



UNIVERSITI PUTRA MALAYSIA

***EFFECTS OF TOCOTRIENOL SUPPLEMENTATION ON PLATELET
AGGREGATION IN SUBJECTS WITH METABOLIC SYNDROME***

GAN YEE LIN

FBSB 2014 16



**EFFECTS OF TOCOTRIENOL SUPPLEMENTATION ON PLATELET
AGGREGATION IN SUBJECTS WITH METABOLIC SYNDROME**

By

GAN YEE LIN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirements for the Degree of Master of Science**

April 2014

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment
of the requirement for the degree of Master of Science

**EFFECTS OF TOCOTRIENOL SUPPLEMENTATION ON PLATELET
AGGREGATION IN SUBJECTS WITH METABOLIC SYNDROME**

By

GAN YEE LIN

April 2014

Chair : Prof. Lai Oi Ming, PhD

Faculty : Biotechnology and Biomolecular Sciences

An occlusive thrombus in vasculature either in arterial or venous, is a pathological condition that predisposes to most cases of cardiovascular diseases (CVDs). As reported by World Health Organisation (WHO), CVDs were among the major cause of deaths for noncommunicable diseases. Prescription of antithrombotic agents (e.g. aspirin, clopidogrel and warfarin) is a standard treatment given to patients to reduce the risk of death from cardiovascular events. However, presence of side effects and inter-individual variability in response towards these antithrombotic agents may limit the prevention of cardiovascular events throughout the world including Malaysia. Tocotrienols are a group of vitamin E besides tocopherols. Previous animal studies showed significant inhibition of platelet aggregation and antithrombotic effect after tocotrienol administration, but results in human trials are controversial. Therefore, this study was designed to ascertain the effect of tocotrienol supplementation on platelet reactivity in subjects with metabolic syndrome (MetS), by taking into account that MetS subjects are associated with prothrombotic state due to higher platelet reactivity and blood coagulability. Thus, an initiation of antithrombotic therapy is important for them to prevent thrombotic events. In this connection, a randomised, double-blinded, crossover, and placebo-controlled human trial was conducted with a total of 31 MetS subjects (15 males and 16 females) completed the study. Subjects recruited received two interventions in a random order, separated by a 15-day washout period in between the interventions. During the intervention, subjects consumed palm mixed tocotrienols 200 mg or placebo twice daily for 14 days followed by a postprandial study day. Post-intervention results demonstrated that reactivity of arachidonic acid and adenosine 5'-diphosphate (ADP) signalling platelet aggregation was not significantly different ($p > 0.05$) between tocotrienol and placebo interventions. For the results in terms of postprandial change, tocotrienols also did not exert inhibitory effect ($p > 0.05$) on these platelet aggregation measurements compared to placebo. No significant differences ($p > 0.05$) were found on the degree of platelet activation induced by thrombin mimic peptide and haemostatic measures (plasma D-dimer, plasminogen activator inhibitor type 1 (PAI-1), fibrinogen and undercarboxylated osteocalcin (ucOC)) between tocotrienol and placebo interventions, both post-intervention and postprandial. Among the inflammatory measures including plasma soluble P-selectin (sP-selectin), plasma

soluble E-selectin (sE-selectin), plasma soluble intracellular adhesion molecules 1 (sICAM-1), plasma soluble vascular cell adhesion molecules 1 (sVCAM-1) and serum high sensitivity C-reactive protein (hsCRP), the results showed that there were no significant differences ($p > 0.05$) between tocotrienol and placebo interventions, both post-intervention and postprandial except for plasma sICAM-1, in which tocotrienols significantly lowered ($p < 0.05$) the plasma sICAM-1 during postprandial. Post-intervention results also showed that brachial systolic blood pressure (SBP) and aortic pulse pressure were significantly lower ($p < 0.05$) in tocotrienol intervention compared to placebo intervention. In conclusion, this study demonstrated that 14 days of 400 mg tocotrienol supplementation did not exert antithrombotic effects on platelet aggregation induced by ADP and arachidonic acid, thrombin mimic peptide induced platelet activation, and haemostatic and inflammatory measures.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
Sebagai memenuhi keperluan untuk ijazah Master Sains

**KESAN PENGAMBILAN TOKOTRIENOL TERHADAP PENGAGREGATAN
PLATELET DALAM SUBJEK DENGAN SINDROM METABOLIK**

By

GAN YEE LIN

April 2014

Pengerusi : Prof. Lai Oi Ming, PhD

Fakulti : Bioteknologi dan Sains Biomolekul

Pembentukan trombus dalam saluran darah, sama ada dalam arteri atau vena, adalah suatu keadaan patologi yang lazimnya punca kepada kes-kes kejadian penyakit kardiovaskular. Menurut laporan WHO, penyakit kardiovaskular merupakan punca utama kematian bagi kategori penyakit tidak berjangkit. Preskripsi agen antitrombotik (contohnya aspirin, clopidogrel and warfarin) merupakan rawatan yang sering diberi kepada pesakit untuk mengurangkan risiko kematian akibat penyakit kardiovascular. Namun demikian, kewujudan kesan-kesan sampingan agen antitrombotik serta keberkesannya terhadap setiap individu yang berbeza mungkin menghadkan keupayaan agen tersebut dalam pencegahan kejadian penyakit kardiovaskular di seluruh dunia termasuk negara Malaysia. Selain tokoferol, tokotrienol merupakan salah satu kumpulan vitamin E. Kajian praklinikal haiwan telah membuktikan bahawa rawatan tokotrienol dapat menghambat platelet agregasi dan menunjukkan kesan antitrombotik yang signifikan, akan tetapi, hasil kajian dalam kajian klinikal manusia adalah kontroversial. Oleh demikian, matlamat kajian ini adalah untuk menentukan kesan pengambilan tokotrienol terhadap kereaktifan platelet dalam subjek sindrom metabolik, dengan mengambil kira bahawa subjek sindrome metabolik adalah berhubung kiat dengan keadaan pratorbotik akibat peningkatan kereaktifan platelet dan koagulabiliti darah. Justeru, langkah permulaan terapi antitrombotik adalah penting bagi mencegah kes kejadian trombotik. Sehubungan ini, satu kajian klinikal secara rawak, rabun dua pihak, bersilang dan kawalan-plasebo telah dilaksanakan dengan sejumlah 31 subjek sindrom metabolik telah mengambil bahagian dalam kajian ini sehingga akhir. Subject dipilih secara rawak untuk pengambilan dua jenis kapsul yang berlainan secara turutan, di mana satu tempoh rehat sekurang-kurangnya 15 hari telah diberikan selepas rawatan pertama dan sebelum sukarelawan mengambil suplemen jenis kedua. Semasa rawatan, subject mengambil satu kapsul tokotrienol (200 mg) atau plasebo sebanyak dua kali sehari selama 14 hari diikuti dengan kajian pasca prandial. Selepas 14 hari rawatan, hasil kajian menunjukkan tiada perbezaan yang signifikan ($p > 0.05$) dalam kereaktifan pengagregatan platelet dengan ransangan dari asid arakidonik atau ADP antara kumpulan tokotrienol dan plasebo. Penemuan pasca prandial turut menunjukkan bahawa pengambilan tokotrienol tidak memberi apa-apa kesan yang

signifikan ($p > 0.05$) terhadap pengagregatan platelet berbanding dengan plasebo. Didapati tiada perbezaan yang signifikan ($p > 0.05$) antara kumpulan tokotrienol dan plasebo dari segi tahap pengaktifan platelet (dirangsang dengan thrombin tiruan peptida) dan paras biopenanda hemostatik (plasma D-dimer, PAI-1, fibrinogen dan ucOC) selepas pengambilan tokotrienol selama 14 hari dan pasca prandial berbanding dengan plasebo. Selain itu, paras biopenanda keradangan termasuk plasma sP-selectin, sE-selectin, sICAM-1 dan sVCAM-1 dan serum hsCRP tidak berbeza signifikan ($p > 0.05$) selepas 14 hari suplementasi dan pasca prandial antara dua kumpulan kecuali biopenanda plasma sICAM-1 yang dilaporkan parasnya agak rendah semasa pasca prandial secara signifikan ($p < 0.05$) selepas pengambilan tokotrienol berbanding dengan plasebo. Tokotrienol juga didapati menurunkan tekanan darah sistolik brakial dan tekanan nadi aorta secara signifikan ($p < 0.05$) berbanding dengan plasebo selepas pengambilan selama 14 hari. Kesimpulannya, kajian ini menunjukkan bahawa dalam subjek sindrom metabolik, pengambilan 400 mg tokotrienol selama 14 hari tidak mempamerkan kesan antitrombotik terhadap pengagregatan platelet yang dirangsang dengan ADP dan asid arakidonik, pengaktifan platelet yang dirangsang dengan thrombin, dan biopenanda-biopenanda hemostatik dan keradangan.

ACKNOWLEDGEMENT

I would like to express my deepest appreciation to Malaysian Palm Oil Board (MPOB) for giving me the chance to carry out this trial and the facilities and grant support. I would also like to extend my greatest gratitude to my supervisor, Prof. Dr. Lai Oi Ming, and co-supervisors Dr. Chew Boon How, Dr. Kalanithi Nesaretnam and Dr. Fu Ju Yen for your valuable guidance, assistance and encouragement in bringing this project to a successful end. A big thank you to my supervisory committee members for their valuable time spent in aiding my thesis writing. It is also my duty to record my thankfulness to Prof. Dr. Yuen Kah Hay, Dr. Teng Kim Tiu, Dr. Kanga Rani Selvaduray, Dr. Maria Madon, Ms. Puvaneswari a/p Meganathan, staffs in Nutrition Unit and Genomic Unit for their kind assistance and valuable opinions along the project. A special note of thanks to the subjects who had participated in this trial and Hovid Bhd. for the dedication of Tocovid™ *SupraBio*™ 200 mg and placebo capsules. I also wanted to thank my labmates: Mo Shuen Yeing, Chang Lin Faun, Doryn Tan, Nur Afiqah bt Kamaruzzaman and the intern students who had lend their helping hands during screening and trial run of this project. Last but not least, I would like to express my heartfelt appreciation to my family members for the endless support, encouragement and understanding showered along this project.

I certify that a Thesis Examination Committee has met on 14 April 2014 to conduct the final examination of Gan Yee Lin on her thesis entitled “Effects of Tocotrienol Supplementation on Platelet Aggregation in Subjects with Metabolic Syndrome” in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Noorjahan Banu binti Mohammed Alitheen, PhD

Associate Professor
Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Chairman)

Amin bin Ismail, PhD

Professor
Faculty of Medicine and Health Science
Universiti Putra Malaysia
(Internal Examiner)

Loh Su Peng, PhD

Associate Professor
Faculty of Medicine and Health Science
Universiti Putra Malaysia
(Internal Examiner)

Nur Aishah binti Mohd Taib, PhD

Professor
University of Malaya
Malaysia
(External Examiner)

NORITAH OMAR, PhD

Associate Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 23 June 2014

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Lai Oi Ming, PhD

Professor

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Chairman)

Chew Boon How, MMed.

