

UNIVERSITI PUTRA MALAYSIA

EXTRACTION AND IDENTIFICATION OF DI(2-ETHYLHEXYL) PHTHALATE FROM SAFED MUSLI (CHLOROPHYTUM BORIVILIANUM L.)

CHUA BEE LIN

FK 2015 73



EXTRACTION AND IDENTIFICATION OF DI(2-ETHYLHEXYL) PHTHALATE FROM SAFED MUSLI (*CHLOROPHYTUM BORIVILIANUM* L.)



By

CHUA BEE LIN

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of Philosophy

October 2015



All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia





Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

EXTRACTION AND IDENTIFICATION OF DI(2-ETHYLHEXYL) PHTHALATE FROM SAFED MUSLI (CHLOROPHYTUM BORIVILIANUM L.)

By

CHUA BEE LIN

October 2015

Chair: Professor Luqman Chuah Abdullah, PhD Faculty: Engineering

Chlorophytum borivilianum (safed musli) is a medicinally important plant. Its roots are being employed in folk medicine. Presently, the crude extract of C. borivilianum has been consumed for the treatment such as anti-diabetic, anti-aging, anti-oxidant, anti-ulcer and anti-inflammatory. Studies have been carried out to further confirm these remarkable bioactivities of C. borivilianum. So far, the isolated chemical constituents are mainly saponins. A fructo-oligosaccharide, three fatty acids, one sterol stigmasterol, hecogenin also have been reported to be isolated from the roots of C. borivilianum. In this research, di(2-ethylhexyl) phthalate (DEHP) was extracted from the aqueous extract of the roots of C. borivilianum. The yield of DEHP was found to be 33.70 mg, which was equivalent to 0.013% with reference to the total weight of root powder (250 g). The structure of DEHP was elucidated based on the spectral data of ¹H-NMR, ¹³C-NMR, distortionless enhancement by polarization transfer (DEPT), correlation spectroscopy (COSY), heteronuclear multiple bond correlation (HMBC) and heteronuclear multiple quantum correlation (HMQC) and also based on the comparison with the previous literature data. This is the first report so far of occurrence and detail spectroscopic description of DEHP from C. borivilianum. Single experimental design and response surface methodology (RSM) was implemented to optimize the extraction conditions for obtaining the maximum yield of DEHP from the roots of C. borivilianum. DEHP was optimized because it could be a starting point to pave a way to isolate and quantify other pure compounds (minor or rare) from this herbaceous plant in order to use it as a tool for quality control and also for the future development of other therapeutic applications. Furthermore, DEHP was reported to possess some remarkable biological activities such as anti-leukemic, anti-microbial, anti-fungal, anti-tumour and antiviral activity against H1N1 disease. In this study, ultrasound-assisted extraction was applied for the effective extraction of DEHP and DEHP was quantified by high performance liquid chromatography (HPLC) analysis. Herein, three independent variables (extraction time, solid to solvent ratio and extraction temperature) with a five level design were evaluated



using the central composite design (CCD), with the yield of DEHP as the response variable. Second-order polynomial model was found to be satisfactory in describing the experimental data for the total DEHP content. The analysis of variance (ANOVA) indicated that the main effect of solid to solvent ratio and the extraction temperature as well as the quadratic effects of all independent variables had significant effect ("Prob>F"<0.05) on the extraction yield of DEHP. The optimal extraction conditions were established as follow: extraction time of 92 min, solvent to solvent ratio of 1:38 (g/mL) and extraction temperature of 51°C. Using these adjusted optimal conditions, the predicted yield of DEHP by model was 0.44 ppm whereas the actual yield of DEHP was 0.43 ± 0.01 ppm which was in close conformity with the predicted values as the relative error was just 2.72%. The extraction kinetic was studied using equilibrium extraction model (EEM) and diffusion extraction model (DEM). The kinetics results revealed EEM model was more suitable in describing the extraction process and the ethanol extraction of the roots of C. borivilianum achieved equilibrium within 70 minutes. Lastly, DEHP extracted from the roots C. borivilianum was tested for its anti-inflammatory activity. However, DEHP had shown a low inhibition effect on the anti-inflammatory activity in HYA assay with a percentage of inhibition of $4.02 \pm 1.17\%$. This study implied that DEHP was not active to inhibit the hyaluronidase enzyme and this indicated that the other compounds in the roots of C. borivilianum might contribute to the antiinflammatory activity as the previous researchers had obtained significant antiinflammatory activity from this plant.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

PENGEKSTRAKAN DAN PENGENALPASTIAN DI(2-ETHYLHEXYL) PHTHALATE DARI SAFED MUSLI (*CHLOROPHYTUM BORIVILIANUM* L.)

Oleh

CHUA BEE LIN

Oktober 2015

Pengerusi: Profesor Luqman Chuah Abdullah, PhD Fakulti: Kejuruteraan

Chlorophytum borivilianum (safed musli) merupakan tumbuhan ubatan yang bernilai. Akarnya telah digunakan dalam perubatan rakyat. Baru-baru ini, ekstrak mentah dari C. borivilianum telah digunakan untuk rawatan seperti penyakit kencing manis, antipenuaan, antioksidan, anti-ulser and anti-radang. Kajian telah dijalani untuk mengesahkan bioaktiviti C. borivilianum. Setakat ini, konstituen kimia yang diasingkan kebanyakannya merupakan saponins. Satu frukto-oligosakarida, tiga asid lemak, satu stigmasterol sterol dan hekogenin telah dilaporkan diasingkan dari akar C. borivil ianum. Dalam penyelidikan ini, di(2-ethylhexyl) phthalate (DEHP) telah diekstrak dari ekstrak akueus akar C. borivilianum.Hasil DEHP adalah 33.70 mg, bersamaan 0.013% merujuk kepada jumlah berat serbuk akar (250 g). Struktur DEHP telah dihuraikan berdasarkan data spektrum seperti ¹H-NMR, ¹³C-NMR, DEPT, COSY, HMBC dan HMQC dan juga berdasarkan kepada perbandingan dengan data kajian lepas. Ini merupakan laporan pertama setakat ini tentang kehadiran dan huraian spektroskopi yang perinci bagi DEHP yang diasingkan dari C. borivilianum. Reka bentuk eksperimen tunggal dan kaedah respons permukaan (RSM) telah digunakan bagi mengoptimumkan syarat-syarat pengekstrakan untuk mendapatkan kadar hasil maksimum DEHP dari akar C. borivilianum. DEHP telah dioptimumkan kerana ia boleh digunakan sebagai titik permulaan bagi mengekstrak and mengenalpasti kandungan kompaun-kompaun yang lain (sikit ataupun jarang) dari tumbuhan herba ini supaya kualiti dapat dikawal dalam aplikasi terapeutik pada masa depan. Tambahan pula, DEHP dilaporkan mempunyai beberapa aktiviti biologi seperti anti-leukemia, anti-mikrob, anti-kulat, anti-tumor dan antivirus terhadap H1N1. Dalam kajian ini, pengekstrakan ultrasonik telah dijalani untuk pengekstrakan DEHP secara berkesan dan DEHP telah dikuantitikan oleh analisis kromatografi cecair berprestasi tinggi (HPLC). Dengan ini, tiga pembolehubah bebas (masa pengekstrakan, nisbah pepejal kepada pelarut dan suhu pengekstrakan) dengan reka bentuk berperingkat lima telah dinilai dengan menggunakan reka bentuk komposit tengah dan hasil DEHP telah dipilih sebagai respons pembolehubah. Model polinomial berkuadratik telah dihasilkan dan didapati memuaskan dalam menghuraikan data eksperimen untuk jumlah kandungan DEHP. Analisis varians (ANOVA) menunjukkan bahawa nisbah pepejal kepada pelarut dan suhu pengekstrakan serta kesan-kesan kuadratik semua pembolehubah bebas mempunyai kesan yang ketara (p<0.05) terhadap hasil pengekstrakan DEHP. Syarat-syarat pengekstrakan yang optimum telah ditetapkan



seperti yang berikut: masa pengekstrakan 92 min, nisbah pepejal kepada pelarut 1:38 (g/mL) dan suhu pengekstrakan 51°C. Dengan meggunakan syarat-syarat optimum yang terlaras, hasil DEHP yang diramalkan oleh model ialah 0.44 ppm manakala hasil sebenar DEHP ialah 0.43±0.01 ppm dan nilai sebenar hampir selaras dengan nilai ramalan kerana ralat relatif hanya 2.72%. Keputusan telah menunjukkan bahawa RSM boleh digunakan bagi mengoptimumkan syarat-syarat pengekstrakan DEHP dari akar *C. borivilianum*. Tambahan pula, kinetik DEHP telah dikaji dan dua model (EEM dan DEM) telah dihasilkan. EEM lebih sesuai bagi menghuraikan proses pengekstrakan dan pengekstrakan etanol akar *C. borivilianum* mencapai keseimbangan dalam 70 minit. Akhir sekali, activiti anti-radang HYA DEHP telah dikaji dan DEHP telah menunjukkan kesan yang tidak ketara dengan hanya 4.02±1.17%. Kajian ini mencadangkan bahawa DEHP tidak aktif terhadap antiviti ini dan mungkin adalah kompaun-kompaun lain yang menyumbang kepada activiti anti-radang. Ini adalan kerana aktiviti anti-radang yang positif telah dilaporkan oleh pengkaji-pengkaji yang sebelumnya.

