



UNIVERSITI PUTRA MALAYSIA

***NUTRITIONAL AND BIOCHEMICAL CHARACTERISTICS OF OIL PALM
(*Elaeis guineensis* Jacq.) SEEDLINGS IN RELATION TO *Ganoderma*
BASAL STEM ROT***

TENGOUA FABIEN FONGUIMGO

ITA 2014 6



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By

TENGOUA FABIEN FONGUIMGO

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirements for the Degree of Doctor of Philosophy**

July 2014

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DEDICATION

This Thesis is dedicated to
My understanding and lovely wife: Madame TENGOUA Josiane

My beloved kids:

SOBZE TENGOUA Melvis

NGUIMGO TENGOUA Ornella

MANEZEM TENGOUA Brynda

TEPIE TENGOUA Vivaldi Ryan

SONGFACK TENGOUA Hensla Warel,
for their love and patience.



Abstract of thesis submitted to the Senate of Universiti Putra Malaysia in fulfilment of the requirements for the degree of Doctor of Philosophy

NUTRITIONAL AND BIOCHEMICAL CHARACTERISTICS OF OIL PALM (*Elaeis guineensis* Jacq.) SEEDLINGS IN RELATION TO *Ganoderma* BASAL STEM ROT

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July 2014

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Institute: Tropical Agriculture

Basal stem rot (BSR) of oil palm caused by the fungus *Ganoderma boninense* is a highly damaging disease in South-east Asia. It is expanding gradually in some oil palm growing countries in Africa and South America. Up to date, available control measures have some limitations. Micronutrients known to have some beneficial effects on disease control have not been assessed on BSR yet. This study investigated the nutritional and biochemical characteristics of six oil palm progenies in relation to BSR. The optimum concentrations of boron (B), copper (Cu) and manganese (Mn) for the growth of oil palm seedlings was determined. Their subsequent effect on nutritional, biochemical and growth parameters of oil palm seedlings was tested prior to evaluating their effects on *Ganoderma* incidence and severity. The six oil palm progenies reported to respond differently to *Ganoderma* attack were found effectively different in many parameters. For instance, progenies were significantly different for their root nutrient content except for Zn. With the exception of leaf Cu, progenies also differed significantly in their leaf nutrient content. No significant difference was observed among progenies at 6-7 months for lignin in roots, but by 16-17 months, lignin content in roots of progenies significantly differed. All enzyme activities were significantly different in roots of oil palm progenies at 6-7 months. At 16-17 months, progenies significantly differed only for peroxidase activity. Two (2) mg B/mL and 2 mg Cu/mL of culture solution were identified as optimum concentrations for the growth of oil palm seedlings. All the tested concentrations of Mn (5, 10, 15 and 20 mg/mL) were phytotoxic, but 2 mg Mn/mL was maintained for subsequent studies to maintain nutrient balance. The single and combined concentrations of the selected micronutrients on oil palm seedlings generally increased SPAD chlorophyll value, plant height, and plant biomass compared with the control (no B, no Cu, and no Mn), suggesting the importance of B, Cu and Mn for the growth of oil palm seedlings. Apart from the control, no treatment was consistently higher or lower than the others for the studied

parameters. Hence, all the treatments were formulated in forms of fertilizers and tested on *Ganoderma* incidence and severity. Treatment T9 (B + Cu + Mn) in general gave the poorest performance for most growth and physiological parameters. Double combinations of treatments, T6 (B + Cu), T7 (B + Mn) and T8 (Cu + Mn) generally performed better than other inoculated treatments for nearly all the parameters assessed. In conclusion, a proper nutritional environment may effectively reduce *Ganoderma* incidence and severity; and the double combination of micronutrients may be more effective than individual nutrients or their triple combination.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

CIRI PEMAKANAN DAN BIOKIMIA ANAK KELAPA SAWIT (*Elaeis guineensis* Jacq.) BERKAITAN REPUT PANGKAL BATANG *Ganoderma*

Oleh

TENGOUA FABIEN FONGUIMGO

Julai 2014

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Reput pangkal batang (BSR) kelapa sawit yang disebabkan oleh kulat *Ganoderma boninense* adalah penyakit sangat serius di Asia Tenggara. Ia berkembang secara beransur-ansur di beberapa negara yang ditanami kelapa sawit di Afrika dan Amerika Selatan. Sehingga kini, langkah-langkah kawalan yang ada sangat terhad dan tidak memberi kesan yang memuaskan. Unsur-unsur pemakanan mikro diketahui memberi kesan yang baik pada kawalan penyakit belum dinilai lagi pada penyakit BSR. Kajian ini ditumpukan kepada ciri-ciri pemakanan dan biokimia enam progeni kelapa sawit berkaitan dengan BSR. Kepekatan optimum boron (B), kuprum (Cu) dan mangan (Mn) untuk pertumbuhan anak kelapa sawit ditentukan. Kesan berikutnya terhadap pemakanan, biokimia dan pertumbuhan parameter anak kelapa sawit telah diuji sebelum menilai kesannya terhadap keterukan penyakit *Ganoderma*. Hasilnya, enam progeni kelapa sawit bertindak balas secara berbeza kepada serangan *Ganoderma* dalam banyak parameter yang disukat. Sebagai contoh, progeni berbeza secara ketara dalam kandungan nutrien akar kecuali Zn. Untuk daun kelapa sawit, kecuali Cu, semua progeni menunjukkan perbezaan yang ketara untuk semua kandungan nutrien. Tiada perbezaan yang ketara diperhatikan di kalangan progeni pada bulan ke 6-7 untuk lignin dalam akar, tetapi pada bulan ke 16-17, kandungan lignin dalam akar progeni berbeza dengan ketara. Semua aktiviti-aktiviti enzim berbeza secara ketara dalam akar progeni kelapa sawit pada bulan ke 6-7. Pada bulan ke 16-17, semua progeni ketara berbeza hanya untuk aktiviti peroksidase. Pada kepekatan 2 mg B/mL dan 2 mg Cu/mL telah dikenal pasti sebagai kepekatan optimum untuk pertumbuhan anak kelapa sawit. Kesemua kepekatan Mn diuji (5, 10, 15 dan 20 mg/mL) didapati fitotoksik, tetapi 2 mg Mn/mL dikekalkan untuk kajian seterusnya bagi keseimbangan nutrien. Ujian kepekatan yang telah dicampur satu mikronutrien dipilih pada anak kelapa sawit memberi hasil keseluruhan yang baik untuk nilai SPAD klorofil, ketinggian tumbuhan, dan biomas tumbuhan, kecuali kawalan (tiada B, Cu, dan Mn), menunjukkan kepentingan unsur-unsur tersebut bagi pertumbuhan anak kelapa sawit. Selain daripada kawalan, rawatan adalah lebih

tinggi secara konsisten atau lebih rendah daripada yang lain untuk parameter tersebut. Oleh itu, kesemua unsur tersebut telah dirumuskan dalam bentuk baja dan diuji ke atas anak kelapa sawit bagi menguji kejadian dan keterukan serangan *Ganoderma*. Rawatan T9 (B + Cu + Mn) secara umumnya memberikan nilai yang tidak memuaskan pada parameter pertumbuhan dan fisiologi. Secara keseluruhannya, gabungan dua rawatan, T6 (B + Cu), T7 (B + Mn) dan T8 (Cu + Mn) memberikan prestasi yang lebih baik daripada lain-lain rawatan yang diinokulat pada hampir semua parameter dinilai. Sebagai kesimpulannya, penambahan B, Cu dan Mn adalah berkesan bagi mengurangkan kejadian dan keterukan penyakit *Ganoderma* dan kombinasi dua antara nutrien lebih berkesan daripada nutrien individu dan/atau gabungan ketiga-ketiga nutrien.



ACKNOWLEDGEMENTS

I would first like to express my deep gratitude to Dr. Claude Bakoume and his wife, Madame Olive Bakoume, whose multipurpose and incommensurable sacrifices made this thesis possible.

Special and heartiest thanks are due to the members of my supervisory committee: Professor Dr. Mohamed Hanafi Musa for his endless advices and guidance, and his availability at anytime and anywhere, to attend to me and answer my concerns; Associate Professor Dr. Syed Omar Syed Rastan for his valuable advice and encouragement and, through him, DIVERSATECH (M) Sdn. Bhd., whose financial support rescued me from the edge of abyss, when I was exhausted and about to give up my PhD programme and go back to my country; Dr. Idris Abu Seman for his valuable advice and assistance, and his devotion to always find a solution to my concerns. I take this opportunity to express my kind appreciation to his staff, Mr. Rosmidi Miswan, Mrs. Noorhasimah Ismail, Mr. Naim Mohammad Shahrul Hasan, Mr. Mazlan Ismail, Mr. Safaruddin Alhamidi, and Mr. Mohd Shukri just to name a few, he always sent to assist me at critical times.

The School of Graduate Studies (SGS) Universiti Putra Malaysia (UPM) is gratefully acknowledged for having granted me with a Graduate Research Assistance (GRA), which, unfortunately, lasted for only four months because of the exhaustion of their funds.

It is a pleasure to record my very grateful thanks to Barbara Ritchie from CABI UK, whose didactic support made my research easy. Associate Professor Dr. Jugah Kadir deserves special thanks for helping in the epidemic analysis. Associate Professor Dr. Husni Ahmad Mohd Hanif also deserves special mention for his continuous advice and encouragement. I am indebted to Professor Dr. Mohd Rafii and Associate Professor Dr. Anuar Abdul Rahim for their precious guidance in the interpretation of statistical results of my data, and to Dr. Tristan Durand-Gasselin from PalmElit France for his availability to supply me with any reference needed.

I am immensely happy to thank Dr. Zeufack Albert Gaspard, my brothers Ateufack Benoît (ATEBE), Kenfack Richard Bilau, Temgoua Joseph (KAHAM), and my sister Madame Ndongmo Florence for their substantial support.

I would like to express my sincere appreciation to Dr. Jose Alvaro Cristancho Rodriguez, Dr. Beyegue Djonko Honore, and Mr. Alagie Bah for their encouragement and assistance in statistical analysis.

I will never forget the encouragement and support of my friends Dr. Naghme Nejat, Dr. Sahar Shahnazi Sangachin, and Dr. Ng Lee Chuen. Special thanks go to my

friends Ms. Tan Siao Hue and Mr. Chen Xingwei for their kind co-operation in field work.

I would like to extend my deep gratitude to the staff of the Institute of Tropical Agriculture (ITA) Central Laboratory, Mr. Zainudin Mohd Ali, Mr. Zahardin Zulkifli, Mrs. Umami Kalthum Abdullah, and Mrs. Nor Rafidah Mohd Yusoff, the staff of ITA office, Mrs. Norashima Sulaiman, Mr. Fadhli Zil Ikram Omar, Mr. Mohd Yusof Ramli, and Ms. Nor Shuhada Mohamad, the staff of Field 2 UPM, Mr. Abdol Rahman Sharif, Mr. Osman Saleh, Mr. Mohd Khalid Ismail, and Mrs. Krishnaveni Lechimanan, for their sincere collaboration.

