

# **UNIVERSITI PUTRA MALAYSIA**

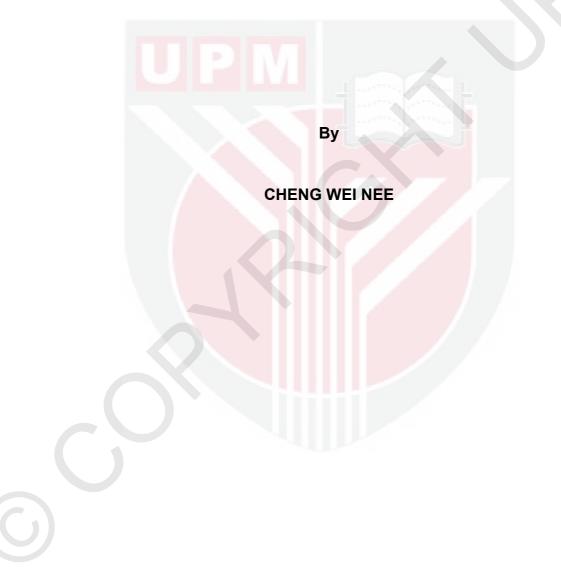
DECOLOURIZATION OF SELECTED TEXTILE DYES BY WHITE ROT FUNGUS CORIOLOPSIS SP. STRAIN AFF17

**CHENG WEI NEE** 

FBSB 2014 34



# DECOLOURIZATION OF SELECTED TEXTILE DYES BY WHITE ROT FUNGUS CORIOLOPSIS SP. STRAIN AFF17



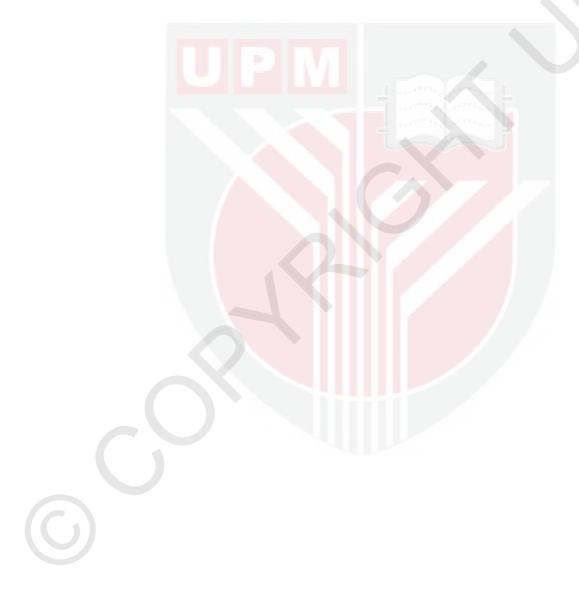
Thesis submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

Oct 2014

# COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artworks, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

## DECOLOURIZATION OF SELECTED TEXTILE DYES BY WHITE ROT FUNGUS CORIOLOPSIS SP. STRAIN AFF17

By

#### **CHENG WEI NEE**

#### October 2014

## Chairman : Professor Mohd. Arif Syed, PhD Faculty : Biotechnology and Biomolecular Sciences

Synthetic dyes are replacing natural dyes in modern textile industries. However, each year, about 10% of dyestuffs wastes enter to the environment and causes water pollution. As synthetic dyes are designed to resist fading therefore they are recalcitrant in the environment. Since physicochemical treatments have limited efficiency and disadvantages, bioremediation of dyestuffs wastes with microorganisms is gaining scientific interest as an alternative pollution treatment. A previously locally isolated white rot fungus, Coriolopsis sp. Strain aff17 (EU863193) from UPM, Serdang was screened for its ability to decolourize azo dyes. It was able to decolourize eleven out of thirteen azo dyes that were chosen. Only seven azo dyes were used in the studies namely Ponceau 2R (C.I. 20470), Amaranth (C.I. 16185), Orange G (C.I. 16230), Naphthol Blue Black (C.I. 20470), Remazol Black B (C.I. 20505), Trypan Blue (C.I. 23850) and Sirius Light Blue BRR (C.I. 34140). Charcaterizations of decolourization ability by Coriolopsis sp. Strain aff17 were investigated. Decolourization rates were shown to be higher in shake cultures as compared to static cultures. The best agitating speed for decolourization of dyes was found to be 75 rpm. The best medium for the decolourization of dyes was distilled water. The presence of chromium, arsenic, cadmium and lead did not inhibit the decolourization of by Coriolopsis sp. Strain aff17, but slightly increase the decolourization process even at a high concentration of 2.0 mg/L. Copper which is essential for laccase, surprisingly showed lower down the decolourization at the concentration above 1.5 mg/L. Mercury inhibited the decolourization even at a low concentration of 0.5 mg/L. The detection of LMEs was studied in 1L and 2L conical flasks. Larger surface area was shown to have a better decolourization rate when 500 mL of working volume was placed into both 2L flask and 1L flask. Two Litre flask's culture has a larger surface area providing more contact with air therefore providing a better condition for dye decolourization. Laccase and Manganese peroxidase were found in Coriolopsis sp. Strain aff17. These Lignin Modifying Enzymes' activities show inverse relationship to the concentration of azo dye. Coriolopsis sp. Strain aff17 was grown on agricultural wastes such as sugarcane bagasse, sugarcane peel and paddy straw, to serve as the alternative inoculum in the 2L decolourizing system to replace the utilization of Potato Dextrose Broth.

*Coriolopsis* sp. Strain aff17 that was grown on paddy straw showed the fastest decolourization, which was six days, followed by sugarcane peels (eight days) and sugarcane bagasse (twelve days). *Coriolopsis* sp. Strain aff17 was then tested on raw wastewaters that were collected from rivers near textile factories of Pahang, Malaysia, to further study its ability to decolourize raw wastewater. *Coriolopsis* sp. Strain aff17 was able to decolourize limited raw wastewater without addition of glucose by 12.74 to 24.89%. However with the addition of glucose, the decolourization percentage can reach over 84%. In conclusion, *Coriolopsis* sp. Strain aff17 is proven to be able to decolourize various azo dyes. More studies should be continued to determine if *Coriolopsis* sp. Strain aff17 is able to decolourize dyes in harsher conditions found in raw wastewater. *Coriolopsis* sp. Strain aff17 also should be tested on other pollutants.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

### PENGURAIAN PERWARNA TEKSTIL AZO OLEH KULAT REPUT-PUTIH **CORIOLOPSIS SP. STRAIN AFF17**

Oleh

#### CHENG WEI NEE

#### Oktober 2014

#### : Profesor Mohd. Arif Syed, PhD Pengerusi : Bioteknologi dan Sains Biomolekul Fakulti

Perwarna sintetik telah menggantikan perwarna semula jadi dalam industri tekstil moden. Walau bagaimanapun, 10% sisa perwarna masuk ke dalam persekitaran dan menyebabkan pencemaran air. Memandangkan perwarna sintetik adalah dicipta untuk menahan pemudaran warna, maka ia akan berada kekal dalam persekitaran. Rawatan beasaskan kimiafizik merupakan cara yang biasa gunakan tetapi ia mempunyai kelemahan dan kecekapan yang terhad. Jadi, biopemulihan sisa perwarna dengan menggunakan mikroorganisma semakin mendapat perhatian saintifik sebagai kaedah rawatan pencemaran alternatif untuk perwarna sintetik. Keupayaan mengurai pewarna tekstil azo oleh kulat-reput putih tempatan Coriolopsis sp. Strain aff17 (EU863193) yang dipencil daripada UPM, Serdang telah kaji tentang keupayaannya mengurai tiga belas perwarna tekstil yang terpilih. Ia boleh menguraikan sebelas perwarna tekstil. Hanya tujuh perwarna tekstil azo, Ponceau 2R (C.I. 20470), Amaranth (C.I. 16185), Orange G (C.I. 16230), Naphthol Blue Black (C.I. 20470), Remazol Black B (C.I. 20505), Trypan Blue (C.I. 23850) dan Sirius Light Blue BRR (C.I. 34140) telah dipilih untuk menjalakan kajian yang seterusnya. Kadar penguraian adalah lebih tinggi dalam kultur goncang apabila dibanding dengan kultur pegun. Kelajuan yang terbaik bagi penguraian perwarna azo adalah 75 rpm. Medium kultur terbaik untuk penguraian perwarna adalah air suling. Kehadiran kromium, arsenik, kadmium dan plumbum tidak mengpengaruhi keupayaan Coriolopsis sp. Strain aff17 untuk mengurai perwarna bahkan ia dapat meningkatkan kadar penguraian walaupun kepekatannya adalah setinggi 2.0 mg/L. Kuprum yang diperlui oleh lakase, tidak meningkatkan kadar penguraian tetapi menjejaskan penguraian pada kepekatan 1.5 mg/L. Raksa menjejaskan kadar penguraian walaupun kepekatannya adalah serendah 0.5 mg/L. Kawasan permukaan yang lebih besar menunjukkan kadar penguraian yang lebih cepat apabila isipadu 500 mL kultur yang diletak dalam 2L kalalang kon dan 1L kelalang kon. Dua Liter kelalang kon mempunyai kawasan permukaan yang lebih besar untuk mendapat lebih banyak udara jadi ia menyediakan persekitaran untuk penguraian perwarna. Enzim yang terdapat dalam Coriolopsis sp. Strain aff17 adalah lakase dan mangan peroksidase. Aktiviti enzim-enzim ini berkadar songsang dengan kepekatan perwarna azo. Coriolopsis sp. Strain aff17 ditumbuhkan ke atas sisa-sisa pertanian seperti



hampas tebu, kulit tebu dan jerami padi supaya mereka boleh menggantikan media Potato Dextrose Broth yang mahal. *Coriolopsis* sp. Strain aff17 yang ditumbuhkan di atas jerami padi mempunyai kadar penguraian yang terpantas, ia itu enam hari, diikuti oleh kulit tebu (lapan hari) dan hampas tebu (dua belas hari). *Coriolopsis* sp. Strain aff17 juga diuji untuk menguraikan air sisa tekstil yang dikutip dari sungai berdekat dengan kilang tekstil di Pahang, Malaysia. *Coriolopsis* sp. Strain aff17 boleh menguraikan perwarna tanpa tambahan glukosa, tetapi pada kadar yang sangat rendah (12.74 ke 24.89%). Walau bagaimanapun dengan tambahan glukosa, peratus penguraian perwarna mencapai 84%. Kesimpulannya, *Coriolopsis* sp. Strain aff17 dibukti boleh menguraikan perwarna tekstil azo. Lebih kajian perlu dijalankan supaya *Coriolopsis* sp. Strain aff17 boleh menguraikan perwarna dalam keadaan yang sukar dalam air sisa tekstil. *Coriolopsis* sp. Strain aff17 juga boleh dicuba atas pencemar-pencemar yang lain.