ACKNOWLEDGEMENTS

First of all, I would like to express my deep sense of gratitude towards my supervisor, Professor Dr. Luqman Chuah Abdullah from the Department of Chemical and Environmental Engineering, Universiti Putra Malaysia for his invaluable guidance, advice, encouragement, inspiration and support during the course of present research and writing of this thesis.

I would like to extend my deepest appreciation to my supervisory committee members, Professor Dr. Thomas Choong Shean Yaw from the Department of Chemical and Environmental Engineering, Professor Umi Kalsom Yusof from the Department of Biology, Dr. Pin Kar Yong and Dr. Zunoliza Abdullah from Forest Research Institute Malaysia (FRIM) who had offered a great meaningful help by providing me some useful ideas, information and lessons throughout the project.

My thanks are also to my senior Nor Fariza Ismail from Universiti of Putra Malaysia, Tn. Haji Abdull Rashih Ahmad and Mr. Mohd Farhan Abdul Razak from Forest Research Institute Malaysia (FRIM) for their technical assistance and kind knowledge sharing.

Furthermore, I am also thankful to my little son, Zyi Hao who has endured very much while I was busy for laboratory works and writing my thesis. I wish to take this opportunity to thank my beloved family for their love, encouragement and support. Finally, special thanks to my husband, Mr. Peng Sher Seong for his love and endless support throughout the course of my studies.

Lastly, I wish to thank all my friends who have been directly or indirectly helped me throughout the course of this project.

I certify that a Thesis Examination Committee has met on (date of viva voce) to conduct the final examination of (student's name)on his (her) thesis entitled ("Title of Thesis")in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the (insert the name of relevant degree).

Members of the Thesis Examination Committee were as follows:

Mohd. Halim Shah bin Ismail, PhD

Associate Professor Faculty of Engineering Universiti Putra Malaysia (Chairperson)

Norhafizah Abdullah, PhD

Associate Professor Faculty of Engineering Universiti Putra Malaysia (Internal Examiner)

Yoshida Hiroyuki, PhD

Professor Faculty of Engineering Universiti Putra Malaysia (Internal Examiner)

Chang Seo Park, PhD

Associate Professor Department of Chemical and Biochemical Engineering College of Engineering Dongguk University Korea (External Examiner)



ZULKARNAIN ZAINAL, PhD Professor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date: 16 February 2016

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Luqman Chuah Abdullah, PhD

Professor Faculty of Engineering Universiti Putra Malaysia (Chairperson)

Thomas Choong Shean Yaw, PhD

Professor, Ir Faculty of Engineering Universiti Putra Malaysia (Member)

Umi Kalsom Yusuf, PhD

Professor Faculty of Science Universiti Putra Malaysia (Member)

Pin Kar Yong, PhD

Research Officer Herbal Technology Center Forest Research Institute Malaysia (Member)

Zunoliza Abdullah, PhD

Research Officer Programme Phytochemistry Forest Research Institute Malaysia (Member)

BUJANG KIM HUAT, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in theUniversiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature:	Date:	
Name and Matric No.:	O	

Declaration by Members of Supervisory Committee

This is to confirm that:

C

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013)are adhered to.

Signature: Name of Chairman of Supervisory Committee:	Prof. Dr. Luqman Chuah Abdullah
Signature: Name of Member of Supervisory Committee:	Prof. Dr. Thomas Choong Shean Yaw
Signature: Name of Member of Supervisory Committee:	Prof. Dr. Umi Kalsom Yusuf
Signature: Name of Member of Supervisory Committee:	Dr. Pin Kar Yong
Signature: Name of Member of Supervisory Committee:	Dr. Zunoliza Abdullah

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xv
LIST OF FIGURES	xvi
LIST OF ABBREVIATIONS	xviii

CHAPTER

1	INTR	ODUCTION	1
	1.1	Introduction to Medicinal Plants	1
	1.2	Background of Study	1
	1.3	Problems statement	2
	1.4	Objectives of the study	2
	1.5	Scopes of Study	3
2	LITE	RATURE REVIEW	4
	2.1	Introduction	4
	2.2	Occurrence and Distribution of <i>C. Borivilianum</i>	4
	2.3	Botanical Description of C. Borivilianum	5
	2.4	Traditional Mode of Application of <i>C. Borivilianum</i>	5
	2.5	Pharmacological Activities of C. Borivilianum	5
		2.5.1 Antioxidant Activity	6
		2.5.2 Immunomodulatory Activity	6
		2.5.3 Anti-Diabetic Activity	7
		2.5.4 Anti-Ulcer Activity	7
		2.5.5 Analgesic Activity	7
		2.5.6 Anti-Microbial Activity	7
		2.5.7 Anti-Stress Activity	8
		2.5.8 Anti-Arthritic Activity	9
		2.5.9 Hepatoprotective Activity	9
		2.5.10 Anthelmintic Activity	9
		2.5.11 Larvicidal Activity	9
		2.5.12 Anti-Inflammatory Activity	9
		2.5.13 Anti-Cancer Activity	10
		2.5.14 Aphrodisiac Activity	10
	2.6	Isolation and Purification of C. Borivilianum	10
		2.6.1 Extraction	11
	2.7	Chromatographic Methods	11
		2.7.1 Thin Layer Chromatography (TLC)	12
		2.7.2 Column Chromatography (CC)	13
	2.8	Physical Chemistry Methods for Structural Elucidation	13
		2.8.1 Nuclear magnetic resonance spectroscopy	13
		(NMR)	
	2.9	Isolated Chemical Constituents of C. Borivilianum	14

2.10	Respon	se Surface Methodology (RSM)	21
	2.10.1	Applications of RSM in the Optimization of	22
		Previous Studies	
	2.10.2	Design of Experiment of RSM	23
	2.10.3	Effect of the Extraction Parameters on the	24
		Extraction Yield	
2.11	Backgro	ound of Di(2-Ethylhexyl) Phthalate (DEHP)	25
МАТ	ERIALS	AND METHODS / METHODOLOGY	27
3.1	Introdu	ction	27
3.2	Extracti	ion, Partition, Fractionation and Isolation of the	28
	Roots o	f C.Borivilianum	
	3.2.1	Plant Collection	28
	3.2.2	Plant Material Preparation and Pretreatment	29
	3.2.3	Chemicals and Reagents	29
	3.2.4	Solid-Liquid Extraction of the Roots of <i>C</i> . <i>Borivilianum</i>	29
	3.2.5	Liquid-Liquid Partition of the Crude Extract	29
	3.2.6	Sample Fractionation and Isolation of	30
		Compound 1	
	3.2.7	Thin Layer Chromatography (TLC) Analysis	30
	3.2.8	Column Chromatography Analysis	30
3.3	Scheme	of Extraction, Partition, Fractionation and	31
	Isolatio	n of Compound 1 from the Roots of C.	
	borivili	anum	
3.4	Structur	re Elucidation of the Compound 1 from the	32
	Roots o	f C. Borivilianum	
3.5	Quantif	ication of the Crude Extract of the Roots of C.	33
	Borivili	anum	
3.6	Quantit	ative Determination of Compound 1 from the	33
	Roots o	f C. Borivilianum	
	3.6.1	Chemicals	33
	3.6.2	Preparation of Stock Solution of Standard Compound 1	34
	3.6.3	High Performance Liquid Chromatography	34
		(HPLC) Analysis	
	3.6.4	Instrumentation and Chromatographic Conditions	34
	3.6.5	Calibration Curve of Standard Compound 1	35
		Isolated from the Roots of C. Borivilianum	
	3.6.6	Quantification of Compound 1 in the Crude Extract of the Roots of <i>C. Borivilianum</i>	35
3.7	Optimiz	zation of Solvents for Effective Extraction of the	36
	Crude E	Extract and Isolation of compound 1 from the	
	Roots o	of C. Borivilianum	
	3.7.1	Preparation of Plant Material and Chemicals	36
	3.7.2	Ultrasound-Assisted Extraction of Compound	36
		1 from the Roots of C. Borivilianum	
	3.7.3	Determination of Total Yield of Compound 1	36
	3.7.4	Experimental Design for the Optimizations of	37
		Solvents	

