

UNIVERSITI PUTRA MALAYSIA

ASSESSMENT OF ORANGE SPOTTING SEVERITY IN OIL PALM USING MULTI SPECTRAL REFLECTANCE APPROACH

SUDHARSAN SELVARAJA

FP 2014 49



ASSESSMENT OF ORANGE SPOTTING SEVERITY IN OIL PALM USING MULTI SPECTRAL REFLECTANCE APPROACH



Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

September 2014

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of Thesis Presented to the Senate of Universiti Putra Malaysia in the Fulfillment of the Requirement for the Degree of Master of Science

ASSESSMENT OF ORANGE SPOTTING SEVERITY IN OIL PALM USING MULTI SPECTRAL REFLECTANCE APPROACH

By

SUDHARSAN SELVARAJA

September 2014

Chairman: Siva K. Balasundram, PhD Faculty: Agriculture

Orange Spotting (OS) caused by a variant of Cadang-Cadang Coconut Viroid (CCCVd) is an emerging problem in oil palm plantations of Malaysia. This work was an effort to study epidemiology of OS in an oil palm plantation using Precision Agriculture (PA) tools. A 4.2 ha plot were established in a commercial plantation situated at Sungai Buloh, Selangor. A total of 587 geo-referenced mature trees were visually observed to identify OS-infected oil palm trees based on symptoms. OS incidence and severity data were acquired from all 587 observed oil palm trees and the data subjected to a series of spatial variability analysis. Forty symptomatic oil palm trees were systematically selected based on OS severity for leaflets sampling. Leaflets from another 10 nonsymptomatic oil palm trees were also sampled. The sampled leaflets were subjected to spectral reflectance acquisition followed by Dot-Blot analysis. A second study plot was established in a mature oil palm stand (approximately 10 km away from the first study site) which was selected based on presence of potassium deficiency and absence of OS. The sampled leaflets were subjected to spectral reflectance acquisition (K) analysis.

The incidence of OS incidence in the study area was 74.3%. OS severity ranged from 0-92.3%. The spatial structure of OS severity was described by an exponential model with an effective range of 29.1 m. Orange spotting severity exhibited a strong spatial dependence with a nugget to sill ratio of 0.15. The spatial variability map of OS severity revealed spatial clustering of kriged values, where 73% of the study area showed low severity (1-30%), 25% showed moderate severity (30-60%) and approximately 2% showed high severity (>60%). This study demonstrates the utility of geo-spatial information in understanding the OS severity scale which could assist in site-specific disease monitoring and intervention.

Dot-blot assay showed CCCVd presence in 40% of the samples obtained from nonsymptomatic oil palm trees. Symptomatic leaf samples showed a significant correlation (r=-0.70) between leaf chlorophyll reflectance and OS severity. Spectral reflectance of symptomatic leaves was significantly lower than that of nonsymptomatic leaves at the 465-711 nm wavelength regions. Spectral reflectance of healthy and asymptomatic leaves, however, did not exhibit significant differences



across all wavelengths investigated. In symptomatic leaves, spectral reflectance showed a decreasing trend with an increase in OS severity of up to 60% at the 555 nm and 780-1000 nm wavelengths. Among the vegetation indices tested, MCARI1 and mSR₇₀₅ performed best in predicting OS severity with a goodness of fit (measured vs. predicted) of 66% and 56%, respectively.

Reflectance between OS-infected and K-deficient leaves showed significant separability (p < 0.05) at 400-538 nm and 667-688 nm wavelengths. Reflectance of K-deficient leaves was significantly different than that of OS-infected leaves from OS severity classes (1-20%, 21-40%, 41-60% and 61-80%). All oil palm leaflets exhibited green peak at 555 nm wavelength, with average reflectance value of 0.15.



Abstrak Tesis yang Dikemukakan kepada Senat Universiti Putra Malaysia Sebagai Memenuhi Keperluan untuk Ijazah Master Sains

PENILAIAN PENYAKIT ORANGE SPOTTING DI TANAMAN KELAPA SAWIT MELALUI PENDEKATAN PANTULAN MULTI SPEKTRUM

Oleh

SUDHARSAN SELVARAJA

September 2014

Pengerusi: Siva K. Balasundram, PhD Fakulti: Pertanian

Penyakit "Orange Spotting" (OS) disebabkan oleh "Cadang-Cadang Coconut Viroid" (CCCVd) kini telah mula menular ladang-ladang kelapa sawit di Malaysia. Penyelidikan ini adalah satu usaha untuk mengkaji epidemiologi penyakit OS di dalam sebuah ladang kelapa sawit dengan menggunakan peralatan "Precision Agriculture" (PA). Plot kajian seluas 4.2 ha telah disediakan di ladang komersial yang terletak di Sungai Buloh, Selangor. Sebanyak 587 pokok matang yang digeorujuk telah dikaji berdasarkan pemerhatian visual untuk mengenal pasti pokok-pokok kelapa sawit OS dijangkiti. Insiden penyakit OS dan data tahap diperolehi daripada semua 587 pokok kelapa sawit diperhatikan dan data tertakluk kepada satu siri analisis kebolehubahan spatial. Empat puluh gejala pokok kelapa sawit telah dipilih secara sistematik berdasarkan keparahan penyakit OS yang simtomatik bagi tujuan persampelan daun. 10 pokok kelapa sawit yang tiada simtom juga telah disampel. Sampel daun tersebut tertakluk kepada pengambilalihan pantulan spektrum diikuti oleh analisis Dot-Blot. Plot kajian kedua ditubuhkan di sebuah kawasan kelapa sawit matang (kira-kira 10 km dari tapak kajian pertama) yang dipilih berdasarkan kepada kehadiran kekurangan kalium dan ketiadaan penyakit OS. Sampel daun tertakluk kepada pengambilalihan pantulan spektrum diikuti dengan daun Kalium (K) analisis.



