



**UNIVERSITI PUTRA MALAYSIA**

***MOLECULAR CLONING, CHARACTERIZATION, AND PROMOTER  
ANALYSIS OF VITAMIN E BIOSYNTHETIC GENES FROM THE OIL PALM***

**KONG SZE LING**

**ITA 2013 8**



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**MASTER OF SCIENCE  
UNIVERSITI PUTRA MALAYSIA**

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By

**KONG SZE LING**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in  
Fulfillment of the Requirements for the Degree of Master of Science**

**July 2013**

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

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By

**KONG SZE LING**

**July 2013**

**Chairman: Prof. Datin Siti Nor Akmar Abdullah, PhD**

**Faculty: Institute of Tropical Agriculture**

Tocopherols and tocotrienols, commonly known as vitamin E, play a crucial role in human and animal nutrition. In recent years, tocotrienols have been reported as a powerful antioxidant agent and linked with various potential health benefits such as anti-angiogenic properties exhibited by palm tocotrienols. Therefore this brings the interest to carry out isolation and characterization of vitamin E biosynthetic genes from the oil palm (*E. guineensis* and *E. oleifera*) since crude palm oil has been well known to be the richest source of tocotrienols in nature. Homogentisate geranylgeranyl transferase (HGGT) and homogentisate phytyltransferase (HPT) are the two key enzymes that catalyse the condensation of homogentisic acid (HGA) with a prenyldiphosphate to produce tocotrienols and tocopherols in plants, respectively. The partial cDNAs encoding HGGT and HPT enzymes were successfully isolated from both oil palm species by PCR amplification using degenerate primers. Subsequently, full length cDNA sequences were obtained by rapid amplification of cDNA ends (RACE) using gene-specific primers. The full length deduced amino acid sequences were further analyzed using various bioinformatics tools available publicly. The analysis revealed the presence of an UbiA prenyltransferase conserved domain in all four protein sequences and suggested that oil palm HGGT and HPT are more evolutionarily related with their counterparts from other monocot plant species based on the result from homologous alignment and phylogenetic analysis. Next, quantitative gene expression analysis was carried out to elucidate the transcript profiles of the oil palm HGGT and HPT genes in different tissues and at different developmental stages of the mesocarp by real-time PCR. Two reference genes that showed to be stably expressed in each experimental set were identified using geNorm software. The expression level of each target gene in each experimental sample was subsequently determined by normalizing to the two validated reference genes. Overall result showed that the oil palm HGGT and HPT transcript production is spatially and temporally regulated. The HPT gene was constitutively expressed in all tested tissues except in 15 w.a.a kernel whereas oil palm HGGT gene showed preferential expression in mesocarp and kernel tissues and highly expressed when active oil deposition occurred in 17 w.a.a mesocarp. This indicates that HGGT expression is regulated by the oil synthesis process in palm fruits. Lastly, genome-

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walking PCR successfully amplified the HGGT promoter region of both oil palm species. By searching in PLACE, PlantCARE and PlantPAN databases, a number of important *cis*-regulatory elements were found and comparison between these data has resulted in the identification of several common motifs which may be involved in coordinating expression of these genes. The motifs basically can be divided into four main groups including phytohormone-responsive, light-responsive, abiotic factor-responsive and endosperm specificity. This suggests that the regulation of HGGT expression in *E. guineensis* and *E. oleifera* involved many similar factors. Further characterization of the potential important motifs would facilitate better understanding on the regulatory mechanism of tocotrienol synthesis in oil palm at the molecular level.



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Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan Ijazah Master Sains

**PENGLONAN, PENCIRIAN, DAN ANALISIS PROMOTER GEN-GEN  
BIOSINTESIS VITAMIN E DARIPADA KELAPA SAWIT**

Oleh

**KONG SZE LING**

**Julai 2013**

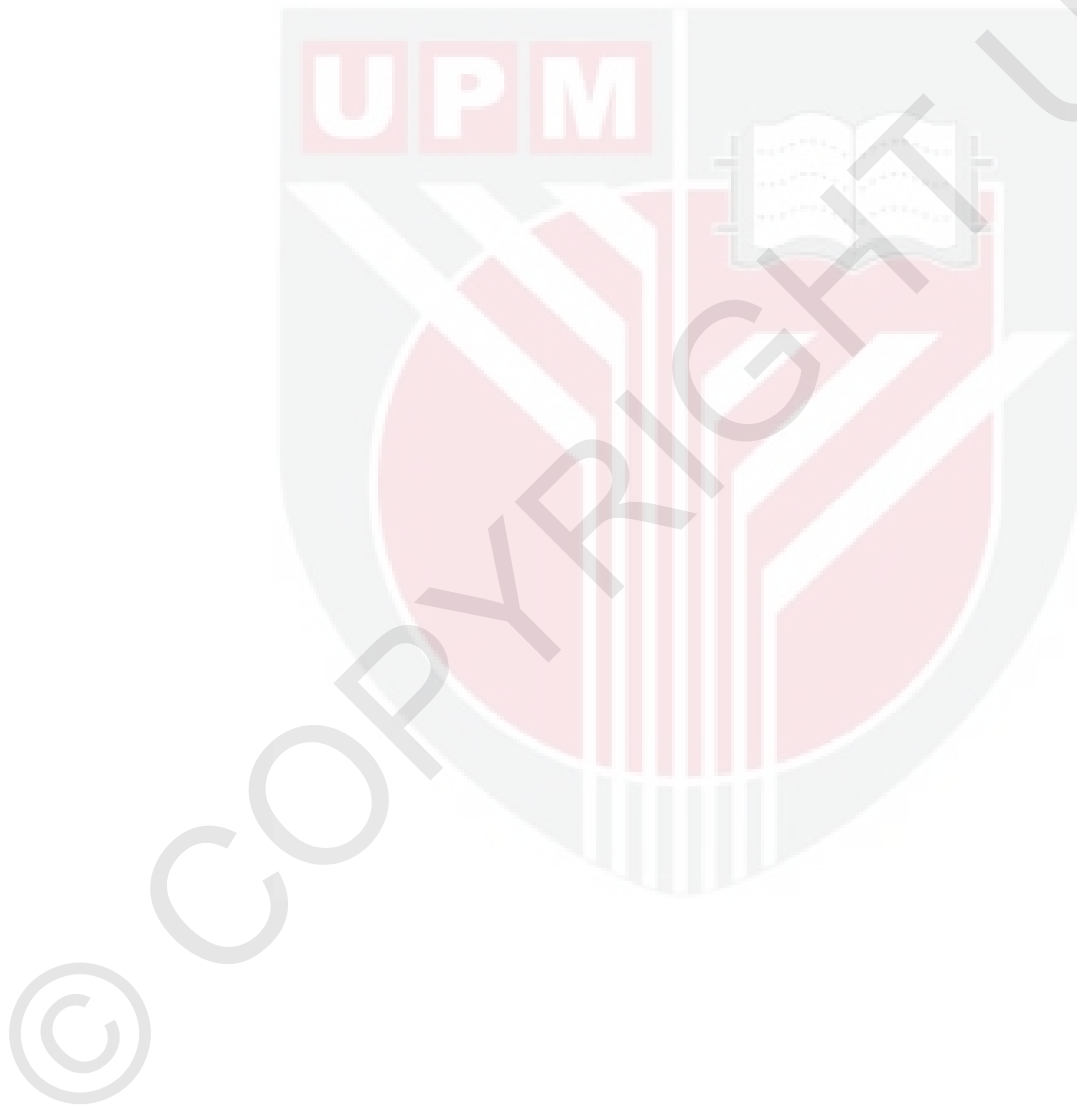
**Pengerusi: Prof. Datin Dr. Siti Nor Akmar Abdullah, PhD**