xi

3.8	Single I	Factor Experimental Design	38
	3.8.1	Preparation of Plant Material and Chemicals	38
	3.8.2	Ultrasound-Assisted Extraction (UAE)	39
	3.8.3	Effect of Extraction Time on Extraction Yield	39
		of Compound 1	
	3.8.4	Effect of Solid to Solvent Ratio on Extraction	39
		Yield of Compound 1	
	3.8.5	Effect of Extraction Temperature on Extraction	39
		Yield of Compound 1	
	3.8.6	Experimental Range of the Single Factor	39
		Experimental Design	
	3.8.7	Single Factor Experimental Design of	40
		Compound 1	
3.9	Optimiz	zation of Response Surface Methodology (RSM)	40
	3.9.1	Extraction Procedure	41
	3.9.2	The Experimental Range of the Central	41
		Composite Design (CCD)	
	3.9.3	Experimental Design of Central Composite	41
		Design (CCD)	
	3.9.4	Analysis of Variance (ANOVA) for the	43
		Response Surface Quadratic Polynomial	
		Model for the Extraction of Compound 1	
	3.9.5	Fitting the Polynomial Models for the	43
		Extraction Yield of Compound 1	
	3.9. <mark>6</mark>	Analysis of Variance for the Fitted Quadratic	44
		Models for the Extraction Yield of Compound	
		1	
	3.9. <mark>7</mark>	Linear Correlation Plot Between the Predicted	44
		and the Actual Values of Extraction Yield	
		Compound 1	
	3.9.8	Effect of Interactive Extraction Parameters	44
3.10	Optimiz	zation Procedure of the Extraction Yield of	45
	Compo	und 1	
3.11	Validati	ion of the Model	46
3.12	Mathen	natical Models for Solid-Liquid Extraction	46
3.13	Mathem	natical Modeling of Solid-Liquid Extraction	46
	(SLE) 0	of C. Borivilianum Roots	10
	3.13.1	Physical Properties of Dried C. Borivilianum	46
	0.10.0	Roots	47
	3.13.2	Mathematical Modeling	47
2.1.4	3.13.3	Fitting the Extraction Models	50
3.14	In-Vitro	Anti-Inflammatory Assay of DEHP from the	51
	Roots o	f C. Borivilianum	7 1
	3.14.1	Chemicals and Reagents	51
	2140	Test Seconds Dresservices	51
	3.14.2	Test Sample Preparation	51
	31/2	Hyaluronidase (HVA) Assay	51
3 1 5	Summa	ry of the Experimental Procedure	52
5.15	Summa		52
RESU	LTS AN	ID DISCUSSION	54

4

RESULTS AND DISCUSSION

xii

4.1	Extracti	on, Partition, Fractionation and Isolation of the	54
	Roots of	C.Borivilianum	
	4.1.1	Solid-Liquid Extraction of the Roots of <i>C</i> . <i>Borivilianum</i>	54
	4.1.2	Liquid-Liquid Partition of the Crude Extract	54
	4.1.3	Sample Fractionation and Isolation of Compound 1	54
	414	Thin Laver Chromatography (TLC) Analysis	54
	415	Liquid-Liquid Partition of the Crude Extract	55
4.2	Scheme	of Extraction Yield of each chromatography	55
	Steps fro	om the Roots of <i>C. Borivilianum</i>	
4.3	Structure	Elucidation of the Isolated Compound from	56
the Roots of <i>C.Borivilianum</i>		s of <i>C.Borivilianum</i>	
	4.3.1	¹ H-NMR Spectrum (CD ₃ OD, 500 MHz) of	57
	4.3.2	Compound 1 ¹³ C-NMR Spectrum (CD ₃ OD, 126 MHz) and	58
		DEPT Experiments of compound 1	
	4.3.3	HMQC and HMBC Experiments of	59
		Compound 1	
	4.3.4	Structure Elucidation of Compound 1 by	59
		Means of 1D and 2D-NMR Experiments	
4.4	Quantita	tive Determination of DEHP from the Roots of	62
	C. Boriv	ilianum by HPLC Analysis	
	4.4.1	Calibration Curve of Standard DEHP Isolated	64
	4.4.2	Quantification of the DEHP from the Roots of	65
		C. Borivilianum	
4.5	Optimiz from the	ation of Solvents for Effective Extraction DEHP Roots of <i>C. Borivilianum</i>	65
4.6	Single F	Factor Experimental Design	67
	4.6.1	Effect of Extraction Time on Extraction Yield	67
		of DEHP	
	4.6.2	Effect of Solid to Solvent Ratio on Extraction	69
		Yield of DEHP	
	4.6.3	Effect of Extraction Temperature on	70
		Extraction Yield of DEHP	
4.7	Optimal	Experimental Range of the Extraction	71
	Paramete	ers	
4.8	Optimiza	ation of Response Surface Methodology (RSM)	71
	4.8.1	Central Composite Design (CCD) of the	72
		Extraction Yield of DEHP	
	4.8.2	Analysis of Variance (ANOVA) for the	73
		Quadratic Polynomial Model	
4.9	Fitting th	ne Polynomial Models	74
	4.9.1	Analysis of Variance for the Fitted Quadratic Model	75
4.10	Linear C	orrelation plot Between the Predicted and the	76
	Actual V	aluesof Extraction Yield	
4.11	Effect of	Interactive Extraction Parameters	77
	4.11.1	Effect of Extraction Time and Solid to Solvent	77

G

		4.11.2	Ratio Upon the Extraction Yield of DEHP Effect of Extraction Time and Extraction Temperature Upon the Extraction Yield of DEHP	78
		4.11.3	Effect of Solid to Solvent Ratio and Extraction Temperature Upon the Extraction Yield of DEHP	79
	4.12	Optimiza DEHP	ation Procedure of the Extraction Yield of	80
	4.13	Validatio	on of the Model	81
	4.14	Fitting th	e Extraction Models of C. Borivilianum	81
		4.14.1	Fitting the Equilibrium Extraction Model (EEM) of <i>C. Borivilianum</i>	82
		4.14.2	Fitting the Diffusion Extraction Model (DEM) of <i>C. Borivilianum</i>	83
		4.14.3	Selection of Extraction Model	84
	4.15	In-Vitro	Anti-Inflammatory Assay of DEHP from the	85
		Roots of	C. Borivilianum	
5	CON	CLUSION		86
-	5.1	Summar	v	86
	5.2	Conclusi	on	87
	5.3	Future S	tudies	87
REFERENC APPENDIC	CES ES			89 98
BIODATA (JF STU	DENT		104
LIST OF PU	BLICA	ATIONS		105

G

LIST OF TABLES

Table		Page
3.1	Experimental design of the selection of solvent for the extraction yield of compound 1	37
3.2	Experimental range of the single factor experimental design	39
3.3	Single factor experimental design of main extraction parameters.	40
3.4	Level of the extraction parameters for the extraction of compound 1 through a central composite design.	42
3.5	The central composite design for the extraction yield of compound 1	43
3.6	Level of the extraction parameters for the constructing the contour plots of extraction of compound 1	45
4.1	¹ H-NMR spectrum data of compound 1 (recorded at 500 MHz in CD ₃ OD); δ in ppm, J in Hz)	57
4.2	13 C-NMR spectrum data of compound 1 (recorded at 126 MHz in CD ₃ OD); δ in ppm	58
4.3	Optimal experimental range of the extraction parameters based on the single factor experimental design results	72
4.4	Experimental results of the central composite design (CCD) of the extraction yield of DEHP	72
4.5	Analysis of variance (ANOVA) for the response surface quadratic polynomial model for the extraction yield of DEHP	74
4.6	Analysis of variance for the fitted quadratic model for the extraction yield of DEHP	76
4.7	Optimum conditions and modified conditions of responses Y_1	81
4.8	The results from EEM model for different extraction temperatures	83
4.9	The results from DEM model for different extraction temperatures	84

