, R., Abrams, J., Bennett, B., Figgdr, C. G., & De Vries, J. E. (1991). Interleukin 10 (IL-10) inhibits cytokine synthesis by human monocytes: an autoregulatory role of IL-10 produced by monocytes. *The Journal of Experimental Medicine*, 174(5), 1209-1220.
- de Waal Malefyt, R., Haanen, J., Spits, H., Roncarolo, M. G., Te Velde, A., Figgdr, C., ... & De Vries, J. E. (1991). Interleukin 10 (IL-10) and viral IL-10 strongly reduce antigen-specific human T cell proliferation by diminishing the antigen-presenting capacity of monocytes via downregulation of class II major histocompatibility complex expression. *The Journal of Experimental Medicine*, 174(4), 915-924.
- Dey, S., Bindu, S., Goyal, M., Pal, C., Alam, A., Iqbal, M. S., ... & Bandyopadhyay, U. (2012). Impact of intravascular hemolysis in malaria on liver dysfunction: Involvement of hepatic free heme overload, NF- κ B activation, and neutrophil infiltration. *Journal of Biological Chemistry*, 287(32), 26630-26646.
- Dietrich, J. B. (2002). The adhesion molecule ICAM-1 and its regulation in relation with the blood-brain barrier. *Journal of Neuroimmunology*, 128(1), 58-68.
- Dinarello, C. A., & Bernheim, H. A. (1981). Ability of human leukocytic pyrogen to stimulate brain prostaglandin synthesis in vitro. *Journal of Neurochemistry*, 37(3), 702-708.
- Dinarello, C. A. (1999). Cytokines as endogenous pyrogens. *Journal of Infectious Diseases*, 179(Supplement 2), S294-S304.
- Dinarello, C. A. (2004). Review: Infection, fever, and exogenous and endogenous pyrogens: some concepts have changed. *Journal of Endotoxin Research*, 10(4), 201-222.
- Dodoo, D., Omer, F. M., Todd, J., Akanmori, B. D., Koram, K. A., & Riley, E. M. (2002). Absolute levels and ratios of proinflammatory and anti-inflammatory cytokine production in vitro predict clinical immunity to *Plasmodium falciparum* malaria. *Journal of Infectious Diseases*, 185(7), 971-979.
- D'Ombrain, M. C., Hansen, D. S., Simpson, K. M., & Schofield, L. (2007). $\gamma\delta$ -T cells expressing NK receptors predominate over NK cells and conventional T cells in the innate IFN- γ response to *Plasmodium falciparum* malaria. *European Journal of Immunology*, 37(7), 1864-1873.

- Dondorp, A. M. (2012). Single-Dose Primaquine as Gametocytocidal Treatment in Patients With Uncomplicated *Falciparum* Malaria. *Clinical Infectious Diseases*, cis962.
- Dondorp, A. M., Kager, P. A., Vreeken, J., & White, N. J. (2000). Abnormal blood flow and red blood cell deformability in severe malaria. *Parasitology Today*, 16(6), 228-232.
- Dondorp, A. M., Nosten, F., Yi, P., Das, D., Phyo, A. P., Tarning, J., ... & White, N. J. (2009). Artemisinin resistance in *Plasmodium falciparum* malaria. *New England Journal of Medicine*, 361(5), 455-467.
- Dondorp, A. M., Pongponratn, E., & White, N. J. (2004). Reduced microcirculatory flow in severe falciparum malaria: pathophysiology and electron-microscopic pathology. *Acta Tropica*, 89(3), 309-317.
- Dondorp, A. M., Yeung, S., White, L., Nguon, C., Day, N. P., Socheat, D., & von Seidlein, L. (2010). Artemisinin resistance: current status and scenarios for containment. *Nature Reviews Microbiology*, 8(4), 272-280.
- Doumbo, O. K., Thera, M. A., Koné, A. K., Raza, A., Tempest, L. J., Lyke, K. E., ... & Rowe, J. A. (2009). High levels of *Plasmodium falciparum* rosetting in all clinical forms of severe malaria in African children. *The American Journal of Tropical Medicine and Hygiene*, 81(6), 987-993.
- Duarte, M. I., Corbett, C. E., Boulos, M., & Amato Neto, V. (1985). Ultrastructure of the lung in *falciparum* malaria. *The American Journal of Tropical Medicine and Hygiene*, 34(1), 31-35.
- Dufour, C., Corcione, A., Svahn, J., Haupt, R., Poggi, V., Béka'ssy, A. N., ... & Pistoia, V. (2003). TNF- α and IFN- γ are overexpressed in the bone marrow of Fanconi anemia patients and TNF- α suppresses erythropoiesis in vitro. *Blood*, 102(6), 2053-2059.
- Dukic-Stefanovic, S., Gasic-Milenkovic, J., Deuther-Conrad, W., & Müncz, G. (2003). Signal transduction pathways in mouse microglia N-11 cells activated by advanced glycation endproducts (AGEs). *Journal of Neurochemistry*, 87(1), 44-55.
- Dybedal, I., Larsen, S., & Jacobsen, S. E. (1995). IL-12 directly enhances in vitro murine erythropoiesis in combination with IL-4 and stem cell factor. *The Journal of Immunology*, 154(10), 4950-4955.
- Ebaid, H., Dkhil, M., Danfour, M., Tohamy, A., & Gabry, M. (2007). Piroxicam-induced hepatic and renal histopathological changes in mice. *The Libyan Journal of Medicine*, 2(2), 82-89.
- Ehrlich, J. H., & Eke, F. U. (2007). Malaria-induced renal damage: facts and myths. *Pediatric Nephrology*, 22(5), 626-637.

- Elghazali, G., Perlmann, H., Rutta, A. S. M., Perlmann, P., & Troye-Blomberg, M. (1997). Elevated plasma levels of IgE in *Plasmodium falciparum*-primed individuals reflect an increased ratio of IL-4 to interferon-gamma (IFN- γ)-producing cells. *Clinical & Experimental Immunology*, 109(1), 84-89.
- Elsheikha, H. M., & Sheashaa, H. A. (2007). Epidemiology, pathophysiology, management and outcome of renal dysfunction associated with plasmodia infection. *Parasitology Research*, 101(5), 1183-1190.
- Engwerda, C. R., Beattie, L., & Amante, F. H. (2005). The importance of the spleen in malaria. *Trends in Parasitology*, 21(2), 75-80.
- Epiphanio, S., Campos, M. G., Pamplona, A., Carapau, D., Pena, A. C., Ata ée, R., ... & Mota, M. M. (2010). VEGF promotes malaria-associated acute lung injury in mice. *PLoS Pathogens*, 6(5), e1000916.
- Farooq, U., & Mahajan, R. C. (2004). Drug resistance in malaria. *Journal of Vector Borne Diseases*, 41(3/4), 45-53.
- Feghali, C. A., & Wright, T. M. (1997). Cytokines in acute and chronic inflammation. *Frontiers in Bioscience*, 2(1), d12-d26.
- Fehrenbach, H., Kasper, M., Tschnig, T., Shearman, M. S., Schuh, D., & Muller, M. (1998). Receptor for advanced glycation endproducts (RAGE) exhibits highly differential cellular and subcellular localisation in rat and human lung. *Cellular and molecular biology (Noisy-le-Grand)*, 44(7), 1147-1157.
- Fell, A. H., & Smith, N. C. (1998). Immunity to asexual blood stages of Plasmodium: is resistance to acute malaria adaptive or innate?. *Parasitology Today*, 14(9), 364-369.
- Felli, N., Pedini, F., Zeuner, A., Petrucci, E., Testa, U., Conticello, C., ... & De Maria, R. (2005). Multiple members of the TNF superfamily contribute to IFN- γ -mediated inhibition of erythropoiesis. *The Journal of Immunology*, 175(3), 1464-1472.
- Fernando, D., Rodrigo, C., & Rajapakse, S. (2011). Primaquine in vivax malaria: an update and review on management issues. *Malaria Journal*, 10(351), 10-1186.
- Fiorentino, D. F., Zlotnik, A., Mosmann, T. R., Howard, M., & O'garra, A. (1991). IL-10 inhibits cytokine production by activated macrophages. *The Journal of Immunology*, 147(11), 3815-3822.
- Fiorentino, D. F., Zlotnik, A., Vieira, P., Mosmann, T. R., Howard, M., Moore, K. W., & O'Garra, A. (1991). IL-10 acts on the antigen-presenting cell to inhibit cytokine production by Th1 cells. *The Journal of Immunology*, 146(10), 3444-3451.

- Fluza, C., Bustin, M., Talwar, S., Tropea, M., Gerstenberger, E., Shelhamer, J. H., & Suffredini, A. F. (2003). Inflammation-promoting activity of HMGB1 on human microvascular endothelial cells. *Blood*, 101(7), 2652-2660.
- Foell, D., Wittkowski, H., Vogl, T., & Roth, J. (2006). S100 proteins expressed in phagocytes: A novel group of damage-associated molecular pattern molecules. *Journal of Leukocyte Biology*, 81(1), 28-37.
- Frevert, U., & Nacer, A. (2013). Immunobiology of Plasmodium in liver and brain. *Parasite Immunology*, 35(9-10), 267-282.
- Galamo, C. D., Jafarshad, A., Blanc, C., & Druilhe, P. (2009). Anti-MSP1 block 2 antibodies are effective at parasite killing in an allele-specific manner by monocyte-mediated antibody-dependent cellular inhibition. *Journal of Infectious Diseases*, 199(8), 1151-1154.
- Gao, X., Zhang, H., Schmidt, A. M., & Zhang, C. (2008). AGE/RAGE produces endothelial dysfunction in coronary arterioles in type 2 diabetic mice. *American Journal of Physiology-Heart and Circulatory Physiology*, 295(2), H491-H498.
- Garnham, P. C. C. (1965). The structure of the early sporogonic stages of *Plasmodium berghei*. *Ann Soc Belges Med Trop Parasitol Mycol*, 45, 259-264.
- Gately, M. K., Renzetti, L. M., Magram, J., Stern, A. S., Adorini, L., Gubler, U., & Presky, D. H. (1998). The interleukin-12/interleukin-12-receptor system: role in normal and pathologic immune responses. *Annual Review of Immunology*, 16(1), 495-521.
- Gazzinelli, R. T., Wysocka, M., Hieny, S., Scharton-Kersten, T., Cheever, A., Kühn, R., ... & Sher, A. (1996). In the absence of endogenous IL-10, mice acutely infected with *Toxoplasma gondii* succumb to a lethal immune response dependent on CD4+ T cells and accompanied by overproduction of IL-12, IFN-gamma and TNF-alpha. *The Journal of Immunology*, 157(2), 798-805.
- Gebhardt, C., Németh, J., Angel, P., & Hess, J. (2006). S100A8 and S100A9 in inflammation and cancer. *Biochemical Pharmacology*, 72(11), 1622-1631.
- Gebhardt, C., Riehl, A., Durchdewald, M., Nemeth, J., Furstenberger, G., Muller-Decker, K., ... Angel, P. (2008). RAGE signaling sustains inflammation and promotes tumor development. *Journal of Experimental Medicine*, 205(2), 275-285.
- Geiger, T., Andus, T., Klaproth, J., Hirano, T., Kishimoto, T., & Heinrich, P. C. (1988). Induction of rat acute-phase proteins by interleukin 6 in vivo. *European Journal of Immunology*, 18(5), 717-721.
- Ghosh, K., & Ghosh, K. (2007). Pathogenesis of anemia in malaria: a concise review. *Parasitology Research*, 101(6), 1463-1469.

