

UNIVERSITI PUTRA MALAYSIA

BIOCONVERSION OF RICE STRAW INTO ACETONE-BUTANOL-ETHANOL BY Clostridium sporogenes A3

NURUL ATIKA BINTI MOHAMAD REMLI

FBSB 2014 20



BIOCONVERSION OF RICE STRAW INTO ACETONE-BUTANOL-ETHANOL BY Clostridium sporogenes A3



NURUL ATIKA BINTI MOHAMAD REMLI

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

January 2014

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia





DEDICATED TO MY MOTHER, HUSBAND, SISTERS AND BROTHERS

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the Degree of Master of Science

BIOCONVERSION OF RICE STRAW INTO ACETONE-BUTANOL-ETHANOL BY Clostridium sporogenes A3

By

NURUL ATIKA BINTI MOHAMAD REMLI

January 2014

Chairman : Associate Professor Umi Kalsom Md Shah, PhD

Faculty : Biotechnology and Biomolecular Sciences

The inevitable depletion of the world's energy supply and unstable oil market have renewed the interest of society in searching for alternative fuels. In addition to depletion of petroleum fuels, environmental issues like greenhouse effect, global warming and climate change, are also the issues to be resolved worldwide. Biobutanol has been considered as a suitable alternative to be a source of fuel. Abundant biomass from various agriculture sectors could be a source for biobutanol production. Rice straw is one of the most abundant lignocellulosic biomass that has a great potential as a cheap and affordable substrate for the production of reducing sugars and biofuel such as biobutanol. The feasibility of rice straw as a source of sugar production was evaluated in this study. The effectiveness of alkaline pretreatment on rice straw was assessed taking into consideration the yield of reducing sugars and changes in the morphological and chemical composition of rice straw. Pretreatment of rice straw by 2% (w/v) NaOH and KOH with autoclaving at 121°C, 15 psi (10 min) could be promising pretreatment methods for sugar production. Alkaline pretreatments with 2% (w/v) KOH and NaOH

followed by thermal pretreatment at 121°C, 15 psi (20 min) resulted in 58.5 to 64.5% higher conversion rate of reducing sugars production than untreated rice straw showing that alkaline pretreatments were effective even when a higher temperature was used. FTIR and SEM investigations showed that alkaline pretreatments caused chemical and morphological changes in the rice straw. The peaks of the cellulose and lignin materials were decreased after alkaline pretreatment, indicating that some cellulose and lignin were degraded. The reducing sugars obtained were then converted to acetone-butanolethanol (ABE) by *Clostridium sporogenes* A3. The total acetone-butanol-ethanol (ABE) production by locally isolated C. sporogenes A3 using rice straw hydrolysate was 1.58±0.11 g/L in which 0.73±0.05 g/L was butanol after 120 h of fermentation. Higher ABE yield was obtained from rice straw hydrolysate when compared to using commercial glucose as a carbon source. Increasing concentration of sugars in the rice straw hydrolysate to 40 g/L did not showed any improvements to the total ABE obtained. A higher level of ABE was obtained (1.72±0.39 g/L) at initial culture pH 5.5 (37°C), in which 0.93±0.22 g/L was butanol. In comparison, higher yield of ABE was obtained when using C. acetobutylicum ATCC824. The ABE yield obtained was 0.33 which corresponds to 64% of ABE increment compared to fermentation using C. sporogenes A3. These results suggested that the reducing sugars obtained from pretreated rice straw could be used as a substrate for ABE fermentation by C. sporogenes A3 and C. acetobutylicum ATCC824. This will reduce carbon emission and our dependency on fossil fuel, and at the same time makes butanol as one of our future energy for many applications.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

PENUKARAN JERAMI PADI KEPADA ASETON-BUTANOL-ETANOL OLEH Clostridium sporogenes A3

Oleh

NURUL ATIKA BINTI MOHAMAD REMLI

Januari 2013

Pengerusi : Prof. Madya Umi Kalsom Md Shah, PhD

Fakulti : Bioteknologi dan Sains Biomolekul

Pengurangan bekalan tenaga dunia yang tidak dapat dielakkan dan ketidakstabilan pasaran minyak telah mendorong masyarakat mencari bahan api alternatif. Di samping kekurangan bahan api petroleum, isu-isu alam sekitar seperti kesan rumah hijau, pemanasan global dan perubahan iklim, adalah isu-isu yang perlu diselesaikan di seluruh dunia. Biobutanol dianggap sebagai alternatif yang lebih sesuai untuk menjadi bahan api. Biomas yang terhasil daripada pelbagai sektor pertanian boleh menjadi sumber pengeluaran biobutanol. Jerami padi merupakan salah satu daripada biomas yang mengandungi banyak lignoselulosa dan mempunyai potensi yang besar sebagai substrat yang murah dan berpatutan untuk pengeluaran gula penurun dan biofuel seperti biobutanol. Kebolehgunaan jerami padi untuk pengeluaran gula telah dikenalpasti dalam kajian ini. Keberkesanan prarawatan beralkali pada jerami padi telah dinilai dengan mengambil kira hasil gula penurun dan perubahan dalam komposisi morfologi dan kimia dalam jerami padi. Prarawatan jerami padi dengan 2% (w/v) NaOH dan KOH beserta autoklaf pada suhu 121°C, 15 psi (10 min) boleh menjadi kaedah rawatan awal yang

boleh menghasilkan gula. Prarawatan jerami padi dengan alkali 2% (w/v) KOH dan NaOH diikuti oleh prarawatan haba pada suhu 121°C, 15 psi (20 min) telah meningkatkan kadar penukaran gula penurun sebanyak 58.5 hingga 64.5% berbanding jerami padi yang tidak dirawat. Ini menunjukkan bahawa prarawatan alkali adalah berkesan walaupun suhu yang lebih tinggi telah digunakan. Kajian FTIR dan SEM menunjukkan bahawa prarawatan alkali menyebabkan perubahan kimia dan morfologi dalam jerami padi. Puncak selulosa dan lignin telah menurun selepas prarawatan alkali, menunjukkan bahawa beberapa selulosa dan lignin telah dihuraikan. Gula penurun yang diperolehi kemudiannya ditukarkan menjadi aseton-butanol-etanol (ABE) oleh Clostridium sporogenes A3. Jumlah penghasilan aseton-butanol-etanol jumlah (ABE) oleh C. sporogenes A3 menggunakan hidrolisat jerami padi adalah sebanyak 1.58 ± 0.11 g/L di mana 0.73 \pm 0.05 g/L adalah butanol selepas 120 jam penapaian. Penghasilan ABE daripada hidrolisat jerami padi adalah lebih tinggi jika dibandingkan dengan menggunakan glukosa komersial sebagai sumber karbon. Peningkatan kepekatan gula dalam hidrolisat jerami padi hingga 40 g / L tidak menunjukkan apa-apa peningkatan kepada jumlah ABE yang diperoleh. Penghasilan ABE yang lebih tinggi telah diperolehi $(1.72 \pm 0.39 \text{ g/L})$ pada pH awal kultur 5.5 (37°C), di mana 0.93 \pm 0.22 g/L adalah butanol. Sebagai perbandingan, ABE yang diperoleh adalah lebih tinggi apabila menggunakan C. acetobutylicum ATCC824. Hasil ABE yang diperolehi ialah 0.33 bersamaan dengan kenaikan sebanyak 64% berbanding fermentasi menggunakan C. sporogenes A3. Kajian ini menunjukkan bahawa gula penurun yang diperoleh daripada jerami padi yang telah dirawat dapat digunakan sebagai substrat untuk penghasilan ABE oleh C. sporogenes A3 dan C. acetobutylicum ATCC824. Ini akan mengurangkan pelepasan karbon dan pergantungan kepada bahan api fosil, dan pada masa yang sama

menjadikan butanol sebagai salah satu bahan api untuk pelbagai aplikasi pada masa depan.



ACKNOWLEDGEMENTS

First of all, I would like to thank Allah Almighty, for his blessing and grace, for me to complete this research project at the Faculty of Biotechnology and Biomolecular Science, Universiti Putra Malaysia, Serdang.