Mr. Jamil Omar from Soil Analytical Laboratory 2, Mrs. Zabedah Tumirin from Soil Chemistry Laboratory 2, Mrs. Umi Kalthum Asmaon from Soil Chemistry Laboratory 3, and Mrs. Siti Samsiah from the Crop Science Laboratory are highly and especially appreciated for their technical and valuable cooperation and help.

The kind assistance of Mr. Suhaimi Aman and Mr. Daud Mustam in lignin staining in the Botany Laboratory, and Mr. Saparin Demin and Mr. Khairul Anwar Bahari for lignin quantification in the Animal Nutrition Laboratory is highly appreciated.

I deeply thank my parents for their affection and moral support.

My friends Mr. Tagni Tepie Samuel, Mr. Tazanou Martin, Dr. Baba Mohamad, Mr. Shamsuddeen Rufai, Mrs. Hasmah Mohidin, Mr. Mohammad Reza Mohammadi, and Dr. Mahbod Sahebi are sincerely acknowledged for their encouragements, assistance in data collection, and collaboration in laboratory work.

My utmost thanks are addressed to my senior brother Tadontsa Edouard, now of late, the only one who used to call me once in a while to know about my progress. Unfortunately, he will never see the output of my devotion, perseverance and endurance, because he was suddenly called to serve the Lord.

At last, but not the least, I say thank you so much to all my other friends, brothers and sisters, who, by one way or another, contributed to make these studies successful.

This thesis was submitted to the Senate of the Universiti Putra Malaysia and has been accepted as fulfilment of the requirements for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

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LIST OF ABBREVIATIONS

%	Percent
°C	Degree Celcius
AA	Auto-analyzer
AAS	Atomic absorption spectrophotomer
ABTS	2, 2'-azino-bis (3-ethylbenzo-thiazoline-6-sulfonic acid)
ADP	Adenosine diphosphate
AIL	Acid insoluble lignin
Al	Aluminium
ANOVA	Analysis of variance
ASL	Acid soluble lignin
AUDPC	Area under the disease progress curve
B	Boron
BRIS	Beach ridges interspersed with swales
BSR	Basal stem rot
Ca	Calcium
Ca(OH) ₂	Copper hydroxide (slaked lime)
CaO	Calcium oxide
CDC	Cameroon development corporation
cm	Centimetre
CPO	Crude palm oil
Cu	Copper
CuCO ₃	Copper carbonate
CuSO ₄	copper (II) sulphate
D × P	<i>Dura × Pisifera</i>
DAB	Diaminobenzidine
DI	Disease incidence
DMRT	Duncan's Multiple Range Test
DOT	Disodium octaborate tetrahydrate
DR	Disease reduction
DSI	Disease severity index
DSIB	Disease severity index for bulb symptoms
DSIF	Disease severity index for foliar symptoms
DSIR	Disease severity index for root symptoms
EC	Enzyme code
EDTA	Ethylene diamine tetraacetic acid
ER	Epidemic rate
FAO	Food and Agricultural Organization of the United Nations
Fe	Iron
FELDA	Federal Land Development Authority
FFB	Fresh fruit bunches
g	Gram
g/L	Gram per litre
GSM	<i>Ganoderma</i> -selective medium
H ₂ O ₂	Hydrogen peroxide
H ₂ SO ₄	Sulphuric acid
ha	Hectare
HCl	Hydrochloric acid
hr	Hour

HRGP	hydroxyproline-rich glycoproteins
IAA	Indol acetic acid
K	Potassium
kg	Kilogram
LAC	Laccase
LSD	Least significant difference
M	Molar
MEA	Malt extract agar
mg	Milligram
Mg	Magnesium
mg/kg	Milligram per kilogram
mg/L	Milligram per litre
min	minute
mL	Millilitre
mM	Millimolar
mm	Millimetre
Mn	Manganese
MnCl ₂	Manganese chloride
MnO(OH)	Manganite
MnO ₂	Pyrolusite
MnSO ₄	Manganese sulphate
Mo	Molybdenum
MPOB	Malaysian Palm Oil Board
N	Nitrogen
NaOH	Sodium hydroxide
NH ₃	Ammonia
nm	Nanometre
O ₂	Oxygen
ODW	Oven-dry weight
P	Phosphorus
PAL	Phenylalanine ammonium-lyase
PDA	Potato dextrose agar
pH	Hydrogen potential
PH	Plant height
<i>P_N</i>	Net photosynthetic rate
POX	Peroxidase
ppm	Part per million
PS I	Photosystem I
PS II	Photosystem II
PVP	polyvinyl pyrrolidone
PVPP	polyvinyl polypyrrolidone
RCBD	Randomized complete block design
RDW	dry weight
RFW	Root fresh weight
RNA	Ribonucleic acid
RS	Root surface
RT	Root tips
RV	Root volume
RWB	Rubber wood block
SDSAS	Sime Darby Seeds and Agricultural Services

SDW	Shoot dry weight
SFS	Severity of foliar symptoms
SFW	Shoot fresh weight
SPAD Chl	SPAD Chlorophyll
TDW	Total dry weight
TL	Total lignin
TLA	Total leaf area
TRL	Total root length
US\$	United States dollars
UV	Ultra violet
Zn	Zinc
μL	Micro litre



CHAPTER 1

INTRODUCTION

1.1 Background Information

Oil palm (*Elaeis guineensis* Jacq.) is a perennial oil crop that exists in wild, semi-wild, and cultivated states in the equatorial tropics of Africa, South-East Asia, and the Americas (Hartley, 1988). The total area planted in oil palms estimated at 11×10^6 ha with 70% exploited by smallholders (Rival, 2007), has rapidly expanded. As a globally important crop, total land under oil palm cultivation has more than quadrupled, moving from less than 4×10^6 ha in 1961 to about 15×10^6 ha across the world (FAO, 2009; Turner et al., 2011; Anonymous; 2011). In many developing countries, oil palm is an alternative to cocoa, coffee and rubber, the traditional cash crops whose prices regularly fluctuate in the world market (Bakoume et al. 2002). In Africa, the oil palm grower is the first consumer of his palm oil or kernel oil and the excess is easily sold in the local market.

In 2008, the major vegetable oil production was 111.127 million tonnes. Palm oil contributed about 40% and ranked first just before soybean oil (33%), and accounted for about 67% of the world exports (Jackson et al., 2009). World palm oil production multiplied 15-fold since 1948 to reach 38×10^6 tonnes in 2007 (Rival, 2007). South-East Asia (Malaysia and Indonesia) contributed 86% of the global palm oil production. Malaysia, the second largest world's palm oil producer after Indonesia contributed 10.3% of the world oils and fats market with 15.82 million tonnes of the 154 million tonnes of oils and fats in 2007 (Global Oil and Fats, 2008). But oil palm is still an important source of income in Africa and Latin America (Billotte, 2004). An increase in world demand for edible palm oil is predicted as a result of future increases in the world population, the increase in per capita consumption of oils and fats, and development of the bio-diesel industry. Palm oil is poised to contribute significantly to meet this demand in view of its high yield of 4-5 tonnes per hectare per year (Barison and Ma, 2000); almost three times the yield of coconut and more than 10 times that of soybean (0.4 tonne per hectare) (Rajanaidu and Jalani, 1994). Furthermore, the production cost of palm oil in its ecosystem is the lowest compared to all other oil crops (Billotte, 2004).

Further improvement in palm oil production in the world is governed not only by the implementation of new plantations, the regeneration of old plantings, and the availability of high yielding planting materials, but also by good pest and disease control measures to fill the yield gap existing between field trials and plantations (Jalani et al., 2003). In South-East Asia, the major constraint to the oil palm industry is *Ganoderma*, a soil-borne fungus which leads to yield reduction to the affected oil palm and also to death. *Ganoderma* was also declared a serious pathogen in Cameroon (Tengoua and Bakoume, 2005) and was becoming serious in replanting areas (Tengoua, 2005).

1.2 Problem Statement

Basal stem rot due to *Ganoderma* spp is the major disease causing serious damage to oil palm in Papua New Guinea and the Pacific Islands (Flood and Hasan, 2004; Pilloti, 2005), and in Southeast Asia, namely in Malaysia and Indonesia (Chenon, 1975; Ariffin, 1990; Singh, 1991). In Africa, Cameroon is currently the only country where *Ganoderma* basal stem rot exists as a major disease. Dead palms due to *Ganoderma* were estimated at 53.22% in 25-year-old or older first generation plantations (Tengoua and Bakoume, 2005). It caused 6.4% of plant losses in 1995 and 1996 replantings in the Cameroon Development Corporation (CDC) plantations (Tengoua, 2005). In Malaysia, Singh (1991) reported plant losses as high as 85% on coastal soils by the time palms were replanted at 25 years. In Latin America, the existence of BSR has been confirmed (Martinez and Arango, 2013) even though the extent of damage is not yet determined. If no appropriate action is taken to control the disease, in the not-too-distant future, BSR will become a great concern in all the oil palm growing countries in the world. Unfortunately, to date, no definitive solution is available. The few control methods being implemented include: (i) Use of a balanced fertilizer, namely N, P, K (Turner and Poon, 1968; Mohd Tayeb et al., 2003), (ii) Manual application of calcium nitrate (Hendry, 1997; Sariah and Zakaria, 2000; Flood and Hasan, 2004), (iii) Excision of infected tissues, (iv) Mounding around the stem base to stimulate root production and provide additional support, the shredding of diseased palms into small fragments and spreading out instead of windrowing (Wan, 2007), (v) Digging of trenches to prevent mycelium spread of the pathogen (Flood and Hasan, 2004), (vi) Use of systemic fungicides (Ramasamy, 1972; Jollands, 1983; Khairudin, 1990a; Khairudin, 1990b; Lim et al., 1990) and natural fungicide (Nurfaezah et al., 2012), and (vii) Cultural techniques during replanting such as sanitation and clean clearing (Idris et al., 2004; Flood et al., 2005).

