

# **UNIVERSITI PUTRA MALAYSIA**

Andrographis paniculata Nees AND Orthosiphon stamineus Benth GROUND LEAF AS ANTIBIOTIC AND ANTIOXIDANT SUPPLEMENTS FOR BROILER CHICKEN

**MASNINDAH BINTI MALAHUBBAN** 

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**MASNINDAH BINTI MALAHUBBAN** 

DOCTOR OF PHILOSOPHY UNIVERSITI PUTRA MALAYSIA

2014



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# By

MASNINDAH BINTI MALAHUBBAN

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Doctor of Philosophy

September 2014

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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirements for the degree of Doctor of Philosophy

### Andrographis paniculata Nees AND Orthosiphon stamineus Benth GROUND LEAF AS ANTIBIOTIC AND ANTIOXIDANT SUPPLEMENTS FOR BROILER CHICKEN

By

#### MASNINDAH BINTI MALAHUBBAN

September 2014

Chairman: Professor Abd. Razak Bin Alimon, Ph.D.

#### Faculty: Institute of Tropical Agriculture

The use of synthetic antibiotics and antioxidants has creates serious obstacles and hazards, to farmers, consumers and the environment. Antibiotics and antioxidant from natural sources can potentially be useful to overcome these problems. Therefore, present study was conducted to evaluate the selected medicinal plants, *Andrographis paniculata* and *Orthosiphon stamineus* for antibiotics and antioxidant potential and subsequently to evaluate the broiler performance by inclusion the *A. paniculata* and *O. stamineus* ground leaf in diets.

Initially, *A. paniculata* and *O. stamineus* ground leaf were evaluated for their *in vitro* antibiotic and antioxidant potential by extracting the ground leaf in four solvents, namely water, methanol, ethanol and chloroform and subsequently testing using the disc diffusion method for antibiotic potential and selected radical scavenging activities for antioxidant potential. Qualitative screening for bioactive compound on both extracts by using methanol as solvent has indicated the presence of alkaloid, saponin, flavonoid, tannin, terpenoid and steroid. High performance liquid chromatography analysis indicated that andrographolide and rosmarinic acid were the major compound from *A. paniculata* and *O. stamineus*, respectively. It was found that the highest yields of these two compounds were obtained by methanol extraction and that substantial antibiotic and antioxidant properties were exhibited by these compounds. In addition, it was showed that the *O. stamineus* extract



The ground leaf were incorporated in broiler diets at levels of 0, 2, 4, and 8 g/kg and the influence on growth performance, carcass characteristics, serum biochemistry, and intestinal and liver morphology was examined in a 42-d feeding trial. It was involved 280 one-day old male broiler chickens, grown, maintained and received ad libitum water and diet. The feeding trial was started from 21 days old and assigned with respective treatments. At day-42, the broiler chickens were slaughtered and analysed and it was found that broilers fed O. stamineus ground leaf at a rate 8 g/kg was the most promising dietary supplement to promote overall growth performance without deleterious effects on carcass characteristics, serum biochemical properties and morphological components of liver and intestine compared with A. paniculata ground leaf and control diets. In addition to promoting weight gain, it reduced abdominal fat and serum cholesterol. It also maintained the integrity of liver, thus indicating that no toxic effect from O. staminues supplementation at a rate up to 0.8 g/kg. Besides that, 8 g/kg O. stamineus supplementation improved intestinal structure, especially in the duodenum. Present study also found that the inclusion of O. stamineus ground leaf at 8 g/kg in the broiler diet increased total tract N retention and apparent metabolizable energy.

Based on these results, O. stamineus ground leaf at 8 g/kg was selected for the next in vivo experiment. Present study was conducted to evaluate the response of broiler due to antibiotic and antioxidant properties of O. staminues by comparing its potential with tetracycline and Vitamin E supplementation in diets, as positive controls. The study was involved 160 of a one-day old male broiler chickens, and had ad libitum water and feed for up to 20-day old. The respective treatments were assigned and initiated at 21day old male broiler chickens. After slaughtering, data were recorded and analysed at 42-day old male broiler chickens, and it was found that O. stamineus ground leaf supplement at a rate of 8 g/kg in broiler diet results in growth performance similar to that of tetracycline and Vitamin E supplementation. In addition, it was also found that 8 g/kg O. stamineus supplementation in diet promoted serum enzymes-lowering effect. In contrast, high serum enzymes activity showed in broiler fed tetracycline supplement. The inclusion of 8 g/kg O. stamineus ground leaf in diet of broiler enhanced meat quality by stabilizing sensory properties, meat colour and meat pH. The results also indicated that the inclusion of O. stamineus leaf ground at 8 g/kg in diet was comparable with 200 mg/kg Vitamin E supplementation in diet. It was found that all dietary treatments maintained intestinal population of Lactobacillus and Escherichia coli. However, the inclusion of 8 g/kg O. stamineus or tetracycline in diet inhibited the population of facultative anaerobe. Therefore, the use of O. stamineus ground leaf as supplement in broiler chicken diet has the potential to promote and maintain growth and gut health and subsequently creates safe and sustainable broiler chicken production.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah.

#### SERBUK DAUN Andrographis paniculata Nees DAN Orthosiphon stamineus Benth SEBAGAI ADITIF ANTIBIOTIK DAN ANTIOKSIDAN UNTUK AYAM PEDAGING

Oleh

#### MASNINDAH BINTI MALAHUBBAN

September 2014

#### Pengerusi: Profesor Abd. Razak Bin Alimon, Ph.D.

#### Fakulti: Institut Pertanian Tropika

Penggunaan antibiotik dan antioksidan sintetik telah menyebabkan masalah dan bencana yang serius kepada petani, pengguna dan persekitaran. Antibiotik dan antioksidan dari sumber semulajadi dilihat berpotensi untuk digunakan bagi mengatasi masalah ini. Maka kajian ini dijalankan untuk menilai tumbuhan perubatan yang terpilih iaitu *Andrographis paniculata* dan *Orthosiphon stamineus* yang berpotensi sebagai sumber antibiotik dan antioksidan, seterusnya kajian ini turut menilai prestasi ayam pedaging kesan dari pengambilan makanan yang mengandungi serbuk daun *A. paniculata* dan *O. stamineus*.

Kajian dimulakan dengan menilai potensi antibiotik dan antioksidan serbuk daun *A. paniculata* dan *O. stamineus* secara *in vitro* dengan mengekstrak serbuk daun tersebut dalam empat jenis bahan pelarut iaitu air, methanol, etanol dan klorofom, dan seterusnya ia diuji menggunakan kaedah sebaran cakera bagi menentukan potensi antibiotik dan aktiviti-aktiviti pencarian radikal bagi penentuan potensi antibiotik dan serbuk daun yang diekstrak menggunakan metanol mengandungi alkaloid, saponin, flavonoid, tannin, terpenoid dan steroid. Analisis kromatografi cecair berprestasi tinggi menunjukkan andrographolida dan asid rosmarinik merupakan kompaun utama masing-masing dari *A. paniculata* dan *O. stamineus*. Kajian mendapati hasilan paling tinggi bagi kedua-dua kompaun terbabit diperoleh daripada pengekstrakan methanol dan ciri-ciri antibiotik dan antioksidan yang menggalakkan ditunjukkan oleh kedua-dua kompaun terbabit.

Serbuk daun berkenaan dicampurkan ke dalam diet ayam pedaging pada beberapa paras kandungan iaitu 0, 2, 4, dan 8 g/kg bagi menentukan pengaruhnya ke atas prestasi pertumbuhan, ciri-ciri karkas, biokimia serum,

dan morfologi usus dan hati, dan kajian cubaan pemakanan ini dilakukan sehingga ayam pedaging berusia 42 hari. Kajian ini melibatkan ayam pedaging jantan yang diperolehi pada ketika usianya satu hari, dipelihara dan dijaga. Cubaan pemakanan dimulakan ketika ayam pedaging berusia 21 hari dan seterusnya disusun mengikut rawatan tersebut. Pada usianya 42 hari, ayam pedaging disembelih dan dianalisis, dan keputusan mendapati ayam pedaging yang makan 8 g/kg serbuk daun O. stamineus merupakan makanan tambahan yang paling berpotensi untuk merangsangkan prestasi pertumbuhan tanpa kesan-kesan negative ke atas ciri-ciri karkas, kandungan biokimia serum dan komponen morfologi hati dan usus berbanding dengan serbuk daun A. paniculata dan diet-diet kawalan. Tambahan kepada penggalakan peningkatan berat, ia telah mengurangkan lemak perut dan kolesterol serum. Ia jua memelihara integriti hati, dan hal ini menunjukkan tiadanya kesan toksik dari O. stamineus yang ditambah di dalam pemakanan sebanyak 8 g/kg. Di samping itu, penambahan 8 g/kg O. stamineus memperbaiki struktur usus, terutamanya duodenum. Kajian ini turut mendapati penambahan 8 g/kg serbuk daun O. stamineus meningkatkan jumlah zon pengumpulan N dan tenaga sebenar yang boleh dimetabolikkan.