Senior Medical Lecturer

Faculty of Medicine and Health Science

Universiti Putra Malaysia

(Member)

Kalanithi Nesaretnam, PhD

Regional Manager

Product Development and Advisory Services Division

Malaysian Palm Oil Board

(Member)

Fu Ju Yen, PhD

Research Officer

Product Development and Advisory Services Division

Malaysian Palm Oil Board

(Member)

BUJANG KIM HUAT, PhD

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date:

DECLARATION

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____

Date: _____

Name and Matric No.: Gan Yee Lin, GS31357

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature:

Signature:

Name of Chairman of
Supervisory Committee:
Prof. Dr. Lai Oi Ming

Name of Member of Supervisory
Committee:
Dr. Chew Boon How

Signature:

Signature:

Name of Member of Supervisory
Committee:
Dr. Kalanithi Nesaretnam

Name of Member of Supervisory
Committee:
Dr. Fu Ju Yen

TABLE OF CONTENTS

	Page
ABSTRACT	ii
ABSTRAK	iv
ACKNOWLEDGEMENT	vi
APPROVAL	vii
DECLARATION	ix
LIST OF TABLES	xiv
LIST OF FIGURES	xvi
LIST OF APPENDICES	xviii
LIST OF ABBREVIATIONS	xix
CHAPTER	
1 INTRODUCTION	1
2 LITERATURE REVIEW	5
2.1 Tocotrienols	5
2.1.1 Homologues and structures	5
2.1.2 Natural sources	6
2.1.3 Extraction processes	7
2.1.4 Metabolism and adsorption	7
2.2 Thrombosis and its pathogenesis	7
2.2.1 Endothelial injury	8
2.2.2 Abnormal blood flow	9
2.2.3 Hypercoagulability	10
2.3 Platelet function tests	11
2.3.1 Traditional platelet function tests	11
2.3.2 New platelet function test	11
2.3.3 VerifyNow system – a new platelet function test	13
2.4 Metabolic syndrome (MetS)	14
2.4.1 Clinical diagnosis	14
2.4.2 MetS components	17
2.4.3 MetS and thrombosis	17
2.5 Antithrombotic effect of vitamin E and its issues	18
2.5.1 Antithrombotic effect of tocopherols	18
2.5.2 Issues of tocopherol supplementation	21
2.5.3 Antithrombotic effect of tocotrienols	21
2.5.3.1 Animal and <i>in vitro</i> studies – antithrombotic effect	22
2.5.3.2 Human studies – antithrombotic effect	27
2.5.3.3 Animal studies – contradict findings	29
2.5.3.4 Human studies – contradict findings	29

2.5.4	Issue of antithrombotic effect of tocotrienols	34
3	MATERIALS AND METHODS	35
3.1	Materials	35
3.1.1	Test compounds	35
3.1.2	High fat breakfast	35
3.1.3	Blood sampling materials	36
3.1.4	Reagents and chemicals	36
3.2	Study Design	36
3.2.1	Description of interventions	37
3.2.2	Double blinding and decoding of interventions	38
3.2.3	Sample size calculation	38
3.2.4	Subjects' eligibility	38
3.2.5	Ethical approval	39
3.2.6	Recruitment methodology	39
3.2.7	Randomisation of subjects	40
3.2.8	Vitamin E-controlled high fat breakfast	40
3.2.8.1	Preparation of vitamin E-stripped palm olein	42
3.2.8.2	Preparation of a high fat breakfast	42
3.3	Standard physical measurement protocols	43
3.3.1	Standing height measurement	43
3.3.2	Waist circumference measurement	43
3.3.3	Body weight measurement	43
3.3.4	Blood pressure measurement	44
3.4	Blood sampling	44
3.4.1	Venepuncture method	44
3.4.2	Intravenous cannulation method	44
3.5	Blood handling	45
3.6	Compliance measurement	46
3.7	Energy intake estimation	46
3.8	Laboratory analyses	46
3.8.1	Vitamin E analysis	46
3.8.1.1	High performance liquid chromatography (HPLC) methodology	46
3.8.1.2	Preparation of standard solutions	46
3.8.1.3	Sample preparation	47
3.8.1.4	Method validation	47
3.8.2	Fatty acids composition analysis	48
3.8.2.1	Gas chromatographic methodology	48
3.8.2.2	Sample preparation	49
3.8.3	Triacylglycerol (TAG) composition analysis	49
3.8.3.1	HPLC methodology	49
3.8.3.2	Sample preparation	49
3.8.4	VerifyNow Aspirin and P2Y12 analyses	50
3.8.5	Platelet activation analysis	50

3.8.6	Plasma and serum analyses	50
3.8.7	Haemodynamic measurement	51
3.9	Safety and tolerance analysis	51
3.10	Statistical analysis	52
4	RESULTS	53
4.1	Validation of vitamin E HPLC quantification analysis	53
4.2	Dietary compounds	56
4.2.1	Dietary supplements	56
4.2.2	Vitamin E-stripped palm olein	56
4.3	Demographic characteristics	58
4.4	Compliance	60
4.5	Post-intervention results	62
4.5.1	Platelet aggregation	62
4.5.2	Platelet activation	63
4.5.3	Haemostatic markers	64
4.5.4	Inflammatory markers	65
4.5.5	Serum lipids and lipoproteins	66
4.5.6	Haemodynamic markers	67
4.6	Postprandial results	68
4.6.1	Platelet aggregation	69
4.6.2	Platelet activation	70
4.6.3	Haemostatic markers	70
4.6.4	Inflammatory markers	71
4.6.5	Haemodynamic markers	73
4.7	Safety and tolerance	74
5	DISCUSSION	77
5.1	Post-intervention	78
5.1.1	Platelet aggregation and activation	78
5.1.2	Haemostatic markers	81
5.1.3	Inflammatory markers	82
5.1.4	Serum lipids and lipoproteins	84
5.1.5	Haemodynamic markers	85
5.2	Postprandial	86
5.3	Implication of the findings	88
5.4	Safety and tolerance	89
6	SUMMARY, CONCLUSION, LIMITATION AND RECOMMENDATIONS FOR FUTURE RESEARCH	91
6.1	Summary	91
6.2	Conclusion	91
6.3	Recommendations for future work	92
	REFERENCES	93
	APPENDICES	113
	BIODATA OF STUDENT	144
	LIST OF PUBLICATIONS	145

LIST OF TABLES

Table		Page
2.1	Hypercoagulable states	10
2.2	New platelet function tests	11
2.3	Metabolic syndrome definitions	15
2.4	The antithrombotic effect of tocotrienols in animal and <i>in vitro</i> studies.	25
2.5	The antithrombotic effect of tocotrienols in human studies.	28
2.6	The oppose findings for the antithrombotic effect of tocotrienols in animal studies.	30
2.7	The oppose findings for the antithrombotic effect of tocotrienols in human studies.	32
3.1	The materials used for the preparation of high fat breakfast.	35
3.2	The materials used in blood sampling.	36
3.3	Biochemical analyses performed during screening.	40
3.4	Orthogonal design for random participant intervention orders.	40
3.5	Estimated energy and macronutrient of muffin and milkshake.	41
3.6	Conditions of short path distillatory.	42
3.7	Amount of ingredients needed to bake one high fat muffin.	43
3.8	The temperature settings of the oven in gas chromatography system.	48
3.9	The evaporative light scattering detector's setting.	49
4.1	Method validation for quantification of vitamin E in lipid matrices.	54
4.2	LOD and LOQ values for quantification of vitamin E in lipid matrices.	55
4.3	Intra-assay and inter-assay precision and accuracy of the assay for vitamin E quantification in lipid matrices.	55

4.4	Method validation for quantification of vitamin E in human plasma.	55
4.5	LOD and LOQ values for quantification of vitamin E in human plasma.	56
4.6	Intra-assay and inter-assay precision and accuracy of the assay for vitamin E quantification in human plasma.	56
4.7	Concentration of tocopherols and tocotrienols in palm olein and vitamin E-stripped palm olein.	57
4.8	Fatty acids composition in palm olein and vitamin E-stripped palm olein.	57
4.9	TAG composition in palm olein and vitamin E-stripped palm olein.	58
4.10	Physical and biochemical characteristics of study population.	60
4.11	Post-intervention of tocotrienol and placebo supplementations on fasting plasma concentration of D-dimer, fibrinogen, PAI-1 and ucOC.	65
4.12	Post-intervention effect of tocotrienol and placebo supplementations on fasting inflammatory markers.	66
4.13	Fasting serum lipid and lipoprotein concentrations after 14 days of tocotrienol and placebo supplementations.	67
4.14	Post-intervention of brachial blood pressure measurement.	67
4.15	Post-intervention of haemodynamic markers.	68
4.16	Postprandial change of inflammatory markers.	73
4.17	Postprandial change of haemodynamic markers.	74
4.18	Fasting haematological profile after 14 days supplementation of mixed tocotrienols and placebo capsules.	75
4.19	Fasting liver function profile after 14 days supplementation of mixed tocotrienols and placebo capsules.	76

LIST OF FIGURES

Figure		Page
2.1	Molecular structures of tocotrienols and tocopherols.	6
2.2	The endothelial injury that contributes to the development of thrombosis.	8
2.3	The abnormal vasculature and blood flow that contribute to the development of arterial and venous thrombosis.	9
2.4	The principle measurement of VerifyNow system.	13
2.5	Correlation of MetS components leads towards cardiovascular events.	17
3.1	Outline of the study design.	37
4.1	HPLC chromatogram of vitamin E composition in human blank plasma.	53
4.2	HPLC chromatogram of vitamin E composition in human plasma spiked with internal standard and vitamin E mix standards.	54
4.3	Consort diagram of study.	59
4.4	Post-intervention concentration of plasma tocotrienols (n = 31; 15 males and 16 females). Intervention effect was examined using Wilcoxon Signed Rank test.	61
4.5	Postprandial concentration of plasma tocotrienols (n = 31; 15 males and 16 females). Intervention effect was examined using Wilcoxon Signed Rank test.	61
4.6	Post-intervention results of platelet function values of PRU (n = 31; 15 males and 16 females). Intervention effect was examined using Student t-test.	63
4.7	Post-intervention results of platelet function values of ARU (n = 31; 15 males and 16 females). Interventions effect was examined using Student t-test.	63
4.8	Dot plot of platelet activation analysis.	64

4.9	Post-intervention results of MFI of platelet activation (n = 28; 14 males and 14 females). Data was logarithmic transformed prior to statistical analysis of Student t-test to determine the intervention effect. There were 28 subjects' samples available for platelet activation analysis due to the electric cutoff event happened during samples analysis.	64
4.10	Postprandial change of platelet function values of PRU (n = 31; 15 males and 16 females). Intervention effect was examined using Student t-test.	69
4.11	Postprandial change of platelet function values of ARU (n = 31; 15 males and 16 females). Intervention effect was examined using Wilcoxon Signed Rank test.	69
4.12	Postprandial change of platelet function values of MFI (n = 28; 14 males and 14 females). Intervention effect was examined using Student t-test. There were 28 subjects' samples available for platelet activation analysis due to the electric cutoff event happened during samples analysis.	70
4.13	Postprandial response of plasma PAI-1 concentrations (n = 31; 15 males and 16 females). Data was logarithmic transformed prior to statistical analysis using repeated measures generalised linear model.	71
4.14	Postprandial change of plasma D-dimer concentrations (n = 31; 15 males and 16 females). Intervention effect was examined using Wilcoxon Signed Rank test.	71
4.15	Postprandial response of plasma sP-selectin concentrations (n = 31; 15 males and 16 females). Data was examined using repeated measures generalised linear model.	72
7.1	HPLC chromatogram of vitamin E composition in human plasma at 4 h after 200 mg of mixed tocotrienols administration.	143

LIST OF APPENDICES

Appendix		Page
1	Individual calendar	113
2	Application record sheet	114
3	Dietary guidelines (English and Malay versions)	115
4	Deviation protocol record sheet	117
5	Diet diary	119
6	Ethics approval letter from JKEUPM, Universiti Putra Malaysia	123
7	Advertisements (English and Malay versions)	124
8	Screening Questionnaire (English and Malay versions)	126
9	Subject's information sheet (English and Malay versions)	130
10	Consent form (English and Malay versions)	136
11	Screening record sheet	138
12	Certificate of analysis of mixed tocotrienols-tocopherols complex 50% oil suspension standard	139
13	Certificate of analysis of Tocovid™ <i>Suprabio</i> ™ 200 mg (batch no: 11901BBA)	140
14	Certificate of analysis of placebo capsules (batch no: 11902BBA)	141
15	HPLC chromatogram of vitamin E composition in human plasma at 4 h after 200 mg of mixed tocotrienols administration	143