I would like to express my sincere appreciation to my main supervisor, Assoc. Prof. Dr. Umi Kalsom Md Shah for giving her advice, motivation and support throughout this research. My sincere gratitude and appreciation also to my co-supervisors, Prof. Dr. Suraini Abd. Aziz and Assoc Prof. Dr. Rosfarizan Mohamad, for their valuable discussion and assistance throughout my research.

I also would like to express my sincere thanks to all Biomass Technology Center and MPP laboratory staffs such as Mr Rosli Aslim, Madam Renuga A/P Panjamurti, Madam Aluyah Marzuki and Madam Azlina for their generous guidance and help since I started my work.

Finally, my deepest appreciation to my beloved husband, Mohd Shaifuddin and my family for their love, patience and encouragement throughout this work. Acknowledgement is also to those who are involved directly and indirectly in the completion of this study.



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Umi Kalsom Md Shah, PhD

Associate Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Chairman)

Suraini Abd. Aziz, PhD

Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Member)

Rosfarizan Mohamad, PhD

Associate Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

Declaration by Graduate Student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

C'	D.4. 15 L	
Signature:	Date: 15 January 2014	
~		-

Name and Matric No.: Nurul Atika binti Mohamad Remli (GS29021)

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature:	Signature:
Name of	Name of
Chairman of	Member of
Supervisory	Supervisory
Committee:	Committee:
Signature:	Signature:
Name of	Name of
Chairman of	Member of
Supervisory	Supervisory
Committee:	Committee:

TABLE OF CONTENTS

			Page
DEDICATIO ABSTRACT ABSTRAK ACKNOWI APPROVAI DECLARAT LIST OF TA	ON F LEDGE L FION ABLES	EMENTS	iii iv vi ix x xii xvii
LIST OF FI	BBREV	VIATIONS	xviii xx
CHAPTER			
1	INT	RODUCTION	1
2	LITI	ERATURE REVIEW	
	2.1	Lignocellulosic biomass	3
		2.1.1 Structure of lignocellulosic biomass	4
	2.2	Paddy rice	6
		2.2.1 Rice industry in Malaysia	7
		2.2.2 Rice by-products	7
		2.2.3 Rice straw as a substrate for reducing sugars production	9
	2.3	Pretreatment methods for lignocellulosic biomass	9
		2.3.1 Physical pretreatment methods	10
		2.3.2 Physio-chemical pretreatment	11
		2.3.3 Chemical pretreatment	11
		2.3.4 Biological pretreatment	12
	2.4	Enzymatic hydrolysis	12
		2.4.1 Cellulolytic enzyme	13
		2.4.2 Mode of action of cellulolytic enzymes	14
	2.5	Acetone-Butanol-Ethanol (ABE) fermentation	16
		2.5.1 Acidogenic pathway	16
		2.5.2 Solventogenic pathway	17
		2.5.3 ABE producing microorganisms	19
		2.5.4 Substrate for ABE fermentation	19
		2.5.5 Application of acetone-butanol-ethanol	20
	2.6	Concluding remarks	21
3	GEN	ERAL MATERIALS AND METHODS	
	3.1	Experimental design	23
	3.2	Chemicals	24
	3.3	Reagents and solution preparation	24

	3.3.1	Sodium citrate buffer	24
	3.3.2	Dinitrosalicyclic acid solution	24
	3.3.3	ρ-nitrophenyl-β-D-glucopyranoside solution	25
	3.3.4	Potassium sodium tartrate solution	25
	3.3.5	Sodium carbonate solution	25
3.4	Pretrea	atment and preparation of rice straw	25
3.5	Saccha	arification of rice straw	25
3.6	Isolati	on of biobutanol-producing microorganism	25
3.7	ABE p	producing microorganism and strain maintenance	26
3.8	Prepar	ation of inoculum medium	26
3.9	Prepar	ation of production medium	26
3.10	Analy	tical procedures	27
	3.10.1	Chemical composition of rice straw	27
	3.10.2	Filter paper unit assay	29
	3.10.3	β-glucosidase assay	29
	3.10.4	Reducing sugar analysis	30
	3.10.5	Monomeric sugars determination using HPLC	30
	3.10.6	Determination of cell morphology by gram staining	; 30
		method	
	3.10.7	Determination of cell density	30
	3.10.8	Determination of ABE and acids concentration	31
		using Gas Chromatography (GC) Analysis	
	3.10.9	Statistical analysis	31

EFFECTS OF CHEMICAL AND THERMAL PRETREATMENTS ON THE ENZYMATIC SACCHARIFICATION OF RICE STRAW FOR SUGARS PRODUCTION

4

4.1	Introdu	uction	32
4.2	Materials and methods		
	4.2.1	Substrate preparation and pretreatments	33
	4.2.2	Effects of alkaline pretreatments at various	33
		temperatures on reducing sugar production	
	4.2.3	Effects of different type of alkalis and their	34
		concentration on reducing sugar production	
	4.2.4	Effect of the addition of β -glucosidase on	34
		reducing sugar production	
	4.2.5	Saccharification of rice straw	34
	4.2.6	Analytical methods	34
4.3	Result	s and Discussion	36
	4.3.1	Effects of alkaline pretreatments at various	36
		temperatures on reducing sugars production	
	4.3.2	Effects of different alkaline concentrations	37
		during pretreatments on reducing sugar production	
	4.3.3	Effects of the addition of β -glucosidase on	39
		reducing sugar production	

		4.3.4 Characterization of untreated and treated	l rice
		40straw	
	4.4	Conclusion	47
5	PROI FROM	DUCTION OF ACETONE-BUTANOL-ETHANOL M RICE STRAW HYDROLYSATE	(ABE)
	5.1	Introduction	48
	5.2	Materials and methods	49
		5.2.1 Source and isolation of biobutanol-producing bacteria	49
		5.2.2 Transmission Electron Microscopy	49
		5.2.3 16S rDNA sequencing and phylogenetic analysis	49
		5.2.4 Acetone-Butanol-Ethanol production using rice straw hydrolysate	49
		5.2.5 Analytical methods	50
	5.3	Results and Discussion	51
		5.3.1 Screening and isolation of bacteria	51
		5.3.2 Identification of bacteria	54
		5.3.3 Production of acetone-butanol-ethanol by <i>Clostridium sporogenes</i> A3	56
	5.4	Conclusion	67
6	CON	CLUSION AND RECOMMENDATIONS FOR FU	J TURE
	WOR	K	
	6.1	Summary	68
	6.2	Conclusion	69
	6.3	Recommendations for future work	69
REFERENC	CES		70
APPENDIC	ES		84
BIODATA (OF STU	DENT	99
LIST OF PU	BLICA	TIONS	100

G

LIST OF TABLES

Ta	able		Page
2	2.1	Composition of lignocellulose in several sources based on dry basis	5
2	2.2	Production quantity of paddy and rice straw	7
2	2.3	Properties of butanol, gasoline, ethanol and methanol	21
3	3.1	Preparation of sodium citrate buffer	24
3	3.2	Composition of Reinforced Clostridial Medium (RCM)	26
3	3.3	Composition of Tryptone-Yeast extract-Acetate (TYA) Medium	26
4	4.1	Total reducing sugar from saccharification of rice straw pretreated with 2% (w/v) alkaline solutions at three different temperatures with an enzyme loading of 10 FPU of cellulase, pH 4.8 at 50° C	37
4	1.2	Total Reducing Sugar Production after Saccharification of Rice Straw Pretreated with 2% and 5% (w/v) of Alkaline Solutions at 121°C, 15 psi with an Enzyme Loading of 10 FPU of Cellulase, pH 4.8 at 50 °C	38
4	1.3	Chemical compositions of the Lignocellulose Components of Rice Straw after pretreatment with various chemicals with autoclaving	41
5	5.1	Bacterial isolates from different sources	52
5	5.2	ABE production from rice straw using different pretreatment technologies.	60
5	5.3	Comparison of ABE fermentation using rice straw hydrolysate by <i>C.sporogenes</i> A3 and <i>C.acetobutylicum</i> ATCC824	61
(C) 5	5.4	Performance of ABE fermentation by <i>C. sporogenes</i> A3 at using different RSH concentration	63
5	5.5	Performance of ABE fermentation by <i>C.sporogenes</i> A3 on rice straw hydrolysate at different initial pH	64
5	5.6	Performance of ABE fermentation by <i>C.sporogenes</i> A3 on rice straw hydrolysate at different temperatures	66