Berdasarkan kepada keputusan tersebut, 8 g/kg serbuk daun O. stamineus telah dipilih untuk eksperimen in vivo. Kajian ini dijalankan untuk menilai tindakbalas ayam pedaging kesan dari kandungan antibiotik dan antioksidan O. stamineus dengan membandingkannya dengan penambahan tetrasiklin dan Vitamin E di dalam pemakanan, sebagai kawalan positif. Kajian ini melibatkan 160 ekor ayam pedaging jantan yang dipelihari sejak berusia satu hari. Rawatan kajian dimulakan semasa ayam pedaging berusia 21 hari. Data mula direkodkan dan dianalisis ketika berusia 42 hari. Kajian menunjukkan serbuk daun O. stamineus pada kadar 8 g/kg mempunyai prestasi pertumbuhan yang menyamai prestasi tetrasiklin dan Vitamin E. Tambahan lagi, ia juga didapati mampu merangsang kesan kekurangan enzim serum. Sebaliknya, aktiviti enzim serum yang tinggi didapati pada ayam pedaging yang dirawat dengan pemakanan tambahan yang mengandungi tetrasiklin. Penambahan 8 g/kg serbuk daun O. stamineus di dalam makanan ayam pedaging telah meningkatkan kualiti daging dengan menstabilkan komponen rasa, warna dan pH daging. Kajian turut menunjukkan ia setanding dengan penambahan sebanyak 200 mg/kg Vitamin E. Selain itu, rawatan tambahan pemakanan ini memelihara populasi Lactobacillus dan Eschericia coli di dalam usus. Walau bagaimanapun, penambahan 8 g/kg serbuk daun O. stamineus merencatkan populasi fakultatif anaerob. Dengan ini, penggunaan serbuk daun O. stamineus sebagai tambahan di dalam pemakanan ayam pedaging berpotensi untuk merangsang dan mengekalkan pertumbuhan dan kesihatan perut ayam pedaging dan seterusnya mewujudkan pengeluaran ayam pedaging yang selamat danlestari.

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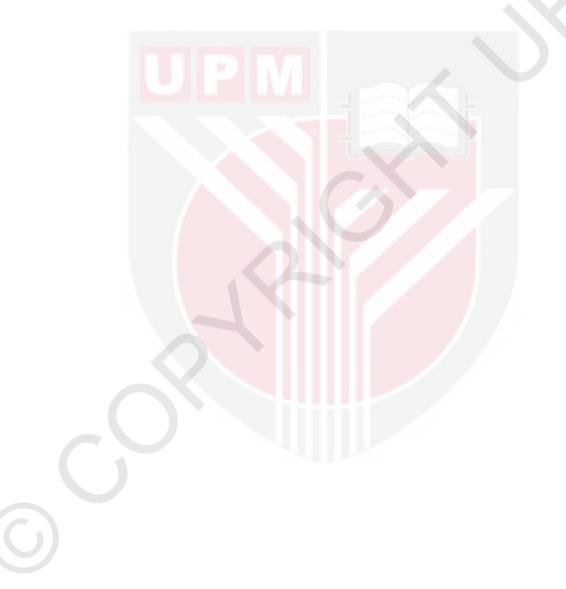
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- 2.1 Andrographis paniculata Nees, common name associated with 14 this medicinal plant including, King of Bitter, and several other names from Malay Archipelago such as *Hempedu Bumi*, *Pokok Cerita, Pasak Bumi* and *Setunjang Bumi*.
- 2.2 Orthosiphon stamineus Benth, common names associated with 17 this medicinal plant including, Cat Whiskers and Kidney Tea Plant, and several other names from Malay Archipelago such as Misai Kucing, Kumis Kucing, Remujung, Ruku Hitam, and Teh Jawa (Java Tea).
- 3.1 HPLC chromatograms (absorbance at 230 nm) of 29 andrographolide (AP) from *Andrographis paniculata* leaves extracted with different solvents: (a) reference marker; (b) water extract; (c) ethanol extract; (d) methanol extract; (e) chloroform extract.
- 3.2 HPLC chromatograms (absorbance at 340 nm) of rosmarinic 32 acid (RA) from *Orthosiphon stamineus* leaves extracted with different solvents: (a) reference marker; (b) water extract; (c) ethanol extract; (d) methanol extract; (e) chloroform extract (the arrow indicates the RA was undetected).
- 3.3 Susceptibility of pathogenic bacteria to Andrographis 34 paniculata and Orthosiphon stamineus extracts prepared with different solvents. Note: Ctrl: Control; Ac: Acetic acid (10 %); Tc: Tetracycline (30 µg/disc). Heights of colored bars represent inhibition diameters.
- 3.4 Total phenol contents of *Andrographis paniculata* and 35 *Orthosiphon stamineus* leaf extracts prepared with different solvents. Values in each column bearing the same letter are not significantly different (P>0.05).
- 3.5 DPPH scavenging activity of *A. paniculata* and *O. stamineus* 36 extracts prepared with different solvents. Values in each column bearing the same letter are not significantly different (P>0.05).
- 3.6 Superoxide scavenging activity of *A. paniculata* and *O.* 37 *stamineus* extracts prepared with different solvents. Values in each column bearing the same letter are not significantly different (P>0.05).

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- 4.1 Histological examination on liver of 42-day old broilers fed 67 control diet, Diet AP2, Diet AP4, Diet AP8, Diet OS2, Diet OS4, and Diet OS8. All figures show normal liver histology where hepatocytes (H), sinusoid (S), central vein (CV) and portal area conditions were at normal (40X magnification).
- 5.1 The pH of raw breast meat of broiler chickens on different 91 diets. Changes over duration of storage at 4°C.



# LIST OF ABBREVIATIONS

ALP	Alkaline Phosphatase
ALT	Alanine Aminotransferase
AME	Apparent metabolizable energy
AP	Andrographis paniculata
AST	Aspartate Aminotransferase
CFU	Colony Forming Unit
СР	Crude Protein
DM	Dry Matter
EU	European Union
FAO	Food and Agriculture Organization
FCR	Food Conversion Ratio
FDA	Food and Drug Association
GE	Gross Energy
GAE	Gallic Acid Equivalent
HPLC	High Performance Liquid Chromatography
IC <sub>50</sub>	Inhibitory Concentration at 50 %
NRC	National Research Council
OS	Orthosiphon stamineus
T20	Tetracycline
USDA	United States Department of Agriculture
VE	Vitamin E

	Units	
	°C	degrees centigrade
	cm	centimeter
	g	gram
	g/day	gram per day
	g/kg	gram per kilogram
	h	hour
	l kg	liter kilogram
	MJ/kg	megajoule per kilogram
	min	minute
	mbar	milibar
	mg	milligram
	ml	milliliter
	mm	millimeter
	mmol/L	milimol per liter
	ml/min	milliliter per minute
	mM	milimol
	mg/ml	milligram per milliliter
	Μ	molar
	μm	micrometer
	μL	microliter
	μg	microgram
	ppm	parts per million
	sec	second

# Common abbreviations

et al.	and others
%	percentage
e.g	for example

# Statistical terms

ANOVA	Analysis of variance
CRD	Completely Randomized Design
SE	Standard Error

# Chemical elements and compounds

ABTS	2,2- azinobis- (3 ethylbenzothiazoline-6-sulfonic acid)
ACN	acetonitrile
DPPH	2, 2- diphenyl -1-picrylhydrazyl
HCI	Hydrochloric acid
$H_2SO_4$	Sulphuric acid
МНА	Muller Hinton Agar
RA	Rogosa Agar

#### CHAPTER 1

#### **GENERAL INTRODUCTION**

Broiler chicken represents 29% of meat production from farmed animals and this value rising each year. Poultry meat and eggs offer considerable potential for meeting human needs for dietary animal protein supply. With regards to the growth of broiler industry, Malaysia ranked seventh place in the world (Anon, 2012). In Malaysia, the broiler industry is expected to grow at the rate of 4% in 2013, outpacing global production growth at 2.5% (Lim, 2013).

To improve broiler production and satisfy market demand, it has been the common practice for farmers to use synthetic antibiotics and antioxidant in the feed as growth promoters. Antibiotics including chlortetracycline, tetracycline, virginiamycin, spiramycin, tylosin phosphate, zinc bacitracin and avopracin as growth promoters have been used for decades in poultry production for improving farm performance and controlling diseases (Huyghebaert et al., 2011). With increasing interests in discontinuing the use of antibiotics due to their harmful effects to environment, actively efforts to search for safe, suitable and viable alternatives to the antibiotic growth promoters has become intensified. Ideally, these alternative growth promoters should improve growth performance, as do the antibiotics, and maintain a sound health of the chickens. Besides the need of healthy grown broilers, meat quality also needs to give much attention. Oxidative stability is a central parameter in the estimation of meat quality because of the susceptibility of this food product to oxidative degeneration, which is one of the main causes of spoilage (Morrissev et al., 1998). The shelf life of meat is related to lipid oxidation reactions which could affect its sensory properties, by causing rancidity, as well as its nutritional characteristics through the formation of potentially toxic compound (Morrissey and Kerry, 2004). Conventionally, oxidation of chicken meat can be prevented by synthetic antioxidants including butylated hydroxyanisole (BHA), butylated hydroxy toluene (BHT) and propyl gallate (PG) but their safety has been questioned (Barlow, 1990).

Moreover, the excessive use of these synthetic antibiotics and antioxidants has led to contamination of broiler meat and environment. For example, the practice of feeding antibiotics to livestock leads to antibiotic-resistant bacteria that are dangerous to human health. Frequently and excessively used of antibiotics cause harmful bacteria become resistant to the drug, and the treatment becomes less effective (Huyghebaert *et al.*, 2011). In fact, bacteria can develop into a completely different strain that cannot be killed by the normally prescribed antibiotic. Synthetic antioxidants have been restricted recently, mainly because of their possible carcinogenicity causing liver swelling and changing liver enzyme activities. With the increasing consciousness of public consumers on safety of food additive, therefore, an urgent need for identifying alternative natural and probably safer sources of antibiotics and antioxidants. Currently, the interest in natural antioxidants and antibiotics have increased dramatically because they are considered to be safer than the synthetics, and have greater application potential for consumers acceptability, palatability, stability and shelf-life of meat products (Kang *et al.*, 2008). The medicinal herbs have been used since ancient times not only for flavouring foods but also for their remedies. The preservative effect of spices and herbs suggests the presence of antimicrobial and antioxidative constituents (Basmacioglu *et al.*, 2004; Al-Marzooqi *et al.*, 2010).

