



**UNIVERSITI PUTRA MALAYSIA**

***OPTIMIZATION OF MEDIUM AND CULTURE CONDITIONS FOR  
PECTINASE PRODUCTION BY LOCALLY ISOLATED BACTERIA  
FROM KENAF STEM***

**PUVANESWARY A/P DEGARAJAN**

**IPTPH 2014 3**



**OPTIMIZATION OF MEDIUM AND CULTURE CONDITIONS FOR  
PECTINASE PRODUCTION BY LOCALLY ISOLATED BACTERIA FROM  
KENAF STEM**

**By**

**PUVANESWARY A/P DEGARAJAN**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,  
in Fulfillment of the Requirements for the Degree of Master of Science**

**August 2014**

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in the fulfillment of the requirement for the degree of Master of Science

**OPTIMIZATION OF MEDIUM AND CULTURE CONDITIONS FOR  
PECTINASE PRODUCTION BY LOCALLY ISOLATED BACTERIA FROM  
KENAF STEM**

By

**PUVANESWARY A/P DEGARAJAN**

**August 2014**

**Chairman : Assoc. Prof. Rosfarizan Mohamad, PhD**

**Institute : Tropical Forestry and Forest Products**

Pectinases are complex and diverse group of enzymes involved in the degradation of pectic substances and have great potential to be applied in biological retting of kenaf plant (*Hibiscus cannabinus*) for high quality fibre production. In this study, bacterial isolates (*Bacillus cereus* T1 and *Enterobacter cloacae* T2) were isolated from rotten kenaf stem and the isolates were identified. Then the bacterial isolates were assayed for potential enzymes production. The best enzyme producer was selected for further medium composition and culture conditions optimization study based on conventional and response surface methodology (RSM) approaches.

The bacterial isolates were isolated from rotten kenaf stem which collected from Taman Pertanian Universiti Putra Malaysia and identified based on morphological analysis, biochemical characteristics and API kit. Then, the bacterial isolates were assayed for cellulolytic enzyme activities and pectinase production chosen. The highest pectinase activity (0.138 U/mL) at 16<sup>th</sup> h of cultivation was exhibited by *B. cereus* T1.

The different media (P1, P2 and P3) were used to analyze pectinase production by *B. cereus* T1. As a result, P3 was chosen as the best medium as P3 showed higher pectinase activity (0.18 U/mL) as compared to P1 and P2 media. In the RSM study, thirty experiments of four factors (inoculum size, temperature, pH and weight of wheat) in response to pectinase biosynthesis were carried out in shake-flask. The estimated optimize conditions of the chosen factors for the growth of *B. cereus* T1 and pectinase biosynthesis as suggested by RSM are as inoculum size (7.6%), temperature (41.85°C), pH (3.41) and weight of wheat (3.67%). By using the optimized conditions as suggested by the RSM, an increment of pectinase activity (6.52 U/mL) by 47-fold was achieved as compared to non optimized conditions.

The optimized conditions were then applied in the cultivation process using 2 L stirred tank bioreactor. However, reduced pectinase activity (2.925 U/mL) was obtained in the bioreactor experiment. Further optimization study considering other bioprocessing factors should increase the pectinase production by the isolate.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia  
sebagai memenuhi keperluan untuk Ijazah Sains Sarjana

**PENGOPTIMUMAN MEDIA DAN KEADAAN KULTUR UNTUK  
PENGHASILAN PEKTINASE OLEH PENGASINGAN BAKTERIA  
DARIPADA BATANG KENAF**

Oleh

**PUVANESWARY A/P DEGARAJAN**

**Ogos 2014**

**Pengerusi : Assoc. Prof. Rosfarizan Mohamad, PhD**

**Institut : Perhutanan Tropika dan Produk Hutan**

Pektinase adalah kumpulan kompleks dan pelbagai enzim yang terlibat dalam degradasi bahan pektik dan mempunyai potensi yang sangat besar untuk diaplikasikan dalam pengeretan secara biologi bagi pokok kenaf (*Hibiscus cannabinus*) untuk penghasilan fiber yang berkualiti tinggi. Dalam kajian ini, jenis bakteria (*Bacillus cereus* T1 dan *Enterobacter cloacae* T2) telah diasingkan daripada batang kenaf yang reput dan bakteria tersebut telah dikenal pasti. Kemudian kedua-dua bakteria tersebut telah disaring untuk pengeluaran enzim yang berpotensi. Bakteria yang terbaik dalam penghasilan enzim telah dipilih untuk digunakan dalam kajian komposisi media dan pengoptimuman keadaan kultur berdasarkan kaedah konvensional dan kaedah ransangan permukaan (RSM).

Bakteria tersebut telah diasingkan daripada batang kenaf reput yang diperolehi dari Taman Pertanian Universiti Putra Malaysia dan dikenalpasti berdasarkan analisis ciri-ciri morfologi, analisis biokimia dan kit API. Kemudian, kedua-dua bakteria telah disaring untuk aktiviti enzim selulase dan pektinase dipilih sebagai enzim yang terbaik. Aktiviti pektinase tertinggi (0.138 U/mL) pada jam ke-16 pengkulturan telah dihasilkan oleh *B. cereus* T1.

Media yang berbeza (P1, P2 dan P3) telah digunakan untuk analisis biosintesis pektinase oleh *B. cereus* T1. P3 dipilih sebagai media terbaik kerana P3 menunjukkan aktiviti pektinase tertinggi (0.18 U/mL) berbanding P1 dan P2. Dalam kajian RSM, tiga puluh eksperimen untuk empat faktor (saiz inokulum, suhu, pH dan berat gandum) yang bertindakbalas terhadap penghasilan pektinase telah dijalankan di dalam kelalang kon. Jangkaan keadaan faktor yang optimum untuk pertumbuhan *B. cereus* T1 dan penghasilan pektinase seperti yang disyorkan oleh RSM adalah seperti berikut; saiz inokulum (7.6%), suhu (41.85°C), pH (3.41) dan berat gandum (3.67%). Dengan menggunakan keadaan optimum seperti yang disarankan RSM,

peningkatan aktiviti enzim pektinase sebanyak gandaan 47 telah dicapai berbanding keadaan yang tidak optimum.

Keadaan optimum kemudiannya diaplikasikan dalam proses pertumbuhan dengan menggunakan bioreaktor 2 L yang berpengaduk. Bagaimanapun, penurunan aktiviti enzim pektinase telah diperoleh melalui eksperimen menggunakan bioreaktor. Kajian pengoptimuman lanjutan yang mengambil kira faktor bioproses lain sepatutnya dapat meningkatkan penghasilan pektinase oleh bakteria tersebut.



