

UNIVERSITI PUTRA MALAYSIA

SYNTHESIS AND CHARACTERIZATION OF SILVER NANOPARTICLES MEDIATED IN GLUTATHIONE AND SODIUM ALGINATE

SEPIDEH KESHAN BALAVANDY

ITMA 2014 3



SYNTHESIS AND CHARACTERIZATION OF SILVER NANOPARTICLES MEDIATED IN GLUTATHIONE AND SODIUM ALGINATE



SEPIDEH KESHAN BALAVANDY

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfillment of the Requirements for the Degree of Master of Science

June 2014

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

To my beloved family



Thank for their support, understanding, love and encouragement.

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the Degree of Master of Science

SYNTHESIS AND CHARACTERIZATION OF SILVER NANOPARTICLES MEDIATED IN GLUTATHIONE AND SODIUM ALGINATE

By

SEPIDEH KESHAN BALAVANDY

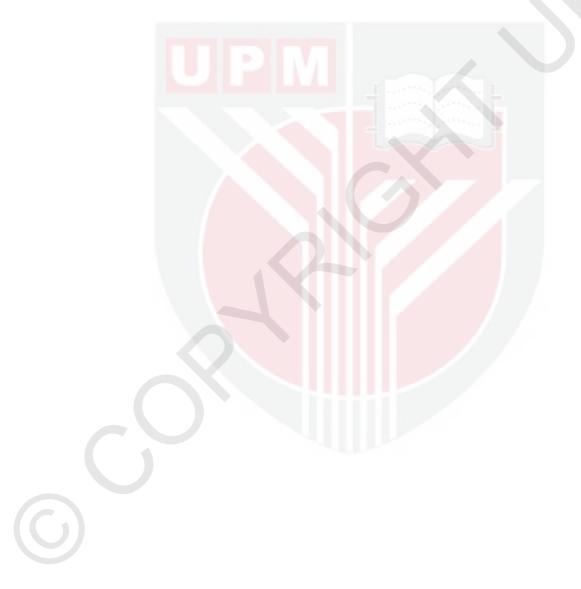
June 2014

Chairman: Associated Professor Zurina Zainal Abidin - PhD Institute: Advanced Technology

Synthesis of silver nanoparticles (Ag NPs) has attracted huge interest from scientists due to its wide applications. Different pathways, such as physical and chemical reduction has been employed regarding the synthesis of Ag NPs. Many of them contain highly reactive chemicals and can cause potential environmental and biological risks. In comparison to those methods, green synthesis of nanoparticles using biopolymer provides a safe way of nanoparticle production with the size and shape of our interest. Nanoparticles formation depends on various factors such as metal ion concentration, pH, time and temperature as well as nature of reducing and stabilizing agent. There is a lack of a comprehensive study in optimizing important parameters for the green synthesis of Ag NPs in the biopolymer substrates. In this study, silver nanoparticles with the small size of 1 to 30 nm were successfully synthesized by green method in the substrate of sodium alginate (Na-Alg) and glutathione (GSH). Specific percentage of AgNO3 and biopolymeric substance were mixed together and stirred for different time at various temperature. Moreover, rapid synthesis of Ag NPs was achieved using an environmental friendly and biodegradable solvent that act as, stabilizing and reducing agent without use of high pressure or temperature, with the help of the accelerator. The silver nitrate, sodium alginate/glutathione, and sodium hydroxide were used as the silver precursor, stabilizer/ reducing agent and accelerator respectively.

The crystalline structure of Ag NPs for all of the samples, the average size and size distributions, surface Plasmon resonance (SPR), surface morphology, and functional groups were studied using X-ray diffraction (XRD), transmission electron microscopy (TEM), UV-visible spectroscopy (UV-vis), scanning electron microscopy (SEM) and Fourier transform infrared (FT-IR) respectively. The XRD analysis confirmed that the crystallographic planes of the silver crystals were the face-centered cubic (fcc) types. The UV-visible absorption spectra showed the peaks characteristic of the surface plasmon resonance (SPR) bands of Ag NPs. The antibacterial activities of Ag NPs were investigated against Gram-negative and Gram-positive bacteria by the disk diffusion method using Mueller-Hinton Agar (MHA) that show highly antibacterial activity of Ag NPs.

The properties of Ag/Na-Alg and Ag/GSH were studied as the function of time, temperature, concentration and pH until a relatively stable size and size distribution were achieved. The results from the UV-visible spectroscopy and TEM demonstrated that the initial concentration of 0.1 M for AgNO₃ ,1.5 wt% for Na-Alg and GSH, in 90°C after 12h stirring time of reaction are the optimum for the synthesis of Ag Nps incorporate in Na-Alg and GSH. In addition, 5 ml is the optimum amount of sodium hydroxide (NaOH) for synthesis of Ag Nps in short time (30 minutes) of the reaction. From the results, all objective was achieved.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

SINTESIS DAN PENCIRIAN NANOPARTICLE PERAK MENGGUNAKAN GLUTATHIONE DAN SODIUM ALGINATE

Oleh

SEPIDEH KESHAN BALAVANDY

Jun 2014

Pengerusi: Associated Professor Zurina Zainal Abidin- PhD Institut: Teknologi Maju

Sintesis nanopartikel perak (Ag NPs) telah menarik minat sebahagian besar saintis disebabkan kegunaannya yang meluas. Pelbagai kaedah berbesa seperti secara pengurangan fizikal dan kimia telah digunakan dalam sintesis Ag NPs. Kebanyakan kaedah ini mengunakan bahan kimia yang sangat reaktif dan boleh menyebabkan risiko pencemaran alam sekitar dan biologikal. Berbanding dengan kaedah-kaedah ini, sintesis "hijau" atau mesra alam menyediakan cara yang selamat untuk menghasilkan nanopartikel dengan menggunakan biopolymer mengikut saiz dan bentuk yang dikehendaki. Pembentukan nanopartikel bergantung kepada pelbagai faktor seperti kepekatan ion logam, pH, masa, suhu dan juga sifat ejen pengurangan dan penstabilan. Daripada kajian, terdapat kekurangan kajian yang komprehensif mengenai keadaan optima bagi penghasilan Ag Nps secara "hijau" dengan menggunakan biopolimer.

Dalam kajian ini, nanopartikel perak bersaiz kecil (1 to 30 nm) telah berjaya dihasilkan melalui kaedah "hijau"dalam subtract natrium alginate (Na-Alg) dan glutasion (GSH). AgNO₃ dan bahan biopolymer dicampur bersama pada peratusan tertentu dan dikacau pada masa dan suhu yang tertentu. Tambahan pula, sintesis Ag NPS telah dicapai melalui pengunaan pelarut yang mesra alam yang bertindak sebagai ejen penstabil dan pengurang tanpa mengunakan suhu atau tekanan yang tinggi dengan bantuan ejen peluncur. Argentum Nitrat, natrium alginat / glutasion dan natrium hidroksida adalah merupakan ejen-ejen "precursor" perak, penstabil/pengurang dan peluncur mengikut urutan yang telah digunakan.

Struktur kristal Ag NPS untuk semua sampel, pengagihan saiz dan saiz purata, permukaan "Plasmon" resonan (SPR), morfologi permukaan dan kumpulan berfungsi telah dikaji dengan teknik pembelauan sinar-X (XRD), mikroskopi electron penghantaran (TEM), UV – spektroskopi (UV -vis), mikroskopi pengimbasan elektron (SEM) dan Fourier inframerah (FT- IR) masing-masing . Analisis XRD mengesahkan bahawa permukaan kristalografi Kristal perak itu adalah jenis berpusat-muka padu (fcc) . UV-spektrum penyerapan menunjukkan persamaan dengan cirri puncak resonan plasma permukaan untuk Ag NPs. Aktiviti antibakteria Ag NPS menunjukkan kadar aktivit yang tinggi setelah dikaji terhadap bakteria Gram –negatif dan Gram- positif melalui kaedah penyebaran "disk" dan Mueller-

Hinton Agar (MHA). Sifat-sifat Ag/ Na- Alg dan Ag/GSH telah dikaji untuk fungsi masa, suhu, kepekatan dan pH sehingga saiz dan taburan saiz yang agak stabil dicapai. Keputusan daripada spektroskopi UV- vis dan TEM menunjukkanbahawa kepekatan awal 0.1 M untuk AgNO3, 1.5 % berat untuk Na- ALG dan GSH, dalam 90°C selepas dikacau 12 jam adalah keadaan yang optima untuk sintesis Ag NPs dalam menggabungkan Na- Alg dan GSH. Di samping itu, 5 ml adalah jumlah natrium hidroksida(NaOH) yang optima untuk sintesis Ag Nps dalam masa tindakbalas yang singkat (30 minit).



ACKNOWLEDGEMENTS

At first I want to thank Allah for all of the things that he has given in my life and then I offer my sincerest gratitude to my chairman, Associate Professor Dr. Zurina Zainal Abidin, who has supported me throughout my thesis from the initial to the final level with her patience and knowledge. I also would like to acknowledge Dr. Dayang Radiah binti Awang Biak and Dr.Mohd Nizar b Hamidon my co-supervisors for their invaluable support and advice. Their encouraging, detailed and constructive comments have enabled me to develop an understanding of the subject. My appreciation goes especially to Dr. Kamyar Shameli, for his guidance and advice throughout the project.

Sincere thanks to Esra Ahmadi for her endless support; we have been studying and working together since September 1998 and she is my best And also Dr. Shima Shayesteh for being my friend, colleagues, and advisor.