Insiden penyakit OS di kawasan kajian ini adalah 74.3%. Tahap penyakit OS adalah diantara 0-92.3%. Struktur ruang tahap penyakit OS digambarkan oleh model eksponen dengan jarak berkesan sejauh 29.1 m. Tahap penyakit OS telah menunjukkan pergantungan spatial yang kukuh dengan nugget kepada nisbah 0.15. Peta spatial kebolehubahan tahap penyakit OS mendedahkan kelompok jangkitan, di mana 73% daripada kawasan kajian menunjukkan tahap rendah (1-30%), 25% menunjukkan tahap sederhana (30-60%) dan kira-kira 2% lagi menunjukkan tahap tinggi (> 60%). Kajian ini menunjukkan utiliti maklumat geo- spatial dalam memahami tahap penyakit OS yang boleh membantu dalam khusus tapak pemantauan penyakit.

Analisis Dol-Blot menunjukkan kehadiran CCCVd sebanyak 40% dalam sampel yang diperolehi daripada pokok kelapa sawit yang tiada simtom. Sampel daun

menunjukkan perkaitan yang signifikan (r = -0.70) antara pantulan klorofil daun dan tahap penyakit OS. Pantulan spektrum daun adalah jauh lebih rendah daripada itu daun yang tiada simtom dalam rantau jarak gelombang sebanyak 465-711 nm. Pantulan spektrum daun yang sihat dan asimptomatik tidak menunjukkan perbezaan yang ketara di semua panjang gelombang disiasat. Dalam daun simptomatik, pantulan spektrum menunjukkan trend yang semakin mengecil dengan peningkatan dalam tahap penyakit OS sehingga 60% pada jarak gelombang di 555 nm dan 780-1000 nm. Antara indeks tumbuh-tumbuhan yang diuji, MCARI1 dan mSR705 dilakukan terbaik dalam meramalkan penyakit OS keterukan dengan keserasian sebanyak 66% dan 56% secara masing-masing.

Pantulan antara daun dijangkiti OS dan kekurangan K menunjukkan yang ia boleh dipisahkan secara signifikan (p <0.05) pada panjang gelombang di 400-538 nm dan 667-688 nm. Pantulan daun kekurangan K adalah jauh berbeza daripada yang daun dijangkiti OS dengan tahap penyakit OS(1-20%, 21-40%, 41-60% dan 61-80%). Semua sampel daun kelapa sawit telah mempamerkan puncak hijau di panjang gelombang 555 nm, dengan purata nilai pantulan sebanyak 0.15.

ACKNOWLEDGEMENT

I would like to take this opportunity to thank all those who contributed for the completion of this work. First and foremost, I would like to extend my deepest gratitude to my supervisor, Assoc. Prof. Dr. Siva K. Balasundram, for his generous guidance, advice and endless support that contributed significantly towards the completion of this work. His careful reviews and constructive criticism have been crucially important for this thesis write up.

My sincere gratitude to my board of co-supervisors, Assoc. Prof. Dr. Ganesan Vadamalai and Assoc. Prof. Dr. Ahmad Husni for their constructive advice and comments throughout the entire course of this research.

I would like to express my heartiest appreciation to my family for their financial and emotional support throughout my postgraduate studies.



I certify that a Thesis Examination Committee has met on 23 September 2014 to conduct final examination on Sudharsan Selvaraja on his thesis entitled "Assessment of Orange Spotting Severity in Oil Palm Using Multi Spectral Reflectance Approach" in accordance with the Universities and University College Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U. (A) 106] 15 March 1998. The committee recommends that the student be awarded the Master of Science. Members of the Thesis Examination Committee were as follows:

Halimi bin Mohd Saud, PhD

Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Helmi Zulhaidi bin Mohd Shafri, PhD

Associate Professor Faculty of Engineering Universiti Putra Malaysia (Internal Examiner)

Jugah bin Kadir, PhD

Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Internal Examinar)

Mahantesh Patil, PhD

Professor, College of Agriculture University of Agriculture Sciences Karnataka, India (External Examiner)

ZULKARNAIN ZAINAL, PhD

Professor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date: 23 November 2014

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Siva Kumar Balasundram, PhD

Associate Professor, Faculty of Agriculture, Department of Agriculture Technology, (Chairman)

Ahmad Husni B Mohd Haniff, PhD

Associate Professor, Faculty of Agriculture, Department of Land Management, (Member)

Ganesan Vadamalai, PhD

Associate Professor, Faculty of Agriculture, Department of Plant Protection, (Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- This thesis is my original work;
- Quotations, illustrations and citation have been duly referenced;
- This thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- Intellectual property from the thesis and copyright of the thesis are fullyowned by Universiti Putra Malaysia as according to the Universiti Putra Malaysia (research) rules 2012;
- Written permission must be obtained from supervisor and the office of the deputy vice-chancellor (research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (research) rules 2012;
- There is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (graduate studies) rules 2003 (revision 2012-2013) and the Universiti Putra Malaysia (research) rules 2012. The thesis has undergone plagiarism detection software.