LIST OF ABBREVIATIONS

ADP	Adenosine 5'-diphosphate
AIx@75	Augmentation index adjusted to heart rate at 75
ALP	Alkaline phosphatase
AIx	Augmentation index
AMP	Adenosine monophosphate
Apo A1	Apolipoprotein A1
Apo B100	Apolipoprotein B100
ARU	Aspirin reactivity unit
ATP	Adenosine triphosphate
cAMP	Cyclic adenosine monophosphate
CRP	C-reactive protein
CVD	Cardiovascular disease
DBP	Diastolic blood pressure
DP	Diastolic pressure
EDTA	Ethylenediaminetetraacetic acid
ELISA	Enzyme-linked immunosorbent assay
FAME	Fatty acid methyl ester
FITC	Fluorescein isothiocyanate
GGT	Gamma-glutanyl transpeptidase
GP IIb/IIIa	Glycoprotein IIb/IIIa
HAECs	Human aortic endothelial cells
Hb	Haemoglobin
HDL-C	High density lipoprotein cholesterol

HMG-CoA	β -hydroxy- β -methylglutaryl-coenzyme A
HPLC	High performance liquid chromatography
hsCRP	High sensitivity C-reactive protein
HUVEC	Human umbilical vein endothelial cells
ICAM-1	Intracellular adhesion molecule 1
IDF	International Diabetes Federation
IOM	Institute of Medicine
iso-TRAP	(DL-Isoser ¹)-thrombin receptor activating peptide-6 trifluoroacetate salt
IV	Intravenous
LDL-C	Low density lipoprotein cholesterol
LLOQ	Lower limit of quantification
LOD	Limit of detection
LOQ	Limit of quantification
MCH	Mean corpuscular haemoglobin
MCHC	Mean corpuscular haemoglobin concentration
MCV	Mean corpuscular volume
MetS	Metabolic syndrome
MFI	Mean fluorescence intensity
NO	Nitric oxide
PAI-1	Plasminogen activator inhibitor type 1
PAR	Protease-activated receptor
PAU	Platelet aggregation unit
PerCP	Peridinin chlorophyll protein complex
PIVKA-II	Protein induced by vitamin K absence-factor II

PKC	Protein kinase C
PMA	Phorbol-12-myristate-13-acetate
PMC	2,2,5,7,8-pentamethyl-6-chromanol
PRP	Platelet rich plasma
PRU	P2Y12 reactivity unit
RBC	Red blood cell
SBP	Systolic blood pressure
SD	Standard deviation
sE-selectin	Soluble E-selectin
SEVR%	Sub-endocardial viability ratio index
SGOT	Serum glutamic oxaloacetic transminase
SGPT	Serum glutamic pyruvic transaminase
sICAM-1	Soluble intracellular adhesion molecules 1
SP	Systolic pressure
sP-selectin	Soluble P-selectin
sVCAM-1	Soluble vascular cell adhesion molecules 1
T1DM	Type 1 diabetes mellitus
T2DM	Type 2 diabetes mellitus
TAG	Triacylglycerol
TC	Total cholesterol
TNF- α	Tumour necrosis factor- α
TRF	Tocotrienol-rich fraction
TxA ₂	Thromboxane A ₂
TxB ₂	Thromboxane B ₂

ucOC	Undercaroxylated osteocalcin
ULOQ	Upper limit of quantification
US FDA	United States Food and Drug Administration
VCAM-1	Vascular cell adhesion molecule 1
VLDL-C	Very low density lipoprotein cholesterol
vWF	von Willebrand factor
WBC	White blood cell
WHO	World Health Organisation
α	Alpha
α -TTP	α -tocopherol transfer protein
β	Beta
δ	Delta
γ	Gamma

CHAPTER 1

INTRODUCTION

Thrombosis is one of the major causes of morbidity, mortality and disability. It is a pathological condition which involves the formation of thrombus in vasculature and thus alters or occludes the normal blood flow. It can take place either in arterial or venous circulation. With respect to arterial thrombosis, thrombus is formed in an area with high shear stress, especially where atherosclerotic plaques are disrupted (Lijfering *et al.*, 2011; Turpie and Esmon, 2011). Several high mortality rate of cardiovascular diseases (CVDs) such as myocardial infarction, cerebrovascular disease and peripheral arterial disease are triggered by arterial thrombosis (Freedman, 2005). On the other hand, venous thrombosis occurs in an intact vein, an area with low blood flow and shear stress. Venous thrombosis is associated with an increased risk of developing deep vein thrombosis or pulmonary embolism (Franchini and Mannucci, 2012; Lijfering *et al.*, 2011). Arterial thrombi comprise predominantly platelet-rich “white thrombi”, whilst venous thrombi consist mainly of red cell-rich “red thrombi”. Nonetheless, both thromboembolic disorders’ thrombi are composed of platelets, fibrin, leukocytes and erythrocytes. Some common pathogenesis biological mechanisms shared between thrombosis of artery and vein included the activation of endothelium, platelets, and coagulation (Franchini and Mannucci, 2012). Recognition of the types of cells, agonists, receptors, enzymes and cellular molecules involved in pathogenesis of thrombosis leads to the major improvement in the development of antithrombotic drugs for the clinical management of patients with thromboembolic disorders.

At present, antithrombotic drugs such as antiplatelets, anticoagulants and fibrinolytic are the standard therapies being applied in the clinical setting for the thrombosis management. They function by targeting the key pathways of coagulation, platelet activation and aggregation including the adenosine 5'-diphosphate (ADP) signalling pathway, thromboxane A₂ (TxA₂) synthesis, activation of glycoprotein IIb/IIIa (GP IIb/IIIa), activity of thrombin and activated factor X, γ -carboxylation of vitamin K and lysis of formed thrombus (Joint Formulary Committee, 2013; Ryan *et al.*, 2013). Some of the antithrombotic drugs have been medicinally available for several decades for the prevention and treatment of arterial and venous thrombosis. However, these antithrombotic drugs possess several drawbacks such as side effects and inter-individual variability in response towards drug's therapeutic effects, may limit their usage or effectiveness (Gross and Weitz, 2009; Sikka and Bindra, 2010; Turpie and Esmon, 2011).

The observation of high residual platelet reactivity after antithrombotic therapies reported in published evidences has indicated that these drugs are insufficient to combat the thromboembolic disorders (Michelson, 2010; Musallam *et al.*, 2011; Sweeny *et al.*, 2009). This may be due to the inter-individual response towards the therapeutic effects of antithrombotic drugs. Growing body of evidences have shown

that some patients have low response towards antiplatelet or anticoagulant therapies of aspirin, clopidogrel, warfarin or heparin (Ancrenaz *et al.*, 2010; Cambria-Kiely and Gandhi, 2002; Hirsh and Raschke, 2004; Musallam *et al.*, 2011; Sinxadi and Blockman, 2008; Sweeny *et al.*, 2009). Similar situation also observed in Malaysia. A study conducted by Ibrahim *et al.* (2013) reported that there were 4.69% low responders for aspirin therapy and 21.9% low responders for clopidogrel therapy in acute coronary syndrome patients. Yet, the mechanisms causing this low response towards antithrombotic drugs have not been fully elucidated. Several potential explanations have been recognised. These included the genetic polymorphisms, pharmacodynamic and pharmacokinetic of drugs interaction and high platelet reactivity (Acikel and Akdemir, 2010; Ancrenaz *et al.*, 2010; Sikka and Bindra, 2010). By taking into consideration the risk-benefit ratio assessment, the presence of side effects of each antithrombotic drug has also limits its usage for the prevention and treatment of thrombosis (Gross and Weitz, 2009; Joint Formulary Committee, 2013; Michelson, 2010; Ryan *et al.*, 2013). Thus, development of new safe and effective antithrombotic agents to strengthen the health care system is an ongoing effort.

Vitamin E, a plant-derived tocopherol vitamin with antioxidant characteristic, is found to be able to improve cardiovascular outcome (Stephens *et al.*, 1996). More interestingly, tocotrienols, a group of vitamin E beyond the commonly known tocopherols, are previously found to possess antithrombotic properties. Previous animal studies have shown that tocotrienols were able to inhibit collagen and ADP induced platelet aggregation and its effects were significantly better than α -tocopherol (Qureshi *et al.*, 2011). Further, in human study, supplementation of tocotrienols has been reported to decrease the collagen and ADP induced platelet aggregation significantly (Qureshi *et al.*, 1991a). In addition, several animal studies also have demonstrated that tocotrienols were able to reduce the platelet activation biomarkers such as plasma thromboxane B₂ (TxB₂) and platelet factor 4 (Qureshi *et al.*, 1991b; 2001a; Qureshi and Peterson, 2001). Similar results were shown by Qureshi *et al.* (1995; 1997) in several other human studies where four weeks of tocotrienol supplementation have significantly decreased the levels of TxB₂ and platelet factor 4 in hypercholesterolemic subjects.

Meanwhile, contradictory findings have been published as well. Different groups of researchers have demonstrated that tocotrienols were not able to inhibit the collagen, adrenaline or ADP induced platelet aggregation in animal and human studies (Koba *et al.*, 1992; Mensink *et al.*, 1999; Tomeo *et al.*, 1995; Wahlqvist *et al.*, 1992; Watkins *et al.*, 1993). Additionally, measurement of platelet activation biomarkers such as plasma and urinary TxB₂ and platelet adenosine triphosphate (ATP) release has shown no difference after tocotrienol supplementation (Mensink *et al.*, 1999; Tomeo *et al.*, 1995; Wahlqvist *et al.*, 1992; Watkins *et al.*, 1993). Hence, the antithrombotic effect of tocotrienols remains uncertain with these inconclusive findings.

Based on the published tocotrienols human studies, it was found that dosages of tocotrienols between the previous human studies were different. Significant suppressive effect on platelet aggregation was observed after four weeks of palm tocotrienol-rich fraction (TRF) supplementation at a daily dose of 200 mg (Qureshi *et al.*, 1991a). Conversely, administration of palm TRF at a daily dose ranged from 160 mg up to 336 mg had showed no significant inhibitory effect on platelet aggregation (Mensink *et al.*, 1999; Tomeo *et al.*, 1995; Wahlqvist *et al.*, 1992). Based on the study conducted by Qureshi *et al.* (2011) in dogs, dosage used in this study was 10 mg TRF/kg of body weight. When the dosage was translated to human equivalent dose with an assumption of 75 kg for human's body weight, it was found that the tocotrienol human equivalent dose was 405 mg. Thus, summarizing from the above studies, in order to ascertain the antithrombotic effect of tocotrienols in current study, a higher dose of 400 mg tocotrienols could be supplemented in humans.

Apart from the dosage of tocotrienols, platelet aggregation methods were also varied between studies. The studies which used whole blood impedance platelet aggregometry for platelet aggregation analyses had demonstrated that tocotrienols had no significant inhibition effect on platelet aggregation. In contrast, previous studies which used light transmission turbidimetric platelet aggregometry showed that tocotrienols significantly reduced ADP and collagen induced platelet aggregation. Light transmission turbidimetric platelet aggregometry is a method that requires a complex sample preparation which may alter platelet behaviour, and a trained operator to perform the test (Harrison, 2005; Harrison *et al.*, 2007). Besides that, results obtained were not reproducible even in the same laboratory (Musallam *et al.*, 2011). On the other hand, whole blood impedance platelet aggregometry also was reported to be poor in standardised method and thus it was difficult to compare findings between the studies (Harrison, 2005). In order to address the drawbacks of these methods, it is crucial to adopt a universal standardised method that requires no sample processing and skillful operator in addition to its ability to produce reproducible results. VerifyNow instrument is a fully automated cartridge-based instrument that fulfills these desirable attributes (Harrison *et al.*, 2007; Harrison and Keeling, 2007).