 \bigcirc

LIST OF FIGURES

Figure		Page
1.1	Plant and Roots of Chlorophytum Borivilianum (Safed	1
	Musli)	
2.1	Structure of the Fructo-Oligosaccharide	14
2.2	Structure of Isolated Fatty Acids	15
2.3	3β-Hydroxy-24-ethyl-5,22-cholestadiene/Stigmasterol	15
2.4	$(3\beta, 5)$ 22R, 23R)-26- $(\beta$ -d-glucopyranosyloxy)-22-	15
	hydroxyfurostan-12-one-3yl O - β -d-galactopyranosyl- $(1 \rightarrow 4)$	
	glucopyranoside / Chlorophytoside-I	
2.5	5-Spirostan-3β-ol-12-one/ Hacogenin	16
2.6	Structure and Name of the Isolated Furostanol and Spirostanol	16
	Steroidal Saponins (Acharya et al., 2008b)	
2.7	Structure and Name of the Isolated Compounds (Acharya et	18
	al., 2008a)	
2.8	Structure and Name of the Isolated Spirostane-type and	19
	Steroid Saponins	
3.1	Scheme of Extraction, Fractionation and Isolation of the	32
	Roots of <i>C.Borivilianum</i>	
3.2	Schematic Diagram of Extraction Process and Solid	48
3.3	Summary of the Experimental Procedure	53
4.1	Scheme of Extraction Yield of each Chromatography Steps	56
	from the Roots of <i>C.Borivilianum</i>	
4.2	Assignments of Proton and Carbon-13 Resonances of	57
	Compound 1	
4.3	¹ H- ¹ H COSY Correlations (Bold Lines) and Key HMBC	59
	Correlations $(H \rightarrow C)$ of Di(2-ethylhexyl) Phthalate (DEHP)	
4.4	HPLC of the Crude Extract of the Roots of C. Borivilianum	62
4.5	HPLC of the Standard DEHP	63
4.6	HPLC of the Spike Test	63
4.7	UV-VIS Spectra of Standard DEHP	63
4.8	UV-VIS Spectra of DEHP in the Crude Extract Sample	64
4.9	Calibration Curve of DEHP	65
4.10	Effect of Type of Solvent on the Extraction Yield of DEHP	66
4.11	Effect of Extraction Time on the Extraction Yield of DEHP	67
4.12	Effect of Solid:Solvent Ratio on the Extraction Yield of	69
	DEHP	
4.13	Effect of Extraction Temperature on the Extraction Yield of	71
	DEHP	
4.14	Comparison Between Actual and Predicted Extraction Yield	76
	of DEHP, Y_1	
4.15	Corresponding Response Surface Graph and Contour Plot	77
	Showing Effects of Extraction time and Solid to Solvent Ratio	
	on the ExtractionYield of DEHP with Extraction	
	Temperature of 50°C	

4.16	Corresponding Response Surface Graph and Contour Plot	78
	Showing Effects of Extraction Time and Extraction	
	Temperature on the Extraction Yield of DEHP with Solid to	
	Solvent Ratio of 1:40 (g/mL)	
4.17	Corresponding Response Surface Graph and Contour	79
	Plot Showing Effects of Solid to Solvent Ratio and	
	Extraction Temperature on the Extraction Yield of	
	DEHP with Extraction Time of 90 min	
4.18	Desirability Ramp for the Optimization of DEHP Yield	81
4.19	Comparison Between Experimental Data and EEM Model	82
	for Different Temperatures	
4.20	Comparison Between Experimental Data and DEM model	83
	for Different Temperatures	
A.1	¹ H-NMR spectrum of di(2-ethylhexyl) phthalate (DEHP)	- 98
	taken at 500 Mhz	
A.2	¹³ C spectrum of di(2-ethylhexyl) phthalate (DEHP) taken at	99
	126 Mhz	
A.3	DEPT spectrum of di(2-ethylhexyl) phthalate (DEHP)	100
A.4	HMQC spectrum of di(2-ethylhexyl) phthalate (DEHP)	101
A.5	HMBC spectrum of di(2-ethylhexyl) phthalate (DEHP)	101
A 6	COSY spectrum of di(2-ethylbexyl) phthalate (DEHP)	103

LIST OF ABBREVIATIONS

1D, 2D	one-dimensional, two-dimensional
ANOVA	analysis of variance
CC	column chromatography
CD ₃ OD	deuterated methanol
BBD	Box-Behnken
CBA	chloroform extract
CBC	hexane extract
CBH	hexane extract
CCD	central composite design
C.V.	coefficient variation
СН	methine carbon atom
CH_2	methylene carbon
CH ₃	methyl carbon atom
COSY	correlation spectroscopy
d	doublet
DEHP	di(2-ethylhexyl) phthalate
DEPT	distortionless enhancement by polarization
FRIM	Forest Research Institute Malaysia
GRAS	Generally Recognized as Safe
HMBC	heteronuclear multiple bond correlation
HPLC	high performance liquid chromatography
HMQC	heteronuclear multiple quantum correlation
i.d.	internal diameter
J	coupling constant
Κ	number of variable
m	multiplet
n	total number of experiments
NMR	nuclear magnetic resonance
MS	mass spectroscopy
ODS	octadecylsilanized silica gel
PDA	photodiode array
PVDF	polyvinylidene difluoride
ppm	parts per million
q	quartet
R^2	correlation coefficient
R_f	retention factor
RSM	response surface methodology
S	singlet
SD	standard deviation
t	triplet
TLC	thin layer chromatography
TMS	tetramethylsilane
UAE	ultrasound-assisted extraction

UV	ultra violet spectroscopy
W_d	weight of the dried extract
W_s	weight of solid
x	independent variable
X_o	number of central points
x_{min}	minimum value of the independent variable
X_{max}	maximum value of the independent variable
Х	coded variable
X_1	coded variable
X_2	extraction time
X_3	solid to solvent ratio
X_1^2	extraction temperature
X_2^2	quadratic effect of extraction time
X_{3}^{2}	quadratic effect of solid to solvent ratio
X_1X_2	quadratic effect of extraction temperature
X_1X_3	interactive effect of extraction time and solid to solvent ratio
X_2X_3	interactive effect of extraction time and extraction temperature
Y_1	interactive effect of solid to solvent ratio and temperature
Y_i	yield of the crude extract
β_0	response variable
Bii	constant coefficient
Bii	linear coefficients
1 ()	

CHAPTER 1

INTRODUCTION

1.1 Introduction to Medicinal Plants

There are a lot of medicinal plants throughout the world with assorted pharmacological activities to treat certain type of diseases. Today, science has isolated the medicinal properties from a large quantity of various types of herbal plants. Most herbal plants remedies are used to treat the health problems and scientific reports have proven success in treating certain chronic conditions. It is crucial to develop a strict standardization procedure and perform pharmacognostical studies of herbal plants in order to avoid accidental herbal medicine misuse due to wrong identification of a medicinal plant and wrong prescription of traditional herbal medicine. Therefore, phytochemistry has become a major branch of pharmacognosy in developing markers for the objective of identification and standardization (Sankh, 2010).

1.2 Background of Study

It has been proven in scientific research that the plants have contributed us the active compounds that have potential therapeutic effects. The plant *Chlorophytum borivilianum* (Safed Musli) shown in Figure 1.1 is a medicinal plant belonging the family Liliaceae. *C. borivilianum* holds an important place in the traditional medicinal system due to its therapeutic importance. The economic part of the herb is its roots. Its roots are powdered and widely used in traditional folk medicines over past decades (Thakur et al., 2009a; Deore and Khadabadi, 2010a).



Figure 1.1: Plant and Roots of Chlorophytum Borivilianum (Safed Musli)

Presently, crude extract of *C. borivilianum* has been utilized for treatment of human diseases especially in the applications such as anti-diabetic (Panda et al., 2007), anti-oxidant (Kenjale et al., 2007), anti-ulcer (Panda et al., 2011a) and anti-inflammatory (Deore and Khadabadi, 2008). *In-vitro* and *in-vivo* studies have confirmed that the crude extract of C. *borivilianum* possesses a wide range of

noteworthy pharmacological activities. Interest is increasing to exploit this herbal plant for the development of therapeutics that could be potentially used as sexual stimulant for impotence and to prevent or treat cancers (Thakur et al., 2009b; Kumar et al., 2010).

1.3 Problems statement

There are extensive studies reported on the biological activities of the crude extract of the roots of C. borivilianum but there were only few studies had been done on the isolation of chemical constituents that responsible for the above reported applications. In addition, there were only few studies regarding the isolation of other chemical compounds since extensive works were carried out in the previous findings were aimed for the isolation of saponins from the roots of the plant. Due to the limited information regarding to the pure compounds other than saponins that have been isolated from this plant, therefore, this research focused on the isolation of any pure compounds other than saponins. At last, DEHP was isolated after few months of DEHP was reported to possess some remarkable biological isolation procedures. activities such as anti-leukemic, anti-microbial, anti-fungal, anti-tumour and antiviral. Therefore, it was selected to be optimized and it could be a starting point to pave a way to isolate and quantify other pure compounds (minor or rare) from this herbaceous plant in order to use it as a tool for quality control and also for the future development of other therapeutic applications. Furthermore, there is a lack of research on the optimization of the extraction parameters in order to maximize the yield of extract isolated from the roots of C. borivilianum. Therefore, it could be used as reference to isolate and elucidate a pure compound other than saponins and thereafter to optimize the isolated compound through response surface methodology (RSM).

To the best of our knowledge, no report was available in the literature regarding the optimization of extraction conditions from *C. borivilianum* through response surface methodology (RSM). Therefore, in this present study, there is a need to isolate and identify the chemical constituents that present in this plant and to identify the optimal conditions on the extraction of the roots of *C. borivilianum* using response methodology method in order to obtain the maximum yield of the isolated compound. Considering the residual toxicity of the extraction solvent, ethanol was opted for the extraction procedure. A single factor experimental design with five levels was studied to determine the preliminary experimental range of the extraction parameters on the extraction yield of DEHP, followed by the optimization of the extraction yield of DEHP by the RSM in combination with central composite design experimental design.