- Gimenez, F., de Lagerie, S. B., Fernandez, C., Pino, P., & Mazier, D. (2003). Tumor necrosis factor α in the pathogenesis of cerebral malaria. *Cellular and Molecular Life Sciences CMSL*, 60(8), 1623-1635.
- Gogos, C. A., Drosou, E., Bassaris, H. P., & Skoutelis, A. (2000). Pro-versus anti-inflammatory cytokine profile in patients with severe sepsis: a marker for prognosis and future therapeutic options. *Journal of Infectious Diseases*, 181(1), 176-180.
- Goldin, A., Beckman, J. A., Schmidt, A. M., & Creager, M. A. (2006). Advanced glycation end products sparking the development of diabetic vascular injury. *Circulation*, 114(6), 597-605.
- Gowda, D. C. (2007). TLR-mediated cell signaling by malaria GPIs. *Trends in Parasitology*, 23(12), 596-604.
- Grau, G. E., Piguet, P. F., Vassalli, P., & Lambert, P. H. (1989). Tumor-necrosis factor and other cytokines in cerebral malaria: experimental and clinical data. *Immunological Reviews*, 112(1), 49-70.
- Grau, G. E., Taylor, T. E., Molyneux, M. E., Wirima, J. J., Vassalli, P., Hommel, M., & Lambert, P. H. (1989). Tumor necrosis factor and disease severity in children with falciparum malaria. *New England Journal of Medicine*, 320(24), 1586-1591.
- Grau, G. E., Mackenzie, C. D., Carr, R. A., Redard, M., Pizzolato, G., Allasia, C., ... & Molyneux, M. E. (2003). Platelet accumulation in brain microvessels in fatal pediatric cerebral malaria. *Journal of Infectious Diseases*, 187(3), 461-466.
- Grobusch, M. P., & Kremsner, P. G. (2005). Uncomplicated malaria. In *Malaria: Drugs, Disease and Post-genomic Biology* (pp. 81-104). Springer Berlin Heidelberg.
- Groux, H., Perraut, R., Garraud, O., Poingt, J. P., & Gysin, J. (1990). Functional characterization of the antibody-mediated protection against blood stages of *Plasmodium falciparum* in the monkey *Saimiri sciureus*. *European Journal of Immunology*, 20(10), 2317-2323.
- Guerra, C. A., Gikandi, P. W., Tatem, A. J., Noor, A. M., Smith, D. L., Hay, S. I., & Snow, R. W. (2008). The limits and intensity of Plasmodium falciparum transmission: implications for malaria control and elimination worldwide. *PLoS Medicine*, 5(2), e38.
- Guo, L., Hu-Li, J., Zhu, J., Watson, C. J., Difilippantonio, M. J., Pannetier, C., & Paul, W. E. (2002). In TH2 cells the IL4 gene has a series of accessibility states associated with distinctive probabilities of IL-4 production. *Proceedings of the National Academy of Sciences*, 99(16), 10623-10628.
- Gysin, J., Gavoille, S., Mattei, D., Scherf, A., Bonnefoy, S., Mercereau-Puijalon, O., ... & da Silva, L. P. (1993). In vitro phagocytosis inhibition assay for the

- screening of potential candidate antigens for sub-unit vaccines against the asexual blood stage of *Plasmodium falciparum*. *Journal of Immunological Methods*, 159(1), 209-219.
- Haldar, K., & Mohandas, N. (2009). Malaria, erythrocytic infection, and anemia. *ASH Education Program Book*, 2009(1), 87-93.
- Hanford, L. E., Enghild, J. J., Valnickova, Z., Petersen, S. V., Schaefer, L. M., Schaefer, T. M., ... Oury, T. D. (2004). Purification and characterization of mouse soluble receptor for advanced glycation end products (sRAGE). *Journal of Biological Chemistry*, 279(48), 50019-50024.
- Hansen, B. D., & Pappas, P. W. (1977). Acute malaria: effects of *Plasmodium berghei* on the metabolic rate of mice. *The Ohio Journal of Science*, 77(4), 189-191.
- Haque, A., Best, S. E., Amante, F. H., Ammerdorffer, A., de Labastida, F., Pereira, T., Ramm, G. A., & Engwerda, C. R. (2011). High parasite burdens cause liver damage in mice following *Plasmodium berghei* ANKA infection independently of CD8+ T cell-mediated immune pathology. *Infection and Immunity*, 79(5), 1882-1888.
- Harpaz, R., Edelman, R., Wasserman, S. S., Levine, M. M., Davis, J. R., & Sztein, M. B. (1992). Serum cytokine profiles in experimental human malaria. Relationship to protection and disease course after challenge. *Journal of Clinical Investigation*, 90(2), 515.
- Hawkes, M., Li, X., Crockett, M., Diassiti, A., Liles, W. C., Liu, J., & Kain, K. C. (2010). Malaria exacerbates experimental mycobacterial infection in vitro and in vivo. *Microbes and Infection*, 12(11), 864-874.
- Hearn, J., Rayment, N., Landon, D. N., Katz, D. R., & de Souza, J. B. (2000). Immunopathology of cerebral malaria: morphological evidence of parasite sequestration in murine brain microvasculature. *Infection and Immunity*, 68(9), 5364-5376.
- Hee, L., Dinudom, A., Mitchell, A. J., Grau, G. E., Cook, D. I., Hunt, N. H., & Ball, H. J. (2011). Reduced activity of the epithelial sodium channel in malaria-induced pulmonary oedema in mice. *International Journal for Parasitology*, 41(1), 81-88.
- Hernandez-Valladares, M., Naessens, J., Musoke, A. J., Sekikawa, K., Rihet, P., Busher, P., & Iraqi, F. A. (2006). Pathology of TNF-deficient mice infected with *Plasmodium chabaudi adami* 408XZ. *Experimental Parasitology*, 114(4), 271-278.
- Herold, K., Moser, B., Chen, Y., Zeng, S., Yan, S. F., Ramasamy, R., ... Schmidt, A. M. (2007). Receptor for advanced glycation end products (RAGE) in a dash to the rescue: Inflammatory signals gone awry in the primal response to stress. *Journal of Leukocyte Biology*, 82(2), 204-212.

- Hirunpetcharat, C., Finkelman, F., Clark, I. A., & Good, M. F. (1999). Malaria parasite-specific Th1-like T cells simultaneously reduce parasitemia and promote disease. *Parasite Immunology*, 21(6), 319-329.
- Hisaeda, H., Maekawa, Y., Iwakawa, D., Okada, H., Himeno, K., Kishihara, K., ... & Yasutomo, K. (2004). Escape of malaria parasites from host immunity requires CD4+ CD25+ regulatory T cells. *Nature Medicine*, 10(1), 29-30.
- Hisaeda, H., Yasutomo, K., & Himeno, K. (2005). Malaria: immune evasion by parasites. *The International Journal of Biochemistry & Cell Biology*, 37(4), 700-706.
- Ho, M., Sexton, M. M., Tongtawe, P., Looareesuwan, S., Suntharasamai, P., & Webster, H. K. (1995). Interleukin-10 inhibits tumor necrosis factor production but not antigen-specific lymphoproliferation in acute *Plasmodium falciparum* malaria. *Journal of Infectious Diseases*, 172(3), 838-844.
- Hoffmann, K. F., Cheever, A. W., & Wynn, T. A. (2000). IL-10 and the dangers of immune polarization: excessive type 1 and type 2 cytokine responses induce distinct forms of lethal immunopathology in murine schistosomiasis. *The Journal of Immunology*, 164(12), 6406-6416.
- Hofmann, M. A., Drury, S., Fu, C., Qu, W., Taguchi, A., Lu, Y., ... Schmidt, A. M. (1999). RAGE mediates a novel proinflammatory axis: A central cell surface receptor for S100/calgranulin polypeptides. *Cell*, 97, 889-901.
- Hofmann, M. A., Drury, S., Hudson, B. I., Gleason, M. R., Qu, W., Lu, Y., ... & Schmidt, A. M. (2002). RAGE and arthritis: the G82S polymorphism amplifies the inflammatory response. *Genes and Immunity*, 3(3), 123-135.
- Hommel, M., & Gilles, H. M. (2005). Malaria. In F. E. G. Cox, D., Wakelin, S. H. Gillespie, and D. D. Despommier (Eds.), *Topley and Wilson's Microbiology and Microbial Infections, Tenth Edition, Volume 5, Parasitology* (pp. 464-518). Washington: ASM Press.
- Hopwood, D. (1996). Fixation and fixatives. *Theory and Practice of Histological Techniques*, 3, 21-142.
- Hori, O., Brett, J., Slattery, T., Cao, R., Zhang, J., Chen, J. X., ... Schmidt, A. M. (1995). The receptor for advanced glycation end products (RAGE) is a cellular binding site for amphotericin: Mediation of neurite outgrowth and co-expression of RAGE and amphotericin in the developing nervous system. *Journal of Biological Chemistry*, 270, 25752-25761.
- Houba, V., Wernsdorfer, W. H., & McGregor, I. (1988). Specific immunity: immunopathology and immunosuppression. *Malaria: principles and practice of malariology*. 1, 621-637.
- Huang, J. S., Guh, J. Y., Chen, H. C., Hung, W. C., Lai, Y. H., & Chuang, L. Y. (2001). Role of receptor for advanced glycation end-product (RAGE) and the

- JAK/STAT-signaling pathway in AGE-induced collagen production in NRK-49F cells. *Journal of Cellular Biochemistry*, 81(1), 102-113.
- Hudson, B. I., Bucciarelli, L. G., Wendt, T., Sakaguchi, T., Lalla, E., Qu, W., ... Schmidt, A. M. (2003). Blockade of receptor for advanced glycation endproducts: A new target for therapeutic intervention in diabetic complications and inflammatory disorders. *Archives of Biochemistry and Biophysics*, 419(1), 80–88.
- Hudson, B. I., Carter, A. M., Harja, E., Kalea, A. Z., Arriero, M., Yang, H., ... Schmidt, A. M. (2007). Identification, classification, and expression of RAGE gene splice variants. *The FASEB Journal*, 22(5), 1572-1580.
- Hugosson, E., Montgomery, S. M., Premji, Z., Troye-Blomberg, M., & Björkman, A. (2004). Higher IL-10 levels are associated with less effective clearance of *Plasmodium falciparum* parasites. *Parasite Immunology*, 26(3), 111-117.
- Hunt, N. H., & Grau, G. E. (2003). Cytokines: accelerators and brakes in the pathogenesis of cerebral malaria. *Trends in Immunology*, 24(9), 491-499.
- Hunter, C. A., Ellis-Neyes, L. A., Slifer, T., Kanaly, S., Grünig, G., Fort, M., ... & Araujo, F. G. (1997). IL-10 is required to prevent immune hyperactivity during infection with *Trypanosoma cruzi*. *The Journal of Immunology*, 158(7), 3311-3316.
- Huttunen, H. J., Fages, C., & Rauvala, H. (1999). Receptor for advanced glycation end products (RAGE)-mediated neurite outgrowth and activation of NF-κB require the cytoplasmic domain of the receptor but different downstream signaling pathways. *Journal of Biological Chemistry*, 274(28), 19919-19924.
- Idro, R., Jenkins, N. E., & Newton, C. R. (2005). Pathogenesis, clinical features, and neurological outcome of cerebral malaria. *The Lancet Neurology*, 4(12), 827-840.
- Idro, R., Marsh, K., John, C. C., & Newton, C. R. (2010). Cerebral malaria: mechanisms of brain injury and strategies for improved neurocognitive outcome. *Pediatric research*, 68(4), 267-274.
- Ishida, H., Imai, T., Suzue, K., Hirai, M., Taniguchi, T., Yoshimura, A., ... & Hisaeda, H. (2013). IL-23 protection against *Plasmodium berghei* infection in mice is partially dependent on IL-17 from macrophages. *European Journal of Immunology*, 43(10), 2696-2706.
- Ishihara, K., Tsutsumi, K., Kawane, S., Nakajima, M., & Kasaoka, T. (2003). The receptor for advanced glycation end-products (RAGE) directly binds to ERK by a D-domain-like docking site. *FEBS Letters*, 550(1), 107-113.
- Ishino, T., Yano, K., Chinzei, Y., & Yuda, M. (2004). Cell-passage activity is required for the malarial parasite to cross the liver sinusoidal cell layer. *PLoS Biology*, 2(1), 77-85.

