

UNIVERSITI PUTRA MALAYSIA

PHYTOCHEMICALS AND BIOACTIVITIES OF Curcuma mangga VALETON AND VAN ZIJP., Boesenbergia prainiana (KING EX BAKER) SCHLTR. AND Bauhinia thonningii SCHUMACH. & THONN.

HALIMATUL SAADIAH BT MOHAMMAD NOOR

FS 2014 36

PHYTOCHEMICALS AND BIOACTIVITIES OF

Curcuma mangga VALETON AND VAN ZIJP., Boesenbergia prainiana (KING EX BAKER) SCHLTR. AND Bauhinia thonningii SCHUMACH. & THONN.

HALIMATUL SAADIAH BT MOHAMMAD NOOR

MASTER OF SCIENCE UNIVERSITI PUTRA MALAYSIA

PHYTOCHEMICALS AND BIOACTIVITIES OF Curcuma mangga VALETON AND VAN ZIJP., Boesenbergia prainiana (KING EX BAKER) SCHLTR. AND Bauhinia thonningii SCHUMACH. & THONN.

By

HALIMATUL SAADIAH BT MOHAMMAD NOOR

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of theRequirements for the Degree of Master of Science

February 2014

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright© Universiti Putra Malaysia

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

PHYTOCHEMICALS AND BIOACTIVITIES OF Curcuma mangga VALETON AND VAN ZIJP., Boesenbergia prainiana (KING EX BAKER) SCHLTR. AND Bauhinia thonningii SCHUMACH. & THONN.

By

HALIMATUL SAADIAH BT MOHAMMAD NOOR

February 2014

Chairman: Mohd Aspollah Bin Hj. Sukari, PhD Faculty: Science

Phytochemical and bioactivity studied on three medicinal plants, *Curcuma mangga* Val. and Van Zijp., *Boesenbergia prainiana* (King ex Baker) Schltr. and *Bauhinia thonningii* Schum. were carried out. *Curcuma mangga* Val. and Van Zijp. and *Boesenbergia prainiana* (King ex Baker) Schltr. belong to the family of Zingiberaceae while *Bauhinia thonningii* Schum. belongs to the family of Cesalpiniaceae. The chemical constituents of these plants were isolated using chromatographic methods while the structure of the compounds were elucidated using various spectroscopics methods including infrared (IR), mass spectrometry (MS) and nuclear magnetic resonance (¹H, ¹³C and 2D NMR). The crude extracts and isolated compounds were screened to biological activity studies which were cytotoxic, antioxidant, total phenolic content, antimicrobial and antifungal.

The isolation work on rhizomes of *Curcuma mangga* have yielded six compounds which were identified as curcumin (4), demethoxycurcumin (5), curcumol (51), curdione (52), zederone (53) and β -sitosterol (22). Curcumol (51), curdione (52) and zederone (53) are reported to be first isolated from this species. For *Boesenbergia prainiana*, phytochemical studies were first reported on isolation of four compounds which were stigmasterol (54), lupeol (55), lupenone (56) and β -sitosterol (22). *Bauhinia thonningii* afforded three compounds elucidated as 9-hydroxytridecyl decosanoate (57), betulinic acid (58) and friedelin (59) which also have never been reported previously on this species.

Crude extracts and isolated compounds were subjected to cytotoxic screening against five human cancerous cell lines; promyelocytic leukemia (HL-60), breast cancer (MCF-7), colonic cancer (HT-29), cervical cancer (HeLa) and T-lymphoblastic (CEM-SS) using MTT assay. Hexane, chloroform, ethyl acetate and methanol extracts of rhizomes of *Curcuma mangga* were active against CEM-SS cell line with IC₅₀ value ranging from 11.35 to 15.41 μ g/mL. This is the first report on cytotoxic activities towards CEM-SS cell line. The hexane and chloroform extracts were also active against MCF-7, HeLa and

HT-29 cell line with IC₅₀ values ranging from 5.75 to 17.9 µg/mL. Isolated compounds, **4** and **5** exhibit promising activities against all selected human cancerous cell. IC₅₀ values of Curcumin (**4**) and demethoxycurcumin (**5**) against HL-60, MCF-7, HeLa and CEM-SS were ranging from 3.14 to 11.04 µg/mL. Furthurmore, curcumol (**51**) was strongly active against MCF-7 and HeLa cell lines with IC₅₀ values of 6.84 and 1.25 µg/mL while curdione (**52**) was active against HT-29 cancer cell line with IC₅₀ of 9.08 µg/mL. In addition, zederone (**53**) was strongly active against MCF-7 cell line with IC₅₀ of 3.43 µg/mL. For *Boesenbergia prainiana*, hexane and chloroform extracts were active against MCF-7 cell line with IC₅₀ of 17.54 and 18.26 µg/mL. As for *Bauhinia thonningii*, hexane, chloroform, ethyl acetate and methanol extracts were strongly active against CEM-SS cell line with IC₅₀ value ranging from 9.21 to 11.12 µg/mL. These cytotoxic properties were also reported for the first time from this species.

For antioxidant assay, chloroform, ethyl acetate and methanol extract of *Curcuma* mangga show strong antioxidant effect with IC_{50} values of 40.58, 30.17 and 24.97 µg/mL, respectively while no antioxidant effect was observe on all crude extracts of *Boesenbergia prainiana*. For *Bauhinia thonningii*, ethyl acetate and methanol extracts show strong antioxidant effect with IC_{50} values of 40.58 and 24.97 µg/mL. Isolated compounds of *Curcuma mangga*, curcumin (4) and demethoxycurcumin (5) showed strong antioxidant activity with IC_{50} values of 26.67 and 31.52 µg/mL respectively. Total phenolic content (TPC) determination indicated chloroform, ethyl acetate and methanol extracts of *Curcuma mangga* have high TPC followed by ethyl acetate and methanol extracts of *Bauhinia thonningii*.

In antimicrobial screening test, methanol extract of *Curcuma mangga* exhibited remarkable antimicrobial activity against Methicillin-resistant *Staphylococcus aureus* (MRSA), *Pseudomonas aeruginosa* and *Bacillus subtillis*. For *Boesenbergia prainiana*, and *Bauhinia thonningii*, its crude extracts indicated low effect against MRSA, *Bacillus subtillis* and *Pseudomonas aeruginosa*. Antimicrobial activity studies have never been reported previously on *Boesenbergia prainiana* and *Bauhinia thonningii*.

Abstrak thesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

FITOKIMIA DAN BIOAKTIVITI Curcuma mangga VALETON DAN VAN ZIJP., Boesenbergia prainiana (KING EX BAKER) SCHLTR. DAN Bauhinia thonnigii SCHUMACH. & THONN.