I certify that a Thesis Examination Committee has met on 20 October 2014 to conduct the final examination of Cheng Wei Nee on her thesis entitled "Decolourization of Selected Textile Dyes by White Rot Fungus Coriolopsis sp. Strain aff17" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Muhajir bin Hamid, PhD Associate Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Chairman)

Janna Ong binti Abdullah, PhD Associate Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Internal Examiner)

Mohd. Puad bin Abdullah, PhD Associate Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Internal Examiner)

Mohd Faiz Foong Abdullah, PhD Associate Professor Faculty of Applied Sciences Universiti Teknologi Mara (External Examiner)

> ZULKARNAIN ZAINAL, PhD Professor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date: 23 January 2015

## ACKNOWLEDGEMENTS

First of all, I would like to extend my greatest and deepest thanks to my supervisor, Prof. Dr. Mohd. Arif Syed for his invaluable guidance throughout the completion of my studies. He has always been my biggest support. My warmest gratitude also goes to my co-supervisor, Assoc. Prof. Dr. Mohd. Yunus Abdul Shukor for his supervision, advice, commentsm patience guidance, constructive suggestions and critical review of my work during the course of this research.

I would like to thank my labmates as well which including Yao Ping, Hui Keem, Aqlima, Zila, Asma, Hana and all members from Lab 115 and 204. And I want to take this opportunity to thank Mr. Ariff and Mr. Sim Han Koh as they gave me a lot of suggestions when I was stuck in my studies.

Last but not least, to all my friends in UPM, Pin Shang, Ka Heng, Haw Eong, Chee Bui, Lik Gin, Wei Tat, Yun Yan. They encourage me everytime I was fed up with my labworks. Thank you so much for everything. This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

# Mohd Arif Syed, PhD

Professor Faculty of Biotechnology and Biomolecular Science Universiti Putra Malaysia (Chairman)

## Mohd. Yunus Abd Shukor, PhD

Associate Professor Faculty of Biotechnology and Biomolecular Science Universiti Putra Malaysia (Member)

> **BUJANG KIM HUAT, PHD** Professor and Dean School of Graduate Studies Univerisiti Putra Malaysia

Date:

# Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fullyowned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholary intergrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The tesis has undergone plagiarism detection Software.

Signature: \_

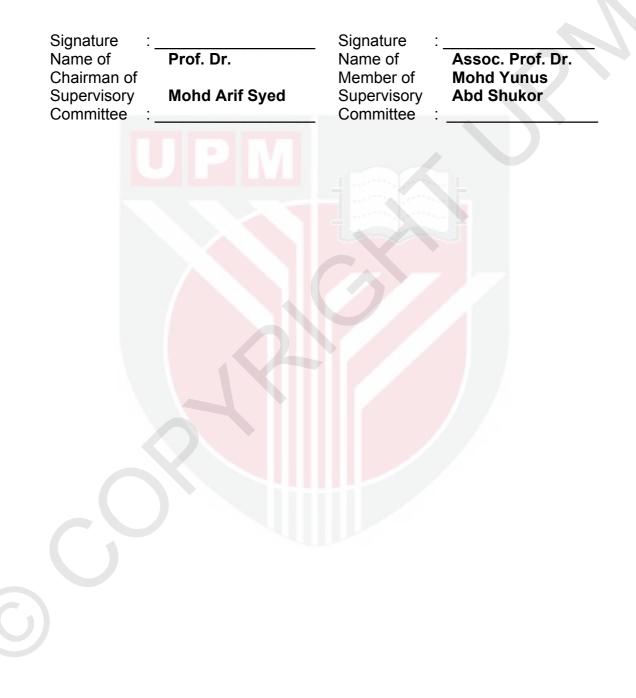
Date:

Name and Matric No: Cheng Wei Nee GS22415

# **Declaration by Members of Supervisory Committee**

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.



# **TABLE OF CONTENTS**

ABSTRACT ABSTRAK ACKNOWLEDGEMENT APPROVAL DECLARATION LIST OF TABLES LIST OF FIGURES LIST ABBREVIATIONS LIST OF APPENDICES

### CHAPTER

1	INTRO	DDUCTION	1
2	LITER	RATURE REVIEWS	3
	2.1	History of Dyes	3
	2.2	Synthetic Dyes	3 3 4
	2.3	Classification of Synthetic Dyes	4
	2.4	Discharge of Dyes into Environment	9
	2.5	Structure and Azo Dye	9
	2.6	Nomenclature of Azo Dye	11
	2.7	Ecotoxicity of Synthetic Dyes	11
	2.8	Removal of Azo Dyes	12
		2.8.1 Physicochemical Methods	12
		2.8.2 Biological Methods	12
	2.9	Bioremediation	13
	2.10	Azo Dyes Bioremediation	13
		2.10.1 Azo Dyes Bioremediation by Bacteria	13
		2.10.2 Azo Dye Biodegradation by Wood-rotting Fungi	14
		2.10.3 Advantages of White-rot Fungi Over Other	16
		Microorganisms in Azo Dye Biodegradation	
		2.10.4 Coriolopsis sp.	17
	2.11	Lignin-Modifying Enzymes	18
		2.11.1 Laccase	19
		2.11.2 Lignin Peroxidase	21
		2.11.3 Manganese peroxidase	21
	2.12	Heavy Metals	22
		2.13 Alternative Growing Substrate for White Rot Fungus	23
3	МАТЕ	RIALS AND METHODS	24
	3.1	General Overiew	24
	3.2	Chemicals and Equipments Used	24
	3.3	Biological Materials	24
	3.4	Subculture and Maintainance of <i>Coriolopsis</i> sp. Strain aff17	24
	3.5	Qualitative Screening of <i>Coriolopsis</i> sp. Strain aff17's	24
		Azo Dye Decolourzing Ability on Solid Medium	
		3.5.1 Inoculum Preparation for Solid Culture	25

Page

		3.5.2 Qualitative Screening	25
	3.6	Quantitative Screening of Coriolopsis sp. Strain aff17's	28
		Azo Dye Decolourzing Ability in Liquid Medium	
		3.6.1 Coriolopsis sp. Strain aff17 Inoculum Preparation	28
		3.6.2 <i>Coriolopsis</i> sp. Strain aff17 Culture Medium	29
		3.6.3 Quantitative Measurement by UV-Visible	30
		Spectrophotometer	00
		• •	31
			51
		Azo Dye Decolourization	04
		3.6.4.1 Static and Agitated Culture of	31
		Coriolopsis sp. Strain aff17	04
		3.6.4.2 Effects of Shaking Speed	31
		3.6.4.3 Effects of pH System	32
		3.6.4.4 Effects of Heavy Metals	32
	3.7	Raw Textile Wastewater Analysis	32
		3.7.1 Measurement of pH	32
		3.7.2 Heavy Metals Analysis by Atomic Absorption Spectrometer	32
		3.7.3 Decolourization of Raw Textile Wastewater	32
		3.7.3.1 Decolourization Without Glucose	33
		3.7.3.2 Decolourization Without Clucose	33
	3.8	Detection of Lignin Modifying Enzyme in Different Lager	33
	5.0		55
		Working Volume	22
		3.8.1 Effects of Lager Working Volume	33
		3.8.2 Detection of Lignin Modifying Enzyme	33
		3.8.2.1 Assay for Lignin Peroxidase Activity	33
		3.8.2.2 Assay for Manganese Peroxidase Activity	34
		3.8.2.3 Assay for Laccase Activity	34
	3.9	Alternative Substrate for <i>Coriolopsis</i> sp. Strain aff17	34
4	RESI	JLTS AND DISCUSSIONS	36
	4.1	Qualitative Screening of Coriolopsis sp. Strain aff17's	36
		Azo Dye Decolourizing Ability on Solid Medium	
	4.2	Characterization of Coriolopsis sp. Strain aff17 in	40
		Azo Dye Decolourization Studies	
		4.2.1 Static and Agitated Culture of <i>Coriolopsis</i> sp.	40
		Strain aff17	40
		4.2.2 Effects of Shaking Speed	44
		4.2.3 Effects of pH	46
		4.2.4 Effects of Heavy Metals	<del>4</del> 0 55
	12		63
	4.3	Raw Textile Wastewater Analysis	
	4.4	Detection of Lignin Modifying Enzyme in Different	66
		Lager Working Volume	~ 7
		4.4.1 Effects of Surface Area on Amaranth	67
		Decolourization	_
		4.4.2 Effects of Surface Area on Enzymes Activities	67
	4.5	Alternative Substrate for Coriolopsis sp. Strain aff17	70