I owe great thanks to my precious parents that leave me soon before my appreciation for all things that they gave me or taught me. And also thanks to my dearest sister Saiedeh and my darling brother Saman, for love and support they gave me always. I owe great thanks to my family in-law specially my father in-law Valiollah from distances far away; this dissertation would not have been possible without their love and encouragement during this tedious journey.

Finally, my greatest appreciation will always go to my loving family for their sacrifices, love, patience, and supports. My dear's husband Bahador Dastorian and my sweetheart daughter Diana for unending support and all their helping in during my study without them I would never able to finish my master.

At long last, I would like to dedicate this thesis to my daughter Diana.

Diana, you will always be the source of my inspiration and a part of me.

I certify that a Thesis Examination Committee has met on 26 June 2014 to conduct the final examination of Sepideh Keshan Balavandy on her thesis entitled "Synthesis and Characterization of Silver Nanoparticles Mediated in Glutathione and Sodium Alginate" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Khamirul Amin bin Matori, PhD

Senior Lecturer Faculty of Science Universiti Putra Malaysia (Chairman)

Mansor bin Hj Ahmad @ Ayob, PhD

Professor Faculty of Science Universiti Putra Malaysia (Internal Examiner)

Mohamad Amran bin Mohd Salleh, PhD

Senior Lecturer Faculty of Engineering Universiti Putra Malaysia (Internal Examiner)

Abdul Rahman Mohamed, PhD Professor Universiti Sains Malaysia Malaysia (External Examiner)

NORITAH OMAR, PhD Associate Professor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date: 19 September 2014

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Zurina binti Zainal Abidin, PhD

Associate Professor Faculty of Engineering Universiti Putra Malaysia (Chairman)

Dayang Radiah binti Awang Biak, PhD

Senior Lecturer Faculty of Engineering Universiti Putra Malaysia (Member)

Mohd Nizar b Hamidon, PhD

Associate Professor Faculty of Engineering Universiti Putra Malaysia (Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

DECLARATION

Declaration by the student

I hereby confirm that:

this thesis is my original work

quotations, illustrations and citations have been duly referenced

the thesis has not been submitted previously or comcurrently for any other degree at any institutions

intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;

written permission must be owned from supervisor and deputy vice –chancellor (Research and innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;

there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software

Signature:

Date: -

Name and Matric No: Sepideh Keshan Balavandy (GS: 31693)

Declaration by Members of Supervisory committee

This is to confirm that:

the research conducted and the writing of this thesis was under our supervision; supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: Name of Chairman of Supervisory Committee:	Signature: Name of Member of Supervisory Committee:
Signature: Name of Member of Supervisory Committee:	

TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	111
ACKNOWLEDGEMENTS	V
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xii
LIST OF FIGURES	xiii
LIST OF ABBREVIATION	xvii

CHAPTER

G

1	INT	TRODUCTION	1
	1.1	Background of the Study	1
	1.2	Research Problems	2
	1.2	Research Approach	2 3 3
		Objectives	3
2	LIT	ERATURE REVIEW	5
-	2.1		5
		Synthesis; of Silver Nanoparticles	5 5
	2.2	2.2.1 Physical Methods	6
		2.2.2 Chemical Methods	7
	23	Silver Nanoparticle incorporated in Sodium Alginate and Glutathion	
	2.5	2.3.1 Alginate and Sodium alginate Structure	10
		2.3.2 Glutathione structure	12
	2.4		13
	2.5		14
	2.6		15
	2.7		10
		Nanoparticles	15
	2.8	Characterization of silver nanoparticles	16
		2.8.1 UV-visible spectroscopy	16
		2.8.2 X-ray diffraction	17
		2.8.3 Transmission Electron Microscopy	17
		2.8.4 Field Emission Scanning Electron Microscopy	17
		2.8.5 Fourier Transform Infrared Spectroscopy	17
3	MA	ATERIALS AND METHODS	19
		Materials	19
	3.2	Preparation of Ag/Na-Alg and Ag/GSH	19
		3.2.1 Effect of different time and temperature	19
		3.2.2 Effect of different concentrations of AgNO ₃	21
		3.2.3 Effect of different concentrations of Na-Alg and GSH	22
		3.2.4 Effect of different volume of NaOH	23
		Evaluation of Antibacterial Activity	24
	3.4	Silver NPs Characterization	24

	3.4.1 UV-visible spectroscopy	24
	3.4.2 X-ray diffraction	25
	3.4.3 Transmission Electron Microscopy	25
	3.4.4 Field Emission Scanning Electron Microscopy	25
	3.4.5 Fourier Transform Infrared Spectroscopy	25
4	RESULTS AND DISCUSSION	28
	4.1 Introduction	28
	4.2 Effect of time and temperature	29
	4.2.1 UV-visible Spectroscopy	30
	4.2.2 Powder X-ray Diffraction	34
	4.2.3 Transmition Electron Microscopy	36
	4.2.4 Field Emission Scanning Electron Microscopy	37
	4.2.5 FT-IR Chemical Analysis	39
	4.2.6 Antibacterial Activity	42
	4.3 Effect of AgNO3 concentration	43
	4.3.1 UV-visible Spectroscopy	44
	4.3.2 Powder X-ray Diffraction	46
	4.3.3 Transmition Electron Microscopy	48
	4.3.4 Field Emission Scanning Electron Microscopy	50
	4.3.5 FT-IR Chemical Analysis	52
	4.3.6 Antibacterial Activity	54
	4.4 Effect of Na-Alg and GSH concentration	56
	4.4.1 UV-visible Spectroscopy	56
	4.4.2 Powder X-ray Diffraction	58
	4.4.3 Transmition Electron Microscopy	60
	4.4.4 Field Emission Scanning Electron Microscopy	64
	4.4.5 FT-IR Chemical Analysis	65
	4.4.6 Antibacterial Activity	67
	4.5 Effect of PH	69
	4.5.1 UV-visible Spectroscopy	70
	4.5.2 Powder X-ray Diffraction	70 74
	4.5.3 Transmition Electron Microscopy	74 76
	4.5.4 Field Emission Scanning Electron Microscopy	78
		80
	4.5.5 FT-IR Chemical Analysis 4.5.6 Antibacterial Activity	83
	4.5.0 Antibacterial Activity	03
5	CONCLUSION AND RECOMMENDATIONS	86
	5.1 Conclusions	86
	5.2 Recommendation for Further Studies	87
DF	FERENCES	00
		88
	ODATA OF STUDENT	102
	ST OF PUBLICATIONS	103

LIST OF TABLES

Table		Page
1.1	Typical nanostructure categories	1
2.1	Common application of alginates	11
3.1	Experimental design for synthesis of Ag/Na-Alg and Ag/GSH throughvarious time and temperature	20
3.2	Experimental design for synthesis of Ag/Na-Alg and Ag/GSH by different concentration of AgNO ₃	21
3.3	Experimental design for synthesis of Ag/Na-Alg by different concentration of Na-Alg and synthesis of Ag/GSH by different concentration of GSH	22
3.4	Experimental design for synthesis of Ag/Na-Alg Nps and Ag/GSH Nps via accelerator (NaOH)	23
4.1	Average inhibition zone for Ag/Na-Alg 90 °C/12 h (S1), Ag/GSH 90 °C/12 h (S2) and Ag/GSH 60 °C/72 h (S3)	43
4.2	Average inhibition zone for Ag/Na-Alg (0.1M AgNO ₃) 90 °C/12 h (S1) and Ag/GSH (0.1M AgNO ₃) 90 °C/12 h (S2)	55
4.3	Average inhibition zone for Ag/Na-Alg (1.5 wt% Na-Alg) 90 °C/12 h (S1), Ag/GSH (1.5 wt% GSH) 90 °C/12 h (S2)	68
4.4	The characteristics of Ag-NPs prepared in different volumes of NaOH	72
4.5	The characteristics of Ag-NPs prepared at different volumes of NaOH	73
4.6	Tverage inhibition zone for Ag/Na-Alg for 5 ml NaOH (S1), Ag/GSH for 5 ml NaOH (S2)	83

LIST OF FIGURES

Figu	re	Page
2.1	Chemical structure of alginate displayed is a polymer chain of 2 mannuronic acid (M) monomers and 2 guluronic acid (G) monomers, with (1–4) linkages	12
2.2	Photograph of brown algae(A) and Chemical structure of sodium alginate(B)	12
2.3	Glutathione structure	13
3.1	Model of laboratory experimental	20
3.2	Research overview	27
4.1	Photograph of Ag/GSH NPs suspension at different time (1, 3, 6, 12, 36, 48 and 72 h) for 60 °C (A) and 90 °C (B)	29
4.2	Photograph of AgNO ₃ /Na-Alg NPs suspension at different time (1, 3, 6, 12, 36, 48 and 72 h) for 90 °C	30
4.3	UV-visible absorption peak of Ag/Na-Alg solution for stirring time of (1, 3, 6, 12, 18, 24, 36, 48 and 72 h) at 28°C (A), 60°C (B) and 90°C (C)	31
4.4	UV-visible absorption spectra of Ag/GSH solution for stirring time of (1, 3, 6, 12, 18, 24, 36, 48 and 72 h) at 28 °C (A), 60 °C (B) and 90 °C (C)	33
4.5	X-ray diffraction patterns of sodium alginate (A) and Ag/Na-Alg NPs (B) at 90 °C after 12 h stirring time of reaction	34
4.6	X-ray diffraction patterns of Glutathione at 60 °C after 72 h stirring time of reaction	35
4.7	X-ray diffraction patterns of GSH (C) and Ag/GSH NPs (D) at 90 °C after 12 h stirring time of reaction	36
4.8	TEM images and corresponding particle size distribution of Ag/Na-Alg at 90 °C/12 h (A), Ag/GSH at 90 °C/12 h (B) and Ag/GSH at 60 °C/72 h (C)	37
4.9	SEM micrographs spectra for the Na-Alg (A), Ag/Na-Alg at 90 °C/12 h (B) and EDX	38
4.10	SEM micrographs spectra for the GSH (A), Ag/GSH at 90 °C/12 h (B), Ag/GSH at 60 °C/72 h (D) and EDXRF (C)	39
4.11	FT-IR spectra for the Na-Alg (A) and Ag/Na-Alg NPs (B) at 90 $^\circ C$ after 12 h stirring time of reaction	40