Oleh

HALIMATUL SAADIAH BINTI MOHAMMAD NOOR

Febuari 2014

Pengerusi: Mohd Aspollah Bin Hj. Sukari, PhD Fakulti: Sains

Tiga jenis tumbuhan yang digunakan untuk tujuan perubatan iaitu *Curcuma mangga* Val. and Van Zijp., *Boesenbergia prainiana* (King ex Baker) Schltr. dan *Bauhinia thonningii* Schum. telah dikaji. *Curcuma mangga* Val. and Van Zijp. dan *Boesenbergia prainiana* (King ex Baker) Schltr. tergolong dalam famili Zingiberaceae manakala *Bauhinia thonningii* Schum. tergolong dalam famili Cesalpiniaceae. Hasil sebatian daripada pokok-pokok ini dipencilkan menggunakan teknik kromatografi. Stereokimia sebatian ini telah ditentukan menggunakan pelbagai teknik spektroskopi termasuk inframerah (IR), spektrometri jisim (MS) dan juga resonan magnet nukleus (¹H, ¹³C dan 2D NMR). Ekstrak mentah dan juga hasil sebatian telah dijalankan ujian sitotoksik, antioksida, jumlah kandungan fenol, dan juga antimikrob dan antifungal.

Kerja pemencilan yang dijalankan ke atas *Curcuma mangga* menghasilkan enam sebatian yang dikenalpasti sebagai curcumin (4), demethoxycurcumin (5), curcumol (51), curdione (52), zederone (53) dan β -Sitosterol (22). Curcumol (51), curdione (52) dan zederone (53) adalah yang pertama berjaya diperolehi daripada spesies ini. Empat sebatian yang pertama dicatatkan diperolehi daripada *Boesenbergia prainiana* iaitu stigmasterol (54), lupeol (55), lupenone (56) dan β -Sitosterol (22). *Bauhinia thonningii* pula berjaya mengasingkan tiga sebatian dikenalpasti sebagai 9-hydroxytridecyl decosanoate (57), betulinic acid (58) dan friedelin (59) yang belum pernah dicatatkan dalam spesies ini.

Semua ekstrak dan sebatian yang diperolehi telah dijalankan ujian sitotoksik terhadap lima jenis sel kanser manusia iaitu sel kanser leukimia (HL-60), sel kanser payudara (MCF-7), sel kanser kolon (HT-29), sel kanser servik (HeLa) dan sel kanser T-lymphoblastic (CEM-SS). Ekstrak heksana, kloroform, etil asetat dan metanol daripada akar *Curcuma mangga* adalah aktif terhadap sel kanser T-limfoblastik (CEM-SS) dengan nilai IC₅₀ dalam julat 11.35 hingga 15.41 µg/mL. Ini adalah catatan aktiviti sitotoksik yang pertama terhadap sel CEM-SS. Ektrak heksana dan kloroform juga aktif terhadap sel MCF-7, HeLa dan HT-29 dengan nilai IC₅₀ antara 5.75 hingga 17.9 µg/mL.

Hasil sebatian (4) dan (5) adalah sangat aktif terhadap semua sel kanser yang dipilih. Nilai IC₅₀ Curcumin (4) dan demethoxycurcumin (5) terhadap HL-60, MCF-7, HeLa dan CEM-SS adalah antara 3.14 to 11.04 µg/mL. Seterusnya, Curcumol (51) adalah sangat aktif terhadap sel MCF-7 dan HeLa dengan nilai IC₅₀ 6.84 dan 1.25 µg/mL. Curdione (52) aktif terhadap sel HT-29 dengan nilai IC₅₀ of 9.08 µg/mL. Tambahan pula, zederone (53) adalah sangat aktif terhadap sel MCF-7 dengan nilai IC₅₀ 3.43. Untuk *Boesenbergia prainiana* pula, ekstrak heksana and kloroform adalah aktif terhadap sel MCF-7 dengan IC₅₀ 17.54 dan 18.26 µg/mL. Untuk *Bauhinia thonningii*, ekstrak heksana, kloroform, etil asetat dan metanol adalah aktif terhadap sel CEM-SS dengan nilai IC₅₀ antara 9.21 hingga 11.12 µg/mL. Catatan aktiviti sitotoksik ini adalah yang pertama terhadap species *Bauhinia thonningii*.

Untuk ujian antioksida, ekstrak kloroform, etil asetat dan metanol daripada *Curcuma* mangga menunjukkan kesan antioksida yang tinggi dengan IC₅₀ 40.58, 30.17 dan 24.97 μ g/mL, manakala tiada kesan antioksida daripada semua ekstrak *Boesenbergia* prainiana. Untuk *Bauhinia thonningii*, ekstrak etil asetat and metanol menunjukkan kesan antioksida yang tinggi dengan IC₅₀ 40.58 dan 24.97 μ g/mL. Sebatian daripada *Curcuma mangga*, curcumin (4) and demethoxycurcumin (5) menunjukkan aktiviti antioksida yang tinggi dengan nilai IC₅₀ 26.67 dan 31.52 μ g/mL. Dalam penentuan jumlah kandungan fenol (TPC), ekstrak kloroform, etil asetat and metanol daripada *Curcuma mangga* mengandungi jumlah kandungan fenol yang tinggi diikuti oleh ekstrak etil asetat dan metanol daripada *Bauhinia thonningii*.

Dalam ujian antimikrob dan antifungal, ekstrak metanol daripada curcuma mangga menunjukkan aktiviti yang memberansangkan terhadap Methicillin-resistant Staphylococcus aureus (MRSA), Pseudomonas aeruginosa and Bacillus subtillis. Bagi Boesenbergia prainiana dan Bauhinia thonningii ekstraknya menunjukkan kesan yang lemah terhadap antimikrob MRSA, Bacillus subtillis dan Pseudomonas aeruginosa. Ujian antimikrob terhadap spesies Boesenbergia prainiana dan Bauhinia thonningii belum pernah dicatatkan sebelum ini.

ACKNOWLEDGEMENTS

First of all, I would like to express my honest appreciation and gratefulness to my project supervisor, Prof. Dr. Mohd Aspollah b. Haji Sukari for his caring and concern on us throughout the course of this project. His guidance, constructive comments, continuous encouragement, advice and suggestion are invaluable and very useful to complete this project. I am also grateful to my co-supervisors, Associate Prof. Dr. Intan Safinar Ismail and Dr. Ahmad Bustamam for their intellectual advices and ideas which help in problem solving during the research.

I also would like to dedicate my special thanks to Zulkhairi Azid, Tang sook Wah, Rahayu Utami, Liyana Ithnin, Noorul Adawiyah Mustahil and Sadikah Ahmad for their guidance, co-operation, and contribution to a good working environment. Thanks for sharing the stress and frustration while completing this project. Very big thank to all the staff of the Department of Chemistry, Faculty of Science in UPM for their guidance and co-operation. Thanks to Universiti Putra Malaysia for awarding the Graduate Research Fellowship for three semesters during my study.

Not forgetting, I would like to express my deep gratitude especially to my lovely and caring family for their continuous support and encouragement. I am indebted too to my friends and colleagues who has influenced my thinking and thus, directly and indirectly, contributing to the completion of this project. This project is hard to accomplish without proper guidance from all of you. Once again, thank you so much for all of the contribution to complete this project.