- Jacobs, P., Radzioch, D., & Stevenson, M. M. (1996). In vivo regulation of nitric oxide production by tumor necrosis factor alpha and gamma interferon, but not by interleukin-4, during blood stage malaria in mice. *Infection and Immunity*, 64(1), 44-49.
- Jakeman, G. N., Saul, A., Hogarth, W. L., & Collins, W. E. (1999). Anaemia of acute malaria infections in non-immune patients primarily results from destruction of uninfected erythrocytes. *Parasitology*, 119(02), 127-133.
- Jambou, R., El-Assaad, F., Combes, V., & Grau, G. E. (2011). In vitro culture of *Plasmodium berghei*-ANKA maintains infectivity of mouse erythrocytes inducing cerebral malaria. *Malaria Journal*, 10(1), 1-5.
- Jaramillo, M., Plante, I., Ouellet, N., Vandal, K., Tessier, P. A., & Olivier, M. (2004). Hemozoin-inducible proinflammatory events in vivo: potential role in malaria infection. *The Journal of Immunology*, 172(5), 3101-3110.
- Jennings, G., & Elia, M. (1987). Effect of *E. coli* endotoxin on temperature, oxygen consumption and brown adipose tissue thermogenesis in rats and mice. *Bioscience Reports*, 7(6), 517-523.
- Jennings, V. M., Lal, A. A., & Hunter, R. L. (1998). Evidence for multiple pathologic and protective mechanisms of murine cerebral malaria. *Infection and Immunity*, 66(12), 5972-5979.
- John, C. C., Opika-Opoka, R., Byarugaba, J., Idro, R., & Boivin, M. J. (2006). Low levels of RANTES are associated with mortality in children with cerebral malaria. *Journal of Infectious Diseases*, 194(6), 837-845.
- John, C. C., Panoskaltsis-Mortari, A., Opoka, R. O., Park, G. S., Orchard, P. J., Jurek, A. M., ... & Boivin, M. J. (2008). Cerebrospinal fluid cytokine levels and cognitive impairment in cerebral malaria. *The American journal of tropical medicine and hygiene*, 78(2), 198-205.
- Jones, K. R., Cottrell, B. J., Targett, G. A., & Playfair, J. H. (1989). Killing of *Plasmodium falciparum* by human monocyte-derived macrophages. *Parasite Immunology*, 11(6), 585-592.
- Joss, A., Akdis, M., Faith, A., Blaser, K., & Akdis, C. A. (2000). IL-10 directly acts on T cells by specifically altering the CD28 co-stimulation pathway. *European Journal of Immunology*, 30(6), 1683-1690.
- Kagi, D., Vignaux, F., Ledermann, B., Burki, K., Depraetere, V., Nagata, S., ... & Golstein, P. (1994). Fas and perforin pathways as major mechanisms of T cell-mediated cytotoxicity. *Science*, 265(5171), 528-530.
- Kai, O. K., & Roberts, D. J. (2008). The pathophysiology of malarial anaemia: where have all the red cells gone?. *BMC Medicine*, 6(1), 24.

- Karunaweera, N. D., Grau, G. E., Gamage, P., Carter, R., & Mendis, K. N. (1992). Dynamics of fever and serum levels of tumor necrosis factor are closely associated during clinical paroxysms in *Plasmodium vivax* malaria. *Proceedings of the National Academy of Sciences*, 89(8), 3200-3203.
- Katsuoka, F., Kawakami, Y., Arai, T., Imuta, H., Fujiwara, M., Kanma, H., & Yamashita, K. (1997). Type II alveolar epithelial cells in lung express receptor for advanced glycation end products (RAGE) gene. *Biochemical and Biophysical Research Communications*, 238(2), 512-516.
- Keller, C. C., Yamo, O., Ouma, C., Ong'echa, J. M., Ounah, D., Hittner, J. B., ... & Perkins, D. J. (2006). Acquisition of hemozoin by monocytes down-regulates interleukin-12 p40 (IL-12p40) transcripts and circulating IL-12p70 through an IL-10-dependent mechanism: in vivo and in vitro findings in severe malarial anemia. *Infection and Immunity*, 74(9), 5249-5260.
- Kern, P., Hemmer, C. J., Damme, J., Gruss, H. J., & Dietrich, M. (1989). Elevated tumor necrosis factor- α and interleukin-6 serum levels as markers for complicated *Plasmodium falciparum* malaria. *The American Journal of Medicine*, 87(2), 139-143.
- Kharazmi, A., Nielsen, H., Rechnitzer, C., & Bendtzen, K. (1989). Interleukin 6 primes human neutrophil and monocyte oxidative burst response. *Immunology Letters*, 21(2), 177-184.
- Kidd, P. (2003). Th1/Th2 balance: the hypothesis, its limitations, and implications for health and disease. *Alternative Medicine Review*, 8(3), 223-246.
- Kim, W., Hudson, B. I., Moser, B., Guo, J., Rong, L. L., Lu, Y., ... & Schmidt, A. (2005). Receptor for advanced glycation end products and its ligands: a journey from the complications of diabetes to its pathogenesis. *Annals of the New York Academy of Sciences*, 1043(1), 553-561.
- Kimura, D., Miyakoda, M., Honma, K., Shibata, Y., Yuda, M., Chinzei, Y., & Yui, K. (2010). Production of IFN- γ by CD4+ T cells in response to malaria antigens is IL-2 dependent. *International Immunology*, 22(12), 941-952.
- Kinyanjui, S. M. (2012). The Immunology of Malaria. In O. Okwa (Ed.), *Malaria Parasites*, (pp. 175-200). Croatia: InTech.
- Kobayashi, F., Ishida, H., Matsui, T., & Tsuji, M. (2000). Effects of in vivo administration of anti-IL-10 or anti-IFN-gamma. Monoclonal antibody on the host defense mechanism against *Plasmodium yoelii yoelii* infection. *Journal of Veterinary Medical Science*, 62(6), 583-587.
- Kochar, D. K., Saxena, V., Singh, N., Kochar, S. K., Kumar, S. V., & Das, A. (2005). *Plasmodium vivax* malaria. *Emerging Infectious Diseases Journal*, 11(1), 132-134.

- Koh, K. H., Chew, P. H., & Kiyu, A. (2004). A retrospective study of malaria infections in an intensive care unit of a general hospital in Malaysia. *Singapore Medical Journal*, 45(1), 28-36.
- Kossodo, S., Monso, C., Juillard, P., Velu, T., Goldman, M., & Grau, G. E. (1997). Interleukin-10 modulates susceptibility in experimental cerebral malaria. *Immunology*, 91(4), 536-540.
- Krishnegowda, G., Hajjar, A. M., Zhu, J., Douglass, E. J., Uematsu, S., Akira, S., ... & Gowda, D. C. (2005). Induction of proinflammatory responses in macrophages by the glycosylphosphatidylinositol of *Plasmodium falciparum* cell signaling receptors, glycosylphosphatidylinositol (GPI) structural requirement, and regulation of GPI activity. *Journal of Biological Chemistry*, 280(9), 8606-8616.
- Kumaratilake, L. M., & Ferrante, A. (1992). IL-4 inhibits macrophage-mediated killing of *Plasmodium falciparum* in vitro. A possible parasite-immune evasion mechanism. *The Journal of Immunology*, 149(1), 194-199.
- Kwiatkowski, D., Sambou, I., Twumasi, P., Greenwood, B. M., Hill, A. V. S., Manogue, K. R., ... & Brewster, D. R. (1990). TNF concentration in fatal cerebral, non-fatal cerebral, and uncomplicated *Plasmodium falciparum* malaria. *The Lancet*, 336(8725), 1201-1204.
- Kwiatkowski, D., Molyneux, M. E., Stephens, S., Curtis, N., Klein, N., Pointaire, P., ... & Greenwood, B. M. (1993). Anti-TNF therapy inhibits fever in cerebral malaria. *QJM*, 86(2), 91-98.
- Lamikanra, A. A., Brown, D., Potocnik, A., Casals-Pascual, C., Langhorne, J., & Roberts, D. J. (2007). Malarial anemia: of mice and men. *Blood*, 110(1), 18-28.
- Lander, H. M., Tauras, J. M., Ogiste, J. S., Hori, O., Moss, R. A., & Schmidt, A. M. (1997). Activation of the receptor for advanced glycation end products triggers a p21 ras-dependent mitogen-activated protein kinase pathway regulated by oxidant stress. *Journal of Biological Chemistry*, 272(28), 17810-17814.
- Langhorne, J. (1994). The immune response to the blood stages of *Plasmodium* in animal models. *Immunology Letters*, 41(2-3), 99-102.
- Langhorne, J., Cross, C., Seixas, E., Li, C., & Von Der Weid, T. (1998). A role for B cells in the development of T cell helper function in a malaria infection in mice. *Proceedings of the National Academy of Sciences*, 95(4), 1730-1734.
- Langhorne, J., Ndungu, F. M., Sponaas, A. M., & Marsh, K. (2008). Immunity to malaria: more questions than answers. *Nature Immunology*, 9(7), 725-732.
- Lee, K. S., Cox-Singh, J., & Singh, B. (2009). Morphological features and differential counts of *Plasmodium knowlesi* parasites in naturally acquired human infections. *Malaria Journal*, 8, 73.