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Tokoferol dan tokotrienol yang dikenali secara am sebagai vitamin E memainkan peranan penting dalam pemakanan manusia dan haiwan. Beberapa tahun kebelakangan ini, tokotrienol telah dilaporkan sebagai agen antioksidan yang hebat dan dikaitkan dengan pelbagai manfaat kesihatan yang berpotensi seperti ciri-ciri anti-angiogenik yang dipamerkan oleh tokotrienol sawit. Oleh itu, perhatian diberikan untuk memencilkan dan mencirikan gen-gen biosintesis vitamin E daripada minyak sawit (*E. guineensis* dan *E. oleifera*) memandangkan minyak sawit mentah merupakan sumber asli yang terkaya dengan tokotrienol. Homogentisat geranilgeranil transferase (HGGT) dan homogentisat fitiltransferase (HPT) adalah dua enzim utama yang menjadi pemangkin dalam pemeluwapan asid homogentisik (HGA) dengan prenildifosfat untuk menghasilkan tokotrienol dan tokoferol dalam tumbuhan. Amplifikasi PCR menggunakan pencetus degenerasi telah berjaya memencilkan cDNA separa lengkap yang mengekod enzim HGGT dan HPT daripada kedua-dua spesies kelapa sawit. Seterusnya, jujukan lengkap cDNA telah diperolehi melalui amplifikasi pantas hujung cDNA dengan penggunaan pencetus spesifik. Jujukan asid amino yang dijangka telah dianalisis dengan menggunakan perisian bioinformatik awam. Analisis tersebut mendedahkan kehadiran domain terpelihara UbiA prenilttransferase dalam kesemua jujukan protein sementara keputusan penjajaran homolog dan analisis filogenetik mencadangkan HGGT dan HPT sawit lebih mempunyai pertalian evolusi dengan tumbuhan monokot lain. Seterusnya, analisis kuantitatif pengekspresan gen dijalankan untuk mendapatkan profil transkrip HGGT dan HPT dalam tisu kelapa sawit yang berlainan dan juga pelbagai peringkat perkembangan dalam mesokarpa melalui kaedah “real-time” PCR. Dua gen rujukan yang menunjukkan tahap pengekspresan yang stabil dalam setiap set eksperimen telah dikenalpasti oleh perisian geNorm. Tahap pengekspresan untuk gen sasaran masing-masing dalam setiap sampel kemudiannya dinormalisasikan oleh kedua-dua gen rujukan yang telah disahkan tersebut. Keputusan keseluruhan menunjukkan bahawa pengekspresan HGGT dan HPT sawit telah dikawalatur dalam tisu dan peringkat perkembangan. HPT sawit diekspreskan secara konstitutif dalam semua tisu kecuali kernel (15 m.s.a) manakala gen HGGT sawit hanya dapat dikesan dalam tisu tertentu, iaitu mesokarpa dan kernel serta menunjukkan tahap pengekspresan yang tinggi semasa pemendapan minyak berlaku secara aktif dalam mesokarpa (17 m.s.a). Ini menunjukkan bahawa pengekspresan HGGT dikawal oleh proses sintesis minyak dalam buah sawit. Akhir sekali, jujukan promoter HGGT sawit telah berjaya diampifikasikan menggunakan teknik “genome-walking PCR”. Pencarian menggunakan pangkalan data

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PLACE, PlantCARE serta PlantPAN, telah mengenalpasti beberapan elemen cis-pengawalatur yang mungkin terabit dalam mengkoordinasi pengekspresan. Secara am motif tersebut dapat dikategorikan kepada empat kumpulan utama termasuk yang respon terhadap fitohormon, cahaya, faktor abiotik dan endosperm spesifik. Ini menunjukkan bahawa pengawalaturan transkripsi HGGT dalam *E. guineensis* dan *E. oleifera* melibatkan banyak faktor yang sama. Pencirian lanjut untuk motif penting yang berpotensi akan memudahkan pemahaman yang lebih baik mengenai mekanisme pengawalaturan sintesis tokotrienol dalam kelapa sawit pada peringkat molekul.





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---

I certify that a Thesis Examination Committee has met on 8 July 2013 to conduct the final examination of Kong Sze Ling on her thesis entitled “Molecular Cloning, Characterization and Promoter Analysis of Vitamin E Biosynthetic Genes from the Oil Palm” in accordance with the Universities and University College Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A). 106] 15 March 1998. The committee recommends that the student be awarded the Master of Science.

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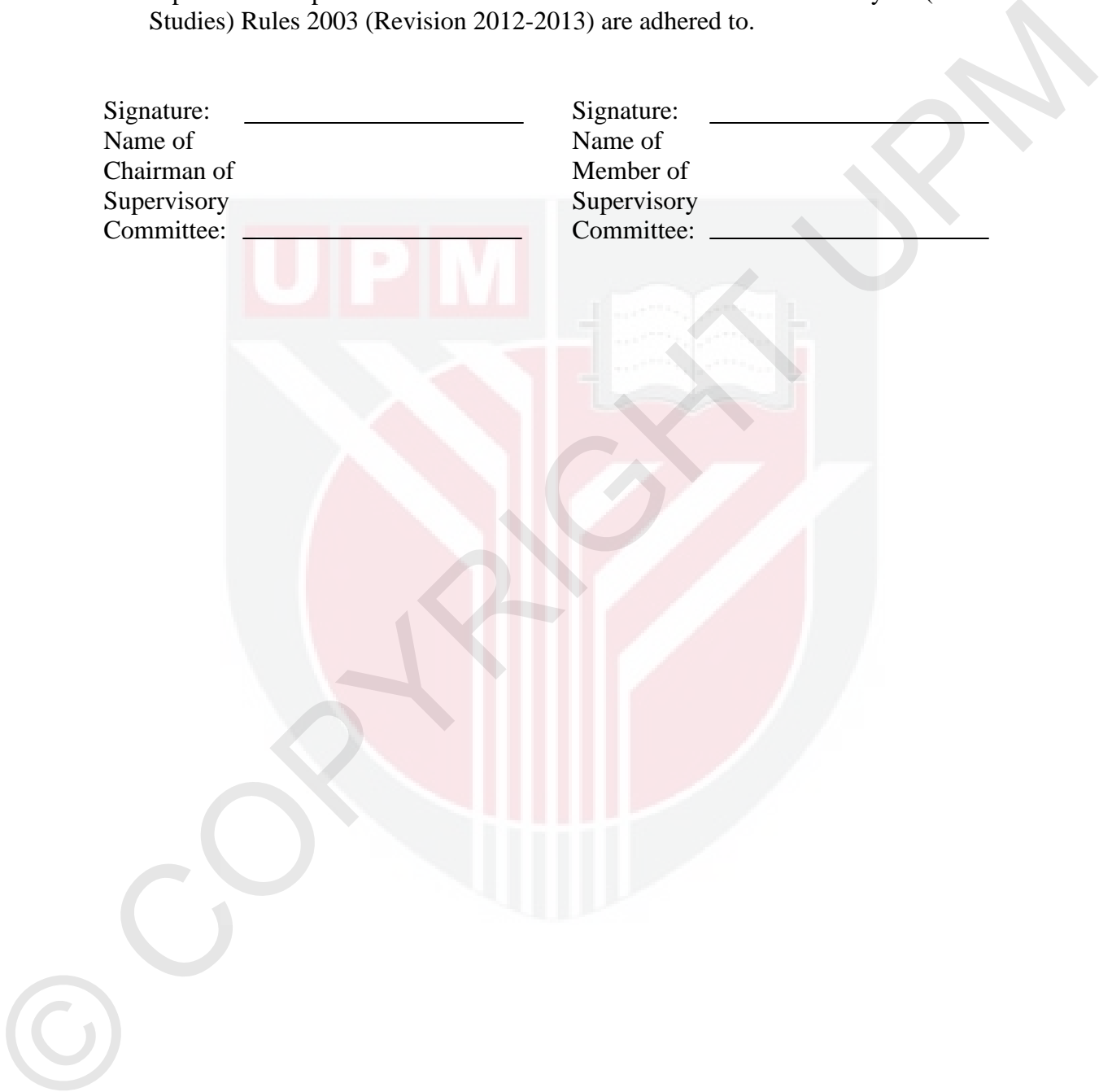
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## TABLE OF CONTENTS

<b>ABSTRACT</b>	<b>Page</b>
<b>ABSTRAK</b>	i
<b>ACKNOWLEDGEMENTS</b>	iii
<b>APPROVAL</b>	v
<b>DECLARATION</b>	vi
<b>LIST OF TABLES</b>	viii
<b>LIST OF FIGURES</b>	xiii
<b>LIST OF ABBREVIATIONS</b>	xiv
	xvii

### CHAPTER

<b>1</b>	<b>INTRODUCTION</b>	<b>1</b>
<b>2</b>	<b>LITERATURE REVIEW</b>	
	The Fruit of Oil Palm	3
	The Composition of Palm Oil	4
	Vitamin E	6
	Chemical Structure and Distribution	6
	Beneficial Properties	8
	Tocotrienols Beyond Tocopherols	9
	Biosynthetic Pathway of Vitamin E	10
	Modification of Vitamin E Contents in Plants	13
	Plant Gene Promoters	14
	Genome Walking for Promoter Isolation	15
	Real-time PCR for Gene Expression Analysis	16
<b>3</b>	<b>MATERIALS AND METHODS</b>	
	Plant Materials	18
	Total RNA Extraction	18
	RNA Quantification	19
	Removal of Genomic DNA from Total RNA	19
	Partial HGGT and HPT Genes Isolation	
	Messenger RNA (mRNA) Isolation	19
	First Strand cDNA Synthesis	20
	Degenerate Primers Design	20
	Primary RT-PCR Amplification	21
	Secondary RT-PCR Amplification	21
	Purification of the Expected Product	22
	Preparation of Competent Cells	22
	Ligation of PCR Product into Vector	23
	Transformation of <i>E. coli</i>	23
	Colony PCR	23