In the present study, two species of medicinal plants have been selected namely, *Andrographis paniculata* Nees, (Acanthaceae) and *Orthosiphon stamineus* Benth, (Lamiaceae), they are common in Southeast Asia, India and China. *A. paniculata* has been traditionally used as an antioxidant, antiviral, anti-inflammatory, immune enhancing agent and hepatoprotective (Prajjal *et al.*, 2003). Its active component has been reported to have anti-cancer (Sheeja and Kuttan, 2007), anti-HIV (Calabrese *et al.*, 2000), and antimicrobial (Roy *et al.*, 2010) properties. The other herb, *Orthosiphon stamineus*, has been used to treat urinary lithiasis, edema, eruptive fever, influenza, rheumatism, hepatitis, jaundice and biliary lithiasis (Akowuah *et al.*, 2005). *O. stamineus* leaf is consumed as Java tea to facilitate body detoxification (Chin *et al.*, 2008). Both plants have been reported containing natural antibiotics and antioxidants (Prajjal *et al.*, 2003; Ho *et al.*, 2010). However, very few information of medicinal plants as dietary supplements in animals, especially *Andrographis paniculata* and *Orthosiphon stamineus*. The hypotheses of the present study were:

- 1. Andrographis paniculata and Orthosiphon stamineus are species of medicinal plants containing antibiotic and antioxidant compounds.
- 2. Supplementation of *A. paniculata* and *O. stamineus* ground leaf in the diets improve growth and meat quality in broiler chickens.

The goal of this research work was to evaluate the possibilities of improving broiler chicken performance using *A. paniculata* and O. *stamineus* leaf preparation as feed supplement as opposed to the synthetic antibiotics and antioxidant. The specific objectives set were:

- 1. To determine the antibiotic and antioxidant properties of *A. paniculata* and *O. stamineus* leaf extracts in different solvents, and to detect and quantify andrographolide and rosmarinic acid presence in the respective species.
- 2. To evaluate the effect of different levels of *A. paniculata* and *O. stamineus* ground leaf supplemented in diets on growth performance, carcass characteristics and blood biochemistry of broiler chickens, and also to determine the morphological and histological changes in gastrointestinal tract and in the liver.
- 3. To evaluate the antibiotic and antioxidant potential of *Orthosiphon stamineus* ground leaf on growth performance, meat characteristics, intestinal microflora and lipid peroxidation of broiler chickens.

#### REFERENCES

- Abdel-Wareth, A.A.A., Kehraus, S., Hippenstiel, F. and Sudekum, K.-H. 2012. Effect of thyme and oregano on growth performance of broilers from 4 to 42 days of age and on microbial counts in crop, small intestine and caecum of 42-day-old broilers. *Animal Feed Science and Technol*ogy 178: 198-202.
- Abu-Dieyeh, Z.H. and Abu-Darwish, M.S. 2008. Effect of feeding powdered black cumin seeds (*Nigella sativa L.*) on growth performance of 4-8 week old broilers. *Journal of Animal and Veterinary Advances* 3: 286-290.
- Ahn, Y.J., Ganesan, P. and Kwak, H.S. 2012. Comparison of polyphenol content and antiradical scavenging activity in methanolic extract of nano-powdered and powdered peanut sprouts. *The Journal of Korean Society of Applied Biological Chemistry* 55: 793-798.
- Akowuah, G.A., Zhari, I., Norhayati, I. and Mariam, A. 2006. HPLC and HPTLC densitometric determination of andrographolides and antioxidant potential of *Andrographis paniculata*. *Journal of Food Composition and Analysis* 19: 118-126.
- Akowuah, G.A., Ismail, Z., Norhayati, I. and Sadikun, A. 2005. The effects of different extraction solvents of varying polarities on polyphenols of *Orthosiphon stamineus* and evaluation of the free radical-scavenging activity. *Food Chemistry* 93: 311-317.
- Akowuah, G.A., Zhari, I., Norhayati, I., Sadikun, A. and Khamsah, S.M. 2004. Sinensetin, eupatorin, 3'-hydroxy-5, 6, 7, 4'-tetramethoxyflavone and rosmarinic acid contents and antioxidative effect of *Orthosiphon stamineus* from Malaysia. *Food Chemistry* 87: 559-566.
- Aksit, M., Yalcin, S., Ozkan, S., Metin, K. and Ozdemir, D. 2006. Effects of temperature during rearing and crating on stress parameters and meat quality of broilers. *Journal of Poultry Science* 85: 1867-1874.
- Al-Bayati, F.A. 2008. Synergistic antibacterial activity between *Thymus vulgaris* and *Pimpinella anisum* essential oils and methanol extracts. *Journal of Ethnopharmacology* 116: 403-406.
- Al-Kassie, G.A.M. and Jameel, Y.J. 2009. The effect of adding *Thyme vulgaris* and *Cinnamomuim zeylanicum* on productive performance in broilers. Proceeding of 9th Veterinary Scientific Conference, Collage University of Veterinary Medical Baghdad, Iraq.
- Al-Marzooqi, W., Al-Farsi, M.A., Kadim, I.T., Mahgoub, O. and Goddard, J.S. 2010. The effect of feeding different levels of sardine fish silage on broiler performance, meat quality and sensory characteristics under closed and open-sided housing systems. *Asian-Australasian Journal of Animal Sciences* 23: 1614-1625.
- Al-Sultan, S.I. 2003. The effect of *Curcuma longa* (Turmeric) on overall performance of broiler chickens. *International Journal of Poultry Science* 2: 351–353.
- Amad, A.A., Manner, K., Wendler, K.R., Neumann, K. and Zentek, J. 2011. Effects of a phytogenic feed additive on growth performance and ileal nutrient digestibility in broiler chickens. *Poultry Science* 90: 2811-2816.

- Amarowicz, R., Naczk, M. and Shahidi, F. 2000. Antioxidant activity of various fraction of non-tanin phenolics of canola hulls. *Journal of Agricultural and Food Chemistry* 48: 2755-2759.
- Ameer, O.Z., Salman, I.M., Asmawi, M.Z., Ibraheem, Z.O. and Yam, M.F. 2012. Orthosiphon stamineus: Traditional Uses, Phytochemistry, Pharmacology, and Toxicology. *Journal of Medicinal Food* 15: 678-690.
- Amos, T.T. 2006. Analysis of backyard poultry production in Ondo state, Nigeria. International Journal of Poultry Science 5: 247-250.
- Anadon, A. 2006. The EU ban of antibiotics as feed additives (2006): alternatives and consumer safety. *Journal of Veterinary and Pharmacology Therapeutics* 29: 41-44.
- Anon. 2012. GLOBAL POULTRY TRENDS 2012 Poultry Increases its Share of Global Meat Production. Retrieved on 5 June 2013. http://www.thepoultrysite.com/articles/2515/global-poultry-trends-2012poultry-increases-its-share-of-global-meat-production.
- Ao, X., Yoo, J.S., Zhou, T.X., Wang, J.P., Meng, Q.W., Yan, L., Cho, J.H. and Kim, I. H. 2011. Effects of fermented garlic powder supplementation on growth performance, blood profiles and breast meat quality in broilers. *Livestock Science* 141: 85-89.
- AOAC International.1995. Official Methods of Analysis of AOAC International. 16th ed. Arlington VA: AOAC International.
- Babatunde, G.M., Pond, W.O., Krook, L., Dvan, L., Walker, E.R. and Chapman, D. 1987. Effect of dietary safflower oil or hydrogenerated coconut oil on growth rate and on swine blood and tissue components of pigs fed fat-free diets. *Journal of Nutrition* 92: 1903-1910.
- Bajpai, V.K., Rahman, A. and Kang, S.C. 2007. Chemical composition and anti-fungal properties of the essential oil and crude extracts of *Metasequoia glyptostroboides* Miki ex Hu. *Industrial Crops and Products* 26: 28-35.
- Barbosa, L.N., Rall, V.L., Fernandes, A.A., Ushimaru, P.I., da Saliva Probst,
  I. and Fernandes, Jr. A. 2009. Essential oils against foodborne pathogens and spoilage bacteria in minced meat. *Foodborne Pathogens Disease* 6: 725-728.
- Barlow, S.N. 1990. Toxicological aspects of antioxidants used as food additives. In *Food antioxidants*, ed. Hudson, B.J.F., pp. 253-307. Amsterdam: Elsevier.
- Barrow, P. A., 1992. Probiotics for chickens. In *Probiotics*, ed. Fuller, R., pp. 225–257. London: Chapman and Hall.
- Basmacioglu, H., Tokusoglu, O. and Ergul, M. 2004. The effect of oregano and rosemary essential oils or alpha-tocopheryl acetate on performance and lipid oxidation of meat enriched with n-3 PUFAs in broilers. *South African Journal of Animal Science* 34: 197-210.
- Bauer, A. W., Kirby, M. M., Sherris, J. C. and Turck, M. 1966. Antibiotic susceptibility testing by a standardized single disk method. *American Journal of Clinical Pathology* 45: 493-496.
- Benz, R. and Bauer, K. 1988. Permeation of hydrophilic molecules through the outer membrane of gram – negative bacteria. *European Journal of Biochemistry* 176: 1-19.