- Li, J., & Schmidt, A. M. (1997). Characterization and functional analysis of the promoter of RAGE, the receptor for advanced glycation end products. *Journal of Biological Chemistry*, 272(26), 16498-16506.
- Li, C., Corraliza, I., & Langhorne, J. (1999). A defect in interleukin-10 leads to enhanced malarial disease in *Plasmodium chabaudi chabaudi* infection in mice. *Infection and Immunity*, 67(9), 4435-4442.
- Li, J., Chang, W. L., Sun, G., Chen, H. L., Specian, R. D., Berney, S. M., ... & van der Heyde, H. C. (2003). Intercellular adhesion molecule 1 is important for the development of severe experimental malaria but is not required for leukocyte adhesion in the brain. *Journal of Investigative Medicine*, 51(3), 128-140.
- Liang, S. C., Tan, X. Y., Luxenberg, D. P., Karim, R., Dunussi-Joannopoulos, K., Collins, M., & Fouser, L. A. (2006). Interleukin (IL)-22 and IL-17 are coexpressed by Th17 cells and cooperatively enhance expression of antimicrobial peptides. *The Journal of Experimental Medicine*, 203(10), 2271-2279.
- Liliensiek, B., Weigand, M. A., Bierhaus, A., Nicklas, W., Kasper, M., Hofer, S., ... & Arnold, B. (2004). Receptor for advanced glycation end products (RAGE) regulates sepsis but not the adaptive immune response. *Journal of Clinical Investigation*, 113(11), 1641-1650.
- Lin, L., Park, S., & Lakatta, E. G. (2009). RAGE signaling in inflammation and arterial aging. *Frontiers in Bioscience-landmark*, 14, 1403-1413.
- Linke, A., Kühn, R., Müller, W., Honarvar, N., Li, C., & Langhorne, J. (1996). *Plasmodium chabaudi chabaudi*: differential susceptibility of gene-targeted mice deficient in IL-10 to an erythrocytic-stage infection. *Experimental Parasitology*, 84(2), 253-263.
- Lokman H. (2011). Malaria trend in Malaysia 1995-2010. Retrieved December 18, 2012, from <http://asmic.akademisains.gov.my/download/tropical/Lokman.pdf>
- Lokuta, M. A., & Huttenlocher, A. (2005). TNF- α promotes a stop signal that inhibits neutrophil polarization and migration via a p38 MAPK pathway. *Journal of Leukocyte Biology*, 78(1), 210-219.
- Lotze, M. T., & Tracey, K. J. (2005). High-mobility group box 1 protein (HMGB1): nuclear weapon in the immune arsenal. *Nature Reviews Immunology*, 5(4), 331-342.
- Lovegrove, F. E., Gharib, S. A., Peña-Castillo, L., Patel, S. N., Ruzinski, J. T., Hughes, T. R., ... & Kain, K. C. (2008). Parasite burden and CD36-mediated sequestration are determinants of acute lung injury in an experimental malaria model. *PLoS Pathogens*, 4(5), e1000068.
- Lutterloh, E. C., Opal, S. M., Pittman, D. D., Keith Jr., J. C., Tan, X., Clancy, B. M., ... Kessimian, N. (2007). Inhibition of the RAGE products increases survival in

- experimental models of severe sepsis and systemic infection. *Critical Care*, 11(6), R122.
- Luty, A. J., Lell, B., Schmidt-Ott, R., Lehman, L. G., Luckner, D., Greve, B., ... & Kremsner, P. G. (1999). Interferon- γ responses are associated with resistance to reinfection with *Plasmodium falciparum* in young African children. *Journal of Infectious Diseases*, 179(4), 980-988.
- Luty, A. J., Perkins, D. J., Lell, B., Schmidt-Ott, R., Lehman, L. G., Luckner, D., ... & Kremsner, P. G. (2000). Low interleukin-12 activity in severe *Plasmodium falciparum* malaria. *Infection and Immunity*, 68(7), 3909-3915.
- Lyke, K. E., Burges, R., Cissoko, Y., Sangare, L., Dao, M., Diarra, I., ... & Sztein, M. B. (2004). Serum levels of the proinflammatory cytokines interleukin-1 beta (IL-1 β), IL-6, IL-8, IL-10, tumor necrosis factor alpha, and IL-12 (p70) in Malian children with severe *Plasmodium falciparum* malaria and matched uncomplicated malaria or healthy controls. *Infection and immunity*, 72(10), 5630-5637.
- Mackey, L. J., Hochmann, A., June, C. H., Contreras, C. E., & Lambert, P. H. (1980). Immunopathological aspects of *Plasmodium berghei* infection in five strains of mice. II. Immunopathology of cerebral and other tissue lesions during the infection. *Clinical and Experimental Immunology*, 42(3), 412-420.
- Mackinnon, M. J., & Read, A. F. (1999). Genetic relationship between virulence and transmissibility in the rodent malaria *Plasmodium chabaudi*. *Evolution*, 53(3), 689-703.
- Mackintosh, C. L., Beeson, J. G., & Marsh, K. (2004). Clinical features and pathogenesis of severe malaria. *Trends in Parasitology*, 20(12), 597-603.
- Maeda, H., & Shiraishi, A. (1996). TGF- β contributes to the shift toward Th2-type response through direct and interleukin-10-mediated pathways in tumor bearing mice. *The Journal of Immunology*, 156(1), 73-78.
- Maegraith, B., & Fletcher, A. (1971). The pathogenesis of mammalian malaria. *Advances in Parasitology*, 10, 49-75.
- Maguire, G. P., Handojo, T., Pain, M. C., Kenangalem, E., Price, R. N., Tjitra, E., & Anstey, N. M. (2005). Lung injury in uncomplicated and severe falciparum malaria: a longitudinal study in Papua, Indonesia. *Journal of Infectious Diseases*, 192(11), 1966-1974.
- Malaguarnera, L., & Musumeci, S. (2002). The immune response to *Plasmodium falciparum* malaria. *The Lancet Infectious Diseases*, 2(8), 472-478.
- Malaguarnera, L., Pignatelli, S., Musumeci, M., Simpore, J., & Musumeci, S. (2002). Plasma levels of interleukin-18 and interleukin-12 in *Plasmodium falciparum* malaria. *Parasite Immunology*, 24(9-10), 489-492.

Malaria Epidemiology in the Western Pacific Malaysia. Retrieved February 16, 2012 from Western Pacific Region World Health Organization (WPRWHO).Website:
http://www.wpro.who.int/sites/mvp/epidemiology/malaria/maa_profile.htm

- Malherbe, P., Richards, J. G., Gaillard, H., Thompson, A., Diener, C., Schuler, A., & Huber, G. (1999). cDNA cloning of a novel secreted isoform of the human receptor for advanced glycation end products and characterization of cells co-expressing cell-surface scavenger receptors and Swedish mutant amyloid precursor protein. *Molecular Brain Research*, 71(2), 159-170.
- Manfredi, A. A., Capobianco, A., Esposito, A., Cobelli, F. D., Canu, T., Monno, A., ... Maschio, A. D. (2008). Maturing dendritic cells depend on RAGE for *in vivo* homing to lymph nodes. *The Journal of Immunology*, 180(4), 2270-2275.
- Martins, Y. C., Smith, M. J., Pelajo-Machado, M., Werneck, G. L., Lenzi, H. L., Daniel-Ribeiro, C. T., & Carvalho, L. J. (2009). Characterization of cerebral malaria in the outbred Swiss Webster mouse infected by *Plasmodium berghei* ANKA. *International Journal of Experimental Pathology*, 90(2), 119-130.
- Mastelic, B., do Rosario, A. P. F., Veldhoen, M., Renauld, J. C., Jarra, W., Sponaas, A. M., ... & Langhorne, J. (2012). IL-22 protects against liver pathology and lethality of an experimental blood-stage malaria infection. *Frontiers in Immunology*, 3, 85.
- Mate-Kole, M. O., Yeboah, E. D., Affram, R. K., & Adu, D. (1996). Blackwater fever and acute renal failure in expatriates in Africa. *Renal Failure*, 18(3), 525-531.
- Mazier, D., Rénia, L., & Snounou, G. (2009). A pre-emptive strike against malaria's stealthy hepatic forms. *Nature Reviews Drug Discovery*, 8(11), 854-864.
- McGilvray, I. D., Serghides, L., Kapus, A., Rotstein, O. D., & Kain, K. C. (2000). Nonopsonic monocyte/macrophage phagocytosis of *Plasmodium falciparum*-parasitized erythrocytes: a role for CD36 in malarial clearance. *Blood*, 96(9), 3231-3240.
- Miller, L. H., Good, M. F., & Milon, G. (1994). Malaria pathogenesis. *Science*, 264(5167), 1878-1883.
- Ministry of Health Malaysia. (2011). Annual report 2011. Retrieved December 18, 2012, from http://www.moh.gov.my/images/gallery/publications/md/ar/2011_en.pdf
- Miura, S., Suematsu, M., Tanaka, S., Nagata, H., Houzawa, S., Suzuki, M., ... & Tsuchiya, M. (1991). Microcirculatory disturbance in indomethacin-induced intestinal ulcer. *American Journal of Physiology-Gastrointestinal and Liver Physiology*, 261(2), G213-G219.
- Mohan, K., Moulin, P., & Stevenson, M. M. (1997). Natural killer cell cytokine production, not cytotoxicity, contributes to resistance against blood-stage

- Plasmodium chabaudi* AS infection. *The Journal of Immunology*, 159(10), 4990-4998.
- Mohan, K., & Stevenson, M. M. (1998). Dyserythropoiesis and severe anaemia associated with malaria correlate with deficient interleukin-12 production. *British Journal of Haematology*, 103, 942-949.
- Mohan, A., Sharma, S. K., & Bollineni, S. (2008). Acute lung injury and acute respiratory distress syndrome in malaria. *Journal of Vector Borne Disease*, 45(3), 179-193.
- Moore, K. W., de Waal Malefyt, R., Coffman, R. L., & O'Garra, A. (2001). Interleukin-10 and the interleukin-10 receptor. *Annual Review of Immunology*, 19(1), 683-765.
- Moore, B. R., Jago, J. D., & Batty, K. T. (2008). *Plasmodium berghei*: parasite clearance after treatment with dihydroartemisinin in an asplenic murine malaria model. *Experimental Parasitology*, 118(4), 458-467.
- Mordmüller, B. G., Metzger, W. G., Juillard, P., Brinkman, B. M., Verweij, C. L., Grau, G. E., & Kremsner, P. G. (1997). Tumor necrosis factor in *Plasmodium falciparum* malaria: high plasma level is associated with fever, but high production capacity is associated with rapid fever clearance. *European Cytokine Network*, 8(1), 29-35.
- Morrot, A., & Zavala, F. (2004). Effector and memory CD8+ T cells as seen in immunity to malaria. *Immunological Reviews*, 201(1), 291-303.
- Moser, B., Desai, D. D., Downie, M. P., Chen, Y., Yan, S. F., Herold, K., ... & Clynes, R. (2007). Receptor for advanced glycation end products expression on T cells contributes to antigen-specific cellular expansion in vivo. *The Journal of Immunology*, 179(12), 8051-8058.
- Moxon, C. A., Heyderman, R. S., & Wassmer, S. C. (2009). Dysregulation of coagulation in cerebral malaria. *Molecular and Biochemical Parasitology*, 166(2), 99-108.
- Mshana, R. N., Boulandi, J., Mshana, N. M., Mayombo, J., & Mendome, G. (1991). Cytokines in the pathogenesis of malaria: levels of IL-1 beta, IL-4, IL-6, TNF-alpha and IFN-gamma in plasma of healthy individuals and malaria patients in a holoendemic area. *Journal of Clinical & Laboratory Immunology*, 34(3), 131-139.
- Muniz-Junqueira, M. I., dos Santos-Neto, L. L., & Tosta, C. E. (2001). Influence of tumor necrosis factor-alpha on the ability of monocytes and lymphocytes to destroy intraerythrocytic *Plasmodium falciparum* in vitro. *Cell Immunology*, 208, 73-79.
- Murphy, G. S., & Oldfield, E. C. (1996). Falciparum malaria. *Infectious disease clinics of North America*, 10(4), 747-775.