Plasmid DNA Miniprep	24
Screening for the Recombinant Plasmids	24
Sequencing Analysis of the Partial Gene Sequences	25
Isolation of partial <i>EoHGGT</i> using gene specific primers	25
Isolation of the Full Length cDNAs of HGGT and HPT	26
Gene Specific Primer Design	26
First Strand cDNA Synthesis	26
Rapid Amplification of cDNA Ends (RACE)	28
Sequencing Analysis of RACE PCR Products	28
Long-Distance PCR (LD-PCR)	29
Full Length cDNA Sequences Analysis	29
Gene Expression Analysis of HGGT and HPT Genes	
Primer Design	30
Determination of the Amplification Efficiency	30
Reference Genes Selection	31
RT-qPCR Analysis	31
Construction of Oil Palm Genome Walker Libraries	
Genomic DNA Extraction	32
Digestion of Genomic DNA	33
DNA Purification	33
Ligation to GenomeWalker™ Adaptors	33
Primer Design for Genome Walking	33
Primary Genome Walking	34
Secondary Genome Walking	34
LD-PCR	36
<i>In Silico</i> Promoter Analysis	36
<b>4 RESULTS</b>	
Total RNA Extraction	37
Design of Degenerate Primers	37
Isolation of the Full Length <i>EgHGGT</i> cDNA Sequence	
Partial Genes Isolation	41
5' and 3' - RACE PCR	46
End-to-End PCR for <i>EgHGGT</i>	46
<i>EgHGGT</i> cDNA Sequence Analysis	50
Isolation of the Full Length <i>EgHPT</i> and <i>EoHPT</i> cDNA Sequences	
Partial Gene Isolation	50
5' and 3' RACE PCR	53
End-to-End PCR for <i>EgHPT</i> and <i>EoHPT</i>	53
<i>EgHPT</i> and <i>EoHPT</i> cDNA Sequences Analysis	57
Isolation of the Full Length <i>EoHGGT</i> cDNA Sequence	
Partial Gene Isolation	57
5' and 3' RACE PCR	59
End-to-End PCR for <i>EoHGGT</i>	59
<i>EoHGGT</i> cDNA Sequence Analysis	65
Comparison of HGGT and HPT from Two Oil Palm Species	65

	Expression Analysis of Oil Palm Vitamin E Biosynthetic Genes	
	Optimization of Real-Time PCR Assays	67
	Selection of Suitable Reference Genes	73
	Relative Quantification of Oil Palm HGGT and HPT Genes	77
	Isolation of the Oil Palm HGGT Promoters	
	Genomic DNA Extraction	77
	Construction of GenomeWalker Libraries	82
	GenomeWalking PCR	82
	LD-PCR	86
	Analysis of <i>EgHGGT</i> and <i>EoHGGT</i> Promoter <i>cis</i> -regulatory Elements	89
<b>5</b>	<b>DISCUSSION</b>	92
	Selection of Tissues for Source of RNA	92
	RT-PCR Using Degenerate Primers	93
	Full Length Oil Palm HGGT and HPT cDNAs Sequence Analysis	94
	Selection of Suitable Reference Genes for Expression Studies	95
	Expression Analysis of Oil Palm HGGT and HPT Genes	96
	<i>In silico</i> Analysis of <i>EgHGGT</i> and <i>EoHGGT</i> Promoters	97
	Future Studies	99
<b>6</b>	<b>CONCLUSION</b>	101
	<b>REFERENCES</b>	103
	<b>APPENDICES</b>	115
	<b>BIODATA OF THE STUDENT</b>	121
	<b>LIST OF PUBLICATIONS</b>	122



---

## LIST OF TABLES

Table		Page
2.1	Ranges in content for various components in the unsaponifiable fraction from palm oil.	5
2.2	The Structures and Chemical Names of the Tocopherols and Tocotrienols.	7
3.1	List of GSPs for RACE PCR.	27
3.2	List of GSP1 and GSP2 used in the Genome Walking PCR.	35
4.1	Spectrophotometric measurement of the total RNA extracted from various tissues of both oil palm species and treated with DNase.	40
4.2	Degenerate primers designed for RT-PCR amplification of oil palm vitamin E biosynthesis genes.	43
4.3	The differences in nucleotide base that lead to the changes in amino acid sequences between <i>EgHGGT</i> and <i>EoHGGT</i> .	66
4.4	The differences in nucleotide base that lead to the changes in amino acid sequences between <i>EgHPT</i> and <i>EoHPT</i> .	68
4.5	List of primers specific for oil palm HGGT, HPT, $\beta$ -actin (ACT), cyclophilin (CYP) and tubulin (TUB) for the quantitative PCR assays.	72
4.6	Ranking of the candidate reference genes in each sample set according to their stability value (M value) using geNorm analysis.	76
4.7	List of <i>cis</i> -regulatory elements found in both oil palm HGGT and HPT promoters where (+) is calculated from the positive strand and (-) is calculated from the negative strand based on the location of the TSS.	90

---

## LIST OF FIGURES

Figure		Page
2.1	Biosynthesis pathway of vitamin E.	11
4.1	Total RNA extracted from different developmental stages of oil palm tissues analyzed on 1% (w/v) agarose gel.	38
4.2	DNase treated total RNA from different developmental stages of mesocarp tissues.	39
4.3	DNase I treated total RNA from different oil palm tissues.	39
4.4	Identification of the conserved regions within the plant HGGT amino acid sequences for degenerate primers synthesis.	42
4.5	Relative location of the degenerate primers to the cDNA sequence of <i>Oryza sativa</i> HGGT gene (Accession # AY222862).	43
4.6	Primary and Secondary RT-PCR amplification of the partial cDNA encoded for <i>E. guineensis</i> HGGT using degenerate primers.	44
4.7	Complete cDNA sequence (717 bp) of fragment encoding the middle region of <i>E. guineensis</i> HGGT.	45
4.8	5'- and 3'- RACE PCR amplification of the <i>E. guineensis</i> HGGT using combination of a gene specific primer and an adaptor primer.	47
4.9	The 1853 bp of consensus cDNA sequence of <i>EgHGGT</i> generated by assembling the 5'-end, middle and 3'-end regions.	48
4.10	End-to-end RT-PCR amplification of the coding region of <i>E. guineensis</i> HGGT using gene specific primers.	49
4.11	RT-PCR amplification of the partial cDNA encoded for <i>E. oleifera</i> HPT using degenerate primers.	51
4.12	Complete cDNA sequence (717 bp) of fragment encoding the middle region of <i>E. oleifera</i> HPT.	52
4.13	RACE PCR products of oil palm HPT gene amplified from 17 w.a.a mesocarp tissues.	54

---

4.14	The 1732 bp of consensus cDNA sequence of <i>EoHPT</i> generated by assembling the 5'-end, middle and 3'-end regions.	55
4.15	The 1762 bp of consensus cDNA sequence of <i>EgHPT</i> generated by assembling the 5'-end, middle and 3'-end regions.	56
4.16	End-to-end PCR amplification of the coding region for <i>EgHPT</i> and <i>EoHPT</i> from <i>E. guineensis</i> and <i>E. oleifera</i> 17 w.a.a mesocarp cDNA, respectively.	58
4.17	Primary and Secondary RT-PCR amplification of the partial cDNA encoded for <i>E. oleifera</i> HGGT using gene specific primers.	60
4.18	Complete cDNA sequence (565 bp) of fragment encoding the middle region of <i>E. oleifera</i> HGGT.	61
4.19	5'-RACE and 3'-RACE PCR products amplified from <i>E. oleifera</i> mesocarp at 17 w.a.a.	62
4.20	The 1732 bp of consensus cDNA sequence of <i>EoHGGT</i> generated by assembling the 5'-end, middle and 3'-end regions.	63
4.21	End-to-end PCR amplification of the coding region for <i>EoHGGT</i> from 17 w.a.a mesocarp cDNA.	64
4.22	Sequence alignment of 3'UTR regions for <i>EgHPT</i> and <i>EoHPT</i> using ClustalW program.	68
4.23	Identification of a highly conserved region across oil palm HGGT amino acid sequences and their homologs using ClustalW alignment tool.	69
4.24	Identification of two highly conserved regions across oil palm HPT amino acid sequences and their homologs using ClustalW alignment tool.	70
4.25	Phylogenetic relationship between the derived amino acid sequences of the oil palm HGGT and HPT with other plants and cyanobacterias.	71
4.26	PCR efficiency test for real-time PCR assays of oil palm $\beta$ -actin (ACT), cyclophilin (CYP), tubulin (TUB), HGGT and HPT gene.	74
4.27	An overlay of melting curve derivative profile following each real-time assay for oil palm $\beta$ -actin (ACT), cyclophilin (CYP), tubulin (TUB), HGGT and HPT gene.	75

