



UNIVERSITI PUTRA MALAYSIA

***ANALYSES OF PUTATIVE GENES INVOLVED IN THE DEFENSE
MECHANISM OF GRACILARIA CHANGII IN RESPONSE TO AGARASE
AND AGAROLYTIC BACTERIA TREATMENTS***

LIM EE LEEN

FBSB 2014 29



**ANALYSES OF PUTATIVE GENES INVOLVED IN THE DEFENSE
MECHANISM OF *GRACILARIA CHANGII* IN RESPONSE TO AGARASE
AND AGAROLYTIC BACTERIA TREATMENTS**

By

LIM EE LEEN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirements for the Degree of Master of Science**

August 2014

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

ANALYSES OF PUTATIVE GENES INVOLVED IN THE DEFENSE MECHANISM OF *GRACILARIA CHANGII* IN RESPONSE TO AGARASE AND AGAROLYTIC BACTERIA TREATMENTS

By

LIM EE LEEN

August 2014

Chair : Ho Chai Ling, PhD
Faculty : Biotechnology and Biomolecular Sciences

Seaweed *Gracilaria* is the main source of agar worldwide. Information on the induced defense mechanisms of seaweed is scarce, particularly in the aspect of seaweed-microbe interactions. A set of differentially expressed genes from *G. changii* in response to agarase were generated from a previous study through next generation sequencing of the seaweed transcriptomes. In this study, *G. changii* was treated with both agarase (which generates the microbe-induced molecular patterns) and a marine agarolytic bacteria, ABS1 isolated from degrading seaweed. The result from the previous study (agarase treatment) was verified and temporal gene expression of candidate genes at 1, 6 and 24 hours post-treatment (hpt) in response to both agarase and bacteria treatments was profiled by quantitative reverse-transcription real-time PCR (qRT-PCR). A total of 20 out of 22 candidates verified have the same gene expression patterns as the next generation sequencing result, demonstrating a 90.9% positive correlation between the two analyses. Four candidates encoding plasma membrane calcium-transporting ATPase (*GcPMCA*), vanadium bromoperoxidase type 1 (*GcVBPO1*), 3-phosphoshikimate 1-carboxyvinyltransferase (*GcEPSP*), and 12-oxophytodienoate reductase (*GcOPR*) showed more than 2-fold up-regulation compared to that of control samples upon agarase treatment, implying their importance in defense response. The gene encoding vanadium bromoperoxidase type 2 (*GcVBPO2*) showed more than 2-fold up-regulation compared to that of the control sample upon bacteria treatment. *GcVBPO1* and *GcVBPO2* displayed different expression profiles in response to the two treatments, indicating the existence of more than one signaling pathways in the transcriptional regulation of vanadium bromoperoxidase. The gene expression of 16 and 10 candidates were further profiled in agarase and bacteria treated samples at different time points, respectively. Most candidates were up-regulated at 1 hpt compared to that of the control sample at the same time point, indicating a rapid modulation of transcription in *G. changii* upon agarase treatment. The gene expression of these candidates displayed different expression profiles in bacteria treated samples. *GcEPSP* and *GcVBPO2*, were found to have the highest fold change when treated by agarase and agarolytic bacteria respectively, at 1 hpt. Temporal gene expression profile of candidates upon agarase and bacteria treatments in *G.*

changii indicated the occurrence of calcium signaling (at early stage), synthesis of tyrosine and phenylalanine through 4-hydroxyphenylpyruvate pathway, production of chorismate and jasmonic acid, increase of bromoperoxidation and heme peroxidation, and repression of photosynthesis upon pathogenic invasion. In conclusion, this study has provided further understanding on the gene expression of putative defense genes in *G. changii*.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**ANALISIS GEN ANGGAPAN YANG TERLIBAT DALAM MEKANISME
PERTAHANAN *GRACILARIA CHANGII* TERHADAP RAWATAN
AGARASE DAN BAKTERIA YANG MENDEGRADASI AGAR**

Oleh

LIM EE LEEN

Ogos 2014

Pengerusi : Ho Chai Ling, PhD
Fakulti : Bioteknologi dan Sains Biomolekul

Rumpai laut *Gracilaria* merupakan sumber agar yang utama di dunia. Maklumat mengenai mekanisme pertahanan teraruh rumpai laut adalah terhad, terutamanya dalam aspek interaksi rumpai laut dengan mikroorganisma. Satu set calon gen yang mempunyai ekpresi yang berbeza dalam *G. changii* yang bertindak balas terhadap agarase telah dijana daripada kajian terdahulu melalui penjujukan generasi baru ke atas transkriptom rumpai laut. Dalam kajian ini, *G. changii* dirawat dengan agarase (yang menjana corak molekul yang teraruh di mikrob) dan bakteria marin yang mendegradasi agar, ABS1 yang diasingkan daripada rumpai laut yang mereput. Keputusan daripada kajian terdahulu (rawatan agarase) telah disahkan dan ekspresi calon-calon pada 1, 6, dan 24 jam selepas rawatan agarase dan bakteria telah diprofil dengan kaedah kuantitatif tindak balas berantai polimeras masa nyata (qRT-PCR). Sebanyak 20 daripada 22 calon yang disahkan mempunyai corak ekspresi gen yang sama dengan keputusan penjujukan generasi baru, menunjukkan 90.9% korelasi positif antara kedua-dua analisis tersebut. Empat calon mengkodkan 'plasma membrane calcium-transporting ATPase' (*GcPMCA*), 'vanadium bromoperoxidase type 1' (*GcVBPO1*), '3-phosphoshikimate 1-carboxyvinyltransferase' (*GcEPSP*), dan '12-oxophytodienoate reductase' (*GcOPR*) menunjukkan peningkatan ekspresi gen yang lebih daripada 2 kali ganda berbanding dengan sampel kawalan dalam rawatan agarase, membayangkan kepentingan mereka dalam tindak balas pertahanan. Gen yang mengkodkan 'vanadium bromoperoxidase type 2' (*GcVBPO2*) menunjukkan peningkatan ekspresi gen yang lebih daripada 2 kali ganda berbanding dengan sampel kawalan dalam rawatan bakteria. *GcVBPO1* dan *GcVBPO2* memaparkan profil ekspresi yang berbeza di bawah dua rawatan tersebut, mencadangkan kewujudan lebih daripada satu laluan isyarat dalam pengawalan transkripsi 'vanadium bromoperoxidase'. Ekspresi 16 calon dari rawatan agarase and 10 calon dari rawatan bakteria pada masa yang berbeza telah diprofil dengan lebih lanjut. Kebanyakan calon menunjukkan peningkatan ekspresi gen pada 1 jam selepas rawatan berbanding dengan sampel kawalan pada masa yang sama, mencadangkan pengawalan yang cepat dalam transkripsi *G. changii* di bawah rawatan agarase. Ekspresi gen calon-calon tersebut memaparkan profil ekspresi yang berbeza di bawah rawatan bakteria. *GcEPSP* dan *GcVBPO* masing-masing didapati

mencatatkan peningkatan ekspresi yang paling tinggi dalam rawatan agarase dan rawatan bakteria, pada 1 jam selepas rawatan. Keputusan profil ekspresi calon-calon dalam rawatan agarase dan bakteria pada *G. changii* pada masa yang berbeza menunjukkan berlakunya isyarat kalsium (pada peringkat awal), sintesis tirosina dan fenilalanina melalui laluan 4-hidroksifenilpiruvat, penghasilan korismat dan asid jasmonik, peningkatan proses pengoksidaan bromo dan heme, dan penindasan fotosintesis di bawah serangan patogen. Kesimpulannya, kajian ini telah meningkatkan pemahaman terhadap ekspresi gen yang dijangka terlibat dalam pertahanan *G. changii*.



ACKNOWLEDGEMENTS

I owe a debt of gratitude to my supervisor, Assoc. Prof. Dr. Ho Chai Ling for her invaluable guidance and patient encouragements. This work would not be possible without her strong support. She is an incredible mentor, and I cherished the time I spent with her. I would also like to express my appreciation for Prof. Dr. Raha Abdul Rahim for being my supervisory committee member.

Sincere thanks are extended to all senior laboratory members especially Teo Swee Sen, Siow Rouh San, Tan Yung Chie, Yeoh Keat Ai, Khew Choy Yuen and Tee Syin Ying for their helpful advices and suggestions during the course of this project. Finally, I wish to thank my parents for their boundless love, and my little sister, for her never-ending encouragements.



I certify that a Thesis Examination Committee has met on 28.08.2014 to conduct the final examination of Lim Ee Leen on her thesis entitled “Analyses of Putative Genes Involved in the Defense Mechanism of *Gracilaria changii* in Response to Agarase and Agarolytic Bacteria Treatments” in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Raja Noor Zaliha Raja Abd. Rahman, PhD

Professor

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Chairman)

Janna Ong binti Abdullah, PhD

Associate Professor

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Internal Examiner)

Noor Azmi Shaharuddin, PhD

Lecturer

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Internal Examiner)

Chu Wan Loy, PhD

Professor

School of Postgraduate Studies

International Medical University

Malaysia

(External Examiner)

Noritah Omar, PhD

Deputy Dean and Associate Professor

School of Graduate Studies

Universiti Putra Malaysia

Date:

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Ho Chai Ling, PhD

Associate Professor
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Chairman)

Raha Abdul Rahim, PhD

Professor
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Member)

BUJANG BIN KIM HUAT, PhD
Dean and Professor
School of Graduate Studies
Universiti Putra Malaysia

Date:

DECLARATION

Declaration by graduate student

I hereby confirm that

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: _____

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____
Name of Chairman of
Supervisory
Committee: _____