- Bera, D., Lahiri, D. and Nag, A. 2006. Studies on natural antioxidant for stabilization of edible oil and comparison with synthetic antioxidants. *Journal of Food Engineering* 74: 542-545.
- Biswas, A.H. and Wakita, M. 2001. Effect of dietary Japanese green tea powder supplementation on feed utilization and carcass profiles in broilers. *Journal of Poultry Science* 38: 50-57.
- Botsoglou, N.A., Florou-Paneri, P., Botsoglou, E., Dotas, V., Giannenas, I., Koidis, A. and Mitrakos, P. 2005. The effect of feeding rosemary, oregano, saffron and alpha-tocopheryl acetate on hen performance and oxidative stability of eggs. *South African Journal of Animal Science* 35: 143-151.
- Botsoglou, N.A., Papageorgiou, G., Nikolakakis, I., Florou-Paneri, P., Giannenas, I., Dotas, V. and Sinapis, E. 2004. Effect of dietary dried tomato pulp on oxidative stability of Japanese quail meat. *Journal of Agricultural and Food Chemistry* 52: 2982–2988.
- Brenes, A. and Roura, E. 2010. Essential oils in poultry nutrition: Main effects and modes of action. *Animal Feed Science and Technology* 158: 1-14.
- Burt, S. 2004. Essential oils: their antibacterial properties and potential applications in food A review. *International Journal of Food Microbiology* 94: 223-253.
- Calabrese C., Berman, S.H., Babish, J.G., Ma, X., Shinto, L., Dorr, M., Wells, K., Wenner C.A. and Standish, L.J. 2000. A Phase I trial of andrographolide in HIV positive patients and normal volunteers. *Phytotherapy Research* 14: 333-338.
- Cao, P.H., Li, Y.F., Peron, P., Schulze, H. and Bento, H. 2010. Effect of essential oils and feed enzymes on performance and nutrient utilization in broilers fed a corn/soy based diet. *International Journal of Poultry Science* 9: 749-755.
- Castanon, J.I.R. 2007. History of the use of antibiotic as growth promoter in European poultry feeds. *Poultry Science* 86: 2466-2471.
- Castellini, C., Mugnai, C. and Dal Bosco, A. 2002. Effect of organic production system on broiler carcass and meat quality. *Meat Science* 60: 219–225.
- Chen, Z.Y., Ma, K.Y., Liang, Y. Peng, C. and Zuo, Y. 2011. Role and classification of cholesterol-lowering functional foods. *Journal of Functional Foods* 3: 61-69.
- Chin, R.P., Van Empel, P.C.M. and Hafez, H.M. 2008. Ornithobacterium rhinotracheale infection. In: Diseases of Poultry, 12th ed. pp. 765–774. Iowa, USA: Blackwell Publishing.
- Choi, S.Y., Lee, M.H., Choi, J.H. and Kim, Y.K. 2012. 2, 3, 22, 23-Tetrahydroxyl-2, 6, 10, 15, 19, 23-hexamethyl-6, 10, 14, 18tetracosatetraene, an Acyclic Triterpenoid Isolated from the Seeds of Alpinia katsumadai, Inhibits Acyl-CoA: Cholesterol Acyltransferase Activity. *Biological and Pharmaceutical Bulletin* 35: 2092-2096.
- Chrubasik, S., Pittler, M.H. and Roufogalis, B.D. 2005. Zingiberis rhizome: Comprehensive review on the ginger effect and efficacy profiles. *Phytomedicine* 12: 684-701.
- Chung, K.T., Stevens, S.E., Jr-Lin, W.F. and Wie, C.I. 1993. Growth inhibition of selected food borne bacteria by tannic acid, propyl gallate and related compounds. *Letters in Applied Microbiology* 17: 29-32.

- Church, J.P., Young, J.T., Kebau, C.W., Kebay, J.C. and Ken, W.W. 1984. Relationships among dietary constituents and specific serum clinical components of subjects eating self-selected diets. *American Journal of Clinical Nutrition* 40: 1338-1344.
- Ciftci, M., Güler, T., Dalkiliç, B. and Ertas, O.N. 2005. The effect of anise oil (*Pimpinella spp.*) on broiler performance. *International Journal of Poultry Science* 4: 851-855.
- Conogullari, S., Baylan, M., Erdogan, Z., Duzguner, V. and Kucukgul, A. 2010. The effect of dietary garlic powder on performance, egg yolk and serum cholesterol concentration in laying quails. *Czech Journal of Animal Science* 55: 286-293.
- Council Regulation (EC) 1999. No. 1804/99 of July 1999 supplementing Regulation (EEC) No. 2092/91 on organic production of agricultural products. Official Journal, L 222 (24/08/1999), pp. 1–28.
- Cross, D.E., Mcdevitt, R.M. and Acamovic, T. 2011. Herbs, thyme essential oil and condensed tannin extracts as dietary supplements for broilers and their effects on performance, digestibility, volatile fatty acids and organoleptic properties. *British Poultry Science* 52: 227-237.
- Cuppett, S. L. and Hall, C. A. 1998. Antioxidant activity of Labitae. Advances in Food and Nutrition Reaserch 42:245-271.
- Daneshyar, M., Kermanshahi, H. and Golian, A. 2012. The effects of turmeric supplementation on antioxidant status, blood gas indices and mortality in broiler chickens with T3-induced ascites. *British Poultry Science* 53: 379-385.
- Das, S., Gautam, N. and Dey, S.K. 2009. Oxidative stress in the brain of nicotine-induced toxicity: protective role of Andrographis Paniculata Nees and Vitamin E. Applied Physiology, Nutrition and Metabolism 34: 124-135.
- Davidson, P.M., Sofos, J.N. and Branen, A. L. 2005. *Antimicrobials in Food* 3<sup>rd</sup> ed. pp 1-9. USA: Taylor and Francis Group.
- Davidson, P.M., and Zivanovic, S., 2003. The use of natural antimicrobials. In *Food Preservation Techniques*, eds. Zeuthen, P., Bøgh-Sørensen, L., pp 5-30. Cambridge: CRC Press.
- Davidson, P.M., 2001. Chemical preservatives and natural antimicrobial compounds. In: *Fundamentals and Frontiers*, 2nd ed. Doyle, M.P., Beuchat, L.R., Montville, T.J., eds. *Food Microbiology*, pp 593-628. Washington: ASM Press.
- Debersac P., Vernevaut, M.F., Amiot, M.J., Suschetet, M. and Siess, M.H. 2001. Effect of a water-soluble extract of rosemary and its purified component rosmarinic acid on xenobiotic metabolizing enzymes in rat liver. *Food Chemistry and Toxicology* 29: 109-117.
- de Padua, L.S., Bunyaprafatsara, N. and Lemmens R.H.M.J. 1999. Plant Resources of South-East Asia: Medicinal and Poisonous Plants eds. pp 167–175. Leiden: Backhuys Publishers.
- Devatkal, S.K., and Naveena, B.M. 2010. Effect of salt, kinnow and pomegranate fruit by-product powders on color and oxidative stability of raw ground goat meat during refrigerated storage. *Meat Science* 85 (2): 306–311.
- Dibner, J.J. and Richards, J.D. 2005. Antibiotic growth promoters in agriculture: history and mode of action. *Poultry Science* 84: 634-643.

- Dibner, J.J., Kitchell, M.L., Atwell, C.A. and Ivey, F.J. 1996. The effects of dietary ingredients and age on the microscopic structure of the gastrointestinal tract in poultry. *Journal of Applied Poultry Research* 5: 70–77.
- Din, M.G., Sunde, M.L. and Bird, H.R. 1979. Measuring metabolisable energy with mature hens. *Poultry Science* 58: 441-445.
- Dong, X.F., Gao, W.W., Tong, J.M., Jia, H.Q., Sa, R.N. and Zhang, Q. 2007. Effect of polysavone (alfalfa extract) on abdominal fat deposition and immunity in broiler chickens. *Poultry Science* 86: 1955-1959.
- Dransfield, E. and Sosnicki, A. A. 1999. Relationship between muscle growth and poultry meat quality. *Poultry Science* 78: 743-746.
- Droleskey, R.E., Oyofo, B.A., Hargis, B.M., Corrier, D.E., and De Loach, J.R. 1994. Effect of mannose on *Salmonella typhimurium*-mediated loss of mucosal epithelial integrity in cultured chick intestinal segments. *Avian Disease* 38: 275–281.
- Dung, N.T., Kim, J.M. and Kang, S.C. 2008. Chemical composition, antimicrobial and antioxidant activities of the essential oil and the ethanol extract of *Cleistocalyx operculatus* (Roxb.) Merr and Perry buds. *Food Chemistry and Toxicology* 46: 3632-3639.
- Du Plessis, S.S., McAllister, D.A., Luu, A., Savia, J., Agarwal, A. and Lampiao, F. 2010. Effects of H<sub>2</sub>O<sub>2</sub> exposure on human sperm motility parameters, reactive oxygen species levels and nitric oxide levels. *Andrologia* 42: 206-210.
- Dwivedi, S., Chansouria, J.P.N. and Somain, P.N. 1987. An experimental model for myocardial ischemia in rabbits. *Indian Journal of Experimental Biology* 25: 753–757.
- Dziezak, J. D. 1986. Antioxidant the ultimate answer to oxidation. *Food Technology* 40: 94-102.
- Ebrahimabadi, A.H., Ebrahimabadi, E.H., Djafari-Bidgoli, Z., Kashi, F.J., Mazoochi, A. and Batooli, H. 2010. Composition and antioxidant and antimicrobial activity of the essential oil and extracts of *Stachys inflate* Benth from Iran. *Food Chemistry* 119: 452-458.
- Edziri, H.L., Smach, M.A., Ammar, S., Mahjoub, M.A., Mighri, Z., Aouni, M. and Mas-touru, M. 2011. Antioxidant, antibacterial, and antiviral effects of *Lactuca sativa* extracts. *Industrial Crops and Products* 34: 1182-1185.
- Elagib, H.A.A., Nabiela, E.M., Abbass, S.A. and Ginawi, T.A.N. 2012. Effect of natural spices on plasma proteins in broiler chicks. *Journal of Nutrition and Food Science* 2:152-160
- Emadi, M. and Kermanshahi, H. 2006. Effect of turmeric rhizome powder on performance and carcass characteristics of broiler chickens. *International Journal of Poultry Science* 5: 1069-1072.
- Emami, N.K., Samie, A., Rahmani, H.R., and Ruiz-feria, C.A. 2012. The effect of peppermint essential oil and fructooligosaccharides, as alternatives to virginiamycin, on growth performance, digestibility, gut morphology and immune response of male broilers. *Animal Feed Science and Technology* 175: 57-64.