---

4.28	Relative abundances of <i>EgHGGT</i> in <i>E. guineensis</i> developing mesocarp tissues of different developmental stages (EGM7-EGM19).	78
4.29	Relative abundances of <i>EoHGGT</i> in <i>E. oleifera</i> developing mesocarp tissues of different developmental stages (EOM7-EOM19).	78
4.30	Relative abundances of <i>EgHPT</i> in <i>E. guineensis</i> developing mesocarp tissues of different developmental stages (EGM7-EGM19).	79
4.31	Relative abundances of <i>EoHPT</i> in <i>E. oleifera</i> developing mesocarp tissues of different developmental stages (EOM7-EOM19).	79
4.32	Relative abundances of <i>EgHGGT</i> in <i>E. guineensis</i> 15 w.a.a kernel (EGK15), spear leaves (EGL) and young root (EGR); <i>EoHGGT</i> in <i>E. oleifera</i> 15 w.a.a kernel (EOK15); <i>EgHPT</i> in EGK15, EGL and EGR and <i>EoHPT</i> in EOK15.	80
4.33	Genomic DNA extracted from spear leaves of <i>E. guineensis</i> and mesocarp of <i>E. oleifera</i> .	81
4.34	Analysis of constructed oil palm ( <i>Elaeis guineensis</i> ) GenomeWalker libraries on 0.6% agarose gel.	83
4.35	Agarose gel electrophoresis analysis of (a) Primary and (b) nested genome walking PCR amplification product of <i>EgHGGT</i> 5' upstream region.	84
4.36	Second attempt of (a) primary and (b) nested genome walking PCR amplification of <i>EgHGGT</i> 5' upstream region.	85
4.37	Third attempt of genome walking PCR amplification of <i>EgHGGT</i> 5' upstream region.	87
4.38	End-to-end PCR amplification of the 5' upstream region for <i>EgHGGT</i> and <i>EoHGGT</i> from <i>E. guineensis</i> and <i>E. oleifera</i> genomic DNA, respectively.	87
4.39	Nucleotide sequence of the <i>EgHGGT</i> promoter region.	88
4.40	Nucleotide sequence of the <i>EoHGGT</i> promoter region.	88

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## LIST OF ABBREVIATIONS

AP	Adaptor Primer
BLAST	Basic Local Alignment Search Tool
bp	base pair
CaCl <sub>2</sub>	calcium chloride
cDNA	complementary deoxuribonucleic acid
Ct	threshold cycle
CTAB	hexadecyl (or cetyl) trimethyl ammonium bromide
Da	Dalton
DNA	deoxyribonucleic acid
DNase I	deoxyribonuclease I
dNTP	deoxynucleoside triphosphate
EDTA	ethylene diamine tetracetate
<i>E. coli</i>	<i>Escherichia coli</i>
GA	gibberillin
GAPDH	glyceraldehydes-3-phosphate dehydrogenase
GGDP	geranylgeranyldiphosphate
GGPP	geranylgeranyl pyrophosphate
GSP	gene-specific primer
HCl	hydrochloric acid
HGA	homogentisic acid
HGGT	homogentisate geranylgeranyl transferase
HPP	p-hydroxyphenylpyruvate

HPPDase	p-hydroxyphenylpyruvate dioxygenase
HPT	homogentisate phytyltransferase
IPTG	Isopropyl $\beta$ -D-1-thiogalactopyranoside
kb	kilobase
LB	Luria-Bertani
LiCl	lithium chloride
M	molar
MgCl <sub>2</sub>	magnesium chloride
min	minutes
mRNA	messenger ribonucleic acid
NaCl	sodium chloride
NaOAc	sodium acetate
ng	nanogram
NCBI	National Center for Biotechnology Information
OD	optical density
ORF	open reading frame
PCR	polymerase chain reaction
PDP	phytyldiphosphate
phytyl-PP	phytyl pyrophosphate
pI	isoelectric point
PrDP	prenyldiphosphate
PVP-40	polyvinylpyrrolidone-40
RACE	Rapid Amplification of cDNA End

RNA	ribonucleic acid
$R^2$	correlation coefficient
RT-PCR	reverse transcription PCR
SDS	Sodium dodecyl sulfate
sec	seconds
SNP	Single nucleotide polymorphism
TAE	tris-acetate-EDTA
TC	tocopherol/tocotrienol cyclase
TE	Tris-EDTA
TG	triacylglycerols
TMT	tocopherol/tocotrienol methyltransferase
TRF	tocotrienol-rich fraction
TSS	transcription start site
UTR	untranslated region
v/v	volume per volume
w/v	weight per volume
w.a.a	week after anthesis
$\mu\text{g}$	microgram
$\mu\text{M}$	micromolar
$\mu\text{l}$	microliter
$g$	relative centrifugal force
$^{\circ}\text{C}$	Degree Celsius

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## CHAPTER 1

### INTRODUCTION

Since it was first planted as a commercial crop in 1917, oil palm cultivation has shown rapid expansion and it is now the main commodity crop of Malaysia. The palm oil industry has contributed significantly to Malaysia economic development and foreign exchange earnings. Palm oil is presently the world's major source of vegetable oil and Malaysia is second only to Indonesia as the world leading exporter of palm oil (CME Group, 2010).

As the world population increases, the demand from the oil palm industry also increases. Among the immediate challenges is the decrease in land availability and labour shortage. Besides, Malaysian palm oil industry also face great competition from other palm oil producing countries especially Indonesia. Thus, appropriate strategies need to be planned in order to ensure agricultural sustainability and to stay competitive in the future. An effective approach is to improve the oil yield per unit area of land with the view of maximizing returns. In addition, improvement of palm oil quality has also been set as one of the priority areas for oil palm research. Oil palm, being naturally rich in the minor components such as carotenoids and vitamin E, offers a great potential to be exploited as a value-added vegetable oil which is an important advantage over other vegetable oils and fats.

Tocochromanols, commonly known as vitamin E, play a crucial role in human and animal nutrition. Belonging to the amphipatic tocochromanol group of molecules, the eight structurally related tocopherols and tocotrienols forms ( $\alpha$ -,  $\beta$ -,  $\gamma$ -  $\delta$ - tocopherols and  $\alpha$ -,  $\beta$ -,  $\gamma$ -  $\delta$ -tocotrienols) collectively constitute the content of vitamin E (Kamal-Eldin and Appelqvist, 1996). Tocotrienols in vitamin E have been reported to possess powerful antioxidant and anti-cancer activities (Ebong *et al.*, 1999). Moreover, palm tocotrienols exhibit anti-angiogenic properties that may inhibit tumour progression (Selvaduray *et al.*, 2012). Besides contributing to human health, tocochromanols are also linked with a number of beneficial properties for cereals which include extending their storage life and contributing to the nutritive value of cereal grains in human and livestock diets.

Oil palm is one of the richest source of vitamin E especially tocotrienols which are not normally present in other edible oil. Crude palm oil extracted from the fruits of *Elaeis guineensis* particularly contains a high amount of tocotrienols (up to 800 mg/kg), mainly consisting of  $\gamma$ -tocotrienol and  $\alpha$ -tocotrienol (Sen *et al.*, 2006). While *E. oleifera* also been reported to contain significantly higher amount of tocotrienol (Choo & Yusof, 1996). Clearly, it has great advantage compared to other plants for genetic manipulation of vitamin E. However, the knowledge on oil palm vitamin E biosynthesis pathway



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which is one of the basic requirements for genetic manipulation is quite limited. This will definitely become an impediment to improve oil palm vitamin E content through genetic engineering, development of molecular markers for cross species breeding and other biotechnological approaches. This work is an initial effort towards the understanding of oil palm vitamin E biosynthetic pathway. This includes molecular characterization of the cDNAs encoding homogentisate geranylgeranyl transferase (HGGT) and homogentisate phytyltransferase (HPT), that catalyse the first committed step in tocotrienol and tocopherol production, respectively. The promoter sequences which regulate the expression of these genes will certainly contribute to the basic platform required for the oil palm vitamin E content and composition improvement.