## ACKNOWLEDGEMENT

First and foremost I would like to thank my advisor, Associate Professor Dr. Rosfarizan binti Mohamad for suggesting this wonderful topic, invaluable guidance, constructive suggestions and her monitoring throughout the project. Without her patience and encouragement, this thesis would not have been possible. I sincerely appreciate invaluable academic and personal support I received from her throughout this thesis. It was a wonderful experience working with such a great advisor.

I would like to extend my gratitude to my co-supervisors; Dr. Wan Zuhainis binti Saad and Professor Dr. Paridah binti Md Tahir for their professional guidance, moral support and helpfulness throughout my research. Special thanks are also due to all of them for giving me full freedom to pursue my research in my own work style.

I would like to grab this golden opportunity to thank all the lab assistance, Mrs. Noriza, Mr. Rosli, Mrs. Renuka, Mrs. Aluyah, Mrs. Ina and Mrs. Aishah for their endless help to carry out all the experiments smoothly. My special appreciation is also expressed to my lovely seniors, Mrs. Kohgilaa and Mr. Sureshkumar and other labmates for their help and support throughout my studies.

Finally, my further gratitude goes to my family; my parents, my siblings, my in-laws and my niece for their moral support, financial support and concern which made me complete this project successfully. Thank you for your love, support, and patience. I am truly blessed to have you as my family.



## APPROVAL SHEET

I certify that a Thesis Examination Committee has met on 6<sup>th</sup> August 2014 to conduct the final examination of Puvaneswary A/P Degarajan on her thesis entitled “Optimization of Medium and Culture Conditions For Pectinase Production by Locally Isolated Bacteria From Kenaf Stem” in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Masters of Science.

Members of the Thesis Examination Committee were as follows:

**Shuhaimi bin Mustafa, PhD**  
Professor  
Halal Products Research Institute  
Universiti Putra Malaysia  
(Chairman)

**Luqman Chuah, PhD**  
Professor  
Faculty of Engineering  
Universiti Putra Malaysia  
(Internal Examiner)

**Nor’ Aini binti Abdul Rahman, PhD**  
Associate Professor  
Faculty of Biotechnology and Biomolecular Science  
Universiti Putra Malaysia  
(Internal Examiner)

**Awang Ahmad Sallehin bin Awang Husaini, PhD**  
Associate Professor  
Faculty of Resource Science and Technology  
Universiti Malaysia Sarawak  
(External Examiner)

---

**SEOW HENG FONG, PhD**  
Professor and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia  
Date:

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirements for the degree of Master of Science. The members of the Supervisory Committee were as follows:

**Rosfarizan binti Mohamad, PhD**

Associate Professor  
Institute of Tropical Forestry and Forest Products  
Universiti Putra Malaysia  
(Chairman)

**Wan Zuhainis Saad, PhD**

Senior lecturer  
Faculty of Biotechnology and Biomolecular Science  
Universiti Putra Malaysia  
(Member)

**Paridah binti Md Tahir, PhD**

Professor and Director  
Institute of Tropical Forestry and Forest Products  
Universiti Putra Malaysia  
(Member)

---

**BUJANG BIN KIM HUAT, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:

## DECLARATION

### Declaration by graduate student

I hereby confirm that:

- This thesis is my original work;
- Quotations, illustrations and citations have been duly referenced;
- This thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- Intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- Written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- There is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: \_\_\_\_\_ Date: \_\_\_\_\_

Name and Matric No:

\_\_\_\_\_

## Declaration by graduate student

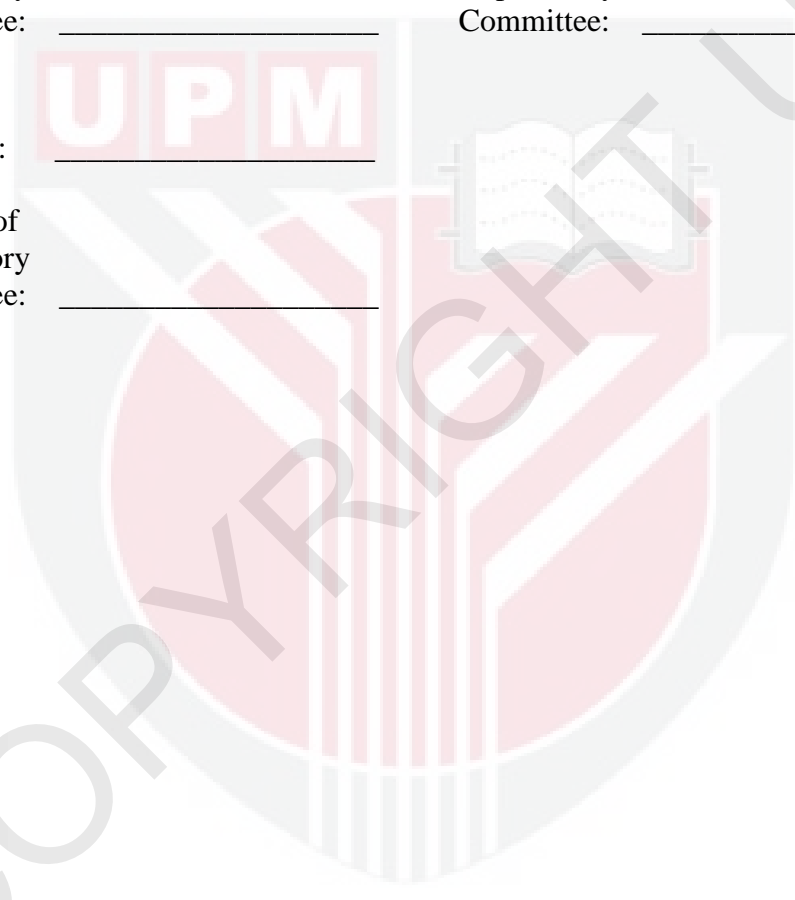
This is to confirm that:

- The research conducted and the writing of this thesis was under our supervision;
- Supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: \_\_\_\_\_  
Name of  
Chairman of  
Supervisory  
Committee: \_\_\_\_\_

Signature: \_\_\_\_\_  
Name of  
Member of  
Supervisory  
Committee: \_\_\_\_\_

Signature: \_\_\_\_\_  
Name of  
Member of  
Supervisory  
Committee: \_\_\_\_\_