- Erel, O. 2004. A novel automated direct measurementmethod for total antioxidant capacity using a new generation, more stable ABTS radical cation. *Clinical and Biochemical*. 37 (4): 277-285
- Esonu, B.O., Emenalom, O.O., Udedibie, A.B.I., Herbert, U., Ekpor, C.F., Okoli, I.C. and Iheukwumere, F.C. 2001. Performance and blood chemistry of weaner pigs fed raw Mucana bean (velvet) meal. *Tropical Animal Production and Investigation* 4: 49-54.
- Fasina, Y.O., Hoerr, F.J., McKee, S.R. and Conner, D. E. 2010. Influence of Salmonella enterica serovar Typhimurium infection on intestinal goblet cells and villous morphology in broiler chicks. Avian Diseases 54: 841-847.
- FAO, 2010, Food and Agriculture Organization of the United Nations (FAO), Livestock and Fish Primary Equivalent, 02 June 2010, FAOSTAT online statistical service, FAO, Rome.
- Fenton J.J., 2002. *Toxicology: a case-oriented approach*. pp. 17-20. New York: CRC Press.
- Frost, A.J. 1991. Antibiotics and animal production. In: World Animal Science Microbiology of Animals and Animals Products, ed. Woolcock J.B. pp. 181-194. New York: Elsevier.
- Fulton, R. M., Nersessian, B. N., and Reed, W. M. 2002. Prevention of *Salmonella enteritidis* infection in commercial duckling by oral chicken egg-derived antibody alone or in combination with probiotics. *Poultry Science* 81: 34-40.
- Fu, Y., Zu, Y., Chen, L., Shi, X., Wang, Z., Sun, S. and Efferth, T. 2007. Antimicrobial activity of clove and rosemary oils alone and in combination. *Phytotheraphy Research* 21: 989-994.
- Furuta, A., Martin, L.J., Lin, C-.L.G., Dykes-Hoberg, M. and Rothstein, J.D. 1997. Cellular and synaptic localization of the neuronal glutamate transporters excitatory amino acid transporters 3 and 4. *Journal of Neuroscience* 81: 1031-1042.
- Gabriel, I., Mallet, S., Leconte, M., Travel, A. and Lalles, J.P. 2008. Effects of whole wheat feeding on the development of the digestive tract of broiler chickens. *Animal Feed Science and Technology* 142: 144-162.
- Gallo, M., Ferracane, R., Graziani, G., Ritieni, A. and Fogliano, V. 2010. Microwave assisted extraction of phenolic compounds from four different spices. *Molecules* 15: 6365-6374.
- Gao, P., Bermejo, R. and Zeigler, H.P. 2001. Whisker differentiation and rodent whisking patterns: behavioral evidence for a central pattern generator. *Journal of Neuroscience* 21: 5374–5380.
- Gaskins, H.R., 2001. Intestinal bacteria and their influence on swine growth. In: *Swine Nutrition*, 2<sup>nd</sup> ed. Lewis AJ and Southern LL. pp. 585-608. Boca Raton FL: CRC Press.
- Gauthier, R. 2002. XVIII Congreso Lantioamericano de Avicultura 2002. Poultry Therapeutics: New alternatives. Retrieved on 3 June 2013 from http://www.jefo.ca/pdf/ALA2003\_en.pdf.
- Giannenas, I.A., Florou-Paneri, P., Botsoglou, N.A., Christaki, E., and Spais, A.B. 2005. Effect of supplementing feed with oregano and/or αtocopheryl acetate on growth of broiler chickens and oxidative stability of meat. *Journal of Animal Feed Science* 14: 521–535.

- Giannenas, A. I., Florou-Paneri, P., Papazahariadou, M., Christaki, E., Botsoglou, N. A., and Spais, A. B. 2003. Dietary oregano essential oil supplementation on performance of broilers challenged with *Eimeria tenella*. *Archives Animal Nutrition* 57: 99-106.
- Goldberg, I.J., Eckel, R.H. and McPherson, R. 2011. Triglycerides and Heart Disease Still a Hypothesis. *Arteriosclerosis, Thrombosis, and Vascular Biology* 31: 1716-1725.
- Goñi, I., Brenes, A., Centeno, C., Viveros, A., Saura-Calixtio, F., and Rebole, A. 2007. Effect of dietary grape pomace and vitamin E on growth performance, nutrient digestibility, and susceptibility to meat lipid oxidation in chickens. *Poultry Science* 86: 508–516.
- Gonzalez, M.T.N.D., Boleman, R.M., Miller, R.K., Keeton, J.T. and Rhee, K.S. 2008. Antioxidant properties of dried plum ingredients in raw and precooked pork sausage. *Journal of Food Science* 73: 63–71.
- Govaris, A., Solomakos, N., Pexara, A. and Chatzopoulou, P.S. 2010. The antimicrobial effect of oregano essential oil, nisin and their combination against Salmonella Enteritidis in minced sheep meat during refrigerated storage. *International Journal of Food and Microbiology* 137:175–180.
- Govaris, A., Botsoglou, N., Papageorgiou, G., Botsoglou, E. and Ambrosiadis, I. 2004. Dietary versus post-mortem use of oregano oil and (or) alpha-tocopherol in turkeys to inhibit development of lipid oxidation in meat during refrigerated storage. *International Journal of Food Science and Nutrition* 55: 115-123.
- Guillen, M.D. and Manzanos, M.J. 1998. Composition of the extract in dichloromethane of the aerial parts of a Spanish wild growing plant *Thymus vulgaris* L. *Journal of Flavour Fragrance* 13: 259–262.
- Gülçin, I., Büyükokuroğlu, M.E., Oktay, M. and Küfrevioğlu, Ö.İ. 2003. Antioxidant and analgesic activities of turpentine of *Pinus nigra* Arn. subsp. *pallsiana* (Lamb.) Holmboe. *Journal of Ethnopharmacology* 86: 51-58.
- Guo, F.C., Savelkoul, H.F.J., Kwakkel, R.P., Williams, B.A. and Verstegen, M.W.A. 2003a. Immunoactive, medicinal properties of mushroom and herbpolysaccharides and their potential use in chicken diets. *Journal of World Poultry Science* 59: 427-440.
- Guo, Y., Zhang, G., Yuan, J. and Nie W. 2003b. Effects of source and level of magnesium and Vitamin E on prevention of hepatic peroxidation and oxidative deterioration of broiler meat. *Animal Feed Science and Technology* 107: 143-150.
- Hammer, K.A., Carson, C.F. and Riley, T.V. 2001. Antimicrobial activity of essential oils and other plant extracts. *Journal of Applied Microbiology* 86: 985-990.
- Han, C.J., Hussin, A.H. and Ismail, S. 2008. Toxicity study of *Orthosiphon stamineus* Benth (Misai Kucing) on Sprague Dawley rats. *Tropical Biomedicine* 25: 9-16.
- Harborne, J.B., 1993. *Introduction to Ecological Biochemistry*. 4th ed. London: Academic Press.
- Hashemi, S.R. Zulkifli, I., Davoodi, H., Zunita, Z. and Ebrahim, M. 2012. Growth performance, intestinal microflora plasma, fatty acid profile in

broiler chickens fed herbal plant (Euphorbia hirta) and mix of acidifiers. *Animal Feed Science and Technology* 178: 167-174.

- Hashemi, S.R. and Davoodi H. 2010. Phytogenics as a new class of feed additives in poultry industry. *Journal of Animal Veterinary Advanced* 9: 2955-2304.
- Hatano, T., Kagawa, H., Yasuhara, T. and Okuda, T. 1988. Two new flavonoids and other constituents in licorice root: their relative astringency and radical scavenging effects. *Chemical and Pharmaceutical Bulletin* 36: 1090-2097.
- Hernandez F., Madrid, J., Gargia, V. Orengo, J. and Megias, M.D. 2004. Influence of two plant extracts on broiler performance, digestibility and digestive organ size. *Poultry Science* 83: 169-174.
- Himani, B.A, Seema, B.B, Bhole, N.C, Mayank, Y.A, Vinod, S.D. and Mamta, S.E. 2013. Misai kuching: A glimpse of maestro. *International Journal of Pharmaceutical Sciences Review and Research* 22 (11): 55-59.
- Ho, C.H., Norhayati, I., Sulaiman, S.F. and Rosma, A. 2010. *In vitro* antibacterial and antioxidant activities of *Orthosiphon stamineus* Benth. extracts against food-borne bacteria. *Food Chemistry* 122: 1168-1172.
- Hong, J.C., Steiner, T., Aufy, A. and Lien, T.F. 2012. Effects of supplemental essential oil on growth performance lipid metabolites and immunity intestinal characteristics microbiota and carcass traits in broilers. *Livestock Science* 144: 253-262.
- Hollman, P.C. and Katan, M.B. 1999. Health effects and bioavailability of dietary flavonols. *Free Radical Research* 31: S75.
- Hossain, M.E., Kim, G.M., Lee, S.K., and Yang, C.J. 2012. Growth performance, meat yield, oxidative stability and fatty acid composition of meat from broilers fed diets supplemented with a medicinal plant and probiotics. *Asian-Australasian Journal of Animal Science* 25(8): 1159-1168.
- Hossain, M.A., Ismail, Z., Rahman, A. and Kang, S.C. 2008. Chemical composition and anti-fungal properties of the essential oils and crude extracts of *Orthosiphon stamineus* Benth. *Industrial Crops Production* 27: 328-334.
- Huff-Lonergan, E. and Lonergan, S.M. 2005. Mechanisms of water-holding capacity of meat: The role of postmortem biochemical and structural changes. *Meat Science* 71: 194-204.
- Huyghebaert, G., Ducatelle, R. and Immerseel, F.V. 2011. An update on alternatives to antimicrobial growth promoters for broilers. *Veterinary Journal* 187: 182-188.
- Ismail, S. 2000. Sayuran traditional ulam dan penyedap rasa. Malaysia: Universiti Kebangsaan Malaysia Publisher.
- Jaganath, I.B., Ng, L.T., Muthuvelu, C. and Razali, A.R. 2000. Herb, Green Pharmacy of Malaysia. Vinpress Sdn. Bhd. Malaysian Agriculture Research and Development Institute (MARDI). pp: 126.
- Jang, A., Liu, X.-D., Shin, M.-H., Lee, B.-D., Lee, S.-K., Lee, J.-H. and Jo, C. 2008. Antioxidative potential of raw breast meat from broiler chicks fed a dietary medicinal herb extract mix. *Poultry Science* 87: 2382-2389.
- Jang I.S., Y.H. Ko, S.Y. Kang and C.Y. Lee. 2007. Effect of a commercial essential oil on growth performance, digestive enzyme activity and

intestinal microflora population in broiler chickens. *Animal Feed Science and Technology* 134: 304-315.