Therefore the objectives of this study are

1. To isolate and characterize full length cDNA sequences encoding homogentisate geranylgeranyl transferase (HGGT) and homogentisate phytyltransferase (HPT) from oil palm (*E. guineensis* and *E. oleifera*).
2. To characterize the transcript expression profile of HGGT and HPT in different oil palm tissues and at different developmental stages through real-time quantitative PCR method in both oil palm species.
3. To isolate and analyze the promoter region of oil palm HGGT and to compare the presence of known *cis*-acting regulatory elements in both promoters.

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## REFERENCES

- Abe, H., Yamaguchi-Shinozaki, K., Urao, T., Iwasaki, T., Hosokawa, D., and Shinozaki, K. 1997. Role of Arabidopsis MYC and MYB Homologs in Drought- and Abscisic Acid-regulated Gene Expression. *Plant Cell* 9: 1859-1868.
- Agarwal, M.K., Agarwal, M.L., Athar, M. and Gupta, S. 2004. Tocotrienol-rich Fraction of Palm Oil Activates p53, Modulates Bax/Bcl2 Ratio and Induces Apoptosis Independent of Cell Cycle Association. *Cell Cycle* 3: 1-7.
- Andersson, S.C., Rumpunen, K., Johansson, E. and Olsson, M.E. 2008. Tocopherols and Tocotrienols in Sea Buckthorn (*Hippophae rhamnoides L.*) Berries during Ripening. *Journal of Agricultural and Food Chemistry* 56: 6701-6706.
- Arango, Y. and Heise, K.P. 1997.  $\alpha$ -Tocopherol Synthesis by Capsicum Fruit Chromoplasts. *Journal of Plant Physiology* 150: 509-513.
- Ashida, O. 2010. Studies on the Transcriptional Regulation of Fatty Acid Biosynthesis in Oil Palm Fruits using Stearoyl-ACP Desaturase & Acyl Carrier Protein Genes. Unpublished master thesis. University Putra Malaysia.
- Bafor, M.E. and Osaige, A.U. 1988. Changes in Non-polar Lipid Composition of Developing Oil Palm Fruit (*Elaeis guineensis*) Mesocarp. *Journal of the Science of Food and Agriculture* 46(4): 325-331.
- Bramley, P.M., Elmadfa, I., Kafatos, A., Kelly, F.J., Manios, Y., Roxborough, H.E., Schuch, W., Sheehy, P.J. A. and Wagner, K.H. 2000. Review-Vitamin E. *Journal of the Science of Food and Agriculture* 80: 913-938.
- Breathnach, R. and Chambon, P. 1981. Organization and Expression of Eucaryotic Split Genes Coding for Proteins. *Annual Review of Biochemistry* 50: 349-383.
- Buchan, D.W., Ward, S.M., Lobley, A.E., Nugent, T.C., Bryson, K. and Jones, D.T. 2010. Protein Annotation and Modelling Servers at University College London. *Nucleic Acids Research* 38: W563-W568.
- Bustin, S.A. 2000. Review-Absolute Quantification of mRNA using Real-Time Reverse Transcription Polymerase Chain Reaction Assays. *Journal of Molecular Endocrinology* 25: 169-193.
- Bustin, S.A., Benes, V., Garson, J.A., Hellemans, J., Huggett, J., Kubista, M., Mueller, R., Nolan, T., Pfaffl, M.W., Shipley, G.L., Vandesompele, J. and Wittwer, C.T. 2009. The MIQE Guidelines: Minimum Information for Publication of Quantitative Real-Time PCR Experiments. *Clinical Chemistry* 55(4): 611-622.

- 
- Bustin, S.A. and Nolan, T. 2004. Pitfalls of Quantitative Reverse transcription Polymerase Chain Reaction. *Journal of Biomolecular Techniques* 5: 155-166.
- Butler, J.E.F. and Kadonaga, J.T. 2002. The RNA Polymerase II Core Promoter: A Key Component in the Regulation of Gene Expression. *Genes Development* 16: 2583-2592.
- Cahoon, E.B., Hall, S.E., Ripp, K.G., Ganzke, T.S., Hitz, W.D., and Coughlan, S.J. 2003. Metabolic Redesign of Vitamin E Biosynthesis in Plants for Tocotrienol Production and Increased Antioxidant Content. *Nature Biotechnology* 21(9): 1082-1087.
- Capell, T. and Christou, P. 2004. Progress in Plant Metabolic Engineering. *Current Opinion in Biotechnology* 15: 148-154.
- Chan, P.L., Siti Nor Akmar, A. and Roohaida, O. 2008. Light-harvesting Chlorophyll a/b Binding Protein (Lhcb) Promoters for Targeting Specific Expression in Oil Palm Leaves. *Journal of Oil Palm Research Special Issue on Malaysia-MIT Biotechnology Partnership Programme: Oil Palm Metabolic Engineering* pp 21-29.
- Chang, W.C., Lee, T.Y., Huang, H.D., Huang, H.Y. and Pan, R.L. 2008. "PlantPAN: Plant Promoter Analysis Navigator, for Identifying Combinatorial *cis*-regulatory Elements with Distance Constraint in Plant Gene Group". *BMC Genomics* 9: 561.
- Chaudhary, N and Khurana, P. 2009. Vitamin E Biosynthesis Genes in Rice: Molecular Characterization, Expression Profiling and Comparative Phylogenetic Analysis. *Plant Science* 177: 479-491.
- Chen, S.Y., Li, H.J. and Liu, G.S. 2006. Progress of Vitamin E Metabolic Engineering in Plants. *Transgenic Research* 15: 655-665.
- Chen, Y.T., Lee, Y.R., Yang, C.Y., Wang, Y.T., Yang, S.F. and Shaw, J.F. 2003. A Novel Papaya ACC oxidase Gene (CP-ACO2) Associated with Late Stage Fruit Ripening and Leaf Senescence. *Plant Science* 164:531-540.
- Choo, Y.M. and Yusof, B. 1996. *Elaeis oleifera* Palm for the Pharmaceutical Industry. PORIM Information Series No. 22: 4 pp.
- Choo, Y.M., Ma, A.N., Chuah, C.H., Khor, H.T. and Bong, S.C. 2004. A Developmental Study on the Appearance of Tocopherols and Tocotrienols in Developing Palm Mesocarp (*Elaeis guineensis*). *Lipids* 39 (6):561-564.
- Collakova, E. and DellaPenna, D. 2001. Isolation and Functional Analysis of Homogentisate Phytyltransferase from *Synechocystis* sp. PCC 6803 and *Arabidopsis*. *Plant Physiology* 127: 1113-1124.

- 
- Collakova, E. and DellaPenna, D. 2003. Homogentisate Phytyltransferase Activity is Limiting for Tocopherol Biosynthesis in Arabidopsis. *Plant Physiology* 13: 632–642.
- Corley, R.H.V. and Tinker, P.B. 2003. The Oil Palm. 4<sup>th</sup> edition. USA: Blackwell Publishing.
- Debeaujon, I. and Koornneef, M. 2000. Gibberellin Requirement for Arabidopsis Seed Germination Is Determined Both by Testa Characteristics and Embryonic Abscisic Acid. *Plant Physiology* 122(2): 415-424.
- Dereeper, A., Audic, S., Claverie, J.M. and Blanc, G. 2010. BLAST-EXPLORER helps you Building Datasets for Phylogenetic Analysis. *BMC Evolutionary Biology* 10: 8.
- Dereeper, A., Guignon, V., Blanc, G., Audic, S., Buffet, S., Chevenet, F., Dufayard, J.F., Guindon, S., Lefort, V., Lescot, M., Claverie, J.M. and Gascuel, O. 2008. Phylogeny.fr: Robust Phylogenetic Analysis for the Non-specialist. *Nucleic Acids Research* 36: W465-9.
- Dörmann, P. 2003. Corn with Enhanced Antioxidant Potential. *Nature biotechnology* 21(9): 1015-1016.
- Dörmann, P. 2007. Functional Diversity of Tocochromanols in Plants. *Planta* 225:269-276.
- Doyle, J.J. and Doyle, J.L. 1990. Isolation of Plant DNA from Fresh Tissues. *FOCUS* 12: 13-15.
- Ebong, P.E., Owu, D.U. and Isong, E.U. 1999. Influence of Palm Oil (*Elaeis guineensis*) on Health. *Plant Foods for Human Nutrition* 53(3): 209-222.
- Edem, D.O. 2002. Palm oil: Biochemical, Physiological, Nutritional, Hematological, and Toxicological Aspects: A Review. *Plant Foods for Human Nutrition* 57: 319-341.
- Ellerström, M., Sffdberg, K., Ezcurra, I. and Rask, L. 1996. Functional Dissection of a Napin Gene Promoter: Identification of Promoter Elements required for Embryo and Endosperm-Specific Transcription. *Plant Molecular Biology* 32: 1019-1027.
- Food and Nutrition Board, Institute of Medicine. 2000. Vitamin E. In “Dietary Reference Intakes for Vitamin C, Vitamin E, Selenium and Carotenoids”. National Academic Press, Washington, DC pp. 186-283.
- Gao, X., Jackson, T.A., Lambert, K.N. and Li, S. 2004. Detection and Quantification of *Fusarium solani* f. sp. *glycines* in Soybean Roots with Real-Time Quantitative Polymerase Chain Reaction. *Plant Disease* 88(12): 1372-1380.