4.12	FT-IR spectra for the GSH (A) and Ag/GSH NPs (B) at 60 $^{\circ}$ C after 72 h stirring time of reaction	41
4.13	FT-IR spectra for the GSH (A) and Ag/GSH NPs (B) at 90 $^{\circ}$ C after 12 h stirring time of reaction	42
4.14	Comparison of the inhibition zone test between Na-Alg, GSH, and Ag/Na-Alg at 90 °C/12 h (S1), Ag/GSH at 90 °C/12 h (S2) and Ag/GSH at 60 °C/72 h (S3) against different bacteria	43
4.15	Photograph of AgNO ₃ /Na-Alg NPs suspension at different concentration of AgNO ₃ (0.05, 0.1, 0.2, 0.5 and 1.0 M) for $(1, 2, 3, 4)$ and 5) respectively	44
4.16	Photograph of AgNO ₃ /GSH NPs suspension at different concentration of AgNO ₃ (0.05, 0.1, 0.2, 0.5 and 1.0 M) for (1, 2, 3, 4 and 5) respectively.	44
4.17	UV-visible absorption spectra of Ag/Na-Alg NPs solution for different AgNO ₃ concentrations (0.05, 0.1, 0.2, 0.5 and 1.0 M)	45
4.18	UV-visible absorption spectra of Ag/GSH NPs solution for different AgNO ₃ concentrations $(0.05, 0.1, 0.2, 0.5 \text{ and } 1.0 \text{ M})$	46
4.19	X-ray diffraction patterns of sodium alginate (A) and Ag/Na-Alg NPs (B) with 0.1 M concentration of AgNO ₃ at 90 °C after 12 h stirring time of reaction	47
4.20	X-ray diffraction patterns of glutathione (A) and Ag/GSH NPs (B) with 0.1 M concentration of AgNO ₃ at 90 °C after 12 h stirring time of reaction	48
4.21	TEM images and relating particle size distribution of Ag/Na-Alg 0.1 M of AgNO ₃ (A) and 0.5 M AgNO ₃ (B) at 90 °C/12 h	49
4.22	TEM images and relating particle size distribution of Ag/GSH 0.1 M of AgNO ₃ (A) and 0.2 M AgNO ₃ (B) at 90 °C/12 h	50
4.23	SEM micrographs spectra for the Na-Alg (A), Ag/Na-Alg at 90 $^{\circ}$ C/12 h with 0.1 M AgNO ₃ concentration (B) and EDXRF (C)	51
4.24	SEM micrographs spectra for the GSH (A), Ag/GSH with 0.1 M AgNO ₃ concentrations at 90 °C/12 h (B), and EDXRF (C)	52
4.25	FTIR spectra for the Na-Alg (A) and Ag/Na-Alg with 0.05 M and 0.1 M AgNO ₃ concentration (B and C) respectively at 90 $^{\circ}$ C/12 h stirring time of reaction	53
4.26	FTIR spectra for the GSH (A) and Ag/GSH with (0.05, 0.1 and 0.2 M) AgNO ₃ concentration for (B, C and D) respectively at 90 $^{\circ}$ C/12 h stirring time of reaction	54

- 4.27 Comparison of the inhibition zone test between Na-Alg, GSH, Ag/Na-Alg (0.1 M AgNO₃) at 90 °C/12 h (S1) and Ag/GSH (0.1M AgNO₃) at 90 °C/12 h (S2) against different bacteria
- 4.28 Photograph of AgNO₃/Na-Alg solution at different concentration of Na-Alg (0.1, 0.5, 1.0, 1.5, 2 and 2.5 wt %)
 56
- 4.29 Photograph of AgNO₃/GSH solution at different concentration of GSH (0.5, 1.0, 1.5, 2.0 and 2.5 wt %)
 56
- UV-visible absorption spectra of Na-Alg, AgNO₃, and Ag/Na-Alg NPs solution for different Na-Alg concentrations (0.1, 0.5, 1.0, 1.5, 2 and 2.5 wt%)
- 4.31 UV-visible absorption spectra of Ag/GSH NPs solution for different GSH concentrations (0.5, 1.0, 1.5, 2 and 2.5 wt%) 58
- 4.32 X-ray diffraction patterns of sodium alginate (A) and Ag/Na-Alg NPs
 (B) with 1.5 wt% concentration of Na-Alg at 90 °C after 12 h stirring time of reaction
- 4.33 X-ray diffraction patterns of GSH (A) and Ag/GSH NPs (B) with 1.5 wt% concentration of GSH at 90 °C after 12 h stirring time of reaction 60
- 4.34 TEM images and corresponding particle size distribution of Ag/Na-Alg 0.1 wt% of Na-Alg (A), 1 wt% (B) and 1.5 wt% (c) at 90 °C/12 h 61
- 4.35 TEM images and corresponding particle size distribution of Ag/GSH 0.5 wt% of GSH (A), 1.5 wt% (B) and 2.5 wt% (c) at 90 °C/12 h 63
- 4.36 SEM micrographs spectra for the Na-Alg (A), Ag/Na-Alg at 90 °C/12 h with 1.5 wt% Na-Alg concentration (B) and EDXRF (C) 64
- 4.37 SEM micrographs spectra for the GSH (A), Ag/GSH at 90 °C/12 h with 1.5 wt% GSH concentration (B) and EDXRF (C) 65
- 4.38 FTIR spectra for the Na-Alg (A) and Ag/Na-Alg (B, C, and D) for (0.1, 1 and 1.5 wt %) Na-Alg concentration respectively at 90 °C/12 h stirring time of reaction
- 4.39 FTIR spectra for the GSH (A) and Ag/GSH (B, C and D) for (1, 1.5 and 2 wt %) GSH concentration respectively at 90 °C/12 h stirring time of reaction 67
- 4.40 Comparison of the inhibition zone test between Na-Alg, GSH, Ag/Na-Alg (1.5 wt% Na-Alg) at 90 °C/12 h (S1) and Ag/GSH (1.5 wt% GSH) at 90 °C/12 h (S2) against different bacteria
- 4.41 Photograph of Ag/Na-Alg NPs solution prepares at different concentration of NaOH (0.5, 1.0, 1.5, 5.0 and 10 ml) for S1, S2, S3, S4 and S5 respectively

2	4.42	Photograph of Ag/GSH NPs solution prepare at different concentration of NaOH (0.5, 1.0, 1.5, 5.0 and 10 ml) for S1, S2, S3, S4 and S5 respectively	70
2	4.43	The UV-visible spectra of Ag-NPs prepared in different volumes of NaOH in Na-Alg mediate	71
2	4.44	The UV-visible spectra of Ag-NPs synthesis in different volumes of NaOH in GSH mediate	73
	4.45	The PXRD patterns of Na-Alg (A) and (B, C, D and E) are Ag-NPs prepared at Na-Alg and different volumes of NaOH (0.5, 1.5, 5.0 and 10 ml) respectively	75
2	4.46	The PXRD patterns of GSH (A) and (B and C) Ag/GSH with different volumes of NaOH (3 and 5 ml) respectively	76
2	4.47	TEM images of Na-Alg /Ag-NPs and related particle size distribution at different volumes of NaOH; 1.5 (A), 5 (B), and 10 ml (C)	77
2	4.48	TEM images of AgNO ₃ /GSH and their particle size distribution at different volumes of NaOH; 3 ml (A) and 5 ml (B)	78
2	4.49	Scanning electron microscopy of sodium alginate (A), S1 (B), S4 (C) and S5 (D)	79
2	4.50	Scanning electron microscopy of Glutathione (A), S4 (B), S3(C) and EDX (D)	80
2	4.51	Fourier transform infrared spectra for Na-Alg (A), (1.5, 5 and 10 ml) are (B, C, and D) respectively	81
2	4.52	Fourier transforms infrared spectra for GSH (A), AgNO ₃ /GSH (3 ml and 5 ml) NaOH B and C respectively	82
2	4.53	Comparison of the inhibition zone test between Na-Alg, GSH, Ag/Na-Alg (5 ml NaOH) S1 and Ag/GSH (5 ml NaOH) S2 against different bacteria	84
	4.54	Reaction design of the synthesis of the Ag NPs in the Na-Alg (A) and GSH (B) solution	85