5	CONCLUSIONS AND RECOMMENDATIONS FOR FUTURE RESEARCH		
	5.1	Conclusions	73
	5.2	Recommendations for Future Research	74
REFERENCES APPENDICES BIODATA OF STUDENT		76 92 140	



LIST OF TABLES

 $\bigcirc$ 

Table		Page
2.1	Classification of synthetic dyes based on chromophore present	4
2.2	Classes of synthetic dyes according to Colour Index (C.I.)	6
2.3	Classification of synthetic dyes based on the method of application and usage	7
2.4	Estimated fixation rate and worldwide sale of dyes	9
2.5	Example of azo dyes that can be decolourized by bacteria	14
2.6	White-rot fungi capable of azo dye biodegradation	16
3.1	List of Common name, C.I. Number, C.I Generic Name and structures of different azo dyes	25
3.2	Absorption maxima of seven azo dyes that were used in quantitative screening	29
4.1	Screening results of <i>Coriolopsis</i> sp. Strain aff17 and <i>Phanerocheate chrysosporium</i> over azo dyes after 10 days of incubation	38
4.2	The percentage of adsorption and percentage of degradation of different azo dyes	44
4.3	Final pH of decolourization culture	46
4.4	Heavy metals levels in water samples from	55
	dye-contaminated waters. Detection with atomic absorption spectroscopy (AAS)	
4.5	Initial pH of raw wastewaters and their maximum absorbance	63
4.6	Comparison of decolourization of actual effluent with or without glucose supply after 10 days of incubation.	66

# LIST OF FIGURES

 $\bigcirc$ 

Figure			
Figur	e	Page	
2.1	Structural representation of a trans- monoazo dye	10	
2.2	Azo dye synthesis via a two-reaction process	10	
2.3	Reduction of azo dyes and formation of aromatic amines	17	
2.4	Proposed mechanism for laccase degradation of azo dyes	20	
3.1	Mycelial mat that formed in culture flasks after seven days of incubation.	29	
3.2	Calculation for the decolourization by Coriolopsis sp. Strain aff17.	31	
4.1	Decolourization of Orange G at day 6 by Phanerochaete	37	
	chrysosporium and Coriolopsis sp. strain aff17 showing different		
	pattern of decolourized zones.		
4.2.	Decolourization of Naphthol Blue Black at day 6 by Phanerochaet	e 37	
	chrysosporium and Coriolopsis sp. strain aff17 showing different		
4.0	pattern of decolourized zones.	11	
4.3	Decolourization percentage of azo dyes by <i>Coriolopsis sp.</i> strain	41	
4.4	aff17 in 50 mL static liquid cultures incubated at room temperature	ə. 43	
4.4	Decolourization percentage of azo dyes by <i>Coriolopsis sp.</i> strain aff17 in 50 mL agitated liquid cultures at 50 rpm incubated at	43	
	room temperature.		
4.5	Effect of shaking speed on the decolourization of Amarant	45	
4.6	Effect of pH on decolourization of Orange G	47	
4.7	Effect of pH on decolourization of Ponceau 2R	48	
4.8	Effect of pH on decolourization of Amaranth	49	
4.9	Effect of pH on decolourization of Direct Blue 71	50	
4.10	Effect of pH on decolourization of Trypan Blue	52	
4.11	Effect of pH on decolourization of Naphthol Blue Black	53	
4.12	Effect of pH on decolourization of Remazol Black B	54	
4.13	Effect of different concentration of chromium (Cr) on	56	
	of Amaranth		
4.14	Effect of different concentration of arsenic (Ar) on	57	
	decolourization of Amaranth	50	
4.15	Effect of different concentration of cadmium (Cd) on	58	
4.16	decolourization of Amaranth Effect of different concentration of lead (Pb) on	60	
4.10	decolourization of Amaranth	00	
4.17	Effect of different concentration of copper (Cu) on	61	
	decolourization of Amaranth	01	
4.18	Effect of different concentration of mercury (Hg) on	62	
	decolourization of Amaranth		
4.19	Percentage of decolourization of absorbance of raw textile	64	
	wastewaters by Coriolopsis sp. Strain aff17 without addition of		
	carbon source (glucose).		
4.20	Percentage of decolourization of absorbance of raw textile	65	
	wastewaters by Coriolopsis sp. Strain aff17 with addition		
	of carbon source (glucose).		

4.21 Percentage of Amaranth decolourization by *Coriolopsis* sp. Strain 68

aff17 in 1L and 2L flask together with laccase and MnP's activity in each flask.

71

4.22 Percentage of decolourization of AMR by *Coriolopsis* sp. Strain aff17 that grown in different substrates.



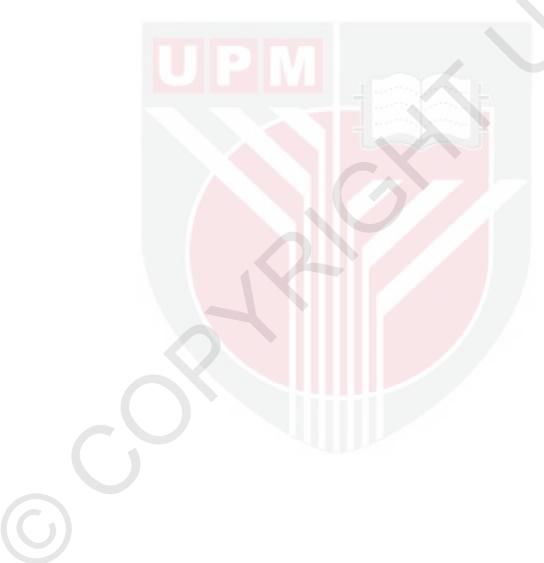
# LIST OF ABBREVIATIONS

2,6-DMP AMR	2,6-dimethoxyphenol Amaranth
BJ	Bukit Juntong
BS	Biebrich Scarlet
CAS C.I.	Chemical Abstracts Service Colour Index
DB71	
EDTA	Direct Blue 71 (alson known as Sirius Light Blue BRR)
IUPAC	Ethylenediaminetetraacetic Acid International Union of Pure Applied Chemistry
KBM	Kirk Basal Medium
KL	Kampung Ladang
LiP	Lignin Peroxidase
LME	Lignin Modifying Enzyme
MnP	Manganese Peroxidase
MR	Methyl Red
MY	Metanil Yellow
NBB	Naphthol Blue Black
OG	Orange G
02	Orange II Sodium Salt
PDA	Potato Dextrose Agar
PDB	Potato Dextrose Broth
PDS	Paddy Straw
PS	Ponceau S
P2R	Ponceau 2R
RBB	Remazol Black B
SB	Sugarcane Bagasse
SP	Sugarcane Peel
TB	Trypan Blue
TE TT7	Trace Elements
TTZ WRF	Tartrazine
	White Rot Fungus

# LIST OF APPENDICES

Appe	ndix	Page
 	List of Chemicals and Equipments Formula for the Calculation of the Enzyme Activities and Specific Activities of LMEs	92 93
III	Qualitative Screening of Coriolopsis sp. Strain aff17	94
IV	Azo Dye Decolourizing Ability on Solid Medium with Amaranth Qualitative Screening of <i>Coriolopsis</i> sp. Strain aff17 Azo Dye Decolourizing Ability on Solid Medium with Biebrich Scarlet	95
V	Qualitative Screening of <i>Coriolopsis</i> sp. Strain aff17 Azo Dye Deccolourizing Ability on Solid Medium with Ponceau 2R	96
VI	Qualitative Screening of <i>Coriolopsis</i> sp. Strain aff17 Azo Dye Deccolourizing Ability on Solid Medium with Ponceau S	97
VII	Qualitative Screening of <i>Coriolopsis</i> sp. Strain aff17 Azo Dye Deccolourizing Ability on Solid Medium with Methyl Red	98
VIII	Qualitative Screening of <i>Coriolopsis</i> sp. Strain aff17 Azo Dye Deccolourizing Ability on Solid Medium with Orange II Sodium S	99 alt
IX	Qualitative Screening of <i>Coriolopsis</i> sp. Strain aff17 Azo Dye Deccolourizing Ability on Solid Medium with Orange G	100
Х	Qualitative Screening of <i>Coriolopsis</i> sp. Strain aff17 Azo Dye Deccolourizing Ability on Solid Medium with Metanil Yellow	101
XI	Qualitative Screening of <i>Coriolopsis</i> sp. Strain aff17 Azo Dye Deccolourizing Ability on Solid Medium with Tartrazine	102
XII	Qualitative Screening of <i>Coriolopsis</i> sp. Strain aff17 Azo Dye Deccolourizing Ability on Solid Medium with Direct Blue 71	103
XIII	Qualitative Screening of <i>Coriolopsis</i> sp. Strain aff17 Azo Dye Deccolourizing Ability on Solid Medium with Remazol Black B	104
XIV	Qualitative Screening of <i>Coriolopsis</i> sp. Strain aff17 Azo Dye Decolourizing Ability on Solid Medium with Naphthol Blue Black	105
XV	Qualitative Screening of <i>Coriolopsis</i> sp. Strain aff17 Azo Dye Decolourizing Ability on Solid Medium with Trypan Blue	106
XVI	Quantitative Screening of Coriolopsis sp. Strain aff17 Azo Dye Decolourizing Ability on Liquid Medium	107
XVII	Standard Curve of Azo Dyes	111
XVIII		114
XIX	Comparison of decolourization of Different Azo Dyes at Agitated Condition (50 rpm)	118
XX	Two-way ANOVA Analysis of Static Culture and Agitated Culture on Different Azo Dyes after 36 hours of Incubation	122
XXI	Two-way ANOVA Analysis on Different Speed of Shaking over 10 Hours of Incubation	123
XXII	Two-way ANOVA Analysis on Decolourization of Orange G in Different pH System	124
XXIII	Two-way ANOVA Analysis on Decolourization of Ponceau 2R in Different pH System	126
XXIV	Two-way ANOVA Analysis on Decolourization of Amaranth in Different pH System	128
XXV	Two-way ANOVA Analysis on Decolourization of Direct	130