- 
- Goh, S.H., Choo, Y.M. and Ong, S.H. 1985. Minor Constituents of Palm Oil. *Journal of the American Oil Chemists' Society* 62 (2): 237-240.
- Gubler, F. and Jacobsen, J.V. 1992. Gibberellin-Responsive Elements in the Promoter of a Barley High-pl  $\alpha$ -Amylase Gene. *The Plant Cell* 4: 1435-1441.
- Guo, J., Liu, G.S., Chen, S.Y. and Amina, A.A. 2009. Vitamin E Metabolic Modulation in Plants. In "Herbal Drugs: Ethnomedicine to Modern Medicine" (Ed. Ramamwat, K. G.) pp. 333-352.
- Guthrie, N., Gapor, A., Chambers, A.F. and Carroll, K.K. 1997. Inhibition of Proliferation of Estrogen Receptor-Negative MDA-MB-435 and -Positive MCF-7 Human Breast Cancer Cells by Palm Oil Tocotrienols and Tamoxifen, Alone and in Combination. *The Journal of Nutrition* 544S-548S.
- Gutierrez, L., Mauriat, M., Gue ñin, S., Pelloux, J., Lefebvre, J.F., Louvet, R., Rusterucci, C., Moritz, T., Guerneau, F., Bellini, C. and Van Wuytswinkel, O. 2008. The Lack of a Systematic Validation of Reference Genes: A Serious Pitfall Undervalued in Reverse Transcription Polymerase Chain Reaction (RT-PCR) Analysis in Plants. *Plant Biotechnology* 6: 609-618.
- Han, X.J., Lu, M.Z., Chen, Y.C., Zhan, Z.Y., Cui, Q.Q. and Wang, Y.D. 2012. Selection of Reliable Reference Genes for Gene Expression Studies Using Real-Time PCR in Tung Tree during Seed Development. *PLOS ONE* 7(8).
- Hartley, C. W. S. 1988. *The Oil Palm*. 3<sup>rd</sup> edition. New York: Longman.
- Harwood, J. and Page, R.A. 1994. Biochemistry of Oil Synthesis. In "Designer Oil Crops" (Eds. Murphy, D. J.) VCH Weinheim. pp. 165-194.
- Havaux, M., Eymery, F., Porfirova, S., Rey, P. and Dörmann, P. 2005. Tocochromanol Protects against Photo Inhibition and Oxidative Stress in *Arabidopsis thaliana*. *Plant Cell* 17: 3451-3469.
- Higo, K., Ugawa, Y., Iwamoto, M. and Korenaga, T. 1999. Plant *cis*-acting Regulatory DNA Elements (PLACE) Database. *Nucleic Acids Research* 27(1): 297-300.
- Hong, S.Y., Seo, P.J., Yang, M.S., Xiang, F. and Park, C.M. 2008. Exploring Valid Reference Genes for Gene Expression Studies in *Brachypodium distachyon* by Real-Time PCR. *BMC Plant Biology* 8: 112.
- Horton, P., Park, K.J., Obayashi, T. and Nakai, K. 2006. Protein Subcellular Localization Prediction with WoLF PSORT. Proceedings of the 4th Annual Asia Pacific Bioinformatics Conference APBC06, Taipei, Taiwan. pp 39-48.

- 
- Horton, P., Park, K.J., Obayashi, T., Fujita, N., Harada, H., Adams-Collier, C.J. and Nakai, K. 2007. WoLF PSORT: Protein Localization Predictor. *Nucleic Acids Research* 35: W585-7.
- Horvath, G., Wessjohann, L., Bigirimana, J., Jansen, M., Guisez, Y., Caubergs, R., Horemans, N. 2006. Differential Distribution of Tocopherols and Tocotrienols in Photosynthetic and Non-photosynthetic Tissues. *Phytochemistry* 67: 1185–1195.
- Hudson, M.E. and Quail, P.H. 2003. Identification of Promoter Motifs Involved in the Network of Phytochrome A-Regulated Gene Expression by Combined Analysis of Genomic Sequence and Microarray Data. *Plant Physiology* 133: 1605-161.
- Hunter, S.C. and Cahoon, E.B. 2007. Enhancing Vitamin E in Oilseeds: Unraveling Tocopherol and Tocotrienol Biosynthesis. *Lipids* 42: 97–108.
- Jalani, B.S., Cheah, S.C., Rajanaidu, N. and Darus, A. 1997. Improvement of Oil Palm through Breeding and Biotechnology. *Journal of the American Oil Chemists' Society* 74: 1451-1455.
- Jeong, M.J. and Shih, M.C. 2003. Interaction of a GATA factor with *cis*-acting Elements involved in Light Regulation of Nuclear Genes encoding Chloroplast Glyceraldehyde-3-phosphate Dehydrogenase in Arabidopsis. *Biochemical and Biophysical Research Communications* 300: 555-562.
- Jones, D.T. 1999. Protein Secondary Structure Prediction based on Position-specific Scoring Matrices. *Journal of Molecular Biology* 292: 195-202.
- Joshi, C.P. 1987. An inspection of the domain between putative TATA box and translation start site in 79 plant genes. *Nucleic Acids Research* 15(16): 6643-6653.
- Kamal-Eldin, A. and Appelqvist, L.A. 1996. The Chemistry and Antioxidant Properties of Tocopherols and Tocotrienols. *Lipids* 31(7): 671-701.
- Karunanandaaa, B., Qi, Q., Hao, M., Baszis, S.R., Jensen, P.K., Wong, Y.H., Jiang, J., Venkatramesh, M., Gruys, K. J., Moshiri, F., Post-Beittenmiller, D., Weiss, J.D. and Valentin, H.E. 2005. Metabolically Engineered Oilseed Crops with Enhanced Seed Tocopherol. *Metabolic Engineering* 7:384-400.
- Kato, A., Yamaoka, M., Gapor, A. and Berger, K.G. 2002. Tocopherols of Oil Palm Leaf. *Journal of the American Oil Chemists' Society* 60(12).
- Khoo, K.M., Belvinder, K.S., Chandram, M.R., Chew, P.S., Chong, G.G., Mohd Yusof, H. 2005. Malaysian Palm Oil- A Success Story. 1<sup>st</sup> edition. Malaysian Palm Oil Promotion Council and Trans-Event Sdn Bhd.