© COPYRIGHT UPM

## TABLE OF CONTENTS

|   | Page        |
|---|-------------|
| <b>ABSTRACT</b>                           | <b>i</b>    |
| <b>ABSTRAK</b>                            | <b>iii</b>  |
| <b>ACKNOWLEDGEMENT</b>                    | <b>v</b>    |
| <b>APPROVAL</b>                           | <b>vi</b>   |
| <b>DECLARATION</b>                        | <b>viii</b> |
| <b>LIST OF TABLES</b>                     | <b>xiii</b> |
| <b>LIST OF FIGURES</b>                    | <b>xiv</b>  |
| <b>LIST OF ABBREVIATIONS</b>              | <b>xvi</b>  |
| <br>                                      |             |
| <b>CHAPTER</b>                            |             |
| <br>                                      |             |
| <b>1.0 INTRODUCTION</b>                   | <b>1</b>    |
| <br>                                      |             |
| <b>2.0 LITERATURE REVIEW</b>              |             |
| 2.1 <i>Hibiscus cannabinus</i>            | 4           |
| 2.1.1 History of Kenaf                    | 4           |
| 2.1.2 Characteristics of Kenaf Plant      | 4           |
| 2.1.3 Chemical Composition of Kenaf       | 6           |
| 2.1.4 Physical Properties of Kenaf        | 9           |
| 2.2 Microorganism                         | 10          |
| 2.2.1 <i>Bacillus</i> spesies             | 10          |
| 2.2.2 <i>Bacillus cereus</i>              | 10          |
| 2.3 Pectinase Biochemistry                | 10          |
| 2.3.1 Chemical Structure                  | 10          |
| 2.3.2 Pectinase Biosynthesis              | 11          |
| 2.4 Sources and Application of Pectinase  | 14          |
| 2.4.1 Sources of Pectinase                | 14          |
| 2.4.2 Application of Pectinase            | 14          |
| 2.5 Cultivation Techniques                | 15          |
| 2.5.1 Batch Culture                       | 15          |
| 2.6 Medium Composition                    | 17          |
| 2.6.1 Carbon Source                       | 17          |
| 2.6.2 Nitrogen Source                     | 17          |
| 2.6.3 Temperature and pH                  | 17          |
| 2.7 Response Surface Methodology          | 18          |
| 2.8 Concluding Remarks                    | 19          |
| <br>                                      |             |
| <b>3.0 MATERIALS AND METHODS</b>          |             |
| 3.1 Isolation of Bacteria from Kenaf Stem | 20          |
| 3.2 Identification of Isolates            | 20          |
| 3.2.1 Gram Staining and Microscopic View  | 20          |
| 3.2.2 Biochemical Tests                   | 20          |

|            |  |           |
|------------|--|-----------|
| 3.2.3      | Identification of Isolates using API Kit   | 22        |
| 3.3        | Assay of Cellulolytic Enzymes Activities   | 22        |
| 3.3.1      | Assay of Carboxymethylcellulase(CMCCase) Enzyme  | 22        |
| 3.3.2      | Assay of Filter Paper Enzyme   | 23        |
| 3.3.3      | Assay of Pectinase Enzyme  | 24        |
| 3.3.4      | Assay of Xylanase Enzyme   | 25        |
| 3.4        | Batch Cultivation  | 25        |
| 3.4.1      | Microorganisms and Maintenance   | 25        |
| 3.4.2      | Inoculum Preparation   | 26        |
| 3.4.3      | Production Medium Preparation  | 26        |
| 3.4.4      | Shake-flask  | 26        |
| 3.4.5      | Stirred Tank Bioreactor  | 27        |
| 3.4.6      | Optimization of Medium Formulation and Culture Conditions  | 29        |
| 3.4.7      | Analytical Methods   | 31        |
| 3.4.8      | Statistical Analysis   | 31        |
| 3.5        | General Experimental Design  | 32        |
| <b>4.0</b> | <b>RESULTS AND DISCUSSION</b>  |           |
| 4.1        | Screening and Isolation of Pectinase Producer from Kenaf Stem  | 34        |
| 4.2        | Preliminary Identification of Bacterial Isolates   | 34        |
| 4.2.1      | Gram staining and Microscopic View   | 34        |
| 4.2.2      | Biochemical Characteristics  | 35        |
| 4.2.3      | Identification of Isolates using API Kit   | 37        |
| 4.3        | Assay of Cellulolytic Enzymes Activities   | 38        |
| 4.3.1      | Carboxymethylcellulase (CMCase) Activity   | 38        |
| 4.3.2      | Filter Paper Activity  | 39        |
| 4.3.3      | Pectinase Activity   | 40        |
| 4.3.4      | Xylanase Activity  | 41        |
| 4.4        | Batch Cultivation  | 43        |
| 4.4.1      | Influence of Carbon Source and Medium Initial pH on Pectinase Biosynthesis by <i>B. cereus</i> T1  | 43        |
| 4.4.2      | Optimization of Medium Formulation and Culture conditions for Pectinase Production by <i>B. cereus</i> T1 using Response Surface Methodology | 47        |
| 4.4.3      | 2 L Stirred Tank Bioreactor  | 55        |
| 4.5        | Comparison on the Performance of <i>B. cereus</i> T1 Batch Cultivation on Different Conditions and Scales for Pectinase Production           | 56        |
| <b>5.0</b> | <b>CONCLUSIONS AND SUGGESTIONS FOR FUTURE WORK</b>   | <b>58</b> |

|                            |    |
|----------------------------|----|
| <b>REFERENCES</b>          | 60 |
| <b>APPENDICES</b>          | 70 |
| <b>BIODATA OF STUDENT</b>  | 72 |
| <b>LIST OF PUBLICATION</b> | 73 |



## LIST OF TABLES

| Table |   | Page |
|-------|---|------|
| 2.1   | The applications and products from kenaf in different industries reported in literature.  | 6    |
| 3.1   | Production medium.  | 26   |
| 3.2   | The dimensions and operating variables of 2 L stirred tank bioreactor.  | 29   |
| 3.3   | Medium composition used for pectinase production.   | 30   |
| 3.4   | Coded and real value of variables selected for CCD.   | 31   |
| 4.1   | Macroscopic and microscopic images of bacterial isolates from kenaf stem (T1 and T2).   | 35   |
| 4.2   | Biochemical test results for bacterial isolates T1 and T2.  | 36   |
| 4.3   | Genus of bacterial isolates T1 and T2 based on Bergey's Manual of Determinative Bacteriology, 8th edition.  | 36   |
| 4.4   | Bacterial isolates identities (%) based on API Kit results.   | 37   |
| 4.5   | The kinetic parameter values of enzyme biosynthesis between isolate T1 and T2.  | 43   |
| 4.6   | Medium composition used for pectinase production.   | 44   |
| 4.7   | The performance and kinetic parameter values of pectinase biosynthesis by <i>B.cereus</i> T1.   | 45   |
| 4.8   | Central Composite Design with a real value and response to pectinase activity (actual and predicted values)   | 48   |
| 4.9   | Analysis of variance (ANOVA) for the quadratic model of pectinase biosynthesis.   | 50   |
| 4.10  | The performance and kinetic parameter values of pectinase biosynthesis by <i>B. cereus</i> T1 in 2 L bioreactor.  | 56   |
| 4.11  | The performance and kinetic parameter values of pectinase biosynthesis by <i>B. cereus</i> T1 cultivation in shake-flask (non optimized and optimized after RSM) and 2 L stirred tank bioreactor. | 57   |