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Mohd Aspollah Bin Hj. Sukari, PhD

Professor Faculty of Science Universiti Putra Malaysia (Chairman)

Ahmad Bustamam Bin Abdul, PhD

UPM-MAKNA Cancer Research Laboratory Institute of Bioscience Universiti Putra Malaysia (Member)

Intan Safinar Ismail, PhD

Associate Professor Faculty of Science Universiti Putra Malaysia (Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

DECLARATION

Declaration by graduate student

I hereby confirm that:

- x This thesis is my original work;
- x Quotations, illustrations and citations have been duly referenced;
- x This thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- x Intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- x Written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- x There is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: 10 February 2014

Name and Matric No.: Halimatul Saadiah Binti Mohammad Noor (GS27511)

Declaration by Members of Supervisory Committee

This is to confirm that:

- x The research conducted and the writing of this thesis was under our supervision;
- x Supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: Name of Chairman of Supervisory Committee:	UPM	Signature: Name of Member of Supervisory Committee:	
Signature: Name of Member of Supervisory Committee:			

TABLE OF CONTENTS

AR	ѕтраст	,	Page
	STRACT STRAK		II iv
AC	KNOWL	EDGEMENTS	vi
API	PROVAI		Vii
DE	CLARAJ	- FION	ix
LIS	T OF TA	ABLES	xiv
LIS	T OF FI	GURES	XV
LIS	T OF AB	BBREVIATIONS	xix
CH	APTER		
1	INTR	RODUCTION	1
	1.1	Natural Product	1
	1.2	Medicinal Plant	1
	1.3	Zingiberaceae	2
		1.3.1 Curcuma	3
		1.3.2 Curcuma mangga Valeton and Van Zijp	3
	1.4	Boesenbergia	3
		1.4.1 <i>Boesenbergia prainiana</i> (King ex Baker) Schltr.	4
	1.5	Leguminosae	4
		1.5.1 Bauhinia	4
		1.5.2 Bauhinia thoninggi Schum.	4
	1.6	Problem statements	5
	1.7	Objective of research	5
2	LITE	RATURE REVIEW	6
	2.1	Literature Review of <i>Curcuma</i>	6
	2.2	Literature Review of Curcuma mangga	7
	2.3	Literature Review of Boesenbergia	10
	2.4	Literarure Review of Bauhinia	14
	2.5	Literature Review of Bauhinia thonningii	14
3	МАТ	ERIALS AND METHODS	16
	3.1	Plant Materials	16
	3.2	Instruments	16
	3.3	Chromatographic Methods	17
	3.4	Experimental Methods	18
		3.4.1 Extraction	18
		3.4.2 Separation and Purification	19
	3.5	Isolation of Chemical Constituents from Rhizomes of	
		Curcuma mangga	19
		3.5.1 Isolation of Curcumin (4)	20

3.5.1 Isolation of Curcumin (4)

		3.5.2 Isolation of Demethoxycurcumin (5)	21
		3.5.3 Isolation of Curcumol (51)	22
		3.5.4 Isolation of Curdione (52)	22
		3.5.5 Isolation of Zederone (53)	23
		3.5.6 Isolation of β -Sitosterol (22)	24
	3.6	Isolation of Chemical Constituents from Rhizomes of	
		Boesenbergia prainiana	25
		3.6.1 Isolation of Stigmasterol (54)	26
		3.6.2 Isolation of Lupeol (55)	27
		3.6.3 Isolation of Lupenone (56)	28
		3.6.4 Isolation of β -sitosterol (22)	29
	3.7	Isolation of Chemical Constituents from Fruit Pulps of	
		Bauhinia thonningii	29
		3.7.1 Isolation of 9-Hydroxytridecyl Decosanoate (57)	30
		3.7.2 Isolation of Betulinic Acid (58)	31
		3.7.3 Isolation of Friedelin (59).	32
	3.8	Bioassays	32
		3.8.1 Cytotoxic Assay	32
		3.8.2 2,2-Diphenyl-1-picrylhydrazyl (DPPH) Radical	
		Scavenging Assay	33
		3.8.3 Total Phenolic Content	33
		3.8.4 Antimicrobial Assay	34
		3.8.4.1 Preparation of Nutrient Broth Culture	34
		3.8.4.2 Medium and plate Preparations	34
4	RESI	ILTS AND DISCUSSION	35
-	4 1	Extraction and Isolation of Chemical Constituent	55
	1.1	from Rhizomes of Curcuma managa	35
		4.1.1 Characterization of Curcumin (4)	35
		4.1.2 Characterization of Demethoxycurcumin (5)	46
		4.1.3 Characterization of Curcumol (51)	58
		414 Characterization of Curdione (52)	64
		4.1.5 Characterization of Zederone (53)	81
		4.1.6 Characterization of B-Sitosterol (22)	88
	4.2	Extraction and Isolation of Chemical Constituents	00
		from Rhizomes of <i>Boesenbergia prainiana</i>	93
		4.2.1 Characterization of Stigmasterol (54)	93
		4.2.2 Characterization of Lupeol (55)	99
		4.2.3 Characterization of Lupenone (56)	115
		4.2.4 Characterization of β -Sitosterol (22)	129
	4.3	Extraction and Isolation of Chemical Constituents	-
		from Rhizomes of <i>Bauhinia thonningii</i>	129
		4.3.1 Characterization of 9-Hydroxytridecyl Decosanoate (57)	129
		4.3.2 Characterization of Betulinic Acid (58)	138
		4.3.2 Characterization of Friedelin (59)	144
			- • •

4	4 Cytotoxic Screening	149
	4.4.1 Cytotoxic Screening Results of Curcuma mangga,	
	Boesenbergia prainiana and Bauhinia thonningii	149
4	5 2,2-Diphenyl-1-picrylhydrazyl (DPPH) Radical Scavengir	ng Assay 152
	4.5.1 DPPH Radical Scavenging Assay Results of Curci	ита
	mangga, Boesenbergia prainiana and Bauhinia	
	thonningii	152
4	5 Total Phenolic Content	153
	4.6.1 Total Phenolic Content of <i>Curcuma mangga</i> ,	
	Boesenbergia prainiana and Bauhinia thonningii	154
4	7 Antimicrobial and Antifungal Screening	154
	4.7.1 Antimicrobial and Antifungal Screening Result of	Curcuma
	mangga, Boesenbergia prainiana and Bauhinia th	honningii 155
(ONCLUSIONS	157
BLIO	GRAPHY	159