- 
- Kusnetsov, V., Landsberger, M., Meurer, J. and Oelmüller, R. 1999. The Assembly of the CAAT-box Binding Complex at a Photosynthesis Gene Promoter Is Regulated by Light, Cytokinin and the Stage of the Plastids. *The Journal of Biological Chemistry* 274(50): 36009-36014.
- Laila, N., Ho, C.L., Tan, S.G. Yusuf, U.K. and Abdullah, F. 2011. Cloning of Transcripts Encoding Chitinases from *Elaeis guineensis Jacq.* and Their Expression Profiles in response to Fungal Infections. *Physiological and Molecular Plant Pathology* 76: 96-103.
- Lata, C., Yadav, A. and Prasad, M. 2011. Role of Plant Transcription Factors in Abiotic Stress Tolerance. In “Abiotic Stress Response in Plants - Physiological, Biochemical and Genetic Perspectives” (Ed. Shanker, A.) pp. 269-295.
- Li, Q., Fan, C.M., Zhang, X.M. and Fu, Y.F. 2012. Validation of Reference Genes for Real-Time Quantitative PCR Normalization in Soybean Developmental and Germinating Seeds. *Plant Cell Report* 31(10): 1799.
- Livak, K.J. and Schmittgen, T.D. 2001. Analysis of Relative Gene Expression Data Using Real-Time Quantitative PCR and the  $2^{-\Delta\Delta C_T}$  Method. *Methods* 25: 402-408.
- Maroufi, A., Bockstaele, E.V. and Loose, M.D. 2010. Validation of Reference Genes for Gene Expression Analysis in Chicory (*Cichorium intybus*) using Quantitative Real-Time PCR. *BMC Molecular Biology* 11: 15.
- Mayes, S., Farah, H., Price, Z., MacDonald, D., Billotte, N. and Roberts, J. 2008. Molecular Research in Oil Palm, the Key Oil Crop for the Future. In “Plant Genetic and Genomics: Genomics of Tropical Crop Plants” (Eds. Moore, P. H. and Ming, R.) pp. 371-404.
- Mizushina, Y., Nakagawa, K., Shibata, A., Awata, Y., Kuriyama, I., Shimazaki, N., Koiwai, O., Uchyama, Y., Sakaguchi, K., Miyazawa, T. and Yoshida, H. 2006. Inhibitory Effect of Tocotrienol on Eukaryotic DNA Polymerase  $\lambda$  and Angiogenesis. *Biochemical and Biophysical Research Communications* 339: 949–955.
- Molina, C. and Grotewold, E. 2005. Genome Wide Analysis of Arabidopsis Core Promoters. *BMC Genomics* 6: 25.
- Munné-Bosch, S. and Alegre, L. 2002. The Function of Tocopherols and Tocotrienols in Plants. *Critical Reviews in Plant Sciences* 21: 31–57.
- Nakamura, M., Tsunoda, T. and Obokata, J. 2002. Photosynthesis Nuclear Genes Generally Lack TATA-boxes; A Tobacco Photosystem I Gene Responds to Light through an Initiator. *The Plant Journal* 29(1): 1-10.

- 
- Nesaretnam, K., Ambra, R., Selvaduray, K.R., Radhakrishnan, K.E., Reimann, K. and Virgili, F. 2004. Tocotrienol-rich Fraction from Palm Oil Affects Gene Expression in Tumors Resulting from MCF-7 Cell Inoculation in Athymic Mice. *Lipids* 39: 459–467.
- Nesaretnam, K., Dorasamy, S. and Dabre, D. 2000. Tocotrienol Inhibit Growth of ZR-75-1 Breast Cancer Cells. *International Journal of Food Science and Nutrition* 51: S95-S105.
- Nesaretnam, K., Guthrie, N., Chambers, A.F. and Carroll, K.K. 1995. Effect of Tocotrienols on the Growth of Human Breast Cancer Cell Line in Culture. *Lipids* 30: 1139-1143.
- Nesaretnam, K., Stephen, R., Dils, R. and Darbre, P. 1998. Tocotrienols Inhibit the Growth of Human Breast Cancer Cells Irrespective of Estrogen Receptor Status. *Lipids* 33: 461–469.
- Neves-Borges, A.C., Guimarães-Dias, F., Cruz, F., Mesquita, R.O., Nepomuceno, A.L., Romano, E., Loureiro, M.E, de Fátima Grossi-de-Sá M. and Alves-Ferreira, M. 2012. Expression Pattern of Drought Stress Marker Genes in Soybean Roots under Two Water Deficit Systems. *Genetics and Molecular Biology* 35(1): 212-21.
- Ogawa, M., Hanada, A., Yamauchi, Y., Kuwahara, A., Kamiya, Y. and Yamaguchi, S. 2003. Gibberellin Biosynthesis and Response during Arabidopsis Seed Germination. *The Plant Cell* 15: 1591-1604.
- Oo, K.C., Teh, S.K., Khor, H.T. and Ong, A.S.H. 1986. Fatty Acid Synthesis in the Oil Palm (*Elaeis guineensis*): Incorporation of Acetate by Tissue Slices of the Developing Fruit. *Lipids* 20 (4): 205-210.
- Packer, L., Weber, S.U. and Rimbach, G. 2001. Molecular Aspects of Alphanatocotrienol Antioxidant Action and Cell Signaling. *Journal of Nutrition* 131: 369S–373S.
- Park, S.C., Kwon, H.B. and Shih, M.C. 1996. Cis-Acting Elements Essential for Light Regulation of the Nuclear Gene Encoding the A Subunit of Chloroplast Glyceraldehyde-3-phosphate Dehydrogenase in *Arabidopsis thaliana*. *Plant Physiology* 112: 1563-1571.
- Pearce, B.C., Parker, R.A., Deason, M.E., Qureshi, A.A. and Wright, J.J. 1992. Hypocholesterolemic Activity of Synthetic and Natural Tocotrienols. *Journal of Medicinal Chemistry* 35(20): 3595–3606.
- Pfaffl, M.W. 2001. A New Mathematical Model for Relative Quantification in Real-Time RT-PCR. *Nucleic Acids Research* 29(9): e45.



- 
- Pfaffl, M.W. 2004. Quantification Strategies in Real-Time PCR. In “A-Z of Quantification PCR” (Ed. Bustin, S. A.) pp. 87-112.
- Pfaffl, M.W., Tichopad, A., Prgomet, C. and Neuvians, T. P. 2004. Determination of Stable Housekeeping Genes, Differently Regulated Target genes and Sample Integrity: BestKeeper - Excel-based Tool using Pair-Wise Correlations. *Biotechnology Letters* 26: 509-515.
- Piechulla, B., Merforth, N. and Rudolph, B. 1998. Identification of Tomato *Lhc* Promoter Regions necessary for Circadian Expression. *Plant Molecular Biology* 38:655-662.
- Prescott, A. and Martin, L. 1987. A Rapid Method for Quantitative Assessment of Levels of Specific mRNAs in Plants. *Plant Molecular Biology Reporter* 4: 219-224.
- Prestridge, D.S. 1991. SIGNAL SCAN: A Computer Program that Scans DNA Sequences for Eukaryotic Transcriptional Elements. *Computer Applications in Biosciences* 7: 203-206.
- Pujade-Renaud, V., Sanier, C., Cambillau, L., Pappusamy, A., Jones, H., Ruengsri, N., Tharreau, D., Chrestin, H., Montoro, P. and Narangajavana, J. 2005. Molecular Characterization of New Members of the *Hevea brasiliensis* Hevein Multigene Family and Analysis of their Promoter Region in Rice. *Biochimica et Biophysica Acta* 1727: 151-161.
- Purseglove, J.W. 1975. Tropical Crops Monocotyledons. London: Longman.
- Rasid, O.A., Wan Nur Syuhada, W.S., Nor Hanin, A., Masura, S.S., Zulqarnain, M., Ho, C. L., Sambanthamurthi, R. and Suhaimi, N. 2008. RT-PCR Amplification and Cloning of Partial DNA Sequence Coding for Oil Palm (*Elaeis oleifera*) Phytoene Synthase Gene. *Asia Pacific Journal of Molecular Biology and Biotechnology* 16 (1): 17-24.
- Rishi, A.S., Nelson, N.D. and Goyal, A. 2004. Genome Walking of Large Fragments: An Improved Method. *Journal of Biotechnology* 111: 9-15.
- Rival, A., Jaligot, E., Beule, T. and Jean Finnegan, E. 2008. Isolation and Expression Analysis of Genes Encoding *MET*, *CMT*, and *DRM* methyltransferases in Oil Palm (*Elaeis guineensis* Jacq.) in Relation to the ‘Mantled’ Somaclonal Variation. *Journal of Experimental Botany* 59(12): 3271-3281.
- Rogers, J.C., Lanahan, M.B. and Rogers, S.W. 1994. The *cis*-acting Gibberellin Response Complex in High-pI-amylase Gene Promoters. *Plant Physiology* 105: 151-158.