- Murphy, K. M., & Reiner, S. L. (2002). The lineage decisions of helper T cells. *Nature Reviews Immunology*, 2(12), 933-944.
- Musumeci, M., Malaguarnera, L., Simpore, J., Messina, A., & Musumeci, S. (2003). Modulation of immune response in *Plasmodium falciparum* malaria: role of IL-12, IL-18 and TGF- β . *Cytokine*, 21(4), 172-178.
- Nacer, A., Movila, A., Baer, K., Mikolajczak, S. A., Kappe, S. H., & Frevert, U. (2012). Neuroimmunological blood brain barrier opening in experimental cerebral malaria. *PLoS Pathogens*, 8(10), e1002982.
- Nagamine, Y., Hayano, M., Kashiwamura, S., Okamura, H., Nakanishi, K., Krudsod, S., Wilairatana, P., Looareesuwan, S., & Kojima, S. (2003). Involvement of interleukin-18 in severe *Plasmodium falciparum* malaria. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 97(2), 236-241.
- Nand, N., Aggarwal, H., Sharma, M., & Singh, M. (2001). Systemic manifestations of malaria. *Journal, Indian Academy of Clinical Medicine*, 2(3), 189-194.
- Nepper, M., Schmidt, A. M., Brett, J., Yan, S. D., Wang, F., Pan, Y. C., ... Shaw, A. (1992). Cloning and expression of a cell surface receptor for advanced glycosylation end products of proteins. *Journal of Biological Chemistry*, 267(21), 14998-15004.
- Neill, A. L., & Hunt, N. H. (1992). Pathology of fatal and resolving *Plasmodium berghei* cerebral malaria in mice. *Parasitology*, 105(02), 165-175.
- Nelms, K., Keegan, A. D., Zamorano, J., Ryan, J. J., & Paul, W. E. (1999). The IL-4 receptor: signaling mechanisms and biologic functions. *Annual Review of Immunology*, 17(1), 701-738.
- Newbold, C., Craig, A., Kyes, S., Rowe, A., Fernandez-Reyes, D., & Fagan, T. (1999). Cytoadherence, pathogenesis and the infected red cell surface in *Plasmodium falciparum*. *International Journal for Parasitology*, 29(6), 927-937.
- Nguansangiam, S., Day, N. P., Hien, T. T., Mai, N. T. H., Chaisri, U., Riganti, M., ... & Pongponratn, E. (2007). A quantitative ultrastructural study of renal pathology in fatal *Plasmodium falciparum* malaria. *Tropical Medicine & International Health*, 12(9), 1037-1050.
- Nie, C. Q., Bernard, N. J., Schofield, L., & Hansen, D. S. (2007). CD4+ CD25+ regulatory T cells suppress CD4+ T-cell function and inhibit the development of *Plasmodium berghei*-specific TH1 responses involved in cerebral malaria pathogenesis. *Infection and Immunity*, 75(5), 2275-2282.
- Nie, C. Q., Bernard, N. J., Norman, M. U., Amante, F. H., Lundie, R. J., Crabb, B. S., ... & Hansen, D. S. (2009). IP-10-mediated T cell homing promotes cerebral inflammation over splenic immunity to malaria infection. *PLoS Pathogens*, 5(4), e1000369.

- Niikura, M., Kamiya, S., Kita, K., & Kobayashi, F. (2008). Coinfection with nonlethal murine malaria parasites suppresses pathogenesis caused by *Plasmodium berghei* NK65. *The Journal of Immunology*, 180(10), 6877-6884.
- Niikura, M., Kamiya, S., Nakane, A., Kita, K., & Kobayashi, F. (2010). IL-10 plays a crucial role for the protection of experimental cerebral malaria by co-infection with non-lethal malaria parasites. *International Journal for Parasitology*, 40(1), 101-108.
- Nussenblatt, V., & Semba, R. D. (2002). Micronutrient malnutrition and the pathogenesis of malarial anemia. *Acta Tropica*, 82(3), 321-337.
- Nyangoto, E. O. (2005). Cell-mediated effector molecules and complicated malaria. *International Archives of Allergy and Immunology*, 137(4), 326-342.
- Oakley, M. S., Gerald, N., McCutchan, T. F., Aravind, L., & Kumar, S. (2011). Clinical and molecular aspects of malaria fever. *Trends in parasitology*, 27(10), 442-449.
- Ochola, L. B., Siddondo, B. R., Ocholla, H., Nkya, S., Kimani, E. N., Williams, T. N., ... & Craig, A. G. (2011). Specific receptor usage in *Plasmodium falciparum* cytoadherence is associated with disease outcome. *PloS One*, 6(3), e14741.
- O'Connor Jr, W., Zenewicz, L. A., & Flavell, R. A. (2010). The dual nature of TH17 cells: shifting the focus to function. *Nature Immunology*, 11(6), 471-476.
- O'Garra, A., & Arai, N. (2000). The molecular basis of T helper 1 and T helper 2 cell differentiation. *Trends in Cell Biology*, 10(12), 542-550.
- Okamura, H., Tsutsui, H., Komatsu, T., Yutsudo, M., Hakura, A., Tanimoto, T., ... & Kurimoto, M. (1995). Cloning of a new cytokine that induces IFN- γ production by T cells. *Nature*, 378(6552), 88-91.
- Oliveira, G. A., Kumar, K. A., Calvo-Calle, J. M., Othoro, C., Altszuler, D., Nussenzwieg, V., & Nardin, E. H. (2008). Class II-restricted protective immunity induced by malaria sporozoites. *Infection and Immunity*, 76(3), 1200-1206.
- Omer, F. M., Kurtzhals, J. A., & Riley, E. M. (2000). Maintaining the immunological balance in parasitic infections: a role for TGF- β ? *Parasitology Today*, 16(1), 18-23.
- Omer, F. M., de Souza, J. B., & Riley, E. M. (2003). Differential induction of TGF- β regulates proinflammatory cytokine production and determines the outcome of lethal and nonlethal *Plasmodium yoelii* infections. *The Journal of Immunology*, 171(10), 5430-5436.
- Orago, A. S. S., & Facer, C. A. (1991). Cytotoxicity of human natural killer (NK) cell subsets for *Plasmodium falciparum* erythrocytic schizonts: stimulation by

- cytokines and inhibition by neomycin. *Clinical & Experimental Immunology*, 86(1), 22-29.
- Orlova, V. V., Choi, E. Y., Xie, C., Chavakis, E., Bierhaus, A., Iharus, E., ... Chavakis, T. (2007). A novel pathway of HMGB1-mediated inflammatory cell recruitment that requires Mac1-integrin. *The EMBO Journal*, 26(4), 1129-1139.
- Ostendorp, T., Leclerc, E., Galichet, A., Koch, M., Demling, N., Weigle, B., ... Fritz, G. (2007). Structural and functional insights into RAGE activation by multimeric S100B. *The EMBO Journal*, 26(16), 3868-3878.
- Ouyang, W., Kolls, J. K., & Zheng, Y. (2008). The biological functions of T helper 17 cell effector cytokines in inflammation. *Immunity*, 28(4), 454-467.
- Overstreet, M. G., Cockburn, I. A., Chen, Y. C., & Zavala, F. (2008). Protective CD8+ T cells against Plasmodium liver stages: Immunobiology of an ‘unnatural’ immune response. *Immunological Reviews*, 225(1), 272-283.
- Palm, N. W., & Medzhitov, R. (2009). Pattern recognition receptors and control of adaptive immunity. *Immunological Reviews*, 227(1), 221-233.
- Park, L., Raman, K. G., Lee, K. J., Lu, Y., Ferran, L. J., Chow, W. S., ... Schmidt, A. M. (1998). Suppression of accelerated diabetic atherosclerosis by the soluble receptor for advanced glycation endproducts. *Nature Medicine*, 4(9), 1025-1031.
- Park, J. S., Arcaroli, J., Yum, H. K., Yang, H., Wang, H., Yang, K. Y., ... & Abraham, E. (2003). Activation of gene expression in human neutrophils by high mobility group box 1 protein. *American Journal of Physiology-Cell Physiology*, 284(4), C870-C879.
- Parroche, P., Lauw, F. N., Goutagny, N., Latz, E., Monks, B. G., Visintin, A., ... & Golenbock, D. T. (2007). Malaria hemozoin is immunologically inert but radically enhances innate responses by presenting malaria DNA to Toll-like receptor 9. *Proceedings of the National Academy of Sciences*, 104(6), 1919-1924.
- Perkins, D. J., Weinberg, J. B., & Kremsner, P. G. (2000). Reduced interleukin-12 and transforming growth factor- β 1 in severe childhood malaria: relationship of cytokine balance with disease severity. *Journal of Infectious Diseases*, 182(3), 988-992.
- Perkins, D. J., Were, T., Davenport, G. C., Kempaiah, P., Hittner, J. B., & Ong'echa, J. M. (2011). Severe malarial anemia: innate immunity and pathogenesis. *International Journal of Biological Sciences*, 7(9), 1427-1442.
- Perlmann, P., & Troye-Bloomberg, M. (2002). Malaria and the immune system in humans. *Malaria Immunology*, 80, 229-242.

- Perry, J. A., Rush, A., Wilson, R. J., Olver, C. S., & Avery, A. C. (2004). Dendritic cells from malaria-infected mice are fully functional APC. *The Journal of Immunology*, 172(1), 475-482.
- Perry, B. C., Soltys, D., Toledo, A. H., & Toledo-Pereyra, L. H. (2011). Tumor necrosis factor- α in liver ischemia/reperfusion injury. *Journal of Investigative Surgery*, 24(4), 178-188.
- Phu, N. H., Day, N., Diep, P. T., Ferguson, D. J., & White, N. J. (1995). Intraleucocytic malaria pigment and prognosis in severe malaria. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 89(2), 200-204.
- Pichyangkul, S., Saengkrai, P., & Webster, H. K. (1994). *Plasmodium falciparum* pigment induces monocytes to release high levels of tumor necrosis factor-alpha and interleukin-1 beta. *The American Journal of Tropical Medicine and Hygiene*, 51(4), 430-435.
- Pied, S., Civas, A., Berlot-Picard, F., Renia, L., Miltgen, F., Gentilini, M., ... & Mazier, D. (1992). IL-6 induced by IL-1 inhibits malaria pre-erythrocytic stages but its secretion is down-regulated by the parasite. *The Journal of Immunology*, 148(1), 197-201.
- Plebanski, M., & Hill, A. V. (2000). The immunology of malaria infection. *Current opinion in immunology*, 12(4), 437-441.
- Pongponratn, E., Turner, G. D., Day, N. P., Phu, N. H., Simpson, J. A., Stepniewska, K., ... & White, N. J. (2003). An ultrastructural study of the brain in fatal *Plasmodium falciparum* malaria. *The American Journal of Tropical Medicine and Hygiene*, 69(4), 345-359.
- Prakash, D., Fesel, C., Jain, R., Cazenave, P. A., Mishra, G. C., & Pied, S. (2006). Clusters of cytokines determine malaria severity in *Plasmodium falciparum*-infected patients from endemic areas of Central India. *Journal of Infectious Diseases*, 194(2), 198-207.
- Price, R. N., Tjitra, E., Guerra, C. A., Yeung, S., White, N. J., & Anstey, N. M. (2007). Vivax malaria: neglected and not benign. *The American journal of tropical medicine and hygiene*, 77(6 Suppl), 79-87.
- Prommano, O., Chaisri, U., Turner, G. D., Wilairatana, P., Ferguson, D. J., Viriyavejakul, P., White, N. J., & Pongponratn, E. (2005). A quantitative ultrastructural study of the liver and the spleen in fatal *falciparum* malaria. *The Southeast Asian Journal of Tropical Medicine and Public Health*, 36(6), 1359-1370.
- Protzer, U., Maini, M. K., & Knolle, P. A. (2012). Living in the liver: hepatic infections. *Nature Reviews Immunology*, 12(3), 201-213.
- Pulido-Mendez, M., Finol, H. J., Giron, M. E., & Aguilar, I. (2005). Ultrastructural pathological changes in mice kidney caused by *Plasmodium berghei*