	Blue 71 in Different pH System	
XXVI	Two-way ANOVA Analysis on Decolourization of Trypan	132
	Blue in Different pH System	
XXVII	Two-way ANOVA Analysis on Decolourization of Naphthol	134
	Blue Black in Different pH System	
XXVIII	Two-way ANOVA Analysis on Decolourization of Remazol	136
	Black B in Different pH System	
XXIX	Two-way ANOVA Analysis on Effect on Mercury (Hg) in	138
	Decolourization of Amaranth	



## CHAPTER 1

#### INTRODUCTION

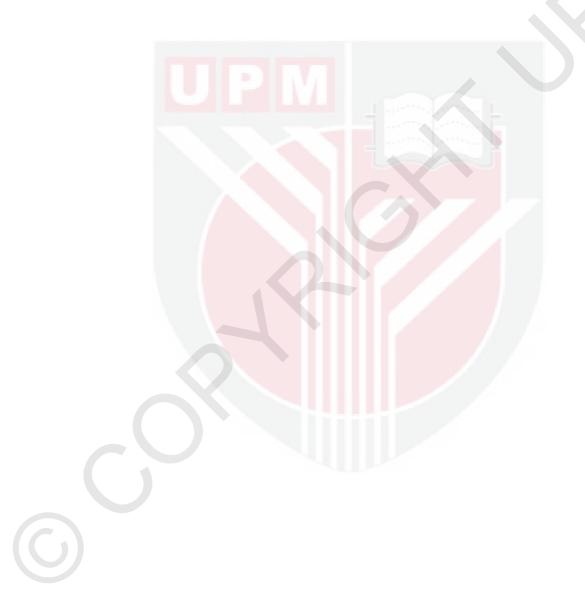
In recent years, synthetic dyes are mostly replaced natural dyes in a number of industries, such as textile dyeing or paper printing. Each year, India, Eastern Europe, China, South Korea and Taiwan consume approximately 800 thousands tons of dyes, and at least 10-15% of dyestuff wastes are enter to the environment (Kumar *et al.*, 2013; Selvam *et al.*, 2012; Xie *et al.*, 2012; Sinha *et al.*, 2012; Palmieri *et al.*, 2005; Levin *et al.*, 2004).

Synthetic dyes are designed to resist fading upon exposure to sweat, light water, many chemicals including oxidizing agents, and microbial attack (Selvam *et al.*, 2012) and therefore, they are recalcitrant in the environment. Regardless of the advantages of synthetic dyes over natural dyes, synthetic dyes present their own new set of problems. The most noticeable is the aesthetic pollution of waterways caused by the presence of dyes leached from textile factories since they are visible even in minute amounts (Stoyanova *et al.*, 2014; Verma *et al.*, 2012; Banat *et al.*, 1996). The production of potentially carcinogenic aromatic amine compounds from the partial cleavage of synthetic dyes are harmful to human (Zheng *et al.*, 2014; Ali, 2010; Yilmaz *et al.*, 2010; Pinheiro *et al.*, 2004). Therefore, the frequently volumetric rate of dyestuff waste discharges raise concern of stringent legislation, the search for appropriate treatment technologies has become an important priority in textile and paper industries (Rodriguez-Couto, 2012; O'Neill *et al.*, 1999).

A number of biotechnological approaches have been suggested by recent researches as of potential interest towards combining this pollution source in a coefficient manner, including the use of bacteria or fungi, often in combination with physicochemical processes (Kumar *et al.*, 2013; Yadav *et al.*, 2013; Zodi *et al.*, 2013; Saratale *et al.*, 2011). By far the single class of microorganisms most efficient in breaking down synthetic dyes are white rot fungi (WRF). *Phanerochaete chrysoporium* is among the first to have been shown to have the ability to degrade azo dyes (Kumar *et al.*, 2013, Priya and Arulmozhi, 2012; Son *et al.*, 2012).

Biodegradation of azo dyes by WRF presents a great potential for large-scale applications after many bioremediation processes being investigated for this purpose. Its biodegradation system, comprising of lignin modifying enzymes are not only efficient but also have a wide substrate range. At this time, most research are focusing on a narrow range of well-known WRF while the rich biodiversity of fungi found in tropical forests such as in Malaysia is ignored most of the time. There are reasons to believe that these undiscovered species might have greater azo dye degrading abilities compared to the ones that are being studied now. Hence, the processes to isolate and screen new white-rot fungi for the biodegradation of azo dyes have to be done intensively.

- The objectives of this research were: I. To characterize the decolourization ability of *Coriolopsis* sp. strain aff17.
  - To determine the ligninolytic enzymes present in Coriolopsis sp. strain II. aff17
  - To evaluate the alternative substrates for *Coriolopsis* sp. strain aff17. III.



#### REFERENCES

- Abadulla, E., Tzanov, T., Costa, S., Robra, K.-H., Cavaco-Paulo, A. and Gubitz, G.M. 2000. Decolourization and detoxification of textile dyes with a laccase from *Trametes hirsuta*. *Applied and Environmental Microbiology* 66: 3357-3362.
- Abdul-Wahab, M.F., Chan, G.F., Yusoff, A.R.M. and Rashid, N.A.A.R. 2013. Reduction of azo dyes by flavin reductase from *Citrobacter freundii* A1. *Journal of Xenobiotics* 3: 9-13.
- Adejoye, O.D. and Fasidi, I.O. 2009. Biodegradation of agro-wastes by some Nigerian white-rot fungi. Bioresources 4: 816-824.
- Adenipekun, C.O., Olanrewaju, O.O.and Ogunjobi, A.A. 2011. Bioaccumulation of heavy metals and nutrient content supplementation by two white rot fungi in crude oil polluted soil. *Researchers* 3: 13-20.
- Adeyemi, A.O. 2009. Bioaccumulation of arsenic by fungi. American Journal of Environmental Sciences 5: 364-370.
- Aksu, Z., Kılıç, N.K., Ertuğrul, S. and Dönmez, G. 2007. Inhibitory effects of chromium(VI) and Remazol Black B on chromium(VI) and dyestuff removals by *Trametes versicolor. Enzyme and Microbial Technology* 40: 1167-1174.
- Albanis, T.A., Hela, D.G., Sakellarides, T.M. and Danis, T.G. 2000. Removal of Dyes from Aqueous Solutions by Adsorption on Mixtures of Fly Ash and Soil in Batch and Column Techniques. *Global Nest: The International Journal* 2: 237–244.
- Aleu, J. and Collado, I.G. (2001). Biotransformations by *Botrytis* species. *Journal of Molecular Catalysis B: Enzymatic* 13: 77-93.
- Ali, H. 2010. Biodegradation of Synthetic Dyes A Review. Water Air Soil Pollution 213: 251-273
- Amaral, P.F.F., Fernandes, D.L.A., Tavares, A.P.M., Xavier, A.B.M.R., Cammarota, M.C., Coutinho, J.A.P. and Coelho, M.A.Z. 2004. Decolorization of dyes from textile wastewater by *Trametes versicolor*. *Environmental Technology* 25: 1313-1320.
- Arslan, M., Sayin, S. and Yilmaz, M. 2013. Removal of carcinogenic azo dyes from water by new cyclodextrin-immobilized iron oxide magnetic nanoparticles. 2012. *Water, Air, Soil Pollution* 224: 1-9.
- Balan, D.S.L and Monteiro, R.T.R. 2001. Decolorization of textile indigo dye by ligninolytic fungi. *Journal of Biotechnology* 89: 141-145.

- Baldrian, P., in der Wiesche, C., Gabriel, J., Nerud, F. and Zadrazil, F. 2000. Influence of cadmium and mercury on activities of ligninolytic enzymes and degradation of polycyclic aromatic hydrocarbons by *Pleurotus ostreatus* in soil. *Applied Environmental Microbiology* 66: 2471-2478.
- Baldrian, P. 2003. Interactions of heavy metals with white-rot fungi. Enzyme Microbial Technology 32: 78-91.
- Banat, I.B., Nigam, P., Singh, D. and Marchant, R. (1996). Microbial decolorization of textile-dye containing effluents: a review. *Bioresource Technology* 58: 217-227.
- Barber, E. J. W. 1991. Prehistoric textiles: the development of cloth in the neolithic and bronze ages with special reference to the aegean. Princeton: Princeton University Press.
- Basha, S.A. and Rajaganesh K. 2014. Microbial bioremediation of heavy metals from textile industry dye effluents using isolated bacterial strains. *International Journal of Current Microbiology and Applied Sciences* 3: 785-794.
- Bizani, E., Fytianos, K., Poulios, I., and Tsiridis, V. 2006. Photocatalytic decolorization and degradation of dye solutions and wastewaters in the presence of titanium dioxide. *Journal of Hazardous Materials* 136: 85– 94.
- Biyik, H., Basbulbul, G., Kalyoncu, F., Kalmis, E. and Oryasin, E. 2012. Biological decolourization of textile dyes from isolated microfungi. *Journal of Environmental Biology* 33: 667-671.
- Boer, C.G., Obici, L., Marques de Souza, C.G. and Peralta, R.M. (2004). Decolorization of synthetic dyes by solid state cultures of *Lentinula* (*Lentinus*) edodes producing manganese peroxidase as the main ligninolytic enzyme. *Bioresource Technology* 94: 107-112.
- Boopathy, R. 2000. Factors limiting bioremediation technologies. *Bioresource Technology* 74: 63-67.
- Bourbonnais, R. and Paice, M.G. 1988. Veratryl alcohol oxidases from the lignin-degrading basidiomycete *Pleurotus sajor-caju. Journal of Biochemistry* 255: 445-450.
- Bromley-Challenor, K.C.A., Knapp, J.S., Zhang, Z., Gray, N.C.C., Hetheridge, M.J. and Evans, M.R. 2000. Decolorization of an azo dye by unacclimated activated sludge under anaerobic conditions. *Water Research* 34: 4410-4418.
- Bumpus, J.A. 1995. Microbial degradation of azo dye. In *Biotransformations: Microbial degradation of health risk compounds*, ed. Singh V.P., pp 157-176. Amsterdam: Elsevier Science Ltd.