Signature: _____
Name of Member of
Supervisory
Committee: _____

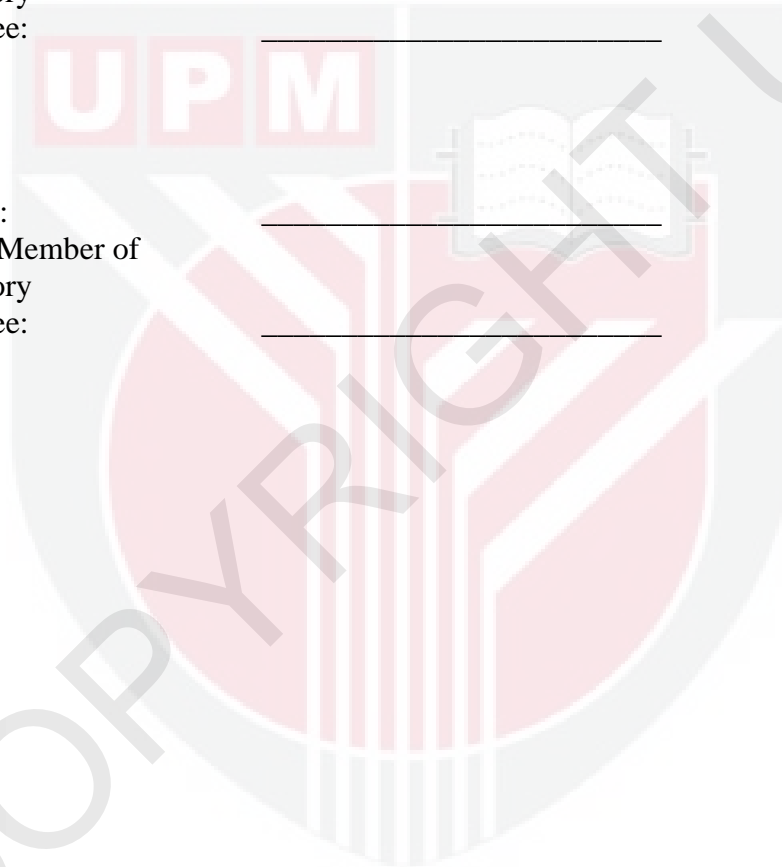


TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiii
LIST OF APPENDICES	xiv
LIST OF FIGURES	xv
LIST OF ABBREVIATIONS	xvi
CHAPTER	
1 INTRODUCTION	1
2 LITERATURE REVIEW	3
2.1 Seaweeds	3
2.1.1 Rhodophyta	7
2.1.1.1 Importance of Rhodophyta	9
2.1.2 <i>Gracilaria</i>	9
2.1.2.1 Cultivation of <i>Gracilaria</i>	10
2.1.2.2 <i>Gracilaria changii</i>	11
2.2 Seaweed diseases	11
2.2.1 Bacteria as pathogens of seaweeds	12
2.2.2 Agarase and agarolytic bacteria	12
2.3 Defense mechanisms of seaweed	13
2.3.1 Recognition of the attack	14
2.3.2 Signaling and transportation	15
2.3.3 Oxidative burst	16
2.3.4 Halogenation	17
2.3.5 Synthesis of inducible secondary metabolites	18
2.3.6 Production of oxylipins	19
2.4 Quantitative reverse transcription real-time PCR (qRT-PCR)	20
2.4.1 Application of qRT-PCR for quantification of gene expression.	20
3 METHODOLOGY	23
3.1 Isolation and identification of agarolytic bacteria	23
3.1.1 Isolation of agarolytic bacteria from <i>Gracilaria changii</i>	23
3.1.2 Construction of bacterial growth curve	23
3.1.3 Genomic DNA extraction	24
3.1.4 Analysis of 16S rDNA	24
3.1.4.1 PCR amplification of 16s rDNA	24
3.1.4.2 Preparation of <i>Escherichia coli</i> competent cell	25
3.1.4.3 Cloning of 16s rDNA	25
3.1.4.4 Sequencing and analysis of sequencing result	26

3.2	Collection and treatments of samples	26
3.2.1	Sample collection	26
3.2.2	Treatment of samples with agarase	26
3.2.3	Treatment of samples with agarolytic bacteria	26
3.3	RNA preparation	27
3.3.1	RNA extraction	27
3.3.2	Quantitative and qualitative analysis of RNA	28
3.4	cDNA synthesis	28
3.4.1	DNase treatment of RNA	28
3.4.2	Reverse transcription and cDNA synthesis	29
3.5	Sequence analysis and primer design for qRT-PCR	29
3.5.1	Sequence analysis	29
3.5.2	Primer design	29
3.6	Quantitative reverse transcription real-time PCR (qRT-PCR)	30
3.6.1	Evaluation of primer specificity	30
3.6.2	Evaluation of PCR amplification efficiency	30
3.6.3	Selection of endogenous control genes for qRT-PCR	34
3.6.4	qRT-PCR	34
3.6.5	Data analysis	34
4	RESULTS AND DISCUSSION	36
4.1	Isolation and identification of agarolytic bacteria	36
4.1.1	Isolation of agarolytic bacteria from <i>Gracilaria changii</i>	36
4.1.2	Analysis of 16S rDNA	36
4.1.3	Construction of bacterial growth curve	39
4.2	RNA preparation and cDNA Synthesis	41
4.2.1	Quantitative and qualitative analysis of RNA after DNase treatment	41
4.2.2	Reverse transcription and cDNA synthesis	43
4.3	Preparation for qRT-PCR	43
4.3.1	Evaluation of primer specificity and PCR amplification efficiency	43
4.3.2	Selection of endogenous control genes for qRT-PCR	44
4.4	Verification of gene expression upon agarase treatment and comparison with agarolytic bacteria treatment	46
4.4.1	Verification of gene expression in response to agarase treatment	46
4.4.2	Gene expression in response to agarolytic bacteria treatment	50
4.5	Temporal gene expression in treated seaweed samples	51
4.5.1	Proteins related to signaling and transportation	55
4.5.2	Proteins related to emission of reactive oxygen species	56
4.5.2.1	Superoxide-generating NADPH oxidase	56
4.5.2.2	L-ascorbate oxidase	56
4.5.2.3	Vanadium bromoperoxidases	57
4.5.2.4	Homologs of animal heme peroxidase	59
4.5.3	Proteins related to biosynthesis of aromatic amino acids	60
4.5.4	Proteins related to production of oxylipin, jasmonic acid	61

and fatty acid	
4.5.5 Proteins related to photosynthesis	63
4.6 Schematic representation of putative defense reactions	64
5 SUMMARY, CONCLUSION AND RECOMMENDATIONS FOR FUTURE RESEARCH	66
5.1 Summary and conclusion	66
5.2 Recommendations for future research	67
REFERENCES	69
APPENDICES	85
BIODATA OF STUDENT	90
LIST OF PUBLICATION	91



LIST OF TABLES

Table		Page
3.1	Primer sequences used in qRT-PCR.	31
4.1	BLASTN result of 16S rRNA gene sequence of ABS1 against nucleotide (nr/nt) database of GenBank.	38
4.2	Quality of <i>G. changii</i> RNA after DNase treatment.	42
4.3	Relative abundance of transcripts measured by next generation sequencing (NGS) and quantitative reverse-transcription real-time PCR (qRT-PCR) in response to agarase and bacteria treatment.	48

LIST OF APPENDICES

Appendix		Page
A	The amplification efficiency (E) and coefficient of determination (R^2) of qRT-PCR generated by each primer pairs of three endogenous control and all candidates.	85
B	Putative functions of defense-related candidates from <i>Gracilaria changii</i> .	86
C	Amino acid sequence alignment of <i>GcVBPO1</i> (GenBank accession no: AGE00855.1) and <i>GcVBPO2</i> .	88
D	Putative role of candidates <i>GcEPSP</i> , <i>GcTAT</i> and <i>GcPDT</i> in the aromatic amino acid biosynthesis pathways of plant.	89

LIST OF FIGURES

Figure		Page
2.1	Seaweed (<i>Gracilaria changii</i>) with turgid and cylindrical thallus.	4
2.2	Life cycles of seaweeds in three general categories: haplontic, diplontic, and alternation of generations.	5
3.1	Healthy and unhealthy <i>Gracilaria changii</i> .	23
4.1	Agarolytic Bacteria Strain 1 (ABS1).	37
4.2	Phylogenetic relationship of Agarolytic bacteria strain 1 (ABS1) and related bacteria in the class of <i>Gammaproteobacteria</i> (with <i>Zoogloea ramigera</i> as an out-group) was inferred using the Neighbor-joining method.	40
4.3	Growth curves of ABS1 in Marine Broth measured at OD ₆₀₀ and OD ₄₂₀ at 25°C.	41
4.4	RNA samples extracted from <i>G. changii</i> .	42
4.5	Melting curve generated by PCR product of <i>GcVBPO2</i> .	44
4.6	Evaluation of PCR amplification efficiency for <i>GcVBPO2</i> .	45
4.7	The geNorm analysis of four candidate endogenous control genes.	47
4.8	The relative abundance of transcript of candidates in response to agarase treatment at different time points.	52
4.9	The relative abundance of transcript of candidates in response to agarolytic bacteria treatment at different time points.	53
4.10	Schematic representation of putative defense reactions induced in <i>G. changii</i> upon agarase and agarolytic bacteria treatment.	65

LIST OF ABBREVIATIONS

AA	ascorbate
AAA	aromatic amino acid
AO	ascorbate oxidase
ATP	adenosine triphosphate
BLAST	Basic Local Alignment Search Tool
BSA	bovine serum albumin
cDNA	complementary deoxyribonucleic acid
Cq	quantification cycle value
DAHP	3-deoxy-d-arabino-heptulosonate- 7-phosphate
dATP	deoxyadenosine triphosphate
DEPC	diethylpyrocarbonate
DHA	dehydroascorbate
DMSO	dimethyl sulfoxide
DNA	deoxyribonucleic acid
DPI	diphenylene iodonium
DSP	dual-specific protein phosphatase
EDTA	ethylenediaminetetraacetic acid
EGTA	ethylene glycol-bis(2-aminoethylether)-N,N,N',N'-tetraacetic acid
EPSP	5-enolpyruvylshikimate 3-phosphate synthase
ERCA	endoplasmic reticulum calcium-transporting ATPase
hpt	hour post treatment
IPTG	isopropyl-beta-D-thiogalactopyranoside
JA	jasmonic acid
LB	Luria-Bertani
LOX	lipoxygenase
LPLAT	lysophospholipid acyltransferase
MAMP	microbe-associated molecular patterns
MAPK	mitogen-activated protein kinase
MEGA	molecular evolutionary genetics analysis
MeJA	methyl jasmonate