Signature:	Date:
Name and Matric No:	

Declaration by Members of Supervisory Committee

This is to confirm that

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature:

Chairman of Supervisory Committee:

Assoc. Prof. Dr. Siva Kumar Balasundram

Signature:

Member of Supervisory Committee:

Assoc. Prof. Dr. Ahmad Husni B Mohd Haniff

Signature:

Member of Supervisory Committee:

Assoc. Prof. Dr. Ganesan Vadamalai

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	V
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xii
LIST OF FIGURES	xiii
LIST OF ABBREVIATIONS	XV

CHAPTER

1	INTR	RODUCTION	1
	1.1	Background	1
	1.2	Problem Statement	2
	1.3	Significance of Study	2
	1.4	Research Objectives	2
2	LITE	RATURE REVIEW	
	2.1	Orange Spotting (OS)	3
		2.1.1 Epidemiology of OS	3
		2.1.2 Symptoms of OS	4
	2.2	Plant Disease Assessment	5
	2.3	Precision Agriculture in Plant Disease Assessment	5
	2.4	Spatial Variability	6
		2.4.1 Application of spatial variability in plant disease assessment	9
	2.5	Spectroscopy	10
		2.5.1 Spectral response of vegetations	10
		2.5.2 Use of spectroscopy in plant disease assessment	12
		2.5.3 Multispectral spectroradiometer	13
3	SPAT	TIAL VARIABILITY OF ORANGE SPOTTING IN OIL	
	PALN	M	15
	3.1	Introduction	15
	3.2	Materials and Methods	16
		3.2.1 Spatial variability	16
		3.2.2 Study site	17
		3.2.3 Disease severity assessment	17
		3.2.4 Exploratory data analysis	17
		3.2.5 Spatial data analysis	17
	3.3	Results and Discussions	18
	3.4	Conclusion	23

4	MUI	LTISPECTRAL REFLECTANCE AS A TOOL TO ASSESS	25
	ORA	NGE SPOTTING IN OIL PALM	
	4.1	Introduction	25
	4.2	Materials and Methods	26
		4.2.1 Study site	26
		4.2.2 Leaf sampling	26
		4.2.3 Leaf spectral reflectance	27
		4.2.4 Leaf chlorophyll reflectance	29
		4.2.5 CCCVd detection	29
		4.2.6 Data analysis	29
	4.3	Results and Discussions	29
		4.3.1 Visual OS symptoms	29
		4.3.2 CCCVd detection using Dot-Blot assav	29
		4.3.3 Relationship between OS severity and leaf	
		chlorophyll reflectance	30
		434 Spectral reflectance of symptomatic nonsymptomatic	
		and asymptomatic leaves	31
		435 Correlation between spectral reflectance and OS	
		4.5.5 Contration between spectral reneetance and OS	34
		4.2.6 Palationship between vegetation indices and OS	
		4.3.0 Relationship between vegetation indices and OS	35
	1 1	Conclusion	27
	4.4	Conclusion	57
5	USE	OF SPECTRAL PEELECTANCE TO DISCRIMINATE	
5	USE	WEEN DOTASSIUM DEFICIENCY AND OD ANCE	20
		TTINC IN OIL DALM (Elegis guingenzie)	39
	SPU	I TING IN OIL PALM (Elaets guineensis)	20
	5.1	Introduction	39
	5.2	Materials and Methods	41
		5.2.1 Study site	41
		5.2.2 Leaf sampling	41
		5.2.3 OS severity assessment	41
		5.2.4 Spectral reflectance	41
		5.2.5 Determination of leaf K content	42
		5.2.7 Data analysis	42
	5.3	Results and Discussions	42
	5.4	Conclusion	45
6	SUM	IMARY, GENERAL CONCLUSION AND	47
	REC	OMMENDATIONS FOR FUTURE RESEARCH	17
	6.1	Summary and General Conclusion	47
	6.2	Recommendations	48
REF	FEREN	ICES	49
APP	PENDI	CES	56
BIO	DATA	OF STUDENT	59
LIS	T OF P	PUBLICATIONS	60

LIST OF TABLES

Table		Page
2.1	Types of spectroscopy and its application in plant disease assessment	13
3.1	Strength of spatial dependence based on nugget to sill ratio	18
3.2	Descriptive statistics of OS severity	18
3.3	Cross validation statistics of interpolated values for OS severity	20
3.4	Cross validation statistics of interpolated values for OS severity from the demarcated sub-plots	20
3.5	Interpolated OS severity categories and corresponding aerial distribution	23
4.1	List of vegetation indices used in this study	28
4.2	Wavelengths that significantly discriminate symptomatic and nonsymptomatic leaves based on contrast analysis	33
4.3	Pearson correlation between spectral reflectance and OS severity	34
4.4	Regression of vegetation index on OS severity (%)	35
5.1	Descriptive statistics of leaf K content and OS severity	43
5.2	Wavelengths which significantly discriminates K-deficient leaves and four classes of OS severity	44