- Jang, J.K., Pham, T.H., Chang, I.S., Kang, K.H., Moon, H., Cho, K.S. and Kim, B.H. 2004. Construction and operation of a novel mediator- and membrane-less microbial fuel cell. *Process Biochemistry* 39: 1007– 1012.
- Janz, J.A.M., Morel, P.C.H., Wilkinson, B.H.P. and Purchas, R.W. 2007. Preliminary investigation of the effects of low-level dietary inclusion of fragrant essential oils and oleoresins on pig performance and pork quality. *Meat Science* 75: 350-355.
- Jamroz, D., Wertelecki, T., Houszka, M., and Kamel. C. 2006. Influence of diet type on the inclusion of plant origin active substances on morphological and histochemical characteristics of the stomach and jejunum walls in chicken. *Journal of Animal Physiology and Animal Nutrition (Berl.)* 90:255–268.
- Jamroz D., Orda, J., Kamel, C., Wiliczkiewicz, A., Wertelecki, T. and Skorupinska, J. 2003. The influence of phytogenetic extracts on performance, nutrient digestibility, carcass characteristics and gut microbial status in broiler chickens. *Journal Animal Feed Science* 12: 583-596.
- Jayathilakan, K., Sharma, G. K., Radhakrishna, K. and Bawa, A. S. 2007. Antioxidant potential of synthetic and natural antioxidants and its effect on warmed-over-flavour in different species of meat. *Food Chemistry* 105: 908–916.
- Kamarudin Mat Salleh and Latiff, A. 2002. *Tumbuhan Ubatan Malaysia*, ed. 524p. Kuala Lumpur: UKM.
- Kang, H. K., Kang, K. H., Na, J. C., Yu, D. U., and Lee, S. J. 2008. Effect of feeding *Rhus verniciflua* extract on egg quality and performance of laying hens. *Korean Journal of Food Science and Animal Resources*. 28(5):610-615.
- Karami, M., Alimon, A.R., Sazili, A.Q., Goh, Y.M. and Ivan, M. 2011. Effects of dietary antioxidants on the quality, fatty acid profile, and lipid oxidation of *longissimus* muscle in Kacang goat with aging time. *Meat Science* 88: 102-108.
- Karou, D., Savadogo A., Canini, A., Yameogo, S., Montesano, C., Simpore, J., Colizzi, V. and Traore, A.S. 2006. Antibacterial activity of alkaloids from Sida acuta. *African Journal of Biotechnology* 5: 195-200.
- Karre, L., Lopez, K., and Getty, K.J.K. 2013. Natural antioxidants in meat and poultry products. *Meat Science* 94: 220-227.
- Kaushik, P. and Goyal, P. 2011. Evaluation of various crude extracts of *Zingiber officinale* rhizome for potential antibacterial acitivity: a study in vitro. *Advanced Microbiology* 1: 7-12.
- Kim, J.S. and Lee, Y.S. 2009. Antioxidant activity of maillard reaction products derived from aqueous glucose/glycine, diglycine, and triglycine model systems as a function. *Food Chemistry* 116: 227–232.
- Kırkpınar, F., Bora Ünlü, H. and Özdemir, G. 2011. Effects of oregano and garlic essential oils on performance, carcase, organ and blood characteristics and intestinal microflora of broilers. *Livestock Science* 137: 219-225.

- Klasing, K.C., Laurin, D.E. Peng, R.K. and Fry, D.M. 1987. Immunologically mediated growth depression in chicks: Influence of feed intake, corticosterone and interleukin-1. *Journal of Nutrition* 117: 1629-1637.
- Klaunig, J.E. and Kolaja, K.L. 1998. Chemical-induced hepatocarcinogenesis. In: *Toxicology of the Liver*, ed. Plaa, G.L. and Hewitt, W.R., pp. 93-123. Washington: Taylor and Francis.
- Krishnan, K.R., Babuskin, S., Babu, P.A.S., Sasikala, M., Sabina, K., Archana, G., Sivarajan, M. and Sukumar, M. 2014. Antimicrobial and antioxidant effects of spice extracts on the shelf life extension of raw chicken meat. *International Journal of Food Microbiology* 171: 32–40.
- Lan, Y., Verstegen, M.W.A., Tamminga, S. and Williams, B.A. 2005. The role of the commensal gut microbial community in broiler chickens. *World's Poultry Science Journal* 61: 95-104.
- Lee, K. W., Everts, H., Kappert, H. J., Frehner, M., Losa, R., and Beynen, A. C. 2003. Effects of dietary essential oil components on growth performance, digestive enzymes and lipid metabolism in female broiler chickens. *British Poultry Science* 44: 450-457
- Leelarasamee, A., Trakulsomboon, S. and Sittisomwong, N. 1990. Undetectable anti-bacterial activity of *Andrographis paniculata* (Burma) wall. ex ness. *Journal of Medical Associated Thailand* 73: 299-304.
- Lim, B.S. 2013. Poultry at top of protein pecking order. TheEdge Online Magazine, published on 25 March 2013. Retrieved 5 June 2013. http://www.theedgemalaysia.com/business-news/233893-poultry-at-topof-protein-pecking-order.html.
- Lin, F.L., Wu, S.J., Lee, S.C. and Ng, L.T. 2009. Antioxidant, antioedema and analgesic activities of *Andrographis paniculata* extracts and their active constituent andrographolide. *Phytotheraphy Research* 23: 958-964.
- Liu, H.W., Tong, J.M. and Zhou, D.W. 2011. Utilization of Chinese herbal feed additives in animal production. *Agriculture Science China* 10: 1262-1272.
- Liu, H.W., Dong, X.F., Tong, J.M. and Zhang, Q. 2010. Alfalfa polysaccharides improve the growth performance and antioxidant status of heat-stressed rabbits. *Livestock Science* 131: 88-93.
- Liu, H.W., Francesco, G., Laura, G., Alberto, B., Carola, L., Guo, K.J., Tong, J.M. and Ivo, Z. 2009. Effects of chestnut tannins on carcass characteristics, meat quality, lipid oxidation and fatty acid composition of rabbits. *Meat Science* 83: 678-683.
- Lu, M., Yuan, B., Zeng, M., and Chen, J. 2011. Antioxidant capacity and major phenolic compounds of spices commonly consumed in China. *Food Research International* 44: 530-536.
- Mahmood, S., Hassan, M.M., Alam, M. and Ahmad, F. 2009. Comparative efficacy of *Nigella sativa* and *Allium sativum* as growth promoters in broilers. *International Journal of Agriculture and Biology* 11: 775–778.
- Malaysian Standard, Halal Food- Production, preparation, handling and storage - General Guidelines. Department of Standard Malaysia MS1500: 2004.
- Mathivanan, R. and Kalaiarasi, K. 2007. Panchagavya and *Andrographis paniculata* as alternatives to antibiotic growth promoters on haematological, serum biochemical parameters and immune status of broilers. *Journal of Poultry Science* 44: 198-204.

Mathivanan, R., Edwin, S.C., Amutha, R. and Viswanathan, K. 2006. Panchagavya and *Andrographis paniculata* as alternatives to antibiotic growth promoter on broiler production and carcass characteristics. *International Journal of Poultry Science* 5: 1144-1150.

Matawalli, A. G., Samuel, A. C., and Yagana, S. 2004. Effects of methanolic leaf extract of Adansonia digitata on serum lipid levels in normal and ethanol fed rats. *Pakistan Journal of Biology and Science* 7: 1094-1095.

Matkowski, A. 2008. Antioxidant activity of extracts and different solvent fractions of *Glechoma hederacea* L. and *Orthosiphon stamineus* (Benth) Kudo. *Advances Clinical and Experimantal Medicine* 17: 615-624.

Mbikay, M. 2012. Therapeutic potential of Moringa oleifera leaves in chronic hyperglycemia and dyslipidemia: a review. Frontier in Pharmacology 3: 1-12.

Mehala, C. and Moorthy, M. 2008. Effect of *Aloe vera* and *Curcuma longa* (turmeric) on carcass characteristics and biochemical parameters of broilers. *International Journal of Poultry Science* 7: 857-861.

Molyneux, P. 2004. The use of the stable free radical diphenylpicryl-hydrazyl (DPPH) for estimation antioxidant activity. *Songklanakarin Journal of Science and Technology* 26: 211–219.

Moon, D.O., Kim, M.O., Lee, J.D., Choi, Y.H. and Kim, G.Y. 2010. Rosmarinic acid sensitizes cell death through suppression of TNFalpha-induced NF-kappaB activation and ROS generation in human leukemia U937 cells. *Cancer Letters* 288: 183-185.