In addition, previous studies had been focused only on the investigation of the long term tocotrienol supplementation effect on the fasting measurement of platelet aggregation. There is lack of study looking into the postprandial effect of tocotrienols on platelet aggregation and thrombotic markers. In fact, humans spend most of their waking hours in postprandial state which triggers a series of biochemical events including platelet activation, hypercoagulable state and endothelial dysfunction (O'Keefe and Bell, 2007).

In view of these gaps in current research, further exploration which includes a daily dose of 400 mg tocotrienols and use of VerifyNow instrument for platelet aggregation measurement is thus needed to fully understand the potential of tocotrienols being the antithrombotic agent and sort out the pathway that tocotrienols are targeting in reducing the thrombotic events. This needs to consider not only the

fasting measurement after a period of supplementation but also postprandially. Seeing that tocotrienol is a natural product extracted from plant, it may be beneficial as a potential supplement to enhance the cardiovascular health and simultaneously circumventing the side effects associated with the current antithrombotic drugs.

As such, this randomised double-blind, crossover and placebo-controlled human clinical trial was designed to determine the antithrombotic effect of tocotrienols in subjects with metabolic syndrome (MetS) at a daily dose of 400 mg. MetS subjects are associated with abdominal obesity, hypertension, atherogenic dyslipidemia, insulin resistance, prothrombotic state and proinflammatory state. This series of conditions alter the haemostasis balance by increasing the platelet reactivity and inducing hypercoagulability and hypofibrinolytic states. Hence, an increased risk of cardiovascular diseases in MetS subjects may be attributed by its high tendency of thrombus formation. Prevention of thrombotic event is thus important for this group of people.

The specific objectives of this study are:

- i. To ascertain the effect of tocotrienol supplementation (400 mg) on platelet aggregation in subjects with MetS.
- ii. To find out the antithrombotic mechanism of tocotrienols via the platelet activation pathway, coagulation pathway, inflammatory pathway, haemodynamic improvement or changes in lipid profile in subjects with MetS.
- iii. To determine the postprandial effect of tocotrienol supplementation on the platelet aggregation and activation, coagulation, inflammatory and haemodynamic markers in subjects with MetS.
- iv. To determine the safety and tolerance of tocotrienol supplementation in subjects with MetS.

REFERENCES

- Abe, Y., El-Masri, B., Kimball, K.T., Pownall, H., Reilly, C.F., Osmundsen, K., Smith, C.W., and Ballantyne, C.M. 1998. Soluble cell adhesion molecules in hypertriglyceridemia and potential significance on monocyte adhesion. *Arteriosclerosis, Thrombosis, and Vascular Biology*. 18:723-731.
- Acikel, S., and Akdemir, R. 2010. The relationship between inflammation, platelet activation and antiplatelet resistance. *Inflammation and Allergy Drug Targets*. 9:364-381.
- Aggarwal, B.B., Sundaram, C., Prasad, S., and Kannappan, R. 2010. Tocotrienols, the vitamin E of the 21st century: its potential against cancer and other chronic diseases. *Biochemical Pharmacology*. 80:1613-1631.
- Ahuja, K.D., Robertson, I.K., and Ball, M.J. 2009. Acute effects of food on postprandial blood pressure and measures of arterial stiffness in healthy humans. *The American Journal of Clinical Nutrition*. 90:298-303.
- Alberti, K.G., and Zimmet, P.Z. 1998. Definition, diagnosis and classification of diabetes mellitus and its complications. Part 1: diagnosis and classification of diabetes mellitus provisional report of a WHO consultation. *Diabetic Medicine: A Journal of the British Diabetic Association*. 15:539-553.
- Alberti, K.G.M.M., Zimmet, P., and Shaw, J. 2006. Metabolic syndrome—a new world-wide definition. A consensus statement from the International Diabetes Federation. *Diabetic Medicine*. 23:469-480.
- Alessi, M.C., and Juhan-Vague, I. 2006. PAI-1 and the metabolic syndrome: links, causes, and consequences. *Arteriosclerosis, Thrombosis, and Vascular Biology*. 26:2200-2207.
- Alessi, M.C., and Juhan-Vague, I. 2008. Metabolic syndrome, haemostasis and thrombosis. *Thrombosis and Haemostasis*. 99:995-1000.
- Altieri, D.C., and Edgington, T.S. 1988. The saturable high affinity association of factor X to ADP-stimulated monocytes defines a novel function of the Mac-1 receptor. *Journal of Biological Chemistry*. 263:7007-7015.
- Ancrenaz, V., Daali, Y., Fontana, P., Besson, M., Samer, C., Dayer, P., and Desmeules, J. 2010. Impact of genetic polymorphisms and drug-drug interactions on clopidogrel and prasugrel response variability. *Current Drug Metabolism*. 11:667-677.

- Armstrong, A.W., and Golan, D.E. 2007. Pharmacology of hemostasis and thrombosis. *In Principles of pharmacology: the pathophysiologic basis of drug therapy.* D.E. Golan, A.H. Tashjian, E.J. Armstrong, and A.W. Armstrong, editors. Lippincott Williams Wilkins, Philadelphia, USA. pp 388-419.
- Badimon, L., and Vilahur, G. 2012. LDL-cholesterol versus HDL-cholesterol in the atherosclerotic plaque: inflammatory resolution versus thrombotic chaos. *Annals of the New York Academy of Sciences.* 1254:18-32.
- Bairati, I., Meyer, F., Jobin, E., Gélinas, M., Fortin, A., Nabid, A., Brochet, F., and Têtu, B. 2006. Antioxidant vitamins supplementation and mortality: a randomized trial in head and neck cancer patients. *International Journal of Cancer.* 119:2221-2224.
- Balkau, B., and Charles, M.A. 1999. Comment on the provisional report from the WHO consultation. European Group for the Study of Insulin Resistance (EGIR). *Diabetic Medicine: A Journal of the British Diabetic Association.* 16:442-443.
- Bardhan, J., Chakraborty, R., and Raychaudhuri, U. 2011. The 21st century form of vitamin E-tocotrienol. *Current Pharmaceutical Design.* 17:2196-2205.
- Batty, P., and Smith, J.G. 2010. Haemostasis. *Surgery (Medicine Publishing).* 28:530-535.
- Becker, R.C. 2008. Platelet biology the role of platelets in hemostasis, thrombosis and inflammation. *In Platelets in cardiovascular disease.* D.L. Bhatt, editor. Imperial College Press, London, U.K. pp 1-36.
- Binkley, N.C., Krueger, D.C., Engelke, J.A., Foley, A.L., and Suttie, J.W. 2000. Vitamin K supplementation reduces serum concentrations of under- γ -carboxylated osteocalcin in healthy young and elderly adults. *The American Journal of Clinical Nutrition.* 72:1523-1528.
- Bisoendial, R.J., Kastelein, J.J.P., Peters, S.L.M., Levels, J.H.M., Birjmohun, R., Rotmans, J.I., Hartman, D., Meijers, J.C.M., Levi, M., and Stroes, E.S.G. 2007. Effects of CRP infusion on endothelial function and coagulation in normocholesterolemic and hypercholesterolemic subjects. *Journal of Lipid Research.* 48:952-960.
- Bjelakovic, G., Nikolova, D., Glud, L.L., Simonetti, R.G., and Glud, C. 2007. Mortality in randomized trials of antioxidant supplements for primary and secondary prevention: systematic review and meta-analysis. *The Journal of the American Medical Association.* 297:842-857.

- Bjelakovic, G., Nikolova, D., Gluud, L.L., Simonetti, R.G., and Gluud, C. 2012. Antioxidant supplements for prevention of mortality in healthy participants and patients with various diseases. *The Cochrane Database of Systematic Reviews*. 3:CD007176.
- Blann, A.D., Nadar, S.K., and Lip, G.Y.H. 2003. The adhesion molecule P-selectin and cardiovascular disease. *European Heart Journal*. 24:2166-2179.
- Bonfigli, A.R., Pieri, C., Manfrini, S., Testa, I., Sirolla, C., Ricciotti, R., Marra, M., Compagnucci, P., and Testa, R. 2001. Vitamin E intake reduces plasminogen activator inhibitor type 1 in T2DM patients. *Diabetes, Nutrition and Metabolism*. 14:71-77.
- Booth, S.L., Golly, I., Satchek, J.M., Roubenoff, R., Dallal, G.E., Hamada, K., and Blumberg, J.B. 2004. Effect of vitamin E supplementation on vitamin K status in adults with normal coagulation status. *The American Journal of Clinical Nutrition*. 80:143-148.
- Borel, P., Preveraud, D., and Desmarchelier, C. 2013. Bioavailability of vitamin E in humans: an update. *Nutrition Reviews*. 71:319-331.
- Cambria-Kiely, J.A., and Gandhi, P.J. 2002. Aspirin resistance and genetic polymorphisms. *Journal of Thrombosis and Thrombolysis*. 14:51-58.
- Chao, J.T., Gapor, A., and Theriault, A. 2002. Inhibitory effect of delta-tocotrienol, a HMG CoA reductase inhibitor, on monocyte-endothelial cell adhesion. *Journal of Nutritional Science and Vitaminology*. 48:332-337.
- Chin, S.-F., Ibahim, J., Makpol, S., Abdul Hamid, N., Abdul Latiff, A., Zakaria, Z., Mazlan, M., Mohd Yusof, Y., Abdul Karim, A., and Wan Ngah, W. 2011. Tocotrienol rich fraction supplementation improved lipid profile and oxidative status in healthy older adults: a randomized controlled study. *Nutrition and Metabolism*. 8:1-14.
- Chun, J., Lee, J., Ye, L., Exler, J., and Eitenmiller, R.R. 2006. Tocopherol and tocotrienol contents of raw and processed fruits and vegetables in the United States diet. *Journal of Food Composition and Analysis*. 19:196-204.
- Clarke, M.W., Ward, N.C., Wu, J.H.Y., Hodgson, J.M., Puddey, I.B., and Croft, K.D. 2006. Supplementation with mixed tocopherols increases serum and blood cell gamma-tocopherol but does not alter biomarkers of platelet activation in subjects with type 2 diabetes. *The American Journal of Clinical Nutrition*. 83:95-102.
- Clemetson, K.J. 2012. Platelets and primary haemostasis. *Thrombosis Research*. 129:220-224.