BIBLIOGRAPHY BIODATA OF STUDENT LIST OF PUBLICATIONS

5

Ś

165 166

LIST OF TABLES

Table		
4.1	¹ H NMR (400 MHz) and ¹³ C NMR (100 MHz) Spectral Data of Curcumin (4)	Page 38
4.2	¹ H NMR (400 MHz) and ¹³ C NMR (100 MHz) Spectral Data of Demethoxycurcumin (5)	49
4.3	¹ H NMR (400 MHz) and ¹³ C NMR (100 MHz) Spectral Data of Curcumol (51)	60
4.4	¹ H NMR (400 MHz) and ¹³ C NMR (100 MHz) Spectral Data of Curdione (52)	67
4.5	Two dimensional (2D) NMR Data of Curdione (52)	68
4.6	¹ H NMR (400 MHz) and ¹³ C NMR (100 MHz) Spectral Data of Zederone (53)	83
4.7	¹ H NMR (400 MHz) and ¹³ C NMR (100 MHz) Spectral Data of β-Sitosterol from <i>Curcuma mangga</i> (22)	90
4.8	¹ H NMR (400 MHz) and ¹³ C NMR (100 MHz) Spectral Data of Stigmasterol (54)	96
4.9	¹ H NMR (400 MHz) and ¹³ C NMR (100 MHz) Spectral Data of Lupeol (55)	101
4.10	¹ H NMR (400 MHz) and ¹³ C NMR (100 MHz) Spectral Data of Lupenone (56)	117
4.11	¹ H NMR (400 MHz) and ¹³ C NMR (100 MHz) Spectral Data of 9-Hydroxytridecyl Decosanoate (57)	132
4.12	¹ H NMR (400 MHz) and ¹³ C NMR (100 MHz) Spectral Data of Betulinic Acid (58)	140
4.13	¹ H NMR (400 MHz) and ¹³ C NMR (100 MHz) Spectral Data of Friedelin (59)	146
4.14	Cytotoxic Test Result of Isolates from Plant Samples	151
4.15	DPPH Radical Scavenging Assay Results of Plant Samples	153
4.16	Total Phenolic Content of Plant Samples	154
4.17	Antimicrobial and Antifungal Screening Result of Plant Samples	156

LIST OF FIGURES

Figu	·e	Page
3.5	Isolation Work on Rhizomes of Curcuma mangga	20
3.6	Isolation Work on Rhizomes of Boesenbergia prainiana	25
3.7	Isolation Work on Fruit Pulps of Bauhinia thonningii	30
4.1	IR Spectrum of Curcumin (4)	36
4.2	EI-MS Spectrum of Curcumin (4)	36
4.3	¹ H NMR Spectrum of Curcumin (4)	40
4.4	COSY Spectrum of Curcumin (4)	41
4.5	¹³ C NMR Spectrum of Curcumin (4)	42
4.6	DEPT Spectrum of Curcumin (4)	43
4.7	HMQC Spectrum of Curcumin (4)	44
4.8	HMBC Spectrum of Curcumin (4)	45
4.9	Mass Fragmentation of Curcumin (4)	39
4.10	IR spectrum of Demethoxycurcumin (5)	46
4.11	EI-MS Spectrum of Demethoxycurcumin (5)	47
4.12	¹ H NMR Spectrum of Demethoxycurcumin (5)	51
4.13	¹ H NMR Spectrum of Demethoxycurcumin (5) [Expansion]	52
4.14	COSY Spectrum of Demethoxycurcumin (5)	53
4.15	¹³ C NMR Spectrum of Demethoxycurcumin (5)	54
4.16	DEPT Spectrum of Demethoxycurcumin (5)	55
4.17	HMQC Spectrum of Demethoxycurcumin (5)	56
4.18	HMBC Spectrum of Demethoxycurcumin (5)	57
4.19	HMBC Correlation of Demethoxycurcumin (5)	48
4.20	Mass Fragmentation of Demethoxycurcumin (5)	50
4.21	IR spectrum of Curcumol (51)	58
4.22	EI-MS Spectrum of Curcumol (51)	59
4.23	¹ H NMR Spectrum of Curcumol (51)	61
4.24	¹ H NMR Spectrum of Curcumol (51) [Expansion]	62
4.25	¹³ C NMR Spectrum of Curcumol (51)	63

4.26	IR spectrum of Curdione (52)	64
4.27	EI-MS Spectrum of Curdione (52)	65
4.28	¹ H NMR Spectrum of Curdione (52)	69
4.29	¹ H NMR Spectrum of Curdione (52) [Expansion a]	70
4.30	¹ H NMR Spectrum of Curdione (52) [Expansion b]	71
4.31	COSY Spectrum of Curdione (52)	72
4.32	COSY Spectrum of Curdione (52) [Expansion]	73
4.33	¹³ C NMR Spectrum of Curdione (52)	74
4.34	DEPT Spectrum of Curdione (52)	75
4.35	HMQC Spectrum of Curdione (52)	76
4.36	HMQC Spectrum of Curdione (52) [Expansion]	77
4.37	HMBC Spectrum of Curdione (52)	78
4.38	HMBC Spectrum of Curdione (52) [Expansion a]	79
4.39	HMBC Spectrum of Curdione (52) [Expansion b]	80
4.40	IR spectrum of Zederone (53)	81
4.41	EI-MS Spectrum of Zederone (53)	82
4.42	¹ H NMR Spectrum of Zederone (53)	85
4.43	¹ H NMR Spectrum of Zederone (53) [Expansion]	86
4.44	¹³ C NMR Spectrum of Zederone (53)	87
4.45	Mass Fragmentation of Zederone (53)	84
4.46	IR spectrum of β-Sitosterol (22)	88
4.47	EI-MS Spectrum of β-Sitosterol (22)	89
4.48	¹ H NMR Spectrum of β -Sitosterol (22)	91
4.49	¹³ C NMR Spectrum of β -Sitosterol (22)	92
4.50	IR spectrum of Stigmasterol (54)	94
4.51	EI-MS Spectrum of Stigmasterol (54)	94
4.52	¹ H NMR Spectrum of Stigmasterol (54)	97
4.53	¹³ C NMR Spectrum of Stigmasterol (54)	98
4.54	IR Spectrum of Lupeol (55)	99
4.55	EI-MS Spectrum of Lupeol (55)	100

4.56	¹ H NMR Spectrum of Lupeol (55)	102
4.57	¹ H NMR Spectrum of Lupeol (55) [Expansion a]	103
4.58	¹ H NMR Spectrum of Lupeol (55) [Expansion b]	104
4.59	¹³ C NMR Spectrum of Lupeol (55)	105
4.60	¹³ C NMR Spectrum of Lupeol (55) [Expansion a]	106
4.61	¹³ C NMR Spectrum of Lupeol (55) [Expansion b]	107
4.62	DEPT Spectrum of Lupeol (55)	108
4.63	DEPT Spectrum of Lupeol (55) [Expansion a]	109
4.64	DEPT Spectrum of Lupeol (55) [Expansion b]	110
4.65	HMQC Spectrum of Lupeol (55)	111
4.66	HMQC Spectrum of Lupeol (55) [Expansion]	112
4.67	HMBC Spectrum of Lupeol (55)	113
4.68	HMBC Spectrum of Lupeol (55) [Expansion]	114
4.69	IR Spectrum of Lupenone (56)	115
4.70	EI-MS Spectrum of Lupenone (56)	116
4.71	¹ H NMR Spectrum of Lupenone (56)	118
4.72	¹ H NMR Spectrum of Lupenone (56) [Expansion]	119
4.73	¹³ C NMR Spectrum of Lupenone (56)	120
4.74	¹³ C NMR Spectrum of Lupenone (56) [Expansion a]	121
4.75	¹³ C NMR Spectrum of Lupenone (56) [Expansion b]	122
4.76	DEPT Spectrum of Lupenone (56)	123
4.77	DEPT Spectrum of Lupenone (56) [Expansion]	124
4.78	HMQC Spectrum of Lupenone (56)	125
4.79	HMQC Spectrum of Lupenone (56) [Expansion a]	126
4.80	HMQC Spectrum of Lupenone (56) [Expansion b]	127
4.81	HMBC Spectrum of Lupenone (56)	128
4.82	IR spectrum of 9-Hydroxytridecyl Decosanoate (57)	130
4.83	EI-MS Spectrum of 9-Hydroxytridecyl Decosanoate (57)	130
4.84	¹ H NMR Spectrum of 9-Hydroxytridecyl Decosanoate (57)	133