- infection. *Journal of Submicroscopic Cytology and Pathology*, 38(2-3), 143-148.
- Purcell, L. A., Wong, K. A., Yanow, S. K., Lee, M., Spithill, T. W., & Rodriguez, A. (2008). Chemically attenuated *Plasmodium* sporozoites induce specific immune responses, sterile immunity and cross-protection against heterologous challenge. *Vaccine*, 26(38), 4880-4884.
- Rajapurkar, M. M. (1994). Renal involvement in malaria. *Journal of Postgraduate Medicine*, 40(3), 132-134.
- Rajkumar, A., Rao, S., & Sundaram, S. (2012). Clinical outcome in malaria - Reiterating the role of parasitic index. *Indian Journal of Clinical Practice*, 22(9), 450-453.
- Ramasamy, R., Yan, S. F., Herold, K., Clynes, R., & Schmidt, A. M. (2008). Receptor for advanced glycation end products: Fundamental roles in the inflammatory response: Winding the way to the pathogenesis of endothelial dysfunction and atherosclerosis. *Annals of The New York Academy of Sciences*, 1126, 7-13.
- Rasheed, Z., Akhtar, N., & Haqqi, T. M. (2010). Advanced glycation end products induce the expression of interleukin-6 and interleukin-8 by receptor for advanced glycation end product-mediated activation of mitogen-activated protein kinases and nuclear factor- κ B in human osteoarthritis chondrocytes. *Rheumatology*, 50, 838-851.
- Raucci, A., Cugusi, S., Antonelli, A., Barabino, S. M., Monti, L., Bierhaus, A., ... Bianchi, M. E. (2008). A soluble form of the receptor for advanced glycation endproducts (RAGE) is produced by proteolytic cleavage of the membrane-bound form by the sheddase a disintegrin and metalloprotease 10 (ADAM10). *The FASEB Journal*, 22(10), 3716-3727.
- Reynolds, P. R., Schmitt, R. E., Kasteler, S. D., Sturrock, A., Sanders, K., Bierhaus, A., ... Hoidal, J. R. (2010). Receptors for advanced glycation end-products targeting protect against hyperoxia-induced lung injury in mice. *American Journal of Respiratory Cell and Molecular Biology*, 42(5), 545-551.
- Rich, S. M., & Ayala, F. J. (2006). Evolutionary origins of human malaria parasites. In *Malaria: Genetic and Evolutionary Aspects* (pp. 125-146). Springer US.
- Richards, A. L. (1997). Tumour necrosis factor and associated cytokines in the host's response to malaria. *International Journal for Parasitology*, 27(10), 1251-1263.
- Riley, E. M. (1999). Is T-cell priming required for initiation of pathology in malaria infections?. *Immunology Today*, 20(5), 228-233.
- Riley, E. M., Wahl, S., Perkins, D. J., & Schofield, L. (2006). Regulating immunity to malaria. *Parasite Immunology*, 28(1-2), 35-49.

- Riley, E. M., Couper, K. N., Helmby, H., Hafalla, J. C., de Souza, J. B., Langhorne, J., ... & Zavala, F. (2010). Neuropathogenesis of human and murine malaria. *Pathology*, 76, 410-415.
- Rockett, K. A., Awburn, M. M., Cowden, W. B., & Clark, I. A. (1991). Killing of *Plasmodium falciparum* in vitro by nitric oxide derivatives. *Infection and Immunity*, 59(9), 3280-3283.
- Rockett, K. A., Awburn, M. M., Aggarwal, B. B., Cowden, W. B., & Clark, I. A. (1992). In vivo induction of nitrite and nitrate by tumor necrosis factor, lymphotoxin, and interleukin-1: possible roles in malaria. *Infection and Immunity*, 60(9), 3725-3730.
- Rockett, K. A., Awburn, M. M., Rockett, E. J., & Clark, I. A. (1994). Tumor necrosis factor and interleukin-1 synergy in the context of malaria pathology. *The American Journal of Tropical Medicine and Hygiene*, 50(6), 735-742.
- Rodrigues, M. M., Cordey, A. S., Arreaza, G., Corradin, G., Romero, P., Maryanski, J. L., ... & Zavala, F. (1991). CD8+ cytolytic T cell clones derived against the *Plasmodium yoelii* circumsporozoite protein protect against malaria. *International Immunology*, 3(6), 579-585.
- Rodriguez-Acosta, A., Finol, H. J., Pulido-Mendez, M., Marquez, A., Andrade, G., Gonzalez, N., ... & Pinto, A. (1998). Liver ultrastructural pathology in mice infected with *Plasmodium berghei*. *Journal of Submicroscopic Cytology and Pathology*, 30(2), 299-307.
- Roll back malaria. (2010). Key malaria facts. Retrieved December 29, 2010, from <http://www.rollbackmalaria.org/keyfacts.html>
- Romano, M., Sironi, M., Toniatti, C., Polentarutti, N., Fruscella, P., Ghezzi, P., ... & Mantovani, A. (1997). Role of IL-6 and its soluble receptor in induction of chemokines and leukocyte recruitment. *Immunity*, 6(3), 315-325.
- Rousset, F., Garcia, E., Defrance, T., Peronne, C., Vezzio, N., Hsu, D. H., ... & Banchereau, J. (1992). Interleukin 10 is a potent growth and differentiation factor for activated human B lymphocytes. *Proceedings of the National Academy of Sciences*, 89(5), 1890-1893.
- Rowe, J. A., Claessens, A., Corrigan, R. A., & Arman, M. (2009). Adhesion of *Plasmodium falciparum*-infected erythrocytes to human cells: molecular mechanisms and therapeutic implications. *Expert Reviews in Molecular Medicine*, 11, e16.
- Rudin, W., Eugster, H. P., Bordmann, G., Bonato, J., Muller, M., Yamage, M., & Ryffel, B. (1997). Resistance to cerebral malaria in tumor necrosis factor- α/β -deficient mice is associated with a reduction of intercellular adhesion molecule- up-regulation and T helper type 1 response. *The American Journal of Pathology*, 150(1), 257-266.

- Rudin, W., Favre, N., Bordmann, G., & Ryffel, B. (1997). Interferon- γ is essential for the development of cerebral malaria. *European Journal of Immunology*, 27(4), 810-815.
- Rui-Mei, L., Kara, A. U., & Sinniah, A. (1998). Dysregulation of cytokine expression in tubulointerstitial nephritis associated with murine malaria. *Kidney International*, 53(4), 845-852.
- Rundi C. *Malaria elimination in Malaysia*. Presented at third annual meeting of the Asia Pacific Malaria Elimination Network (APMEN), Sabah, Malaysia. May 2011. Retrieved May 3, 2012, from <http://apmen.org/apmen-iii-meeting-proceedings/>
- Ryan-Payseur, B., Ali, Z., Huang, D., Chen, C. Y., Yan, L., Wang, R. C., ... & Chen, Z. W. (2011). Virus infection stages and distinct Th1 or Th17/Th22 T-cell responses in malaria/SHIV coinfection correlate with different outcomes of disease. *Journal of Infectious Diseases*, 204(9), 1450-1462.
- Sam, H., & Stevenson, M. M. (1999). Early IL-12 p70, but not p40, production by splenic macrophages correlates with host resistance to blood-stage *Plasmodium chabaudi* AS malaria. *Clinical and Experimental Immunology*, 117, 343-349.
- Sanni, L. A., Fonseca, L. F., & Langhorne, J. (2002). Mouse models for erythrocytic-stage malaria. *Methods in Molecular Medicine*, 72, 57-76.
- Sanni, L. A., Jarra, W., Li, C., & Langhorne, J. (2004). Cerebral edema and cerebral hemorrhages in interleukin-10-deficient mice infected with *Plasmodium chabaudi*. *Infection and Immunity*, 72(5), 3054-3058.
- Santilli, F., Vazzana, N., Bucciarelli, L. G., & Davi, G. (2009). Soluble forms of RAGE in human diseases: clinical and therapeutical implications. *Current Medicinal Chemistry*, 16(8), 940-952.
- Sarangi, A., Mohapatra, P. C., Dalai, R. K., & Sarangi, A. K. (2014). Serum IL-4, IL-12 and TNF-alpha in malaria: a comparative study associating cytokine responses with severity of disease from the Coastal Districts of Odisha. *Journal of Parasitic Diseases*, 38(2), 143-147.
- Sarfo, B. Y., Wilson, N. O., Bond, V. C., & Stiles, J. K. (2011). *Plasmodium berghei* ANKA infection increases Foxp3, IL-10 and IL-2 in CXCL-10 deficient C57BL/6 mice. *Malaria Journal*, 10, 69.
- Schmidt, A. M., Vianna, M., Gerlach, M., Brett, J., Ryan, J., Kao, J., ... Stern, D. (1992). Isolation and characterization of two binding proteins for advanced glycosylation end products from bovine lung which are present on the endothelial cell surface. *Journal of Biological Chemistry*, 267(21), 14987-14997.
- Schmidt, A. M., Yan, S. D., Brett, J., Mora, R., Nowyngrod, R., & Stern, D. (1993). Regulation of human mononuclear phagocyte migration by cell surface-