LIST OF FIGURES

Figure		Page
2.1	Cellulose structure made up of from glucose monomers	4
2.2	Chemical structure of hemicelluloses	5
2.3	Chemical structure of lignin	6
2.4	Schematic goals of pretreatment on lignocellulosic material	9
2.5	Schematic of the mechanism for an enzyme binding with a substrate	13
2.6	Schematic Representation of Amorphogenesis of Cellulose	15
2.7	Fermentation pathways employed by <i>C. acetobutylicum</i>	18
3.1	General layout of experimental work	23
4.1	The effects of different NaOH and KOH concentrations ranging from 0.5 to 5.0% (w/v) on the performance of saccharification of rice straw using 10 FPU of Celluclast 1.5L, at pH 4.8 and 50°C	39
4.2	The effects of addition of β -glucosidase (Novozyme) into cellulase enzyme preparation with low level of β -glucosidase (Celluclast) on the performance of saccharification of rice straw pretreated with different types of alkaline solutions at 121°C, 15 psi	40
4.3	Chemical changes in rice straw determined by FTIR wavelength ranged from 750 to 4000 (cm-1)	43
4.5	Scanning electron microscope images of rice straw (3000×)	45-46
5.1	Electron micrographs depicting cell morphology and flagella	54
5.2	Neighbour-joining inferred tree based on 16S rDNA gene sequences showing the evolutionary relationship of isolates within previously characterized species	55
5.3	Growth cycle of <i>C.sporogenes</i> A3 in a batch fermentation using commercial glucose	56
5.4	Products produced by <i>C.sporogenes</i> A3 in batch ABE fermentation using glucose as a carbon source	57

5.5 Batch ABE fermentation of *C.sporogenes* A3 using rice straw 58 hydrolysate as carbon source



LIST OF ABBREVIATIONS

ABE	Acetone-butanol-ethanol
ADF	Acid detergent fiber
ADL	Acid detergent lignin
AFEX	Ammonia fiber explosion
ATCC	American Type Culture Collection
ATR	Attenuate total reflectance
bp	boiling point
CaC1.2H ₂ O	Calcium chloride dehydrate
СВМ	Carbohydrate binding module
CoC1 ₂	Cobalt chloride
DNA	Deoxyribonucleic acid
DNS	Dinitrosalicyclic acid
FAO	Food and agriculture organization
FAOSTAT	The statistics division of food and agriculture organization
FeSO ₄	Ferrous sulfate
FeSO ₄ .7H ₂ O	Ferrous sulfate heptahydrate
FID	Flame ionization detector
FPU	Filter paper unit
FPU/mL	Filter paper unit per milliliter
FTIR	Fourier Transform Infrared
g/g	Gram per gram substrate
g/L	Gram per liter
g/L/h	Gram per liter per hour
GNI	Gross national income
H_2SO_4	Sulphuric acid

HCl	Hydrochloric acid
HPLC	High performance liquid chromatography
IU	International unit
LHW	Liquid hot water
Kg/ha	Kilogram per hectare
KH ₂ PO ₄	Potassium dihydrogen phosphate
K ₂ HPO ₄	Potassium hydrogen phosphate
kV	kiloVolt
MgC1 ₂	Magnesium chloride
MgSO ₄	Magnesium sulphate
MgSO ₄ .7H ₂ O	Magnesium sulphate heptahydrate
MnSO ₄ .H ₂ O	Magnesium sulphate hydrate
MOA	Ministry of agriculture
mp	melting point
NaOCl	Sodium hypochlorite
NaOH	Sodium hydroxide
NCBI	National center for biotechnology information
NDF	Neutral detergent fiber
OPEFB	Oil palm empty fruit bunch
PCR	Polymerase chain reaction
POME	Palm oil mill effluent
RCM	Reinforced clostridial medium
rRNA	Ribosomal ribonucleic acid
SEM	Scanning electron microscope
T ha ⁻¹	Tons per hectare
TEM	Transmission electron microscope
TYA	Tryptone-yeast extract-acetate medium

v/v	volume per volume
UV	Ultraviolet
w/v	weight per volume
ZnSO ₄	Zinc sulphate hydrate



CHAPTER 1

INTRODUCTION

1.1 Background

Rising prices of crude oil and continuous depletion of existing fossil fuel reserves, combined with concerns over global climate change, have created the need for alternative biofuels to replace fossil fuels. Among alternative fuels, biobutanol is considered as the most potential biofuels as it has many chemical and physical features that are particularly attractive for application as a liquid fuel. Various properties of biobutanol have proved its potential of being alternative fuel over ethanol such as giving higher energy, less sensitivity to temperature, less corrosivity, and the absence of any requires modification in combustion engines (Jang *et al.*, 2012; Lee *et al.*, 2008)

Like ethanol, butanol can be produced petrochemically or fermentatively. Biobutanol is produced by acetone-butanol-ethanol (ABE) fermentation using a variety type of bacteria, particularly clostridia. The most commonly used strains for ABE fermentation are *C. acetobutylicum* and *C. beijerinckii* (Badr *et al.*, 2001; Qureshi *et al.*, 2010). The process was known as ABE fermentation since acetone and ethanol were also produced during the fermentation. The main steps of the fermentation are acidogenesis and solventogenesis phase. ABE fermentation ranked second only to ethanol fermentation in the first part of the 20th century, but disappeared in the second part due to the rise of the petrochemical industry. With the depletion of fossil fuels ABE fermentation becomes interesting again.

Fermentation substrate is the most important factor influencing the cost of biobutanol production (Qureshi and Blaschek, 2000). In order to produce a relatively cheap biobutanol, use of more economical substrates must be identified and evaluated. In this case, lignocelluloses have been identified as the most suitable candidate to expand the biobutanol production capacity with low capital since it is the most abundant renewable resource on the planet and not fully exploited (Qureshi and Ezeji, 2008; Zhang and Shahbazi, 2011).

Malaysia has a lot of agricultural waste that has not been exploited for its usage as a substrate for fermentation. Rice straw is particularly attractive as feedstock for developing a cost-competitive cellulosic biobutanol conversion process since it is available in significant quantities from the rice processing industries. It was estimated about 1.3 million tonnes of rice straw is generated annually from the 684000 hectares of rice fields (MOA, 2004). Commonly, the practice of agricultural wastes are left in the field to rot or discarded through open burning. Since large amount of rice straw produced is disposed by burning which is not only causing environmental hazards but also can cause severe impacts on human health (Gadde *et al.*, 2009; Nori *et al.*, 2008), utilizing the rice straw as substrate for alternative biofuels can help in solving environmental problems associated with its disposal (Binod *et al.*, 2010).



Bioconversion of rice straw into biobutanol is a multi-step process consisting of pretreatment, enzymatic hydrolysis, and fermentation (Mosier *et al.*, 2005; Zhang and Cai, 2008; Lee *et al.*, 2008). To initiate the production of biobutanol from cellulosic biomass, bioconversion of the cellulosic components into reducing sugars is necessary (Kumar *et al.*, 2008). The key obstacle for reducing sugars production is the nature recalcitrant of the raw biomass and therefore pretreatment is particularly crucial to alter the cellulosic biomass by physical, thermal or chemical mean to improve the efficiency of enzymatic hydrolysis of carbohydrates to fermentable sugars (Chang and Holtzapple, 2000).

Despite being considered as a crucial step in the biological conversion to biofuels, biomass pretreatment is one of the main economic costs in the process. In fact, it has been described as the second largest expense in the conversion of lignocellulosic biofuels after the feedstock cost (Mosier *et al.*, 2005). Therefore, developing a cost-effective and efficient biomass pretreatment technology is required to improve the production efficiencies and reduce the costs of lignocellulosic biofuels production (Yang and Wyman, 2008).