LIST OF ABBREVIATIONS

Ag NPs	Silver nanoparticles
Na-Alg	Sodium alginate
GSH	Glutathione
Ag/Na-Alg	Silver/Sodium alginate
Ag/GSH	Silver/Glutathione
fcc	face-centred cubic
SPR	Surface Plasmon Resonance
MHA	Mueller-Hinton Agar
S. aureus	Staphylococcus aureus
MRSA	methicillin-resistant Staphylococcus aureus
E. coli	Escherichia coli
TEM	Transmission electron microscopy
UV-vis	UV-visible spectroscopy
SEM	Scanning electron microscopy
FT-IR	Fourier transform infrared
PXRD	powder X-ray diffraction
XRD	X-ray diffraction
EDXRF	energy dispersive X-ray fluorescence spectrometer
SPR	Surface pelasmon resonance

CHAPTER 1

INTRODUCTION

1.1 Background of the Study

Nanotechnology is a rapidly growing field with its aim in science and technology for expanding new materials at the nano-scale level (Albrecht et al., 2006). The term nanotechnology was created by Professor Norio Taniguchi of Tokyo Science University in the year 1974 to indicate accuracy manufacturing of materials at the nanometer level (Rai et al., 2009) Nanotechnology and Nanoscience relate to the interrogation, characterization, utilization and exploitation creation. of nanostructured materials, which, at least are characterized by one dimension in the nanometer $(1 \text{ nm} = 10^{-9} \text{ m})$ range. Therefore, nanotechnology has been established as a new multidisciplinary science (Shabatina and Sergeev, 2007). The nanomaterials envelop different categories of nanostructured materials, including nanocrystals, nanowires, nanotubes, clusters, and quantum dots, while collections of nanostructures involve arrays, assemblies, and superlatives of the individual nanostructures (Rao and Cheetham, 2001).

The properties of materials with nanometer sizes are meaningfully altered from those of atoms and bulk materials. Due to their small dimensions, nanomaterials have an extremely large surface area to volume ratio, which results in more desirable properties of surface dependent material. Table 1.1 lists some of the typical dimensions of nanostructures (Jortner and Rao, 2002). In nanotechnology, a particle is described as a small object that acts as a full unit in the relations of the properties and transport. It is more classified according to size: in terms of diameter, fine particles cover a range between 100 and 2500 nm, while nanoparticles (NPs) and ultrafine particles, are between 1 and 100 nm. The nanocrystals may exhibit size-related properties that differ significantly from those detected in fine particles or bulk materials (Buzea et al., 2007).

Structure	Size Diameter (nm)	Materials	
Nanocrystals and clusters		Insulators, Metals,	
(quantum dots)	Radius: 1-10 nm	Semiconductors, Magnetic	
		materials	
Other Nanoparticles	Radius: 1-100 nm	Ceramic oxides	
Nanowires	Radius: 1-100 nm	Metals, Semiconductors,	
		oxide, sulphides, nitrides	
Nanotubes	Radius: 1-100 nm	Carbon, Layered,	
		Chalcogenides	
Nanoporous solids (pore)	0.5-30 nm	Zeolite, phosphates (etc.)	
2-Dimensional arrays	Area: Several nm2- µm2	Metals, semiconductors,	
of Nanoparticles		magnetic materials	
Surfaces and thin films	thickness 1-1000 nm	Insulators, metal, DNA	
3-Dimensional	several nm	Metals, semiconductors,	
structures		magnetic materials	

Table1.1:Typical nanostructure categories (Jortner and Rao, 2002).

In addition, metallic NPs demonstrate unusual properties that are as a result of an arrangement of high energy surface of atoms compared to bulk solid or isolated atoms (Murphy, 2008). A challenge in nanotechnology is to modify the antibacterial, electrical, and optical behaviors of nanoparticles by monitoring their size and shape. Monodisperse metal NPs are of course ideal, but unusual properties are to be expected, even if the ideality is not perfectly realized (Panigrahi et al., 2004).

Benefits of compatibility and eco-friendliness for pharmaceutical and other biomedical applications as they do not usage toxic chemicals for the synthesis procedure, grab significant attention.

Silver has long been recognized as having an inhibitory effect on microbes and bacteria present in industrial and medical process (Rai et al., 2009). The most significant application of silver nanoparticles and silver is in the medical industry such as topical ointments to prevent infection against burn and open wounds (Singh et al., 2010).

Herein, this research reports for the first time synthesis of silver nanoparticles, reducing the silver ions present in the solution of silver nitrate by sodium alginate and glutathione. Further, these biologically synthesized nanoparticles were found highly toxic against different pathogenic bacteria.

1.2 Research Problems

Silver nanoparticles often synthesized using chemical and physical methods that involves toxic chemical reagents, hazardous procedures or toxic and potentially harmful by-products involved in most of these methods thus; it has been difficult to employ these methods on large scale production (Panigrahi et al., 2004). There is a strong requirement for green, economic, commercially possible as well as environmental friendly procedure for synthesis of Ag NPs. In order to approach green synthetic techniques, several methods have been developed for the synthesis of desired Ag NPs. In spite of uncountable research, comprehensive investigations in green chemistry frame work are much more demanded. So far there has been no thoroughly investigation to optimize the most important factor (time, temperature, concentration and PH) on the green synthesis of Ag NPs by the ecofriendly and organic polymer substrates that exhibit highly antibacterial properties.

1.2 Research Approach

Green synthesis offers improvement over physical and chemical method as it is environment friendly, cost effective, easily scaled up for large scale production and in this method there is no need to use high temperature, high pressure, toxic chemicals and energy (Panigrahi et al., 2004). In these researches, using the green method Ag NPs was effectively prepared in aqueous solution of Glutathione (GSH) and Sodium Alginate (Na-Alg). The possibilities for manipulating the geometry of silver nanoparticles by altering the key growth parameters such as pH, temperature, concentrations and time have been explored. The Ag ions were reduced to the Ag NPs into the space of GSH and Na-Alg by using thermal condition. Furthermore, different time, temperature, concentrations of either silver nitrate or polymers completely investigated as well as pH effect.

Finally, Ag NPs synthesized into the biodegradable and organic mediator via green method in H₂O solvent. The crystalline structure of NPs average size, size distributions, surface morphology, surface Plasmon resonance and functional groups were characterized using powder X-ray diffraction (PXRD), transmission electron microscopy (TEM), scanning electron microscopy (SEM), UV-visible spectroscopy and Fourier transform infrared (FT-IR) respectivly.

The antibacterial activities for Ag NPs in organic substrates were investigated against Gram positive and Gram negative bacterium at different size and amount of Ag NPs.

1.3 Objectives

The main objectives of this research are:

1. To optimize effects of time, temperature, concentration and PH on the synthesis of Ag NPs by two different biopolymers, Glutathione and Sodium Alginate via green frame work of chemical reduction method.

2. To characterize the crystalline structure, average size and size distribution, surface morphology, functional groups and the surface plasmon resonance of nano particles in the optimum point

3. To investigate the antibacterial behavior of synthesized Ag NPs against Gram negative and Gram positive bacteria by Mueller-Hinton Agar (MHA) test.



REFERENCES

- Abid, J. P., Wark, a W., Brevet, P. F., and Girault, H. H. (2002). Preparation of silver nanoparticles in solution from a silver salt by laser irradiation. *Chemical Communications* 7: 792 \pm 3.
- Abu Bakar, N. H. H., Ismail, J., and Abu Bakar, M. (2007). Synthesis and characterization of silver nanoparticles in natural rubber. *Materials Chemistry and Physics*, 104: 276 £83.
- Agnihotri, S., Mukherji, S., and Mukherji, S. (2014). Size-controlled silver nanoparticles synthesized over the range 5 ± 100 nm using the same protocol and their antibacterial efficacy. *Royal Society of Chemistry*, 4: 3974 ±3983.
- Aguado, J., Arsuaga, J. M., Arencibia, A., Lindo, M., and Gascón, V. (2009). Aqueous heavy metals removal by adsorption on amine-functionalized mesoporous silica. *Journal of Hazardous Materials*, 163: 213 ±21.
- Ahmad, M., Shameli, K., and Darroudi, M. (2009). Antibacterial activity of silver/clay/chitosan bionanocomposites. *Res J Biol* ..., 4: 1156 ±1161.
- Ahmad, M., Shameli, K., Darroudi, M., Wan Md. Zin Wan, Y., Ibrahim, N. A., Rustaiyan, A., and Abdollahi, Y. (2009). Synthesis and Characterization of Silver / Clay / Chitosan Bionanocomposites by UV-Irradiation Method. *International Journal of Nanomedicine*, 6: 2030 ±2035.
- Ahmad, M., Tay, M. Y., Shameli, K., Hussein, M. Z., and Lim, J. J. (2011). Green Synthesis and Characterization of Silver/Chitosan/Polyethylene Glycol Nanocomposites without any Reducing Agent. *International Journal of Molecular Sciences*, 12: 4872 ±84.
- Aihara, N., Torigoe, K., and Esumi, K. (1998). Preparation and Characterization of Gold and Silver Nanoparticles in Layered Laponite Suspensions. *Langmuir*, 14: 4945 ±4949.
- Albrecht, M. a., Evans, C. W., and Raston, C. L. (2006). Green chemistry and the health implications of nanoparticles. *Green Chemistry*, 8: 417.
- Amanullah, M., and Yu, L. (2005). Environment friendly fluid loss additives to protect the marine environment from the detrimental effect of mud additives. *Journal of Petroleum Science and Engineering*, 48: 199 ±08.
- Article, R. (2010). Textile-based smart wound dressings. *Indian Journal of Fiber & Textile Research*, 35: 174 ±187.
- Augustine, R. (2012). Synthesis and characterization of silver nanoparticles and its immobilization on alginate coated sutures for the prevention of surgical wound infections and the in vitro release studies. *International Journal of Nano Dimension*, 2: 205 ± 212 .