MIMP	microbe-induced molecular pattern
MOPS	3-(N-morpholino) propanesulfonic acid
mRNA	messenger ribonucleic acid
NADPH	nicotinamide adenine dinucleotide phosphate
NCBI	National Center for Biotechnology Information
NGS	next-generation sequencing
OD	optical density
p.p.t.	parts per thousand
PCR	polymerase chain reaction
Phe	phenylalanine
PMCA	plasma membrane calcium transporting ATPase
PUFA	polyunsaturated fatty acids
PUFA	polyunsaturated fatty acid
qRT-PCR	quantitative reverse-transcription real-time PCR
RNA	ribonucleic acid
ROS	reactive oxygen species
rRNA	ribosomal ribonucleic acid
SDS	sodium dodecyl sulfate
SSW	synthetic sea salt water
TAE	Tris/ acetate/ EDTA
Trp	tryptophan
Tyr	tyrosine
VBPO	vanadium bromoperoxidase
VHOC	volatile halogenated organic compound
VHPO	vanadium-dependent haloperoxidase

CHAPTER 1

INTRODUCTION

“Seaweeds” (or macroalgae) are macroscopic, multicellular marine algae that usually dwell on the coastal region of oceans (Lobban and Harrison, 1996; Khan *et al.*, 2009). They are photosynthetic but the absence of roots, leafy shoots, flowers, and vascular tissues distinguishes them from other marine plants (Diaz-Pulido and McCook, 2008). Generally, seaweeds are categorized into three groups, viz. red algae (Rhodophyte), green algae (Chlorophyte) and brown algae (Phaeophyte), primarily based on the composition of their photosynthetic pigments (Khan *et al.*, 2009).

Rhodophytes are characteristically red in colour, which is caused by the red pigment phycoerythrin. They are the major source of agar and carrageenans (Chapman, 2013; Lopez-Bautista, 2010). The second largest genus of Rhodophyta is *Gracilaria*. A total of 20 species of *Gracilaria* have been recorded for Malaysia, and one of them is *Gracilaria changii* (Yow *et al.*, 2011). *G. changii* is an edible seaweed that is rich in omega fatty acid and various other nutrients essential for health (Norziah and Ching, 2000). It is also the main source of high-quality agar and agarose with good gel strength (Armisen, 1995; Phang *et al.*, 1996). As the utilization for agar in various industries increases, expansion in the cultivation of *Gracilaria* is expected to meet the increasing demand. In light of the great commercial value of *Gracilaria*, diseases that threaten their cultivation are of paramount concern.

Seaweed is constantly being challenged by microorganisms such as viruses, bacteria, fungi and etc. (Potin *et al.*, 2002; Cosse *et al.*, 2008). Many bacteria that rely on seaweeds as a source of nutrients are able to enzymatically decompose seaweed cell wall. Their pathogenic attacks cause wounds and diseases on seaweed, generating symptoms such as rot, bleaching, lesion, and malformation (Weinberger, 2007). To overcome the attack of these pathogens, seaweeds have evolved a variety of defense mechanisms. Continuous synthesis of new cell wall and chemical deterrents are part of the defensive tactics of seaweeds against pathogens (Cosse *et al.*, 2008; Weinberg and Potin, 2010).

Besides constitutive defense mechanisms, recent studies reveal defense of seaweed that varies with time or environmental factors, indicating the existence of induced defense mechanisms (Weinberger and Potin, 2010). This mode of defense mechanism is initiated by recognition of pathogenic invasion through the perception of pathogenic origin elicitors or host endogenous elicitors such as oligosaccharides (Weinberger *et al.*, 1999). Endogenous elicitors of host are the cell wall fragments released during enzymatic attacks of pathogens (e.g. agarase). Upon recognition of the attack, induced defense mechanisms evoke a series of responses such as oxidative burst, emission of volatile halogenated organic compounds (VHOCs), production of oxylipins, synthesis of inducible secondary metabolites and etc. (Potin, 2008).

Information on the induced defense mechanisms of seaweed is scarce compared to terrestrial plants (Cosse *et al.*, 2008). The majority of investigations on seaweed defense mechanisms focused on seaweed-herbivore interactions, while the seaweed-microbe interactions have been neglected (Weinberger, 2007). Furthermore, the seaweed-pathogen interactions have hardly been studied beyond the phenomenology of infection; the regulation of defense at molecular level remains largely unexplored (Weinberg and Potin, 2010). Hence, there is an urgent need to investigate the gene regulation involved in the induced defense mechanisms of seaweed in response to pathogenic invasion. A set of differentially expressed candidates from *G. changii* in response to agarase were generated from a previous study through next generation sequencing of the seaweed transcriptomes (unpublished data). In this study, the expression of those candidates in *G. changii* upon both agarase and agarolytic bacteria treatment was analyzed with quantitative reverse-transcription real-time PCR (qRT-PCR).

The objectives of this study were:

- 1) To verify the gene expression of candidate defense genes from *Gracilaria changii* in response to agarase treatment.
- 2) To compare the gene expression of candidate genes in response to agarase and agarolytic bacteria treatment.
- 3) To profile the gene expression of selected genes in response to agarase and agarolytic bacteria treatment at different time points.

The results from this study may provide further understanding on the defense mechanisms of seaweed in response to cell-wall degrading enzymes and agarolytic bacteria invasion. This may in turn facilitate prevention and action against bacterial disease in seaweeds.

REFERENCES

- Adir, N. (2005). Elucidation of the molecular structures of components of the phycobilisome: reconstructing a giant. *Photosynthesis Research*, 85: 15-32.
- Aldea, M., Hamilton, J. G., Resti, J. P., Zangerl, A. R., Berenbaum, M. R., Frank, T. D. and DeLucia, E. H. (2006). Comparison of photosynthetic damage from arthropod herbivory and pathogen infection in understory hardwood saplings. *Oecologia*, 149: 221-232.
- Amsler, C. D. (2012). Chemical Ecology of Seaweeds. In C. Wiencke and K. Bischof. *Seaweed Biology: Novel Insights Into Ecophysiology, Ecology and Utilization* (pp. 177-188). Berlin Heidelberg: Springer.
- Apt, K. E., Hoffman, N. E. and Grossman, A. R. (1993). The gamma subunit of R-phycoerythrin and its possible mode of transport into the plastid of red algae. *Journal of Biological Chemistry*, 268: 16208-16215.
- Armisen, R. (1995). World-wide use and importance of *Gracilaria*. *Journal of Applied Phycology*, 7: 231-243.
- Armstrong, E., Yan, L., Boyd, K. G., Wright, P. C. and Burgess, J. G. (2001). The symbiotic role of marine microbes on living surfaces. *Hydrobiologia*, 461: 37-40.
- Auer, M., Gruber, C., Bellei, M., Pirker, K. F., Zamocky, M., Kroiss, D., Teufer, S. A., Hofbauer, S., Soudi, M., Battistuzzi, G., Furtmüller, G. F. and Obinger, C. (2013). A stable bacterial peroxidase with novel halogenating activity and an autocatalytically linked heme prosthetic group. *Journal of Biological Chemistry*, 288: 27181-27199.
- Austin, M. B. and Noel, J. P. (2003). The chalcone synthase superfamily of type III polyketide synthases. *Natural Product Reports*, 20: 79-110.
- Baharum, H., Chu, W. C., Teo, S. S., Ng, K. Y., Rahim, A. R. and Ho, C. L. (2013). Molecular cloning, homology modeling and site-directed mutagenesis of vanadium-dependent bromoperoxidase (GcVBPO1) from *Gracilaria changii* (Rhodophyta). *Phytochemistry*, 92: 49-59.
- Barbas, C. F., Burton, D. R., Scott, J. K. and Silverman, G. J. (2007). Quantitation of DNA and RNA. *Cold Spring Harbor Protocols*, 2007, pdb-ip47.
- Barsanti, L. and Gualtieri, P. (2005). *Algae: Anatomy, Biochemistry, and Biotechnology*. Florida: CRC press.
- Battistuzzi, G., Bellei, M., Bortolotti, C. A. and Sola, M. (2010). Redox properties of heme peroxidases. *Archives of Biochemistry and Biophysics*, 500: 21-36.
- Benes, V. and Castoldi, M. (2010). Expression profiling of microRNA using real-time quantitative PCR, how to use it and what is available. *Methods*, 50: 244-249.
- Berger, S., Benediktyová, Z., Matouš, K., Bonfig, K., Mueller, M. J., Nedbal, L. and Roitsch, T. (2007). Visualization of dynamics of plant-pathogen interaction by novel combination of chlorophyll fluorescence imaging and statistical analysis: differential effects of virulent and avirulent strains of *P. syringae* and of oxylipins on *A. thaliana*. *Journal of Experimental Botany*, 58: 797-806.