Biological control of the fungus is one of the pest management strategies with bright a prospect compared to chemical pesticides. It is also an environmentally safe alternative. Some microorganisms shown to have antagonistic action against *Ganoderma* include *Trichoderma* spp (Ilias, 2000; Sariah et al., 2005; Shamala, 2005; Izzati and Faridah, 2008; Siddiquee et al., 2009; Shamala et al., 2013; Susanto, 2013), *Aspergillus*, *Penicillium* spp, Arbuscular mycorrhiza (Idris and Ariffin, 2004; Shamala et al., 2013), *Gliocladium* (Flood and Hasan, 2004) and a non-pathogenic strain of *Ganoderma*, GanoEF1 (Idris et al., 2010). Bacteria, such as *Pseudomonas fluorescens* and *Bacillus* sp (Susanto et al., 2005; Susanto, 2013), *Pseudomonas aeruginosa* and *Burkholderia cepacia* (Zaiton et al., 2008; Bivi et al., 2010; Shamala et al., 2011) and actinomycetes (Lim et al., 2013) have also been involved in biological control of *Ganoderma* BSR. Plant extracts have also been tested against this pathogen (Noor Pahtiwati et al., 2013); but none of the above mentioned methods has yet been satisfactory in maintaining disease incidence at an acceptable threshold. Many of these methods efficiently work at laboratory and nursery levels, but face serious limitations for field application probably due to the variation in environmental conditions. This holds true especially for the use of *Trichoderma* as a bio-control agent whose population drastically drops in oil palm plantations from 10^6 to less than 10^3 cfu, rendering the application of the method ineffective or its maintenance at an efficient level uneconomical. Likewise, implementation of

chemical control measures is not cost effective and is environmentally unfriendly with regard to the amount of chemicals needed to treat a few million hectares of oil palm plantations. Breeding for tolerance to *Ganoderma* is an optimistic prospect since there are putative resistant materials in African *Elaeis guineensis* collections (Idris *et al.*, 2004; Durand-Gasselino *et al.*, 2005), but much is still required before the release of *Ganoderma* tolerant planting materials. Diversity of strains due to mutations requires a wide range of putative tolerant oil palm materials. Unfortunately, current sources of partial tolerance are found only in limited oil palm materials to permit an effective breeding programme. Owing to these limitations, thinking about developing alternative cost effective and environmentally sound control measures such as improvement of the oil palm defence system through balanced fertilizer (with required quantity and quality of micronutrients) and lignification becomes an imperative.

Ganoderma is a white rot fungus that degrades lignin to water and carbon dioxide and uses cellulose as a nutrient (Siti *et al.*, 2004; Paterson, 2007; Paterson *et al.*, 2008). Hence, a comprehensive study of the mode of action of *Ganoderma* and setting up of new strategies would allow the development of an efficient integrative control measure. This includes the reinforcement of cell walls by improving the lignification process to create a stronger physical barrier against pathogen penetration. The differences in susceptibility observed in some oil palm progenies may be related to differences in their lignin content (Paterson *et al.*, 2008). Lignification can be improved through the manipulation of certain nutritional factors directly or indirectly involved in the process. Boron, copper and manganese play an important role in lignin biosynthesis, in addition to their known biocidal effect on a number of plant pathogens and their other functions in the plant.

1.3 Research Objectives

The general objective was to examine nutritional status of oil palm seedlings with special reference to micronutrients B, Cu, and Mn to see whether their manipulation can reduce BSR disease. The specific objectives were: (i) to determine the nutrient status and biochemical characteristics of six oil palm progenies reported to behave differently toward *Ganoderma*, (ii) to determine the optimum concentrations of B, Cu and Mn for the growth of oil palm seedlings, (iii) to assess the effects of single and combined optimum concentrations of B, Cu and Mn on nutritional, biochemical and growth parameters of oil palm seedlings, and (iv) to test the effects of single and combined optimum concentrations of B, Cu and Mn on *Ganoderma* incidence and severity.

1.4 Outline of the Thesis

Since developing partially resistant material is a long term approach, it is imperative and essential that oil palm scientists continue to investigate other alternative

management strategies. Over many years, micronutrient application has been totally overlooked in oil palm fertilization programmes. This appears to have weakened oil palms through exhaustion of soil reserves of these nutrients, and predisposed this crop to certain diseases that could be controlled by a balanced nutrient status. This study was carried out to identify the missing nutritional factors with emphasis on micronutrients that could be manipulated to control *Ganoderma* BSR in oil palm. With the nutrients identified, a genetic approach as stated by Amtmann et al. (2008), can be applied to establish a causal relationship between susceptibility/tolerance on the one hand, and nutrient assimilation capability on the other hand. When clearly identified, this information can be used to design agricultural strategies that support the nutritional status of the oil palm while exploiting their inherent potential for defence against BSR disease. After stating the problem with some background information in Chapter 1, a brief review on oil palm, *Ganoderma*, lignin, boron, copper, manganese, and their importance in plant disease is presented in Chapter 2. In Chapter 3, six oil palm progenies reported to behave differently towards *Ganoderma* BSR were examined to identify nutritional and biochemical characteristics that could explain differences observed when challenged with *Ganoderma boninense*. In view toward advising the incorporation of micronutrients in the oil palm fertilization programme with regard to their importance in growth and their potential role in plant defence against pests and diseases, different concentrations of boron, copper and manganese were tested in Chapter 4 to identify their optimum for the growth of oil palm seedlings. In Chapter 5, the optimum concentrations of selected micronutrients identified in Chapter 4 were tested singly and in different possible combinations on nutritional, biochemical and growth parameters of oil palm seedlings to select the best treatment (s) to be assessed with *Ganoderma*. With no treatment being distinctively and consistently better than others for the parameters tested, both single and combined optimum concentrations were formulated in forms of fertilizer treatments and examined for their effects on *Ganoderma* incidence and severity in Chapter 6. Chapter 7 summarizes the results obtained in this work, presents the conclusions and proposes some recommendations for further research.

REFERENCES

- Ariffin, D. and Idris, A. S. 1992. The *Ganoderma*-selective medium (GSM). Palm Oil Research Institute of Malaysia (PORIM) Information Series ISSN 0128-5726. 2 pp.
- Ariffin, D., Idris, A. S. and Singh, G. 2000. Status of *Ganoderma* in oil palm. In: Flood, J., Bridge, P. D. and Holderness, M. (eds). *Ganoderma disease of perennial crops*. CABI Publishing, UK. pp 49-68.
- Bakoume C., Jannot C., Rafflegeau S., Ndigui B. and Weise S. 2002. Etudes complémentaires pour la relance des filières hévéa et palmier. Revue du secteur rural. Rapport palmier. Institut de la Recherche Agricole pour le Développement. Yaoundé. 80 p.
- Barison, Y. and Ma, A. N. 2000. Malaysian palm oil industry: Moving towards the future. *Palm Oil Development*, 32: 1-9.
- Basiron, Y. 2007. Palm oil production through sustainable plantations. *European Journal of Lipid Science and Technology*, 109: 289-295.
- Beardmore, J., Ride, J. P. and Granger, J. W. 1983. Cellular lignifications as a factor in the hypersensitive resistance of wheat to stem rot. *Physiological Plant Pathology*, 22: 209-220.
- Bechinger, C., Giebel, K. F., Schnell, M., Leiderer, P., Deising, H. B. and Bastmeyer, M. 1999. Optical measurements of invasive forces exerted by appressoria of a plant pathogenic fungus. *Science*, 285: 1896-1899.
- Benton Jones, J. Jr. 2003. *Agronomic handbook: Management of crops, soils, and their fertility*. CRC Press LLC, Florida, USA. 450 p.
- Bhuiyan, N. H., Selvaraj, G., Wei, Y. and King, J. 2009a. Gene expression profiling and silencing reveal that monolignol biosynthesis plays a critical role in penetration defence in wheat against powdery mildew invasion. *Journal of Experimental Botany*, 60 (2): 509-521.
- Bhuiyan, N. H., Selvaraj, G., Wei, Y. and King, J. 2009b. Role of lignification in plant defense. *Plant Signalling and Behaviour*, 4 (2): 158 – 159.
- Billotte, N. 2004. Recherche et étude des locus contrôlant les caractères à déterminisme génétique complexe (QTL) du palmier à huile (*Elaeis guineensis* Jacq.) par cartographie génétique multiparentale. Thèse de Doctorat. Ecole Nationale Supérieure Agronomique de Montpellier. 102 p.
- Bivi, M. R., Farhana, M. S. N., Khairulmazmi, A. and Idris, A. 2010. Control of *Ganoderma boninense*: A causal agent of basal stem rot disease in oil palm with endophyte bacteria in vitro. *International Journal of Agriculture and Biology*, 12: 833-839.

- Blanchette RA. 1994. Lignin biodegradation in cell walls of woody plants. In: Petrini O, Ouellette GB (eds). *Host wall alterations by parasitic fungi*. APS Press, St Paul Minnesota, USA. pp 55-65.
- Boerjan, W., Ralph, J. and Baucher, M. 2003. Lignin biosynthesis. *Annual Review of Plant Biology*, 54: 519-546.
- Boudet, A. M., Lapierre, C. and Grima-Pettenati, J. 1995. Biochemistry and molecular biology of lignification. *New Phytologist*, 129: 203-236.
- Boudet A. M. 2000. Lignins and lignification: selected issues. *Plant Physiology and Biochemistry*, 38 (1/2): 81 – 96.
- Breton, F., Hasan, Y., Hariadi., Lubis, Z. and de Franqueville, H. 2006. Characterization of parameters for the development of an early screening test for basal stem rot tolerance in oil palm progenies. *Journal of Oil Palm Research*, Special Issue-April 2006: 24-36.
- Broadley, M., Brown, P., Cakmak, I., Rengel, Z. and Zhao, F. 2012. Functions of nutrients: Micronutrients. In: Marschner, P. (ed). *Marschner's Mineral nutrition of higher plants*. Third Edition. Academic Press., USA. pp 191-248.
- Brown, G. E. and Barmore, C. R. 1983. Resistance of healed citrus exocarp to penetration by *Penicillium digitatum*. *Phytopathology*, 73: 691-694.
- Brown, G. E. 1989. Host defenses at the wound site on harvested crops. *Phytopathology*, 79 (5): 1381-1384.
- Brown, L. R. 2006. Plan B 2.0: *Rescuing a planet under stress and a civilization in trouble*. NY: W. W. Norton and Co. Earth Policy Institute. 343 p.
- Brown, P. H., Bellaloui, N., Wimmer, M. A., Bassil, E. S., Ruiz, J., Hu, H., Pfeffer, H., Dannel, F. and Romheld, V. 2002. Boron in plant biology. *Plant Biology*, 4: 205-223.
- Burnell, J. N. 1988. The biochemistry of manganese in plants. In: Graham, R. D., Hannam, R. J. and Uren, N. C. (eds). *Manganese in soils and plants*. Kluwer Academic Publishers, Dordrecht, The Netherlands. pp 125-137.
- Cadena-Gomez, G. and Nicholson, R. L. 1987. Papilla formation and associated peroxidase activity: A non-specific response to attempted fungal penetration of maize. *Physiology and Molecular Plant Pathology*, 31: 51-67.
- Cahill, D. M. and McComb, J. A. 1992. A comparison of changes in phenylalanine ammonia-lyase activity, lignin and phenolic synthesis in the roots of *Eucalyptus calophylla* (field resistant) and *E. marginata* (susceptible) when infected with *Phytophthora cinnamomi*. *Physiological and Molecular Plant Pathology*, 40: 315-332.