4.85	¹ H NMR Spectrum of 9-Hydroxytridecyl Decosanoate (57) [Expansion]	134
4.86	¹³ C NMR Spectrum of 9-Hydroxytridecyl Decosanoate (57)	135
4.87	¹³ C NMR Spectrum of 9-Hydroxytridecyl Decosanoate (57) [Expansion a]	136
4.88	¹³ C NMR Spectrum of 9-Hydroxytridecyl Decosanoate (57) [Expansion b]	137
4.89	IR spectrum of Betulinic Acid (58)	138
4.90	EI-MS Spectrum of Betulinic Acid (58)	139
4.91	¹ H NMR Spectrum of Betulinic Acid (58)	141
4.92	¹³ C NMR Spectrum of Betulinic Acid (58)	142
4.93	DEPT Spectrum of Betulinic Acid (58)	143
4.94	IR spectrum of Friedelin (59)	144
4.95	EI-MS Spectrum of Friedelin (59)	145
4.96	¹ H NMR Spectrum of Friedelin (59)	147
4.97	¹³ C NMR Spectrum of Friedelin (59)	148

C

LIST OF ABBREVIATIONS

α	Alpha
β	Beta
δ	Chemical shift in ppm
^{13}C	Carbon-13
CHCl ₃	Chloroform
COSY	Correlation Spectroscopy
cm	Centimeter
I	Coupling constant in Hertz
°C	Degree in Celcius
	Deuterated Chloroform
CD_2OD	Deuterated Methanol
d	Doublet
u DEPT	Distortionless Enhancement by Polarization Transfer
DMSO	Distortionless Emancement by Foranzation Hanster
FIMS	Electron Emission Mass Spectroscopy
$EtO\Delta c$	Ethyl Acetate
	Gamma
r CC	Gas Chromatography
GC MS	Cas Chromatography Mass Spectrometry
0C-1015	Gram
g UMBC	Hotoropuoloor Multiple Bond Connectivity
HMOC	Heteronuclear Multiple Dond Connectivity
	Heteronuclear Multiple Quantum Correlation
	Hellz
С	Hydroxy
	Infinition Concentration
IK V a	lilla- Red Kilogram
Kg	Kilogram
	Literature
m/z	Mass per charge
MS M-OU	Mass spectrum/spectra/spectrometer/spectrometry
MeOH	Methanol
OCH ₃	Methoxy
m.p.	Melting point
mL	Millilitre
mg	Miligram
μg	Microgram
M	Molecular ion
m	Multiplet
nm	Nanometer
NMR	Nuclear Magnetic Resonance
ppm	Parts per million
Ή ₩	Proton
KBr	Potassium Bromide
q	Quartet

C

S	Singlet
TLC	Thin Layer Chromatography
t	Triplet
UV	Ultra Violet
WHO	World Health Organization



CHAPTER ONE

INTRODUCTION

1.1 Natural Products

Many important basic discoveries in organic chemistry were carried out in the field of organic chemistry research. Natural products refer to those organic compounds, which are found in nature and are associated with living organisms. Most natural product used for studies have usually been obtained from plants and microorganisms since the practical difficulties in extracting them from animal are much greater (Paul, 1992). Organic natural products are constructed of carbon, hydrogen and oxygen atoms; frequently nitrogen atoms are also involved, and less frequently sulphur, phosphorous, chlorine, bromine, and iodine atoms (Tedder *et al.*, 1972). These compounds are divided into two classes; primary and secondary metabolites. Natural products researchers are more interested in secondary metabolite compounds that are formed from the process which does not show the cell functions clearly such as alkaloids, steroids, terpenoids, phenolic compounds, glycosides and others. This is due to the existence of many outstanding compounds from secondary metabolites which display interesting biological activities.

Natural product chemistry has lately undergone explosive growth in isolation techniques, synthetic methods, physic-chemical measurements, and new concept. On the other hand, it is precisely the chemistry of the natural product which has fostered many of the new developments because of the variety of compound available (Nakanishi *et al.*, 1974).

In recent years, modern strategies have been employed in which bioassay-guided (mainly *in vitro*) isolation and identification of active "lead" compounds from natural sources was employed, besides production of natural products libraries, production of active compounds in cell or tissue cultures, genetic manipulation, natural combinatorial chemistry and others. Natural product research is now more focused on bioactivity and the concepts of dereplication, chemical fingerprinting, and metabolomics have been introduced. In addition, selection of organisms also included those randomly selected (Sarker *et al.*, 2006). In general, the plant extract contains low concentration of active compounds and a large number of compounds, requiring the use of sensitive bioassay suitable for the wide chemical variety and small amount of the tested samples. Test must be simple, reproducible, fast and cheap (Souza and krish, 1996).

1.2 Medicinal plant

The study of natural products continuous to be a major force in the development of the field of organic chemistry and medicinal chemistry. Higher plants used in traditional medicine provided some of the first prototype drugs used clinically in the treatment of a

wide variety of diseases. The need to purify natural products from complex mixtures and to determine the structures has driven the development of more sophisticated methods for separation of compound and for their structural analysis by chemical and subsequently, spectroscopy means. Medicinal plants provide a cost-effective means of primary health care to millions of people around the world. The demand for medicinal plants is steadily increasing in both developing and developed countries due to the growing recognition of drugs based on natural product, food supplements and flavours. Being non-narcotic, having less side effects and easy availability at affordable prices makes these products sometimes the only source of health care available to the poor (Ramawat and Merillon, 2008).

Plant drugs (also called phytomedicines of phytopharmaceuticals) are plant-derived medicines that contain a chemical compound or more usually mixtures of chemical compound that act individually or in combination on the human body to prevent disorders and to restore or maintain health. Chemical entities are pure chemical compounds (isolated from natural sources such as plants, or produced by chemical synthesis) that are use for medicinal purposes (usually with a clearly defined and tested mode of action).

1.3 Zingiberaceae

Zingiberaceae is among the plant families which are widely distributed throughout the tropics particularly in Southeast Asia. Zingiberaceae is one of the largest plant families from the order Zingiberales, with approximately 50 genera and over 1,000 species. In Peninsular Malaysia, the Zingiberaceae are a component of the herbaceous ground flora of the rainforest. It is estimated that there are 150 species of ginger belonging to 23 genera found in Peninsular Malaysia. Zingiberaceae species grow naturally in damp, shaded parts of the low-land or on hill slopes, as scattered plants of thickets. Most members of the family are easily recognized by the characteristics aromatic leaves and fleshy rhizome when both of them are crushed and also by elliptic to elliptic-oblong leaves arranged in two ranks on the leaf-shoot (Holtum, 1950).