DEHP was reported to possess biological activities such as anti-leukemic, antimicrobial, anti-fungal, anti-tumour and antiviral. Considering the medicinal and pharmacological importance of *C. borivilianum* and the reported biological activities of DEHP, it is worth to optimize the extraction yield of DEHP in order to develop a systematic approach for the optimization of the other future isolated active compounds which are responsible for the reported pharmacological activities.

1.4 Objectives of the Study

With this respect, the objectives of this study are:

- 1. Extraction, isolation, purification and structure elucidation of pure compound from the roots of *C. borivilianum* by a combination of various chromatographic methods and different spectroscopic techniques, respectively.
- 2. Identification and quantification of the isolated compound by the mean of high performance liquid chromatography (HPLC) method.
- 3. Optimization of the extraction conditions and extraction kinetic of the isolated compound.

1.5 Scopes of Study

The work embodied in the present thesis is divided into following stages:

1. Isolation and structure elucidation of the pure compound from the roots of *C*. *borivilianum*

The process of extraction and isolation of the pure compound from the roots of the C. borivilianum was carried out by mean of chromatography method such as open column chromatography and thin layer chromatography. The structure of the pure compound isolated from the roots of C. borivilianum was elucidated by means of nuclear magnetic resonance (NMR) spectroscopy analysis. NMR spectroscopy of the structure elucidation included Proton $(^{13}C-NMR),$ NMR (¹H-NMR), Carbon-13 NMR distortionless enhancement by polarization transfer (DEPT), correlation spectroscopy heteronuclear multiple bond correlation (HMBC) and (COSY), heteronuclear multiple quantum correlation (HMQC). An in-vitro antiinflammatory assay was conducted for the C. borivilianum extract using Hyaluronidase (HYA) assay.

2. Identification and quantification of the isolated compound

A high performance liquid chromatography (HPLC) method was developed and implemented to identify the compound appeared in the standard and in the crude extract. The yield of the compound was quantified based on the calibration curve of the pure isolated compound.

3. Optimization of the extraction conditions and extraction kinetic of the isolated compound

Four operating parameters including extraction solvent, extraction time, solid to solvent ratio and extraction temperature were studies. Firstly, the selection of suitable solvent system was made and it was evaluated by quantifying the amount of extraction yield. Then, the experimental range of extraction time, solid to solvent ratio and extraction temperature was identified by a single factor experimental design. Also, a polynomial model for the extraction yield of compound from the roots of C. *borivilianum* was developed by mean of response surface methodology (RSM) using a central composite design (CCD) by Design Expert software. The optimum values for each of extraction parameter were established and the model was validated by performing the extraction of the compound under the optimal extraction conditions. Based on the optimization process, the extraction kinetics of the *C. borivilianum* was investigated and mathematical models for solid-liquid extraction of the *C. borivilianum* extract were applied to predict the extraction process.

REFERENCES

- Acharya, D., Mitaine-Offer, A.-C., Kaushik, N., Miyamoto, T., Paululat, T., and Lacaille-Dubois, M.-A. (2008a). Furostane-type steroidal saponins from the roots of chlorophytum borivilianum. *Helvetica Chimica Acta*, 91(12):2262–2269.
- Acharya, D., Mitaine-Offer, A.-C., Kaushik, N., Miyamoto, T., Paululat, T., and Lacaille-Dubois, M.-A. (2008b). Steroidal saponins from the roots of chlorophytum borivilianum. *Planta Medica*, 74(09):177–181.
- Acharya, D., Mitaine-Offer, A.-C., Kaushik, N., Miyamoto, T., Paululat, T., Mirjolet, J.-F., Duchamp, O., and Lacaille-Dubois, M.-A. (2009). Cytotoxic spirostane-type saponins from the roots of chlorophytum borivilianum. *Journal of Natural Products*, 72(1):177–181.
- Agil, R., Oomah, D., Mazza, G., and Hosseinian, F. (2012). Optimization of alkylresorcinols extraction from triticale bran using response surface methodology. *Food and Bioprocess Technology*, 5(7):2655–2664.
- Ba, D. and Boyaci, I. (2007). Modeling and optimization i: Usability of response surface methodology. *Journal of Food Engineering*, 78(3):836–845.
- Ballesteros, L., Teixeira, J., and Mussatto, S. (2014). Selection of the solvent and extraction conditions for maximum recovery of antioxidant phenolic compounds from coffee silverskin. *Food and Bioprocess Technology*, 7(5):1322–1332.
- Banik, R. and Pandey, D. (2008). Optimizing conditions for oleanolic acid extraction from lantana camara roots using response surface methodology. *Industrial Crops* and Products, 27(3):241–248.
- Bathoju, G. and Giri, A. (2012). Production of medicinally important secondary metabolites (stigmasterol and hecogenin) from root cultures of chlorophytum borivilianum (safed musli). *Recent Research In Science & Technology*, 4(5):45–48.
- Bezerra, M., Santelli, R., Oliveira, E., Villar, L., and Escaleira, L. (2008). Response surface methodology (rsm) as a tool for optimization in analytical chemistry. *Talanta*, 76(5):965–977.
- Bimakr, M., Rahman, R., Taip, F., Adzahan, N., Islam Sarker, M., and Ganjloo, A. (2013). Supercritical carbon dioxide extraction of seed oil from winter melon (benincasa hispida) and its antioxidant activity and fatty acid composition. *Molecules*, 18(1):997–1014.
- Chakraborthy, G., Dwivedi, P., and Purwar, S. (2009). Phytochemical and antimicrobial studies of chlorophytum borivilianum. *International Journal of Pharmaceutical Sciences and Drug Research*, 5(3):343–347.
- Chakraborthy, G., Zafar, R., and Aeri, V. (2008). Anti-inflammatory activity of methanolic extract of *Chlorophytum borivilianum*. Journal of Pharmacy Research, 1(1):58–60.
- Şahin, S., Aybastier, O., and Işik, E. (2013). Optimisation of ultrasonic-assisted extraction of antioxidant compounds from artemisia absinthium using response surface methodology. *Food Chemistry*, 141(2):1361–1368.
- Şahin, S. and Şamli, R. (2013). Optimization of olive leaf extract obtained by ultrasound-assisted extraction with response surface methodology. *Ultrasonics Sonochemistry*, 20(1):595–602.
- Dahmoune, F., Nayak, B., Moussi, K., Remini, H., and Madani, K. (2015). Optimization of microwave-assisted extraction of polyphenols from myrtus communis l. leaves. *Food Chemistry*, 166:585–595.
- Dang, J., Chen, C., Shao, Y., Mei, L., Zhang, H., Zhong, Z., Wang, Q., and Tao, Y. (2014). Optimization of extraction technology of gentiopicroside from gentiana

straminea maxim using response surface methodology on account of hplc. *Journal of Liquid Chromatography and Related Technologies*, 37(14):1940–1952.

- Deepak, V., Kalishwaralal, K., Ramkumarpandian, S., Babu, S. V., Senthilkumar, S., and Sangiliyandi, G. (2008). Optimization of media composition for nattokinase production by bacillus subtilis using response surface methodology. *Bioresource Technology*, 99(17):8170–8174.
- Deore, S. and Khadabadi, S. (2008). Antiinflammatory and antioxidant activity of chlorophytum borivilianum root extracts. *Asian Journal of Chemistry*, 20(2):983– 986.
- Deore, S. and Khadabadi, S. (2009a). Larvicidal activity of the saponin fractions of chlorophytum borivilianum santapau and fernandes. *Journal of Entomology and Nematology*, 1(5):64–66.
- Deore, S. and Khadabadi, S. (2009b). Screening of antistress properties of chlorophytum borivilianum tuber. *Pharmacologyonline*, 1:320–328.
- Deore, S. and Khadabadi, S. (2010a). Development and evaluation of safed musli formulation. *Archives Appl. Sci. Res*, 2(1):324–328.
- Deore, S. and Khadabadi, S. (2010b). In vitro anthelmintic studies of chlorophytum borivilianum sant. and fernandez tubers. *Indian Journal of Natural Products and Resources*, 1(1):53–56.
- Deore, S. and Khadabadi, S. (2010c). Isolation and characterization of phytoconstituents from chlorophytum borivilianum. *Pharmacognosy Research*, 2(6):343–349.
- Desale, P. (2013). Safed musli: Herbal viagra for male impotence. Journal of Medicinal Plants Studies, 1(3):91–97.
- Dirar, A. I., Mohamed, M. A., Ahmed, W. J., Mohammed, M. S., Khalid, H. S., and Garelnabi, E. A. (2014). Isolation and characterization of potential cytotoxic leads from ambrosia maritima l.(asteraceae). *Journal of Pharmacognosy and Phytochemistry*, 3(4):38–41.
- Dodke, J., Sao, S., and Sahu, P. (2013). Phytochemical screening of root of chlorophytum borivilianum l. (safed musli): A medicinal plant. *International Journal of Pharma and Bio Sciences*, 4(4):B237–B241.
- El-Sayed, M., Abdel-Aziz, M., Abdel-Gawad, M., Abdel-Hameed, E.-S., Ahmed, W., and Abdel-Lateef, E. (2013). Chemical constituents and cytotoxic activity of cassia glauca lan. leaves. *Life Science Journal*, 10(3):1617–1625.
- El-Sayed, O., Asker, M., Shash, S., and Hamed, S. (2015). Isolation, structure elucidation and biological activity of di- (2-ethylhexyl) phthalate produced by penicillium janthinellum 62. *International Journal of ChemTech Research*, 8(1):58– 66. cited By 2.
- Fang, X., Wang, J., Wang, Y., Li, X., Zhou, H., and Zhu, L. (2014). Optimization of ultrasonic-assisted extraction of wedelolactone and antioxidant polyphenols from eclipta prostrate l using response surface methodology. *Separation and Purification Technology*, 138:55–64.
- Fariza, N. (2014). Extraction, Characterization, Optimization And Modeling of Phalerin from Phaleria Macrocarpa (Scheef.) Boerl Leaves. Doctoral thesis, Universiti Putra Malaysia.
- Feng, S., Cheng, H., Fu, L., Ding, C., Zhang, L., Yang, R., and Zhou, Y. (2014). Ultrasonic-assisted extraction and antioxidant activities of polysaccharides from camellia oleifera leaves. *International Journal of Biological Macromolecules*, 68:7– 12.
- Ferreira, S., Bruns, R., da Silva, E., dos Santos, W., Quintella, C., David, J., de Andrade, J., Breitkreitz, M., Jardim, I., and Neto, B. (2007). Statistical designs and