In order to reduce the cost of lignocellulosic biofuels, the present study was focused on the feasibility of rice straw as a substrate for bioconversion to biobutanol. Thus, the objectives of the study were:

- i) To evaluate the effects of alkaline pretreatment on the lignocellulosic structure and chemical composition of rice straw in enhancement of reducing sugar production from rice straw.
- ii) To screen, isolate and identify acetone-butanol-ethanol (ABE) producing strain.
- iii) To optimize ABE production from rice straw hydrolysate by a locally isolated *Clostridium sporogenes* A3.

REFERENCES

- Abd-Alla, M.H., El-Enany, A.W.E. (2012) Production of acetone-butanol-ethanol from spoilage date palm (*Phoenix dactylifera L.*) fruits by mixed culture of *Clostridium acetobutylicum* and *Bacillus subtilis*. *Biomass and Bioenergy* 42:172–178.
- Abedinifar, S., Karimi, K., Khanahmadi, M. and Taherzadeh, M.J. (2009). Ethanol production by *Mucor indicus* and *Rhizopus oryzae* from rice straw by separate hydrolysis and fermentation. *Biomass and Bioenergy* 33: 828–833.
- Agbor, V.A., Cicek, N., Sparling, R., Berlin, A and Levin, D.B. (2011). Biomass pretreatment: Fundamentals toward application. *Biotechnology Advances* 29: 675-685.
- Ahmed, S., Bashir, A., Saleem, H., Saadia, M. and Jamil, A. (2009). Production and purification of cellulose-degrading enzymes from filamentous fungus *Trichoderma harzianum. Pakistan Journal of Botany* 41: 1411-1419.
- Ambarsari, H. and Sonomoto, K. (2012). Enhanced acetone, butanol, and ethanol fermentation by *Clostridium saccharoperbutylacetonicum* N1-4 (ATCC 13564) in a chemically defined medium: Effect of iron and initial pH on ABE ratio. *Microbiology Indonesia* 6(4): 139-147.
- Alvira, P., PejÓ, E.T., Ballesteros, M. and Negro, M.J. (2010). Pretreatment technologies for an efficient bioethanol production process based on enzymatic hydrolysis: A review. *Bioresource Technology* 101: 4851-4861.
- Apun, K., Salleh, M.A. and Jong, B.C. (2000). Screening and isolation of a cellulolytic and amylolytic *Bacillus* from sago pith waste. *Journal of Genetic Application Microbiology* 46: 263-267.
- Arantes, V. and Saddler, J.N. (2010). Access to cellulose limits the efficiency of enzymatic hydrolysis: the role of amorphogenesis. *Biotechnology for Biofuels* 3:4.
- Atsumi, S., Hanai, T. and Liao, J.C. (2008). Non-fermentative pathways for synthesis of branched-chain higher alcohols as biofuels. *Nature* 451(7174): 86-9.
- Badr, H.R., Toledo, R. and Hamdy, M.K. (2001). Continuous acetone-ethanolbutanol fermentation by immobilized cells of *Clostridium acetobutylicum*. *Biomass and Bioenergy* 20: 119-132.
- Baharuddin, A.S., Wakisaka, M., Shirai, Y., Abd Aziz, S., Abdul Rahman, N.A. and Hassan, M.A. (2009). Co-composting of empty fruit bunches and partially treated palm oil mill effluents in pilot scale. *International Journal of Agriculture Research* 4(2): 69-78.
- Bahrin, E.K., Baharuddin, A.S., Ibrahim, M.F., Razak, M.N.A., Sulaiman, A., Abd-Aziz, S., Hassan, M.A., Shirai, Y. and Nishida, H. (2012). Physicochemical

- property changes and enzymatic hydrolysis enhancement of oil palm empty fruit bunches treated with superheated steam. *Bioresources* 7(2): 1784-1801.
- Ballongue, J., Amine, J., Petitdemange, H. and Gay, R. (1986). Regulation of acetate kinase and butyratekinase by acid in *Clostridium acetobutylicum*. *Microbiology* 35: 295–30.
- Bals, B., Rogers, C., Jin, M., Balan, V., and Dale, B. (2010). Evauation of ammonia fibre expansion (AFEX) pretreatment for enzymatic hydrolysis of switchgrass harvested in different seasons and locations. *Biotechnology for Biofuels* 3(1)
- Bak, J.S., Ko, J.K., Han, Y.H., Lee, B.C., Choi, I.G. and Kim, K.H. (2009). Improved enzymatic hydrolysis yield of rice straw using electron beam irradiation pretreatment. *Bioresource Technology* 100: 1285-1290.
- Bakri, Sanusi, D. and Muin, M. (2009). ASR of rice husk and the potential use of RHA to mitigate ASR in cement composite. *Jurnal Perennial* 6(1): 25-32.
- Banerjee, S. (May 2012). FAO expects rice production to rise by 1.7% in 2012. Thailand Business News[Online]. Available : <u>http://thailand-business-news.com/news/top-stories/37983-fao-expects-rice-production-to-rise-by-1-7-in-2012.</u>
- Binod, P., Sindhu, R., Singhania, R.R., Vikram, S., Devi, L., Nagalakshmi, S., Kurien, N., Sukumaran, R.K. and Pandey, A. (2010). Bioethanol production from rice straw: An overview. *Bioresource Technology* 101: 4767-4774.
- Brodeur, G., Yau, E., Badal, K., Collier, J., Ramachandran, K.B. and Ramakrishnan,S. (2011). Chemical and physicochemical pretreatment of lignocellulosic biomass: A review, *Enzyme Research*.
- Bryant, D.L. and Blaschek, H.P. (1988). Buffering as a means for increasing growth and butanol production by *Clostridium acetobutylicum*. *Journal of Industrial Microbiology* 3(1): 49-55.
- Cara, C., Ruiz, E., Ballesteros, M., Manzanares, P., Negro, J.M. and Castro, E. (2008). Production of ethanol from steam-explosion pretreated olive oil tree pruning. *Fuel* 87: 692-700.
- Cardona, C. A. and Sanchez, O.J. (2007). Fuel ethanol production: Process design trends and integration opportunities. *Bioresource Technology* 98(12): 2415-2457.
- Carere, C.R., Sparling, R., Cicek, N. and Levin, D.B. (2008). Third generation biofuels via direct cellulose fermentation. *International Journal of Molecular Sciences* 9:1342–60.
- Chandra, R.P., Bura, R., Mabee, W.E., Berlin, A., Pan, X. and Saddler, J.N. (2007). Substrate pretreatment: the key to effective enzymatic hydrolysis of lignocellulosics? *Advances in Biochemical Engineering/ Biotechnology* 108:67–93.

- Chandrasekhar, S., Satyanarayana, K.G. and Pramada, P.N. (2003). Review processing, properties and applications of reactive silica from rice husk-an overview. *Journal of Material Science* 38: 3159-3168.
- Chang, V.S. and Holtzapple, M.T. (2000). Fundamental factors affecting biomass enzymatic reactivity. *Applied Biochemistry and Biotechnology* (84–86): 5-37.
- Cheng, J., Su, H., Zhou, J., Song, W. and Cen, K. (2010). Microwave-assisted alkali pretreatment of rice straw to promote enzymatic hydrolysis and hydrogen production in dark- and photo-fermentation. *International Journal of Hydrogen Energy* 36: 2093-2101.
- Chittitabu, S., Rajendran, K. and Santhanmuthu, M. (2011). Optimization of microwave assisted alkali pretreatment and enzymatic hydrolysis of Banana pseudostem for bioethanol production. *2nd International Conference on Environmental Science and Technology* IPCBEE. IACSIT Press, Singapore.
- Clark, T.A. and Mackie, K.L. (1987) Steam explosion of the soft-wood Pinus radiate with sulfphur dioxide addition. Process optimization. *Journal of Wood Chemistry and Technology* 7:373–403.
- Corrẽa, P.C., Schwanz da Silva, F, Jareb, C., Afonso junior, P.C. and Arana, I. (2007). Physical and mechanical properties in rice processing. *Journal of Food Engineering* 79: 137-142.
- Csōke, B., Bokanyi, L., Bōhm, J and Pethō, S.Z. (2003). Selective grindability of lignites and its application for producing an advanced fuel. *Applied Energy* 74: 359-368.
- Dashtban, M., Schraft, H. and Qin, W. (2009). Fungal bioconversion of lignocellulosic residues; opportunities & perspectives. *International Journal of Biological Sciences* 5(6): 578-595.
- Demirbas, M.F., Balat, M. and Balat, H. (2011). Biowastes-to-biofuels. *Energy Conversion and Management* 52: 1815-1828.
- Donaldson, V.N., Gail, K., Eliot, A.C., Flint, D. and Maggio-Hall, A. (2011). Fermentative production of four carbone alcohols. *Patent application publication*, p. 1-326,
- Drapcho, C.M., Nhuan, N.P., Walker, T.H. (2008). Biofuels Engineering Process Technology. Mc Graw Hill Companies, Inc..
- Durre, P. (1998). New insights and novel developments in clostridial acetonebutanol-isopropanol fermentation. *Applied microbiology and biotechnology* 49: 639-648.
- Durre P. (2005). Sporulation I clostridia. In: Durre P, editor. *Handbook on clostridia*. *Boca Raton*: CRC Press. p 659-699.
- Durre P. (2007) Biobutanol: an attractive biofuel. *Biotechnology Journal* 2:1525–34.