- Baia, L., and Simon, S. (2007). UV-VIS and TEM assessment of morphological features of silver nanoparticles from phosphate glass matrices. *Modern Research and Educational Topics in Microscopy*. 576 ±583.
- Bar, H., Bhui, D. K., Sahoo, G. P., Sarkar, P., De, S. P., and Misra, A. (2009). Green synthesis of silver nanoparticles using latex of Jatropha curcas. *Colloids and Surfaces 339*: 134 ± 139.
- Baruwati, B., Polshettiwar, V., and Varma, R. S. (2009). Glutathione promoted expeditious green synthesis of silver nanoparticles in water using microwaves. *Green Chemistry*, 11: 926.
- Baruwati, B., Polshettiwar, V., and Varma, R. S. (2009). Glutathione promoted expeditious green synthesis of silver nanoparticles in water using microwaves. *Green Chemistry*, 11: 926 ±930.
- % HOOLQD % & RPSDJQRQ , % HUWRUHOOH) % UR\HU 0 (2011). Structural and Optical Properties of Isolated Noble Metal À Glutathione
 & RPSOH[HV×, QVLJKW LQWR WKH & KHPhe Vowmbal, oRI /LJD Physical Chemistry, 115: 24549 ±24554.
- % K D L Q V D . & D Q G ' ¶ 6 R X] D 6) ([WUDFHOOXODU E using the fungus Aspergillus fumigatus. *Colloids and Surfaces*. 47: 160 ±164.
- Ahmad, M., Lim, J. J., Shameli, K., Ibrahim, N. A., and Tay, M. Y. (2011). Synthesis of silver nanoparticles in chitosan, gelatin and chitosan/gelatin bionanocomposites by a chemical reducing agent and their characterization. *Molecules*, *16*: 7237 ±48.
- Ahmad, M., Lim, J. J., Shameli, K., Ibrahim, N. A., Tay, M. Y., and Chieng, B. W. (2012). Antibacterial activity of silver bionanocomposites synthesized by chemical reduction route. *Chemistry Central Journal*, 6: 101.
- Ahmad, M., Shameli, K., Darroudi, M., Yunus, W. M. Z. W., and Ibrahim, N. A. (2009). Synthesis and Characterization of Silver / Clay Nanocomposites by Chemical Reduction Method. *American Journal of Applied Sciences*, 6: 1909 ±1914.
- Birkholz, M. 2006. Principles of X-ray Diffraction. In *Thin Film Analysis by X-Ray Scattering* (pp. 1 ±42). Verlag Wiley
- Brown, K. R., and Natan, M. J. (1998). Hydroxylamine Seeding of Colloidal Au Nanoparticles in Solution and on Surfaces. *Langmuir*, 14: 726 ±728.
- Buzea, C., Pacheco, I. I., and Robbie, K. (2007). Nanomaterials and nanoparticles: Sources and toxicity. *Biointerphases*, 2: MR17 ±71.
- Chai, F., Wang, C., Wang, T., Li, L., and Su, Z. (2010). Colorimetric detection of Pb2+ using glutathione functionalized gold nanoparticles. *ACS Applied Materials & Interfaces*, 2: 1466 ±70.

- Chang, C., Duan, B., and Zhang, L. (2009). Fabrication and characterization of novel macroporous cellulose ±alginate hydrogels.*Polymer*, *50*: 5467 ±5473.
- Chen, C.-Y., and Chiang, C.-L. (2008). Preparation of cotton fibers with antibacterial silver nanoparticles. *Materials Letters*, 62: 3607 ±3609.
- & K H Q 3 6 R Q J / / L X < D Q G) D Q J < $6 \setminus Q$ -WayK H V L V irradiation in acetic water solution containing chitosan. *Radiation Physics and Chemistry*, 76:1165 ±1168.
- Chen, S. F., Li, J. P., Qian, K., Xu, W. P., Lu, Y., Huang, W. X., and Yu, S. H. (2010). Large scale photochemical synthesis of M@TiO2 nanocomposites (M = Ag, Pd, Au, Pt) and their optical properties, CO oxidation performance, and antibacterial effect. *Nano Research*, 3: 244 ±255.
- Cho, M., Chung, H., Choi, W., and Yoon, J. (2005). Different Inactivation Behaviors of MS-2 Phage and Escherichia coli in TiO 2 Photocatalytic Disinfection Different Inactivation Behaviors of MS-2 Phage and Escherichia coli in TiO 2 Photocatalytic Disinfection. Applied and Environmental Microbiology, 71: 270 ±275.
- Choi, S., Kim, K.-S., Yeon, S.-H., Cha, J.-H., Lee, H., Kim, C.-J., and Yoo, I.-D. (2007). Fabrication of silver nanoparticles via self-regulated reduction by 1-(2-hydroxyethyl)-3-methylimidazolium tetrafluoroborate. *Korean Journal of Chemical Engineering*, 24: 856 ±859.
- Chudasama, B., Vala, A. K., Andhariya, N., Upadhyay, R. V., and Mehta, R. V. (2010). Enhanced antibacterial activity of bifunctional Fe₃O₄-Ag core-shell nanostructures. *Nano Research*, 2: 955 ±965.
- Corazza, a, Harvey, I., and Sadler, P. J. (1996). 1H,13C-NMR and X-ray absorption studies of copper(I) glutathione complexes. *European Journal of Biochemistry* 236: 697 ±705.
- Crooks, R. M., Zhao, M., Sun, L., Chechik, V., and Yeung, L. K. (2001). Dendrimerencapsulated metal nanoparticles: synthesis, characterization, and applications to catalysis. *Accounts of Chemical Research*, 34: 181 ±90.
- Dallas, P., Niarchos, D., Vrbanic, D., Boukos, N., Pejovnik, S., Trapalis, C., and Petridis, D. (2007). Interfacial polymerization of pyrrole and in situ synthesis of polypyrrole/silver nanocomposites. *Polymer*, 48: 2007 ±2013.
- Darroudi, M., Ahmad, M. Bin, Shameli, K., Abdullah, A. H., and Ibrahim, N. A. (2009). Synthesis and characterizationof UV-irradiated silver/ montmorillonite nanocomposites. *Solid State Sciences*, 11: 1621 ±1624.
- Desai, R., Mankad, V., Gupta, S. K., and Jha, P. K. (2012). Size Distribution of Silver Nanoparticles: UV-Visible Spectroscopic Assessment. *Nanoscience and Nanotechnology Letters*, 4: 30 ±34.
- Dubas, S. T., and Pimpan, V. (2008). Optical switch from silver nanocomposite thin films. *Materials Letters*, 62: 3361 ±3363.

- Duran, N., Marcato, P. D., De Souza, G. I. H., Alves, O. L., and Esposito, E. (2007). Antibacterial Effect of Silver Nanoparticles Produced by Fungal Process on Textile Fabrics and Their Effluent Treatment. *Journal of Biomedical Nanotechnology*, 3: 203 \pm 208.
 - \hat{a} so X Evas, I. P. C., and Ius, S. T. C. (2009). Investigation of Silver Nanoparticles Formation Kinetics During Reduction of Silver Nitrate with Sodium Citrate. *Materials Science*, 15: 1 ±7.
- Eptember, S., and Iego, S. Guidelines for UV-vis Analysis, (2012), http:// www.Nanocompcsix.com(Accessed 7 Julay 2014).
- Eustis, S., Krylova, G., Eremenko, A., Smirnova, N., Schill, A. W., and El-Sayed, M. (2005). Growth and fragmentation of silver nanoparticles in their synthesis with a fs laser and CW light by photo-sensitization with benzophenone. *Photochemical & Photobiological Sciences*, 4: 154 ±9.
- Gacesa, P., Alginates *Carbohydrate Polymers*, 1988, (accessed 8 May. 2013). http://linkinghub.elsevier.com
- George, M., and Abraham, T. E. (2006). Polyionic hydrocolloids for the intestinal delivery of protein drugs: alginate and chitosan--a review. *Journal of Controlled Release*, 114: 1 ± 14 .
- Gittins, D. I., Bethell, D., Nichols, R. J., and Schiffrin, D. J. (2000). Diode-like electron transfer across nanostructured Blms containing a redox ligand. *Journal of Materials Chemistry*, 10: 79 ±83.
- Goh, C. H., Heng, P. W. S., and Chan, L. W. (2012). Alginates as a useful natural polymer for microencapsulation and therapeutic applications. *Carbohydrate Polymers*, 88:1 ±12.
- Guével, X., Spies, C., Daum, N., Jung, G., and Schneider, M. (2012). Highly fluorescent silver nanoclusters stabilized by glutathione: a promising fluorescent label for bioimaging. *Nano Research*, 5: 379 ±387.
- Guo, J.-Z., Cui, H., Zhou, W., and Wang, W. (2008). Ag nanoparticle-catalyzed chemiluminescent reaction between luminol and hydrogen peroxide. *Journal of Photochemistry and Photobiology*, 193: 89 ±96.
- Hadad, L., Perkas, N., Gofer, Y., Calderon-moreno, J., Ghule, A., and Gedanken, A. (2006). Sonochemical Deposition of Silver Nanoparticles on Wool Fibers. *Journal of Applied Polymer Science*, *104*: 1732 ±1737.
- Han, M. Y., Quek, C. H., Huang, W., Chew, C. H., and Gan, L. M. (1999). A Simple and Effective Chemical Route for the Preparation of Uniform Nonaqueous Gold Colloids. *Chem.mater*, *11*: 1144 ±1147.
- Harris, W., White, N. 2007. X-ray Diffraction Techniques for Soil Mineral Identification. In *Methods of Soil Analysis*, PP. 1-36, Gainesville, University of Florida: USA.