- Bessho, K. and Iwasa, Y. (2009). Heteromorphic and isomorphic alternations of generations in macroalgae as adaptations to a seasonal environment. *Evolutionary Ecology Research*, 11: 691-711.
- Bilgin, D. D., Zavala, J. A., Zhu, J. I. N., Clough, S. J., Ort, D. R. and DeLucia, E. H. (2010). Biotic stress globally downregulates photosynthesis genes. *Plant, Cell and Environment*, 33: 1597-1613.
- Bolwell, G. P. (1999). Role of active oxygen species and NO in plant defence responses. *Current Opinion in Plant Biology*, 2: 287-294.
- Borchardt, S. A., Allain, E. J., Michels, J. J., Stearns, G. W., Kelly, R. F. and McCoy, W. F. (2001). Reaction of acylated homoserine lactone bacterial signaling molecules with oxidized halogen antimicrobials. *Applied and Environmental Microbiology*, 67: 3174-3179.
- Bouarab, K., Adas, F., Gaquerel, E., Kloareg, B., Salaün, J. P. and Potin, P. (2004). The innate immunity of a marine red alga involves oxylipins from both the eicosanoid and octadecanoid pathways. *Plant Physiology*, 135: 1838-1848.
- Bouarab, K., Potin, P., Correa, J. and Kloareg, B. (1999). Sulfated oligosaccharides mediate the interaction between a marine red alga and its green algal pathogenic endophyte. *The Plant Cell*, 11: 1635-1650.
- Boursiac, Y. and Harper, J. F. (2007). The origin and function of calmodulin regulated Ca²⁺ pumps in plants. *Journal of Bioenergetics and Biomembranes*, 39: 409-414.
- Bozsó, Z., Maunoury, N., Szatmari, A., Mergaert, P., Ott, P. G., Zsíros, L. R., Szabó, E., Kondorosi, E. and Klement, Z. (2009). Transcriptome analysis of a bacterially induced basal and hypersensitive response of *Medicago truncatula*. *Plant Molecular Biology*, 70: 627-646.
- Buchholz, C. M., Krause, G., and Buck, B. H. (2012). Seaweed and Man. In C. Wiencke and K. Bischof. *Seaweed Biology: Novel Insights into Ecophysiology, Ecology and Utilization* (pp. 471-493). Berlin Heidelberg: Springer.
- Bustin, S. A., Benes, V., Garson, J. A., Hellems, J., Huggett, J., Kubista, M., Mueller, R., Nolan, T., Pfaffl, M. W., Shipley, G. L., Vandesompele, J. and Wittwer, C. T. (2009). The MIQE guidelines: minimum information for publication of quantitative real-time PCR experiments. *Clinical Chemistry*, 55: 611-622.
- Butler, A. and Carter-Franklin, J. N. (2004). The role of vanadium bromoperoxidase in the biosynthesis of halogenated marine natural products. *Natural Product Reports*, 21: 180-188.
- Castro, P. and Huber, M. (2007). *Marine Biology* (6th ed.). New York: McGraw-Hill Higher Education.
- Chan, C. X., Teo, S. S., Ho, C. L., Othman, R. Y. and Phang, S. M. (2004). Optimisation of RNA extraction from *Gracilaria changii* (Gracilariales, Rhodophyta). *Journal of Applied Phycology*, 16: 297-301.
- Chapman, R. L. (2013). Algae: the world's most important "plants"—an introduction. *Mitigation and Adaptation Strategies for Global Change*, 18: 5-12.

- Chapman, V. J. (1970). *Seaweeds and Their Uses* (2nd ed.). London: Methuen Co. Ltd.
- Cheong, J. J. and Choi, Y. D. (2003). Methyl jasmonate as a vital substance in plants. *Trends in Genetics*, 19: 409-413.
- Chi, W. J., Chang, Y. K. and Hong, S. K. (2012). Agar degradation by microorganisms and agar-degrading enzymes. *Applied Microbiology and Biotechnology*, 94: 917-930.
- Chou, L. S., Liu, C. S. J., Boese, B., Zhang, X., and Mao, R. (2010). DNA sequence capture and enrichment by microarray followed by next-generation sequencing for targeted resequencing: neurofibromatosis type 1 gene as a model. *Clinical Chemistry*, 56: 62-72.
- Cole, K. M. and Sheath, R. G. (1990). *Biology of The Red Algae*. Cambridge: Cambridge University Press.
- Collén, J., Hervé, C., Guisle-Marsollier, I., Léger, J. J. and Boyen, C. (2006). Expression profiling of *Chondrus crispus* (Rhodophyta) after exposure to methyl jasmonate. *Journal of Experimental Botany*, 57: 3869-3881.
- Collén, J., Porcel, B., Carré, W., Ball, S. G., Chaparro, C., Tonon, T., Barbeyron, T., Michel, G., Noel, B., Valentin, K., Elias, M., Artiguenave, F., Arun, A., Aury, J., Barbosa-Neto, J. F., Bothwell, J. H., Bouget, F., Brilllet, L., Cabello-Hurtado, F., Capella-Gutiérrez, S., Charrier, B., Cladière, L., Cock, J. M., Coelho S. M., Colleoni, C., Czjzek, M., Silva, C. D., Delage, L., Denoëud, F., Deschamps, P., Dittami, S. M., Gabaldón, T., Gachon, C. M. M., Groisillier, A., Hervé, C., Jabbari, K., Katinka, M., Kloareg, B., Kowalczyk, N., Labadie, K., Leblanc, C., Lopez, P. J., McLachlan, D. H., Meslet-Cladiere, L., Moustafa, A., Nehr, A., Collén, P. N., Panaud, O., Partensky, F., Poulain, J., Rensing, S. A., Rousvoal, S., Samson, G., Symeonidi, A., Weissenbach, J., Zambounis, A., Wincker, P. and Boyen, C. (2013). Genome structure and metabolic features in the red seaweed *Chondrus crispus* shed light on evolution of the Archaeplastida. *Proceedings of the National Academy of Sciences USA*, 110: 5247-5252.
- Cosse, A., Leblanc, C. and Potin, P. (2008). Dynamic defense of marine macroalgae against pathogens: from early activated to gene-regulated responses. *Advances in Botanical Research*, 46: 221-266.
- Cosse, A., Potin, P. and Leblanc, C. (2009). Patterns of gene expression induced by oligoguluronates reveal conserved and environment-specific molecular defense responses in the brown alga *Laminaria digitata*. *New Phytologist*, 182: 239-250.
- Coveney, M. F. and Wetzel, R. G. (1989). Bacterial metabolism of algal extracellular carbon. *Hydrobiologia*, 173: 141-149.
- Cramer, M. J., Haghshenas, N., Bagwell, C. E., Matsui, G. Y. and Lovell, C. R. (2011). *Celerinatantimonas diazotrophica* gen. nov., sp. nov., a nitrogen-fixing bacterium representing a new family in the *Gammaproteobacteria*, *Celerinatantimonadaceae* fam. nov. *International Journal of Systematic and Evolutionary Microbiology*, 61: 1053-1060.

- Danhorn, T. and Fuqua, C. (2007). Biofilm formation by plant-associated bacteria. *Annual Review of Microbiology*, 61: 401-422.
- De Tullio, M., Guether, M. and Balestrini, R. (2013). Ascorbate oxidase is the potential conductor of a symphony of signaling pathways. *Plant Signaling and Behavior*, 8: e23213.
- Denny, M. and Gaylord, B. (2002). The mechanics of wave-swept algae. *Journal of Experimental Biology*, 205: 1355-1362.
- Desikan, R., Hancock, J. T., Ichimura, K., Shinozaki, K. and Neill, S. J. (2001). Harpin induces activation of the Arabidopsis mitogen-activated protein kinases AtMPK4 and AtMPK6. *Plant Physiology*, 126: 1579-1587.
- Dhargalkar, V. K. and Pereira, N. (2005). Seaweed: promising plant of the millennium. *Science and Culture*, 71: 60-66.
- Diaz-Pulido, G. (2008). Macroalgae. In P. A. Hutchings, M. Kingsford and O. Hoegh-Guldberg. *The Great Barrier Reef: Biology, Environment and Management* (pp. 145-154). Collingwood: Csiro Publishing.
- Diaz-Pulido, G. and McCook, L. J. (2008). Environmental Status: Macroalgae (Seaweeds). In A. Chin. *The State of The Great Barrier Reef* (pp. 1-44). Townsville: Great Barrier Reef Marine Park Authority.
- Djami-Tchatchou, A. T. and Straker, C. J. (2012). The isolation of high quality RNA from the fruit of avocado (*Persea americana* Mill.). *South African Journal of Botany*, 78: 44-46.
- Dodd, A. N., Kudla, J. and Sanders, D. (2010). The language of calcium signaling. *Annual Review of Plant Biology*, 61: 593-620.
- Druehl, L. and Druehl, L. D. (2001). *Pacific Seaweeds: A Guide to Common Seaweeds of The West Coast*. Canada: Harbour Publishing.
- Du, Z. J., Lv, G. Q., Rooney, A. P., Miao, T. T., Xu, Q. Q. and Chen, G. J. (2011). *Agarivorans gilvus* sp. nov. isolated from seaweed. *International Journal of Systematic and Evolutionary Microbiology*, 61: 493-496.
- Egan, S., Fernandes, N. D., Kumar, V., Gardiner, M. and Thomas, T. (2013a). Bacterial pathogens, virulence mechanism and host defence in marine macroalgae. *Environmental Microbiology*, doi:10.1111/1462-2920.12288.
- Egan, S., Harder, T., Burke, C., Steinberg, P., Kjelleberg, S. and Thomas, T. (2013b). The seaweed holobiont: understanding seaweed–bacteria interactions. *FEMS Microbiology Reviews*, 37: 462-476.
- Fang, Z. and Cui, X. (2011). Design and validation issues in RNA-seq experiments. *Briefings in Bioinformatics*, 12: 280-287.
- Ferrari, S., Galletti, R., Denoux, C., De Lorenzo, G., Ausubel, F. M. and Dewdney, J. (2007). Resistance to *Botrytis cinerea* induced in Arabidopsis by elicitors is independent of salicylic acid, ethylene, or jasmonate signaling but requires PHYTOALEXIN DEFICIENT3. *Plant Physiology*, 144: 367-379.
- Farrell, R. E. (2010). *RNA Methodologies: A Laboratory Guide for Isolation and Characterization* (4th ed.). New York: Elsevier.