- Durre, P. (2008). Fermentative butanol production: bulk chemical and biofuel. Annals of the New York Academy of Sciences. 1125: 353–362
- Eriksson, K-EL., Bermek, H. (2009). Lignin, lignocellulose, ligninase. *Applied and Industrial Microbiology* 373–84.
- Eriksson, T., Karlsson, J. and Tjerneld, F. (2002). A model explaining declining rate in hydrolysis of lignocellulose substrates with cellobiohydrolase I (Cel7A) endoglucanase ! (Cel7B) of *Trichoderma reesei*. Applied Biochemistry and Biotechnology 101: 41-60.
- Esteghalian, A.R., Srivastava, V., Gilkes, N., Gregg, D.J. and Saddler, J.N. (2000). An overview of factors influencing the enzymatic hydrolysis of lignocellulosic feedstocks. Glycosyl Hydrolases for Biomass Conversion Washington DC.
- FAOSTAT F. Agriculture Organization of the United Nations. Statistical database; 2009.
- Fatimah, M.A., Nik Mustapha, R.A., Bisant, K. and Amin, M.A. (2007). 50 years of Malaysian Agriculture Transformational Issues Challenges & Direction,1st ed. Universiti Putra Malaysia Press 11: 281-308.
- Feng, Q., Lin, Q., Gong, F., Sugita, S. and Shoya, M. (2004). Adsorption leas and mercury by rice husk ash. *Journal of Colloid and Interface Sicence* 1(1):1-8.
- Fond, O., Matta-Ammouri, G., Petitdemange, H. and Engasser, J. M. (1985). The role of acid on the production of acetone and butanol by *Clostridium acetobutylicum*. *Applied Microbiology and Biotechnology* 22: 195–200.
- Formanek, J., Mackie, R. and Blaschek, H.P. (1997). Enhanced butanol production by *Clostridium beijerinckii* BA101 grown in semidefined P2 medium contining 6 percent maltodextrin or glucose. *Applied and Environmental Microbiology* 63: 2306-2310.
- Gadde, B., Bonnet, S., Menke, C. and Garivait, S. (2009). Air pollution emissions from rice straw open field burning in India, Thailand and the Philippines. *Environmental Pollution* 157: 1554-1558.
- Galbe, M., and Zacchi, G. (2007). Pretreatment of lignocellulosic materials for efficient bioethanol production. *Biofuels* 108: 41–65.
- Gheshlaghi, R., Scharer, J.M., Young, M.M. and Chou, C.P. (2009). Metabolic pathways of Clostridia for producing butanol. *Biotechnology Advances* 27:764–81.
- Gidde, M.R. and Jivani, A.P. (2007). Waste to wealth-Potential of rice husk in India a literature review. *Proceedings of the International Conference on Cleaner Technologies and Environemtal Management PEC, Pondicherry, India,* January 4-6 586-590.
- Gnansounou, E. and Dauriat, A. (2005). Ethanol fuel from biomass: A review. Journal of Scientific & Industrial Research 64: 809-821.

- Govunami, S.P., Koti, S., Kothagouni, S.Y., Venkateshhwar, S., and Linga, V.R. (2013). Evaluation of pretreatment methods for enzymatic saccharification of wheat straw for bioethanol production. *Carbohydrate Polymers* 91: 646-650.
- Guo, G.L., Chen, W.H., Men, L.C., and Hwang, W.S. (2008). Characterization of dilute acid pretreatment of silvergrass for ethanol production. *Bioresouce*. *Technology* 99: 6046-6053.
- He,Y., Pang, Y., Liu, Y., Li, X. and Wang, K. (2008). Physicochemical characterization of rice straw pretreated with sodium hydroxide in the solid state for enhancing biogas production. *Energy & Fuels* 22: 2775-2781.
- Hock, L.S., Baharuddin, A.S., Ahmad, M.N., Shah, U.K.M., Abdul Rahman, N.A., Abd-Aziz, S., Hassan, M.A. and Shirai, Y. (2009). Physicochemical changes in windrow co-composting process of oil palm mesocarp fiber and palm oil mill effluent anaerobic sludge. *Australian Journal of Basic and Applied Sciences* 3(3): 2809-2816.
- Howard, R.L., Abotsi, E., Jansen van Rensburg, E.L. and Howard, S. (2003). Lignocellulose biotechnology: issues of bioconversion and enzyme production. *African Journal of Biotechnology* 2(12): 802-819.
- Hsu, T.C., Guo, G.L., Chen, W.H., and Hwang, W.S. (2010). Effect of dilute acid pretreatment of rice straw on structural properties and enzymatic hydrolysis. *Bioresource Technology* 101: 4907–4913.
- Ibrahim, M.F., Abd-Aziz, S., Razak, M.N.A and Phang, L.Y. (2012). Oil palm empty fruit bunch as alternative substrate for acetone-butanol-ethanol production by *Clostridium butyricum EB6. Applied Biochemistry and Biotechnology* 166: 1615-1625.
- Ibrahim, M.M., El–Zawawy, W.K., Abdel–Fattah, Y.R., Soliman, N.A., and Agblevor, F.A. (2011). Comparison of alkaline pulping with steam explosion for glucose production from rice straw. *Carbohydrate Polymers* 83(2): 720–726.
- Ioelovich, M. and Morag, E. (2012). Study of enzymatic hydrolysis of mild pretreated lignocellulosic biomass. *BioResources* 7(1): 1040-1052.
- Jang, Y.S., Lee, J., Malaviya, A., Seung do, Y., Cho, J.H. and Lee, S.Y. (2012). Butanol production from renewable biomass: Rediscovery of metabolic pathways and metabolic engineering. *Biotechnology Journal* 7(2): 186-98.
- Jiang, Y., Xu, C., Dong, F., Yang, Y., Jiang, W. and Yang, S. (2009). Disruption of the *acetoacetate decarboxylase* gene insolvent-producing *Clostridium acetobutylicum* increases the butanol ratio. *Metabolic Engineering* 11:284–9.
- Jin, C., Yao, M., Liu, H., Lee, C.F. and Ji, J. (2011) Progress in the production and application of n-butanol as a biofuel. *Renewable and Sustainable Energy Reviews*. 15: 4080-4106.