Morrissey, P.A. and Kerry, J.P. 2004. Lipid oxidation and the shelf life of muscle foods. In *Understanding and measuring the shelf life of food, ed.* Steele, R. P 150. Boca, Raton: CRC Press.

Morrissey J.P. and Osbourn A.E., 1999. Fungal resistance to plant antibiotics as a mechanism of pathogenesis. *Microbiology Molecular and Biology Reviews* 63: 708-724.

Morrissey, P.A., Sheehy, P.J.A., Galvin, K., Kerry, J.P. and Buckley, D.J. 1998. Lipid stability in meat and meat products. *Meat Science* 49: 73– 86.

Musa, Y. Azimah, K. and Zaharah, H. 2009. Tumbuhan Ubatan Popular Malaysia, ed. 69p. Serdang: MARDI.

Najafi, P. and Torki, M. 2010. Performance, blood metabolites and immunocompetaence of broiler chicks fed diets included essential oils of medicinal herbs. *Journal of Animal and Veterinary Advances* 9: 1164-1168.

Narimani-Rad, M., Nobakht, A., Shahryar, H.A., Kamani, J. and Lotfi, A. 2011. Influence of dietary supplemented medicinal plants mixture (Ziziphora, oregano and peppermint) on performance and carcass characterization of broiler chickens. *Journal of Medicinal Plant Research* 5(23): 5626-5629.

National Research Council (NRC), 1996. *Guide for the care and use of laboratory animals*. Washington: National Academy Press.

Naveena, B. M., Sen, A. R., Kingsly, R. P., Singh, D. B. and Kondaiah, N. 2008. Antioxidant activity of pomegranate rind powder extract in cooked chicken patties. *International Journal of Food Science and Technology* 43: 1807–1812.

- Negi, P.S., Chauhan, A.S., Sadia, G.A., Rohinishree, Y.S. and Ramteke, R. S. 2005. Antioxidant and antibacterial activities of various seabuckthorn (*Hippophae rhamnoides* L.) seed extracts. *Food Chemistry* 92: 119-124.
- Neish, A.S. 2002. The gut microflora and intestinal epithelial cells: a continuing dialogue. *Microbes and Infection* 4: 309-317.
- Newton, S.M., Lau, C., Gurcha, S.S., Besra, G.S. and Wright, C.W. 2002. The evaluation of forty-three plant species for *in vitro* antimycobacterial acitivities: Isolation of active constituents from *Psoralea corylifolia* and *Sanguinaria canadensis*. *Journal of Ethnopharmacology* 79: 57-67.
- Nobakht, A., Mansoub, N.H. and Nezhady, M.A.M. 2012. Effect of *Melissa* officinalis L., *Tanacetum balsamita* L. and *Ziziphora clinopodioides* L. on performance, blood biochemical and immunity parameters of laying hens. *Asian Journal of Animal and Veterinary Advances* 7: 74-79.
- Noraida, A. 2005. *Penyembuhan semulajadi dengan herba*. Malaysia: PTS Millennia Sdn. Bhd.
- Parekh, J. and Chanda, S. 2010. Antibacterial and phytochemical studies on twelve species of Indian medicinal plants. *African Journal of Biomedical Research* 10: 175-181.
- Parekh, J. and Chanda, S.V. 2007. *In vitro* antimicrobial activity and phytochemical analysis of some Indian medicinal plants. *Turkey Journal of Biology* 31: 53-58.
- Petracci, M., Betti, M., Bianchi, M. and Cavani, C. 2004. Color variation and characterization of broiler breast meat during processing in Italy. *Poultry Science* 83: 2086-2092.
- Phillips, I. 2007. Withdrawal of growth-promoting antibiotics in Europe and its effects in relation to human health. *International Journal of Antimicrobial Agents* 30: 101-107.
- Platel, K. and Srinivasan, K. 2004. Digestive stimulant action of spices: a myth or reality. *Indian Journal of Medical and Research* 119(5):167-79.
- Pokorny, J., 1999. Antioxidants in food preservation. In: Handbook of Food Preservation, ed. Rahman, M.S., pp 309-338. New York: Marcel Dekker.
- Pourmorad, F., Hosseinimehr, S.J., and Shahabimajd, N. 2006. Antioxidant activity, phenol and flavonoid contents of some selected Iranian medicinal plants. *African Journal of Biotechnology* 5: 1142-1145.
- Prajjal K., Singhaa, S., Royb, S. and Deya. 2003. Antimicrobial activity of Andrographis paniculata. Fitoterapia 74: 692–694.
- Prasad, N.K., Yang, B., Yang, S., Chen, Y., Zhao, M. and Ashraf M. 2009. Identification of phenolic compounds and appraisal of antioxidant and antityrosinase activities from litchi (*Litchi sinensis* Sonn.) seeds. *Food Chemistry* 116: 1–7.
- Premanath, R. and Devi, N.L. 2011. Antibacterial, antifungal and antioxidant activities of *Andrographis paniculata* Nees. leaves. *International Journal of Pharmaceutical Science and Research* 2: 2091-2099.
- Qwele K., Hugo, A., Oyedemi, S.O., Moyo, B., Masika, P.J. and Muchenje, V. 2013. Chemical composition, fatty acid content and antioxidant potential of meat from goats supplemented with Moringa (*Moringa oleifera*) leaves, sunflower cake and grass hay. *Meat Science* 93: 455–462.

- Rao, K.Y., Vimalamma, G., Venkata, Rao, C. and Tzeng, Y.M. 2004. Flavonoids and andrographolides from *Andrographis paniculata*. *Phytochemistry* 65: 2317–2321.
- Reyes, B.A.S., Bautista, N.D., Tanquilut, N.C., Anunciado, R.V., Leung. A.B., Sanchez, G.C., Magtoto, R.L. Castronuevo, P., Tsukamura, H. and Maeda, K.I. 2006. Anti-diabetic potentials of *Momordica charantia* and *Andrographis paniculata* and their effect on estrous cyclicity of alloxaninduced diabetic rats. *Journal of Ethnopharmacology* 105: 196-200.
- Rinttilä, T. and Apajalahti, J. 2013. Intestinal microbiota and metabolites— Implications for broiler chicken health and performance. *The Journal of Applied Poultry Research* 22: 647-658.
- Rowe, L.J., Maddock, K.R., Lonergan, S.M. and Huff-Lonergan, E. 2004. Oxidative environments decrease tenderization of beef steaks through inactivation of μ-calpain. *Journal of Animal Science* 82: 3254-3266.
- Rovinsky, S.A. and Cizadlo, G.R. 1998. *Salvia divinorum* Epling et Játiva-M. (Labiatae): An ethnopharmacological investigation. *The McNair Scholarly Review* 3: 142-156.
- Roy, S., Rao, K., Bhuvaneswari, C., Giri, A. and Mangamoori, L.N. 2010. Phytochemical analysis of *Andrographis paniculata* extract and its antimicrobial activity. *World Journal of Microbiology and Biotechnology*. 26: 85-91.
- Saad B., Dakwar, S., Said, O., Abu-Hijleh, G., Al Battah, F., Kmeel A. and Aziazeh, H. 2006. Evaluation of medicinal plant hepatotoxicity in cocultures of hepatocytes and monocytes. *Evidence-based Complementary and Alternative Medicine* 3: 93-98.
- Saha, P.K., Kojima, H., Martinez-Botas, J., Sunehag, A.L. and Chan, L. 2004. Metabolic Adaptations in the Absence of Perilipin increased β-oxidation and decreased hepatic glucose production associated with peripheral insulin resistance but normal glucose tolerance in perilipin-null mice. *Journal of Biology and Chemistry* 279: 35150-35158.
- Sahib, H.B., Aisha, A.F., Yam, M.F., Asmawi, M.Z., Ismail, Z., Salhimi, S.M., Othman, N.H. and Abdul Majid, A.M.S. 2009. Anti-angiogenic and antioxidant properties of *Orthosiphon stamineus* Benth methanolic leaves extract. *International Journal of Pharmacology* 5: 162-167.
- Sample, I. 2013. Antibiotic-resistant diseases pose 'apocalyptic' threat, top expert says. The Guardian Online Magazine, published on 23 January 2013, Retrieved 10 March 2013. http://www.theguardian.com/society/2013/jan/23/antibiotic-resistantdiseases-apocalyptic-threat.
- Sapcota D.R., Islam and Medhi, A.K. 2005. Efficacy of Andrographis paniculata in ameliorating aflatoxicosis in broilers. *Indian Veterinary Journal* 82: 529-532.
- Scalbert, A. 1991. Antimicrobial properties of tannins. *Phytochemistry* 30: 3875-3883.
- Schiavone, A., Righi, F., Quarantelli, A., Bruni, R., Serventi, P. and Fusari, A. 2007. Use of Sibyllum marianum fruit extract in broiler chicken nutrition: Influence on performance and meat quality. *Journal of Animal Physiology and Animal Nutrition* 91: 256–267.
- Schulte-Hermann R. 1979. Adaptive liver growth induced by xenobiotic compounds: its name and mechanism. In: *Mechanism of Toxic Action*

*on Some Target Organs*, ed. Chambers PL and Günzel P., pp.113-124. Berlin: Springer-Verlag.