LIST OF FIGURES

Figure		Page
2.1	An example of semivariogram using exponential model with range, nugget (C_0) and sill $(C_0 + C_1)$	7
2.2	Spatial prediction is a process of quantitative estimation of properties value at unsampled site within the area covered by existing observations	9
2.3	Vegetation spectral reflectance curve showing main four wavelength regions	11
2.4	Leaf morphology influencing the spectral reflectance from visible and infrared wavelengths	12
3.1	Distribution of OS severity	18
3.2	Spatial structure of OS severity computed using i) total dataset (n = 587) and ii) symptom-based dataset (n = 436)	19
3.3	Spatial structure and attributes of OS severity from the four 1-ha sub-plots	21
3.4	Spatial variability of OS severity from the demarcated sub-plots (based on measured and kriged values)	22
3.5	Spatial variability of OS severity (based on measured and kriged values)	23
4.1	Ten leaves were sub-sampled from each of the three sections within the sampled frond	27
4.2	CCCVd detection in leaf samples using the Dot-Blot analysis $(n=50)$; where +/k denotes presence of CCCVd, -/w denotes absence of CCCVd	30
4.3	Correlation between OS severity and leaf chlorophyll reflectance	31
4.4	Comparison of spectral reflectance between nonsymptomatic $(n=10)$ and symptomatic $(n=40)$ fronds	32
4.5	Comparison of spectral reflectance between healthy $(n=6)$ and asymptomatic $(n=4)$ fronds	32

 \bigcirc

- 4.6 Spectral reflectance of symptomatic (based on various severity 34 clusters), asymptomatic and healthy leaves
- 4.7 Comparison between measured and predicted OS severity based 36 on seven vegetation indices used in this study
- 5.1 Comparison of symptoms between OS-infected and K-deficient 40 leaves in mature (> 6 years old) oil palm trees
- 5.2 Comparison of spectral reflectance between OS-infected and K- 44 deficient oil palm leaves
- 5.3 Comparison of spectral reflectance from OS-infected (four 45 classes of severity), K-deficient and healthy leaves



LIST OF ABBREVIATIONS

CCCVd	Coconut Cadang-Cadang Viroid
et al.	et alia
GIS	Geographical Information System
GM	Gitelson and Merzlak Index
GPS	Global Positioning System
MCARI	Modified Chlorophyll Absorption in Reflectance Index
mND	Modified Normalized Difference
MPOB	Malaysian Palm Oil Board
MS	Mean Square
MSE	Mean Square Error
mSR	Modified Spectral Ratio
NDCI	Normalized Difference Cloud Index
NDI	Chlorophyll Normalized Difference Index
NDRE	Normalized Difference Red Edge
NDVI	Normalized Difference Vegetation Index
NPCI	Normalized Pigment Chlorophyll Index
OS	Orange Spotting
PSSR	Pigment Specific Simple Ratio
RI	Response Index
SIPI	Structural Independent Pigment Index
SPAD	Soil Plant Analysis Development
SRPI	Simple Ratio Pigment Index
VI	Vegetation Index
ZM	Zarco-Tejada and Miller Index



CHAPTER 1

INTRODUCTION

1.1 Background

The oil palm, *Elaeis guineensis*, originates from Africa. Palm oil is mainly processed and refined as edible oil. Palm oil is also extensively used to make soap, detergent and margarine in an industrial scale. Crude palm oil is the most exported commodity in Malaysia with a constantly increasing demand. Total oil palm planted area in Malaysia increased by 4.5% to 4.69 million hectares in 2009. Among the regions, Sarawak registered the largest increase in planted area at 12.8%, followed 3.3% and 2.1% in Peninsular Malaysia and Sabah, respectively. Sabah still stands as the state with the largest oil palm acreage, accounting for 1.36 million hectares or 29% of the total planted area in Malaysia (MPOB, 2012). Currently, Malaysia produces 17.73 million tonnes of palm oil and 2.13 tonnes of palm kernel oil. Malaysia is one the largest producers and exporters of palm oil in the world, accounting for 11% of the world's oils & fats production and 27% of export trade of oils & fats (USDA, 2013). The industry provides employment to more than half a million people and livelihood to an estimated one million people.

However, oil palm trees are prone to diseases which may significantly affect oil palm yields (Corley, 2008). Kamarudin and Arshad (2006) had reported oil palm yield affecting pests and diseases such as foliar damage caused by *Metisa plana* (bagworms) and fruit bunch damage caused by *Oryctes rhinoceros* (Asiatic rhinoceros beetle). This study revealed that 50% of defoliation damage in an oil palm plantation results in loss of 4 tonne per hectare of fresh fruit bunches. Another well-known disease of oil palm such as basal stem rot disease caused by *Ganoderma sp.* also affects oil palm yield significantly (Naher et al., 2013). These damages had been well documented and its' epidemiology are known, thus, control methods could be conducted timely and efficiently by removing the infected palm. Recently, OS disease incidence had been reported in several oil palm plantations situated along the west coast of Malaysia. Even though, presence of OS in Malaysian oil palm plantations known, epidemiology of OS in oil palm is not documented. Thus, lack of understanding in epidemiology of OS disrupts plantation managements in taking timely control actions.