- Calvo, A.M., Galletti, G.C. and González, A.E. 1995. Paper waste-water analyses by pyrolysis-gas chromatography/mass spectrometry during biological decolorization with the fungi *Coriolopsis gallica* and *Paecilomyces variotii*. *Journal of Analytical and Applied Pyrolysis* 33: 39-50.
- Camassola, M. and Dillon, A.J.P. 2009. Biological pretreatment of sugarcane bagasse for the production of cellulases and xylanases by *Penicillium echinulatum*. *Industrial Crops and Products 29: 642-647*.
- Cardona, C.A., Quintero, J.A. and Paz, I.C. Production of bioethanol from sugarcane bagasse: Status and perspectives. *Bioresource Technology* 101: 4754-4766.
- Chagas, E. P., and L. R. Durrant. 2001. Decolorization of Azo Dyes by Phanerochaete Chrysosporium and Pleurotus Sajorcaju. Enzyme and Microbial Technology 29: 473–477.
- Cook, S.W. and Chancey, J.R. 2005. *By native hands: woven treasures from the Lauren Rogers Museum of Art*. Lauren Rogers Museum of Art.
- Chen, K.C., Wu, J.Y., Liou, D.J. and Hwang, S.J. 2003. Decolorization of textile dyes by newly isolated bacterial strains. *Journal of Biotechnology* 101: 57–68.
- Chivukula, M. and Renganathan, V. (1995). Phenolic azo dye oxidation by laccase from *Pyricularia oryzae. Applied and Environmental Microbiology* 61: 4374-4377.
- Christie, R.M. 2001. Colour chemistry. *In Azo Dyes and Pigments* pp 45-68. The Royal Society of Chemistry, Cambridge, UK.
- Collins, P.J., Dobson, A.D.W. and Field, J.A. (1998). Reduction of the 2,2'-Azinobis(3-Ethylbenzthiazoline-6-Sulfonate) cation radical by physiological organic acids in the absence and presence of manganese. *Applied and Environmental Microbiology* 64: 2026-2031.
- Crini, G. 2006. Non-conventional low-cost adsorbents for dye removal: A review. *Bioresource Technology* 97: 1061-108.
- Dâassi, D., Belbahri, L., Vallat, A., Woodward, S., Nasri, M. and Mechichi, T. 2013. Enhanced reduction of phenol content and toxicity in olive mill wastewaters by a newly isolated strain of *Coriolopsis gallica*. *Environmetal Science Pollution Research* 21: 1746-1758.
- Daneshvar, N., Oladegaragoze, A. and Djafarzadeh, N. 2006. Decolorization of basic dye solutions by electrocoagulation: An investigation of the effect of operational parameters. *Journal of Hazardous Materials* 129: 116-122.

- Dayaram, P. and Dasgupta, D. 2008. Decolorisation of synthetic dyes and textile wastewater using Polyporus rubidus. *Journal of Environmental Biology* 29: 831-836.
- Deacon, J.W. (1984). Volume 7, Introduction to modern mycology, pp 22-23, 160-166. London: Blackwell Scientific Publications.
- De Jong, E., De Vries, F.P., Field, J.A., Van Der Zwan, R.P. and De Bont, J.A.M. 1992. Isolation and screening of basidiomycetes with high peroxidative activity. *Mycological Research* 96: 1098-1104.
- Deniz, F. and Karaman, S. 2011. Removal of Basic Red 46 dye from aqueous solution by pine tree leaves. *Chemical Engineering Journal* 170: 67-74.
- Eichlerova, I., Homolka, L., Benada, O., Kofronova', O., Huba'lek, T. and Nerud, F. 2007. Decolorization of orange G and remazol brilliant blue R by the white rot fungus *Dichomitus squalens*: toxicological evaluation and morphological study. *Chemosphere* 69, 795–802.
- Elisashvili, V., Penninckx, M., Kachlishvili, E., Tsiklauri, N., Metreveli, E., Kharziani, T. and Kvesitadze, G. 2008. *Lentinus edodes* and *Pleurotus* species lignocellulolytic enzymes activity in submerged and solid-state fermentation of lignocellulosic wastes of different composition. *Bioresource Technology* 99: 457-462.
- El-Mashad, H.M., Zeeman, G., van Loon, W.K.P., Bot, G.P.A. and Lettinga, G. 2004. Effect of temperature and temperature fluctuation on thermophilic anaerobic digestion of cattle manure. *Bioresource Technology* 95: 191-201.
- Ersahin, M.E., Ozgun, H., Dereli, R.K., Ozturk, I., Roest, K. and Lier, J.B.V. 2012. A review on dynamic membrane filtration: materials, applications and future perspectives. *Bioresource Techonology* 122: 196-206.
- Forgacs, E., Cserháti, T., Oros, G. 2004. Removal of synthetic dyes from wastewaters: A review. Environment International 30: 953-971.
- Fuh, M.R. and Chia, K.J. 2002. Determination of sulphonated azo dyes in food by ion-pair liquid chromatography with photodiode array and electrospray mass spectrometry detection. *Talanta* 56: 663–671.
- Gianfreda, L. and Rao, M.A. 2004. Potential of extra cellular enzymes in remediation of polluted soils: a review. *Enzyme and Microbial Technology* 35: 339-354.
- Glenn, J.K. and Gold, M.H. 1983. Decolorization of several polymeric dyes by the lignin-degrading basidiomycete *Phanerochaete chrysosporium*. *Applied and Environmental Microbiology* 45: 1741-1747.

- Glenn, J.K. and Gold, M.H. 1985. Purification and characterization of an extracellular Mn(II)-dependent peroxidase from the lignin-degrading basidiomycete, *Phanerochaete chrysosporium*. *Archives of Biochemistry and Biophysics* 242: 329-341.
- Goodwin, J. 1982. A Dyer's Manual. In *Analysis, Treatment, and Techniques: Fibers and Textiles*. London: Pelham Books Ltd.
- Gopinath, K.P., Murugesan, S., Abraham, J. and Muthukumar, K. 2009. Bacillus sp. mutant for improved biodegradation of Congo red: Random mutagenesis approach. *Bioresource Technology* 100: 6295-6300.
- Gültekin, I. and Ince, N.H. 2006. Degradation of aryl-azo-naphthol dyes by ultrasound, ozone and their combination: Effect of α -substituents. *Ultrasonics Sonochemistry* 13: 208-214.
- Harvey, P.J. and Thurston, C.F. (2001). The biochemistry of ligninolytic fungi. *In* Fungi in bioremediation, ed. Gadd. G.M. pp 27-51. Cambridge: Cambridge University Press.
- Hatakka, A. 1994. Lignin-modifying enzymes from selected white-rot fungi: production and role from in lignin degradation. *FEMS Microbiology Reviews* 13: 125-135.
- Heinfling, A., Martinez, M.J., Martinez, A.T., Bergbauer, M. and Szewzyk, U. (1998). Transformation of industrial dyes by manganese peroxidases from *Bjerkandera adusta* and *Pleurotus eryngii* in a manganeseindependent reaction. *Applied and Environmental Microbiology* 64: 2788-2793.
- Heinzkill, M., Bech, L., Halkier, T., Schneider, P. and Anke, T. 1998. Characterization of laccases and peroxidases from wood-rotting fungi (Family *Coprinaceae*). *Applied and Environmental Microbiology* 64: 1601–1606.
- Hofrichter, M., Vare, T., Kalsi, M., Galkin, S., Scheibner, K., Fritsche, W. and Hatakka, A. 1999. Production of manganese peroxidase and organic acids and mineralization of <sup>14</sup>C-labelled lignin (14C-DHP) during solidstate fermentation of wheat straw with the white rot fungus *Nemataloma frowardii. Applied and Environmental Microbilogy* 65: 1864-1870.
- Hossain, S.M. and Anantharaman, N. 2006. Activity enhancement of ligninolytic enzymes of *Trametes versicolor* with bagasse powder. *African Journal of Biotechnology* 5: 189-194.
- Hou, H., Zhou, J., Wang, J., Du, C. and Yan, B. (2004). Enhancement of laccase production by *Pleurotus ostreatus* and its use for the decolorization of anthraquinone dye. *Process Biochemistry* 39: 1415-1419.