- Scramlin, S.M., Newman, M.C., Cox, R.B., Sepe, H.A., Alderton, A.L., O'Leary, J., Mikel, W.B. 2010. Effects of oregano oil brine enhancement on quality attributes of beef longissimus dorsi and semimembranosus muscles from various age animals. *Journal of Food Science* 75: 89–94.
- Seifried, H.E., Anderson, D.E., Fisher, E.I. and Milner J.A. 2007. A review of the interaction among dietary antioxidants and reactive oxygen species. *Journal of Nutrition and Biochemistry* 18(19): 567-579.
- Shahidi, F., 1997. Natural antioxidant: an overview. In: *Natural Antioxidants: Chemistry, Health Effects, and Applications,* ed. Shahidi, F., pp 1-11. Champaign: AOCS Press.
- Shahidi, F., Janitha, P.K., Wanasundara, P.D., 1992. Phenolic antioxidants. *Critical Reviews Food Science and Nutrition* 32: 67-103.
- Shan, B., Cai, Y.Z., Brooks, J.D. and Corke, H. 2009. Antibacterial and antioxidant effects of five spice and herb extracts as natural preservatives of raw pork. *Journal of Science of Food and Agriculture* 89: 1879-1885.
- Shanoon, A.K., Jassim, M.S., Amin, Q.H., and Ezaddin, I.N. 2012. Effects of ginger (*Zingiber officinale*) oil on growth performance and microbial population of broiler Ross 308. *International Journal of Poultry Science* 11: 589-593.
- Sharma, A.L., Krishnan and Handa, S.S. 1992. Standardization of Indian crude drug Kalmegh by High pressure liquid chromatographic determination of andrographaloide. *Phytochemistry Analysis* 3: 129-131.
- Sheeja K. and Kuttan, G. 2007. Activation of cytotoxic T lymphocyte responses and attenuation of tumor growth in vivo by Andrographis paniculata extract and andrographolide. *Immunopharmacology and Immunotoxicology* 29: 81-93.
- Si, W, Gong, J., Tsao, R., Zhou, T., Yu, H., Poppe, C., Johnson R. and Du, Z. 2006. Antimicrobial activity of essential oils and structurally related synthetic food additives towards selected pathogenic and beneficial gut bacteria. *Journal of Applied Microbiology* 100: 296-305.
- Simitzis, P.E., Deligeorgis, S.G., Bizelis, J.A., Dardamani, A., Theodosiou, I. and Fegeros, K. 2008. Effect of dietary oregano oil supplementation on lamb meat characteristics. *Meat Science* 79: 217–223.
- Singha, P.K., Roy, S. and Dey S. 2003. Antimicrobial activity of *Andrographis* paniculata. *Fitoterapia* 74: 692-694.
- Smeti, S., Atti, N., Mahouachi, M. and Munoz, F. 2013. Use of dietary rosemary (*Rosmarinus officinalis L.*) essential oils to increase the shelf life of Barbarine light lamb meat. *Small Ruminant Research* 113: 340–345
- Sokmen, A., Jones, B.M. and Erturk M. 1999. The in vitro antibacterial activity of Turkish plants. *Journal of Ethnopharmacology* 67: 79–86.
- SPSS Version 21. 2013. *Statistical Package for Social Science*. Chicago, Illinois: IBM SPSS.
- Sriplang, K., Adisakwattana, S., Rungsipipat A. and Yibchok-Anun, S. 2007. Effect of *Orthosiphon stamineus* aqueous extract in plasma glucose

concentration and lipid profile in normal and streptozotocin-induced diabetic rats. *Journal of Ethanopharmacology* 109: 510-514.

- Stef, D.S., Stef, L., Mot, D., Pop, C. and Hegedus, M.G. 2012. The effect of medicinal plants on broilers immunological profile and productive performances. *Journal of Food, Agriculture and Environment* 10(1): 434-437.
- Steiner, T. 2009. *Phytogenic in animal nutrition (Natural concept to optimize gut health and performance)*, ed. Nottingham: Nottingham University Press.
- Sturkie, P.O., Hazel, W. and Wood, R. 2000. *Avian Physiology*. 3<sup>rd</sup> ed. New York: Springer-Vallock.
- Tawaha, K., Alali, F.Q., Gharaibeh, M., Mohammad, M. and El-Elimat, T. 2007. Antioxidant activity and total phenolic content of selected Jordanian plant species. *Food Chemistry.* 104: 1372-1378.
- Teguia, A. and Beynen, A.C. 2004. Nutritional aspects of broiler production in small-holder farms in Cameroon. *Livestock Research for Rural Development* 16: 1-5.
- Tezuka Y., Stampoulis, P., Banskota, A.H., Awale, S., Tran, K.Q., Saiki, I., and Kadota S. 2000. Constituents of the Vietnamese medicinal plant *Orthosiphon stamineus*. *Chemical and Pharmaceutical Bulletin* 48: 1711-1719.
- Tong, W.Y., Darah, I. and Latifah, Z. 2011. Antimicrobial activities of endophytic fungal isolates from medicinal herb *Orthosiphon stamineus* Benth. *Journal of Medicinal Plants Research* 5: 831-836.
- Trivedi N.P., Rawal, U.M. and Patel, B.P. 2007. Hepatoprotective effect of andrographolide against hexachlorocyclohexane-induced oxidative injury. *Integrative Cancer Therapies*. 6: 271-280.
- Walker, J.R.L., 1994. Antimicrobial compounds in food plants. In *Natural Antimicrobial Systems and Food Preservation*, ed. Dillon, V.M. and Board, R.G., pp 205-264. England: CABI.
- Wan Hassan, W. E. 2007. *Healing Herbs of Malaysia,* ed., pp 145-164. Malaysia: Federal Land and Development Authority Malaysia (FELDA).
- Wan Zaki, W. M., Zaharah, A and Musa, Y. 2005. Hempedu bumi (*Andrographis paniculata*) In: Penanaman Tumbuhan Ubatan dan Beraroma, ed. Musa, Y. Muhammad, Ghawas, M. and Mansor, P. Pp 36-42. Serdang: MARDI.
- Wei, A. and Shibamoto, T. 2007. Antioxidant activities and volatile constituents of various essential oils. *Journal of Agriculture of Food and Chemistry* 55(5):1737-42.
- Wiart, C.2002. *Medicinal plants of South East Asia*, ed. 264p. Kuala Lumpur: Pearson Malaysia Sdn. Bhd.
- Windisch W.M., Schedle K., Plitzner, C. and Kroismayr A. 2008. Use of phytogenic products as feed additives for swine and poultry. *Journal of Animal Science* 86: 140-148.
- Wong, C., Li, H., Cheng, K. and Chen, F. 2006. A systematic survey of antioxidant activity of 30 Chinese medicinal plants using the ferric reducing antioxidant power assay. *Food Chemistry* 97: 705-711.
- Xiao, Z., Lester, G.E., Luo, Y. and Wang, Q. 2012. Assessment of vitamin and carotenoid concentrations of emerging food products: edible microgreens. *Journal of Agriculture and Food Chemistry* 60: 7644-7651.

- Xu, J., Bjursell, M.K., Himrod, J., Deng, S., Carmichael, L.K., Chiang, H.C., Hooper, L.V. and Gordon, J.I. 2003. A genomic view of the human-Bacteroides thetaiotaomicron symbiosis. *Science* 299: 2074–2076.
- Yam M.F., Basir, R., Asmawi, M.Z. and Ismail Z. 2007. Antioxidant and hepatoprotective effects of *Orthosiphon stamineus* Benth. standardized extract. *American Journal of Chinese Medicine* 35: 115-141.
- Yang, Y., Iji, P.A. and Choct, M. 2009. Dietary modulation of gut microflora in broiler chickens: a review of the role of six kinds of alternatives to in-feed antibiotics. *World's Poultry Science Journal* 65: 97-114.
- Yanishlieva, N.V., Marinova, E. and Pokorny, J. 2006. Natural antioxidants from herbs and spices. *European Journal of Lipid Science and Technology* 108 (9): 776–793.
- Yanishlieva-Maslarova, N.V. 2001. Inhibiting oxidation. In Antioxidants in food. Practical applications, ed. Pokorny, J., Yanishlieva, N. and Gordon, M., pp. 22-70. England: CRC Press.
- Yao, Y. and Ren, G. 2011. Effect of thermal treatment on phenolic composition and antioxidant activities of two celery cultivars. *LWT-Food Science and Technology*. 44: 181-185.
- Yesil-Celiktas, O., Sevimli, C., Bedir, E. and Vardar-Sukan, F. 2010. Inhibitory effects of rosemary extracts, carnosic acid and rosmarinic acid on the growth of various human cancer cell lines. *Plant Foods for Human Nutrition* 65: 158-163.
- Zaidan, M.R., Noor Rain, A., Badrul, A.R., Adlin, A., Norazah, A. and Zakiah, I. 2005. In vitro screening of five local medicinal plants for antibacterial acitivity using disc diffusion method. *Tropical Biomedicine* 22: 165-170.
- Zainol, M.K., Abd-Hamid, A., Yusof, S. and Muse, R. 2003. Antioxidative activity and total phenolic compounds of leaf, root and petiole of four accessions of *Centella asiatica* (L.) Urban. *Food Chemistry* 81: 575-581.
- Zaharah, A. 2005. Misai kucing (*Orthosiphon stamineus*). In: *Penanaman tumbuhan ubatan dan beraroma,* ed. Musa, Y., Muhammad Ghawas, M. and Mansor, P. pp 14-20. Serdang: MARDI.
- Zaharah, A., Wan Zaki, W. M., Yahya, H. and Musa, Y. 2005. Manual Teknologi Penanaman Hempedu Bumi. Serdang: Mardi
- Zargar, M., Azizah, A.H., Roheeyati, A.M., Fatimah, A.B., Jahanshiri, F. and Pak-Dek, M.S. 2011. Bioactive compounds and antioxidant activity of different extracts from *Vitex negundo* leaf. *Journal of Medicinal Plants Research* 5: 2525-2532.
- Zhang, W., Xiao, S., Samaraweera, H., Lee, E.J. and Ahn, D.U. 2010. Improving functional value of meat products. *Meat Science* 86: 15-31.
- Zhang, X.F. and Tan, B.K. 2000. Antihyperglycaemic and anti-oxidant properties of *Andrographis paniculata* in normal and diabetic rats. *Clinical and Experimental Pharmacology and Physiology* 27: 358–363.