Orange Spotting (OS) is becoming a significant problem in oil palm plantations in Malaysia based on the symptom exhibited in several oil palm plantations. Vadamalai (2009) had associated OS disease with several variants of Coconut Cadang-Cadang Viroid (CCCVd) in oil palms in Malaysia. OS disease epidemiology in an oil palm plantation is still unclear since there are no publications available to date. Therefore, this study was focused on understanding the epidemiology of OS via plant disease assessment. Potential of contemporary techniques in plant disease assessment had been evaluated in an effort to accurately quantify and understanding epidemiology of OS. This study may serve as a stepping stone towards understanding epidemiology of OS in oil palm.

1.2 Problem Statement

In Malaysia, studies on epidemiology of OS disease in oil palms have not been documented. Unknown epidemiology, mainly distribution and phytopathometry, of OS in Malaysian oil palm plantations would affect disease control actions in case of an outbreak of the disease.

Symptoms of OS are exhibited on oil palm leaves (Hanold and Randles, 1998). Conventional plant disease assessment via visual assessment could yield erroneous result due to high canopy position in matured oil palm trees. Plant disease assessment requires an accurate and robust method for quick and effective disease control actions (Nutter et al., 2006).

OS of oil palm could be asymptomatic (Turner, 1981). Asymptomatic oil palm trees do not exhibit OS symptoms while acting as CCCVd host. Asymptomatic oil palm trees could act as a loci or a source point of OS transmission which may not be detected by visual assessment.

Symptoms of OS and Potassium (K) deficiency are similar via visual assessment. Trained skilled workers required for the discrimination of OS and potassium deficiency in oil palm. Hence, disease assessment may vary between raters causing unreliable disease assessment results (Nutter *et al.*, 2006).

Plant disease assessment of OS in oil palms of Malaysia had not been conducted.

1.3 Significance of Study

Firstly, the distribution of OS in an oil palm plantation was evaluated using spatial variability technique. A geostatistical analysis has been adopted in this spatial variability OS study to accurately estimate the distribution of OS. Spatial variability study of OS severity may reveal if the disease is random or aggregated in an oil palm plantation.

Secondly, there was a need to accurately quantify OS severity (Nutter *et al.*, 2006). Potential of spectral reflectance to quantify OS severity had been evaluated this study. Use of spectral reflectance in quantifying OS may replace the need of erroneous visual assessment in OS assessment.

Thirdly, spectral reflectance is used to discriminate OS and K deficiency in oil palm trees. This may serve as an alternative method in discriminating these disorders.

1.4 Research Objectives

- 1. To assess the incidence and severity of OS in an oil palm plantation using spatial variability technique.
- 2. To quantify and discriminate OS severity into significant clusters using multispectral reflectance and several published vegetation indices.
- 3. To evaluate and analyze the performance of multispectral reflectance in discriminating OS and K deficiency symptoms in oil palm trees.

REFERENCES

- Albayrak, S. (2008). Use of reflectance measurements for the detection of N, P, K, ADF and NDF contents in sainfoin pasture. Sensors, 8(11): 7275-7286.
- Apan, A., Datt, B. and Kelly, R. (2005). Detection of pests and diseases in vegetable crops using hyperspectral sensing: A comparison of reflectance data for different sets of symptoms. Proceedings of SSC 2005 Spatial Intelligence, Innovation and Praxis. Spatial Sciences Institute. Melbourne, Australia. pp. 10-18.
- Araus, J.L., Casadesus, J. and Bort, J. (2001). Recent tools for the screening of physiological traits determining yield. In: Application of Physiology in Wheat Breeding. Reynolds, M.P., Ortiz-Monasterio, J.I. and McNab, A (eds). CIMMYT, D.F, Mexico. pp. 59-77.
- Balasundram, S.K., Mulla, D.J. and Robert, P.C. (2006a). Accounting for spatial variability in a short-term fertilizer trial for oil palm. International Journal of Soil Science, 1(3): 184-195.
- Balasundram, S.K., Mulla, D.J. and Robert, P.C. (2006b). Relationship between oil palm yield and soil fertility as affected by topography in an Indonesian plantation. Communications in Soil Science and Plant Analysis, 37 (9-10): 1321-1337.
- Balasundram, S.K., Husni, M.H.A. and Ahmed, O.H. (2008). Application of geostatistical tools to quantify spatial variability of selected soil chemical properties from a cultivated tropical peat. Journal of Agronomy, 7: 82-87.
- Baranoski, G.V.G. and Rokne, J.G. (2005). A practical approach for estimating the red edge position of plant leaf reflectance. International Journal of Remote Sensing, 26 (3): 503-521.
- Basayigit, L., and Senol, H. (2009). Prediction of plant nutrient contents in deciduous orchards fruits using spectroradiometer. International Journal of ChemTech Research, 1(2): 212-224.
- Belasque Jr, J., Gasparoto, M.C.G., and Marcassa, L. G. (2008). Detection of mechanical and disease stresses in citrus plants by fluorescence spectroscopy. Applied Optics, 47(11): 1922-1926.
- Björkman, O., and Demmig-Adams, B. (1994). Regulation of photosynthetic light energy capture, conversion, and dissipation in leaves of higher plants. In: Ecophysiology of Photosynthesis. Schulze, E.D. and Caldwell, M.M. (eds). Springer Heidelberg. Berlin, Germany. pp. 17-47.
- Blackburn, G.A. (1998). Spectral indices for estimating photosynthetic pigment concentrations: A test using senescent tree leaves. International Journal of Remote Sensing, 19 (4): 657-675.