response surface techniques for the optimization of chromatographic systems. *Journal of Chromatography A*, 1158(1-2):2–14.

- Fliniaux, O., Corbin, C., Ramsay, A., Renouard, S., Beejmohun, V., Doussot, J., FalguiÃÍres, A., Ferroud, C., Lamblin, F., LainÃľ, E., Roscher, A., Grand, E., Mesnard, F., and Hano, C. (2014). Microwave-assisted extraction of herbacetin diglucoside from flax (linum usitatissimum 1.) seed cakes and its quantification using an rp-hplc-uv system. *Molecules*, 19(3):3025–3037.
- Gia"o, M., Pereira, C., Fonseca, S., Pintado, M., and Malcata, F. (2009). Effect of particle size upon the extent of extraction of antioxidant power from the plants agrimonia eupatoria, salvia sp. and satureja montana. *Food Chemistry*, 117(3):412– 416.
- Govindarajan, R., Sreevidya, N., Vijayakumar, M., Thakur, M., Dixit, V., Mehrotra, S., and Pushpangadan, P. (2005). In vitro antioxidant activity of ethanolic extract of chlorophytum borivilianum. *Natural Product Sciences*, 11(3):165–169.
- Guo, H., Zhang, Z., Qian, J., and Liu, Y. (2013a). Optimization of the liquid-liquid extraction of phosphatidyl-choline from rapeseed oil gums by response surface methodology. *Industrial Crops and Products*, 42(1):500–506.
- Guo, L., Zhu, W., Xu, F., Liu, M., Xie, Y., and Zhang, J. (2014). Optimized ultrasonicassisted extraction of polysaccharides from cyclina sinensis and evaluation of antioxidant activities in vitro. *CYTA - Journal of Food*, 12(1):32–39.
- Guo, L., Zhu, W.-C., Liu, Y.-T., Wu, J.-Y., Zheng, A.-Q., and Liu, Y.-L. (2013b). Response surface optimized extraction of flavonoids from mimenghua and its antioxidant activities in vitro. *Food Science and Biotechnology*, 22(5):1–8.
- Habib, M. and Karim, M. (2009). Antimicrobial and cytotoxic activity of di-(2ethylhexyl) phthalate and anhydrosophoradiol-3-acetate isolated from calotropis gigantea (linn.) flower. *Mycobiology*, 37(1):31–36.Habib, M. and Karim, M. (2012). Antitumour evaluation of di-(2-ethylhexyl) phthalate (dehp) isolated from calotropis gigantea l. flower. *Acta Pharmaceutica*, 62(4):607–615.
- Handa, S. (2008). Extraction Technologies for Medicinal and Aromatic Plants. International Centre for Science and High Technology. Retrieved Jan 4th, 2015. Hossain, M., Brunton, N., Patras, A., Tiwari, B., O'Donnell, C., Martin-Diana,
- A., and Barry-Ryan, C. (2012). Optimization of ultrasound assisted extractionof antioxidant compounds from marjoram (origanum majorana 1.) using response surface methodology. *Ultrasonics Sonochemistry*, 19(3):582–590.
- Hu, T., Guo, Y.-Y., Zhou, Q.-F., Zhong, X.-K., Zhu, L., Piao, J.-H., Chen, J., and Jiang, J.-G. (2012). Optimization of ultrasonic-assisted extraction of total saponins from eclipta prostrasta l. using response surface methodology. *Journal of Food Science*, 77(9):C975–C982.
- Ilbay, Z., Şahin, S., and Kirbaşlar, S. (2013). Optimisation of ultrasound-assisted extraction of rosehip (rosa canina l.) with response surface methodology. *Journal of the Science of Food and Agriculture*, 93(11):2804–2809.
- Jiménez-Contreras, E., Torres-Salinas, D., Moreno, R., BaÃsos, R., and LÃspez-CÃszar, E. (2009). Response surface methodology and its application in evaluating scientific activity. *Scientometrics*, 79(1):201–218.
- Jovanovic-Malinovska, R., Kuzmanova, S., and Winkelhausen, E. (2015). Application of ultrasound for enhanced extraction of prebiotic oligosaccharides from selected fruits and vegetables. *Ultrasonics Sonochemistry*, 22:446–453.
- Katade, S., Pawar, P., Tungikar, V., Tambe, A., Kalal, K., Wakharkar, R., and Deshpande, N. (2006). Larvicidal activity of bis(2-ethylhexyl) benzene-1,2dicarboxylate from sterculia guttata seeds against two mosquito species. *Chemistry* and Biodiversity, 3(1):49–53.

- Kavitha, A., Prabhakar, P., Vijayalakshmi, M., and Venkateswarlu, Y. (2009). Production of bioactive metabolites by nocardia levis mk-vl-113. *Letters in Applied Microbiology*, 49(4):484–490.
- Kenjale, R., Shah, R., and Sathaye, S. (2007). Anti-stress and anti-oxidant effects of roots of chlorophytum borivilianum santa pau and fernandes. *Indian Journal of Experimental Biology*, 45(11):974–979.
- Kenjale, R., Shah, R., and Sathaye, S. (2008). Effects of chlorophytum borivilianum on sexual behaviour and sperm count in male rats. *Phytotherapy Research*, 22(6):796– 801.
- Keshani, S., Luqman Chuah, A., Nourouzi, M., Russly, A., and Jamilah, B. (2010). Optimization of concentration process on pomelo fruit juice using response surface methodology (rsm). *International Food Research Journal*, 17(3):733–742.
- Kim, H., Lee, A., Jo, J., Moon, B., Chun, J., Choi, G., and Kim, H. (2014). Optimization of ultrasound-assisted extraction of quercitrin from houttuynia cordata thunb. using response surface methodology and uplc analysis. *Food Science and Biotechnology*, 23(1):1–7.
- Kim, H., Lee, A., Jo, J., Moon, B., Ji, Y., and Kim, H. (2013). Optimization of ultrasonic-assisted extraction of continentalic acid from the root of aralia continentalis by using the response surface methodology. *Archives of Pharmacal Research*, pages 1–8.
- Kim, S., Kim, H., Yang, E., Lee, K., Kim, S., Kim, Y., and Sung, S. (2010). Optimization of pressurized liquid extraction for spicatoside a in liriope platyphylla. *Separation and Purification Technology*, 71(2):168–172.
- King, S. L. (1998). Structure Elucidation and Oxidation Chemistry of Natural Products. Doctoral thesis, The University of Hong Kong. Retrieved Jun 16th, 2014.
- Kumar, M., Meena, P., Verma, S., Kumar, M., and Kumar, A. (2010). Anti-tumour, anti- mutagenic and chemomodulatory potential of chlorophytum borivilianum. *Asian Pacific Journal of Cancer Prevention*, 11(2):327–334.
- Lai, J., Wang, H., Wang, D., Fang, F., Wang, F., and Wu, T. (2014). Ultrasonic extraction of antioxidants from chinese sumac (rhus typhina l.) fruit using response surface methodology and their characterization. *Molecules*, 19(7):9019–9032.
- Lan, G., Chen, H., Wang, Z., Zhang, W., and Zhang, L. (2011). Extraction of polygonatum odoratum polysaccharides using response surface methodology and preparation of a compound beverage. *Carbohydrate Polymers*, 86(3):1175–1180.
- Lee, K., Kim, J., Lim, D., and Kim, C. (2000). Anti-leukaemic and anti-mutagenic effects of di(2-ethylhexyl)phthalate isolated from aloe vera linne. *Journal of Pharmacy and Pharmacology*, 52(5):593–598.
- Li, H., Zhu, H., Qiao, J., Du, J., and Zhang, H. (2012). Optimization of the main liming process for inulin crude extract from jerusalem artichoke tubers. *Frontiers of Chemical Science and Engineering*, 6(3):348–355.
- Liu, B., Ma, Y., Liu, Y., Yang, Z., and Zhang, L. (2013). Ultrasonic-assisted extraction and antioxidant activity of flavonoids from adinandra nitida leaves. *Tropical Journal of Pharmaceutical Research*, 12(6):1045–1051.
- Mavar-Manga, H., Haddad, M., Pieters, L., Baccelli, C., Penge, A., and Quetin-Leclercq, J. (2008). Anti-inflammatory compounds from leaves and root bark of alchornea cordifolia (schumach. & thonn.) mÃijll. arg. *Journal of Ethnopharmacology*, 115(1):25–29.
- Mehrnoush, A., Mustafa, S., Sarker, M., and Yazid, A. (2011). Optimization of the conditions for extraction of serine protease from kesinai plant (streblus asper) leaves using response surface methodology. *Molecules*, 16(11):9245–9260.