- He, R., Qian, X., Yin, J., and Zhu, Z. (2002). Preparation of polychrome silver nanoparticles in different solvents. *Journal of Materials Chemistry*, *12*: 3783 ±3786.
- Henglein, A. (1993). Physicochemical properties of small metal particles in solution: ³ P L F U R H O H F W U R G H ′ U H D F W L R Q V F K H P L V R U S W-to-R Q F R P metal transition. *J.phys.chem*, 97: 5457 ±5471.
- Henglein, A., and Giersig, M. (2000). 5 H G X F W L R Q R I 3 Wiffects oE Citrate and NaOH and Reaction Mechanism. *The Journal of Physical Chemistry B*, 104:6767 ±6772.
- Hu, B., Wang, S.-B., Wang, K., Zhang, M., and Yu, S.-H. (2008). Microwave-Assisted Rapid
) D F L O H ³ * U H H Q ^ć 6 \ QoWnKSHIWetL MandparticSeQ LSelf-Assembly into Multilayered Films and Their Optical Properties. *Journal of Physical Chemistry C*, 112: 11169 ±11174.
- Huggett, J. M. (1997). Field Emission Scanning Electron Microscopy A High-Resolution Technique for the Study of Clay Minerals in Sediments. *Clay Minerals*, 32: 197 ±203.
- Hussain, I., Brust, M., Papworth, A. J., and Cooper, A. I. (2003). Preparation of Acrylate-Stabilized Gold and Silver Hydrosols and Gold - Polymer Composite Films. *Langmuir*, 19: 4831 ±4835.
- Ishikawas, T. (1993). Glutathione-associated cis-Diamminedichloroplatinum (II) Metabolism and ATP-dependent Efflux from Leukemia Cells. *The Journal of Biological Ch Emistry*, 268: 20116 ±20125.
- Jacob, J. a, Mahal, H. S., Biswas, N., Mukherjee, T., and Kapoor, S. (2008). Role of phenol derivatives in the formation of silver nanoparticles. *Journal of Surfaces and Colloids*, 24: 528 ±33.
- Jain, P., and Pradeep, T. (2005). Potential of silver nanoparticle-coated polyurethane foam as an antibacterial water filter. *Biotechnology and Bioengineering*, 90: 59 ±63.
- James Meschino DC, MS, N. Ultimate Glutathione. 2009, (Acces 17 January. 2014) http://adeeva.com/index.php/products/ult_glutathione
- Jana, N. R., Gearheart, L., and Murphy, C. J. (2001). Evidence for Seed-Mediated Nucleation in the Chemical Reduction of Gold Salts to Gold Nanoparticles. *Chem.mater*, *13*: 2313 ± 2322.
- Jeong, S. H., Hwang, Y. H., and Yi, S. C. (2005). Antibacterial properties of padded PP/PE nonwovens incorporating nano-sized silver colloids. *Journal of Materials Science*, 40: 5413 ±5418.
- Jortner, J., and Rao, C. N. R. (2002). Nanostructured advanced materials. Perspectives and directions. *Pure and Applied Chemistry*, 74: 1491 ±1506.
- Jung, J. H., Cheol Oh, H., Soo Noh, H., Ji, J. H., and Soo Kim, S. (2006). Metal nanoparticle generation using a small ceramic heater with a local heating area. *Journal of Aerosol Science*, *37*: 1662 ±1670.

- Kaempfe, M., Graener, H., Kiesow, a., and Heilmann, a. (2001). Formation of metal particle nanowires induced by ultrashort laser pulses. *Applied Physics Letters*, 79:1876.
- Kalishwaralal, K., Deepak, V., Ramkumarpandian, S., Nellaiah, H., and Sangiliyandi, G. (2008). Extracellular biosynthesis of silver nanoparticles by the culture supernatant of Bacillus licheniformis. *Materials Letters*, 62: 4411 ±4413.
- Kelly, K. L., Coronado, E., Zhao, L. L., and Schatz, G. C. (2003). The Optical Properties of Metal Nanoparticles: The Influence of Size, Shape, and Dielectric Environment. J. phys.chem.B, 107:668 ±677.
- Kensuke, Akamatsu, Shingo, Ikeda. and Hidemi, N. (2003). Site-Selective Direct Silver Metallization on Surface-Modified Polyimide LayersNo Title. *Langmuir*, 19: 10366 ± 10371.
- Khomutov, G. ., and Gubin, S. . (2002). Interfacial synthesis of noble metal nanoparticles. *Materials Science and Engineering*, 22: 141 ±146.
- Köhler, J. M., Abahmane, L., Wagner, J., Albert, J., and Mayer, G. (2008). Preparation of metal nanoparticles with varied composition for catalytical applications in microreactors. *Chemical Engineering Science*, 63: 5048 ±5055.
- Korchev, a S., Bozack, M. J., Slaten, B. L., and Mills, G. (2004). Polymer-initiated photogeneration of silver nanoparticles in Speek/PVA films: direct metal photopatterning. *Journal of the American Chemical Society*, *126*: 10 ±1.
- Krishnaraj, C., Jagan, E. G., Rajasekar, S., Selvakumar, P., Kalaichelvan, P. T., and Mohan, N. (2010). Synthesis of silver nanoparticles using Acalypha indica leaf extracts and its antibacterial activity against water borne pathogens. *Colloids and Surfaces. B*, *Biointerfaces*, 76: 50 ±6.
- Kruis, F. E., Fissan, H., and Rellinghaus, B. (2000). Sintering and evaporation characteristics of gas-phase synthesis of size-selected PbS nanoparticles. *Materials Science and Engineering*, 69-70:329 ±334.
- Kvttek Libor, Panacek, Ales & Soukupova, J. (2008). Effect of Surfactants and Polymers on Stability and Antibacterial Activity of Silver Nanoparticles. *J. Phys. Chem*, 112:
- Lei, G. (2007). Synthesis of Nano-Silver Colloids and Their Anti- Microbial Effects S, master thesis, University Blacksburg, Virginia.
- Li, H., Cui, Z., and Han, C. (2009). Glutathione-stabilized silver nanoparticles as colorimetric sensor for Ni2+ ion. *Sensors and Actuators*, *143*: 87 ±92.
- Li, Q., Mahendra, S., Lyon, D. Y., Brunet, L., Liga, M. V, Li, D., and Alvarez, P. J. J. (2008). Antimicrobial nanomaterials for water disinfection and microbial control: potential applications and implications. *Water Research*, 42: 4591 ±602.

- Li, Z., Huang, H., Shang, T., Yang, F., Zheng, W., Wang, C., and Manohar, S. K. (2006). Facile synthesis of single-crystal and controllable sized silver nanoparticles on the surfaces of polyacrylonitrile nanofibres. *Nanotechnology*, *17*: 917 ±920.
- Lim, B., Jiang, M., Yu, T., Camargo, P. H. C., and Xia, Y. (2010). Nucleation and growth mechanisms for Pd-Pt bimetallic nanodendrites and their electrocatalytic properties. *Nano Research*, *3*: 69 ±80.
- Lin, S., Huang, R., Cheng, Y., Liu, J., Lau, B. L. T., and Wiesner, M. R. (2013). Silver nanoparticle-alginate composite beads for point-of-use drinking water disinfection. *Water Research*, 47: 3959 ±65.
- Link, S., and Wang, Z. L. (1999). Alloy Formation of Gold Silver Nanoparticles and the Dependence of the Plasmon Absorption on Their Composition. *The Journal of Physical Chemistry*, *103*: 3529 ±3533.
- Liu, P., and Zhao, M. (2009). Silver nanoparticle supported on halloysite nanotubes catalyzed reduction of 4-nitrophenol (4-NP). *Applied Surface Science*, 255: 3989 ±3993.
- Liu, Y., Chen, S., Zhong, L., and Wu, G. (2009). Preparation of high-stable silver nanoparticle dispersion by using sodium alginate as a stabilizer under gamma radiation. *Radiation Physics and Chemistry*, 78: 251 ±255.
- Liu, Y., Chen, S., Zhong, L., and Wu, G. (2009). Preparation of high-stable silver nanoparticle dispersion by using sodium alginate as a stabilizer under gamma radiation. *Radiation Physics and Chemistry*, 78: 251 ±255.
- Liu, Z., Wang, H., Li, H., and Wang, X. (1998). Red shift of plasmon resonance frequency due to the interacting Ag nanoparticles embedded in single crystal SiO[sub 2] by implantation. *Applied Physics Letters*, 72: 1823 ±1825.
- Lu, Y., and Chen, W. (2012). Sub-nanometre sized metal clusters: from synthetic challenges to the unique property discoveries. *Chemical Society Reviews*, 41: 3594 ±623.
- Martensson, J., Jain, A., and Meister, A. (1990). Glutathione is required for intestinal function. *PANS*, 87: 1715 ±9.
- Mayer, A. B. R., Grebner, W., and Wannemacher, R. (2000). Preparation of Silver Latex Composites. J. phys.chem.B, 104: 7278 ±7285.
- Mbhele, Z. H., Salemane, M. G., Sittert, C. G. C. E. Van, Nedeljkovic, J. M., and Luyt, A. S. (2003). Fabrication and Characterization of Silver Polyvinyl Alcohol Nanocomposites. *Chem. Mater*, *15*: 5019 ±024.
- Mohammed Fayaz, a, Balaji, K., Girilal, M., Kalaichelvan, P. T., and Venkatesan, R. (2009). Mycobased synthesis of silver nanoparticles and their incorporation into sodium alginate

films for vegetable and fruit preservation. *Journal of Agricultural and Food Chemistry*, 57: 6246 ±52.