- Colette, C., Pares-Herbute, N., Monnier, L.H., and Cartry, E. 1988. Platelet function in type I diabetes: Effects of supplementation with large doses of vitamin E. *The American Journal of Clinical Nutrition*. 47:256-261.
- Collet, J.P., Montalescot, G., Vicaut, E., Ankri, A., Walylo, F., Lesty, C., Choussat, R., Beygui, F., Borentain, M., Vignolles, N., and Thomas, D. 2003. Acute release of plasminogen activator inhibitor-1 in ST-Segment elevation myocardial infarction predicts mortality. *Circulation*. 108:391-394.
- Cook, N.R., Albert, C.M., Gaziano, J.M., Zaharris, E., MacFadyen, J., Danielson, E., Buring, J.E., and Manson, J.E. 2007. A randomized factorial trial of vitamins C and E and beta carotene in the secondary prevention of cardiovascular events in women: results from the women's antioxidant cardiovascular study. *Archives of Internal Medicine*. 167:1610-1618.
- Davi, G., Alessandrini, P., Mezzetti, A., Minotti, G., Bucciarelli, T., Costantini, F., Cipollone, F., Bon, G.B., Ciabattini, G., and Patrono, C. 1997. In vivo formation of 8-epi-prostaglandin F₂ α is increased in hypercholesterolemia. *Arteriosclerosis, Thrombosis, and Vascular Biology*. 17:3230-3235.
- Demerath, E., Towne, B., Blangero, J., and Siervogel, R.M. 2001. The relationship of soluble ICAM-1, VCAM-1, P-selectin and E-selectin to cardiovascular disease risk factors in healthy men and women. *Annals of Human Biology*. 28:664-678.
- Dereska, N.H., McLemore, E.C., Tessier, D.J., Bash, D.S., and Brophy, C.M. 2006. Short-term, moderate dosage vitamin E supplementation may have no effect on platelet aggregation, coagulation profile, and bleeding time in healthy individuals. *The Journal of Surgical Research*. 132:121-129.
- Derhaschnig, U., Testori, C., Riedmueller, E., Aschauer, S., Wolzt, M., and Jilma, B. 2013. Hypertensive emergencies are associated with elevated markers of inflammation, coagulation, platelet activation and fibrinolysis. *Journal of Human Hypertension*. 27:368-373.
- Desideri, G., Croce, G., Marinucci, M.C., Masci, P.G., Stati, M., Valeri, L., Santucci, A., and Ferri, C. 2002a. Prolonged, low dose alpha-tocopherol therapy counteracts intercellular cell adhesion molecule-1 activation. *Clinica Chimica Acta; International Journal of Clinical Chemistry*. 320:5-9.
- Desideri, G., Marinucci, M.C., Tomassoni, G., Masci, P.G., Santucci, A., and Ferri, C. 2002b. Vitamin E supplementation reduces plasma vascular cell adhesion molecule-1 and von Willebrand factor levels and increases nitric oxide concentrations in hypercholesterolemic patients. *The Journal of Clinical Endocrinology and Metabolism*. 87:2940-2945.
- Devaraj, S., Chan, A.V.C., Jr., and Jialal, I. 2002. Alpha-tocopherol supplementation decreases plasminogen activator inhibitor-1 and P-selectin levels in type 2 diabetic patients. *Diabetes Care*. 25:524-529.

- Devaraj, S., Tang, R., Adams-Huet, B., Harris, A., Seenivasan, T., de Lemos, J.A., and Jialal, I. 2007. Effect of high-dose alpha-tocopherol supplementation on biomarkers of oxidative stress and inflammation and carotid atherosclerosis in patients with coronary artery disease. *The American Journal of Clinical Nutrition*. 86:1392-1398.
- Devaraj, S., Xu, D.Y., and Jialal, I. 2003. C-reactive protein increases plasminogen activator inhibitor-1 expression and activity in human aortic endothelial cells: Implications for the metabolic syndrome and atherothrombosis. *Circulation*. 107:398-404.
- Dian, N.L.H.M., Sundram, K., and Idris, N.A. 2007. Effect of chemical interesterification on triacylglycerol and solid fat contents of palm stearin, sunflower oil and palm kernel olein blends. *European Journal of Lipid Science and Technology*. 109:147-156.
- Doggen, C.J.M., Smith, N.L., Lemaitre, R.N., Heckbert, S.R., Rosendaal, F.R., and Psaty, B.M. 2004. Serum lipid levels and the risk of venous thrombosis. *Arteriosclerosis, Thrombosis, and Vascular Biology*. 24:1970-1975.
- Duttaroy, A.K. 2005. Postprandial activation of hemostatic factors: role of dietary fatty acids. *Prostaglandins, Leukotrienes and Essential Fatty Acids*. 72:381-391.
- Dyszkiewicz-Korpanty, A.M., Frenkel, E.P., and Sarode, R. 2005. Approach to the assessment of platelet function: comparison between optical-based platelet-rich plasma and impedance-based whole blood platelet aggregation methods. *Clinical and Applied Thrombosis/Hemostasis*. 11:25-35.
- EFSA. 2008. Opinion on mixed tocopherols, tocotrienol tocopherol and tocotrienols as sources for vitamin E added as a nutritional substance in food supplements. Scientific opinion of the scientific panel on food additives, flavourings, processing aids and materials in contact with food. (Question No EFSA Q-2005-146, Q-2005-172, Q-2006-265). *European Food Safety Authority Journal*. 640:1-34.
- Einhorn, D., Reaven, G.M., Cobin, R.H., Ford, E., Ganda, O.P., Handelsman, Y., Hellman, R., Jellinger, P.S., Kendall, D., Krauss, R.M., Neufeld, N.D., Petak, S.M., Rodbard, H.W., Seibel, J.A., Smith, D.A., and Wilson, P.W.F. 2003. American College of Endocrinology position statement on the insulin resistance syndrome. *Endocrine Practice: Official Journal of the American College of Endocrinology and the American Association of Clinical Endocrinologists*. 9:237-252.
- Elmas, E., Kälsch, T., Suvajac, N., Leweling, H., Neumaier, M., Dempfle, C.-E., and Borggrefe, M. 2007. Activation of coagulation during alimentary lipemia under real-life conditions. *International Journal of Cardiology*. 114:172-175.

- Esmon, C.T. 2005. The interactions between inflammation and coagulation. *British Journal of Haematology*. 131:417-430.
- Evans, H.M. and Bishop, K.S. 1922. On the existence of a hitherto unrecognized dietary factor essential for reproduction. *Science*. 56:650-651.
- Expert Panel on Detection, Evaluation, and Treatment of High Blood Cholesterol in Adults. 2001. Executive summary of the third report of the national cholesterol education program (NCEP) expert panel on detection, evaluation, and treatment of high blood cholesterol in adults (adult treatment panel iii). *The Journal of the American Medical Association*. 285:2486-2497.
- Fairus, S., Nor, R.M., Cheng, H.M., and Sundram, K. 2006. Postprandial metabolic fate of tocotrienol-rich vitamin E differs significantly from that of α -tocopherol. *The American Journal of Clinical Nutrition*. 84:835-842.
- Ferroni, P., Martini, F., Riondino, S., La Farina, F., Magnapera, A., Ciatti, F., and Guadagni, F. 2009. Soluble P-selectin as a marker of in vivo platelet activation. *Clinica Chimica Acta*. 399:88-91.
- Fischbach, F., and Dunning, M.B. 2009. A manual of laboratory and diagnostic tests. Wolters Kluwer Health/Lippincott Williams & Wilkins, Philadelphia. pp 57-183.
- Fong, J.S. 1976. Alpha-tocopherol: its inhibition on human platelet aggregation. *Experientia*. 32:639-641.
- Franchini, M., and Mannucci, P.M. 2012. Association between venous and arterial thrombosis: clinical implications. *European Journal of Internal Medicine*. 23:333-337.
- Franchini, M., Targher, G., Montagnana, M., and Lippi, G. 2008. The metabolic syndrome and the risk of arterial and venous thrombosis. *Thrombosis Research*. 122:727-735.
- Freedman, J.E. 2005. Molecular regulation of platelet-dependent thrombosis. *Circulation*. 112:2725-2734.
- Freedman, J.E., Farhat, J.H., Loscalzo, J., and Keaney, J.F., Jr. 1996. Alpha-tocopherol inhibits aggregation of human platelets by a protein kinase C-dependent mechanism. *Circulation*. 94:2434-2440.
- Frimodt-Møller, M., Nielsen, A.H., Kamper, A.L., and Strandgaard, S. 2008. Reproducibility of pulse-wave analysis and pulse-wave velocity determination in chronic kidney disease. *Nephrology Dialysis Transplantation*. 23:594-600.
- Fulop, T., Tessier, D., and Carpentier, A. 2006. The metabolic syndrome. *Pathologie Biologie*. 54:375-386.

- Furman, M.I., Kereiakes, D.J., Krueger, L.A., Mueller, M.N., Broderick, T.M., Schneider, J.F., Howard, W.L., Fox, M.L., Barnard, M.R., Frelinger Iii, A.L., and Michelson, A.D. 2003. Quantification of abciximab-induced platelet inhibition is assay dependent: a comparative study in patients undergoing percutaneous coronary intervention. *American Heart Journal*. 145:M1-M6.
- Gachet, C., and Aleil, B. 2008. Testing antiplatelet therapy. *European Heart Journal Supplements*. 10:A28-A34.
- Gawaz, M. 2001. Blood platelets: physiology, pathophysiology, membrane receptors, antiplatelet principles, and therapy for atherothrombotic diseases. Georg Thieme Verlag, Germany. pp 2-24.
- Gisinger, C., Jeremy, J., Speiser, P., Mikhailidis, D., Dandona, P., and Schernthaner, G. 1988. Effect of vitamin E supplementation on platelet thromboxane A2 production in type I diabetic patients. Double-blind crossover trial. *Diabetes*. 37:1260-1264.
- Gomes, J.A.C., Venkatachalapathy, D., and Haft, J.I. 1976. The effect of vitamin E on platelet aggregation. *American Heart Journal*. 91:425-429.
- Gross, P.L., and Weitz, J.I. 2009. New antithrombotic drugs. *Clinical Pharmacology & Therapeutics*. 86:139-146.
- Grundey, S.M., Cleeman, J.I., Daniels, S.R., Donato, K.A., Eckel, R.H., Franklin, B.A., Gordon, D.J., Krauss, R.M., Savage, P.J., Smith, S.C., Spertus, J.A., and Costa, F. 2005. Diagnosis and management of the metabolic syndrome: an American Heart Association/National Heart, Lung, and Blood Institute scientific statement. *Circulation*. 112:2735-2752.
- Hagberg, I.A., Roald, H.E., and Lyberg, T. 1997. Platelet activation in flowing blood passing growing arterial thrombi. *Arteriosclerosis, Thrombosis, and Vascular Biology*. 17:1331-1336.
- Harding, S.A., Sommerfield, A.J., Sarma, J., Twomey, P.J., Newby, D.E., Frier, B.M., and Fox, K.A.A. 2004. Increased CD40 ligand and platelet-monocyte aggregates in patients with type 1 diabetes mellitus. *Atherosclerosis*. 176:321-325.
- Harrison, P. 2005. Platelet function analysis. *Blood Reviews*. 19:111-123.
- Harrison, P., Frelinger Iii, A.L., Furman, M.I., and Michelson, A.D. 2007. Measuring antiplatelet drug effects in the laboratory. *Thrombosis Research*. 120:323-336.
- Harrison, P., and Keeling, D. 2007. Chapter 23 - Clinical tests of platelet function. *In Platelets (Second Edition)*. A.D. Michelson, editor. Academic Press, Burlington. pp 445-474.