- Campbell, C. L. and Madden, L. V. 1990. *Introduction to plant disease epidemiology*. John Wiley and Sons, Inc. USA. 532 p.
- Carter, C., Finley, W., Fry, J., Jackson, D. and Willis, L. 2007. Palm oil markets and future supply. *European Journal of Lipid Science and Technology*, 109: 307-314.
- Carver, T. L. W., Robbins, M. P. and Zeyen, R. J. 1991. Effects of two PAL inhibitors on the susceptibility and localized autofluorescent host cell responses of oat leaves attacked by *Erisiphe graminis* D. C. *Physiological and Molecular Plant Pathology*, 39: 269-287.
- Carver, T. L.W., Robbins, M. P., Zeyen, R. J., and Dearne, G. A. 1992a. Effects of PAL-specific inhibition on suppression of activated defence and quantitative susceptibility of oats to *Erisiphe graminis* D. C. *Physiological and Molecular Plant Pathology*, 41: 149-163.
- Carver, T. L.W., Zeyen, R. J., Robbins, M. P. and Dearne, G. A. 1992b. Effects of PAL inhibitor, AOPP, on oat, barley and wheat cell responses to appropriate and inappropriate formae specialis of *Erisiphe graminis* D. C. *Physiological and Molecular Plant Pathology*, 41: 397-409.
- Carver, T. L.W., Zeyen, R. J., Robbins, M. P., Vance C. P. and Boyles, D. A. 1994a. Suppression of host cinnamyl alcohol dehydrogenase and phenylalanine ammonia-lyase increase oat epidermal cell susceptibility to powdery mildew penetration. *Physiological and Molecular Plant Pathology*, 44: 243-259.
- Carver, T.L.W., Zeyen, R.J., Bushnell W.R. and Robbins, M.P. 1994b. Inhibition of phenylalanine ammonia-lyase and cinnamyl alcohol dehydrogenase increases quantitative susceptibility of barley to powdery mildew (*Erisiphe graminis* D.C). *Physiological and Molecular Plant Pathology*, 44: 261-272.
- Cervilla, L. M., Rosales, M. A., Rubio-Wilhelmi, Sánchez-Rodríguez, E., Blasco, B., Ríos, J. J., Romero, L. and Ruiz, J. M. 2009. Involvement of lignification and membrane permeability in the tomato root response to boron toxicity. *Plant Science*, 176: 545-552.
- Chen, E-L., Chen, Y-A., Chen, L-M. and Liu, Z-H. 2002. Effect of copper on peroxidase activity and lignin content in *Raphanus sativus*. *Plant Physiology and Biochemistry*, 40: 439-444.
- Chen, F., Reddy, M.S.S., Temple, S., Jackson, L., Shadle, G. and Dixon, R.A. 2006. Multi-site genetic modulation of monolignol biosynthesis suggests new routes for formation of syringyl lignin and wall-bound ferulic acid in alfalfa (*Medicago sativa* L.). *The Plant Journal*, 48: 113-124.
- Chenon de, R. D. 1975. Presence in Indonesia and Malaysia of a Lepidoptera oil palm root miner, *Sufetula sunidesalis* Walker, and its relationship to attacks by *Ganoderma*. *Oléagineux*, 30 (11): 449-456.

- Chigrin, V. V., Rozu, L. V. and Zaprometov, M. N. 1973. Phenolcarboxylic acid and lignin in leaves of resistant and sensitive varieties of spring wheat during infection with stem rust. *Sovietic Plant Physiology*, 20 (5): 801-806.
- Chmielowska J., J. Deckert and J. Diaz, 2008. Activity of peroxidases and phenylalanine ammonia-lyase in lupine and soybean seedlings treated with copper and an ethylene inhibitor. *Biological Lett*, 45: 59 – 67.
- Chmielowska, J., Veloso, J., Gutiérrez, J., Silvar, C. and Díaz, J. 2010. Cross-protection of pepper plants stressed by copper against a vascular pathogen is accompanied by the induction of a defence response. *Plant Science*, 178: 176-182.
- Chong, K. P. 2010. The roles of phenolics in the interaction between oil palm and *Ganoderma boninense* the causal agent of basal stem rot. PhD Thesis, University of Nottingham. 178 p.
- Corley, R. H. V. and Tinker, P. B. 2003. *The oil palm*. Fourth edition. Blackwell Science Ltd. 562 p.
- Dean, R. A. and Kuć, J. 1987. Rapid lignification in response to wounding and infection as a mechanism for induced systemic protection in cucumber. *Physiological and Molecular Plant Pathology*, 31: 69-81.
- Dogbo, D. O., Gogbeu, S. J., N'zue, B., Yao, K. A., Zohouri, G. P., Bekro, J. A. M. and Bekro, Y.-A. 2012. Comparative activities of phenylalanine ammonia-lyase and tyrosine ammonia-lyase and phenolic compounds accumulated in cassava. *African Crop Science Journal*, 20 (2): 85-94.
- Dordas, C. 2008. Role of nutrients in controlling plant diseases in sustainable agriculture. A review. *Agronomy for Sustainable Development*, 28: 33-46.
- Douglas, C. J. 1996. Phenylpropanoid metabolism and lignin biosynthesis: From weeds to trees. *Trends in Plant Science*, 1: 171-178.
- Durand-Gasselin, T., Asmady, H., Flori, A., Jacquemard, J. C., Hayun, Z., Breton, F. and de Franqueville, H. 2005. Possible sources of genetic resistance in oil palm (*Elaeis guineensis* Jacq.) to basal stem rot caused by *Ganoderma boninense* – Prospects for future breeding. *Mycopathologia*, 159: 93-100.
- Engels, C., Kirkby, E. and White, P. 2012. Mineral nutrition, yield and source-sink relationships. In: Marschner, P. (ed). *Marschner's Mineral nutrition of higher plants*. Third Edition. Academic Press., USA. pp 85-133.
- Epstein, E. and Bloom, A. J. 2005. *Mineral nutrition of plants: Principles and perspectives*. Second edition. Sinauer Associates, Inc. Massachusetts, USA. 400 p.
- Eschbach J. M., 1980. Micronutrients in oil palm. *Oleagineux*, 35: 291 – 294.

- Evans, I., Solberg, E. and Huber, D. M. 2007. Copper and plant disease. In: Datnoff, L. E., Elmer, W. H. and Huber, D. M. (eds) *Mineral nutrition and plant disease*. APS Press St. Paul, Minnesota, USA. pp 177-188.
- Fageria, V. D. 2001. Nutrient interactions in crops. *Journal of Plant Nutrition*, 24 (8): 1269-1290.
- Fairhurst T. and R. Härdter, 2003. *Oil palm: Management for large and sustainable yields*. Potash and Phosphate Institute, Potash and Phosphate Institute of Canada and International Potash Institute. 382 p.
- Fairhurst, T., Caliman, J.-P., Härdter, R. and Witt, C. 2005. *Oil palm: Nutrient disorders and nutrient management*. Potash and Institute (PPI)/Potash and Institute of Canada (PPIC) and International Potash institute (IPI); French Agricultural Research Centre for International Development (CIRAD); and Pacific Rim Palm Oil Limited (PRPOL). 67 p.
- FAO. 2009. FAOSTAT online statistical service. Food and Agriculture Organization of the United Nations, Rome. <http://faostat.fao.org/> (Accessed August 2011).
- Fitzherbert, E. B., Struebig, M. J., Morel, A., Danielsen, F., Brühl, C. A., Donald, P. F. and Phalan, B. 2008. How will oil palm expansion affect biodiversity? *Trends in Ecology and Evolution*, 23 (10): 538-545.
- Flood, J. and Hasan Y. 2004. Basal stem rot – Taxonomy, biology, epidemiology, economic status and control in South East Asia and Pacific Islands. Paper 8. International conference on pests and diseases of importance to the oil palm industry. Fostering Global Cooperation in Instituting Quarantine Shield. Kuala Lumpur, Malaysia. 18-19 May 2004. 18 p.
- Flood, J., Keenan, L., Wayne, S. and Hasan Y. 2005. Studies of oil palm trunks as sources of infection in the field. *Mycopathologia*, 159: 153-157.
- Gavnholt, B. and Larsen, K. 2002. Molecular biology of plant laccases in relation to lignin formation. *Physiologia Plantarum*, 116: 273-280.
- Glenn, J. K., Akileswaran, L. and Gold, M. H. 1986. Mn (II) oxidation is the principal function of the extracellular Mn-peroxidase from *Phanerochaete chrysosporium*. *Archives of Biochemistry and Biophysics*, 251: 688-696.
- Global Oil and Fats. 2008.
- Goh, K.-J. and Härdter, R. 2003. General oil palm nutrition. In: Fairhurst, T. and Härdter, R. (eds). *Oil palm: Management for large and sustainable yield*. Potash and Institute (PPI), Potash and Institute of Canada (PPIC) and International Potash institute (IPI), pp 191-228.

- Grabber, J. H., Ralph, J., Hatfield, R. D. and Quideau, S. 1997. p-Hydroxyphenyl, guaiacyl, and syringyl lignins have similar inhibitory effects on cell wall degradability. *Journal of Agriculture and Food Chemistry*, 45: 2530-2532.
- Graham, R. D. 1980. Susceptibility to powdery mildew of wheat plants deficient in copper. *Plant and Soil*, 56: 181-185.
- Graham, R. D. 1983. Effects of nutrient stress on susceptibility of plants to disease with particular reference to the trace elements. *Advances in Botanical Research*, 10: 221-276.
- Graham, R. D. and Webb, M. J. 1991. Micronutrients and disease resistance and tolerance in plants. In: Mortvedt, J. J., Cox, F. R., Shuman, L. M. and Welch, R. M. (eds). *Micronutrients in agriculture*. Second edition. Soil Science Society of America, Inc. Madison, Wisconsin, USA. pp 329-370.
- Guo, D., Chen, F., Inoue, K., Blount, J. W. and Dixon, R. A. 2001. Downregulation of caffeic acid 3-O-methyltransferase and caffeoyl CoA 3-O-methyltransferase in transgenic alfalfa: Impacts on lignin structure and implications for the biosynthesis of G and S lignins. *The Plant Cell*, 13: 73-88.
- Gupta U. C., 2007. Boron. In: Barker, A.V. and Pilbeam, D.G. (eds). *Handbook of plant nutrition*. Taylor and Francis Group, LLC. CRC Press, New York, USA. pp 241-277.
- Gupta, P.K. 2007. *Soil, plant, water and fertilizer analysis*. Second edition. AGROBIOS (India). 350 p.
- Halpin, C., Holt, K., Chojecki, J., Oliver, D., Chabbert, B., Monties, B., Edwards, K., Barakate, A. and Foxon, G.A. 1998. Brown-midrib maize (bm 1) - a mutation affecting the cinnamyl alcohol dehydrogenase gene. *The Plant Journal*, 14 (5): 545-553.
- Hamid, M. and Khalil-ur-Rehman. 2009. Potential applications of peroxidases. *Food Chemistry*, 115: 1177-1186.
- Hartley, C.W.S. 1988. *The Oil Palm (Elaeis guineensis Jacq.)*. New York: Longman Scientific and Technical Publication.
- Hasan, Y. and Turner, P. D. 1998. The comparative importance of different oil palm tissue as infection sources for basal stem rot in replantings. *The Planter*, 74: 119-135.
- Hatfield, R. and Fukushima, R. S. 2005. Can lignin be accurately measured? *Crop Science*, 45: 832-839.
- Havlin, J. L., Beaton, J. D., Tisdale, S. L. and Nelson, W. L. 1999. *Soil fertility and fertilizers-An introduction to nutrient management*. Sixth edition. Prentice Hall, Inc. New Jersey, USA. 499 p.