- Cambardella, C.A., Moorman, T.B., Novak, J.M., Parkin, T.B., Karlen, D.L., Turco, R.F. and Konopka, A.E. (1994). Field-scale variability of soil properties in central Iowa soils. Soil Science Society of American Journal, 58(5): 1501-1511.
- Carter, G.A. (1993). Responses of leaf spectral reflectance to plant stress. American Journal of Botany, 80(3): 239-243.
- Cibula, W.G. and Carter, G.A. (1992). Identification of a far-red reflectance response to ectomycorrhizae in slash pine. International Journal of Remote Sensing, 13(5): 925-932.
- Corley, R.H.V. and Tinker, P.B.H. (2008). The Oil Palm. World Agriculture Series. John Wiley & Sons. p. 592.
- Daughtry, C.S.T., Walthall, C.L., Kim, M.S., Brown de Colstoun, E. and McMurtrey III, J.E. (2000). Estimating corn leaf chlorophyll from leaf and canopy reflectance. Remote Sensing of Environment, 74(2): 229-239.
- Delhomme, J.P. (1978). Kriging in the hydrosciences. Advances in Water Resources, 1(5): 251-266.
- Flatman, G.T. and Yfantis, A.A. (1984). Geostatistical strategy for soil sampling: The survey and the census. Environmental Monitoring and Assessment, 4(4): 335-349.
- Forde, S.C.M. and Leyritz, M.J.P. (1968). A study of confluent orange spotting of the oil palm in Nigeria. Journal of the Nigerian Institute for Oil Palm Research, 4: 371-380.
- Franke, J. and Menz, G. (2007). Multi-temporal wheat disease detection by multispectral remote sensing. Precision Agriculture, 8(3): 161-172.
- Gitelson, A. and Merzlyak, M.N. (1994). Spectral reflectance changes associated with autumn senescence of *Aesculus hippocastanum* and *Acer platanoides* leaves. Journal of Plant Physiology, 143(3): 286-292.
- Gitelson, A. and Merzlyak, M.N. (1998). Remote sensing of chlorophyll concentration in higher plant leaves. Advanced Space Research, 22(5): 689-692.
- Graeff, S., Link, J., and Claupein, W. (2006). Identification of powdery mildew (*Erysiphe graminis* sp. tritici) and take-all disease (*Gaeumannomyces graminis* sp. tritici) in wheat (*Triticum aestivum* L.) by means of leaf reflectance measurements. Central European Journal of Biology, 1(2): 275-288.
- Gupta, R.K., Vijayan, D. and Prasad, T.S. (2003). Comparative analysis of red edge hyperspectral indices. Advanced Space Research, 32(11): 2217-2222.

- Haboudane, D., Miller, J.R., Pattey, E., Zarco-Tejada, P.J. and Strachan, I.B. (2004). Hyperspectral vegetation indices and novel algorithms for predicting green LAI of crop canopies: Modeling and validation in the context of precision agriculture. Remote sensing of Environment, 90(3): 337-352.
- Hanold, D. and Randles, J.W. (1998). Report on ACIAR-Funded Research on Viroids and Viruses of Coconut Palm and Other Tropical Monocotyledons 1985-1993. ACIAR Monograph No. 45. Australian Centre for International Agricultural Research, Canberra, Australia. p. 45.
- Hanold, D. and Randles, J.W. (1991). Detection of coconut cadang-cadang viroidlike sequences in oil and coconut palm and other monocotyledons in South-West Pacific. Annals of Applied Biology, 118(1): 139-151.
- Hansen, P.M. and Schjoerring, J.K. (2003). Reflectance measurement of canopy biomass and nitrogen status in wheat crops using normalized difference vegetation indices and partial least squares regression. Remote Sensing of Environment, 86(4): 542-553.
- Isaaks, E.H. and Srivastava, R.M. (1989). An Introduction to Applied Geostatistics. Oxford University Press. New York. USA. p. 561.
- Jensen, J.R. (2009). Remote sensing of the environment: An earth resource perspective. Pearson Education. India. p. 592.
- Joo, G. K., Soon, C.P. and Kiang, K.K. (1994). K nutrition for mature oil palm in Malaysia. IPI Research Topics No. 17. International Potash Institute. Basel, Switzerland. p. 31.
- Kamarudin, N. and Arshad, O. (2006). Potentials of using a pheromone trap monitoring and controlling the bagworm, *Metisa plana* Wlk. (Lepidoptera: Psychidae) on young oil palm in a smallholder plantation. Journal of Asia-Pacific Entomology, 9(3): 281-285.
- Liu, Z.Y., Shi, J.J., Zhang, L.W. and Huang, J.F. (2010). Discrimination of rice panicles by hyperspectral reflectance data based on principal component analysis and support vector classification. Biomedicine and Biotechnology, 11(1): 71-78.
- Mahlein, A.K., Steiner, U., Hillnhütter, C., Dehne, H.W. and Oerke, E.C. (2012). Hyperspectral imaging for small-scale analysis of symptoms caused by different sugar beet diseases. Plant Methods, 8(1): 1-3.
- Mahlein, A.K. (2010). Detection, identification, and quantification of fungal diseases of sugar beet leaves using imaging and non-imaging hyperspectral techniques. Inaugural dissertation. Institute of Crop Science and Resource Conservation, University of Bonn. Bonn, Germany. p. 147.
- Marshak, A., Knyazikhin, Y., Davis, A., Wiscombe, W. and Pilewskie, P. (2000). Cloud-vegetation interaction: Use of normalized difference cloud index for