- Hayes, K.C., Pronczuk, A., and Liang, J.S. 1993. Differences in the plasma transport and tissue concentrations of tocopherols and tocotrienols: observations in humans and hamsters. *Proceedings of the Society for Experimental Biology and Medicine. Society for Experimental Biology and Medicine (New York, N.Y.)*. 202:353-359.
- Heng, E., Karsani, S., Abdul Rahman, M., Abdul Hamid, N., Hamid, Z., and Wan Ngah, W. 2013. Supplementation with tocotrienol-rich fraction alters the plasma levels of apolipoprotein a-I precursor, apolipoprotein E precursor, and C-reactive protein precursor from young and old individuals. *European Journal of Nutrition*. 52:1811-1820.
- Hirsh, J., and Raschke, R. 2004. Heparin and low-molecular-weight heparin: the seventh accp conference on antithrombotic and thrombolytic therapy. *CHEST Journal*. 126:188S-203S.
- Ho, D.S.S. 2009. Recovery of phytonutrient from oils. *In United States Patent*. Vol. US7544822 B2. Carotech Bhd., United States.
- Ho, D.S.S., Yuen, K.H., and Yap, S.P. 2003. Drug delivery system: formulation for fat-soluble drugs. *In United States Patent*. Vol. US6596306 B1. Ho, D. S. S. and Yuen, K. H. and Yap, S. P., United States.
- Huijgens, P.C., van den Berg, C.A.M., Imandt, L.M.F.M., and Langenhuijsen, M.M.A.C. 1981. Vitamin E and platelet aggregation. *Acta Haematologica*. 65:217-218.
- Ibrahim O, Oteh M, A Syukur A, Che Hassan H H, S Fadilah W, and Rahman, M.M. 2013. Evaluation of aspirin and clopidogrel resistance in patients with acute coronary syndrome by using adenosine diphosphate test and aspirin test. *Pakistan Journal Medical Sciences*. 29:97-102.
- Ima-Nirwana, S., Nurshazwani, Y., Nazrun, A.S., Norliza, M., and Norazlina, M. 2011. Subacute and subchronic toxicity studies of palm vitamin E in mice. *Journal of Medicinal Food*. 7:45-51.
- Institute of Medicine. 2000. Dietary reference intakes for vitamin C, vitamin E, selenium, and carotenoids. National Academy Press, Washington, DC. pp 21-34.
- Jain, S.K., Krueger, K.S., McVie, R., Jaramillo, J.J., Palmer, M., and Smith, T. 1998. Relationship of blood thromboxane-B2 (TxB2) with lipid peroxides and effect of vitamin E and placebo supplementation on TxB2 and lipid peroxide levels in type 1 diabetic patients. *Diabetes Care*. 21:1511-1516.
- Jang, M.J., Choi, W.-i., Bang, S.-M., Lee, T., Kim, Y.-K., Ageno, W., and Oh, D. 2009. Metabolic syndrome is associated with venous thromboembolism in the Korean population. *Arteriosclerosis, Thrombosis, and Vascular Biology*. 29:311-315.

- Jekel, J.F., Katz, D.L., Elmore, J.G., and Wild, D. 2007. Epidemiology, biostatistics and preventive medicine. Saunders Philadelphia, PA, USA. pp 197-212.
- Jerjes-Sanchez, C. 2005. Venous and arterial thrombosis: a continuous spectrum of the same disease? *European Heart Journal*. 26:3-4.
- Joint Formulary Committee. 2013. British national formulary. BMJ Publishing Group Ltd and Royal Pharmaceutical Society, London, U.K. pp 84-176.
- Kayden, H.J., and Traber, M.G. 1993. Absorption, lipoprotein transport, and regulation of plasma concentrations of vitamin E in humans. *Journal of Lipid Research*. 34:343-358.
- Katsanidis, E., and Addis, P.B. 1999. Novel HPLC analysis of tocopherols, tocotrienols, and cholesterol in tissue. *Free Radical Biology and Medicine*. 27:1137-1140.
- Khalili, H., Dayyeh, B., and Friedman, L. 2011. Assessment of liver function in clinical practice. In *Chronic liver failure*. P. Ginès, P.S. Kamath, and V. Arroyo, editors. Humana Press. pp 47-76.
- Khosla, P., Patel, V., Whinter, J.M., Khanna, S., Rakhkovskaya, M., Roy, S., and Sen, C.K. 2006. Postprandial levels of the natural vitamin E tocotrienol in human circulation. *Antioxidants and Redox Signaling*. 8:1059-1068.
- Kim, J.-E., Han, M., Hanl, K.-S., and Kim, H.K. 2011. Vitamin E inhibition on platelet procoagulant activity: involvement of aminophospholipid translocase activity. *Thrombosis Research*. 127:435-442.
- Klein, E.A., Thompson, I.M., Jr., Tangen, C.M., Crowley, J.J., Lucia, M.S., Goodman, P.J., Minasian, L.M., Ford, L.G., Parnes, H.L., Gaziano, J.M., Karp, D.D., Lieber, M.M., Walther, P.J., Klotz, L., Parsons, J.K., Chin, J.L., Darke, A.K., Lippman, S.M., Goodman, G.E., Meyskens, F.L., Jr., and Baker, L.H. 2011. Vitamin E and the risk of prostate cancer: the selenium and vitamin E cancer prevention trial (SELECT). *The Journal of the American Medical Association*. 306:1549-1556.
- Koba, K., Abe, K., Ikeda, I., and Sugano, M. 1992. Effects of alpha-tocopherol and tocotrienols on blood pressure and linoleic acid metabolism in the spontaneously hypertensive rat (SHR). *Bioscience, Biotechnology, and Biochemistry*. 56:1420-1423.
- Koren-Morag, N., Goldbourt, U., and Tanne, D. 2005. Relation between the metabolic syndrome and ischemic stroke or transient ischemic attack: a prospective cohort study in patients with atherosclerotic cardiovascular disease. *Stroke*. 36:1366-1371.

- Kramer, J.G., Blais, L., Fouchard, R., Melnyk, R., and Kallury, K.R. 1997. A rapid method for the determination of vitamin E forms in tissues and diet by high-performance liquid chromatography using a normal-phase diol column. *Lipids*. 32:323-330.
- Kumar, A., Kar, S., and Fay, W.P. 2011. Thrombosis, physical activity, and acute coronary syndromes. *Journal of Applied Physiology*. 111:599-605.
- Kunisaki, M., Umeda, F., Inoguchi, T., Watanabe, J., and Nawata, H. 1990. Effects of vitamin E administration on platelet function in diabetes mellitus. *Diabetes Research (Edinburgh, Scotland)*. 14:37-42.
- Lakka, H., Laaksonen, D.E., Lakka, T.A., Niskanen, L.K., Kumpusalo, E., Tuomilehto, J., Salonen, J.T. 2002. The metabolic syndrome and total and cardiovascular disease mortality in middle-aged men. *The Journal of the American Medical Association*. 288:2709-2716.
- Lau, D.C.W., Yan, H., and Dhillon, B. 2006. Metabolic syndrome: a marker of patients at high cardiovascular risk. *Canadian Journal of Cardiology*. 22, Supplement B:85B-90B.
- Laugesen, E., Rossen, N.B., Høyem, P., Christiansen, J.S., Knudsen, S.T., Hansen, K.W., Hansen, T.K., and Poulsen, P.L. 2013. Reproducibility of pulse wave analysis and pulse wave velocity in patients with type 2 diabetes. *Scandinavian Journal of Clinical and Laboratory Investigation*. 73:428-435.
- Lemkes, B.A., BÄHler, L., Kamphuisen, P.W., Stroobants, A.K., Van Den Dool, E.J., Hoekstra, J.B., Nieuwland, R., Gerdes, V.E., and Holleman, F. 2012. The influence of aspirin dose and glycemic control on platelet inhibition in patients with type 2 diabetes mellitus. *Journal of Thrombosis and Haemostasis*. 10:639-646.
- Libby, P. 2006. Inflammation and cardiovascular disease mechanisms. *The American Journal of Clinical Nutrition*. 83:456S-460S.
- Lijfering, W.M., Flinterman, L.E., Vandenbroucke, J.P., Rosendaal, F.R., and Cannegieter, S.C. 2011. Relationship between venous and arterial thrombosis: a review of the literature from a causal perspective. *Seminars Thrombosis Hemostasis*. 37:885-896.
- Liu, M., Wallmon, A., Olsson-Mortlock, C., Wallin, R., and Saldeen, T. 2003. Mixed tocopherols inhibit platelet aggregation in humans: potential mechanisms. *The American Journal of Clinical Nutrition*. 77:700-706.
- Lonn, E., Bosch, J., Yusuf, S., Sheridan, P., Pogue, J., Arnold, J.M.O., Ross, C., Arnold, A., Sleight, P., Probstfield, J., and Dagenais, G.R. 2005. Effects of long-term vitamin E supplementation on cardiovascular events and cancer: a randomized controlled trial. *The Journal of the American Medical Association*. 293:1338-1347.

- López, J.A., and Chen, J. 2009. Pathophysiology of venous thrombosis. *Thrombosis Research*. 123, Supplement 4:S30-S34.
- Lowe, G.D.O. 2010. Fibrinogen assays for cardiovascular risk assessment. *Clinical Chemistry*. 56:693-695.
- LÜThje, J., Schomburg, A., and Ogilvie, A. 1988. Demonstration of a novel ecto-enzyme on human erythrocytes, capable of degrading ADP and of inhibiting ADP-induced platelet aggregation. *European Journal of Biochemistry*. 175:285-289.
- Mabile, L., Bruckdorfer, K.R., and Rice-Evans, C. 1999. Moderate supplementation with natural alpha-tocopherol decreases platelet aggregation and low-density lipoprotein oxidation. *Atherosclerosis*. 147:177-185.
- Maccarana, M., and Lindahl, U. 1993. Mode of interaction between platelet factor 4 and heparin. *Glycobiology*. 3:271-277.
- Mahalingam, D., Radhakrishnan, A.K., Amom, Z., Ibrahim, N., and Nesaretnam, K. 2011. Effects of supplementation with tocotrienol-rich fraction on immune response to tetanus toxoid immunization in normal healthy volunteers. *European Journal of Clinical Nutrition*. 65:63-69.
- Malinin, A., Pokov, A., Swaim, L., Kotob, M., and Serebruany, V. 2006. Validation of a VerifyNow-P2Y12 cartridge for monitoring platelet inhibition with clopidogrel. *Methods and Findings in Experimental and Clinical Pharmacology*. 28:315-322.
- Martinelli, I., Bucciarelli, P., and Mannucci, P.M. 2010. Thrombotic risk factors: basic pathophysiology. *Critical Care Medicine*. 38:S3-S9.
- Mayer, H., Metzger, J., and Isler, O. 1967. Über die chemie des vitamins E. 8. mitteilung [1]. Die Stereochemie von natürlichem γ -tocotrienol (plastochromanol-3), plastochromanol-8 und plastochromanol-8. *Helvetica Chimica Acta*. 50:1376-1393.
- McRae, M.P. 2006. Is vitamin C an effective antihypertensive supplement? A review and analysis of the literature. *Journal of Chiropractic Medicine*. 5:60-64.
- Mensink, R.P., van Houwelingen, A.C., Kromhout, D., and Hornstra, G. 1999. A vitamin E concentrate rich in tocotrienols had no effect on serum lipids, lipoproteins, or platelet function in men with mildly elevated serum lipid concentrations. *The American Journal of Clinical Nutrition*. 69:213-219.
- Michelson, A.D. 2010. Antiplatelet therapies for the treatment of cardiovascular disease. *Nature Reviews Drug Discovery*. 9:154-169.