- 
- Rombauts, S., Dhais, P., Van Montagu, M. and Rouzé P. 1999. PlantCARE, a Plant *cis*-acting Regulatory Element Database. *Nucleic Acids Research* 27(1):2 95-6.
- Sambanthamurthi, R., Sundram, K. and Tan, Y.A. 2000. Chemistry and Biochemistry of Palm Oil. *Progress in Lipid Research* 39: 507-558.
- Sambrook, J., Fritsch, E.F. and Maniatis, T. 2001. Molecular Cloning: A Laboratory Manual, third ed. Cold Spring Harbor Laboratory Press, NY.
- Savidge, B., Weiss, J.D., Wong, Y.H., Lassner, M.W., Mitsky, T.A., Shewmaker, C.K., Post-Beittenmiller, D. and Valentin H.E. 2002. Isolation and Characterization of Homogentisate Phytyl-transferase Genes from *Synechocystis* sp. PCC 6803 and *Arabidopsis*. *Plant Physiology* 129:321–332.
- Schultz, G. 1990. Biosynthesis of  $\alpha$ -tocopherols in Chloroplasts of Higher Plants. *European Journal of Lipid Science and Technology* 92: 86-91.
- Selvaduray, K.N., Radhakrishnan, A.K., Kutty, M.K. and Nesaretnam, K. 2012. Palm Tocotrienols Decrease Levels of Pro-angiogenic Markers in Human Umbilical Vein Endothelial Cells (HUVEC) and Murine Mammary Cancer Cells. *Genes & Nutrition* 7(1): 53-61.
- Sen, C.K., Khanna, S. and Roy, S. 2006. Tocotrienols: Vitamin E Beyond Tocopherols. *Life Sciences* 78: 2088–2098.
- Serbinova, E.A. and Packer, L. 1994. Antioxidant Properties of  $\alpha$ -tocopherol and  $\alpha$ -tocotrienol. *Methods in Enzymology* 234: 354-366.
- Serbinova, E.A., Kagan, V., Han, D., Packer, L., 1991. Free radical recycling and intramembrane mobility in the antioxidant properties of alpha-tocopherol and alpha-tocotrienol. *Free Radical Biology & Medicine* 10 (5), 263–275.
- Shahmuradov, I.A., Gammerman, A.J., Hancock, J.M., Bramley, P.M. and Solovyev, V.V. 2003. PlantProm: A Database of Plant Promoter Sequences. *Nucleic Acids Research* 31(1): 114-117.
- Shahmuradov, I.A., Solovyev, V.V. and Gammerman, A.J. 2005. Plant Promoter Prediction with Confidence Estimation. *Nucleic Acids Research*. 33(3): 1069-1076.
- Shiple, G.L. 2006. An Introduction to Real-Time PCR. In “Real-time PCR” (Ed. Dorak, M. T.) pp. 1-37.
- Silveira, E.D., Alves-Ferreira, M., Guimarães, L.A., Rodrigues da Silva, F. and Tavares de Campos Carneiro, V. 2009. Selection of Reference Genes for Quantitative Real-

- 
- Time PCR Expression Studies in the Apomictic and Sexual Grass *Brachiaria brizantha*. *BMC Plant Biology* 9: 84.
- Siti Nor Akmar, A. and Zubaidah, R. 2008. Mesocarp-specific Metallothionein-like Gene Promoter for Genetic Engineering of Oil Palm. *Journal of Oil Palm Research* 2:1-8.
- Skriver, K., Olsen, F.L., Rogers, J.C., and Mundy, J. 1991. *cis*-acting DNA Elements Responsive to Gibberellin and Its Antagonist Abscisic Acid. *Proceedings of the National Academy of Sciences of the United States of America* 88: 7266-7270.
- Srivastava, J.K. and Gupta, S. 2006. Tocotrienol-rich Fraction of Palm Oil Induces Cell Cycle Arrest and Apoptosis Selectively in Human Prostate Cancer Cells. *Biochemical and Biophysical Research Communications* 346: 447-453.
- Suzuki, Y.J., Tsuchiya, M., Wassall, S.R., Choo, Y.M., Govil, G., Kagan, V.E. and Packer, L. 1993. Structural and Dynamic Membrane Properties of Alpha-tocopherol and Alpha-tocotrienol: Implication to the Molecular Mechanism of Their Antioxidant Potency. *Biochemistry* 32(40): 10692-10699.
- Sundram, K. and Nor, R.M. 2002. Analysis of Tocotrienols in Different Sample Matrixes by HPLC. In: Armstrong A, ed. *Methods in Molecular Biology*, vol. 186: Oxidative Stress Biomarkers and Antioxidant Protocols. Totowa, New Jersey: Humana Press Inc. pp 221-232.
- Sundram, K., Sambanthamurthi, R. and Tan, Y.A. 2003. Palm Fruit Chemistry and Nutrition. *Asia Pacific Journal Clinical Nutrition* 12 (3): 355-362.
- Terauchi, R. and Kahl, G. 2000. Rapid isolation of Promoter Sequences by TAIL-PCR: the 5'-flanking Regions of *Pal* and *Pgi* Genes from Yams (*Dioscorea*). *Molecular and General Genetics* 263: 554-560.
- Toplak, I., Grom, J., Hostnik, P. and Barlic-Meaganja, D. 2004. Phylogenetic Analysis of Type 2 Porcine Cicoviruses Identified in wild Boar in Slovenia. *Veterinary Record* 155(6): 178-180.
- Toyofuku, K., Umemura, T. and Yamaguchi, J. 1998. Promoter Elements required for Sugar-repression of the *RAmy3D* Gene for K-amylase in Rice. *FEBS Letters* 428: 275-280.
- Unni, S.C., Vivek, P.J., Maju, T.T., Varghese, R.T. and Soniya, E.V. 2012. Molecular Cloning and Characterization of Fruit Specific Promoter from *Cucumis sativus L.* *American Journal of Molecular Biology* 2: 132-139.
- Vandesompele, J., Preter, K.D., Pattyn, F., Poppe, B., Roy, N.V., Paepe, A.D. and Speleman, F. 2002. Accurate Normalization of Real-Time Quantitative RT-PCR

---

Data by Geometric Averaging of Multiple Internal Control Genes. *Genome Biology* 3(7): 34.1–34.11.

- Van Eenennaam, A.L., Lincoln, K., Durrett, D.P., Valentin, H.E., Shewmaker, C.K., Thorne, G.M., Jiang, J., Baszis, S.R., Levering, C.K., Aasen, E.D., Hao, M., Stein, J.C., Norris, S.R. and Last, R.L. 2003. Engineering Vitamin E Content: From Arabidopsis Mutant to Soy Oil. *The Plant Cell* 15: 3007–3019.
- Washida, H., Wu, C.Y., Suzuki, A., Yamanouchi, U., Akihama, T., Harada, K. and Takaiwa, F. 1999. Identification of *cis*-regulatory Elements required for Endosperm Expression of the Rice Storage Protein Glutelin Gene *GluB-1*. *Plant Molecular Biology* 40: 1-12.
- Xu, M., Zhang, B., Su, X.H., Zhang, S.G. and Huang, M.R. 2011. Reference Gene Selection for Quantitative Real-Time Polymerase Chain Reaction in *Populus*. *Analytical Biochemistry* 408: 337-339.
- Xu, W.F. and Shi, W.M. 2006. Expression Profiling of the 14-3-3 Gene Family in Response to Salt Stress and Potassium and Iron Deficiencies in Young Tomato (*Solanum lycopersicum*) Roots: Analysis by Real-time RT-PCR. *Annals of Botany* 98: 965-974.
- Yamaguchi-Shinozaki, K. and Shinozaki, K. 2005. Organization of *cis*-acting Regulation Elements in Osmotic- and Cold-stress-responsive Promoters. *Trends in Plant Science* 10(2): 88-94.
- Yamamoto, Y.Y., Ichida, H., Abe, T., Suzuki, Y., Sugano, S. and Obokata, J. 2007. Differentiation of Core Promoter Architecture between Plants and Mammals Revealed by LDSS Analysis. *Nucleic Acids Research* 35(18): 6219-6226.
- Yang, W.Y., Cahoon, R.E., Hunter, S.C., Zhang, C.Y., Han, J.X., Borgschulte, T. and Cahoon, E.B. 2011. Vitamin E Biosynthesis: Functional Characterization of the Monocot Homogentisate Geranylgeranyl Transferase. *The Plant Journal* 65: 206–217.
- Yeoh, K.A., Othman, A., Meon, S., Abdullah, F. and Ho, C.L. 2012. Sequence Analysis and Gene Expression of Putative Exo- and Endo-Glucanases from Oil Palm (*Elaeis guineensis*) during Fungal Infection. *Journal of Plant Physiology* 169(15): 1565-1570.
- Yuan, Y.J., Liang, Y.X., Zheng, Y.S. and Li, D.D. 2012. Cloning, Characterization and Expression Analysis of a 7S Globulin Gene in Mesocarp of Oil Palm (*Elaeis guineensis* Jacq.). *Scientia Horticulturae* 143: 167-175.
- Zhong, H.Y., Chen, J.W., Li, C.Q., Chen, L., Wu, J.Y., Chen, J.Y., Lu, W.J. and Li, J.G. 2011. Selection of Reliable Reference Genes for Expression Studies by Reverse

---

Transcription Quantitative Real-Time PCR in Litchi under Different Experimental Conditions. *Plant Cell Report* 30: 641-653.