- Mohan, Y. M., Raju, K. M., Sambasivudu, K., Singh, S., and Sreedhar, B. (2007). Preparation of Acacia- 6 W D E L O L] H G 6 L O Y H U 1 D Q R S D U Wourka Odd Xpplied * U H H O Polymer Science, 106: 3375 ±3381.
- Morones, J. R., Elechiguerra, J. L., Camacho, A., Holt, K., Kouri, J. B., Ramírez, J. T., and Yacaman, M. J. (2005). The bactericidal effect of silver nanoparticles. *Nanotechnology*, *16*: 2346 ±53.
- Mueller, B. B. L., and Ii, H. (2006). BBL TM Mueller Hinton II Agar. *BD(medical technology company)*, 16: 5-9.
- Murphy, C. J. (2008). Sustainability as an emerging design criterion in nanoparticle synthesis and applications. *Journal of Materials Chemistry*, 18: 2173 ±2176.
- Nicolet, T., and All, C., Introduction to Fourier Transform Infrared Spectrometry (*Thermo nicolet* corporation), 2001, mmrc.caltech.edu/FTIR/FTIRintro (accessed 15 Julay. 2014)
- Nimrodh Ananth, a, Umapathy, S., Ghosh, G., Ramprasath, T., and Jothi Rajan, M. a. (2012). Peptide assisted synthesis and functionalization of gold nanoparticles and their adsorption by chitosan particles in aqueous dispersion. *Advances in Natural Sciences: Nanoscience and Nanotechnology*, *3*: 045010.
- Pal, A., Esumi, K., and Pal, T. (2005). Preparation of nanosized gold particles in a biopolymer using UV photoactivation. *Journal of Colloid and Interface Science*, 288: 396 ±401.
- Pal, S., Tak, Y. K., and Song, J. M. (2007). Does the antibacterial activity of silver nanoparticles depend on the shape of the nanoparticle. A study of the Gram-negative bacterium Escherichia coli. *Applied and Environmental Microbiology*, 73: 1712 ±20.
- Pal, S., Tak, Y. K., and Song, J. M. (2007b). Does the antibacterial activity of silver nanoparticles depend on the shape of the nanoparticle. A study of the Gram-negative bacterium Escherichia coli. *Applied and Environmental Microbiology*, 73: 1712 ±20.
- Panigrahi, S., Kundu, S., Ghosh, S., Nath, S., and Pal, T. (2004). General method of synthesis for metal nanoparticles. *Journal of Nanoparticle Research*, 6: 411 ±414.
- Paper, R., Vasireddy, R., Paul, R., and Mitra, A. K. (2012). Green Synthesis of Silver Nanoparticles and the Study of Optical Properties Regular Paper. *Nanomaterials and Nanotechnology*, 2: 1 ±6.
- Pathak, S., Greci, M. T., Kwong, R. C., Mercado, K., Prakash, G. K. S., Olah, G. A., and Thompson, M. E. (2000). Synthesis and Applications of Palladium-Coated Poly (vinylpyridine) Nanospheres. *Chem.mater*, 146: 1985 ±1989.

- Pavel A Kossyrev, Aijun Yin, Sylvain G Cloutier, David A Cardimona, Danhong Huang, Paul M Alsing, J. M. X. (2005). Electric field tuning of plasmonic response of nanodot array in liquid crystal matrix.No Title. *Nano Letters*, 5: 1978 ±81.
- Peng, S., McMahon, J. M., Schatz, G. C., Gray, S. K., and Sun, Y. (2010). Reversing the sizedependence of surface plasmon resonances. *Proceedings of the National Academy of Sciences of the United States of America*, 107: 14530 ±4.
- Pileni, M. P. (1997). Nanosized Particles Made in Colloidal Assemblies. *Langmuir*, 13: 3266 ±3276.
- Pillai, Z. S., and Kamat, P. V. (2004). What Factors Control the Size and Shape of Silver Nanoparticles in the Citrate Ion Reduction Method. *The Journal of Physical Chemistry B*, 108: 945 ±951.
- Polavarapu, L., Manna, M., and Xu, Q.-H. (2011). Biocompatible glutathione capped gold clusters as one- and two-photon excitation fluorescence contrast agents for live cells imaging. *Nanoscale*, *3*: 429 ±34.
- Popa, M., Pradell, T., Crespo, D., and Calderón-Moreno, J. M. (2007). Stable silver colloidal dispersions using short chain polyethylene glycol. *Colloids and Surfaces A*, 303: 184 ± 190.
- Rai, M., Yadav, A., and Gade, A. (2009). Silver nanoparticles as a new generation of antimicrobials. *Biotechnology Advances*, 27: 76 ±83.
- Rajeshkumar, S., Kannan, C., and Annadurai, G. (2012). Synthesis and Characterization of Antimicrobial Silver Nanoparticles Using Marine Brown Seaweed Padina tetrastromatica. *Drug Invention Today*, 4: 511 ±513.
- Rao, C., and Cheetham, A. (2001). Science and technology of nanomaterials: current status and future prospects. J. Mater. Chem., 2887 ±2894.
- Raveendran, P., Fu, J., and Wallen, 6 / & R P S O H W H O \ ³ J U H H Q ´ V ` stabilization of metal nanoparticles. *Journal of the American Chemical Society*, *125*: 13940 ±13941.
- Rifai, S., Breen, C. a., Solis, D. J., and Swager, T. M. (2006). Facile in Situ Silver Nanoparticle Formation in Insulating Porous Polymer Matrices. *Chemistry of Materials*, 18: 21 ±25.
- Sabra, W., Zeng, a.-P., and Deckwer, W.-D. (2001). Bacterial alginate: physiology, product quality and process aspects. *Applied Microbiology and Biotechnology*, 56: 315 ±325.
- Saha, S., Pal, A., Kundu, S., Basu, S., and Pal, T. (2010). Photochemical green synthesis of calcium-alginate-stabilized Ag and Au nanoparticles and their catalytic application to 4-nitrophenol reduction. *Langmuir*, *26*: 2885 ±93.
- Sartori, C., Finch, D. S., and Ralph, B. (1997). Determination of the cation content of alginate by thin films. *Polymer*, *38*: 43 ±51.

- Schultz, S., Smith, D. R., Mock, J. J., and Schultz, D. a. (2000). Single-target molecule detection with nonbleaching multicolor optical immunolabels. *Proceedings of the National Academy of Sciences of the United States of America*, 97: 996 ±1001.
- Selvi, K. V., and Sivakumar, T. (2012). Isolation and characterization of silver nanoparticles from Fusarium oxysporum. *Int.J.Curr.Microbiol.App.Sci*, 1: 56 ±62.
- Shabatina, T., and Sergeev, G. (2007). Cryochemistry of nanometals. *Colloids and surfaces*, *11*: 185 ±197.
- Shameli, K., Ahmad, M. Bin, Jazayeri, S. D., Sedaghat, S., Shabanzadeh, P., Jahangirian, H., « \$EGROODKL < 6\QWKHVLVy@hQleheEkcolth@hEatwalHUL]DW silver nanoparticles by the green method. *International Journal of Molecular Sciences*, 13: 6639 ±50.
- Shameli, K., Ahmad, M. Bin, Yunus, W. Z. W., Ibrahim, N. A., and Darroudi, M. (2010). Synthesis and characterization of silver/talc nanocomposites using the wet chemical reduction method. *International Journal of Nanomedicine*, 5: 743 ±51.
- Shameli, K., Ahmad, M. Bin, Zamanian, A., Sangpour, P., Shabanzadeh, P., Abdollahi, Y., and Zargar, M. (2012). Green biosynthesis of silver nanoparticles using Curcuma longa tuber powder. *International Journal of Nanomedicine*, 7:5603 ±10.
- Shameli, K., Ahmad, M. Bin, Zargar, M., Yunus, W. M. Z. W., and Ibrahim, N. A. (2011). Fabrication of silver nanoparticles doped in the zeolite framework and antibacterial activity. *International Journal of Nanomedicine*, 6: 331 ±41.
- Shameli, K., Ahmad, M. Bin, Zargar, M., Yunus, W. M. Z. W., Rustaiyan, A., and Ibrahim, N. A. (2011). Synthesis of silver nanoparticles in montmorillonite and their antibacterial behavior. *International Journal of Nanomedicine*, 6: 581 ±90.
- Shameli, K., Bin Ahmad, M., Jaffar Al-Mulla, E. a, Ibrahim, N. A., Shabanzadeh, P., $5 X V W D L \setminus D Q$ \$ $\ll = L G D Q$ 0 $\ast U H H Q E L R V \setminus Q W K H V L$ Callicarpa maingayi stem bark extraction. *Molecules (Basel, Switzerland)*, 17: 8506 ±17.
- Shameli, K., Bin Ahmad, M., Zargar, M., Yunus, W. M. Z. W., Ibrahim, N. A., Shabanzadeh, P., and Moghaddam, M. G. (2011). Synthesis and characterization of silver/montmorillonite/chitosan bionanocomposites by chemical reduction method and their antibacterial activity. *International Journal of Nanomedicine*, 6: 271 ±284.
- Shankar, S. S., Ahmad, A., Pasricha, R., and Sastry, M. (2003). Bioreduction of chloroaurate ions by geranium leaves and its endophytic fungus yields gold nanoparticles of different shapes. *Journal of Materials Chemistry*, *13*: 1822.
- Sharma, V. K., Yngard, R. a, and Lin, Y. (2009). Silver nanoparticles: green synthesis and their antimicrobial activities. *Advances in Colloid and Interface Science*, 145: 83 ±96.
- Shen, J., Shi, M., Li, N., Yan, B., Ma, H., Hu, Y., and Ye, M. (2010). Facile synthesis and application of Ag-chemically converted graphene nanocomposite. *Nano Research*, *3*: 339 ±349.