- Mei, L., Zhen-Chang, W., Hao-Jie, D., Li, C., Qing-Gang, X., and Jing, L. (2009). Response surface optimization of polysaccharides extraction from liriope roots and its modulatory effect on sjogren syndrome. *International Journal of Biological Macromolecules*, 45(3):284–288.
- Meziane, S. (2013). Optimization of oil extraction from olive pomace using response surface methodology. *Food Science and Technology International*, 19(4):315–322. Moslavac, T., JokiÄĞ, S., ÅăubariÄĞ, D., AladiÄĞ, K., Vukoja, J., and Prce, N. (2014). Pressing and supercritical co2 extraction of camelina sativa oil. *IndustrialCrops and Products*, 54:122–129.
- Myers, R., Montgomery, D., Geoffrey Vining, G., Borror, C., and Kowalski, S. (2004). Response surface methodology: A retrospective and literature survey. *Journal of Quality Technology*, 36(1):53–78.
- Nair, J., Ndhlala, A., Chukwujekwu, J., and Van Staden, J. (2012). Isolation of di(2ethylhexyl) phthalate from a commercial south african cognate herbal mixture. *South African Journal of Botany*, 80:21–24.
- Narasimhan, S., Govindarajan, R., Madhavan, V., Thakur, M., Dixit, V., Mehrotra, S., and Madhusudanan, K. (2006). Action of (2→1)fructo-oligopolysaccharide fraction of chlorophytum borivilianum against streptozotocin-induced oxidative stress. *Planta Medica*, 72(15):1421–1424.
- Panda, S., Das, D., and Tripathy, N. (2011a). Studies on anti-ulcer activity of roottubers of chlorophytum borivilianum santapau and fernandes. *International Journal of Pharmaceutical Sciences Review and Research*, 9(2):65–68.
- Panda, S., Das, D., and Tripathy, N. (2011b). A study on antipyretic activity of chlorophytum borivilianum sant. and fern. root tubers. *International Journal of Pharmaceutical Research and Development*, 3(3):153–156.
- Panda, S., Si, S., and Bhatnagar, S. (2007). Studies on hypoglycaemic and analgesic activities of chlorophytum borivilianum sant and ferz. *Journal of Natural Remedies*, 7(1):31–36.
- Pandey, D. and Banik, R. (2012). Optimization of extraction conditions for colchicine from gloriosa superba tubers using response surface methodology. *Journal of Agricultural Technology*, 8(4):1301–1315.
- Peng, L.-X., Zou, L., Zhao, J.-L., Xiang, D.-B., Zhu, P., and Zhao, G. (2013). Response surface modeling and optimization of ultrasound-assisted extraction of three flavonoids from tartary buckwheat (fagopyrum tataricum). *Pharmacognosy Magazine*, 9(35):210–215.
- Pin, K., Luqman Chuah, A., Abdull Rashih, A., Rasadah, M., Law, C., and Choong, T. (2011). Solid-liquid extraction of betel leaves (piper betle 1.). *Journal of Food Process Engineering*, 34(3):549–565. cited By 8.
- Prakash Maran, J. and Manikandan, S. (2012). Response surface modeling and optimization of process parameters for aqueous extraction of pigments from prickly pear (opuntia ficus-indica) fruit. *Dyes and Pigments*, 95(3):465–472.
- Prasad, K., Hassan, F., Yang, B., Kong, K., Ramanan, R., Azlan, A., and Ismail, A. (2011). Response surface optimisation for the extraction of phenolic compounds and antioxidant capacities of underutilised mangifera pajang kosterm. peels. *Food Chemistry*, 128(4):1121–1127.
- Prasad, K., Kong, K., Ramanan, R., Azlan, A., and Ismail, A. (2012). Determination and optimization of flavonoid and extract yield from brown mango using response surface methodology. *Separation Science and Technology*, 47(1):73–80.
- Prasad, K., Yang, E., Yi, C., Zhao, M., and Jiang, Y. (2009). Effects of high pressure extraction on the extraction yield, total phenolic content and antioxidant activity of

longan fruit pericarp. Innovative Food Science and Emerging Technologies, 10(2):155–159.

- Prommajak, T., Surawang, S., and Rattanapanone, N. (2014). Ultrasonic-assisted extraction of phenolic and antioxidative compounds from lizard tail (houttuynia cordata thunb.). Songklanakarin Journal of Science and Technology, 36(1):65–72. Purohit, S., Dave, A., and Kukda, G. (1994). Micropropagation of safed musli (chlorophytum borivilianum), a rare indian medicinal herb. *Plant Cell, Tissue and Organ Culture*, 39(1):93–96.
- Qi, S.-H., Xu, Y., Gao, J., Qian, P.-Y., and Zhang, S. (2009). Antibacterial and antilarval compounds from marine bacterium pseudomonas rhizosphaerae. *Annals* of *Microbiology*, 59(2):229–233.
- Rice, G. R. and Do, D. D. (1999). Applied mathematics and modelling for chemical engineers. John Wiley & Son, Inc., USA.
- Rizvi, M., Kukreja, A., and Khanuja, S. (2007). In vitro culture of chlorophytum borivilianum sant. et fernand. in liquid culture medium as a cost-effective measure. *Current Science*, 92(1):87–90.
- Robert, C., Devillers, T., Wathelet, B., Van Herck, J.-C., and Paquot, M. (2006). Use of a plackett-burman experimental design to examine the impact of extraction parameters on yields and compositions of pectins extracted from chicory roots (chicorium intybus l.). *Journal of Agricultural and Food Chemistry*, 54(19):7167– 7174.
- Sankh, A. C. (2010). Phytochemical Investigation and Antidiabetic Activity of Roots of Rotula Aquatica Lour. Master thesis, K.L.E.SOs College of Pharmacy, Belgaum. Retrieved December 31st, 2014.
- Sathishkumar, T., Baskar, R., Shanmugam, S., Rajasekaran, P., Sadasiyam, S., and Manikandan, V. (2008). Optimization of flavonoids extraction from the leaves of tabernaemontana heyneana wall. using 116 orthogonal design. *Nature and Science*, 6:10–19.
- Satija, J. and Singh, G. (2005). Safed musli: The golden root. *Indian Journal of Pharmaceutical Education and Research*, 39(4):180–185.
- Shao, Q., Deng, Y., Liu, H., Zhang, A., Huang, Y., Xu, G., and Li, M. (2014a). Essential oils extraction from anoectochilus roxburghii using supercritical carbon dioxide and their antioxidant activity. *Industrial Crops and Products*, 60:104–112. Shao, Y., Wu, Q.-N., Duan, J.-A., Yue, W., Gu, W., and Wang, X. (2014b). Optimisation of the solvent extraction of bioactive compounds from lophatherum gracile brongn. using response surface methodology and hplc-pad coupled with precolumn antioxidant assay. *Analytical Methods*, 6(1):170–177.
- Sharma, G. and Kumar, M. (2012). Antioxidant and modulatory role of chlorophytum borivilianum against arsenic induced testicular impairment. *Journal of Environmental Sciences (China)*, 24(12):2159–2165.
- Sharma, S. and Kumar, M. (2011). Hepatoprotective effect of chlorophytum borivilianum root extract against arsenic intoxication. *Pharmacologyonline*, 3:1021–1032.
- Shi, D., Ding, H., and Xu, S. (2014). Optimization of microwave-assisted extraction of wedelolactone from eclipta alba using response surface methodology. *Frontiers of Chemical Science and Engineering*, 8(1):34–42.
- Singh, D., Pokhriyal, B., Joshi, Y. M., and Kadam, V. (2012). Phytopharmacological aspects of chlorophytum borivilianum (safed musli): A review. *Int J Res Pharm Chem*, 2(3):853–859.
- Smaoui, S., Mellouli, L., Lebrihi, A., Coppel, Y., Fguira, L., and Mathieu, F. (2011). Purification and structure elucidation of three naturally bioactive molecules from

the new terrestrial streptomyces sp. tn17 strain. *Natural Product Research*, 25(8):806–814.