- Fleige, S. and Pfaffl, M. W. (2006). RNA integrity and the effect on the real-time qRT-PCR performance. *Molecular Aspects of Medicine*, 27: 126-139.
- Fleige, S., Walf, V., Huch, S., Prgomet, C., Sehm, J. and Pfaffl, M. W. (2006). Comparison of relative mRNA quantification models and the impact of RNA integrity in quantitative real-time RT-PCR. *Biotechnology Letters*, 28: 1601-1613.
- Fotopoulos, V., Sanmartin, M. and Kanellis, A. K. (2006). Effect of ascorbate oxidase over-expression on ascorbate recycling gene expression in response to agents imposing oxidative stress. *Journal of Experimental Botany*, 57: 3933-3943.
- Fu, X. T. and Kim, S. M. (2010). Agarase: review of major sources, categories, purification method, enzyme characteristics and applications. *Marine Drugs*, 8: 200-218.
- Funk, C. D. (2001). Prostaglandins and leukotrienes: advances in eicosanoid biology. *Science*, 294: 1871-1875.
- Gachon, C. M., Sime-Ngando, T., Strittmatter, M., Chambouvet, A. and Kim, G. H. (2010). Algal diseases: spotlight on a black box. *Trends in Plant Science*, 15: 633-640.
- Gallie, D. R. (2013). L-ascorbic acid: a multifunctional molecule supporting plant growth and development. *Scientifica*, 2013.
- García-Pineda, E., Castro-Mercado, E. and Lozoya-Gloria, E. (2004). Gene expression and enzyme activity of pepper (*Capsicum annuum* L.) ascorbate oxidase during elicitor and wounding stress. *Plant Science*, 166: 237-243.
- Gerwick, W. H. (1999). Eicosanoids in Nonmammals. In O. Meth-Cohn, S. D. Barton and K. Nakanishi. *Comprehensive Natural Products Chemistry Volume 1* (pp. 207-254). Oxford: Elsevier Science.
- Gerwick, W. H., Roberts, M. A., Vulpanovici, A. and Ballantine, D. L. (1999). Biogenesis and Biological Function of Marine Algal Oxylipins. In S. Nigam and C. R. Pace-Asciak. *Lipoxygenases and Their Metabolites* (pp. 211-218). New York: Kluwer Academic Plenum Publishers.
- Git, A., Dvinge, H., Salmon-Divon, M., Osborne, M., Kutter, C., Hadfield, J., Bertone, P. and Caldas, C. (2010). Systematic comparison of microarray profiling, real-time PCR, and next-generation sequencing technologies for measuring differential microRNA expression. *Rna*, 16: 991-1006.
- Goecke, F. R., Labes, A., Wiese, J. and Imhoff, J. F. (2010). Chemical interactions between marine macroalgae and bacteria. *Marine Ecology Progress Series*, 409: 267-299.
- Görlach, J., Raesecke, H. R., Rentsch, D., Regenass, M., Roy, P., Zala, M., Keel, C., Boller, T., Amrhein, N. and Schmid, J. (1995). Temporally distinct accumulation of transcripts encoding enzymes of the prechorismate pathway in elicitor-treated, cultured tomato cells. *Proceedings of the National Academy of Sciences USA*, 92: 3166-3170.

- Green, E. D., Birren, B., Klapholz, S., Myers, R. M. and Roskam, J. (1998). *Genome Analysis: A Laboratory Manual, Volume 2, Detecting Genes*. New York: Cold Spring Harbor Laboratory Press.
- Gurgel, C. F. D., Fredericq, S. (2004). Systematics of the *Gracilariaceae* (Gracilariales, Rhodophyta): a critical assessment based on rbcL sequence analyses. *Journal of Phycology*, 40: 138–159.
- Hall, T. A. (1999). BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symposium Series*, 41: 95-98.
- Hall-Stoodley, L., Costerton, J. W. and Stoodley, P. (2004). Bacterial biofilms: from the natural environment to infectious diseases. *Nature Reviews Microbiology*, 2: 95-108.
- Halliwell, B. and Gutteridge, J. M. (1999). *Free Radicals in Biology and Medicine* New York: Oxford University Press.
- Hallman, A. (2007). Algal transgenics and biotechnology. *Transgenic Plant Journal*, 1: 81-89.
- Hammerschmidt, R. (1999). Phytoalexins: what have we learned after 60 years?. *Annual Review of Phytopathology*, 37: 285-306.
- Harvell, C. D., Mitchell, C. E., Ward, J. R., Altizer, S., Dobson, A. P., Ostfeld, R. S. and Samuel, M. D. (2002). Ecology-climate warming and disease risks for terrestrial and marine biota. *Science*, 296: 2158–2162.
- Hervé, C., Tonon, T., Collén, J., Corre, E. and Boyen, C. (2006). NADPH oxidases in eukaryotes: red algae provide new hints!. *Current Genetics*, 49: 190-204.
- Hoffmann, M., Monday, S. R., Allard, M. W., Strain, E. A., Whittaker, P., Naum, M., McCarthy, P. J., Lopez, J. V., Fischer, M. and Brown, E. W. (2012). *Vibrio caribbeanicus* sp. nov., isolated from the marine sponge *Scleroderma cyanea*. *International Journal of Systematic and Evolutionary Microbiology*, 62: 1736-1743.
- Hollants, J., Leliaert, F., Clerck, O. and Willems, A. (2013). What we can learn from sushi: a review on seaweed–bacterial associations. *FEMS Microbiology Ecology*, 83: 1-16.
- Hosoya, S., Adachi, K. and Kasai, H. (2009). *Thalassomonas actiniarum* sp. nov. and *Thalassomonas haliotis* sp. nov., isolated from marine animals. *International Journal of Systematic and Evolutionary Microbiology*, 59: 686-690.
- Huggett, J., Dheda, K., Bustin, S. and Zumla, A. (2005). Real-time RT-PCR normalisation; strategies and considerations. *Genes and Immunity*, 6: 279-284.
- Hunter, T. (1995). Protein kinases and phosphatases: the yin and yang of protein phosphorylation and signaling. *Cell*, 80: 225-236.
- Iriti, M. and Faoro, F. (2009). Chemical diversity and defence metabolism: how plants cope with pathogens and ozone pollution. *International Journal of Molecular Sciences*, 10: 3371-3399.

- Ivanova, E. P., Romanenko, L. A., Matte, M. H., Matte, G. R., Lysenko, A. M., Simidu, U., Kita-Tsukamoto, K., Sawabe, T., Vysotskii, M. V., Frolova, G. M., Mikhailov, V., Christen, R. and Colwell, R. R. (2001). Retrieval of the species *Alteromonas tetraodonis* Simidu *et al.* 1990 as *Pseudoalteromonas tetraodonis* comb. nov. and emendation of description. *International Journal of Systematic and Evolutionary Microbiology*, 51: 1071-1078.
- Ivanova, E. P., Shevchenko, L. S., Sawabe, T., Lysenko, A. M., Svetashev, V. I., Gorshkova, N. M., Satomi, M., Christen, R. and Mikhailov, V. V. (2002). *Pseudoalteromonas maricaloris* sp. nov., isolated from an Australian sponge, and reclassification of [*Pseudoalteromonas aurantia*] NCIMB 2033 as *Pseudoalteromonas flavipulchra* sp. nov. *International Journal of Systematic and Evolutionary Microbiology*, 52: 263-271.
- Jaffray, A. E. and Coyne, V. E. (1996). Development of an in situ assay to detect bacterial pathogens of the red alga *Gracilaria gracilis* (Stackhouse) Steentoft, Irvine et Farnham. *Journal of Applied Phycology*, 8: 409-414.
- Janda, J. M. and Abbott, S. L. (2007). 16S rRNA gene sequencing for bacterial identification in the diagnostic laboratory: pluses, perils, and pitfalls. *Journal of Clinical Microbiology*, 45: 2761-2764.
- Jean, W. D., Huang, S. P., Liu, T. Y., Chen, J. S. and Shieh, W. Y. (2009). *Aliagarivorans marinus* gen. nov., sp. nov. and *Aliagarivorans taiwanensis* sp. nov., facultatively anaerobic marine bacteria capable of agar degradation. *International Journal of Systematic and Evolutionary Microbiology*, 59: 1880-1887.
- Jin, G., Wang, S., Yu, M., Yan, S. and Zhang, X. H. (2010). Identification of a marine antagonistic strain JG1 and establishment of a polymerase chain reaction detection technique based on the *gyrB* gene. *Aquaculture Research*, 41: 1867-1874.
- Jozefczuk, J. and Adjaye, J. (2011). Quantitative real-time PCR-based analysis of gene expression. *Methods in Enzymology*, 500: 99.
- Jukes, T. H. and Cantor, C. R. (1969). Evolution of Protein Molecules. In H. N. Munro. *Mammalian Protein Metabolism* (pp. 21-132). New York: Academic Press.
- Karleskint, G., Turner, R. and Small, J. (2009). *Introduction to Marine Biology* (3rd ed.). Belmont: Cengage Learning.
- Kerk, D., Conley, T. R., Rodriguez, F. A., Tran, H. T., Nimick, M., Muench, D. G. and Moorhead, G. B. (2006). A chloroplast-localized dual-specificity protein phosphatase in *Arabidopsis* contains a phylogenetically dispersed and ancient carbohydrate-binding domain, which binds the polysaccharide starch. *The Plant Journal*, 46: 400-413.
- Key, J., Marcus, M., Jeffries, W. B., Voris, H. K. and Yang, C. M. (1996). Epizoic bryozoans, horseshoe crabs, and other mobile benthic substrates. *Bulletin of Marine Science*, 58: 368-384.