- Miller, E.R., 3rd, Pastor-Barriuso, R., Dalal, D., Riemersma, R.A., Appel, L.J., and Guallar, E. 2005. Meta-analysis: high-dosage vitamin E supplementation may increase all-cause mortality. *Annals of Internal Medicine*. 142:37-46.
- Mineo, C., Deguchi, H., Griffin, J.H., and Shaul, P.W. 2006. Endothelial and antithrombotic actions of HDL. *Circulation Research*. 98:1352-1364.
- Ministry of Health Malaysia. 2009. Clinical practice guideline: management of type 2 diabetes mellitus in Malaysia. Ministry of Health Malaysia, Putrajaya, Malaysia. pp 36-37.
- Morange, P.E., and Alessi, M.C. 2013. Thrombosis in central obesity and metabolic syndrome: mechanisms and epidemiology. *Thrombosis and Haemostasis*. 110:669-680.
- Mottillo, S., Filion, K.B., Genest, J., Joseph, L., Pilote, L., Poirier, P., Rinfret, S., Schiffrin, E.L., and Eisenberg, M.J. 2010. The metabolic syndrome and cardiovascular risk: a systematic review and meta-analysis. *Journal of the American College of Cardiology*. 56:1113-1132.
- Mullan, B.A., Young, I.S., Fee, H., and McCance, D.R. 2002. Ascorbic acid reduces blood pressure and arterial stiffness in type 2 diabetes. *Hypertension*. 40:804-809.
- Musallam, K.M., Charafeddine, K., Bitar, A., Khoury, M., Assaad, S., Beresian, J., Alam, S., and Taher, A.T. 2011. Resistance to aspirin and clopidogrel therapy. *International Journal of Laboratory Hematology*. 33:1-18.
- Mustad, V.A., Smith, C.A., Ruey, P.P., Edens, N.K., and DeMichele, S.J. 2002. Supplementation with 3 compositionally different tocotrienol supplements does not improve cardiovascular disease risk factors in men and women with hypercholesterolemia. *The American Journal of Clinical Nutrition*. 76:1237-1243.
- Naito, Y., Shimozaawa, M., Kuroda, M., Nakabe, N., Manabe, H., Katada, K., Kokura, S., Ichikawa, H., Yoshida, N., Noguchi, N., and Yoshikawa, T. 2005. Tocotrienols reduce 25-hydroxycholesterol-induced monocyte-endothelial cell interaction by inhibiting the surface expression of adhesion molecules. *Atherosclerosis*. 180:19-25.
- Nakamura, H., Furukawa, F., Nishikawa, A., Miyauchi, M., Son, H.Y., Imazawa, T., and Hirose, M. 2001. Oral toxicity of a tocotrienol preparation in rats. *Food and Chemical Toxicology*. 39:799-805.
- Nesaretnam, K., Selvaduray, K.R., Abdul Razak, G., Veerasenan, S.D., and Gomez, P.A. 2010. Effectiveness of tocotrienol-rich fraction combined with tamoxifen in the management of women with early breast cancer: a pilot clinical trial. *Breast Cancer Research: BCR*. 12:R81-R81.

- Ng, M.H., and Yuen May, C. 2012. Chromatographic analyses of tocopherols and tocotrienols in palm oil. *Journal of Chromatographic Science*. 50:283-286.
- Nicholson, N.S., Panzer-Knodle, S.G., Haas, N.F., Taite, B.B., Szalony, J.A., Page, J.D., Feigen, L.P., Lansky, D.M., and Salyers, A.K. 1998. Assessment of platelet function assays. *American Heart Journal*. 135:S170-S178.
- Nielsen, H.L., Kristensen, S.D., Thygesen, S.S., Mortensen, J., Pedersen, S.B., Grove, E.L., and Hvas, A.-M. 2008. Aspirin response evaluated by the VerifyNow™ aspirin system and light transmission aggregometry. *Thrombosis Research*. 123:267-273.
- Nieuwdorp, M., Stroes, E.S.G., Meijers, J.C.M., and Büller, H. 2005. Hypercoagulability in the metabolic syndrome. *Current Opinion in Pharmacology*. 5:155-159.
- Norsidah, K.-Z., Asmadi, A.Y., Azizi, A., Faizah, O., and Kamisah, Y. 2013. Palm tocotrienol-rich fraction improves vascular proatherosclerotic changes in hyperhomocysteinemic rats. *Evidence-Based Complementary and Alternative Medicine: Ecam*. 2013:976967-976967.
- O'Byrne, D., Grundy, S., Packer, L., Devaraj, S., Baldenius, K., Hoppe, P.P., Kraemer, K., Jialal, I., and Traber, M.G. 2000. Studies of LDL oxidation following alpha-, gamma-, or delta-tocotrienyl acetate supplementation of hypercholesterolemic humans. *Free Radical Biology and Medicine*. 29:834-845.
- O'Keefe, J.H., and Bell, D.S.H. 2007. Postprandial hyperglycemia/hyperlipidemia (postprandial dysmetabolism) is a cardiovascular risk factor. *The American Journal of Cardiology*. 100:899-904.
- O'Rourke, M., and Frohlich, E.D. 1999. Pulse pressure: is this a clinically useful risk factor? *Hypertension*. 34:372-374.
- Ong, S., Xu, C.H., Teo, V.H., Yap, W.N., Chen, A., Lacuesta, V., Zaiden, N., X.W., Z., Shiba, S., and Yap, Y.L. 2010. Comprehensive quantification of palm vitamin E by normal phase high performance liquid chromatography. *Palm Oil Developments*. 52:10-24.
- Oo, S.L., Chang, P., and Chan, K.E. 1992. Toxicological and pharmacological studies on palm vitee. *Nutrition Research*. 12, Supplement 1:S217-S222.
- Oostrom, A.J.H.H.M.v., Rabelink, T.J., Verseyden, C., Sijmonsma, T.P., Plokker, H.W.M., De Jaegere, P.P.T., and Cabezas, M.C. 2004. Activation of leukocytes by postprandial lipemia in healthy volunteers. *Atherosclerosis*. 177:175-182.

- Osakada, F., Hashino, A., Kume, T., Katsuki, H., Kaneko, S., and Akaike, A. 2004. α -Tocotrienol provides the most potent neuroprotection among vitamin E analogs on cultured striatal neurons. *Neuropharmacology*. 47:904-915.
- Packer, L., Weber, S.U., and Rimbach, G. 2001. Molecular aspects of alpha-tocotrienol antioxidant action and cell signalling. *The Journal of Nutrition*. 131:369S-373S.
- Parker, R.A., Pearce, B.C., Clark, R.W., Gordon, D.A., and Wright, J.J. 1993. Tocotrienols regulate cholesterol production in mammalian cells by post-transcriptional suppression of 3-hydroxy-3-methylglutaryl-coenzyme A reductase. *Journal of Biological Chemistry*. 268:11230-11238.
- Patel, V., Rink, C., Gordillo, G.M., Khanna, S., Gnyawali, U., Roy, S., Shneker, B., Ganesh, K., Phillips, G., More, J.L., Sarkar, A., Kirkpatrick, R., Elkhmmas, E.A., Klatter, E., Miller, M., Firstenberg, M.S., Chiocca, E.A., Nesaretnam, K., and Sen, C.K. 2012. Oral tocotrienols are transported to human tissues and delay the progression of the model for end-stage liver disease score in patients. *The Journal of Nutrition*. 142:513-519.
- Patrono, C., Ciabattoni, G., and Davi, G. 1990. Thromboxane biosynthesis in cardiovascular diseases. *Stroke*. 21:IV130-133.
- Patrono, C., and FitzGerald, G.A. 1997. Isoprostanes: potential markers of oxidant stress in atherothrombotic disease. *Arteriosclerosis, Thrombosis, and Vascular Biology*. 17:2309-2315.
- Pennock, J.F., Hemming, F.W., and Kerr, J.D. 1964. A reassessment of tocopherol chemistry. *Biochemical and Biophysical Research Communications*. 17:542-548.
- Praticò, D., Smyth, E.M., Violi, F., and FitzGerald, G.A. 1996. Local amplification of platelet function by 8-epi prostaglandin F₂alpha is not mediated by thromboxane receptor isoforms. *The Journal of Biological Chemistry*. 271:14916-14924.
- Qureshi, A., Bradlow, B., Brace, L., Manganello, J., Peterson, D., Pearce, B., Wright, J., Gapor, A., and Elson, C. 1995. Response of hypercholesterolemic subjects to administration of tocotrienols. *Lipids*. 30:1171-1177.
- Qureshi, A.A., Bradlow, B.A., Salser, W.A., and Brace, L.D. 1997. Novel tocotrienols of rice bran modulate cardiovascular disease risk parameters of hypercholesterolemic humans. *The Journal of Nutritional Biochemistry*. 8:290-298.
- Qureshi, A.A., Karpen, C.W., Qureshi, N., Papasian, C.J., Morrison, D.C., and Folts, J.D. 2011. Tocotrienols-induced inhibition of platelet thrombus formation and platelet aggregation in stenosed canine coronary arteries. *Lipids in Health and Disease*. 10.

- Qureshi, A.A., Pearce, B.C., Nor, R.M., Gapor, A., Peterson, D.M., and Elson, C.E. 1996. Dietary α -tocopherol attenuates the impact of γ -tocotrienol on hepatic 3-hydroxy-3-methylglutaryl coenzyme a reductase activity in chickens. *The Journal of Nutrition*. 126:389-394.
- Qureshi, A.A., and Peterson, D.M. 2001. The combined effects of novel tocotrienols and lovastatin on lipid metabolism in chickens. *Atherosclerosis*. 156:39-47.
- Qureshi, A.A., Peterson, D.M., Hasler-Rapacz, J.O., and Rapacz, J. 2001a. Novel tocotrienols of rice bran suppress cholesterogenesis in hereditary hypercholesterolemic swine. *The Journal of Nutrition*. 131:223-230.
- Qureshi, A., Qureshi, N., Wright, J., Shen, Z., Kramer, G., Gapor, A., Chong, Y., DeWitt, G., Ong, A., and Peterson, D. 1991a. Lowering of serum cholesterol in hypercholesterolemic humans by tocotrienols (palmvitee). *The American Journal of Clinical Nutrition*. 53:1021S-1026S.
- Qureshi, A.A., Qureshi, N., Hasler-Rapacz, J.O., Weber, F.E., Chaudhary, V., Crenshaw, T.D., Gapor, A., Ong, A.S., Chong, Y.H., Peterson, D., and et al. 1991b. Dietary tocotrienols reduce concentrations of plasma cholesterol, apolipoprotein B, thromboxane B2, and platelet factor 4 in pigs with inherited hyperlipidemias. *The American Journal of Clinical Nutrition*. 53:1042S-1046S.
- Qureshi, A.A., Sami, S.A., Salser, W.A., and Khan, F.A. 2001b. Synergistic effect of tocotrienol-rich fraction (TRF25) of rice bran and lovastatin on lipid parameters in hypercholesterolemic humans. *The Journal of Nutritional Biochemistry*. 12:318-329.
- Qureshi, A.A., Sami, S.A., Salser, W.A., and Khan, F.A. 2002. Dose-dependent suppression of serum cholesterol by tocotrienol-rich fraction (TRF25) of rice bran in hypercholesterolemic humans. *Atherosclerosis*. 161:199-207.
- Rasool, A.H.G., Rahman, A.R.A., Yuen, K.H., and Wong, A.R. 2008. Arterial compliance and vitamin E blood levels with a self emulsifying preparation of tocotrienol rich vitamin E. *Archives of Pharmacal Research*. 31:1212-1217.
- Rasool, A.H.G., Yuen, K.H., Yusoff, K., Wong, A.R., and Rahman, A.R.A. 2006. Dose dependent elevation of plasma tocotrienol levels and its effect on arterial compliance, plasma total antioxidant status, and lipid profile in healthy humans supplemented with tocotrienol rich vitamin E. *Journal of Nutritional Science and vitaminology*. 52:473-478.
- Reaven, G.M. 1988. Banting lecture 1988. Role of insulin resistance in human disease. *Diabetes*. 37:1595-1607.
- Reinhart, W.H. 2003. Fibrinogen - marker or mediator of vascular disease? *Vascular Medicine*. 8:211-216.