- binding proteins for advanced glycation end products. *Journal of Clinical Investigation*, 91(5), 2155–2168.
- Schmidt, A. M., Yan, S. D., Yan, S. F., & Stern, D. M. (2000). The biology of the receptor for advanced glycation end products and its ligands. *Biochimica Et Biophysica Acta-molecular Cell Research*, 1498, 99-111.
- Schmidt, A. M., Yan, S. D., Yan, S. F., & Stern, D. M. (2001). The multiligand receptor RAGE as a progression factor amplifying immune and inflammatory responses. *Journal of Clinical Investigation*, 108(7), 949-955.
- Schofield, L., & Hackett, F. (1993). Signal transduction in host cells by a glycosylphosphatidylinositol toxin of malaria parasites. *The Journal of Experimental Medicine*, 177(1), 145-153.
- Schofield, L., & Grau, G. E. (2005). Immunological processes in malaria pathogenesis. *Nature Reviews Immunology*, 5(9), 722-735.
- Schwartz, J. E., Scuderi, P., Wiggins, C., Rudolph, A., & Hersh, E. M. (1989). A phase I trial of recombinant tumor necrosis factor (rTNF) administered by continuous intravenous infusion in patients with disseminated malignancy. *Biotherapy*, 1(3), 207-214.
- Seixas, E., Gozzelino, R., Chora, Â., Ferreira, A., Silva, G., Larsen, R., ... & Soares, M. P. (2009). Heme oxygenase-1 affords protection against noncerebral forms of severe malaria. *Proceedings of the National Academy of Sciences*, 106(37), 15837-15842.
- Serghides, L., Smith, T. G., Patel, S. N., & Kain, K. C. (2003). CD36 and malaria: friends or foes?. *Trends in Parasitology*, 19(10), 461-469.
- Shah, S., Ali, L., Sattar, R. A., Aziz, T., Ansari, T., & Ara, J. (2009). Malaria hepatopathy in falciparum malaria. *Journal of the College of Physicians and Surgeons Pakistan*, 19(6), 367-70.
- Sherman, I. W. (2009). Reflections on a century of malaria biochemistry. *Advances in Parasitology*, 67, 25-47.
- Sherry, B. A., Alava, G., Tracey, K. J., Martiney, J., Cerami, A., & Slater, A. F. (1994). Malaria-specific metabolite hemozoin mediates the release of several potent endogenous pyrogens (TNF, MIP-1 alpha, and MIP-1 beta) in vitro, and altered thermoregulation in vivo. *Journal of Inflammation*, 45(2), 85-96.
- Shi, Y. P., Nahlen, B. L., Kariuki, S., Urdahl, K. B., McElroy, P. D., Roberts, J. M., & Lal, A. A. (2001). Fcy receptor IIa (CD32) polymorphism is associated with protection of infants against high-density *Plasmodium falciparum* infection. VII. Asembo Bay Cohort Project. *Journal of Infectious Diseases*, 184(1), 107-111.
- Shibui, A., Hozumi, N., Shiraishi, C., Sato, Y., Iida, H., Sugano, S., & Watanabe, J. (2009). CD4+ T cell response in early erythrocytic stage malaria: *Plasmodium*

- berghei* infection in BALB/c and C57BL/6 mice. *Parasitology Research*, 105(1), 281-286.
- Shio, M. T., Kassa, F. A., Bellemare, M. J., & Olivier, M. (2010). Innate inflammatory response to the malarial pigment hemozoin. *Microbes and Infection*, 12(12), 889-899.
- Simpson, K. J., Henderson, N. C., Bone-Larson, C. L., Lukacs, N. W., Hogaboam, C. M., & Kunkel, S. L. (2003). Chemokines in the pathogenesis of liver disease: so many players with poorly defined roles. *Clinical Science*, 104(1), 47-63.
- Singh, B., Sung, L. K., Matusop, A., Radhakrishnan, A., Shamsul, S. S., Cox-Singh, J., ... & Conway, D. J. (2004). A large focus of naturally acquired *Plasmodium knowlesi* infections in human beings. *The Lancet*, 363(9414), 1017-1024.
- Sinniah, R., Rui-Mei, L., & Kara, A. U. (1999). Up-regulation of cytokines in glomerulonephritis associated with murine malaria infection. *International journal of Experimental Pathology*, 80(2), 87-95.
- Skorokhod, O. A., Caione, L., Marrocco, T., Migliardi, G., Barrera, V., Arese, P., ... & Schwarzer, E. (2010). Inhibition of erythropoiesis in malaria anemia: role of hemozoin and hemozoin-generated 4-hydroxynonenal. *Blood*, 116(20), 4328-4337.
- Snounou, G., Jarra, W., & Preiser, P. R. (2000). Malaria multigene families: the price of chronicity. *Parasitology Today*, 16(1), 28-30.
- Song, S., Ling-Hu, H., Roebuck, K. A., Rabbi, M. F., Donnelly, R. P., & Finnegan, A. (1997). Interleukin-10 inhibits interferon- γ -induced intercellular adhesion molecule-1 gene transcription in human monocytes. *Blood*, 89(12), 4461-4469.
- Sparvero, L. J., Asafu-Adjei, D., Kang, R., Tang, D., Amin, N., Im, J., ... & Lotze, M. T. (2009). RAGE (Receptor for Advanced Glycation Endproducts), RAGE ligands, and their role in cancer and inflammation. *Journal of Translational Medicine*, 7(1), 17.
- Stevenson, M. M., Tam, M. F., Wolf, S. F., & Sher, A. (1995). IL-12-induced protection against blood-stage *Plasmodium chabaudi* AS requires IFN-gamma and TNF-alpha and occurs via a nitric oxide-dependent mechanism. *The Journal of Immunology*, 155(5), 2545-2556.
- Stevenson, M. M., & Riley, E. M. (2004). Innate immunity to malaria. *Nature Reviews Immunology*, 4(3), 169-180.
- Strickland, G. T. (1991). Malaria. In G. T. Strickland (Eds.), *Hunter's Tropical Medicine*, seventh edition. (pp. 586-617). Philadelphia: WB Saunders.
- Su, Z., & Stevenson, M. M. (2002). IL-12 is required for antibody-mediated protective immunity against blood-stage *Plasmodium chabaudi* AS malaria infection in mice. *The Journal of Immunology*, 168(3), 1348-1355.

- Sugaya, K., Fukagawa, T., Matsumoto, K., Mita, K., Takahashi, E., Ando, A., ... Ikemura, T. (1994). Three genes in the human MHC class III region near the junction with the class II: Gene for receptor of advanced glycosylation end products, PBX2 homeobox gene and a notch homolog, human counterpart of mouse mammary tumor gene int-3. *Genomics*, 23(2), 408-419.
- Suh, K. N., Kain, K. C., & Jay, S. (2004). Keystone. *Malaria: CMAJ*, 170(11), 1693-1702.
- Sullivan, A. D., Ittarat, I., & Meshnick, S. R. (1996). Patterns of haemozoin accumulation in tissue. *Parasitology*, 112(03), 285-294.
- Stüss, G., Eichmann, K., Kury, E., Linke, A., & Langhorne, J. (1988). Roles of CD4- and CD8-bearing T lymphocytes in the immune response to the erythrocytic stages of *Plasmodium chabaudi*. *Infection and Immunity*, 56(12), 3081-3088.
- Sutton, C. E., Mielke, L. A., & Mills, K. H. (2012). IL-17-producing $\gamma\delta$ T cells and innate lymphoid cells. *European Journal of Immunology*, 42(9), 2221-2231.
- Tanaka, N., Yonekura, H., Yamagishi, S., Fujimori, H., Yamamoto, Y., & Yamamoto, H. (2000). The receptor for advanced glycation end products is induced by the glycation products themselves and tumor necrosis factor-alpha through nuclear factor-kappa B, and by 17beta-estradiol through Sp-1 in human vascular endothelial cells. *Journal of Biological Chemistry*, 275, 25781-25790.
- Tangpukdee, N., Krudsood, S., Kano, S., & Wilairatana, P. (2012). Falciparum malaria parasitemia index for predicting severe malaria. *International Journal of Laboratory Hematology*, 34(3), 320-327.
- Tangteerawatana, P., Pichyangkul, S., Hayano, M., Kalambaheti, T., Looareesuwan, S., Troye-Bloomberg, M., & Khusmith, S. (2007). Relative levels of IL4 and IFN- γ in complicated malaria: Association with IL4 polymorphism and peripheral parasitemia. *Acta Tropica*, 101(3), 258-265.
- Taramelli, D., Basilico, N., De Palma, A. M., Saresella, M., Ferrante, P., Mussoni, L., & Olliaro, P. (1998). The effect of synthetic malaria pigment (β -haematin) on adhesion molecule expression and interleukin-6 production by human endothelial cells. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 92(1), 57-62.
- Taylor, W. R., Hanson, J., Turner, G. D., White, N. J., & Dondorp, A. M. (2012). Respiratory manifestations of malaria lung in malaria. *Chest Journal*, 142(2), 492-505.
- Taylor-Robinson, A. W., Phillips, R. S., Severn, A., Moncada, S., & Liew, F. Y. (1993). The role of TH1 and TH2 cells in a rodent malaria infection. *Science*, 260(5116), 1931-1934.
- Taylor-Robinson, A. W. (1995). Regulation of immunity to malaria: valuable lessons learned from murine models. *Parasitology Today*, 11(9), 334-342.

- Taylor-Robinson, A. W., & Looker, M. (1998). Sensitivity of malaria parasites to nitric oxide at low oxygen tensions. *The Lancet*, 351(9116), 1630.
- Tebo, A. E., Kremsner, P. G., & Luty, A. J. (2001). *Plasmodium falciparum*: a major role for IgG3 in antibody-dependent monocyte-mediated cellular inhibition of parasite growth in vitro. *Experimental Parasitology*, 98(1), 20-28.
- Tham, W. H., Wilson, D. W., Reiling, L., Chen, L., Beeson, J. G., & Cowman, A. F. (2009). Antibodies to reticulocyte binding protein-like homologue 4 inhibit invasion of *Plasmodium falciparum* into human erythrocytes. *Infection and Immunity*, 77(6), 2427-2435.
- Tian, J., Avalos, A. M., Mao, S., Chen, B., Senthil, K., Wu, H., ... Coyle, A. J. (2007). Toll-like receptor 9-dependent activation by DNA-containing immune complexes is mediated by HMGB1 and RAGE. *Nature Immunology*, 8(5), 487-496.
- Timms, R., Colegrave, N., Chan, B. H. K., & Read, A. F. (2001). The effect of parasite dose on disease severity in the rodent malaria *Plasmodium chabaudi*. *Parasitology*, 123(01), 1-11.
- Torre, D., Speranza, F., Giola, M., Matteelli, A., Tambini, R., & Biondi, G. (2002). Role of Th1 and Th2 cytokines in immune response to uncomplicated *Plasmodium falciparum* malaria. *Clinical and Diagnostic Laboratory Immunology*, 9(2), 348-351.
- Trampuz, A., Jereb, M., Muzlovic, I., & Prabhu, R. M. (2003). Clinical review: Severe malaria. *Critical Care*, 7(4), 315-323.
- Treutiger, C. J., Mullins, G. E., Johansson, A. S., Rouhiainen, A., Rauvala, H. M. E., Erlandsson-Harris, H., ... & Palmblad, J. E. W. (2003). High mobility group 1 B-box mediates activation of human endothelium. *Journal of Internal Medicine*, 254(4), 375-385.
- Trinchieri, G. (1998). Interleukin-12: a cytokine at the interface of inflammation and immunity. *Advances in Immunology*, 70, 83-243.
- Tripathi, A. K., Sullivan, D. J., & Stins, M. F. (2006). Plasmodium falciparum-infected erythrocytes increase intercellular adhesion molecule 1 expression on brain endothelium through NF- κ B. *Infection and Immunity*, 74(6), 3262-3270.
- Tripp, C. S., Wolf, S. F., & Unanue, E. R. (1993). Interleukin 12 and tumor necrosis factor alpha are costimulators of interferon gamma production by natural killer cells in severe combined immunodeficiency mice with listeriosis, and interleukin 10 is a physiologic antagonist. *Proceedings of the National Academy of Sciences*, 90(8), 3725-3729.
- Troye-Blomberg, M., Riley, E. M., Kabilan, L., Holmberg, M., Perlmann, H., Andersson, U., ... & Perlmann, P. (1990). Production by activated human T cells of interleukin 4 but not interferon-gamma is associated with elevated