- Khan, S. T. and Harayama, S. (2007). *Paraferrimonas sedimenticola* gen. nov., sp. nov., a marine bacterium of the family Ferrimonadaceae. *International Journal of Systematic and Evolutionary Microbiology*, 57: 1493-1498.
- Khan, W., Rayirath, U. P., Subramanian, S., Jithesh, M. N., Rayorath, P., Hodges, D. M., Critchley, A. T., Craigie, J. S., Norrie, J. and Prithiviraj, B. (2009). Seaweed extracts as biostimulants of plant growth and development. *Journal of Plant Growth Regulation*, 28: 386-399.
- Kim, J. H., Yu, J., Mahoney, N., Chan, K. L., Molyneux, R. J., Varga, J., Bhatnagar, D., Cleveland, T. E., Nierman, W. C. and Campbell, B. C. (2008). Elucidation of the functional genomics of antioxidant-based inhibition of aflatoxin biosynthesis. *International Journal of Food Microbiology*, 122: 49-60.
- Kim, S. H. and Hamada, T. (2005). Rapid and reliable method of extracting DNA and RNA from sweetpotato, *Ipomoea batatas* (L). Lam. *Biotechnology Letters*, 27: 1841-1845.
- Klein, D. (2002). Quantification using real-time PCR technology: applications and limitations. *Trends in Molecular Medicine*, 8: 257-260.
- Krieg, P. A. and Johnson, A. D. (1996). In Vitro Synthesis of mRNA. In P. A. Krieg. *A Laboratory Guide to RNA: Isolation, Analysis, and Synthesis* (pp. 141-154). New York: John Wiley and Sons.
- Küpper, F. C., Gaquerel, E., Boneberg, E. M., Morath, S., Salaün, J. P. and Potin, P. (2006). Early events in the perception of lipopolysaccharides in the brown alga *Laminaria digitata* include an oxidative burst and activation of fatty acid oxidation cascades. *Journal of Experimental Botany*, 57: 1991-1999.
- Küpper, F. C., Kloareg, B., Guern, J. and Potin, P. (2001). Oligoguluronates elicit an oxidative burst in the brown algal kelp *Laminaria digitata*. *Plant Physiology*, 125: 278-291.
- Kurahashi, M. and Yokota, A. (2004). *Agarivorans albus* gen. nov., sp. nov., a γ -proteobacterium isolated from marine animals. *International Journal of Systematic and Evolutionary Microbiology*, 54: 693-697.
- Lachnit, T., Blümel, M., Imhoff, J. F. and Wahl, M. (2009). Specific epibacterial communities on macroalgae: phylogeny matters more than habitat. *Aquatic Biology*, 5: 181-186.
- Lane, A. L. and Kubanek, J. (2008). Secondary Metabolite Defenses Against Pathogens and Biofoulers. In C. D. Amsler. *Algal Chemical Ecology* (pp. 229-243). Berlin Heidelberg: Springer.
- Lane, D. J. (1991). 16S/23S rRNA Sequencing. In E. Stackebrandt and M. Goodfellow. *Nucleic Acid Techniques in Bacterial Systematics* (pp. 115-175). New York: John Wiley and Sons.
- Lecourieux, D., Mazars, C., Pauly, N., Ranjeva, R. and Pugin, A. (2002). Analysis and effects of cytosolic free calcium increases in response to elicitors in *Nicotiana glauca* cells. *The Plant Cell*, 14: 2627-2641.
- Lee, R.E. (2008). *Phycology*. Cambridge: Cambridge University Press.

- Lengeler, J. W., Drews, G., and Schlegel, H. G. (1999). *Biology of the Prokaryotes*. Germany: Georg Thieme Verlag.
- Lewmanomont, K. A Review Paper on The Taxonomy of The *Gracilaria* in Asian Countries. In *Technical Sessions*. Regional Study and Workshop on the Taxonomy, Ecology and Processing of Economically Important Red Seaweeds, Bangkok, Thailand, January 24-27, 1995. Kongkeo H. (Ed.); Food and Agriculture Organization of the United Nations, Network of Aquaculture Centres in Asia-Pacific: Bangkok 1996.
- Lister, R., Chew, O., Lee, M. N., Heazlewood, J. L., Clifton, R., Parker, K. L., Millar, A. H. and Whelan, J. (2004). A transcriptomic and proteomic characterization of the Arabidopsis mitochondrial protein import apparatus and its response to mitochondrial dysfunction. *Plant Physiology*, 134: 777-789.
- Lion, U., Wiesemeier, T., Weinberger, F., Beltrán, J., Flores, V., Faugeron, S., Correa, J. and Pohnert, G. (2006). Phospholipases and galactolipases trigger oxylipin-mediated wound-activated defence in the red alga *Gracilaria chilensis* against epiphytes. *European Journal of Chemical Biology*, 7: 457-462.
- Liu, C., Yang, Z. and Tang, X. (2002). Generality of production of reactive oxygen species under infection of alginic acid decomposing bacteria in *Laminaria japonica*. *Mar. Fish. Res. Haiyang Shuichan Yanjiu*, 23: 33-36.
- Lobban, C. S. and Harrison, P. J. (1996). *Seaweed Ecology and Physiology*. Cambridge: Cambridge University Press.
- Longford, S. R., Tujula, N. A., Crocetti, G. R., Holmes, A. J., Holmström, C., Kjelleberg, S., Steinberg, P. D. and Taylor, M. W. (2007). Comparisons of diversity of bacterial communities associated with three sessile marine eukaryotes. *Aquatic Microbial Ecology*, 48: 217-229.
- Lopez-Bautista, J. M. (2010). Red Algal Genomics: A Synopsis. In J. Seckbach and D. J. Chapman. *Red Algae in the Genomic Age* (pp. 227-240). Berlin Heidelberg: Springer.
- Luan, S. (2003). Protein phosphatases in plants. *Annual Review of Plant Biology*, 54: 63-92.
- Lüning, K. and Pang, S. (2003). Mass cultivation of seaweeds: current aspects and approaches. *Journal of Applied Phycology*, 15: 115-119.
- Macián, M. C., Ludwig, W., Schleifer, K. H., Garay, E. and Pujalte, M. J. (2001). *Thalassomonas viridans* gen. nov., sp. nov., a novel marine gamma-proteobacterium. *International Journal of Systematic and Evolutionary Microbiology*, 51: 1283-1289.
- Mackey, D. and McFall, A. J. (2006). MAMPs and MIMPs: proposed classifications for inducers of innate immunity. *Molecular Microbiology*, 61: 1365-1371.
- Maeda, H. and Dudareva, N. (2012). The shikimate pathway and aromatic amino acid biosynthesis in plants. *Annual Review of Plant Biology*, 63: 73-105.

- Mashiguchi, K., Sasaki, E., Shimada, Y., Nagae, M., Ueno, K., Nakano, T., Yoneyama, K., Suzuki, Y. and Asami, T. (2009). Feedback-regulation of strigolactone biosynthetic genes and strigolactone-regulated genes in *Arabidopsis*. *Bioscience, Biotechnology, and Biochemistry*, 73: 2460-2465.
- Matsuyama, H., Minami, H., Kasahara, H., Kato, Y., Murayama, M. and Yumoto, I. (2013). *Pseudoalteromonas arabiensis* sp. nov., a marine polysaccharide-producing bacterium. *International Journal of Systematic and Evolutionary Microbiology*, 63: 1805-1809.
- Mazid, M., Khan, T. A. and Mohammad, F. (2011). Role of secondary metabolites in defense mechanisms of plants. *Biology and Medicine*, 3: 232-249.
- McHugh, D. J. (2003). *A Guide to The Seaweed Industry*. Rome: Food and Agriculture Organization of the United Nations.
- Meng, X. and Zhang, S. (2013). MAPK cascades in plant disease resistance signaling. *Annual Review of Phytopathology*, 51: 245-266.
- Milat, M. L., Ricci, P., Bonnet, P. and Blein, J. P. (1991). Capsidiol and ethylene production by tobacco cells in response to cryptogin, an elicitor from *Phytophthora cryptogea*. *Phytochemistry*, 30: 2171-2173.
- Mohapatra, P. K. (2008). *Textbook of Environmental Microbiology*. New Delhi: I.K. International Pvt. Ltd.
- Mtolera, M. S., Collén, J., Pedersén, M., Ekdahl, A., Abrahamsson, K. and Semesi, A. K. (1996). Stress-induced production of volatile halogenated organic compounds in *Eucheuma denticulatum* (Rhodophyta) caused by elevated pH and high light intensities. *European Journal of Phycology*, 31: 89-95.
- Muge, E., Burg, K., Kadu, C., Muchugi, A., Lemurt, S. and Jamnadass, R. (2009). Isolation of high quality DNA and RNA from cambium of the East African Greenheart (*Warburgia ugandensis*). *African Journal of Biotechnology*, 8: 3036-3040.
- Munn, C. B. (2011). *Marine Microbiology: Ecology and Applications* (2nd ed.). New York: Taylor and Francis.
- Najiah, M. and Lee, S. (2008). Bacteria attached on cultured seaweed *Gracilaria changii* at Mengabang Telipot, Terengganu. *Academic Journal of Plant Sciences*, 1: 1-4.
- Niu, J. F., Wang, G. C. and Tseng, C. K. (2006). Method for large-scale isolation and purification of R-phycoerythrin from red alga *Polysiphonia urceolata* Grev. *Protein Expression and Purification*, 49: 23-31.
- Norziah, M. H. and Ching, C. Y. (2000). Nutritional composition of edible seaweed *Gracilaria changgi*. *Food Chemistry*, 68: 69-76.
- Nürnberg, T., Brunner, F., Kemmerling, B. and Piater, L. (2004). Innate immunity in plants and animals: striking similarities and obvious differences. *Immunological Reviews*, 198: 249-266.
- Overbergh, L., Giulietti, A., Valckx, D. and Mathieu, C. (2010). Chapter 7 - Real-Time Polymerase Chain Reaction. In G. P. Patrinos and W. J. Ansoorge. *Molecular Diagnostics* (2nd ed.) (pp. 87-105). San Diego: Academic Press.