- Heldt H-W. 2005. *Plant biochemistry*. Third edition. Elsevier Academic Press. California, USA. 630 p.
- Hendry, J. 1997. The effect of calcium nitrate (Ca(NO₃)₂) on the incidence of basal stem rot (BSR) on oil palm seedlings (*Elaeis guineensis* Jacq.). Bachelor of Agricultural Science Project Report. Faculty of Agriculture, Universiti Putra Malaysia, Serdang, Selangor Malaysia. 44 p.
- Hess, D. 1975. *Plant physiology. Molecular, biochemical, and physiological fundamentals of metabolism and development*. Springer-Verlag. New York, Inc. USA. 333 p.
- Ho, Y. W. and Nawawi, A. 1985. *Ganoderma boninense* Pat. from basal stem rot of oil palm (*Elaeis guineensis*) in Peninsula Malaysia. *Pertanika Journal of Tropical Agricultural Science*, 8: 425-428.
- Hoagland D. R. and D. I. Arnon, 1950. The water – culture method for growing plants without soil. California Agricultural Experiment Station, Circular 347. 32 p.
- Hoffland, E., Jeger, M. J. and van Beusichem, M. L. 2000. Effect of nitrogen supply rate on disease resistance in tomato depends on the pathogen. *Plant and Soil*, 218: 239-247.
- Huber, D. M. and Watson, R. D. 1974. Nitrogen form and plant disease. *Annual Review of Phytopathology*, 12: 139-165.
- Huber, D. M. and Wilhelm, N. S. 1988. The role of manganese in resistance to plant diseases. In: Graham, R. D., Hannam, R. J. and Uren, N. C. (eds). *Manganese in soils and plants*. Kluwer Academic Publishers, Dordrecht, The Netherlands. pp 155-173.
- Huber, D. M. 1989. The role of nutrition in the take-all disease of wheat and other small grains. In: Engelhard, A. W. (ed). *Soilborne plant pathogens: Management of diseases with macro- and microelements*. APS Press, St Paul, Minnesota, USA. pp 46-74.
- Huber, D. M. and Jones, J. B. 2013. The role of magnesium in plant disease. *Plant and Soil*, 368: 73-85.
- Hückelhoven, R. 2007. Cell wall-associated mechanisms of disease resistance and susceptibility. *Annual Review of Plant Pathology*, 45: 101-127.
- Idris, A. S. 1999. Basal stem rot (BSR) of oil palm (*Elaeis guineensis* Jacq.) in Malaysia: Factors associated with variation in disease severity. PhD Thesis. Wye College, University of London, UK.
- Idris, A. S., Ariffin, D., Swinburne. T. R. and Watt T. A. 2000. The identification of *Ganoderma* species responsible for basal stem rot (BSR) disease of oil palm

in Malaysia – Morphological characteristics. MPOB Information series No 102, TT No 77a. 4 p.

- Idris, A. S. and Ariffin, D. 2004. Basal stem rot-Biology, detection, and control. aper 9. International conference on pests and diseases of importance to the oil palm industry. Fostering Global Cooperation in Instituting Quarantine Shield. Kuala Lumpur, Malaysia. 18-19 May 2004. 35 p.
- Idris, A. S., Kushairi, A., Ismail, S. and Ariffin, D. 2004. Selection for partial resistance in oil palm progenies to *Ganoderma* basal stem rot. *Journal of Oil Palm Research*, 16 (2): 12-18.
- Idris, A. S. 2009. Basal stem rot in Malaysia-Biology, economic importance, epidemiology, detection and control. In: Proceedings of MPOB-IOPRI International Workshop on Awareness, Detection and Control of Oil Palm Devastating Diseases. November 2009, Kuala Lumpur Convention Centre (KLCC), Kuala Lumpur Malaysia. 48 p.
- Idris, A. S., Noor Haida, S. and Nur Rashyeda, R. 2010. GanoEF1-A fungal biocontrol agent for *Ganoderma* in oil palm. MPOB Information Series No.501. 4 p.
- Idris, A. S. 2011. Further advances in oil palm research. In: Basri, M. W., Choo, Y. M. and Chan, K. W. (eds). *Other devastating diseases of oil palm*. Malaysian Palm Oil Board (MPOB). pp 522-542.
- Idris, A. S., Mior, M. H. A. Z., Maizatul, S. M. and Kushairi, A. 2011. Survey on status of *Ganoderma* disease of oil palm in Malaysia 2009-2010. In: Proceedings of the PIPOC 2011 International Palm Oil Conference. “Palm Oil: Fortifying and Energizing the World”. Agriculture, Biotechnology and Sustainability Conference. pp 235-238.
- Ilias, G. N. M. 2000. *Trichoderma* Pers. Ex Fr. and its efficacy as a bio-control agent of basal stem rot of oil palm (*Elaeis guineensis* Jacq.). PhD Thesis, Faculty of Science and Environmental Studies, Universiti Putra Malaysia, Selangor, Malaysia. 283 p.
- Izzati, N. A.M. Z. and Faridah, A. 2008. Disease suppression in *Ganoderma*-infected oil palm seedlings treated with *Trichoderma harzianum*. *Plant Protection Science*, 44 (3): 101-107.
- Jackson, D., Fry, J., Fitton, C., Hickingbottom, S., Hogg, M., Fereday, N. and Finley, W. 2009. *Oil-Seeds, Oils and Meals Analysis, March 2009*. LMC International Ltd. 16 p.
- Jalani, B. S., Chan, K. W. and Ranajaidu, N. 2003. Contributions of Breeding and agronomy in increasing oil palm yield. Paper presented at ISOPB seminar on The Progress of Oil Palm Breeding and Selection. Medan, Sumatra, Indonesia. 6-9 October 2003.

- Janezic, T. S. 2002. Small diameter forest residues for soil rehabilitation. In: Grover, V. I., Grover, V. K. and Hogland, W. (eds). *Recovering energy from waste: Various aspects*. Science Publishers, Inc. Enfield (NH), USA. pp 49-73.
- Johnson, D. 1980. Tree crops and tropical development: the oil palm as a successful example. *Agricultural Administration*, 7: 107-112.
- Jollands, P. 1983. Laboratory investigations on fungicides and biological agents to control three diseases of rubber and oil palm and their potential applications. *Tropical Pest Management*, 29: 33-38.
- Jones, J. P. and Woltz, S. S. 1970. *Fusarium* wilt of tomato: Interaction of soil liming and micronutrient amendment on disease development. *Phytopathology*, 60: 812-813.
- Jones, Jr. J. B., Wolf B. and Mills, H. A. 1991. *Plant analysis handbook. A practical sampling, preparation, analysis and interpretation guide*. Micro-Macro Publishing, Inc. Athens, Georgia, USA. 213 p.
- Jourdan, C. and Rey, H. 1997. Modelling and simulation of the architecture and development of the oil palm (*Elaeis guineensis* Jacq.) root system. *Plant and Soil*, 190: 235-246.
- Judin, J., Hamid, N. and Weng, C. C. 2009. Preliminary screening for *Ganoderma* tolerant oil palm progenies in FELDA. In: Proceedings of Agriculture, biotechnology and sustainability conference. Malaysian Palm Oil Board (MPOB) International Palm Oil Congress (PIPOC) 2009, Palm Oil-Balancing Ecologies with Economics, 9-12 November 2009, Kula Lumpur Convention Centre (KLCC), Kuala Lumpur Malaysia. pp 779-789.
- Junejo, N., Khanif, M.Y., Hanafi, M. M., Wan, M. Z. W. Y. and Dharejo, K. A. 2010. Maize response to biodegradable polymer and urease inhibitor coated urea. *Intational Journal of Agriculture and Biology*, 12: 773-776.
- Kandan, A., Radjacommar, R., Ramanathan, A., Raguchander, T., Balasubramanian, P. and Samiyappan, R. 2009. Molecular biology of *Ganoderma* pathogenicity and diagnosis in coconut seedlings. *Folia Microbiology*, 54 (2): 147-152.
- Keča, N. and Keča, L. 2012. The efficiency of rotstop and sodium borate to control primary infections of *Heterobasidion annosum* to *Picea abies* stumps: A Serbia study. *Baltic Forestry*, 18 (2) 35: 247-254.
- Khairudin, H. 1990a. Basal stem rot of oil palm: Incidence, etiology and control. Master of Agricultural Science Thesis, Faculty of Agriculture, University Pertanian Malaysia, Selangor, Malaysia. 147 p.
- Khairudin, H. 1990b. Results of four trials on *Ganoderma* basal stem rot of oil palm in Golden Hope Estates. In: Ariffin, D. and Jalani, S. (eds). *Proceedings of*

the Ganoderma Workshop, 11 September 1990, Palm Oil Research Institute of Malaysia, Bangi, Selangor, Malaysia. pp 113-131.