- Howard, P.H., Boethling, R.S., Jarvis, W.F., Meylan, W.M. and Michalenko, E.M. 1991. Handbook of Environmental Degradation Rates, pp 318-319, 624-625. Washington D.C: Lewis Publishers.
- Hunger, K. 2003. Industrial Dyes, Chemistry, Properties, Applications. In *Dyes, General Survey*, pp. 1-7. Weinheim, Germany: Wiley-VCH.
- Işık, M. and Sponza, D.T. 2008. Anaerobic/aerobic treatment of a simulated textile wastewater. *Separation and Purification Technology* 60: 64-72.
- Iwamoto, T. and Nasu, M. (2001). Current bioremediation practice and perspective. *Journal of Bioscience and Engineering* 92: 1-8.
- Jadhav, S.U., Kalme, S.D., Govindwar, S.P. 2008. Biodegradation of methyl red by *Galactomycytes geotrichum* MTCC 1360. International Biodeterioration and Biodegradation 62: 135-142.
- Jonsson, L., Sjostrom, K., Haggstrom, I., Nyman, P.O. 1995. Characterization of a laccase gene from the white-rot fungus Trametes versicolor and structural features of basidiomycete laccases. *Biochimica et Biophysica Acta - Protein Structure and Molecular Enzymology* 1251: 210-215.
- Joo, D.J, Shin, W.S., Choi, J.H., Choi, S.J., Kim, M.C., Han, M.H., Ha, T.W. and Kim, Y.H. 2007. Decolorization of reactive dyes using inorganic coagulants and synthetic polymer. *Dyes and Pigments* 73: 59-64.
- Kalme, S., Ghodake, G. and Govindwar, S. 2007. Red HE7B degradation using desulfonation by *Pseudomonas desmolyticum* NCIM 2112. *International Biodeteriotation & Biodegradation* 60: 327-333.
- Kalpana, D., Shim, J.H., Oh, B., Senthil, K. and Lee, Y.S. 2011.
  Bioremediation of the heavy metal complex dye Isolan Dark Blue 2SGL-01 by white rot fungus *Irpex lacteus*. *Journal of Hazardous Materials* 198: 198-205
- Kapdan, I.K., Kargi, F., McMullan, G. and Marchant, R. 2000. Effect of environmental conditions on biological decolorization of textile dyestuff by *C. versicolor. Enzyme and Microbial Technology* 26: 381-387.
- Kasinath, A., Novotny, C., Svobodova, K., Patel. K.C. and Sasek, V. 2003. Decolourization of synthetic dyes by *Irpex lacteus* in liquid cultures and packed-bed bioreactor. *Enzyme and Microbial Technology* 32: 167-163.
- Keck, A., Klein, J., Kudlich, M., Stolz, A., Knackmuss, H.J. and Mattes, R. 1997. Reduction of azo dyes by redox mediators originating in the naphthalenesulfonic acid degradation pathway of *Sphingomonas* sp. strain BN6. *Applied and Environmental Microbiology* 63: 3684-3690.

- Kelly, R.L. 1988. Ligninolytic activity of *Phanerochaete chrysosporium* measured as ethylene production from α-keto-γ-methylthiolbutyric acid. *Methods in Enzymology* 161B: 79-82.
- Khehra, M.S., Saini, H.S., Sharma, D.K., Chadha, B.S. and Chimni, S.S. 2005. Comparative studies on potential of consortium and constituent pure bacterial isolates to decolorize azo dyes. *Water Research* 39: 5135-5141.
- Kiernan, J.A. 2001. Classification and naming of dyes, stains and fluorochromes. *Biotechnic & Histochemistry* 76: 261-278.
- Koshy, J. and Nambisan, P. 2011. Biopulping of Paddy Straw by *Pleurotus eous. Advanced BioTech 11:* 44-46.
- Kumar, S.S., Balasubramanian, P. and Swaminathan, G. 2013. Degradation potential of free and immobilized cells of white rot fungus Phanerochaete chrysosporium on synthetic dyes. *International Journal* of Chem Tech Research 2013: 565-571
- Levin, L., Forchiassin, F. and Ramos, A.M. 2002. Copper induction of ligninmodifying enzymes in the white-rot fungus *Trametes trogii*. *Mycologia* 94: 377-383.
- Levin, L., Papinutti, L. and Forchiassin, F. 2004. Evaluation of Argentinean white rot fungi for their ability to produce lignin-modifying enzymes and decolourise industrial dyes. *Bioresource Technology* 94: 169-176.
- Li, X., Lin, X., Zhang, J., Wu, Y., Yin, R., Feng, Y. and Wang, Y. 2010. Degradation of polycyclic aromatic hydrocarbons by crude extracts from spent mushroom substrate and its possible mechanisms. *Current Microbiology* 60: 336-342.
- Maas, R. and Chaudhari, S. 2005. Adsorption and biological decolourization of azo dye Reactive Red 2 in semicontinuous anaerobic reactors. *Process Biochemistry 40*: 699-705.
- Maceiras, R., Rodriguez-Couto, S., and Sanroman, A. 2001. Influence of several activities on the extracellular laccase activity and *in vivo* decolourization of Poly R-478 by semi-solid-state cultures of *Trametes versicolor*. *Acta Biotechnology* 21: 255–264.
- Machuca, A., Napoleao, D. and Milagres, A.M.F. 2001. Detection of metalchelating compounds from wood-rotting fungi *Trametes versicolor* and *Wolfiporia cocos*. *World Journal of Microbiology & Biotechnology* 17: 687-690.
- Maheswari, S. and Murugesan, A.G. 2009. Biosorption of arsenic(III) ion from aqueous solution using *Aspergillus fumigatus* isolated from arsenic contaminated site. *Desalination & Water Treatment* 11: 294-301.

- Maier, J., Kandelbauer, A., Erlacher, A., Cavaco-Paulo, A. and Gubitz, G.M. 2004. A new alkali-thermostable azoreductase from *Bacillus* sp. Strain SF. *Applied and Environmental Microbiology* 70: 837-844.
- Maximo, C., Amorim, M.T.P. and Costa-Ferreira, M. 2003. Biotransformation of industrial reactive azo dyes by *Geotrichum* sp. CCMI 1019. *Enzyme and Microbial Technology* 32: 145-151.
- Mayer, A.M. and Staples, R.C. 2002. Laccase: a new function for an old enzyme. *Phytochemistry* 60: 551-565.
- Meehan, C., Banat, I.M., McMullan, G., Nigam, P., Smyth, F. and Marchant, R. 2000. Decolorization of Remazol Black-B using a thermotolerant yeast, *Kluyveromyces marxianus* IMB3. *Environment International* 26: 75-79.
- Mezohegyi, G., Bengoa, C., Stuber, F., Font, J., Fabregat, A. and Fortuny, A. 2008. Novel bioreactor design for decolourisation of azo dye effluents. *Chemical Engineering Journal* 143: 293-298.
- Michizoe, J., Uchimura, Y., Ichinose, H., Maruyama, T., Kamiya, N., Wariishi, H., Furusaki, S. and Goto, M. (2004). Activation of manganese peroxidase in an organic medium using a mediator. *Biochemical Engineering Journal* 19: 43-46.
- Mittal, A., Jhare, A. and Mittal, J. 2013. Adsorption of hazardous dye Eosin Yellow from aqueous solution onto waste material De-oiled Soya: Isotherm, kinetics and bulk removal. *Journal of Molecular Liquids* 179: 133-140.
- Morikawa Y, Shiomi K, Ishihara Y, Matsuura N. 1998. Triple primary cancers involving kidney, urinary bladder and liver in a dye workers. *American Journal of Industrial Medicine* 31: 44-49.
- Moyson, E. and Verachtert, H. 1991. Growth of higher fungi on wheat straw and their impact on the digestibility of the substrate. *Applied Microbiology & Biotechnology* 36: 421-424.
- Munoz, C., Guillen, F., Martinez, A.T. and Martinez, M.J. 1997. Laccase isoenzymes of *Pleurotus eryngii*: characterization, catalytic properties, and participation in activation of molecular oxygen and Mn<sup>2+</sup> oxidation. *Applied and Environmental Microbiology* 63: 2166-2174.
- Munro, J. H. 2003. The Cambridge Hstory of Western Texties. In *Medieval Woollens: Textiles, Technology, and Organisation, c. 800-1500, ed.* D. Jenkins, pp. 181-227. Cambridge: Cambridge University Press.

- Munro, J. H. 2007. Medieval Clothing and Textiles. In *The Anti-Red Shift To the 'Dark Side': Colour Changes in Flemish Luxury Woollens, 1300-1500*, ed. R. Netherton, and G.R. Owen-Crocker, pp. 55-96. Trowbridge, Wiltshire: Cromwell Press.
- Ngah, W.S.W., Teong, L.C. and Hanafiah, M.A.K.M. 2011. Adsorption of dyes and heavy metal ions by chitosan composites: A review. *Carbohydrate Polymers* 83: 1446-1456.
- Nigam, P., Armour, G.I., Banat, M., Singh, D. and Marchant, R. 2000. Physical Removal of Textile Dyes from Effluents and Solid-state Fermentation of Dye-adsorbed Agricultural Residues. *Bioresource Technology* 72: 219–226.
- Nikazar M., Davarpanah L. and Vahabzadeh F. 2008. Biosorption of aqueous chromium (VI) by living mycelium of *Phanerochaete chrysosporium*. *Chemical Engineering Transactions* 14: 475-480.
- Nilsson, I., Möller, A., Mattiasson, B., Rubindamayugi, M.S.T. and Welander, U. 2006. Decolorization of synthetic and real textile wastewater by the use of white-rot fungi. Enzyme & Microbial Technology 38: 94-100.
- Novotný, Č., Svobodová, K., Erbanová, P., Cajthaml, T., Kasinath, A., Lang, E. and Šašek, V. 2004. Ligninolytic fungi in bioremediation: extracellular enzyme production and degradation rate. *Soil Biology and Biochemistry* 36: 1545-1551.
- Nozaki, K., Beh, C.H., Mizuno, M., Isobe, T., Shiroishi, M., Kanda, T. and Amano, Y. 2008. Screening and investigation of dye decolorization activities of basidiomycytes. *Journal of Bioscience and Bioengineering* 105: 69-72.
- Nyanhongo, G.S., Gomes, J., Gubitz, G.M., Zvauya, R., Read, J. and Steiner, W. 2002. Decolorization of textile dyes by laccases from a newly isolated strain of *Trametes modesta*. *Water Research* 36: 1449-1456.
- Okitsu, K., Iwasaki, K., Yobiko, Y., Bandow, H., Nishimura, R. and Maeda, Y. 2005. Sonochemical degradation of azo dyes in aqueous solution: a new heterogeneous kinetics model taking into account the local concentration of OH radicals and azo dyes. *Ultrasonic Sonochemistry* 12: 255-262.
- Ollgaard, H., Frost, L., Galster, J., Hansen, O.C., 1998. Survey of azocolorants in Denmark: consumption, use, health and environmental aspects (no. XX 1998). Denmak: Ministry of Environment and Energy, Danish Environmental Protection Agency.
- O'Neill, C., Hawkes, F.R., Lourenco, N.D., Pinheiro, H.M. and Delee, W. 1999. Colour in textile effluents-sources, measurements, discharge