- Steinberg, D. and Bursztyn, D. (2010). Response surface methodology in biotechnology. *Quality Engineering*, 22(2):78–87.
- Stroescu, M., Stoica-Guzun, A., Ghergu, S., Chira, N., and Jipa, I. (2013). Optimization of fatty acids extraction from portulaca oleracea seed using response surface methodology. *Industrial Crops and Products*, 43(1):405–411.
- Sui, Z., Li, L., Liu, B., Gu, T., Zhao, Z., Liu, C., Shi, C., and Yang, R. (2013). Optimum conditions for radix rehmanniae polysaccharides by rsm and its antioxidant and immunity activity in uvb mice. *Carbohydrate Polymers*, 92(1):283– 288.
- Sulyok, E. (2013). Isolation and Structure Elucidation of Diterpenes from Euphorbia pannonica, E. esula and E. falcata. Doctoral thesis, University of Szeged. Retrieved December 16th, 2014.
- Sundaram, S. and Aeri, V. (2009). Antibacterial activities of crude extracts of chlorophytum borivilianum to bacterial pathogens. *Research Journal of Medicinal Plant*, 1(2):110–112.
- Swami, U., Lande, A., Ghadge, P., Adkar, P., and Ambavade, S. (2014). Pharmacological evaluation of chlorophytum borivilianum sant. & fern. for anxiolytic activity and effect on brain gaba level. *Oriental Pharmacy and Experimental Medicine*, 14(2):169–180.
- Tahmouzi, S. (2014). Optimization of polysaccharides from zagros oak leaf using rsm: Antioxidant and antimicrobial activities. *Carbohydrate Polymers*, 106(1):238–246.
 Tasharrofi, N., Adrangi, S., Fazeli, M., Rastegar, H., Khoshayand, M., and Faramarzi, M. (2011). Optimization of chitinase production by bacillus pumilus using plackett-burman design and response surface methodology. *Iranian Journal of*

Pharmaceutical Research, 10(4):759–768.

- Teng, H. and Choi, Y. (2014). Optimization of ultrasonic-assisted extraction of bioactive alkaloid compounds from rhizoma coptidis (coptis chinensis franch.) using response surface methodology. *Food Chemistry*, 142:299–305.
- Tesso, H. (2005). *Isolation and Structure Elucidation of Natural Products from Plants*. Doctoral thesis, University of Hamburg. Retrieved Jun 16th, 2014.
- Thakur, G., Bag, M., Sanodiya, B., Debnath, M., Zacharia, A., Bhadauriya, P., Prasad, G., and Bisen, P. (2009a). Chlorophytum borivilianum: A white gold for biopharmaceuticals and neutraceuticals. *Current Pharmaceutical Biotechnology*, 10(7):650–666.
- Thakur, M., Bhargava, S., and Dixit, V. (2007). Immunomodulatory activity of chlorophytum borivilianum sant. f. *Evidence-based Complementary and Alternative Medicine*, 4(4):419–423.
- Thakur, M., Bhargava, S., Praznik, W., Loeppert, R., and Dixit, V. (2009b). Effect of chlorophytum borivilianum santapau and fernandes on sexual dysfunction in hyperglycemic male rats. *Chinese Journal of Integrative Medicine*, 15(6):448–453. Thakur, M., Connellan, P., Deseo, M., Morris, C., and Dixit, V. (2011). Immunomodulatory polysaccharide from chlorophytum borivilianum roots. *Evidence-based Complementary and Alternative Medicine*, 2011.
- Vyawahare, N., Kagathara, V., Kshirsagar, A., Rajendran, R., Patil, M., Jagtap, A., and Sadar, S. (2009). Effect of hydroalcoholic extract of *Chlorophytum borivilianum* tubers in alleviating the diabetic impotency in streptozotocin induced male diabetic rats. *Pharmacognosy Research*, 1(5):314–319.

- Wang, C., Shi, L., Fan, L., Ding, Y., Zhao, S., Liu, Y., and Ma, C. (2013a). Optimization of extraction and enrichment of phenolics from pomegranate (punica granatum l.) leaves. *Industrial Crops and Products*, 42(1):587–594.
- Wang, L., Wang, Z., and Li, X. (2013b). Optimization of ultrasonic-assisted extraction of phenolic antioxidants from malus baccata (linn.) borkh. using response surface methodology. *Journal of Separation Science*, 36(9-10):1652–1658.
- Wang, X., Wu, Y., Chen, G., Yue, W., Liang, Q., and Wu, Q. (2013c). Optimisation of ultrasound assisted extraction of phenolic compounds from sparganii rhizoma with response surface methodology. *Ultrasonics Sonochemistry*, 20(3):846–854.
- Wong, C. (2017). Isolation of Saponins from Solanum Mammosum and Characterization of Their Anticancer Activity by Proteomics. Master thesis, The University of Hong Kong. Retrieved Jun 19th, 2014.
- Wu, Z., Li, H., Tu, D., Yang, Y., and Zhan, Y. (2013a). Extraction optimization, preliminary characterization, and in vitro antioxidant activities of crude polysaccharides from finger citron. *Industrial Crops and Products*, 44:145– 151.
- Wu, Z., Ruan, H., Wang, Y., Chen, Z., and Cui, Y. (2013b). Optimization of microwave- assisted extraction of puerarin from radix puerariae using response surface methodology. *Separation Science and Technology (Philadelphia)*, 48(11):1657–1664.
- Xie, D.-T., Wang, Y.-Q., Kang, Y., Hu, Q.-F., Su, N.-Y., Huang, J.-M., Che, C.-T., and Guo, J.-X. (2014a). Microwave-assisted extraction of bioactive alkaloids from stephania sinica. *Separation and Purification Technology*, 130:173–181.
- Xie, Z., Sun, Y., Lam, S., Zhao, M., Liang, Z., Yu, X., Yang, D., and Xu, X. (2014b). Extraction and isolation of flavonoid glycosides from flos sophorae immaturus using ultrasonic-assisted extraction followed by high-speed countercurrent chromatography. *Journal of Separation Science*, 37(8):957–965.
- Xu, W., Chu, K., Li, H., Chen, L., Zhang, Y., and Tang, X. (2011). Extraction of lepidium apetalum seed oil using supercritical carbon dioxide and anti-oxidant activity of the extracted oil. *Molecules*, 16(12):10029–10045.
- Xynos, N., Papaefstathiou, G., Gikas, E., Argyropoulou, A., Aligiannis, N., and Skaltsounis, A.-L. (2014). Design optimization study of the extraction of olive leaves performed with pressurized liquid extraction using response surface methodology. *Separation and Purification Technology*, 122:323–330.
- Yu, P. and Chao, X. (2013). Statistics-based optimization of the extraction process of kelp polysaccharide and its activities. *Carbohydrate Polymers*, 91(1):356–362.
- Zakia, K., Singh, O., Singh, R., and Bhat, I. U. H. (2013). Safed musli (chlorophytum borivilianum): A review of its botany, ethnopharmacology and phytochemistry. *Journal of Ethnopharmacology*, 150(2):421 441.
- Zhang, H., Zhang, L., Hu, X., Zhou, Y., Ding, C., Yang, R., Wang, X., and Li, D. (2014a). Optimization of ultrasound-assisted extraction of artemisinin from artemisia annua l. by response surface methodology. *Separation Science and Technology (Philadelphia)*, 49(5):673–681.
- Zhang, Y., Wang, L., Zhang, D., Zhou, L., and Guo, Y. (2014b). Ultrasound-assisted extraction and purification of schisandrin b from schisandra chinensis (turcz.) baill seeds: Optimization by response surface methodology. *Ultrasonics Sonochemistry*, 21(2):461–466.
- Zheng, Y., Li, Y., and Wang, W.-D. (2014). Optimization of ultrasonic-assisted extraction and in vitro antioxidant activities of polysaccharides from trametes orientalis. *Carbohydrate Polymers*, 111:315–323.

Zou, T., Wu, H., Li, H., Jia, Q., and Song, G. (2013). Comparison of microwave-assisted and conventional extraction of mangiferin from mango (mangifera indica l.) leaves. *Journal of Separation Science*, 36(20):3457–3462.Zou, T.-B., Xia, E.-Q., He, T.-P., Huang, M.-Y., Jia, Q., and Li, H.-W. (2014). Ultrasound-assisted extraction of mangiferin from mango (mangifera indica l.) leaves using response surface methodology. *Molecules*, 19(2):1411–1421.