In the Southeast Asia region, several species of zingiberaceae are used as spices, medicines, flavouring agents and as source of certain dyes. Several species from the genera *Boesenbergia*, *Curcuma*, *Alpinia*, *Amomum*, *Costus*, *Kaempferia* and *Zingiber* are major ingredients in traditionally prepared tonics, locally called 'jamu' which are commercially available. Various ginger rhizobia provide health-promoting effects and have been utilized to treat certain illnesses such as nausea, motion sickness, stomachic, asthma, diarrhea, digestive, disorder, vomiting, rheumatism, swelling, common cold, cough and other disorder (Habsah *et al.*, 2000).

1.3.1 Curcuma

The genus *Curcuma* (Figure 1.1) belongs to the tribe Zingibereae and consists of about 80 species of rhizomatous herbs. It is native to the warm and humid environments. It has widespread adaptation from the sea level to altitude as high as 2000 m in the Western Ghats and Himalayas. Species such as *C. longa*, *C. angustifolia*, *C. neilgberrensis*, *C. kudagensis*, *C. pseudomontana* and *C. coriacea* are confined to hills at 1000-2500 metre altitude. It is considered to have originated in the Indo-Malayan Region and widely distributed in the tropics of Asia to Africa and Australia. About 40 out of the 100 or so species reported in the Genus are of Indian origin (Sasikumar, 2005).

1.3.2 Curcuma mangga Valeton and Van Zijp

Curcuma mangga is locally known as "mango ginger" or "manggo turmeric" while in Indonesia, it is recognized as *těmu mangga* (general), *těmu poh, těmu banjangan, těmu lalab, konèng lalab* (Sundanese), *těmu pauh* (Java), *temo pao* (Madurese), *konèng joho* and *konèng parè* (Burkill, 1966; Abas *et al.*, 2005 and Bos *et al.*, 2007). This plant commonly grown in Thailand, Peninsular Malaysia, Bengal, North, Eastern India and Java. The rhizomes are very similar to ginger but have a raw mango smell when the fresh rhizomes are cut. So it becomes popular vegetable which the tips of young rhizomes and shoots are consumed raw with rice and also used in making pickles in south India (Abas *et al.*, 2005 and Liu and Nair, 2010). Medicinally, the rhizomes are used as a stomachic and for chest pains, fever, gastric ulcer and general debility. It is also used in postpartum care, specifically to aid womb healing (Abas *et al.*, 2005 and Ruangsang *et al.*, 2010).

1.4 Boesenbergia

Boesenbergia belongs to ginger family, Zingiberaceae in the order of Zingiberales. The genus of about 80 species is distributed from India to South East Asia. Borneo, is one of the two distribution centres apart from Thailand, which is estimated to have 25 species (Larsen *et al.*, 2003). *Boesenbergia* species is extremely rare compare to other genera. Mostly, they are found in very damp, shaded areas and are usually close to streams or in boggy conditions. Thus, there is an urgency to document the plants before facing extinction. Many researchers have shown that the rhizome of *Boesenbergia* displayed health-benefits properties. For instance, the rhizome of *B. rotunda* is generally used as a culinary spice in Thailand and also has been used for the treatment of oral diseases (that is dry mouth), stomach discomfort, stomach pain, leucorrhoea, diuretic, dysentery, and inflammation. The rhizomes are used in traditional medicine as antiseptic and for the treatment of stomach ache (Hasnah *et al.*, 1995), diarrhea, dermatitis, dry cough and mouth ulcers (Burkill, 1935).

1.4.1 Boesenbergia prainiana (King ex Baker) Schltr.

Boesenbergia prainiana (Figure 1.2) is grown in peninsular Malaysia and it is distributed in the lowland forest in the states of Terengganu, Perak, Pahang and Johor. It is a few-leaved herb up to 30 cm tall and have inflorescence terminal on leafy shoots. It flowers from the top, flower parts are delicate and short-lived.

1.5 Leguminosae

Many plants from the Leguminosae family are medicinal herbs which are easily found in Malaysia. There are mostly tropical and subtropical trees and shrubs comprising about 150 genera and 2,200 species. The leaves are alternate but may be bipinnate or simple. They are grown as weeds (i.e. *Mimosa pudica*), woody shrubs (*Peltophorum pterocarpum*), crops (*Arachis hypogaea*) and vines (*Bauhinia kockiana*). Some of these plants are edible, and hence they are utilised for various purposes in food, beverages, and food colouring agent (Goh, 2004 and Ong, 2006). These plants are commonly used as traditional medicines to treat various health complications

1.5.1 Bauhinia

The genus *Bauhinia* belongs to the Leguminosae family, sub-family Cesalpiniaceae and comprises about 300 species distributed in tropical and subtropical regions. *B. kockiana*, a tropical vine is cultivated as a garden ornamental plant because of its bright orange–red magnificent inflorescences. This plant originates from the Malaysia tropical forest and its roots are used by the Kelabit ethnic group in Sarawak to treat gonorrhoea (Fasihuddin *et al.*, 1995). Besides, the infusion of the roots is consumed orally to treat nervous debility, insomnia and fatigue. The bark and root are also used traditionally to treat toothache (Ong, 2006).

1.5.2 Bauhinia thonningii Schum.

Bauhinia thonningii (Figure 1.3) is a savanna deciduous plant which has a wide distribution range in tropical Africa and extending from West Africa to the Sudan and south wards to east and central southern Africa, including countries such as Mozambique, Malawi, Zimbabwe, Zambia, Botswana, Tanzania and Namibia. *B. thonningii* trees typically grow to a height of 6 to 12 m and their branches spread 3 to 6 m outwards. The flowers are usually five-petaled and are 7 to 12 cm in diameter, generally in shades of red, pink, purple, orange or yellow colour. The species is widely used in sub-Saharan Africa for poles, firewood, charcoal and its pods are eaten by wild animals. The plant is also used to make ropes, dyes and gums and are used in ethnomedicine (Chidumayo, 2007 and Chidumayo, 2008).

1.6 Problem Statements

Plants with ethobotanical or ethnopharmaceutical activities are acceptable as healthygiving supplements. Plants have a long history of use in the treatment of cancer but many claims for the efficacy of such treatment. These have prompted us to carry out investigation of the three medicinal plant species which are *Curcuma mangga* Valeton and Van Zijp., Boesenbergia prainiana (King ex Baker) Schltr. and Bauhinia thonnigii Schumach. & Thonn. These plants have long been utilized as traditional medicine either consume internally or apply externally and also consumed as food or vegetable.So further investigation been carry out in term of phytochemistry and bioactivity studies. For *curcuma mangga*, there were not much previous research on the chemical constituents and biological activity studies while for *Boesenbergia prainiana* and *Bauhinia thonningii* there were no phytochemical and biological activity studies previously.