- Khairudin H., 1993. Basal stem rot of oil palm caused by *Ganoderma boninense*. An update. In: Jalani, B. S., Ariffin, D., Rajanaidu, N., Mohd Tayed, D., Paranjothy, K., Mohd Basri, W., Henson, I. E. and Chong, K. C. (eds). *Proceedings of the 1993 PORIM International Palm Oil Congress "Update and Vision" Agriculture*, pp: 735 – 738. Palm Oil Research Institute of Malaysia, Kuala Lumpur, Malaysia.
- Khairudin, H. and Tey, C. C. 2008. An overview of the current status of *Ganoderma* basal stem rot and its Management in a large scale plantation group in Malaysia. *The Planter*, 84 (988): 469-482.
- Koh, L. P. and Wilcove, D. S. 2008. Is oil palm agriculture really destroying tropical biodiversity? *Conservation Letters*, Blackwell Publishing, Inc. 5 p.
- Kongsager, R. and Reenberg, A. 2012. Contemporary land-use transitions: The global oil palm expansion. Global Land Project (GLP) Report No. 4. Global Land Project (GLP)-International Project Office (IPO), Copenhagen. 39 p.
- Kruger, W. M., Carver, T. L. W. and Zeyen, R.J. 2002. Effects of inhibiting phenolic biosynthesis on penetration resistance of isolines containing seven powdery mildew resistance genes or alleles. *Physiology and molecular Plant Pathology*, 1: 41-51.
- Kunamneni, A., Ballesteros, A., Plou, F.J. and Alcalde, M. 2007. Fungal laccase-Aversatile enzyme for biotechnological applications. In: Méndez-Vilas, A. (ed). *Communicating current research and educational topics and trends in applied microbiology*.
- Kunamneni, A., Camarero, S., García-Burgos, C., Plou, F.J., Ballesteros, A. and Alcalde, M. 2008. Engineering and applications of fungal laccasee for organic synthesis. *Microbial Cell Factories*. 7:32:<http://www.microbialcellfactories.com/content/7/1/32>.
- Lewis, D. H. 1980. Boron, lignification and the origin of vascular plants-A unified hypothesis. *New Phytologist*, 84: 209-229.
- Li, L., Popko, J. L., Zhang, X.-H., Osakabe, K., Tsai, C.-J., Joshi, C. P. and Chiang, V. L. 1997. A novel multifunctional O-methyltransferase implicated in a dual methylation pathway associated with lignin biosynthesis in loblolly pine. *Proceeding of National Academy of Science USA*, 94: 5461-5466.
- Li, X., Weng, J.-K. and Chapple, C. 2008. Improvement of biomass through lignin modification. *The Plant Journal*, 54: 569-581.
- Lim, P. K., Jualang, A. G. and Chong, K. P. 2013. Potential of microorganisms isolated from forest soil to control *Ganoderma* spp. of oil palm. In: *Proceedings of the 5th MPOB-IOPRI International Seminar: Sustainable*

management of pests and diseases in oil palm-The way forward. 22-23 November 2013, Kuala Lumpur Convention Centre (KLCC), Kuala Lumpur, Malaysia. pp 345-351.

- Lim, T.K., Hamm, R. and Mohamad, R. 1990. Persistency and volatile behavior of selected chemical in treated soil against three basidiomycetes root disease pathogens. *Tropical Pest Management*, 36: 23-26.
- Lin, C-C., Chen L-M. and Liu, Z-H. 2005. Rapid effect of copper on lignin biosynthesis in soybean root. *Plant Science*, 168 (3): 855-861.
- Lin, J., He, X., Hu, Y., Kuang, T. and Ceulemans, R. 2002. Lignification and lignin heterogeneity for various age classes of bamboo (*Phyllostachys pubescens*) stems. *Physiologia Plantarum*, 114: 296-302.
- Liu, L., Dean, J. F. D. and Friedman, W. E. 1994. A laccase-like phenoloxidase is correlated with lignin biosynthesis in *Zinnia elegans* stem tissues. *The Plant Journal*, 6 (2): 213-224.
- Lloyd, J. D. and Pratt, J. E. 1996. The use of disodium octaborate tetrahydrate to control conifer butt rot caused by *Heterobasidion annosum*. In: Proceedings of Crop Protection in Northern Britain. pp: 135-139.
- Lybeer, B. and Koch, G. 2005. A topochemical and semiquantitative study of the lignification during ageing of bamboo culms (*Phyllostachys viridiglaucescens*). *IAWA Journal*, 26 (1): 99-109.
- Mäder, M. and Füssl. R. 1982. Role of peroxidase in lignification of tobacco cells. *Plant Physiology*, 70: 1132-1134.
- Madhavi, V. and Lele, S. S. 2009. Laccase: properties and applications. *BioResources*, 4(4): 1694-1717.
- Maher, E. A., Bate, N. J., Ni, W., Elkind, Y., Dixon, R. A. and Lamb, C. J. 1994. Increased disease susceptibility of transgenic tobacco plants with suppressed levels of preformed phenylpropanoid products. *Proceedings of the National Academy of Science USA*, 91: 7802-7806.
- Marschner, H. 1986. *Mineral nutrition of higher plants*. First edition. Academic Press, London, UK. 674 p.
- Marschner, P. 2012. *Marschner's Mineral nutrition of higher plants*. Third edition. Academic Press, Waltham, USA. 651 p.
- Martinez, G. and Arango, M. 2013. Detection of basal stem rot in infected oil palms using a Picus Sonic Tomograph. In: Proceedings of the 5th MPOB-IOPRI International Seminar: Sustainable management of pests and diseases in oil palm-The way forward. 22-23 November 2013, Kuala Lumpur Convention Centre (KLCC), Kuala Lumpur, Malaysia. pp 309-313.

- Masoero, F., Moschini, M., Rossi, F., Prandini, A. and Pietri, A. 1999. Nutritive value, mycotoxin contamination and in vitro rumen fermentation of normal and genetically modified corn (cry1A(b)) grown in northern Italy. *Maydica*, 44 (3): 205-209.
- Mauch-Mani, B. and Slusarenko, A. J. 1996. Production of salicylic acid precursors is a major function of phenylalanine ammonia-lyase in the resistance of Arabidopsis to *Peronospora parasitica*. *The Plant Cell*, 8: 203-212.
- Mayer, A. M. and Staples, R. C. 2002. Laccase: new functions for an old enzyme. *Phytochemistry*, 60: 551-565.
- Mielke, T. 2012. *Oil World Annual 2012*, Vol. 1. ISTA Mielke GmbH, Hamburg, Germany.
- Mills, H. A. and Jones Jr. J. J. B. 1996. *Plant analysis handbook II. A practical sampling, preparation, analysis and interpretation guide*. Micro-Macro Publishing, Inc. Athens, Georgia, USA. 422 p.
- Mlíčková, K., Luhová, L., Peč, P. and Lebeda, A. 2004. The role of active oxygen species in defense of *Lycopersicon* spp. against *Oidium neolycopersici*. Acta Fytotechnica et Zootechnica. Vol. 7, Special Number. Proceedings of the XVI Slovak and Czech Plant Protection Conference organized by Slovak Agricultural University in Nitra, Slovakia.
- Moerschbacher, B. M., Noll, U., Gorrichon, L. and Reisener, H.-J. 1990. Specific inhibition of lignifications breaks hypersensitive resistance of wheat to stem rust. *Plant Physiology*, 93: 465-470.
- Mohd Tayeb, D. and Hamdan, A. B. 1999. Relation of fertilizer nutrient to *Ganoderma*. In: Proceedings of the 1999 PORIM International Palm Oil Congress, PORIM, Bangi. pp 422-452.
- Mohd Tayeb, D., Idris, A. S. and Mohd Haniff, H. 2003. Reduction of *Ganoderma* infection I oil palm through balanced fertilizer in peat. In: Proceedings of Agricultural Conference PIPOC 2003. 24-24 August 2003 Putrajaya Marriott Hotel, Malaysia. pp 193-219.
- Mohsina Hamid and Khalil-ur-Rehman. 2009. Potential applications of peroxidases. *Food Chemistry*, 115: 1177-1186.
- Monferrán, M. V., Sánchez Agudo, J. A., Pignata, M. L. and Wunderlin, D. A. 2009. Copper-induced response of physiological parameters and antioxidant enzymes in aquatic macrophytes *Potamogeton pusillus*. *Environmental Pollution*, 157: 2570-2576.
- Morrison, T. A., Kessler, J. R., Hatfield, R. D. and Buxton, D. R. 1994. Activity of two lignin biosynthesis enzymes during development of a maize internode. *Journal of Science, Food and Agriculture*, 65: 133 – 139.

- Mortvedt, J. J., Fleischfresser, M. H., Berger, K. C. and Darling, H. M. 1961. The relation of soluble manganese to the incidence of common scab in potatoes. *American Potato Journal*, 38: 95-100.
- Mortvedt, J. J., Berger, K. C. and Darling, H. M. 1963. Effect of manganese and copper on the growth of *Streptomyces scabies* and the incidence of potato scab. *American Potato Journal*, 40: 96-102.
- Motsara, M. R. and Roy, R. N. 2008. Guide to laboratory establishment for nutrient analysis. FAO Fertilizer and plant nutrition bulletin 19. Food and Agriculture Organization of United Nations. Rome, Italy. 204 p.
- Moura, J. C. M. S., Bonie, C. A. V., Viana, J. O. F., Dornelas, M. C. and Mazzafera, P. 2010. Abiotic and biotic stresses and changes in the lignin content and composition in plants. *Journal of Integrative Plant Biology*, 52 (4): 360-376.
- Munevar M. F., 2001. Fertilization of oil palm to obtain high yields. *Palmas*, 22: 9-17.
- Myren, D. T. 1973. The influence of experimental conditions on a test of borax and sodium nitrate as stump treatments against infection by *Fomes annosus*. Information report O-X-191, Canadian Forestry Service, Department of the Environment, Great Lakes Forest Research Centre. 13 p.
- Naher, L., Tan, S. G., Umi Kalsom, Y., Ho, C. L. and Siddiquee, S. 2012. Activities of chitinase enzymes in oil palm (*Elaeis guineensis* Jacq.) in interactions with pathogenic and non-pathogenic fungi. *Plant Omics Journal*, 5 (4): 333-336.
- Ng, S. K., Cheah, T. E., Tamboo, S. and de Souza, P. 1968a. Nutrient contents of oil palms in Malaya. II. Nutrients in vegetative tissues. *The Malaysian Agricultural Journal*, 46 (3): 332-391.
- Ng, S. K., Cheah, T. E. and Tamboo S. 1968b. Nutrient contents of oil palms in Malaya. III. Micronutrient contents in vegetative tissues. *The Malaysian Agricultural Journal*, 46 (4): 421-434.
- Nicholson, R. L. and Hammerschmidt, R. 1992. Phenolic compounds and their role in disease resistance. *Annual Review of Plant Pathology*, 30: 369 – 389.
- Noor Pahtiwati, B., Harny, C. and Tiong, C. Y. 2013. Effect of Bunga Ta'ang essential oil volatile vapour on growth of the plant pathogenic fungus: *Ganoderma boninense*. In: Proceedings of the 5th MPOB-IOPRI International Seminar: Sustainable management of pests and diseases in oil palm-The way forward. November 2013, Kuala Lumpur Convention Centre (KLCC), Kuala Lumpur, Malaysia. pp 304-308.
- Nur Sabrina, A. A. 2011. Effects of calcium and copper on lignin biosynthesis and suppression of *Ganoderma boninense* infection in oil palm seedlings. MSc Thesis, Institute of Tropical Agriculture, University Putra Malaysia. 132 p.