## LIST OF FIGURES

| Figure |  | Page |
|--------|--|------|
| 2.1    | Chemical structure of cellulose.   | 7    |
| 2.2    | Chemical structure of hemicellulose.   | 8    |
| 2.3    | Chemical structure of pectin.  | 11   |
| 2.4    | Action mode of pectinases.   | 12   |
| 2.5    | Growth profile of a microorganism under batch culture condition.   | 15   |
| 2.6    | Central composite design (CCD) for 3 design variables at 2 levels.   | 19   |
| 3.1    | Schematic diagram of a stirred tank bioreactor for the growth of <i>Bacillus cereus</i> T1 for pectinase production.   | 28   |
| 3.2    | The flow of general experimental design.   | 33   |
| 4.1    | Profile of CMCase activity during batch cultivation of bacterial <i>B.cereus</i> T1 and <i>E. cloacae</i> T2 in shake-flask culture using nutrient broth supplemented with carboxymethylcellulose (1%) as substrate.   | 39   |
| 4.2    | Profile of FPase activity during batch cultivation of bacterial <i>B.cereus</i> T1 and <i>E. cloacae</i> T2 in shake-flask culture using nutrient broth supplemented with Whatman No 1 filter paper (1%) as substrate. | 40   |
| 4.3    | Profile of Pectinase activity during batch cultivation of bacterial <i>B.cereus</i> T1 and <i>E. cloacae</i> T2 in shake-flask culture using nutrient broth supplemented with polygalacturonic acid (1%) as substrate. | 41   |
| 4.4    | Profile of Xylanase activity during batch cultivation of bacterial <i>B.cereus</i> T1 and <i>E. cloacae</i> T2 in shake-flask culture using nutrient broth supplemented with xylan birchwood (0.25%) as substrate.     | 42   |
| 4.5    | Time course of pectinase activity using different medium by <i>B. cereus</i> T1.   | 45   |
| 4.6    | Time course of cell concentration on pectinase production by <i>B. cereus</i> T1.  | 46   |
| 4.7    | Time course of substrate concentration on pectinase production by <i>B. cereus</i> T1.   | 47   |

|      |   |    |
|------|---|----|
| 4.8  | Surface plot of response pectinase activity of <i>B. cereus</i> T1 with factors of inoculum size (A) and temperature (B). | 52 |
| 4.9  | Surface plot of response pectinase activity of <i>B. cereus</i> T1 with factors of inoculum size (A) and pH (C).          | 52 |
| 4.10 | Surface plot of response pectinase activity of <i>B. cereus</i> T1 with factors of inoculum size (A) and wheat (D).       | 53 |
| 4.11 | Surface plot of response pectinase activity of <i>B. cereus</i> T1 with factors of temperature (B) and pH (C).            | 53 |
| 4.12 | Surface plot of response pectinase activity of <i>B. cereus</i> T1 with factors of temperature (B) and wheat (D).         | 54 |
| 4.13 | Surface plot of response pectinase activity of <i>B. cereus</i> T1 with factors of pH (C) and wheat (D).                  | 54 |
| 4.14 | Time course of pectinase activity, cell and substrate concentration by <i>B. cereus</i> T1 in 2 L bioreactor.             | 55 |

## LIST OF ABBREVIATIONS

|            |   |
|------------|---|
| NA         | Nutrient Agar                                   |
| PGA        | Polygalacturonic Acid                           |
| rpm        | Rotation per minutes                            |
| m          | meter   |
| mm         | Millimeter                                      |
| RSM        | Response surface methodology                    |
| CCD        | Central composite design                        |
| h          | Hour  |
| min        | Minutes   |
| ml         | milliliter                                      |
| OD         | Optical density                                 |
| L          | Liter   |
| $X_{\max}$ | Maximum cell concentration (g/L)                |
| $P_{\max}$ | Pectinase activity (U/mL)                       |
| $Y_{x/s}$  | Growth yield coefficient (g/g)                  |
| $Y_{p/s}$  | Pectinase yield per substrate utilized (U/mL/g) |

## CHAPTER 1

### INTRODUCTION

Enzymes are known as biological catalysts with high selectivity, which has been used in the food industry and also important for other industries such as washing agents, textile manufacturing, pharmaceuticals, pulp and paper for hundreds of years. Recently, enzymes are becoming important in sustainable technology and green chemistry and according to many researchers and studies, 20% of world chemical products will be produced by biotechnological approach in a worth of 300 billion US dollar. Microorganisms, fungus or bacteria are commonly used as sources as enzyme producer industrially. Microorganisms have a few advantages; interaction or involved in various substrates and selection of method on biosynthesis through the suitable conditions cultivation (Bhardwaj and Garg, 2010). One of the industrially important enzymes and intensively studied is pectinase. Pectinases are complex and diverse group of enzymes involved in the degradation of pectic substances (Sathyanarayana et al., 2007).

Pectinases are declared as one of the upcoming enzymes of the commercial sector. According to (Jayani et al., 2005), the microbial pectinases account for 25% of global food enzymes. These pectinases are known as heterogeneous group related enzymes which breakdown pectin substances that present in plants. Pectin is complex polysaccharide consisting mainly of esterified D-galacturonic acid residues in  $\alpha(1-4)$  chain (Shembekar and Dhotre, 2009).

Pectinases or pectinolytic enzymes are naturally distributed in microorganisms and higher plants (Whitaker, 1990). They are crucial for plants because aid in extension of cell wall and also play the important role as softening the plants tissues during maturation and storage (Ward and Moo-Young, 1989; Sakai, 1992; Aguilar and Huirton, 1990). Besides, it helps to maintain the ecological balance by decomposition and recycling of waste plant materials (Jayani et al., 2005).

Pectinases have potential applications in cotton scouring and degumming of plant fibres to improve the quality of fibre (Salazar and Jayasinghe, 1999). Pectinases also widely used in industry for retting of natural fibre and extraction of oils from vegetables and citrus peels (Bacarat et al., 1989; Brumano et al., 1993). Treatment of jute, flax, ramie and kenaf with pectinase improves the mechanical properties of the fibres, increasing flexibility with good retention of tensile strength. Besides, pectinases also used for clarification of juices, increases of filtration efficiency (Brawman, 1981), in maceration and liquefaction (Arunachalam and Asha, 2010).