- Johansson, L., Virkki, L., Anttila, H., Esselstrom, H., Tuomainen, P. and Sontag-Strohm, T. (2006). Hydrolysis of b-glucan. *Food Chemistry* 97: 71-79.
- Johnson, J.L., Toth, J., Santiwatanakul, S., Chen, J.S. (1997). Cultures of *'Clostridium acetobutylicum''* from various collections comprise *Clostridium acetobutylicum, Clostridium beijerinckii*, and two other distinct types based on DNA–DNA reassociation. *International Journal of Systematic Bacteriology* 47(2):420–4.
- Jones, D.T. and Woods, D.R. (1986). Acetone–butanol fermentation revisited. *Microbiology Revision* 50(4):484–524.
- Jørgensen, H., Kristensen, J.B., and Felby, C. (2007). Enzymatic conversion of lignocellulose into fermentable sugars: challenges and opportunities. *Biofuels Bioproducts and Biorefining* 1: 119-134.
- Kadam, K.L., Forrest, L.H. and Jacobson, W.A. (2000). Rice straw as a lignocellulosic resource: collection, processing, transportation, and environmental aspetcs. *Biomass and Bioenergy* 18: 369-389.
- Karunanithy, C. and Muthukumarappan, K. (2011). Optimization of alkali, big bluestem particle size and extruder parameters for maximum enzymatic sugar recovery using response surface methodology. *BioResources* 6(1): 762-790.
- Khaw, T.S., and Ariff, A.B. (2009). Optimization of enzymatic saccharification of palm oil effluent solid and oil palm fruit fibre to fermentable sugars. *Journal of Tropical Agriculture and Food Sciences* 37(1): 85-94.
- Ko, J.K., Bal, J.S., Jung, M.W., Lee, H.J., Choi, I.G., Kim, T.H. and Kim, K.H. (2009). Ethanol production from rice straw nusing ammonia soaking pretreatment and simultaneous saccharification and fermentation processes. *BioResources* 100: 4374-4380.
- Kristensen, J.B. (2008): Enzymatic hydrolysis of lignocellulose. Substrate interactions and high solids loadings. Forest & Landscape Research No. 42-2008. Forest & Landscape Denmark. Frederiksberg. 130 pp.
- Kuhad, R.C., Gupta, R. and Singh, A. (2011). Microbial cellulases and their industrial applications. *Enzyme Research*. Article ID 280696, 10 pages, 2011. doi:10.4061/2011/280696
- Kumar, M. and Gayen, K. (2011). Developments in biobutanol production: New insights. *Applied Energy* 88: 1999-2012.
- Kumar N, Das D (2000) Enhancement of hydrogen production by *Enterobacter* cloaca IIT-BT 08. Process Biochem 35:589–593.
- Kumar, P., Barret, D.M., Delwiche, M.J. and Stroeve, P. (2009). Methods for pretreatment of lignocellulosic biomass for efficient hydrolysis and biofuel production. *Industrial Engineering Chemistry Research* 48: 3713-3729.

- Kumar, R., Singh, S. and Singh, O.V. (2008). Bioconversion of lignocellulosic biomass: biochemical and molecular perspectives. *Journal of Industrial Microbiology and Biotechnology* 35:377-391.
- Kumar, S., Kothari, U., Kong, L., Lee, Y.Y. and Gupta, R.B. (2011). Hydrothermal pretreatment of switchgrass and corn stover for production of ethanol and microspheres. *Biomass & Bioenergy* 35: 956-968
- Laureano-Perez, L., Teymouri, F., Alizadeh, H., Dale, B.E. (2005). Understanding Factors that Limit Enzymatic Hydrolysis of Biomass. In: *Twenty-Sixth Symposium on Biotechnology for Fuels and Chemicals* 1081–1099.
- Lee, S.Y., Park, J.H., Jang, S.H., Nielsen, L.K., Kim, J., Jung, K.S. (2008). Fermentative butanol production by Clostridia. *Biotechnology and Bioengineering* 101:209–228
- Liew, S.T., Arbakariya, A., Rosfarizan, M. and Raha, A.R. (2006). Production of solvent (acetone-butanol-ethanol) in continuous fermentation by *Clostridium saccharobutylicum* DSM 13864 using gelatinized sago starch as a carbon source. *Malaysian Journal of Microbiology* 2(2): 42-50.
- Lim, J.S., Manan, Z.A., Alwi, S.R.W. and Hashim, H. (2012). A review on utilization of biomass from rice industry as a source of renewable energy. *Renewable and Sustainable Energy Reviews* 16: 3084-3094.
- Liu, X. and Yang, S. T. (2006). Kinetics of butyric acid fermentation of glucose and xylose by *Clostridium tyrobutyricum* wild type and mutant. *Process Biochemistry* 41: 801–808.
- Lynd, L.R., Cushman, J.H., Nichols, R.J. and Wyman, C.E. (1991). Fuel ethanol from cellulosic biomass. *Science*.
- Lynd, L.R., Weimer, P.J., van Zyl, W.H. and Isak, S. (2002). Microbial cellulose utilization: Fundamentals and biotechnology. *Microbiology and Molecular Biology Reviews* 66(3): 506-577.
- Maddox, I.S., Steiner, E., Hirsch, S., Wessner, N.A., Grapes, J.R. and Schuster, K.C. (2000). The cause of "acid crash" and "acidogenic fermentations" during batch acetone-butanol-ethanol (ABE) fermentation process. *Journal of Molecular Microbiology and Biotechnology* 2(1): 95-100.
- Mahmud, M. and Shafie, S.H.M. (2012). Comparison of air quality between 2002 and 2003 in the area during the burning of rice straws in Mergong, Kedah. GEOGRAFIA OnlineTM *Malaysia Journal of Society and Space* 8(3): 33-42.
- Mansfield, S.D., Mooney, C. and Saddler, J.N. (1999). Substrate and enzyme characteristics that limit cellulose hydrolysis. *Biotechnology Progress* 15: 804-816.
- Miller, G.L. (1959). Use of DNS reagent for the determination of reducing sugars. Analytical Chemistry 31: 426-428.

- Mirahmadi, K., Kabir, M.M., Jeihanipour, A., Karimi, K. and Taherzadech, M.J. (2010). Alkaline pretreatment of spruce and birch to improve bioethanol and biogas production. *BioResouces* 5(2): 928-938.
- Mitchell, W.J. (1998). Physiology of carbohydrate to solvent conversion by Clostridia. *Journal of Advanced Microbiology and Physiology* 39:31-130.
- MOA. 2004. Buletin Kementerian Pertanian dan Industri Asas Tani Malaysia. Bil. 1: Jan-April, 2004. *Kementerian Pertanian dan Industri Asas Tani Malaysia*.
- Mohanty S. Rice crisis: the aftermath. Rice Today 2008;7.
- Monot F, Martin J-R, Petitdemange H, Gay R. 1982. Acetone and butanol production by *Clostridium acetobutylicumin* a synthetic medium. *Applied and Environmental Microbiology* 44:1318–1324
- Montoya, D., Spitia, S., Silva, E. and Schwarz, W.H. (2000). Isolation of mesophilic solvent-producing clostridia from Colombian source: physiological characterization, solvent production and polysaccharide hydrolysis. *Journal of Biotechnology* 79: 117-126.
- Mora-pale, M., Meli, L., Doherty, T.V., Linhardt, R.J. and Dordick, J.S. (2011). Room temperature ionic liquids as emerging solvents for the pretreatment of lignocellulosic biomass. *Biotechnology and Bioengineering* 108: 1229-1245.
- Moradi, F., Amiri, H., Soleimanianzad, S., Ehsani, M.R. and Karimi, K. (2013). Improvement of acetone, butanol and ethanol production from rice straw by acid and alkaline pretreatments. *Fuel* 112: 8–13.
- Mosier, N., Wyman, C., Dale, B., Elander, R., Lee, Y.Y., Holtzapple, M. and Ladisch, M. (2005). Features of promising technologies for pretreatment of lignocellulosic biomass. *Bioresource Technology* 96: 673-686.
- Munaf, E. and Zein, R. (1996). The use of rice husk for removal; of toxic metals from wastewater. *Environmental Technology* 18 (3): 359-362.
- Najim, M.M.M., Lee, T.S., Haque, M.A. and Esham, M. (2007). Sustainability of rice production: A Malaysian perspective. *The Journal of Agricultural Sciences* 3(1): 1-12.
- Neethirajan, S., Gordon, R., and Wang, L. (2009). Potential of silica bodies (phytoliths) for nanotechnology. *Trends in Biotechnology* 27(8): 461-467.
- Niven, R.K. (2005). Ethanol in gasoline: environmental impacts and sustainability review article. *Renewable and Sustainable Energy Reviews*. 9(6); 535-555
- Nori, H., Halim, R.A. and Ramlan, M.F. (2008). Effects of nitrogen fertilization management practice on the yield and straw nutritional quality of commercial rice varieties. *Malaysia Journal of Mathematical Sciences* 2(2): 61-71.
- Norsida, M., & Sami, I. S. (2009). Off-farm employment participation among paddy farmers in the Muda Agricultural Development Authority and Kemasin