- Shin, H. S., Yang, H. J., Kim, S. Bin, and Lee, M. S. (2004). Mechanism of growth of colloidal silver nanoparticles stabilized by polyvinyl pyrrolidone in gamma-irradiated silver nitrate solution. *Journal of Colloid and Interface Science*, 274: 89 ±94.
- Sies, H. (1999). Forum Glutathione and its role in Cellular Functions. *Free Radical Biology and Medicine*, 27: 916 ±921.
- Singh, A., Jain, D., Upadhyay, M. K., and Khandelwal, N. (2010). Green synthesis of silver nanoparticles using argemone mexicana leaf extract and evaluation of their antibacterial activity. *Digest Journal of Nanomaterials and Biostructures*, 5: 483 ±489.
- Smitha, B., Sridhar, S., and Khan, a. a. (2005). Chitosan ±sodium alginate polyion complexes as fuel cell membranes. *European Polymer Journal*, *41*: 1859 ±1866.
- Song, H. Y., Ko, K. K., Oh, I. H., and Lee, B. T. (2006). Fabrication of Silver Nanoparticles and Their Antimicrobial Mechanisms. *European Cells and Materials*, 11, 58.
- Stamplecoskie, K. G., and Scaiano, J. C. (2010). Light emitting diode irradiation can control the morphology and optical properties of silver nanoparticles. *Journal of the American Chemical Society*, *132*: 1825 ±7.
- 7 D J O L H W W L \$ 'L D]) H U Q D Q G H] < D \$ P D W R (& X F F D M. (2012). Antibacterial activity of glutathione-coated silver nanoparticles against Gram positive and Gram negative bacteria. *Langmuir*, 28: 8140 ±8.

/

- Tan, Y., Li, Y., and Zhu, D. (2003). Preparation of silver nanocrystals in the presence of aniline. *Journal of Colloid and Interface Science*, 258: 244 ±251.
- Tao, A., Sinsermsuksakul, P., and Yang, P. (2006). Polyhedral silver nanocrystals with distinct scattering signatures. *Angewandte Chemie*, 45: 4597 ±601.
- Tarun, K., and Gobi, N. (2012). Calcium alginate / PVA blended nano fibre matrix for wound dressing. *Indian Journal of Fiber & Textile Research*, *37*: 127 ±132.
- Torigoe, K., Suzuki, A., and Esumi, K. (2001). Au(III) ±PAMAM Interaction and Formation of Au ±PAMAM Nanocomposites in Ethyl Acetate. *Journal of Colloid and Interface Science*, 241: 346 ±356.
- Torres, E., Mata, Y. N., Blázquez, M. L., Muñoz, J. a, González, F., and Ballester, A. (2005). Gold and silver uptake and nanoprecipitation on calcium alginate beads. *Langmuir*, 21(17), 7951 ±8.
- Tsuji, T., Thang, D.-H., Okazaki, Y., Nakanishi, M., Tsuboi, Y., and Tsuji, M. (2008). Preparation of silver nanoparticles by laser ablation in polyvinylpyrrolidone solutions. *Applied Surface Science*, 254: 5224 ±5230.
- Umadevi, M., Shalini, S., and Bindhu, M. R. (2012). Synthesis of silver nanoparticle using D. carota extract. Advances in Natural Sciences: Nanoscience and Nanotechnology, 3: 025008.

- Underhill, R. S., and Liu, G. (2000). Preparation and Performance of Pd Particles Encapsulated in Block Copolymer Nanospheres as a Hydrogenation Catalyst. *Chemistry* of Materials, 12: 3633 ±3641.
- Vanaja, M., Gnanajobitha, G., Paulkumar, K., Rajeshkumar, S., Malarkodi, C., and Annadurai, G. (2013). Phytosynthesis of silver nanoparticles by Cissus quadrangularis: influence of physicochemical factors. *Journal of Nanostructure in Chemistry*, *3*: 17.
- Vansant, E. F. (2002). Novel Composites of TiO 2 (Anatase) and Silicate Nanoparticles. *Chem.mater*, 2: 5037 ±5044.
- Vasileva, P., Donkova, B., Karadjova, I., and Dushkin, C. (2011). Synthesis of starchstabilized silver nanoparticles and their application as a surface plasmon resonancebased sensor of hydrogen peroxide. *Colloids and Surfaces*, *382*: 203 ±210.
- Vilchis-Nestor, A. R., Sánchez-Mendieta, V., Camacho-López, M. a., Gómez-Espinosa, R. M., Camacho-López, M. a., and Arenas-Alatorre, J. a. (2008). Solventless synthesis and optical properties of Au and Ag nanoparticles using Camellia sinensis extract. *Materials Letters*, 62: 3103 ±3105.
- Wang, H., Qiao, X., Chen, J., and Ding, S. (2005). Preparation of silver nanoparticles by chemical reduction method. *Colloids and Surfaces*, 256: 111 ±115.
- Wang, J., Neoh, K. G., and Kang, E. T. (2001). Preparation of Nanosized Metallic Particles in Polyaniline. *Journal of Colloid and Interface Science*, 239: 78 ±86.
- Wang, Y., Ren, J., Deng, K., Gui, L., and Tang, Y. (2000). Preparation of Tractable Platinum, Rhodium, and Ruthenium Nanoclusters with Small Particle Size in Organic Media. *ChemInform*, 31(41), no no.
- Wiley, B., Sun, Y., Mayers, B., and Xia, Y. (2005). Shape-controlled synthesis of metal nanostructures: the case of silver. *Chemistry*, 11: 454 ±63.
- X = DQG LQ 5Nano Letters, 10: 2568 ±73.
- Xu, G., Qiao, X., Qiu, X., and Chen, J. (2008). Preparation and characterization of stable monodisperse silver nanoparticles via photoreduction. *Colloids and Surfaces*, 320: 222 ± 226.
- Yang, J., and Pan, J. (2012). Hydrothermal synthesis of silver nanoparticles by sodium alginate and their applications in surface-enhanced Raman scattering and catalysis. *Acta Materialia*, 60: 4753 ±4758.
- Yang, Q., Wang, Z., and Weng, J. (2012). Self-assembly of natural tripeptide glutathione triggered by graphene oxide. *Soft Matter*, 8(38), 9855.
- Yang, X., and Lu, Y. (2005). Preparation of polypyrrole-coated silver nanoparticles by onestep UV-induced polymerization. *Materials Letters*, 59: 2484 ±2487.

- Yang, Huaming; Du, Chunfang; Jin, Shengming; Tang, A. (2007). Preparation and characterization of SnO2 nanoparticles incorporated into talc porous materials *Materials Letters*, *61*: 3736 ±3739.
- Yeo, S. Y., Lee, H. J. O. O., and Jeong, S. H. (2003). Preparation of nanocomposite fibers for permanent antibacterial effect. *Journal of Materials Science*, *38*: 2143 ±2147.
- Yin, Y., Li, Z.-Y., Zhong, Z., Gates, B., Xia, Y., and Venkateswaran, S. (2002). Synthesis and characterization of stable aqueous dispersions of silver nanoparticles through the Tollens processElectronic supplementary information (ESI) available: photographs of silver mirror, and of stable dispersions of silver nanoparticles from mixi. *Journal of Materials Chemistry*, 12: 522 ±527.
- Yoosaf, K., James, P. V., Ramesh, A. R.Suresh, C.H. and George Thomas, K. (2007). Self-Organization of Phenyleneethynylene into Wire-Like Molecular Materials on SurfacesNo Title. *Journal of Physical Chemistry*, 111: 14933 ±14936.
- Yuvarani, I., Kumar, S. S., Venkatesan, J., Kim, S.-K., and Sudha, P. N. (2012). Preparation and Characterization of Curcumin Coated Chitosan-Alginate Blend for Wound Dressing Application. *Journal of Biomaterials and Tissue Engineering*, 2: 54 ±60.
- Zhang, L., Yu, J. C., Yip, H. Y., Li, Q., and Kwong, K. W. (2003). Ambient Light Reduction Strategy to Synthesize Silver Nanoparticles and Silver-Coated TiO 2 with Enhanced Photocatalytic and Bactericidal Activities. *Langmuir*, 19: 10372 ±10380.
- Zheng, M., Gu, M., Jin, Y., and Jin, G. (2001). Optical properties of silver-dispersed PVP thin film. pp. 853 ±859. Mater. Res. Bull. books.google.com.my
- zhu, H. Y., orthman, J. A. and li, J. Y. (2002). Novel composites of TiO2 (anatase) and silicate nanoparticles. *Chem. Mater*, 14: 5037 ±5044.