- Nur Sabrina, A. A., Sariah, M. and Zaharah, A. R. 2012. Suppression of basal stem rot progress in oil palm (*Elaeis guineensis*) after copper and calcium supplementation. *Pertanika Journal of Tropical Agricultural Science*, 35 (S): 13-24.
- Nurfaezah, S., Wong, M. Y. and Dzolkhifli, O. 2012. Effects of pyraclostrobin, a natural fungicide on *Ganoderma boninense* mycelia growth and basal stem rot in oil palm seedlings. In: Book of abstracts International Agriculture Congress. Putrajaya Marriot Hotel, Malaysia, 4-6 September 2012. 2 p.
- O'Malley, D.M., Whetten, R., Bao, W., Chen, C-L. and Sederoff, R.R. 1993. The role of laccase in lignifications. *The Plant Journal*, 4 (5): 751-757.
- Panhwar, Q. A., Radziah, O., Khanif, M. Y. and Naher, U. A. 2011. Application of boron and zinc in the tropical soils and its effect on maize (*Zea mays*) growth and soil microbial environment. *Australian Journal of Crop Science*, 5 (12): 1649-1654.
- Paramanathan, S. 2000. *Soils of Malaysia – Their characteristics and identification*. Volume 1. Academic of Science Malaysia. 616 p.
- Paterson, R. R. M. 2007. *Ganoderma* disease of oil palm-A white rot perspective necessary for integrated control. *Crop Protection*, 26: 1369-1375.
- Paterson, R. R. M., Sariah M. and Zainal A. 2008. Altered lignin in oil palm: a novel approach to *Ganoderma* control. *The Planter*, 84 (985): 219-228.
- Pedras, M. S. C. and Yu, Y. 2008. Stress-driven discovery of metabolites from the phytopathogenic fungus *Leptosphaeria maculans*: structure and activity of leptomaculins A-E. *Bioorganic and Medicinal Chemistry*, 16: 8063-8071.
- Pillary, A-E., Williams, J. R., El Mardi, M. O., Hassan, S. M. and Al-Hamdi, A. 2005. Boron and the alternate-bearing phenomenon in the date palm (*Phoenix dactylifera*). *Journal of Arid Environment*, 62: 199-207.
- Pilotti, C. A. 2005. Stem rot of oil palm caused by *Ganoderma boninense*: pathogen biology and epidemiology. *Mycopathologia*, 159: 129-137.
- Polle, A., Otter, T. and Seifert F. 1994. Apoplastic peroxidase and lignification in needles of Norway spruce (*Picea abies* L.). *Plant Physiology*, 106: 53-60.
- Pomar, F., Merino, F. and Barceló, A.R. 2002. O-4-linked coniferyl and sinapyl aldehydes in lignifying cell walls are the main targets of the Wiesner (phloroglucinol-HCl) reaction. *Protoplasma*, 220: 17-28.
- Pratt, J. E. and Quill, K. 1996. A trial of disodium octaborate tetrahydrate for the control of *Heterobasidion annosum*. *European Journal of Forest Pathology*, 26: 297-305.

- Pratt, J. E. 2000. Effect of inoculum density and borate concentration in a stump treatment trial against *Heterobasidion annosum*. *Forest Pathology*, 30: 277-283.
- Quiroga, M., Guerrero, C., Botella, M.A., Barceló, A., Amaya, I., Medina, M.I., Alonso, F.J., de Forchetti, S.M., Tigier, H. and Valpuesta, V. 2000. A tomato peroxidase involved in the synthesis of lignin and suberin. *Plant Physiology*, 122: 1119-1127.
- Rahioui, B., Aissam, S., Messaouri, H., Moukhli, A., Khadari, B. and El Modafar, C. 2013. Role of phenolic metabolism in the defense of the olive-tree against leaf-spot disease caused by *Siplocaea oleagina*. *International Journal of Agriculture and Biology*, 15: 273-278.
- Raaskila S. 2008. The effect of lignin content and lignin modification on Norway spruce wood properties and decay resistance. Academic Dissertation 68. Department of Biological and Environmental Sciences, Faculty of Biosciences, University of Helsinki, Finland. 34 p.
- Rajanaidu, N. and Jalani, B. S. 1994. Prospects for breeding for kernels in oil palm (*Elaeis guineensis*). *The Planter*, 70 (820): 309 – 318.
- Ralph, J., Lundquist, K., Brunow, G., Lu, F., Kim, H., Schatz, P. F., Marita, J. M., Hatfield, R. D., Ralph, F. A., Christensen, J. H. and Boerjan, W. 2004. Lignins: Natural polymers from oxidative coupling of 4-hydroxyphenylpropanoids. *Phytochemistry Reviews*, 3: 29-60.
- Ramasamy, S. 1972. Cross-infection and decay ability of *Ganoderma* species parasitic to rubber, oil palm and tea. Bachelor of Agriculture Science. Project Report, niversity of Malaya. 49 p.
- Ranade-Malvi, U. 2011. Interaction of micronutrients with major nutrients with special reference to potassium. *Karnataka Journal of Agricultural Science*, 24 (1): 106-109.
- Ranocha, P., McDougall, G., Hawkins, S., Sterjiades, R., Borderies, G., Stewart, D., Cabanes-Macheteau, M., Boudet, A-M. and Goffner, D. 1999. Biochemical characterization, molecular cloning and expression of laccases-a divergent gene family-in poplar. *European Journal of Biochemistry*, 259: 485-495.
- Rao, V., Lim, C. C., Chia, C. C. and Teo, K. W. 2003. Studies on *Ganoderma* spread and control. *The Planter*, 79 (929): 367-383.
- Rees, R. W., Flood, J., Hasan, Y., Potter, U. and Cooper, R. M. 2009. Basal stem rot of oil palm (*Elaeis guineensis*); mode of root infection and lower stem invasion by *Ganoderma boninense*. *Plant Physiology*, 58: 982-989.
- Reuter, D. J. and Robinson, J. B. 1986. *Plant analysis. An interpretation manual*. Inkata Press. 218 p.

- Rice, R. W. 2007. The physiological role of minerals in plant. In: Datnoff, L. E., Elmer, W. H. and Huber, D. M. (eds). *Mineral nutrition and plant disease*. APS Press St. Paul, Minnesota, USA. pp 9-29.
- Ritchie, D. 2004. Copper-containing fungicides/bactericides and their use in management of bacterial spot on peaches. North Carolina State University as printed in Southeast Regional Newsletter, Vol.4, No. 1, March 2004. 4 pp. www.caes.uga.edu/commodities/fruits/gapeach/pdf/copperformulations.pdf (Accessed on the 08 August 2013).
- Rival A., 2007. Oil palm. In: Pua, E. C. and Davey, M. R. (eds). *Transgenic crops VI. Biotechnology in agriculture*, Vol. 61. Springer - Verlag Berlin Heidelberg. pp 59-80.
- Robson, A. D., Hartley, R. D. and Jarvis, S. C. 1981. Effect of copper deficiency on phenolic and other constituents of wheat cell walls. *New Phytologist*, 89: 361-371.
- Rouhi, A. M. C. and Washington, E. N. 2000. Lignin and lignan biosynthesis. *Science/Technology*, 78 (46): 29-32.
- Salisbury, F. B. and Ross, C. W. 1992. *Plant physiology*. Fourth edition. Wadsworth Publishing Company, California, USA. 682 p.
- Sariah, M., Hussin, M. Z., Miller, R. N. G. and Holderness, M. 1994. Pathogenicity of *Ganoderma boninense* tested by inoculation of oil palm seedlings. *Plant Pathology*, 43: 507-510.
- Sariah, M. and Zakaria, H. 2000. The use of soil amendments for the control of basal stem rot in oil palm seedlings. In: Flood, J., Bridge, P. D. and Holderness, M. (eds). *Ganoderma disease of perennial crops*. CABI Publishing, UK. pp 89-112.
- Sariah, M., Choo, C. W., Zakaria, H. and Norihan, M. S. 2005. Quantification and characterization of *Trichoderma* spp. from different ecosystems. *Mycopathologia*, 159: 113-117.
- Saxena, D. and Stotzky, G. 2001. Bt corn has a higher lignin content than non-Bt corn. *American Journal of Botany*, 88 (9): 1704-1706.
- Sayer, J., Ghazoul, J., Nelson, P. and Boedhihartono, A. K. 2012. Oil palm expansion transforms tropical landscapes and livelihoods. *Global Food Security*, 1: 114-119.
- Scăețeanu, G. V., Ilie, L. and Călin, C. 2013. An overview of manganese in nature. *American Chemical Science Journal*, 3 (3): 247-263.
- Sels, J., Mathys, J., De Coninck, B. M. A., Cammue, B. P. A. and De Bolle, M. F. C. 2008. Plant pathogenesis-related (PR) proteins: A focus on PR peptides. *Plant Physiology and Biochemistry*, 46: 941-950.

- Shamala, S. 2005. Performance of *Trichoderma harzianum* Rifai as a biological control agent for basal stem rot of oil palm (*Elaeis guineensis* Jacq.) caused by *Ganoderma boninense* Pat. MSc Thesis, Faculty of Science, University Putra Malaysia, Serdang, Selangor Malaysia. 182 p.
- Shamala, S., Sariah, M., Idris, A. S. and Radziah, O. 2011. Symbiotic interaction of endophytic bacteria with arbuscular mycorrhizal fungi and its antagonistic effect on *Ganoderma boninense*. *The Journal of Microbiology*, 49 (4): 551-557.
- Shamala, S., Idris, A. S., Nur Rasyeda, R. and Shariffah, M. S.A. 2013. Exploring the potentials of biological control agents against *Ganoderma* basal stem rot disease. In: Proceedings of the 5th MPOB-IOPRI International Seminar: Sustainable management of pests and diseases in oil palm-The way forward. 22-23 November 2013, Kuala Lumpur Convention Centre (KLCC), Kuala Lumpur, Malaysia. pp 178-191.
- Shamshuddin, J. and Darus, A. 1979. Mineralogy and genesis of soils in Universiti Pertanian Malaysia, Serdang, Selangor. *Pertanika*, 2 (2): 141-148.
- Sharma, C. P. 2006. *Plant micronutrients*. Science Publishers, New Hampshire, USA. 265 p.
- Sharma, M. 2007. Challenges facing the Malaysian palm oil industry-Multi-pronged strategies for raising oil yield, productivity and profitability. *The Planter*, 83 (981): 797-833.
- Shimomura, T. 1982. Effect of boron on the formation of local lesions and accumulation of callose in French bean and Samsun NN tobacco leaves inoculated with tobacco mosaic virus. *Physiology and Plant Pathology*, 20: 257-261.
- Siddiquee, S., Umi Kalsom, Y., Hossain K. and Jahan, S. 2009. In vitro studies on the potential *Trichoderma harzianum* for antagonistic properties against *Ganoderma boninense*. *Journal of Food, Agriculture and Environment*, 7 (3 and 4): 970-976.
- Singh, G. 1991. *Ganoderma* – The scourge of oil palm in coastal areas. *The Planter*, 67 (786): 421-444.
- Siti, R. A. A., Nor S. Y., Idris, A. S. and Mohd, B. W. 2004. Oil palm cellulose and lignin degradation of different *Ganoderma* sp based on ASTM standard rotting experiment. Poster paper 8. Unedited. International conference on pests and diseases of importance to the oil palm industry. Fostering global cooperation in instituting quarantine shield. 18-19 May 2004 Mutiara Hotel, Kuala Lumpur, Malaysia. 15 p.