consents and simulation: a review. *Journal of Chemical Technology & Biotechnology* 74: 1009-1018.

- Orna M.V. 1980. Chemistry and artists' colors. Part III. Preparation and properties of artists' pigments. *Journal of Chemical Education* 57: 267.
- Pagès, D., Sanchez, L., Conrod, S., Gidrol, X., Fekete, A., Schmitt-Kopplin, P., Heulin, T. and Achouak, W. 2007. Exploration of intraclonal adaptation mechanisms of *Pseudomonas brassicacearum* facing cadmium toxicity. *Environmental Microbiology* 9: 2820-2835.
- Palmans, E., Mares, M., Poppe, J. and Höfte, M. 1995. Biodegradation of xenobiotics by heavy metal resistant higher fungi. Gent, Belgium. Ninth Forum for Applied Biotechnology, September 27-29. 1995. Proceedings part II: 2593-2594.
- Palmieri, G., Cennamo, G. and Sannia G. 2005. Remazol Brilliant Blue R decolourisation by the fungus *Pleurotus ostreatus* and its oxidative enzymatic system. *Enzyme and Microbial Technology* 36: 17-24.
- Pandey, A., Singh, P., and Iyengar, L. 2007. Bacterial decolourization and degradation of azo dyes. *International Biodeterioration and Biodegradation* 59: 73-84.
- Pant, D., Singh, A., Satyawali, Y. and Gupta, R.K. 2008. Effect of carbon and nitrogen source amendment on synthetic dyes decolourizing efficiency of white-rot fungus, *Phanerochaete chrysosporium*. *Journal of Environmental Biology* 29: 79-84.
- Paszczynski, A., Crawford, R.L. and Huynh, V.B. 1988. Manganese peroxidase of *Phanerochaete chrysosporium*: purification. *Methods in Enzymology* 161: 264-270.
- Pearce, C.I., Lloyd, J.R. and Guthrie, J.T. (2003). The removal of colour from textile wastewater using whole bacterial cells: a review. *Dyes and Pigments* 58: 179-196.
- Pickard, M.A., Roman, R., Tinoco, R. and Vazquez-Duhalt, R. 1999. Polycyclic aromatic hydrocarbon metabolism by white rot fungi and oxidation by *Coriolopsis gallica* UAMH 8260 laccase. *Applied and Environmental Microbiology* 65: 3805-3809.
- Pinheiro, H.M., Touraud, E. and Thomas, O. 2004. Aromatic amines from azo dye reduction: status review with emphasis on direct UV spectrophotometric detection in textile industry wastewaters. *Dyes and Pigments* 61: 121-139.

- Pointing, S.B., Bucher, L.P. and Vrijmoed, L.L. 2000. Dye decolourization by subtropical basidiomycetous fungi and the effect of metals on decolourizing ability. *World Journal of Microbiology and Biotechnology* 16: 199-205.
- Priya, S.V. and Arulmozhi, M. 2012. Bioremediation of colored industrial effluent using the white-rot fungi *Phanerochaete Chrysoporium*. *Advances in Enigineering, Science and Management*: 199-203.
- Raghukumar, C., D'Souza, T.M., Thorn, R.G. and Reddy, C.A. 1999. Ligninmodifying enzymes of *flavodon flavus*, a basidiomycete isolated from a coastal marine environment. *Applied and Environmental Microbiology* 65: 2103-2111.
- Rani, C., Jana A.K. and Bansal, A. 2011. Studies on the biodegradation of azo dyes by white rot fungi *Daedalea flavida* in the absence of external carbon source. 2011 2nd International Conference on Environmental Science and Technology IPCBEE 6: 147-150. Singapore: IACSIT Press.
- Rieger, P.G., Meier, H.M. and Gerle, M. 2002. Xenobiotics in the Environment: Present and Future Strategies to Obviate the Problem of Biological Persistence. *Journal of Biotechnology* 94: 101–123.
- Rodriguez-Couto, S. 2012. A promising inert support for laccase production and decolouration of textile wastewater by the white-rot fungus *Trametes pubescesns. Journal of Hazardous Materials* 233-234: 158-162.
- Rossbach, S., Kukuk, M.L., Wilson, T.L., Feng, S.F., Pearson, M.M. and Fisher, M.A. 2000. Cadmium-regulated gene fusions in *Pseudomonas fluorescens*. *Environmental Microbiology* 2: 373-382.
- Rothschild, N., Levkowitz, A., Hadar, Y. and Dosoretz, C.G. (1999). Manganese deficiency can replace high oxygen levels needed for lignin peroxidase formation by *Phanerochaete chrysosporium*. *Applied and Environmental Microbiology* 65: 483-488.
- Russ, R., Rau, J. and Stolz, A. 2000. The function of cytoplasmic flavins reductases in the reduction of azo dyes by bacteria. *Applied and Environmental Microbiology* 66: 1429-1434.
- Saha, B.C. 2003. Hemicellulose bioconversion. *Journal of Industrial Microbiology & Biotechnology* 30: 29-291.
- Saratale, R.G., Saratale, G.D., Chang, J.S. and Govindwar, S.P. 2011. Bacterial decolorization and degradation of azo dyes: a review. *Journal of the Taiwan Institute of Chemical Engineers* 2011: 138-157.

- Saravanan, P., Sivakumar, P., Suganya, T., Gandhi, N.N. and Renganathan, S. 2012. Bioaccumulation of reactive red 11 using live yeast *Rhodotorula glutinis. Indian Journal Environmental Protection* 32: 249-255.
- Sarkar, N. and Aikat, K. 2012. Alkali pretreatment of rice straw and enhanced cellulase production by a locally isolated fungus *Aspergillus fumigatus* NITDGPKA3. *Journal of Microbiology & Biotechnology Research* 2: 717-726.
- Sarkar, S., Satheshkumar, A., Pradeepa, N. and Premkumar, R. 2010. Hexavalent chromium (Cr (VI)) removal by live mycelium of a *Trichoderma harzianum* strain. *Asian Journal of Experimental Biological Sciences* 1: 606-613.
- Sathishkumar, P., Arulkumar, M. and Palvannan. 2012. Utilization of agroindustrial waste *Jatropha curcas* pods as an activated carbon for the adsorption of reactive dye Remazol Brilliant Blue R (RBBR). *Journal of Cleaner Production* 22: 67-75.
- Satyanarayana, K.G., Guimaraes, J.L. and Wypych, F. 2008. Studies on lignocellulosic fibers of Brazil. Part I: source, production, morphology properties and applications. *Composites Part A: Applied Science & Manufacturing* 38: 1694-1709.
- Say, R., Denizli, A. and Arica, M.Y. 2001. Biosorption of cadmium(II), lead(II) and copper(II) with the filamentous fungus *Phanerochaete chrysosporium. Bioresource Technology* 76: 67-70.
- Seidenari, S., Giusti, F., Massone, F. and Mantovani, L. 2002. Sensitization to disperse dyes in a patch test population over a five-year period. *American Journal of Contact Dermatitis* 13: 101-107.
- Selvam, K., Arungandhi, K., Rajenderan, G. and Yamuna, M. 2012. Biodegradation of azo dyes and textile industry effluent by newly isolated white rot fungi. *Open Access Scientific Reports* 1. <u>http://www.omicsonline.org/scientific-reports/2155-6199-SR-564.pdf</u>.
- Sen, S., Singh, P.P., Rathore, V.S. and Pereira, B.M.J. 2004. Sugarcane bagasse improves the activity of ligninolytic enzymes and decolourization of dyes by the white-rot fugus *Pleurotus ostreatus*. *Journal of Scientific & Industrial Research* 63: 739-746.
- Sequin-Frey, M. 1981. The Chemistry of Plant and Animal Dyes. *Journal of Chemical Education* 58: 301-305.
- Shah, M.P. and Patel, K.A. 2014. Microbial decolourization and degradation of remazol black & mordant orange by microbial consortia isolated from common effluent treatment plant. International Journal of Environmental *Bioremediation & Biodegradation* 2: 117-124.