In Malaysia, kenaf is an emerging new crop to the agriculture industry (Kassim, 2001). Research on kenaf typically have been done by the Institute of Tropical Forestry and Forest Products (INTROP) and Malaysian Agricultural Research and Development Institute (MARDI). INTROP identified nine varieties of kenaf which can be categorized as early flowering varieties (V132, V36, V19, V12, Q-Ping, KK60), intermediate flowering varieties (V133, NS) and late peak varieties (TK). In Malaysia, variety TK as the potential fibre source to be used in pulp and paper process, particle board and fibreboard (H'ng et al., 2009).

Currently, microbial pectinase production has been widely studied by researchers and experts (Kashyap et al., 2001; Torres et al., 2006). Pectinolytic enzymes have been reported to produce by bacteria and fungi such as *Bacillus* spp., *Clostridium* spp., *Pseudomonas* spp., *Penicillium* spp., *Aspergillus* spp., *Monilla laxa*, *Fusarium* spp., *Verticillium* spp., *Sclerotinia libertiana*, *Coniothyrium diplodiella* and *Polyporus squamosus* (Bhardwaj and Garg, 2010).

Higher cost of production is probably the critical constraint in commercialization of new sources of enzymes. The cost of enzyme production can be reduced in industrial processes due to several factors; using high producing strain, optimum fermentation conditions and cheap raw material (Murad and Azzaz., 2011) or organic nature of agro-industrial crops and residues as a carbon source (Pandey, 2001). In this study, wheat has been used as inexpensive carbon source for pectinase production by locally isolated bacteria of *Bacillus cereus* T1.

Optimization of fermentation conditions is an important step in the development of feasible industrial bioprocess. Response surface methodology (RSM) is a tool that used to analyze the optimized conditions which enables the experts or researchers to design the experimental and the RSM initially introduced by (Box and Wilson, 1951). Nowadays, researchers used this tool to provide maximum yield production through an experimental design that suggested by RSM (Raissi, 2009). Besides, RSM is a combination of good experimental design, regression modeling techniques and optimization for process improvement. Therefore, RSM is a most significant use approach prior to industrial level production (Montgomery and Myers, 1997)

The present study focuses on the search of local bacterial pectinase producer and strategies in improving the pectinase production via optimization of medium and culture conditions of the selected producer cultivation process. The optimized environmental conditions were applied in the bioreactor system cultivation equipped with a process control system. Hence, the specific objectives of this study were;

1. To screen, isolate and identify the bacterial pectinase producer isolated from rotten *Hibiscus cannabinus* (kenaf) stem.
2. To optimize the medium and culture conditions for pectinase production by the selected bacterial isolate via conventional and RSM approaches.
3. To evaluate the performance of the bacterial isolate in batch cultivation process for pectinase production using 2 L stirred tank bioreactor.



## REFERENCES

- Abdul Khalil, H.P.S., Ireana Yusra, A.F., Bhat, A.H. and Jawaid, M. (2010). Cell wall ultrastructure, anatomy, lignin distribution and chemical composition of Malaysian cultivated kenaf fiber. *Industrial Crops and Product* 31(1): 113 – 121.
- Adamsen, A., Akin, D. and Rigsby, L. (2002). Chelating agents and enzyme retting of flax. *Textile Research Journal* 72: 296 – 302.
- Aguilar, G. and Huirton, C. (1990). Constitutive exo-pectinase produced by *Aspergillus* species CH- Y- 1043 on different carbohydrate source. *Biotechnology Letters* 12: 655-660.
- Ahlawat, S., Dhiman, S.S., Battan, B., Mandhan, R.P. and Sharma, J. (2009). Pectinase production by *Bacillus subtilis* and its potential application in biopreparation of cotton and micropoly fabric. *Process Biochemistry* 44(5): 521-526.
- Alexopoulou, E., Papatheohari, Y. and Kipriotis, E. (2007). Response of kenaf (*Hibiscus cannabinus* L.) growth and yield to fertilization. *Journal of Food, Agriculture and Environment* 5(2): 228 – 232.
- Alkorta, I., Gabirsu, C., Lhama, M.J. and Serra, J.L. (1998). Industrial applications of pectic enzymes: a review. *Process Biochemistry* 33: 21-28.
- Angayarkanni, J., Palaniswamy, M., Murugesan, S. and Swaminathan, K. Improvement of tea leaves fermentation with *Aspergillus* species pectinase. *Journal Bioscience Bioengineering* 94: 299-303.
- Ariogele, K.H., Sano, Y., Uraki, Y., Sasaya, N. and Sameshima, K. (1993). *Separation and utilization of kenaf components obtained by continuous acetic acid pulping under normal pressure*. Paper presented at Japan Wood Research Society, Hokkaido Branch.
- Arunachalam, C. and Asha, S. (2010). Pectinolytic enzyme. A review of new studies. *Journal of Advanced Biotechnology*.
- Ashori, A., Jalaludin, H., Raverty, W.D. and Mohammad Nor, M.Y. (2006). Chemical and morphological characteristics of Malaysian cultivated kenaf (*Hibiscus cannabinus*) fibre. *Polymer Plastics Technology and Engineering* 45: 131 – 134.
- Bacarat, M.C., Valentin, C., Muchovej, J. and Silva, D.O. (1989). Selection of pectinolytic fungi for degumming of natural fibres. *Biotechnology Letters* 11: 899 – 902.



- Bailey, M.J. and Poutanen, K. (1989). Production of xylanotic enzymes by strain of *Aspergillus*. *Applied Microbiology Biotechnology* 30: 5 – 10.
- Bajaj, B.K., Wani, M.A., Sharma, A. and Pangotra, H. (2009). Partial purification and characterization of highly thermostable PH stable endoglucanase from a newly isolated *Bacillus* strain M-9. *Indian Journal of Chemical Technology* 16: 382 – 387.
- Bayoumi, R.A., Yassin, H.M., Swelim, M.A. and Abdel-All, E.Z. (2008). Production of bacterial pectinases from agro-industrial wastes under solid state fermentation conditions. *Journal of Applied Sciences Research* 4(12) : 1708-1721.
- Bhardwaj, V. and Garg, N. (2010). Exploitation of microorganisms for isolation and screening of pectinase from environment. Globelics 2010 8<sup>th</sup> International Conference, University of Malaya, Kuala Lumpur. November 2010.
- Benjamin, C. (2004). Pearson Education Inc.
- Berg, B. and Petersson, G. (1977). Location and formation of cellulases in *Trichoderma viride*. *Journal of Applied Bacteriology* 42: 65 – 75.
- Berner, R.A. (2003). The long-term carbon cycle, fossil fuels and atmospheric composition. *Nature* 426: 323 – 326.
- Biermann, C.J. (1996). Handbook of pulping and paper making (Second ed.). San Diego, California. Academic Press.
- Blanco, P., Sieiro, C. and Villa, T.G. (1999). Production of pectic enzymes in yeast. *FEMS Microbiology Letters* 175: 1 – 9.
- Blanco, P., Sieiro, C., Diaz, A. and Villa, T.G. (1994). Production and partial characterization of an endopolygalacturonase from *Saccharomyces cerevisiae*. *Canada Journal Microbiology* 40: 974 – 977.
- Blandino, A., Dravillas, K., Pandiella, S.S. and Webb, C. (2001). Utilisation of whole wheat flour for the production of extracellular pectinases by some fungal strains. *Process Biochemistry* 37: 497 – 503.
- Box, G.E.P. and Wilson, K.B. (1951). On the experimental attainment of optimum conditions. *Journal Royal Statistic Society* 1: 16 – 29.
- Brawman, J.W. (1981). Application of enzymes in fruit juice technology. In: Birch, G.G., Blakcorough, N. and Barker, J.K. (Editors). *Enzymes and food processing*. London. *Applied Science* : 247-261.