- Semerak granary areas of Malaysia. *Asia-Pacific Development Journal* 16(2): 141-153.
- Oh, S.Y., Yoo, D.I., Shin, Y., and Seo, G. (2005). FTIR analysis of cellulose treated with sodium hydroxide and carbon dioxide. *Carbohydrate Research* 340: 417–42.
- Ong, L.G.A., Chuah, C. and Chew, A.L. (2010). Comparison of sodium hydroxide and potassium hydroxide followed by heat treatment on rice straw for cellulase production under solid state fermentation. *Journal of Applied Sciences* 10(21): 2608-2612.
- Palmqvist, E. and Hahn-Hagerdal, B. (2000). Fermentation of lignocellulosic hydrolysates. II: inhibitors and mechanisms of inhibition. *Bioresource Technology* 74 (1): 25–33.
- Papoutsakis, E.T. (2008). Engineering solventogenic Clostridia. *Current Opinion in Biotechnology* 19:420–9.
- Pereira, L., Amado, A.M., Critchley, A.T., van de Velde, F. and Ribeiro-Claro, P.J.A. (2009). Identification of selected seaweed polysaccharides (phycocolloids) by vibrational spectroscopy (FTIR-ATR and FT-Raman). *Food Hydrocolloids* 23(7): 1903-1909.
- Pereira Ramos, L. (2003). The chemistry involved in steam treatment of lignocellulosic materials. *Quim. Nova* 26(6): 863-871.
- Qureshi, N. and Blaschek, H.P. (2000). Economics of butanol fermentation using hyper-butanol producing *Clostridium beijerinckii* BA101, *Trans IChemE* 78: 139-144.
- Qureshi, N. and Ezeji, T.C. (2008). Butanol, 'a superior biofuel' production from agricultural residues (renewable biomass): recent progress in technology. *Biofuels, Bioproducts & Biorefining* 2: 319-330.
- Qureshi, N. and Maddox, I.S. (2005). Reduction of butanol inhibition by perstraction: utilization of concentrated lactose/ whey permeate by *Clostridium acetobutylicum* to enhance butanol fermentation economics. *Food and Bioproducts Processing* 83: 43-52.
- Qureshi, N., Saha, B.C and Cotta, M.A. (2007). Butanol production from wheat straw hydrolysate using *Clostridium beijerinckii*. *Bioprocess and Biosystems Engineering* 30:419-427
- Qureshi, N., Saha, B.C., Dien, B., Hector, R.E. and Cotta, M.A. (2010). Production of butanol (a biofuel) from agricultural residues: Part 1 Use of barley straw hydrolysate. *Biomass and Bioenergy* 34: 559-565.
- Rahnama, N., Mamat, S., Shah, U.K.M., Ling, F.H., Rahman, N.A.A. and Ariff, A.B. (2013). Effects of alkali pretreatment of rice straw on cellulose and xylanase production by local *Trichoderma harzianum* SNRS3 under solid state fermentation. *BioResources* 8(2): 2881-2896.

- Ramli, N.N., Shamsudin, M.N., Mohamad, Z. and Radam, A. (2012). Impact of price support on Malaysian rice industry. UMT 11th International Annual Symposium on Sustainability Science and Management 799-804.
- Ranjan, A. and Moholkar, V., (2013). Comparative study of various pretreatment techniques for rice straw saccharification for the production of alcoholic biofuels. *Fuel* 112: 567–571.
- Rodrigues, A.L., Cavalett, A. and Lima, A.O.S. (2010). Enhancement of *Escherichia coli* cellulolytic activity by co-production of beta-glucosidase and endoglucanase enzymes. *Electronic Journal of Biotechnology* 13(5).
- Roslan, A.M., Hassan, m.A., Abd-Aziz, S. and Yee, P.L. (2009). Effect of palm oil mill effluent supplementation on cellulose production from rice straw by local fungal isolates. *International journal of Agricultural Research* 4(5): 185-192.
- Ruiz, E., Cara, C., Manzanares, P., Ballesteros, M., and Castro, E. (2008). Evaluation of steam explosion pre-treatment for enzymatic hydrolysis of sunflower stalks. *Enzyme and Microbial Technology* 42: 160-166.
- Saha, B.C. (2000). Alpha-L-arabinofuranosidases: Biochemistry, molecular biology and application in biotechnology. *Biotechnology Advances* 18:403-423.
- Saha, B.C. (2003). Hemicellulose bioconversion. *Journal of Industrial Microbiology* and Biotechnology 30:279-91.
- Sahare, P., Singh, R., Laxman, R.S. and Rao, M. (2012). Effect of alkali pretreatment on the structural properties and enzymatic hydrolysis of corn cob. *Applied Biochemistry and Biotechnology* 168: 1806-1819.
- Sanchez, O. J. and Cardona, C. A. (2008). Trends in biotechnological production of fuel ethanol from different feedstocks. *Bioresource Technology* 99: 5270-5295.
- Schwanninger, M, Hinterstoisser, B, Gradinger, C, Messner, K, and Fackler, K. (2004). Examination of spruce wood biodegraded by *Ceriporiopsis* subvermispora using near and mid infrared spectroscopy. Journal of Near Infrared Spectroscopy 12: 397-409.
- Shahriarinour, M., Wahab, M.N.A., Mustafa, S., Mohamad, R. and Ariff, A.B. (2011). Effect of various pretreatments of oil palm empty fruit bunch fibers for subsequent use as substrate on the performance of cellulase production by *Aspergillus Terreus. BioResource* 6(1): 291-307.
- Shamsudin, S., Shah, U.K.M., Zainudin, H., Abd-Aziz, S., Kamal, S.M.M., Shirai, Y. and Hassan, M.A. (2011). Effect of steam pretreatment on oil palm empty fruit bunch for the production of sugars. *Biomass and Bioenergy* 36: 280-288.
- Sharma, R. (2012). Novel pretreatment methods of switchgrass for fermentable sugar generation. *Master Thesis, North Carolina State University*.

- Shanmugam, S. and Satishkumar, T. (2009). Enzyme Technology. *Enzyme Kinetics* (pp 48-108) I.K. International Publishing House Pvt. Ltd.
- Sheehan, J.S. and Himmel, M.E. (2001). Outlook for bioethanol production from lignocellulosic feedstocks: technology hurtles. *Agro Food Industry Hi-Tech* 12(5):54-57.
- Shiratori, H., Sasaya, K., Ohiwa, H., Ikeno, H., Ayame, S., Kataoka, N., Miya, A., Beppu, T. and Ueda, K. (2009). *Clostridium clariflavum sp. nov.* and *Clostridium caenicola sp. nov.*, moderately thermophilic, cellulose-/cellobiose-digesting bacteria isolated from methanogenic sludge. *International Journal of systematic and Evolutionary Microbiology* 59: 1764-1770.
- Sierra, R., Smith, A., Granda, C., Holtzapple, M.T. (2008). Producing fuels and chemicals from lignocellulosic biomass. *Chemical Engineering Progress* 104:S10–S18.
- Sillers, R., Chow, A., Tracy, B. and Papoutsakis, E.T. (2008). Metabolic engineering of the non-sporulating, non-solventogenic *Clostridium acetobutylicum* strain M5 to produce butanol without acetone demonstrate the robustness of the acid-formation pathways and the importance of the electron balance. *Metabolic Engineering* 10:321–32.
- Sills, D.L., and Gossett, J.M. (2011). Assessment of commercial hemicellulases for saccharification of alkaline pretreated perennial biomass. *Bioresource Technology*. 102(2): 1389–1398.
- Silverstein, R.A., Chen, Y., Sharma-Shiyappa, R.R., Boyette, M.D., and Osborne, J. (2007). A comparison of chemical pretreatment methods for improving saccharification of cotton stalks. *Bioresource Technology* 98: 3000–3011.
- Singh, A. and Bishnoi, N.R. (2012). Enzymatic hydrolysis optimization of microwave alkali pretreated wheat straw and ethanol production by yeast *Bioresource Technology* 108: 94-101.
- Singh, D.P., and Trivedi, R.K. (2013). Acid and alkali pretreatment of lignocellulosic biomass to produce ethanol as biofuel. *International Journal of ChemTech Research* 5(2): 727-734.
- Sipalan, J. (Feb 2012). Full rice yield still unrealized. The Star online [Online]. Available:<u>http://thestar.com.my/news/story.asp?file=/2012/2/15/nation/10736</u> 889&sec=nation
- Somerville, C., Bauer, S., Brininstool, G., Facette. M. and Hamann, T. (2004). Toward a systems approach to understanding plant cell walls. *Science* 306:2206–11.
- Srinokutara, T., Suttikul, S. and Boonvitthya, N. (2013). Effect of different pretreatment methods on the enzymatic saccharification and ethanol production from sugarcane shoots and leaves. *Journal of Food Science and Engineering* 3:309-316.