- Sharma, R.K. and Arora, D.S. 2010. Changes in biochemical constituents of paddy straw during degradation by white rot fungi and its impact on in vitro digestibility. *Journal of Applied Microbiology* 109: 679-686.
- Shin, M., Nguyen, T. and Ramsay, J. 2002. Evaluation of support materials for the surface immobilization and decolourization of amaranth by *Trametes versicolor. Applied Microbial Biotechnology* 60: 218-223.
- Sing, C. and Yu, J. 1998. Copper adsorption and removal from water by living mycelium of white rot fungus *Phanerochaete chrysosporium*. *Water Research* 32: 2746-2752.
- Singh, D., Zeng, J.J., Laskar, D.D., Deobald, L., Hiscox, W.C. and Chen, S.L. 2011. Investigation of wheat straw biodegradation by *Phanerochaete chrysosporium*. *Biomass Bioenergy* 35: 1030-1040.
- Sinha, K., Saha, P.D. and Datta, S. 2012. Response surface optimization and artificial and neural network modelling of microwave assisted natural dye extraction from pomegranate rind. *Industrial Crops and Products* 37: 408-414.
- Soltani, T. and Entezari, M.H. 2013. Photolysis and photocatalysis of methylene blue by ferrite bismuth nanoparticles under sunlight irradiation. *Journal of Molecular Catalysis A: Chemical* 377: 197-203.
- Son, Y., Kim, H., Thiyagarajan, S., Xu, J.J. and Park, S. 2012. Heterologous expression of Phanerochaete chrysosporium glyoxal oxidase and its application for the coupled reaction with manganese peroxidase to decolorize malachite green. *Mycobiology* 40: 258-262.
- Songulashvili, G., Jimenéz-Tobón, G.A., Jaspers, C. Gratia, J., Debaste, F. and Penninckx M.J. 2014. Immobilized Coriolopsis sp. laccase for continuous elimination and transformation of phenolic micropollutants. *Water Quality Research Journal of Canada* 49: 328-338.
- Spadaro, J.T., Gold, M.H. and Renganathan, V. 1992. Degradation of azo dyes by the lignin-degrading fungus *Phanerochaete chrysosporium*. *Applied and Environmental Microbiology* 58: 2397-2401.
- Spencer, J. 2007. China Pays Steep Price As Textile Exports Boom. *The Wall Street Journal Online.* <u>http://online.wsj.com/news/articles/SB118580938555882301</u>
- Stoyanova, M., Slavova, I., Christoskova, St. and Ivanova, V. 2014. Catalytic performance of supported nanosized cobalt and iron-cobalt mixed oxides on MgO in oxidative degradation of Acid Orange 7 azo dye with peroxymonosulfate. *Applied Catalysis A: General* 476: 121-132.

- Sumit, P. and Vimala, Y. 2012. Comparative study on bioremediation of chromium in fortified solutions by viable, biosorbent and immobilized cells of *Phanerochaete chrysosporium*-MTCC787. *International Journal of Microbiology Research* 4: 240-248.
- Swamy, J. and Ramsay, J.A. 1999. The evaluation of white rot fungi in the decoloration of textile dyes. *Enzyme and Microbial Technology* 24: 130-137.
- Tekere, M., Mswaka, A.Y., Zvauya, R. and Read, J.S. 2001. Growth, dye degradation and ligninolytic activity studies on Zimbabwean white rot fungi. *Enzyme and Microbial Technology* 28: 420-426.
- Thassitou, P.K. and Arvanitoyannis, I.S. (2001). Bioremediation: a novel approach to food waste management. *Food Science and Technology* 12: 185-196.
- Thompson, F and Thompson, Tony. 1987. Synthetic Dyeing: for Spinners, Weavers, Knitters and Embroiderers. Newton Abbot, Devon: David & Charles Publishers.
- Tien, M. and Kirk, T.K. 1983. Lignin-degrading enzyme from the Hymenomycete *Phanerochaete chrysosporium* Burds. *Science* 221: 661-663.
- Toh, Y.C., Yen, J.J.L., Obbard, J.P. and Ting, Y.P. 2003. Decolourisation of azo dyes by white rot fungi (WRF) isolated in Singapore. *Enzyme and Microbial Technology* 33: 569-575.
- Torres, E., Bustos-Jaimes, I. and Le Borgne, S. (2003). Potential use of oxidative enzymes for the detoxification of organic pollutants. *Applied Catalysis B: Environmental* 46: 1-15.
- Umasaravanan, D., Jayapriya, J. and Rajendran, R. 2011. Comparison of lignocelluloses biodegradation in solid state fermentation of sugarcane bagasse and rice straw by *Aspergillus tamari*. *Ceylon Journal of Science* 40: 65-68.
- Ürek, R.O. and Pazarlioglu, N.K. 2004. Purification and partial characterization of manganese peroxidase from immobilized *Phanerochaete chrysosporium. Process Biochemistry* 39: 2061-2068.
- Valmaseda, M., Martinez, M.J. and Martinez, A.T. 1991. Kinetics of wheat straw solid-state fermentation with *Trametes versicolor* and *Pleurotus ostreatus*: lignin and polysaccharide alteration and production of related enzymatic activities. *Applied Microbiology & Biotechnology* 15: 817-823.
- Verma, A.K., Dash, R.R. and Bhunia, P. 2012. A review on chemical coagulation/flocculation technologies for removal of colour from textile wastewaters. *Journal of Environmental Management* 93: 154-168.

- Wang, M., Funabiki, K. and Matsui, M. 2003. Synthesis and properties of bis (hetaryl) azo dyes. *Dyes and Pigments* 57: 77-86.
- Wang, Y., Wang, A., Liu, W., Kong, D., Tan, W. and Liu, C. 2013. Accelerated azo dye removal by biocathode formation in singlechamber biocatalyzed electrolysis systems. *Biosource Technology* 146: 740-743.
- Welham, A. 2000. The theory of dyeing (and the secret of life). *Journal of the Society of Dyers and Colourists* 116: 140-143.
- Wesenberg, D., Kyriakides, I. and Agathos, S.N. 2003. White rot fungi and their enzymes for the treatment of industrial dye effluents. *Biotechnology Advances* 22: 161-187.
- Wong, P.K. and Yuen, P.Y. 1998. Decolourization and biodegradation of N,N'-dimethyl-p-phenylenediamine by *Klebsiella pneumonia* RS-13 and *Acetobacter liquefaciens* S-1. *Journal of Applied Microbiology* 85: 79-87.
- Xie, H.C., Li, C.R., Wang, L. and Li, H. 2012. Phytotoxicity, adsorption, uptake and removal of azo dyes in sunflowers. *Advanced Materials Research* 573-574: 1090-1095.
- Yadav, A., Mukherji, S. and Garg, A. 2013. Removal of chemical oxygen demand and color from simulated textile wastewater using a combination of chemical/physiochemical processes. *Industrial & Engineering Chemistry Research* 52: 10063-10071.
- Yang, X.W., Zeng, Y.L., Ma, F.Y., Zhang, X.Y. and Yu, H.B. 2010a. Effect of bipretreatment on thermogravimetric and chemical characteristics of corn stover by different white-rot fungi. *Bioresource Technology* 101: 5457-5479.
- Yang, X.W., Ma, F.Y., Zeng, Y.L., Yu, H.B., Xu, C.Y. and Zhang, X.Y. 2010b. Structure alteration of lignin in corn stover degraded by white-rot fungus *Irpex lactues* CD2. *International Biodeterioration & Biodegradation* 64: 119-123.
- Yesilada, O., Asma, D. and Cing, S. (2003). Decolorization of textile dyes by fungal pellets. *Process Biochemistry* 38: 933-938.
- Yilmaz, E., Memon, S. and Yilmaz, M. 2010. Removal of direct azo dyes and aromatic amines from aqueous solutions using two β -cyclodextrinbased polymers. *Journal of Hazardous Materials* 174: 592-597.
- Yun, M.A., Yeon K.M., Park, J.S., Lee, C.H., Chun, J. and Lim D.J. 2006. Characterization of biofilm structure and its effect on membrane permeability in MBR for dye wastewater treatment. *Water Research* 40: 45-52.

- Zee, F. P. van der. 2002. Anaerobic azo dye reduction. ISBN: 90-5808-610-0 Thesis Wageningen University, Wageningen, The Netherlands. In Production and discharge statistic of dyes pp 7
- Zeikus, J.G. 1981. Lignin metabolism and the carbon cycle. Polymer biosynthesis, biodegradation and environmental recalcitrance. *Advances in Microbial Ecology* 5: 211-243.
- Zheng, M., Chi, Y., Yi, H. and Shao, S. 2014. Decolorization of Alizarin Red and other synthetic dyes by a recombinant laccase from *Pichia pastoris*. *Biotechnology Letters* 36: 39-45.
- Zhou, X., Gou, W., Yang, S., Zheng, H. and Ren, N. 2013. Ultrasonicassisted ozone oxidation process of triphenylmethane dye degradation: Evidence for the promotion effects of ultrasonic on malachite green decolorization and degradation mechanism. *Bioresource Technology* 128: 827-830.
- Zhou, X., Wen, X. and Feng, Y. 2007. Influence of glucose feeding on the ligninolytic enzyme production of the white-rot fungus *Phanerochaete chrysosporium*. *Frontiers of Environmental Science* & *Engineering in China* 1: 89-94.
- Zodi, S., Merzouk, B., Potier, O., Lapicque, F. and Leclerc, J-P. 2013. Direct red 81 dye removal by a continuous flow electrocoagulation/flotation reactor. *Seperation and Purification Technology* 108: 215-222.
- Zollinger, H. 2003. Color chemistry syntheses, properties, and applicatios of organic dyes and pigments. 3<sup>rd</sup> revised edition. In Azo Dyes and Pigments, pp 165-253. VCH, New York.