- Brown, N. and Dowds, B. (2001). Heat and salt stress in the food pathogen *Bacillus cereus*. *Journal of Applied Microbiology* 91(6): 1085 – 1094.
- Brumano, M.H.N., Coelho, J.L.C., Araujo, E.F. and Silva, D.O. (1993). Pectin lyase activity of *Penicillium griseoroseum* related to degumming of ramie. *Review Microbiology* 24(3): 175 – 178.
- Candilo, M.D, Bonatti, P.M., Guidetti, C., Focher, B., Grippo, C., Tamburini, E. and Mastromei, G. (2009). Effects of pectinolytic bacterial strains on water-retting of hemp and fibre properties. *Journal of Applied Microbiology* 108: 194 – 203.
- Carmano, E.C., Pedrolli, D.B., Monteiro, E.C. and Gomes, E. (2009). Production, characterization and industrial application of microbial pectinolytic enzymes. *Open Biotechnology Journal* 3 : 9-18.
- Coutinho, P.M. and Henrissat, B. (1999). Carbohydrate-active enzymes: an inte-grated database approach. In "Recent Advances in Carbohydrate Bioengineering". Gilbert, H.J., Davies, G., Henrissat, B. and Svensson, B. (Editors). Cambridge: *The Royal Society of Chemistry*: 3-12.
- Dempsey, J.M. (1967). An economic comparison of kenaf and other crops. Tehran, Iran: U.S Agency for International Development.
- Dutt, D., Upadhay, J.S., Singh, B. and Tyagi, C. H. (2009). Studies on *Hibiscus cannabinus* and *Hibiscus sabdariffa* as an alternative pulp blend for softwood: An optimization of kraft delignification process. *Industrial Crops and Products* 29(1): 16 – 26.
- Elliot, S. (1953). Progress of the Mexican kenaf industry. *Textile Quart (Belfast)* 3: 243 – 246.
- Esquivel, J.C.C., Hours, R.A., Voget, C.E. and Mignone, C.F. (1999). *Aspergillus kawachii* produces an acidic pectin releasing enzyme activity. *Journal of Bioscience and Bioengineering* 88(1): 48 – 52.
- Falkowski, P., Scholes, R.J., Boyle, E., Canadell, J., Canfield, D., Elser, J. (2000). The global carbon cycle: a test of our knowledge of earth as a system. *Science* 290: 291 – 296.
- FAO. (2006). Jute, Kenaf, Sisal, Abaca, Coir and Allied Fibers Statistics. Retrieved on 20 January 2011 from [http://www.fao.org/es/esc/en/20953/21005/highlight\\_51023en.html](http://www.fao.org/es/esc/en/20953/21005/highlight_51023en.html).

- Fisher, G. (1994). *Availability of kenaf fibers for the US paper industry*. Paper presented at the Tappi pulping conference proceeding, Book 2, November 6-10, Sheraton Harbor Island, San Diego, California, Tappi Press, Atlanta, GA, USA.
- Goncalves, D.B., Teixeira, J.A., Bazzali, D.M.S., De Queiroz, M.V. and De Araujo, E.F. (2010). Use of response surface methodology to optimize production of pectinase by recombinant *Penicillium griseoroseum* T20 1: 140 – 146.
- Goring, D.A.I. (1971). Polymer properties of lignin and lignin derivatives. *Lignins: Occurrence, formation, structure and reactions*: 695-768.
- Hao, X.C., Yui, X.B. and Yan, Z.L.(2006). Optimization of the medium for the production of cellulase by the mutant *Trichoderma reesei* WX-112 using response surface methodology. *Food Technology and Biotechnology* 44: 89-94.
- Helga, D., Mats, A. and Jens, N. (2003). Reconstruction of the central carbon metabolism of *Aspergillus Niger*. *Journal Biochemistry* 270: 4243 -4253.
- H'ng, P.S., Nobuchi, T., Khor, B.N, Chin, K.L., Abdul Shukor, N.A. and Paridah, M.T. (2009). Anatomical structure, fiber morphology and chemical constituents of different kenaf varieties. *7<sup>th</sup> Pacific Regional Wood Anatomy Conference (PRWAC)*, 57.
- Hoondal, G.S., Tiwari, R.P., Tiwari, R., Dahiya, N. and Beg, Q. K. (2002). Microbial alkaline pectinases and their industrial applications: a review. *Applied Microbiology Biotechnology* 59: 409 – 418.
- Hoskisson, P.A and Hobbs, G. (2005). Continuous culture making a comeback. *Microbiology* 151 : 3153 – 3159.
- Jalaluddin, H. (2001). Progress report of national kenaf technical committee on kenaf fibre utilization. In: Proceedings of the second technical review meeting on the national kenaf research project, Sungai Petani, Kedah.
- Jayani, R.S., Saxena, S. and Gupta, R. (2005). Microbial pectinolytic enzymes of a review. *Process Biochemistry* 40: 2931-2944.
- Jianqiang, Lin., Sang-mok Lee, Ho-joon Lee, Yoon-mo Koo. 2000. Modeling of typical microbial cell growth in batch culture. *FEMS Microbiology Reviews*, 5, 382-385.
- Kaldor, A.F., Kerlgren, C. and Werwest, H. (1990). Kenaf-a fast growing fiber source for paper making. *Tappi* 73(11): 205 – 209.
- Kamra, D.N., Agarwal, N. and McAllister, T.A. (2010). Screening for compounds enhancing fibre degradation. In *In Vitro Screening of Plant Resources for Extra-*

*Nutritional Attributes in Ruminants: Nuclear and Related Methodologies*, ed. P.E. Vercoe, pp 87 – 105. Dordrecht, Springer Science.