estimation of cloud optical thickness. Geophysical Research Letters, 27(12): 1695-1698.

- Merino, G.G., Jones, D., Stooksbury, D.E. and Hubbard, K.G. (2001). Determination of semivariogram models to krige hourly and daily solar irradiance in Western Nebraska. Journal of Applied Meteorology, 40(6): 1085-1094.
- Mohammed, G.H., Noland, T.L., Irving, D., Sampson, P.H., Zarco-Tejada, P.J. and Miller, J.R. (2000). Natural and stress-induced effects on leaf spectral reflectance in Ontario species. Forest Research Report No. 156. Ontario Forest Research Institute, Canada. p. 42.
- Moshou, D., Bravo, C., Oberti, R., West, J., Bodria, L., McCartney, A. and Ramon, H. (2005). Plant disease detection based on data fusion of hyper-spectral and multi-spectral fluorescence imaging using Kohonen maps. Real-Time Imaging, 11: 75-83.
- MPOB. (2012). Overview of the Malaysian Oil Palm Industry 2012. Retrieved 2 March 2013. http://bepi.mpob.gov.my/images/overview/Overview_of_Industry_2012.pdf.
- Naher, L., Yusuf, U.K., Ismail, A., Tan, S.G. and Mondal, M.M.A. (2013). Ecological status of *Ganoderma* and basal stem rot disease of oil palms (*Elaeis guineensis* Jacq.). Australian Journal of Crop Science, 7(11): 1723-1727.
- Naidu, R.A., Perry, E.M., Pierce, F.J., and Mekuria, T. (2009). The potential of spectral reflectance technique for the detection of grapevine leafroll-associated virus in two red-berried wine grape cultivars. Computers and Electronics in Agriculture, 66(1): 38-45.
- Nutter Jr, F.W., Esker, P.D., Netto, R.A.C. (2006). Disease assessment concepts and the advancements made in improving the accuracy and precision of plant disease data. European Journal of Plant Pathology, 115: 95-103.
- Owens, R.A. (1998). Viroids and viroid diseases of plants. In: Studies in Viral Ecology. Flores, R., Serios, F.D., Navarro, B., Vila, N.D. and Owens R.A. (eds). John Wiley & Sons. pp. 307-342.
- Paulitz, T.C., Zhang, H. and Cook, R.J. (2003). Spatial distribution of *Rhizoctonia oryzae* and rhizoctonia root rot in direct-seeded cereals. Canadian Journal of Plant Pathology, 25(3): 295-303.
- Peňuelas, J., Baret, F. and Filella, I. (1995). Semi-empirical indices to assess carotenoids/chlorophyll a ratio from leaf spectral reflectance. Photosynthetica, 31(2): 221-230.
- Peňuelas, J., Gamon, J.A., Fredeen, A.L., Merino, J. and Field, C.B. (1994). Reflectance indices associated with physiological changes in nitrogen and water-limited sunflower leaves. Remote Sensing Environment, 48(2): 135-146.