- Rosenson, R.S., and Lowe, G.D.O. 1998. Effects of lipids and lipoproteins on thrombosis and rheology. *Atherosclerosis*. 140:271-280.
- Russo, I. 2012. The prothrombotic tendency in metabolic syndrome: focus on the potential mechanisms involved in impaired haemostasis and fibrinolytic balance. *Scientifica*. 2012:17.
- Ryan, J., Bolster, F., Crosbie, I., and Kavanagh, E. 2013. Antiplatelet medications and evolving antithrombotic medication. *Skeletal Radiology*. 42:753-764.
- Sanders, T.A.B., Filippou, A., Berry, S.E., Baumgartner, S., and Mensink, R.P. 2011. Palmitic acid in the sn-2 position of triacylglycerols acutely influences postprandial lipid metabolism. *The American Journal of Clinical Nutrition*. 94:1433-1441.
- Sanders, T.B. 2003. Dietary fat and postprandial lipids. *Current Atherosclerosis Reports*. 5:445-451.
- Savoia, C., Sada, L., Zezza, L., Pucci, L., Lauri, F.M., Befani, A., Alonzo, A., and Volpe, M. 2011. Vascular inflammation and endothelial dysfunction in experimental hypertension. *International Journal of Hypertension*. 2011.
- Schauss, A., Endres, J.R., and Clewell, A. 2012. Safety of unsaturated vitamin E tocotrienols and their isomers. In *Tocotrienols: vitamin E beyond tocopherols*. B. Tan, R.R. Watson, and V.R. Preedy, editors. CRC Press and AOCS Press, United States. pp 17-35.
- Sen, C.K., Khanna, S., and Roy, S. 2006. Tocotrienols: vitamin E beyond tocopherols. *Life Sciences*. 78:2088-2098.
- Serebruany, V., Malinin, A., Ong, S., and Atar, D. 2008. Patients with metabolic syndrome exhibit higher platelet activity than those with conventional risk factors for vascular disease. *Journal of Thrombosis and Thrombolysis*. 25:207-213.
- Sikka, P., and Bindra, V. 2010. Newer antithrombotic drugs. *Indian Journal of Critical Care Medicine*. 14:188-195.
- Singh, B., Arora, S., Goswami, B., and Mallika, V. 2009. Metabolic syndrome: A review of emerging markers and management. *Diabetes & Metabolic Syndrome: Clinical Research and Reviews*. 3:240-254.
- Singh, I., Turner, A.H., Sinclair, A.J., Li, D., and Hawley, J.A. 2007. Effects of gamma-tocopherol supplementation on thrombotic risk factors. *Asia Pacific Journal of Clinical Nutrition*. 16:422-428.
- Sinxadi, P., and Blockman, M. 2008. Warfarin resistance. *Cardiovascular Journal Africa*. 19:215-217.

- Smith, A., Patterson, C., Yarnell, J., Rumley, A., Ben-Shlomo, Y., and Lowe, G. 2005. Which hemostatic markers add to the predictive value of conventional risk factors for coronary heart disease and ischemic stroke?: the caerphilly study. *Circulation*. 112:3080-3087.
- Stampfer, M.J., Jakubowski, J.A., Faigel, D., Vaillancourt, R., and Deykin, D. 1988. Vitamin E supplementation effect on human platelet function, arachidonic acid metabolism, and plasma prostacyclin levels. *The American Journal of Clinical Nutrition*. 47:700-706.
- Standeven, K.F., and Grant, P.J. 2011. Atherothrombosis and the metabolic syndrome. *In The metabolic syndrome*. Wiley-Blackwell, New York, USA. pp 194-209.
- Stec, J.J., Silbershatz, H., Tofler, G.H., Matheney, T.H., Sutherland, P., Lipinska, I., Massaro, J.M., Wilson, P.F.W., Muller, J.E., and D'Agostino, R.B. 2000. Association of fibrinogen with cardiovascular risk factors and cardiovascular disease in the Framingham offspring population. *Circulation*. 102:1634-1638.
- Steiner, M. 1983. Effect of alpha-tocopherol administration on platelet function in man. *Thrombosis and Haemostasis*. 49:73-77.
- Steinhubl, S.R. 2007. Chapter 27 - The Verifynow system. *In Platelets (Second Edition)*. A.D. Michelson, editor. Academic Press, Burlington. pp 509-518.
- Stephens, N.G., Parsons, A., Schofield, P.M., Kelly, F., Cheeseman, K., and Mitchinson, M.J. 1996. Randomised controlled trial of vitamin E in patients with coronary disease: cambridge heart antioxidant study (CHAOS). *Lancet*. 347:781-786.
- Sundram, K., Sambanthamurthi, R., and Tan, Y.A. 2003. Palm fruit chemistry and nutrition. *Asia Pacific Journal of Clinical Nutrition*. 12:355-362.
- Sweeny, J.M., Gorog, D.A., and Fuster, V. 2009. Antiplatelet drug 'resistance'. Part 1: mechanisms and clinical measurements. *Nature Reviews Cardiology*. 6:273-282.
- Szczeklik, A., Gryglewski, R.J., Domagala, B., Dworski, R., and Basista, M. 1985. Dietary supplementation with vitamin E in hyperlipoproteinemias: effects on plasma lipid peroxides, antioxidant activity, prostacyclin generation and platelet aggregability. *Thrombosis and Haemostasis*. 54:425-430.
- Tan, B. 2005. Appropriate spectrum vitamin E and new perspectives on desmethyl tocopherols and tocotrienols. *The Journal of the American Nutricutical Association*. 8:35-42.
- Tan, B., and Saleh, M.H. 1992. Integrated process for recovery of carotenoids and tocotrienols from oil. *In United States Patent*. Vol. US5157132 A. Carotech Associates, United States.

- Tan, D.T., Khor, H.T., Low, W.H., Ali, A., and Gapor, A. 1991. Effect of a palm-oil-vitamin E concentrate on the serum and lipoprotein lipids in humans. *The American Journal of Clinical Nutrition*. 53:1027S-1030S.
- Tarrago-Trani, M.T., Phillips, K.M., Lemar, L.E., and Holden, J.M. 2006. New and existing oils and fats used in products with reduced trans-fatty acid content. *Journal of the American Dietetic Association*. 106:867-880.
- Tasaki, M., Umemura, T., Inoue, T., Okamura, T., Kuroiwa, Y., Ishii, Y., Maeda, M., Hirose, M., and Nishikawa, A. 2008. Induction of characteristic hepatocyte proliferative lesion with dietary exposure of wistar hannover rats to tocotrienol for 1 year. *Toxicology*. 250:143-150.
- Therriault, A., Chao, J.-T., and Gapor, A. 2002. Tocotrienol is the most effective vitamin E for reducing endothelial expression of adhesion molecules and adhesion to monocytes. *Atherosclerosis*. 160:21-30.
- Thor, H.T., and Ng, T.T. 2000. Effects of administration of α -tocopherol and tocotrienols on serum lipids and liver HMG CoA reductase activity. *International Journal of Food Sciences and Nutrition*. 51:s3-s11.
- Tomeo, A.C., Geller, M., Watkins, T.R., Gapor, A., and Bierenbaum, M.L. 1995. Antioxidant effects of tocotrienols in patients with hyperlipidemia and carotid stenosis. *Lipids*. 30:1179-1183.
- Top, A.G.M., Leong, L.W., Ong, A.S.H., Kawada, T., Watanabe, H., and Tsuchiya, N. 1993. Production of high concentration tocopherols and tocotrienols from palm-oil by-products. *In United States Patent*. Vol. US5190618 A. Bioindustry Development Centre & Palm Oil Research and Development Board, United States.
- Traber, M.G., Burton, G.W., and Hamilton, R.L. 2004. Vitamin E trafficking. *Annals of the New York Academy of Sciences*. 1031:1-12.
- Turpie, A.G.G., and Esmon, C. 2011. Venous and arterial thrombosis – pathogenesis and the rationale for anticoagulation. *Thrombosis and Haemostasis*. 105:586-596.
- van Dam, B., van Hinsbergh, V.W.M., Stehouwer, C.D.A., Versteilen, A., Dekker, H., Buytenhek, R., Princen, H.M., and Schalkwijk, C.G. 2003. Vitamin E inhibits lipid peroxidation-induced adhesion molecule expression in endothelial cells and decreases soluble cell adhesion molecules in healthy subjects. *Cardiovascular Research*. 57:563-571.
- Wagner, D.D., and Burger, P.C. 2003. Platelets in inflammation and thrombosis. *Arteriosclerosis, Thrombosis, and Vascular Biology*. 23:2131-2137.

- Wahlqvist, M.L., Krivokuca-Bogetic, Z., Lo, C.S., Hage, B., Smith, R., and Lukito, W. 1992. Differential serum responses of tocopherols and tocotrienols during vitamin supplementation in hypercholesterolaemic individuals without change in coronary risk factors. *Nutrition Research*. 12, Supplement 1:S181-S201.
- Waters, D.D., Alderman, E.L., Hsia, J., Howard, B.V., Cobb, F.R., Rogers, W.J., Ouyang, P., Thompson, P., Tardif, J.C., Higginson, L., Bittner, V., Steffes, M., Gordon, D.J., Proschan, M., Younes, N., and Verter, J.I. 2002. Effects of hormone replacement therapy and antioxidant vitamin supplements on coronary atherosclerosis in postmenopausal women: a randomized controlled trial. *The Journal of The American Medical Association*. 288:2432-2440.
- Watkins, M.T., Patton, G.M., Soler, H.M., Albadawi, H., Humphries, D.E., Evans, J.E., and Kadowaki, H. 1999. Synthesis of 8-epi-prostaglandin F2alpha by human endothelial cells: role of prostaglandin H2 synthase. *Biochemical Journal*. 344:747-754.
- Watkins, T., Lenz, P., Gapor, A., Struck, M., Tomeo, A., and Bierenbaum, M. 1993. γ -tocotrienol as a hypocholesterolemic and antioxidant agent in rats fed atherogenic diets. *Lipids*. 28:1113-1118.
- Wichitsranoi, J., Weerapreeyakul, N., Boonsiri, P., Settasatian, C., Settasatian, N., Komanasin, N., Sirijaichingkul, S., Teerajetgul, Y., Rangkadilok, N., and Leelayuwat, N. 2011. Antihypertensive and antioxidant effects of dietary black sesame meal in pre-hypertensive humans. *Nutrition Journal*. 10:82.
- Williams, J.C., Forster, L.A., Tull, S.P., Wong, M., Bevan, R.J., and Ferns, G.A. 1997. Dietary vitamin E supplementation inhibits thrombin-induced platelet aggregation, but not monocyte adhesiveness, in patients with hypercholesterolaemia. *International Journal of Experimental Pathology*. 78:259-266.
- Wolberg, A.S.P.F., Aleman, M.M.B.S., Leiderman, K.P., and Machlus, K.R.P. 2012. Procoagulant activity in hemostasis and thrombosis: virchow's triad revisited. [Review]. *Anesthesia and Analgesia February*. 114:275-285.
- Wong, R.S.Y., and Radhakrishnan, A.K. 2012. Tocotrienol research: past into present. *Nutrition Reviews*. 70:483-490.
- Yap, S.P., and Yuen, K.H. 2004. Influence of lipolysis and droplet size on tocotrienol absorption from self-emulsifying formulations. *International Journal of Pharmaceutics*. 281:67-78.
- Yap, S.P., Yuen, K.H., and Wong, J.W. 2001. Pharmacokinetics and bioavailability of alpha-, gamma- and delta-tocotrienols under different food status. *The Journal of Pharmacy and Pharmacology*. 53:67-71.

Yuen, K.H., Wong, J., W., Lim, A.B., Ng, B.H., and Choy, W.P. 2011. Effect of mixed-tocotrienols in hypercholesterolemic subjects. *Functional Foods in Health Disease*. 3:106–117.

Zaiden, N., Yap, W.N., Ong, S., Xu, C.H., Teo, V.H., Chang, C.P., Zhang, X.W., Nesaretnam, K., Shiba, S., and Yap, Y.L. 2010. Gamma delta tocotrienols reduce hepatic triglyceride synthesis and VLDL secretion. *Journal of Atherosclerosis and Thrombosis*. 17:1019-1032.