1.7 Objectives of Research

Objectives of the research are:

- 1. To extract and isolate the chemical constituents of *Curcuma mangga* Valeton and Van Zijp., *Boesenbergia prainiana* (King ex Baker) Schltr. and *Bauhinia thonnigii* Schumach. & Thonn.
- 2. To elucidate the structures of isolated compounds using various spectroscopic methods such as IR, MS and NMR.
- 3. To screen the bioactivities of the crude extracts and isolated compounds through various bioactivy studies such as cytotoxic, antioxidant, total phenolic content, antimicrobial and antifungal.



BIBLIOGRAPHY

- Abas, F., Lajis, N., Shaari, K., Israf, D. A., Stanslass, J., Yusof, U.K. and Raor, S. (2005). A Labdane Diterpene Glucoside from the Rhizomes of *Curcuma mangga*. *Journal of Natural Products*, 68: 1090-1093.
- Achenbach, H., Stocker, M. and Constenla, M. A. (1988). Flavonoid and others Constituents of *Bauhinia manca*. *Phytochemistry*, 27: 1835-184.
- Ali, M. S., Mahmud, S., Perueen, S., Ahmad, V. V. and Rizwani, G. H. (1999). Epimers from The Leaves of *Calophyllum inophyllum*, *Phytochemistry*, 50(8): 1385-1389.
- Asakawa, Y., Takahashi, H., and Toyota, M. (1991). Biotransformation of Germacrane Type Sesquiterpenoids By *Aspergillus Niger*. *Phytochemistry* 30: 3993-3997.
- Atta-ur Rahman. 1992. Nuclear Magnetic Resonance: Basic Principle. New York.
- Bos, R., Rein, J., Windono, L., Tri, G., Woerdenbag, W., Herman, J., Boersma, Y., Kelein, L., Koulman, H., Albert, W., Kayser, G., Oliver, D. (2007). HPLCphotodiode Array Detection Analysis of Curcuminoids in *Curcuma* Species Indigenous to Indonesia. *Phytochemical Analysis* 18(2): 118-122.
- Burkill., I.H. (1935). A dictionary of the Economic Product of the Malay Peninsula. *The Malaya Nature Society*. Vol 1, Kuala Lumpur.
- Burkill, I.H. (1966). A Dictionary of the Economic Product of the Malay Peninsula, Vol. A-H & I-Z, Kuala Lumpur: The Ministry of Agriculture & Cooperatives.
- Butler, M. S. (2008). Natural Products to drugs: Natural Product-Derived Compounds in Clinical Trials. *Natural Product Reports*. 25: 475-516.
- Cheenpracha, S., Karalai, C., Ponglimanont, C., Subhadhirasakul, S. and Tewtrakul, S. (2006). Anti-HIV-1 Protease Activity of Compounds from *Boesenbergia* pandurata. *Bioorganic and Medical Chemistry*. 16: 1710-1714.
- Chew, Y. L., Lim, Y. Y., Omar, M. and Khoo, K. S. (2008). Antioxidant Activity of Three Edible Seaweeds from Two Areas in South East Asia. *Food Science and Technology*, 41: 1067-1072.
- Chidumayo, E. N. (2007). Growth Responses of an African Savanna Tree *Bauhinia thonningii* to Defoliation, Fire and Climate Trees, 21: 231-238.
- Chidumayo, E.N. (2008). Growth of *Bauhinia thonningii* Trees and Saplings Over a Decade in Savanna in Zambia: Interaction of Climate, Fire and Source of

Regeneration. J. Trop. Ecol. The Malaysian Journal of Analytical Sciences. 11: 154-159.

- Ching, A. Y. L., Wah, T. S., Sukari, M. A., Lian, G. E. C., Rahmani, M. and Khalid, K. (2007). Characterization of Flavanoid Derivatives from *Boesenbergia rotunda* (L.). *The Malaysian Journal of Analytical Sciences*, 11(1): 154-159.
- Duarte-Almeida, J. M., Negri, G. and Salatino, A. (2004) Volatile oils in Leaves of *Bauhinia* (Fabaceae Caesalpinioideae). *Biochem. Syst. Ecol.* 32: 947–753.
- Fasuhuddin, B. A., Ipor, I. B. and Din, L. B. (1995). Medical plants used by the Kelabit community in Bario, Sarawak. *Chemical prospecting in the malaysian forest*, 43-46.
- Goh, K. L. (2004), Malaysian herbs, Klang.
- Gowela, J. P. (2003). Status of Medicinal Trees Used in Maternal and Child Health in Malawi: A Case of Dzalanyama Forest Reserve. MSc thesis. University of Malawi, Bunda College of Agriculture, 116.
- Habsah, M., Amran, M., Mackeen, M. M., Lajis, N. H., Kikuzaki, H., Nakatani, N., Rahman, A. A. and Ali, A.M. (2000). Screening of Zingiberaceae Extracts for Antimicrobia and Antioxidant Activities. *Journal of Ethnopharmacology*. 72: 403-410.
- Hasnah, M. S., Shajarahtunnur, J., Neelavany, M. (1995). Chemical Constituents of *Boesenbergia* species. Chemical prospecting in the Malaysian Forest.
- Heyne, K. (1987). Tumbuhan Berguna Indonesia. Jilid IV. Jakarta: Badan Litbang Kehutanan, 592-594.
- Holland, H. L., Diakow, P. R. P. and Taylor, G. J. (1976). Microbial hydroxylation of Steroids. *Can J. Chem.*, 56: 3121.
- Holtum, R. E. (1950). The Zingiberaceae of the Malay Peninsula. *Gardens Bulletin of Singapore*, 13: 1-249.
- Huang, Y. T., Hwang, J. J., Lee, P. P., and Ke F. C. (1999). Effects of Luteolin and Quercetin, Inhibitors of Tyrosine Kinase, on Growth and Metastasis-associated Properties in A431 Cells Over Expressing Epidermal Growth Factor Receptor. *Br. J. Pharmacol.* 128: 999-1010.
- Jamal, A. K., Yaacob, W. A. and Din, L. B. (2008). A Chemical Study on *Phyllanthus reticulates*. *Journal of Physical Science*, 19(2): 45-50