- Sukumaran, R.K., Singahnia, R.R. and Pandey, A. (2005). Microbial cellulases-Production, application and challenges. *Journal of Scientific & Industrial Research* 64: 832-844.
- Sun, R.C., Tomkinson, J., Sun, X.F. and Wang, N.J. (2000). Fractional isolation and physic-chemical characterization of alkaline-soluble lignins from fastgrowing poplar woods. *Polymer*. 41:8409-8417.
- Sun, Y. and Cheng, Y. (2002). Hydrolysis of lignocellulosic materials for ethanol production: a review. *Bioresource Technology* 83: 1-11.
- Taherzadeh, M. and Karimi, K. (2008). Pretreatment of lignocellulosic wastes to improve ethanol and biogas production: A review. *International Journal of Molecular Sciences* 9: 1621–1651.
- Taherzadeh, M.J., Eklund, R., Gustafsson, L., Niklasson, C., and Liden, G. (1997). Characterization and fermentation of dilute-acid hydroyzates from wood. *Industrial & Engineering Chemistry Research* 369(11): 4659-4665.
- Tengborg, C., Galbe, M., and Zacchi, G. (2001). Influence of enzyme loading and physical parameters on the enzymatic hydrolysis of steam-pretreated softwood. *Biotechnology Progress* 17(1):110-117.
- Tengerdy, R. P. and Szakacs, G. (2003). Bioconversion of lignocellulose in solid substrate fermentation. *Biochemical Engineering Journal* 13(2-3): 169-179.
- Tomas-Pejo, E., Alvira, P., Ballesteros, M. and Negro, M.J. (2011). Pretreatment technologies for lignocellulose-to-bioethanol conversion. *Biofuels: Alternative Feedstocks and Conversion Process.* 7:149-176.
- Torget, R., Walter, P., Himmel, M. and Grohmann, K. (1991). Dilute-acid pretreatment of corn residues and short-rotation woody crops. *The Humana Press Inc.*
- Väljamäe, P., Kipper, K., Pettersson, G. and Johansson, G. (2003) Synergestic cellulose hydrolysis can be described in terms of fractal-like kinetics. *Biotechnology and Bioengineering* 84: 254-257.
- Van Dyk, J.S. and Pletschke, B.I. (2012). A review of lignocellose bioconversion using enzymatic hydrolysis and synergistic cooperation between enzymes-Factors affecting enzymes, conversion and synergy. *Biotechnology Advances* 30: 1458-1480.
- Wang,Z., Keshwani, D.R., Redding, A.P. and Cheng, J.J. (2010). Sodium hydroxide pretreatment and enzymatic hydrolysis of coastal Bermuda grass. *Biological Systems Engineering: Papers and Publications*. Paper 143.
- Wang, Z., Keshwani, D.R., Redding, A.P. and Cheng, J.J. (2008). Alkaline pretreatment of Coastal Bermuda grass for bioethanol production. *Conference Presentations and White Papers: Biological Systems Engineering*. Paper 38.
- Wyman, C.E. (1994). Ethanol from lignocellulosic biomass: Technology, economics, and opportunities. *Bioresource Technology* 50: 3-16.

- Wyman, C.E., Dale, B.E., Elander, R.T., Holtzapple, M., Ladisch, M.R. and Lee, Y.Y. (2005). Coordinated development of leading biomass pretreatment technologies. *Bioresource Technology* 96: 1959–1966.
- Xiao, B., Sun, X.F., and Sun, R.C. (2001). Chemical, structural, and thermal characterizations of alkali-soluble lignins and hemicelluloses, and cellulose from maize stems, rye straw, and rice straw. *Polymer Degradation and Stability* 74: 307-319
- Yang, B. and Wyman, C.E. (2008). Pretreatment: the key to unlocking low-cost cellulosic ethanol. *Biofuels, Bioproducts and Biorefining* 2: 26-40.
- Yang, B., Dai, Z., Ding, S.Y. and Wyman, C.E. (2011). Enzymatic hydrolysis of cellulosic biomass. *Biofuels* 2(4): 421-450.
- Yang, H., Yan, R., Chen, H., Lee, D.H., and Zheng, C. (2007). Characteristics of hemicelluloses, cellulose and lignin pyrolysis. *Fuel* 86: 1781-1788.
- Yang, X., Tu, M., Xie, R., Adhikari, S. and Tong, Z. (2013). A comparison of three pH control methods for revealing effects of undissociated butyric acid on specific butanol production rate in batch fermentation of *Clostridium acetobutylicum*. *AMB Express* 3(3)
- Zhang, B., Shahbazi, A., Wang, L., Diallo, O., and Whitmore, A. (2010). Alkali pretreated and enzymatic hydrolysis of cattails from constructed wetlands. *American Journal of Engineering and Applied Sciences* 3(2): 328-332.
- Zhang, B. and Shahbazi, A. (2011). Recents developments in pretreatment technologies for production of lignocellulosic biofuels. *Petroleum & Environmental Biotechnology* 2:111.
- Zhang, Q. and Cai, W. (2008). Enzymatic hydrolysis of alkali-pretreated rice straw by *Trichoderma reesei* ZM4-F3. *Biomass and Bioenergy* 32: 1130–1135.
- Zhang, Y.H. (2008). Reviving the carbohydrate economy via multiproduct lignocellulosic biorefineries. Journal of Industrial Microbiology and Biotechnology 35:367-375.
- Zhang, Y.H.P., Ding, S.Y., Mielenz, J.R., Elander, R., Laser, M., Himmel, M., McMillan, J.D. and Lynd, L.R. (2007). Fractionating recalcitrant lignocellulose at modest reaction conditions. *Biotechnology and Bioengineering* 97: 214-223.
- Zhao, J., Wang, Y.H., Chu, J., Luo, L.Z., Zhuang, Y.P., and Zhang, S.L. (2009). Optimization of cellulase mixture for efficient hydrolysis of steam-exploded corn stover by statiscally designed experiments. *Bioresource Technology* 100: 819-825.
- Zhao, X., Zhang, L. and Liu, D. (2011). Biomass recalcitrance. Part 1: the chemical compositions and physical structures affecting the enzymatic hydrolysis of lignocellulose. *Biofuels, Bioproducts & Biorefining* 6: 465-482.

- Zhao, Y., Wang, Y., Zhu, J.Y., Ragauskas, A., Deng, Y (2007). Enhanced enzymatic hydrolysis of spruce by alkaline pretreatment at low temperatures. *Biotechnology and Bioengineering* 99(6): 1321-1328.
- Zheng, Y., Pan, Z. and Zhang, R. (2009), Overview of biomass pretreatment for cellulosic ethanol production, *International Journal of Agriculture and Biological Engineering* 2: 51–68.
- Zhu, Y. and Yang, S.T. (2004). Effect of pH on metabolic pathway shift in fermentation of xylose by *Clostridium tyrobutyricum*. *Journal of Biotechnology* 110: 143-157.
- Zigova, J. and Sturdik, E. (2000). Advances in biotechnological production of butyric acid. *Journal of Industrial Microbilogy and Biotechnology* 24: 153–160.