- levels of serum antibodies to activating malaria antigens. *Proceedings of the National Academy of Sciences*, 87(14), 5484-5488.
- Tsunawaki, S., Sporn, M., Ding, A., & Nathan, C. (1988). Deactivation of macrophages by transforming growth factor- β . *Nature*, 334(6179), 260-262.
- Turner, G. D., Morrison, H., Jones, M., Davis, T. M., Looareesuwan, S., Buley, I. D., ... & Berendt, A. R. (1994). An immunohistochemical study of the pathology of fatal malaria: evidence for widespread endothelial activation and a potential role for intercellular adhesion molecule-1 in cerebral sequestration. *The American Journal of Pathology*, 145(5), 1057-1069.
- Urquhart, A. D. (1994). Putative pathophysiological interactions of cytokines and phagocytic cells in severe human falciparum malaria. *Clinical Infectious Diseases*, 19(1), 117-131.
- Van den Steen, P. E., Geurts, N., Deroost, K., Van Aelst, I., Verhenne, S., Heremans, H., ... & Opdenakker, G. (2010). Immunopathology and dexamethasone therapy in a new model for malaria-associated acute respiratory distress syndrome. *American Journal of Respiratory and Critical Care Medicine*, 181(9), 957-968.
- Van der Heyde, H. C., Nolan, J., Combes, V., Gramaglia, I., & Grau, G. E. (2006). A unified hypothesis for the genesis of cerebral malaria: sequestration, inflammation and hemostasis leading to microcirculatory dysfunction. *Trends in Parasitology*, 22(11), 503-508.
- Van Snick, J. (1990). Interleukin-6: an overview. *Annual Review of Immunology*, 8(1), 253-278.
- Van Vugt, M., Van Beest, A., Sicuri, E., Van Tulder, M., & Grobusch, M. P. (2011). Malaria treatment and prophylaxis in endemic and nonendemic countries: evidence on strategies and their cost-effectiveness. *Future Microbiology*, 6(12), 1485-1500.
- Van Zoelen, M. A., Schouten, M., de Vos, A. F., Florquin, S., Meijers, J. C., Nawroth, P. P., ... van der Poll, T. (2009). The receptor for advanced glycation end products impairs host defense in pneumococcal pneumonia. *Journal of Immunology*, 182(7), 4349-4356.
- Vinetz, J. M., Kumar, S., Good, M. F., Fowlkes, B. J., Berzofsky, J. A., & Miller, L. H. (1990). Adoptive transfer of CD8+ T cells from immune animals does not transfer immunity to blood stage *Plasmodium yoelii* malaria. *The Journal of Immunology*, 144(3), 1069-1074.
- Viriyavejakul, P., Khachonsaksumet, V., & Punsawad, C. (2014). Liver changes in severe *Plasmodium falciparum* malaria: histopathology, apoptosis and nuclear factor kappa B expression. *Malaria Journal*, 13(1), 106.
- Visser, B. J., van Vugt, M., & Grobusch, M. P. (2014). Malaria: an update on current chemotherapy. *Expert Opinion on Pharmacotherapy*, 15(15), 2219-2254.

- Vogetseder, A., Ospelt, C., Reindl, M., Schober, M., & Schmutzhard, E. (2004). Time course of coagulation parameters, cytokines and adhesion molecules in *Plasmodium falciparum* malaria. *Tropical Medicine & International Health*, 9(7), 767-773.
- Walther, M., Tongren, J. E., Andrews, L., Korbel, D., King, E., Fletcher, H., ... & Hill, A. V. (2005). Upregulation of TGF- β , FOXP3, and CD4+ CD25+ regulatory T cells correlates with more rapid parasite growth in human malaria infection. *Immunity*, 23(3), 287-296.
- Walther, M., Jeffries, D., Finney, O. C., Njie, M., Ebonyi, A., Deininger, S., ... & Riley, E. M. (2009). Distinct roles for FOXP3+ and FOXP3- CD4+ T cells in regulating cellular immunity to uncomplicated and severe *Plasmodium falciparum* malaria. *PLoS Pathogens*, 5(4), e1000364.
- Wang, H., Bloom, O., Zhang, M., Vishnubhakat, J. M., Ombrellino, M., Che, J., ... & Tracey, K. J. (1999). HMG-1 as a late mediator of endotoxin lethality in mice. *Science*, 285(5425), 248-251.
- Wautier, M. P., Chappey, O., Corda, S., Stern, D. M., Schmidt, A. M., & Wautier, J. L. (2001). Activation of NADPH oxidase by AGE links oxidant stress to altered gene expression via RAGE. *American Journal of Physiology-Endocrinology And Metabolism*, 280(5), E685-E694.
- Wells, S., Diap, G., & Kiechel, J. R. (2013). The story of artesunate-mefloquine (ASMQ), innovative partnerships in drug development: case study. *Malaria Journal*, 12, 68.
- Wendt, T. M., Tanji, N., Guo, J., Kislinger, T. R., Qu, W., Lu, Y., ... & Schmidt, A. M. (2003). RAGE drives the development of glomerulosclerosis and implicates podocyte activation in the pathogenesis of diabetic nephropathy. *The American Journal of Pathology*, 162(4), 1123-1137.
- Wenisch, C., Linnau, K. F., Looaresuwan, S., & Rumpold, H. (1999). Plasma levels of the interleukin-6 cytokine family in persons with severe *Plasmodium falciparum* malaria. *Journal of Infectious Diseases*, 179(3), 747-750.
- White, N. J., & Ho, M. (1992). The pathophysiology of malaria. *Advances in Parasitology*, 31, 83-173.
- White, N. J. (2011). Determinants of relapse periodicity in *Plasmodium vivax* malaria. *Malaria journal*, 10(1), 297.
- Willems, F., Marchant, A., Delville, J. P., Gérard, C., Delvaux, A., Velu, T., ... & Goldman, M. (1994). Interleukin-10 inhibits B7 and intercellular adhesion molecule-1 expression on human monocytes. *European Journal of Immunology*, 24(4), 1007-1009.

- Wilson, E. H., Wille-Reece, U., Dzierszinski, F., & Hunter, C. A. (2005). A critical role for IL-10 in limiting inflammation during toxoplasmic encephalitis. *Journal of Neuroimmunology*, 165(1), 63-74.
- Winkler, S., Willheim, M., Baier, K., Schmid, D., Aichelburg, A., Graninger, W., & Kremsner, P. G. (1998). Reciprocal regulation of Th1-and Th2-cytokine-producing T cells during clearance of parasitemia in *Plasmodium falciparum* malaria. *Infection and Immunity*, 66(12), 6040-6044.
- World Health Organization. (2000). Severe falciparum malaria. *Transactions of the Royal Society of Tropical Medicine and Hygiene*, 94, 1-90.
- World Health Organization. (2003, December). Fixed-dose combinations for HIV/AIDS, tuberculosis and malaria. In *Report of a meeting held* (pp. 16-18).
- World Health Organization (Ed.). (2006). *Guidelines for the treatment of malaria*. Geneva: World Health Organization.
- World Health Organization. (2012). World malaria report 2012 fact sheet. Retrieved December 17, 2012, from http://www.who.int/malaria/publications/world_malaria_report_2012/wmr2012_factsheet.pdf?ua=1
- Wu, Y., Wang, Q. H., Zheng, L., Feng, H., Liu, J., Ma, S. H., & Cao, Y. M. (2007). *Plasmodium yoelii*: distinct CD4+ CD25+ regulatory T cell responses during the early stages of infection in susceptible and resistant mice. *Experimental Parasitology*, 115(3), 301-304.
- Yamamoto, Y., Kato, I., Doi, T., Yonekura, H., Ohashi, S., Takeuchi, M., ... & Yamamoto, H. (2001). Development and prevention of advanced diabetic nephropathy in RAGE-overexpressing mice. *Journal of Clinical Investigation*, 108(2), 261-268.
- Yan, S. D., Zhu, H., Fu, J., Yan, S. F., Roher, A., Tourtellotte, W. W., ... Schmidt, A. M. (1997). Amyloid-beta peptide-Receptor for Advanced Glycation Endproduct interaction elicits neuronal expression of macrophage-colony stimulating factor: A proinflammatory pathway in Alzheimer disease. *Proceedings of the National Academy of Sciences*, 94(10), 5296-5301.
- Yan, S. S., Wu, Z. Y., Zhang, H. P., Furtado, G., Chen, X., Yan, S. F., ... & Jiang, H. (2003). Suppression of experimental autoimmune encephalomyelitis by selective blockade of encephalitogenic T-cell infiltration of the central nervous system. *Nature Medicine*, 9(3), 287-293.
- Yan, S. D., Chen, X., Walker, D. G., Schmidt, A. M., Arancio, O., & Lue, L. F. (2007). RAGE: A potential target for A β -mediated cellular perturbation in Alzheimer's disease. *Current Molecular Medicine*, 7(8), 735-742.
- Yang, H., Ochani, M., Li, J., Qiang, X., Tanovic, M., Harris, H. E., ... & Tracey, K. J. (2004). Reversing established sepsis with antagonists of endogenous high-mobility group box 1. *Proceedings of the National Academy of Sciences*, 101(1), 296-301.

- Yonekura, H., Yamamoto, Y., Sakurai, S., Petrova, R. G., Li, H., Yasui, K., ... Yamamoto, H. (2003). Novel splice variants of the receptor for advanced glycation end-products expressed in human vascular endothelial cells and pericytes, and their putative roles in diabetes-induced vascular injury. *Biochemical Journal*, 370(3), 1097–1109.
- Yoneto, T., Yoshimoto, T., Wang, C. R., Takahama, Y., Tsuji, M., Waki, S., & Nariuchi, H. (1999). Gamma interferon production is critical for protective immunity to infection with blood-stage *Plasmodium berghei* XAT but neither NO production nor NK cell activation is critical. *Infection and Immunity*, 67(5), 2349-2356.
- Yoshimoto, T., Takeda, K., Tanaka, T., Ohkusu, K., Kashiwamura, S., Okamura, H., Akira, S., & Nakanishi, K. (1998). IL-12 up-regulates IL-18 receptor gene expression on T cells, Th1 cells and B cells: synergism with IL-18 for IFN gamma production. *The Journal of Immunology*, 161(7), 3400-3407.
- Zeh III, H. J., & Lotze, M. T. (2005). Addicted to death: invasive cancer and the immune response to unscheduled cell death. *Journal of Immunotherapy*, 28(1), 1-9.
- Zhang, L. Y. and Wang, C. X. (1984). Histopathological and histochemical studies on toxic effect of brodifacoum in mouse liver. *Acta Academiae Medicinae Sinicae*, 6(5), 386-388.
- Zhang, L., Bukulin, M., Kojro, E., Roth, A., Metz, V. V., Fahrenholz, F., ... Postina, R. (2008). Receptor for advanced glycation end products is subjected to protein ectodomain shedding by metalloproteinases. *Journal of Biological Chemistry*, 283(51), 35507-35516.
- Zhu, J., Yamane, H., Cote-Sierra, J., Guo, L., & Paul, W. E. (2006). GATA-3 promotes Th2 responses through three different mechanisms: induction of Th2 cytokine production, selective growth of Th2 cells and inhibition of Th1 cell-specific factors. *Cell Research*, 16(1), 3-10.
- Zuzarte-Luis, V., Mota, M. M., & Vig ário, A. M. (2014). Malaria infections: What and how can mice teach us. *Journal of Immunological Methods*, 410, 113-122.