- Polischuk, V.P., Shadchina, T.M., Kompanetz, T.I., Budzanivskaya, I.G., Boyko, A.L. and Sozinov, A.A. (1997). Changes in reflectance spectrum characteristic of *Nicotiana debneyi* plant under the influence of viral infection. Archives of Phytopathology and Plant Protection, 31(1): 115-119.
- Price, W.C. and Bigornia, A.E. (1971). Incidence of cadang-cadang in three varieties of coconut trees of different ages. FAO Plant Protection Bulletin, 19(6): 136-137.
- Price, W.C. and Bigornia, A.E. (1972). Evidence for spread of cadang-cadang disease of coconut from tree to tree. FAO Plant Protection Bulletin, 20(6): 133-135.
- Qin, Z. and Zhang, M. (2005). Detection of rice sheath blight for in-season disease management using multispectral remote sensing. International Journal of Applied Earth Observation and Geoinformation, 7(2): 115-128.
- Randles, J.W. (1998). CCCVd-related sequences in species other than coconut. In: Report on ACIAR-Funded Research on Viroids and Viruses of Coconut Palm and Other Monocotyledons 1985-1993, 51st edition. Hanold, D. and Randles, J.W. (eds.). Centre for International Agricultural Research. Canberra, Australian. pp. 144-152.
- Randles, J.W. and Rodriguez, M.J.B. (2003). Coconut cadang-cadang viroid.
 In: Viroids. Hadidi, A., Flores, R., Randles, J.W. and Semancik, J.S. (eds.).
 Commonwealth Scientific and Industrial Research Organisation. Collingwood, Australia. pp. 233-241.
- Rekah, Y., Shtienberg, D. and Katan, J. (1999). Spatial distribution and temporal development of fusarium crown and root rot of tomato and pathogen dissemination in field soil. Phytopathology, 89(9): 831-839.
- Rodriguez, D., Fitzgerald, G.J., Belford, R. and Christensen, L. (2006). Detection of nitrogen deficiency in wheat from spectral reflectance indices and basic crop eco-physiological concepts. Australian Journal of Agriculture Research, 57(7): 781-789.
- Sankaran, S., Mishra, A., Ehsani, R., and Davis, C. (2010). A review of advanced techniques for detecting plant diseases. Computers and Electronics in Agriculture, 72(1): 1-13.
- Shibusawa, S. (1998). Precision farming and terra-mechanics. In: The fifth ISTVS Asia-Pacific regional conference in Korea. pp. 20-22.
- Sill Jr., W.H., Bigornia, A.E. and Pacumbaba, R.P. (1964). Incidence of cadangcadang disease of coconut trees of different ages and its relationship to practical control. Philippines Journal of Plant Industry, 28: 87-102.

- Sims, D.A. and Gamon, J.A. (2002). Relationships between leaf pigment content and spectral reflectance across a wide range of species, leaf structures and developmental stages. Remote Sensing of Environment, 81(2): 331-354.
- Steddom, K., Bredehoeft, M.W., Khan, M., Rush, C.M. (2005). Comparison of visual and multispectral radiometric disease evaluations of cercospora leaf spot of sugar beet. Plant Disease, 89(2): 153-158.
- Thomas, J.R. and Gausman, H.W. (1977). Leaf reflectance vs. leaf chlorophyll and carotenoid concentrations for eight crops. Agronomy Journal, 69(5): 799-802.
- Tian, Y.C., Yao, X., Yang, J., Cao, W.X., Hannaway, D.B. and Zhu, Y. (2011). Assessing newly developed and published vegetation indices for estimating rice leaf nitrogen concentration with ground-and space-based hyperspectral reflectance. Field Crops Research, 120(2): 299-310.
- Turner, P.D. (1981). Oil Palm Diseases and Disorders. Oxford University Press. Kuala Lumpur, Malaysia. p. 280.
- USDA. (2013). Oilseeds: World Markets and Trade. Circular Series. Foreign Agriculture Services. Retrieved 17th November 2013. http://www.fas.usda.gov/oilseeds/Current/
- Vadamalai, G., Perera, A., Hanold, D., Rezaian, M.A. and Randles, J.W. (2009). Detection of coconut cadang-cadang viroid sequences in oil and coconut palm by ribonuclease protection assay. Annals of Applied Biology, 154(1): 117-125.
- Vadamalai, G., Hanold, D., Rezaian, M.A., and Randles, J.W. (2006). Variants of Coconut cadang-cadang viroid isolated from an African oil palm (*Elaies guineensis* Jacq.) in Malaysia. Archives of virology, 151(7): 1447-1456.
- Vigier, B.J., Pattey, E. and Strachan, I.B. (2004). Narrowband vegetation indexes and detection of disease damage in soybeans. Geoscience and Remote Sensing Letters IEEE, 1(4): 255-259.
- Villate, L., Fievet, V., Hanse, B., Delemarre, F., Plantard, O., Esmenjaud, D. and van Helden, M. (2008). Spatial distribution of the dagger nematode *Xiphinema index* and its associated grapevine fanleaf virus in French vineyard. Phytopathology, 98(8): 942-948.
- West, J.S., Bravo, C., Oberti, R., Lemaire, D., Moshou, D. and McCartney, H.A. (2003). The potential of optical canopy measurement for targeted control of field crop diseases. Annual Review of Phytopathology, 41(1): 593-614.
- Xu, H.R., Ying, Y.B., Fu, X.P. and Zhu, S.P. (2007). Near-infrared spectroscopy in detecting leaf miner damage on tomato leaf. Biosystems Engineering, 96(4): 447-454.

- Xue, L.H., Cao, W.X., Luo, W.H., Dai, T.B. and Zhu, Y. (2004). Monitoring leaf nitrogen status in rice with canopy spectral reflectance. Agronomy Journal, 96(1): 135-142.
- Zarco-Tejada, P.J., Miller, J.R., Noland, T.L., Mohammed, G.H. and Sampson, P. (2001). Scaling up and model inversion methods with narrow-band optical indices for chlorophyll content estimation in closed forest canopies with hyperspectral data. Geoscience Remote Sensing IEEE Transactions, 39(7): 1491-1501.
- Zhu, Y., Zhou, D.Q., Yao, X., Tian, Y.C. and Cao, W.X. (2007). Quantitative relationships of leaf nitrogen status to canopy spectral reflectance in rice. Australian Journal of Agriculture Research, 58(11): 1-9.