- Park, Y. D., Baik, K. S., Yi, H., Bae, K. S. and Chun, J. (2005). *Pseudoalteromonas byunsanensis* sp. nov., isolated from tidal flat sediment in Korea. *International Journal of Systematic and Evolutionary Microbiology*, 55(6): 2519-2523.
- Pelletreau, K. N. and Targett, N. M. (2008). New Perspectives for Addressing Patterns of Secondary Metabolites in Marine Macroalgae. In C. D. Amsler. *Algal Chemical Ecology* (pp. 121-146). Berlin Heidelberg: Springer.
- Peng, C., Hong-BO, S., Di, X. and Song, Q. (2009). Progress in *Gracilaria* biology and developmental utilization: main issues and prospective. *Reviews in Fisheries Science*, 17: 494-504.
- Perez, R. and Barbaroux, O. Cultivation and Uses of *Gracilaria*. In *Technical Sessions. Regional Study and Workshop on the Taxonomy, Ecology and Processing of Economically Important Red Seaweeds*, Bangkok, Thailand, January 24-27, 1995. Kongkeo H. (Ed.); Food and Agriculture Organization of the United Nations, Network of Aquaculture Centres in Asia-Pacific: Bangkok 1996.
- Petti, C. A. (2007). Detection and identification of microorganisms by gene amplification and sequencing. *Clinical Infectious Diseases*. 44: 1108–1114.
- Phang, S. M., Shaharuddin, S., Noraishah, H. and Sasekumar, A. (1996). Studies on *Gracilaria changii* (Gracilariales, Rhodophyta) from Malaysian mangroves. *Hydrobiologia*, 326: 347-352.
- Pignocchi, C. and Foyer, C. H. (2003). Apoplastic ascorbate metabolism and its role in the regulation of cell signalling. *Current Opinion in Plant Biology*, 6: 379-389.
- Pignocchi, C., Kiddle, G., Hernández, I., Foster, S. J., Asensi, A., Taybi, T., Barnes, J. and Foyer, C. H. (2006). Ascorbate oxidase-dependent changes in the redox state of the apoplast modulate gene transcript accumulation leading to modified hormone signaling and orchestration of defense processes in tobacco. *Plant Physiology*, 141: 423-435.
- Potin, P. (2008). Oxidative Burst and Related Responses in Biotic Interactions of Algae. In C. D. Amsler. *Algal Chemical Ecology* (pp. 245-271). Berlin Heidelberg: Springer.
- Potin, P. (2012). Intimate Associations Between Epiphytes, Endophytes, and Parasites of Seaweeds. In C. Wiencke, K. Bischof. *Seaweed Biology* (pp. 203-234). Berlin Heidelberg: Springer.
- Potin, P. and Leblanc, C. (2006). Phenolic-based Adhesives of Marine Brown Algae. In A. M. Smith and J. A. Callow. *Biological Adhesives* (pp. 105-124). Berlin Heidelberg: Springer.
- Potin, P., Bouarab, K., Salaiün, J. P., Pohnert, G. and Kloareg, B. (2002). Biotic interactions of marine algae. *Current Opinion in Plant Biology*, 5: 308-317.
- Prabhu, P. R. and Hudson, A. O. (2010). Identification and partial characterization of an L-tyrosine aminotransferase (TAT) from *Arabidopsis thaliana*. *Biochemistry Research International*, 2010, doi:10.1155/2010/549572.
- Prescott, L. M., Harley, J. P. and Klein, O. A. (2005). *Microbiology* (6th ed.). New York: Mc-Graw Hills.

- Pugin, A., Frachisse, J. M., Tavernier, E., Bligny, R., Gout, E., Douce, R. and Guern, J. (1997). Early events induced by the elicitor cryptogein in tobacco cells: involvement of a plasma membrane NADPH oxidase and activation of glycolysis and the pentose phosphate pathway. *The Plant Cell*, 9: 2077-2091.
- Radonić, A., Thulke, S., Mackay, I. M., Landt, O., Siegert, W. and Nitsche, A. (2004). Guideline to reference gene selection for quantitative real-time PCR. *Biochemical and Biophysical Research Communications*, 313: 856-862.
- Ranall, M. V., Butler, M. S., Blaskovich, M. A. and Cooper, M. A. (2012). Resolving biofilm infections: current therapy and drug discovery strategies. *Current Drug Targets*, 13: 1375-1385.
- Raugei, S. and Carloni, P. (2006). Structure and function of vanadium haloperoxidases. *The Journal of Physical Chemistry B*, 110: 3747-3758.
- Relman, D. A. (2008). 'Til death do us part': coming to terms with symbiotic relationships. *Nature Reviews Microbiology*, 6: 721-724.
- Riewe, D., Koochi, M., Lisec, J., Pfeiffer, M., Lippmann, R., Schmeichel, J., Willmitzer, L. and Altmann, T. (2012). A tyrosine aminotransferase involved in tocopherol synthesis in *Arabidopsis*. *The Plant Journal*, 71: 850-859.
- Rio, D. C., Ares, M., Hannon, G. J., and Nilsen, T. W. (2010). Nondenaturing agarose gel electrophoresis of RNA. *Cold Spring Harbor Protocols*, doi:10.1101/pdb.prot5445.
- Roeder, V., Collén, J., Rousvoal, S., Corre, E., Leblanc, C. and Boyen, C. (2005). Identification of stress gene transcripts in *Laminaria digitata* (Phaeopjyceae) protoplast cultures by expressed sequence tag analysis. *Journal of Phycology*, 41: 1227-1235.
- Rorrer, G. L. (2005). *Metabolic Engineering of Seaweeds for the Detoxification of TNT-Contaminated Marine Waters*. Oregon: Oregon State University, Department of Chemical Engineering.
- Ruhfel, B. R., Gitzendanner, M. A., Soltis, P. S., Soltis, D. E., and Burleigh, J. G. (2014). From algae to angiosperms - inferring the phylogeny of green plants (*Viridiplantae*) from 360 plastid genomes. *BMC Evolutionary Biology*, 14: 23.
- Saad, R. Malaysia. In *Country Reports. Regional Study and Workshop on the Taxonomy, Ecology and Processing of Economically Important Red Seaweeds*, Bangkok, Thailand, January 24-27, 1995. Kongkeo H. (Ed.); Food and Agriculture Organization of the United Nations, Network of Aquaculture Centres in Asia-Pacific: Bangkok 1996.
- Sajiki, J. and Kakimi, H. (1998). Identification of eicosanoids in the red algae, *Gracilaria asiatica*, using high-performance liquid chromatography and electrospray ionization mass spectrometry. *Journal of Chromatography A*, 795: 227-237.
- Sambrook, J., Russell, D. W. and Russell, D. W. (2001). *Molecular Cloning: A Laboratory Manual* (3rd ed.). New York: Cold Spring Harbor Laboratory Press.

- Sandy, M., Carter-Franklin, J. N., Martin, J. D. and Butler, A. (2011). Vanadium bromoperoxidase from *Delisea pulchra*: enzyme-catalyzed formation of bromofuranone and attendant disruption of quorum sensing. *Chemical Communications*, 47: 12086-12088.
- Schneider, I. and Bucar, F. (2005). Lipoxygenase inhibitors from natural plant sources. Part 1: Medicinal plants with inhibitory activity on arachidonate 5-lipoxygenase and 5-lipoxygenase cyclooxygenase. *Phytotherapy Research*, 19: 81-102.
- Schroeder, D. C., Jaffer, M. A. and Coyne, V. E. (2003). Investigation of the role of a β (1–4) agarase produced by *Pseudoalteromonas gracilis* B9 in eliciting disease symptoms in the red alga *Gracilaria gracilis*. *Microbiology*, 149: 2919-2929.
- Seal, S., Patel, M. V., Collins, C., Colvin, J. and Bailey, D. (2012). Next generation transcriptome sequencing and quantitative real-time PCR technologies for characterisation of the *Bemisia Asia* 1 mtCOI phylogenetic clade. *Journal of Integrative Agriculture*, 11: 281-292.
- Shanmughapriya, S., Manilal, A., Sujith, S., Selvin, J., Kiran, G. S. and Natarajaseenivasan, K. (2008). Antimicrobial activity of seaweeds extracts against multiresistant pathogens. *Annals of Microbiology*, 58: 535-541.
- Shetty, N. P., Lyngs Jørgensen, H. J., Jensen, J. D., Collinge, D. B. and Shekar Shetty, H. (2008). Roles of reactive oxygen species in interactions between plants and pathogens. *European Journal of Plant Pathology*, 121: 267-280.
- Shimizu, T., Satoh, K., Kikuchi, S. and Omura, T. (2007). The repression of cell wall-and plastid-related genes and the induction of defense-related genes in rice plants infected with rice dwarf virus. *Molecular Plant-Microbe Interactions*, 20: 247-254.
- Shindou, H., Hishikawa, D., Harayama, T., Yuki, K. and Shimizu, T. (2009). Recent progress on acyl CoA: lysophospholipid acyltransferase research. *Journal of Lipid Research*, 50(Supplement), S46-S51.
- Shipley, G. L. (2006). An Introduction to Real-Time PCR. In M. T. Dorak. *Real-Time PCR* (pp. 1-37). Abingdon: Taylor and Francis Group.
- Smirnoff, N. (2000). Ascorbic acid: metabolism and functions of a multi-faceted molecule. *Current Opinion in Plant Biology*, 3: 229-235.
- Stephenson, F. H. (2010). *Calculations for Molecular Biology and Biotechnology: A Guide to Mathematics in the Laboratory* (2nd ed.). Oxford: Academic press.
- Sun, L., Wang, S., Gong, X., Zhao, M., Fu, X. and Wang, L. (2009). Isolation, purification and characteristics of R-phycoerythrin from a marine macroalga *Heterosiphonia japonica*. *Protein Expression and Purification*, 64: 146-154.
- Surzycki, A. (2000). *Basic Techniques in Molecular Biology*. Berlin Heidelberg: Springer-Verlag.
- Suthiphongchai, T., Boonsiri, P. and Panijpan, B. (2008). Vanadium-dependent bromoperoxidases from *Gracilaria* algae. *Journal of Applied Phycology*, 20: 271-278.