- Jayaprakasha, G. K., Rao, L. J. M. and Sakiriah, K. K. (2002). Improved HPLC Method for Determination of Curcumin, Demethoxycurcumin and Bisdemethoxycurcumin. *Journal of Agricultural and Food Chemistry*. 50(13): 3668-3672.
- Jimoh, F.O. and Oladiji A.T. (2005). Preliminary studies on *Piliostigma thonningii* seeds: Preliminary Analysis, Mineral Composition and Phytochemical Screening. *African Journal of Biotechnology*. 4: 1439-1442.
- Jing, L.L., Mohamed, M., Rahmat, A. and Abu Bakar, M. F. (2010). Phytochemicals, antioxidant properties and anticancer investigations of the different parts of several gingers species (*Boesenbergia rotunda*, *Boesenbergia pulchella var attenuata* and *Boesenbergia armeniaca*). Journal of Medicinal Plants Research, 4(1): 27-32.
- Kaewkroek, K., Wattanapiromsakul, C. and Tewtrakul, S. (2009). Nitric Oxide Inhibitory Substancew from *Curcuma mangga* Rhizomes. *Songklanakarin Journal* of Science and Technology, 31(3): 293-297.
- Khine, M. M. (2006). Isolation and Characterization of Phytoconstituents from Myanmar Medicinal Plants. *Electronisches Document*, 15: 25-77.
- Kim, D. S. H. L., Chen, Z., Nguyen, V.T., Pezzuta, J. M., Qiu, S. and Lu, Z.Z. (1997). A Concise Semi-synthetic Approach to Betulinic Acid from Betulin. Synthetic Communications: An International Journal for Rapid Communication of Synthetic Organic Chemistry. 27(9): 1607-1612.
- Kirana, C., Record, I. R., McIntosh, G. H. and Jones, G. P. (2003). Screening for Antitumor Activity of 11 Species of Indonesian Zingiberaceae Using Human MCF-7 and HT-29 Cancer cells, *Pharmaceutical Biology*, 41(4): 271-276.
- Kirana, C., Jones, G. P., Record, I. R. and McIntosh, G.H. (2007). Anticancer Properties of Panduratin A Isolated from *Boesenbergia pandurata* (Zingiberaceae). *Journal of Natural Medicine*, 61: 131-137.
- Klass, J., Tinto, W. F., McLean, S. and Reynolds, W. F. (1992). Friedelin Triterpenoids from *Peritassa Compta*: Complete ¹H and ¹³C Assignments by 2D NMR Spectroscopy. *Journal of Natural Products*, 55(11): 1626-1630.
- Kojima, H., Sato, N., Hatano, A., Ogura, H. (1990). Sterol Glucosides from *Prunella* vulgaris. *Phytochemistry*, 29(7):2351-2355.
- Kosuge, T., Yokota, M., Sugiyama, K., Yamamoto, T., Ni, M. and Yan, S. (1985). Studies on Antitumor Activities and Antitumor Principles of Chinese herbs. Yakugaku Zasshi, 105(8): 791-795.

- Ong, H.C. (2006). *Tanaman Hiasan: Khasiat makanan dan Ubatan*. Selangor: Utusan Publications and Distributors Sdn. Bhd.
- Paul, S. 1992. Guide to Medical Plant.Ferdinan, Phytochemistry, 2696-2703.
- Pandji, C., Grimm, C., Wrays, V. and Prokschs, P. (1993). Insectidal Constituents Four Species of the Zingiberaceae. *Phytochemistry*, 34(2): 415-419.
- Phillip, K., Abdul Malek, S. N., Sani, W., Sim, K. S., Kumar, S., Hong, S. L., Lee, G. S. and Rahman, S. N. S. A. (2009). Antimicrobial Activity of Some Medicinal Plants from Malaysia. *American Journal of Applied Sciences*, 6(8): 1613-1617.
- Prachayasittikul, S., Saraban, P., Cherdtrakulkiat, R., Ruchirawat S. and Prachayasittikul, V. (2010). New Bioactive Triterpeniods and Antimalarial activity of Diospyros Rubra Lec. *EXCLI Journal*, 9: 1-10.
- Prior, R. L., Wu, X., Schaich, K. (2005). Standardized Methods for Detemination of Antioxidant Capacity and Phenolics in Foods and Dietary Agric. Food Chem, 53: 4290-4303
- Ramawat, K. and Merillon, S. (2008). Secondary metabolite production fromplant cell cultures : The Success Stories of Rosmarininc Acid. Plant Cell Tiss. Org. Cult. 16: 85-100.
- Ruangsang, P., Tewtrakul, S. and Reanmongkol, W. (2010). Evaluation of the Analgesic and Anti-inflammatory Activities of *Curcuma mangga* Val. And Van Zijp. Rhizomes. *Journal of Natural Medicine*. 64(1): 36-41.
- Sarkel, S. D., Latif, Z. and Gray, A. I. (2006). Natural Product Isolation- An Overview. In Sarker, S. D., Latif, Z. and Gray, A. I. *Natural Products Isolation* (pp.1-3). 2nd Edition. New Jersey: Humana Press Inc.
- Sartorelli, P. and Correa, D. S. (2007). Constituents of Essential Oil from *Bauhinia* forficata Link . J. Essent. Oil Res. 19: 468–469.
- Sasikumar, B. (2005).Genetic Resources of *Curcuma*: Diversity, Characterization and Utilization. *Plant Genetic Resources*, 3(2): 230-251.
- Schwartz, J.J. and Wall, L. E. (1995). Isolation of the Sterols of the White Potato. *Journal of America Chemistry Society*, 77: 5442-5443.
- Sauza, B., Krish, J. (1996). How to Study the Pharmacology of Medical Plants in Underdeveloped Countries. *Journal of Ethno pharmacology* 54: 131-138.

- Tapondjou, A.L., Miyamoto, T. and Lacaille-Dubois, M.A. (2006). Glucuronide triterpene Saponins from *Bersama engleriana*. *Phytochemistry*. 67(19): 2126-2132.
- Tedder, J. M., Nechvatal, A., Murray, A. W. and Carnduff, J. (1972). Basic Organic Chemistry-Part 4: Natural Products, Chichester:John Wiley & Sons Ltd.
- Tewtrakul, S., Subhadhirasakul, S., Puripattanavong, J. and Panphadung, T. (2003). HIV-1 Protease inhibitory substances from *Boesenbergia pandurata* Holtt. *Songklanakarin Journal of Science and Technology*. 25(4): 503-508.
- Viana, E. P., Santa-Rosa, R. S., Almeida, S. S. M. S. and Santos, L. S. (1999). Constituents of the Stem Bark of *Bauhinia guianensis*. *Fitoterapia*. 70: 111–112
- Van Wyk, B. E. and Wink, M. (2004). Medicinal plants of the World. Singapore: Times Editions-Marshall Cerendish.
- Wei, Y.Q., Zhao, Y. Kariya, H. Fukata, K. Teshigawara. and Uchida, A. (1994). Induction of apoptosis by quercetin: Involvement of heat shock protein. *Cancer Res.* 54: 4952-4957.
- Yang, F.Q., Li, S. P., Chen, Y., Lao, S. C., Wang, Y. T., Dong, T. T. X., Tsim, K. W. K. (2005). Identification and Quantitation of Eleven Sesquiterpenes in Three Species of *Curcuma* Rhizomes by Pressurized Liquid Extraction and Gas Chromatographymass Spectrometry. *Journal of Pharmaceutical and Biomedical Analysis*. 39: 552-558
- Zhang, X., Geoffray, P., Miesch, M., Julien-David, D., Raul, F., Aoude-Werner, D. and Marchioni, E.2005. Gram-Scale Chromatographic Purification of β-sitosterol Synthesis and characterization of β-sitosterol Oxides. *Steroids*. 70: 886-895