- Kapoor, M., Beg, Q.K., Bhushan, B., Singh, K., Dadhich, K.S. and Hoondal, G.S. (2001). Application of an alkaline and thermostable polygalacturonase from *Bacillus* species MG-cp-2 in degumming of ramie (*Boehmeria nivea*) and sunn hemp (*Crotalaria juncea*) bast fibers. *Process Biochemistry* 36: 803-807.
- Kasyap, D.R., Chandra, S. Kaul, A. and Tewari, R. (2000). Production, purification and characterization of pectinase from *Bacillus* species. DT7. *World Journal of Microbiology Biotechnology* 16: 277 – 282.
- Kashyap, D.R., Vohra, P.K., Chopra, S. and Tewari, R. (2001). Applications of pectinases in the commercial sector. A review. *Bioresources Technology* 77: 215-227.
- Kashyap, D.R., Soni, S.K. and Tewari, R. (2003). Enhanced production of pectinase by *Bacillus* species DT7 using solid state fermentation. *Bioresources Technology* 88 : 251-254.
- Kassim, B. (2001). Water management requirement for kenaf production in Malaysia. In: Proceedings of the first technical review meeting on the national kenaf research project. MARDI, Selangor.
- Kawai, S. (2005). Development of high performance kenaf bast oriented fibreboard and kenaf core binderless particleboard. *Sustainable Humanospere* 1:12.
- Kenneth, T. (2008). *Online Text Book of Bacteriology*, University of Wisconsin, Madison, US.
- Lang, C. and Dornenburg, H. (2000). Perspectives in the biological function and the technological application of polygalacturonases. *Applied Microbiology Biotechnology* 53: 366- 75.
- Laxman, R.S., Sonawane, A.P., Mores, S.V., Rao, B.S., Rele, M.V. and Jogdand, V.V. (2005). Optimization and scale up of production of alkaline protease from *Conidio boluscoronatus*. *Process Biochemistry* 40(9): 3152 -3158.
- Lin, J., Takagi, M., Qu, Y., Gao, P. and Yoshida, T. (1999). Metabolic flux change in hybridoma cells under osmotic pressure. *Journal Bioscience Bioengineering* 87 : 255 – 257.
- Malvessi, E. and Silveira, M.M. (2004), Influence of medium composition and pH on the production of polygalacturonases by *Aspergillus oryzae*. *Brazilian Archives Biology Technology* 47 : 693-702.

- Mandels, M. and Stenberg, D. (1976). Recent advances in cellulase technology. *Journal of Fermentation Technology* 54: 267 – 286.
- Melillo, J.M., Steudler, P.A., Aber, J.D., Newkirk, K., Lux, H., Bowles F.P. (2002). Soil warming and carbon-cycle feedbacks to the climate system. *Science* 298: 2173 – 2176.
- Montgomery, D.C. and Myers, R.H. (1997). Response Surface Methodology: Process and Product Optimized Using Designed Experiments, John and Wiley sons, New York, USA : 427 – 510.
- Morrison, S.J., Carpenter, C.E., Morris, S.A., Cromwell, V.A. and Peterson, E.U. (1999). Chemistry of permeable reactive barrier for uranium containment, Monticello, Utah. *EOS Trans Am Geophysic Union, Fallmeet* 80 : 366.
- Murad, H.A. and Azzaz, H.H. (2011). Microbial pectinases and ruminant nutrition. *Research Journal of Microbiology* 6 (3) : 246-269.
- Myres, R.H. and Montgomert, D.C. (1997). A tutorial on generalized linear models. *Journal of Quality Technology* 29 : 274-291.
- Ohtani, Y., Mazumder, B.B. and Sameshima, K. (2001). Influence of chemical composition of kenaf bast and core on the alkaline pulping response. *Journal of Wood Science* 47(1): 30 – 35.
- Outtrup, H. and Jorgensen, S.T. (2002). The importance of *Bacillus* species in the production of industrial enzymes .In Applications and systems of *Bacillus* and relatives. Edited by R, R .Berkley. Blackwell Science Inc., Malden, Mass : 206– 218.
- Panda, T., Naidu, G.S.N. and Sinha, J. (1999). Multiresponse analysis of microbiological parameters affecting the production of pectinolytic enzymes by *Aspergillus niger* : A statistical review. *Process Biochemistry* 35: 187 – 195.
- Pandey, A., Soccol, C.R., Nigam, P. and Soccol, V.T. (2000). Biotechnological potential of agro-industrial residual I. Sugar cane bagasse. *Bioresources Technology* 74: 69 – 80.
- Pandey, A. (2001). Production of enzymes by solid state fermentation. In: Pandey, A., Soccol, C.R., Rodriguez-Leon, J.A. and Nigam, P. (Editors). Solid state fermentation in biotechnology. Fundamentals and Applications. Asiatech Publisher, New Delhi : 98-100.

- Paridah, M.T., Hafizah, A.W.N., Zaidon, A., Azmi, I., Nor, M.Y.M. and Yuziah, M.Y.N. (2009). Bonding properties and performance of multi layered kenaf board. *Journal of Tropical Forest Science* 21(2): 113 – 122.
- Pastor, F.I.J., Gallardo, O., Aparicio, J.S. and Diaz, P. (2007). Xylanases: molecular properties and applications. In *Industrial Enzymes Structure, Function and Applications*, ed. J. Polaine, and A.P. MacCabe, pp. 65 – 82. The Netherlands: Springer.
- Prasanthi, V., Nikku, M.Y., Vuddaraju, S.P., Nalla, K.K., Raju, C.A.I. and Donthireddy, S.R.R. (2008). Optimization of the fermentation media using statistical approach and artificial neural networks for the production of an alkaline protease from *Bacillus subtilis*. *Journal International of Natural and Engineering Science* 2: 51 – 56.
- Raissi, S. (2009). Developing new processes and optimizing performance using response surface methodology. *World Academic Science Engineering Technology* 49: 1039-1042.
- Ramaswamy, G.N., Craft, S. and Wartelle, L. (1995). Uniformity and softness of kenaf fibers for textile products. *Textile Research Journal* 65 : 765-770.
- Rosfarizan, M. (2000). Kinetics and modeling of kojic acid fermentation by *Aspergillus flavus* using different carbon sources. Ph.D thesis. Universiti Putra Malaysia. pp. 91 – 104.
- Rosfarizan, M., Madihah, S. and Ariff, A.B. (1998). Isolation of a kojic acid producing fungus capable of using starches as a carbon source. *Letters Applied Microbiology* 26: 27-30.
- Roubroeks, J.P., Andersson, R., Mastromauro, D.I., Christensen, B.E. and Aman, P. (2001). Molecular weight, structure and shape of oat (1→3),(1→4)-b-D-glucan fractions obtained by enzymatic degradation with (1→4)-b-D-glucan 4-glucanohydrolase from *Trichoderma reesei*. *Carbohydrate Polymer* 46: 275-285.
- Rowell, R.M., Young, R.A. and Rowell, J.K. (1997). Paper and composites from agro based resources. New York, Lewis.
- Rowell, R.M. and Stout, H.P. (1998). Jute and Kenaf. In *Handbook of Fiber Chemistry*, ed. M. Lewin and E. M. Pearce, pp. 466 – 502. USA, Marcell Dekker Inc.
- Rowell, R.M., Han, j.S. and Rowell, J.S. (2000). Characterization and factors effecting fibre properties. In *Natural Polymer and Agrofibre Composites*, eds, F. Elisabete, L.L. Alcides, and H.C. Matusso, pp. 115 – 134. Emrapa Instrumentacao Agropecuaria, Brasil.