- Tamura, K., Peterson, D. , Peterson, N. , Stecher, G. , Nei, M. and Kumar, S. (2011). MEGA5: molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. *Molecular Biology and Evolution*, 28: 2731-2739.
- Tanaka, M., Moriya, Y., Goto, S. and Kanehisa, M. (2010). Analysis of a lipid biosynthesis protein family and phospholipid structural variations. *Japanese Society for Bioinformatics*, 22: 191-201.
- Taylor, S., Wakem, M., Dijkman, G., Alsarraj, M. and Nguyen, M. (2010). A practical approach to RT-qPCR-publishing data that conform to the MIQE guidelines. *Methods*, 50: S1-S5.
- Trewavas, A. (2000). Signal Perception and Transduction. In B. B. Buchanan, W. Gruissem and R. L. Jones. *Biochemistry and molecular biology of plants* (pp. 930-987). Rockville: American Society of Plant Physiologists.
- Tuteja, N. and Mahajan, S. (2007). Calcium signaling network in plants: an overview. *Plant Signaling and Behavior*, 2: 79-85.
- Tzin, V. and Galili, G. (2010). New insights into the shikimate and aromatic amino acids biosynthesis pathways in plants. *Molecular Plant*, 3: 956-972.
- Vadassery, J. and Oelmüller, R. (2009). Calcium signaling in pathogenic and beneficial plant microbe interactions: what can we learn from the interaction between *Piriformospora indica* and *Arabidopsis thaliana*. *Plant Signaling and Behavior*, 4: 1024-1027.
- Valasek, M. A. and Repa, J. J. (2005). The power of real-time PCR. *Advances in Physiology Education*, 29: 151-159.
- Vandesompele, J., De Preter, K., Pattyn, F., Poppe, B., Van Roy, N., De Paepe, A. and Speleman, F. (2002). Accurate normalization of real-time quantitative RT-PCR data by geometric averaging of multiple internal control genes. *Genome Biology*, 3: research0034.
- VanGuilder, H. D., Vrana, K. E. and Freeman, W. M. (2008). Twenty-five years of quantitative PCR for gene expression analysis. *Biotechniques*, 44: 619-626.
- Voelkerding, K. V., Dames, S. A. and Durtschi, J. D. (2009). Next-generation sequencing: from basic research to diagnostics. *Clinical Chemistry*, 55: 641-658.
- Weber, H. (2002). Fatty acid-derived signals in plants. *Trends in Plant Science*, 7: 217-224.
- Weinberger, F. (1999). *Epiphyte-Host Interactions: Gracilaria conferta (Rhodophyta) and Associated Bacteria*. Doctoral dissertation, Christian-Albrechts-Universität zu Kiel, Germany.
- Weinberger, F. (2007). Pathogen-induced defense and innate immunity in macroalgae. *The Biological Bulletin*, 213: 290-302.
- Weinberger, F. and Friedlander, M. (2000a). Endogenous and exogenous elicitors of a hypersensitive response in *Gracilaria conferta* (Rhodophyta). *Journal of Applied Phycology*, 12: 139-145.

- Weinberger, F. and Friedlander, M. (2000b). Response of *Gracilaria conferta* (Rhodophyta) to oligoagars results in defense against agar-degrading epiphytes. *Journal of Phycology*, 36: 1079-1086.
- Weinberger, F. and Potin, P. (2010). Red Algal Defenses in The Genomics Age. In J. Seckbach and D. Chapman. *Red Algae in the Genomics Age* (pp. 457-477). Berlin Heidelberg: Springer.
- Weinberger, F., Coquemot, B., Forner, S., Morin, P., Kloareg, B. and Potin, P. (2007). Different regulation of haloperoxidation during agar oligosaccharide-activated defence mechanisms in two related red algae, *Gracilaria* sp. and *Gracilaria chilensis*. *Journal of Experimental Botany*, 58: 4365-4372.
- Weinberger, F., Friedlander, M. and Hoppe, H. G. (1999). Oligoagars elicit a physiological response in *Gracilaria conferta* (Rhodophyta). *Journal of Phycology*, 35: 747-755.
- Weinberger, F., Guillemin, M. L., Destombe, C., Valero, M., Faugeron, S., Correa, J. A., Pohnert, G., Pehlke, C., Kloareg, B. and Potin, P. (2010). Defense evolution in the Gracilariaceae (Rhodophyta): substrate-regulated oxidation of agar oligosaccharides is more ancient than the oligoagar-activated oxidative burst. *Journal of Phycology*, 46: 958-968.
- Weinberger, F., Hoppe, H. G. and Friedlander, M. (1997). Bacterial induction and inhibition of a fast necrotic response in *Gracilaria conferta* (Rhodophyta). *Journal of Applied Phycology*, 9: 277-285.
- Weinberger, F., Leonardi, P., Miravalles, A., Correa, J. A., Lion, U., Kloareg, B. and Potin, P. (2005). Dissection of two distinct defense-related response to agar oligosaccharides in *Gracilaria chilensis* (Rhodophyta) and *Gracilaria conferta* (Rhodophyta). *Journal of Phycology*, 41: 863-873.
- Weinberger, F., Richard, C., Kloareg, B., Kashman, Y., Hoppe, H. G. and Friedlander, M. (2001). Structure-activity relationships of oligoagar elicitors toward *Gracilaria conferta* (Rhodophyta). *Journal of Phycology*, 37: 418-426.
- Wever, R. (2012). Structure and Function of Vanadium Haloperoxidases. In H. Michibata. *Vanadium: Biochemical and Molecular Biological Approaches* (pp. 95-125). Berlin Heidelberg: Springer.
- Wu, Y. H., Shen, Y. Q., Xu, X. W., Wang, C. S., Oren, A. and Wu, M. (2009). *Pseudidiomarina donghaiensis* sp. nov. and *Pseudidiomarina maritima* sp. nov., isolated from the East China Sea. *International Journal of Systematic and Evolutionary Microbiology*, 59: 1321-1325.
- Xia, B. M. and Abbott, I. A. (1987). New species of *Polycarvernosa* Chang and Xia (Gracilariaceae, Rhodophyta) from the western Pacific. *Phycologia*, 26: 405-418.
- Xu, X. W., Wu, Y. H., Wang, C. S., Gao, X. H., Wang, X. G. and Wu, M. (2010). *Pseudoalteromonas lipolytica* sp. nov., isolated from the Yangtze River estuary. *International Journal of Systematic and Evolutionary Microbiology*, 60: 2176-2181.

- Yang, C., Guo, R., Jie, F., Nettleton, D., Peng, J., Carr, T., Yeakley, J. M., Fan, J. B., and Whitham, S. A. (2007). Spatial analysis of *Arabidopsis thaliana* gene expression in response to Turnip mosaic virus infection. *Molecular Plant-Microbe Interactions*, 20: 358-370.
- Yang, E. C., Kim, M. S., Geraldino, P. J. L., Sahoo, D., Shin, J. A. and Boo, S. M. (2008). Mitochondrial *cox1* and plastid *rbcL* genes of *Gracilaria vermiculophylla* (Gracilariaceae, Rhodophyta). *Journal of Applied Phycology*, 20: 161-168.
- Yeoh, K. A., Othman, A., Meon, S., Abdullah, F., and Ho, C. L. (2012). Sequence analysis and gene expression of putative exo- and endo-glucanases from oil palm (*Elaeis guineensis*) during fungal infection. *Journal of Plant Physiology*, 169: 1565-1570.
- Yoon, H. S., Zuccarello, G. C. and Bhattacharya, D. (2010). Evolutionary History and Taxonomy of Red Algae. In J. Seckbach, D. J. Chapman, A. Weber. *Red Algae in the Genomic Age* (pp. 25-42). New York: Springer.
- Yoon, H.S., Müller, K.M., Sheath, R.G., Ott, F.D. and Bhattacharya, D. (2006). Defining the major lineages of red algae (Rhodophyta). *Journal of Phycology*, 42: 482-492.
- Yoshimoto, T., Furukawa, M., Yamamoto, S., Horie, T. and Watanabe-Kohno, S. (1983). Flavonoids: potent inhibitors of arachidonate 5-lipoxygenase. *Biochemical and Biophysical Research Communications*, 116: 612-618.
- Yow, Y. Y., Lim, P. E. and Phang, S. M. (2011). Genetic diversity of *Gracilaria changii* (Gracilariaceae, Rhodophyta) from west coast, Peninsular Malaysia based on mitochondrial *cox1* gene analysis. *Journal of Applied Phycology*, 23: 219-226.
- Zámocký, M. and Obinger, C. (2010). Molecular Phylogeny of Heme Peroxidases. In E. Torres and M. Ayala. *Biocatalysis Based on Heme Peroxidases* (pp. 7-35). Berlin Heidelberg: Springer.
- Zhang, R., Chan, D., Jessica, S., Iskander, G., Black, D. S. and Kumar, N. (2009). Synthesis of new aryl substituted 5-alkylidenefuran-2 (5H)-ones. *ARKIVOC*, 5: 102-115.
- Zhang, S. and Klessig, D. F. (2001). MAPK cascades in plant defense signaling. *Trends in Plant Science*, 6: 520-527.
- Zhou, X., Ren, L., Li, Y., Zhang, M., Yu, Y. and Yu, J. (2010). The next-generation sequencing technology: a technology review and future perspective. *Science China Life Sciences*, 53: 44-57.
- Zou, J., Rodriguez-Zas, S., Aldea, M., Li, M., Zhu, J., Gonzalez, D. O., Vodkin, L. O., DeLucia, E. and Clough, S. J. (2005). Expression profiling soybean response to *Pseudomonas syringae* reveals new defense-related genes and rapid HR-specific downregulation of photosynthesis. *Molecular Plant-Microbe Interactions*, 18: 1161-1174.