- Sluiter, A., Hames, B., Ruiz, R., Scarlata, C., Sluiter, J., Templeton, D. and Crocker, D. 2007. Determination of structural carbohydrate and lignin in biomass. Laboratory analytical procedure. National Bioenergy Center, Department of Energy, USA. 14 p.
- Spann, T. and Schumann, A. W. 2013. Mineral nutrition contributes to plant disease and pest resistance. Horticultural Science series 1181 (HS1181), Department of Horticultural Sciences, Institute of Food and Agricultural Sciences (IFAS), University of Florida/IFAS Extension. 4 pp. edis.ifas.ufl.edu/hs1181 (Accessed on the 01.08.2013).
- Srivastava, S., Mishra, S., Tripathi, R. D., Dwivedi, S. and Guptadk. 2006. Copper-induced oxidative stress and responses of antioxidants and phytochelatins in *Hydrilla verticillata* L.f.) Royle. *Aquatic Toxicology*, 80 (4), 405-415.
- Stange, R. R. Jr. and McDonald, R. E. 1999. A simple and rapid method for determination of lignin in plant tissues-its usefulness in elicitor and comparison to the thioglycolic method. *Postharvest Biology and Technology*, 15: 185-193.
- Stangoulis J. C. R. and Graham, R. D. 2007. Boron and plant disease. In: Datnoff, L. E., Elmer, W. H. and Huber, D. M. (eds). *Mineral nutrition and plant disease*, APS Press St Paul, Minnesota, USA. pp 207-214.
- Sterjiades, R., Dean, J.F.D., Gamble, G., Himmelsbach, D.S. and Eriksson, K-E.L. 1993. Extracellular laccases and peroxidises from synamore maple (*Acer pseudoplanatus*) cell-suspension cultures. *Planta*, 190: 75-87.
- Sticknothe, H. and Schuchardt, F. 2011. Life cycle assessment of two palm oil production systems. *Biomass and Bioenergy*, 35: 3976-3984.
- Stone, E. 1990. Boron deficiency and toxicity in forest trees: A review. *Forest Ecology and Management*, 37: 49-75.
- Suleman, K. M. and Kausar, N. 1990. Effect of age, locality and sampling position on chemical composition of *Eucalyptus camaldulensis* Dehn. wood. *Pakistan Journal of Forestry*, 40 (1): 61-70.
- Sumathi, S., Chai, S. P. and Mohamed, A. R. 2008. Utilization of oil palm as a renewable energy in Malaysia. *Renewable and Sustainable Energy Reviews*, 12: 2404-2421.
- Sun, R., Tomkinson, J. and Bolton, J. 1999. Separation and characterization of lignins from the black liquor of oil palm trunk fiber pulping. *Separation Science and Technology*, 34(15): 3045-3058.
- Susanto, A., Sudharto, P. S. and Purba, R. Y. 2005. Enhancing biological control of basal stem rot disease (*Ganoderma boninense*) in oil palm plantations. *Mycopathologia*, 159: 153-157.

- Susanto, A. 2009. Basal stem rot in Indonesia-Biology, economic importance, epidemiology, detection and control. In: Proceedings of MPOB-IOPRI International Workshop on Awareness, Detection and Control of Oil Palm Devastating Diseases. November 2009, Kuala Lumpur Convention Centre (KLCC), Kuala Lumpur Malaysia. 18 p.
- Susanto, A. 2013. Biocontrol of Ganoderma basal stem rot disease of oil palm in Indonesia: Application and challenges. In: Proceedings of the 5th MPOB-IOPRI International Seminar: Sustainable management of pests and diseases in oil palm-The way forward. 22-23 November 2013, Kuala Lumpur Convention Centre (KLCC), Kuala Lumpur, Malaysia. pp 164-177.
- Taiz, L. and Zeiger, E. 2006. *Plant physiology*. Fourth edition. Sinauer. Associates, Inc., Publishers, Massachusetts, USA. 764 p.
- Tengoua F. F. and Bakoume, C. 2005. Basal stem rot and vascular wilt, two threats for oil palm sector in Cameroon. *The Planter*, 81 (947): 97-105.
- Tengoua, F. F. 2005. Report of phytosanitary inspection in Cameroon Development Corporation (CDC) Mungo and Mondoni Palm Estates. 4 p.
- Thompson, I. A. and Huber, D. M. 2007. Manganese and plant disease. In: Datnoff, L. E., Elmer, W. H. and Huber, D. M. (eds). *Mineral nutrition and plant disease*. APS Press, St Paul, Minnesota, USA. pp 139-153.
- Thurston, C.F. 1994. The structure and function of fungal laccases. *Microbiology*, 140: 19-26.
- Timonen, S. and Sen, R. 1998. Heterogeneity of fungal and plant enzyme expression in intact Scots pine-Suillus bovinus and-Paxillus involutus mycorrhizospheres developed in natural forest humus. *New Phytologist*, 138: 355-366.
- Turner, P. D. and Poon, Y. C. 1968. Effects of *Ganoderma* infection on the inorganic nutrient status of oil palm tissues. *Oléagineux*, 23 (6): 367-370.
- Turner, P. D. and Gillbanks, R. A. 2003. *Oil palm cultivation and management*. The Incorporated Society of Planters, Kuala Lumpur Malaysia. 633 p.
- Turner, E. C., Snaddon, J. L., Ewers, R. M., Fayle, T. M. and Foster, W. A. 2011. The impact of oil palm expansion on environmental change: Putting environmental research in context. In: Dos Santos Bernardes, M. A. (ed). *Environmental impact of biofuels*. In Tech, Croatia. pp 1-40.
- USDA, NRCS, 2007. Statistix 8 user guide for the plant materials program. United State Department of Agriculture and National Resources Conservation Service.

- Utomo, C. Werner, S., Niepold, F. and Deising, H. B. 2004. Specific primer design to detect *Ganoderma* associated with basal stem rot disease in oil palm. Poster Paper 9. International conference on pests and diseases of importance to the oil palm industry. Fostering Global Cooperation in Instituting Quarantine Shield. Kuala Lumpur, Malaysia. 18-19 May 2004. 14 p.
- Utomo, C. Werner, S., Niepold, F. and Deising, H. B. 2005. Identification of *Ganoderma*, the causal agent of basal stem rot disease in oil palm using a molecular method. *Mycopathologia*, 159: 159-170.
- Vance, C. P., Kirk, T. K. and Sherwood, R. T. 1980. Lignification as mechanism of disease resistance. *Annual Review of Phytopathology*, 18: 259-288.
- Veresoglou, S. D., Barto, E.K., Menexes, G. and Rillig, M. C. 2013. Fertilization affects the severity of disease caused by fungal plant pathogen. *Plant Pathology*, 62: 961-969.
- Vermerris, W., Thompson, K.J. and McIntyre, L.M. 2002. The maize Brown midrib 1 locus affects cell wall composition and plant development in a dose-dependent manner. *Heredity*, 88: 450-457.
- Vicent, A., Armengal, J. and García-Jiménez, J. 2009. Protectant activity of reduced concentration copper sprays against *Aternaria* brown spot on “Fortune” mandarin fruit in Spain. *Crop Protection*, 28: 1-6.
- Vidhyasekaran, P. 2008. *Fungal pathogenesis in plants and crops. Molecular biology and host defense mechanism*. Second edition. CRC Press. Taylor and Francis Group. New York, USA.
- Wahida, N. H., Anuar, A. R., Fauziah, C. I. and Osumanu, H. A. 2013. Response of *Brassica rapa* var. *parachinensis* grown on copper contaminated oxisol, inceptisol and histosol. *Malaysian Journal of Soil Science*, 17: 99-110.
- Wan, H. H. 2007. *Ganoderma* disease of oil palm in Sabah. *The Planter*, 83 (974): 299-313.
- Wang, G.-D., Li, Q.-J., Luo, B. and Chen, X.-Y. 2004. Ex-planta phytoremediation of trichlorophenol and phenolic allelochemicals via an engineered secretory laccase. *Nature Biotechnology*, 22 (7): 893-897.
- Wang, J., Wang, C., Zhu, M., Yu, Y., Zhang, Y. and Wei, Z. 2008. Generation and characterization of transgenic poplar plants overexpressing a cotton laccase gene. *Plant Cell Tissue and Organ Culture*, 93: 303-310.
- Webster, M. A. and Dixon, G. R. 1991. Boron, pH and inoculum concentration influencing colonization by *Plasmodiophora brassicae*. *Mycological Research*, 95 (1); 74-79.

- Whetten, R.W. and Sederoff, R.R. 1992. Phenylalanine ammonia-lyase from loblolly pine. Purification of the enzyme and isolation of complementary DNA clone. *Plant Physiology*, 98: 380-386.
- Whetten, R. and Sederoff, R. 1995. Lignin biosynthesis. *The Plant Cell*, 7: 1001-1013.
- Whetten, R. W., MacKay, J. J. and Sederoff, R. R. 1998. Recent advances in understanding lignin biosynthesis. *Annual Review of Plant Physiology and Plant Molecular Biology*, 49: 585-609.
- Wiedenhoeft, A. C. and Hopkins, W. G. 2006. *Plant nutrition*. Infobase Publishing, New York, USA. 144 p.
- Wilcove, D. S. and Koh, L. P. 2010. Addressing the threats to biodiversity from oil-palm agriculture. *Biodiversity Conservation*, 19: 999-1007.
- Wong, L.-C., Bong, C.-F. J. and Idris, A. S. 2012. *Ganoderma* species associated with basal stem rot disease of oil palm. *American Journal of Applied Sciences*, 9 (6): 879-885.
- Yao, K., De Luca, V. and Brisson, N. 1995. Creation of a metabolic sink for tryptophan alters the phenylpropanoid pathway and the susceptibility of potato to *Phytophthora infestans*. *The Plant Cell*, 7: 1787-1799.
- Yoshihara, K., Kobayasi, T., Fujii, T. and Akamatsu, I. 1984. A novel modification of Klason lignin quantitative method. *Journal Japan Tappi*, 38: 466-475.
- Yusoff, S. and Hansen, S. B. 2007. Feasibility study of performing a life cycle assessment on crude palm oil production in Malaysia. *International Journal of Life Cycle Assessment*, 12 (1): 50-58.
- Zaiton, S., Sariah, M. and Zainal, A. M. A. 2008. Effect of endophytic bacteria on growth and suppression of *Ganoderma* infection in oil palm. *International Journal of Agriculture and Biology*, 10: 127-132.
- Zhong, R., Morrison III, W. H., Negrel, J. and Ye, Z.-H. 1998. Dual methylation pathways in lignin biosynthesis. *The Plant Cell*, 10: 2033-2045.
- Zitter, T. A. 2012. Copper fungicides-A comprehensive list of products used for vegetable disease control. [www.vegetablemdonline.ppath.cornell.edu/NewsArticles/Copper Fungicides/2012 OCT.pdf](http://www.vegetablemdonline.ppath.cornell.edu/NewsArticles/Copper_Fungicides/2012_OCT.pdf) (Accessed on the 13 August 2013). 6 p.