- Sakai, T. (1992). Degradation of pectins. In: *Microbial Degradation of Natural Products*. Winkellmann, G. (Editor). VCH, Weinheim, Germany: 57-81.
- Sakai, T., Sakamoto, T., Hallaert, J. and Vandamme E. (1993) Pectin, pectinase and protopectinase: production, properties and applications. *Advanced Applied Microbiology* 39: 213-94.
- Salazar, L. and Jayasinghe, U. (1999). Fundamental of purification of plant viruses. In *Techniques in Plant, Virology, CIP, Training Manual, J.O. Virus Purification*. International Potato Centre, Peru.
- Samira, S. and Majid, M. (2011). Pretreatment of wool/polyester blended fabrics to enhance titanium dioxide nanoparticles adsorption and self cleaning properties. *Colouration Technology* 127(5): 322 – 327.
- Sathyanarayana, N.G., Manoj, N. and Sunil Kumar, D. (2007). Structural and biochemical properties of pectinases. *Industrial Enzymes*: 99 – 115.
- Schallmey, M., Singh, A. and Ward, O.P. (2004). Development in the use of *Bacillus* species for industrial production. *Journal Microbiology* 50 : 1-17
- Semenova, M., Sinitsyna, O. and Morozova, V.(2006). Use of a preparation from fungal pectin lyase in the food industry. *Applied Biochemistry Microbiology* 42: 598-602.
- Shakhes, J., Dehghani-Firouzabadi, M.R. and Rezayati Charani, P. (2010). Evaluation of harvesting time effects and cultivars of kenaf on papermaking. *Bioresources* 5(2): 1268 – 1280.
- Shamsudin, A. and Mahboob, H. (1998). Breeding for kenaf varieties suitable for round the year cultivation. In: Proceedings of the Workshop and Second Project Coordination Committee Meeting. Bangladesh.
- Shembekar, V.S. and Dhotre, A. (2009). Studies of pectin degrading microorganism from soil. *Journal of Microbial World* 11(2): 216-222.
- Shevchik, V.E. and Hugouvieux-Cotte-Pattat, N. (1997). Identification of a bacterial pectin acetyl esterase in *Erwinia chrysanthemi* 3937. *Molecular Microbiology* 24(6) : 1285-1301.
- Shirwaikar, A., Shirwaikar, A.N., Prabu, S.L. and Kumar, G.A. (2008). Herbal Excipients in Novel Drug Delivery Systems. *Indian Journal Pharmaceutical Sciences* 70:415-22.

- Sjostrom, E. (1993). *Wood Chemistry: Fundamentals and Applications* 2<sup>nd</sup> Edition. San Diego, USA: Academic.
- Sunnotel, O. and Nigam, P. (2002). Pectinolytic activity of bacteria isolated from soil and two fungal strains during submerged fermentation. *World Journal of Microbiology and Biotechnology* 18: 835 – 839.
- Torres, E.F., Supelveda, T.V. and Gonzalez, G.V. (2006). Production of hydrolytic depolymerising pectinases. *Food Technology Biotechnology* 44: 221-227.
- Tari, C., Gogus, N. and Tokatli, F. (2007). Optimization of biomass, pellet size and polygalacturonase production by *Aspergillus sojae* ATCC 20235 using response surface methodology. *Enzyme and Microbiology Technology* 40: 1108 – 1116.
- Terzaghi, B.E. and Sandine, W.E. (1975). Improved medium for lactic streptococci and their bacteriophages. *Applied Microbiology* 29: 807 -813.
- Villar, J.C., Revilla, E., Gomez, N. Carbajo, J.M. and Simon, J. L. (2009). Improving the use of kenaf for kraft pulping by using mixtures of bast and core fibres. *Industrial Crops and Products* 29(2-3): 301 – 307.
- Ward, O.P, and Moo-Young, M. (1989). Enzymatic degradation of cell wall and related plant polysaccharides. *CRC Critical Review Biotechnology* 8: 237-274.
- Webber, C.L. (1992). The effects of kenaf cultivars and harvest dates on plant grown, protein content, and dry matter yields. In: *New Industrial Crops and Products. Proceedings of The First International Conference on New Industrial Crops and Products*, eds. H.H. Naqvi, A. Estila, I.P. Ting, pp. 147 – 152. Riverside, CA. Office of Arid Lands Studies, College of Agriculture, The University of Arizona, Tucson, AZ.
- Webber (III), C.L. and Bledsoe, V.K. (2002). Plant maturity and kenaf yield component. *Industrial Crop and Product* 16: 81 – 88.
- Whitaker, J.R. (1990). Microbial pectinolytic enzymes. In: *Microbial Enzymes and Biotechnology*. Fogarty, W.M. and Kelly, C.T (Editors). 2<sup>nd</sup> Edition. Elsevier Science Ltd, London: 133-176.
- Wong, C.C., Nizaum, M., Najib, M., Vijaysegaran, S. and Yin, M.S. (2001). Effects of defoliation (cutting) on regrowth, forage yield and quality of selected kenaf accessions. In: *Proceedings of the first technical review meeting on the national kenaf research project*.

Yu, H.Q. and Yu, C.W. (2007). Study on microbe retting of kenaf fiber. *Enzyme and Microbial Technology* 40: 180-1809.

Zhang, J. (2006). *Biochemical study and technical applications of fungal pectinases*. PhD Thesis, Uppsala Universitet, Sweden.

Zhang, J., Henriksson, G. and Johansson, G. (2000). Polygalacturonase is the key component in enzymatic retting of flax. *Journal of Biotechnology* 81: 85 – 89.

Zhang, Y.H.P., Himmel, M.E. and Mielenz, J.R. (2006). Outlook for cellulase improvement: Screening and selection strategies. *Biotechnology Advances* 24: 452 – 481.

