



**UNIVERSITI PUTRA MALAYSIA**

***FUNCTIONAL CHARACTERIZATION OF ALCOHOL  
DEHYDROGENASE GENES IN ARABIDOPSIS PLANTS  
GROWN UNDER DROUGHT CONDITION***

**THAWDA MYINT**

**FBSB 2013 47**



**UPM**  
UNIVERSITI PUTRA MALAYSIA  
BERILMU BERBAKTI

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CONDITION**

**By**

**THAWDA MYINT**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of  
the Requirement for the Degree of Doctor of Philosophy**

**June, 2013**

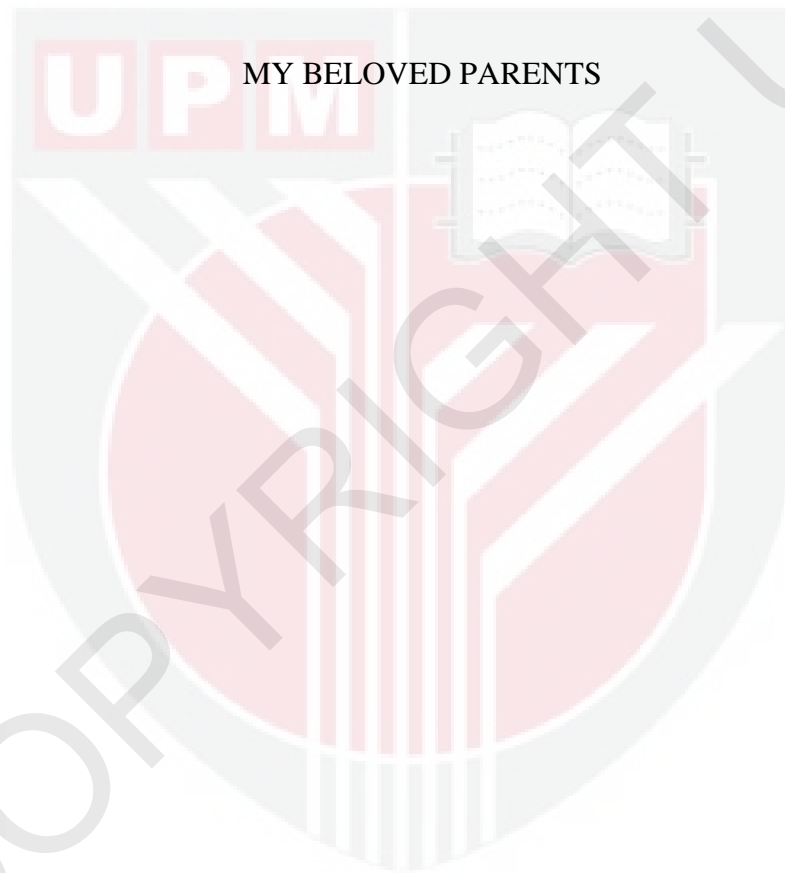
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Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

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By

**THAWDA MYINT**

**June, 2013**

**Chairman: Assoc. Professor Mohd Puad Abdullah, PhD**

**Faculty: Biotechnology and Biomolecular Sciences**

In response to drought, plants change their metabolic activities towards limiting cellular water consumption and loss. One metabolic process that is affected by this stress is ethanolic fermentation. In plants, ethanolic fermentation occurs during limited oxygen condition and under certain environmental stresses. The effects of ethanol fermentation on plant growth and survival under drought stress are not well explained. In addition, previous studies on ethanolic fermentation in plants were limited to alcohol dehydrogenase (EC.1.1.1.1) enzyme activity and gene expression. In this study, it was hypothesized that ethanolic fermentation is required to enhance plant ability to retain cellular water under drought. Enhancing the capacity of ethanolic fermentation in a plant would improve the plant ability to retain cellular water; thus, retain the plant's photosynthetic capacity. To test the hypothesis, this study was carried out with the following objectives: i) to identify the specific *ADH* genes responding to drought in *Arabidopsis* plants, ii) to evaluate the effects of defective *ADH* on growth and drought-related parameters, iii) to evaluate the effects of enhanced ethanolic fermentation on growth and drought-related parameters. The objectives were achieved by a combination of the gain-and the loss-of-function approaches. For the gain-of-function approach, an *Arabidopsis* plant over-expressing the *ADH1* transgene was developed using the Gateway technology where fully characterized homozygous lines were used for the analysis. As for the loss-of-function approach, the T-DNA insertion mutant lines with impaired *ADH* genes were used. The plants were exposed to polyethylene glycol-induced drought stress, and their responses at physiological, biochemical and molecular levels were analysed together with their overall growth performance.

In the present study, the level of relative water content (RWC) of *Arabidopsis* plants dropped to 75% from the initial level of 85% when treated with 5% (w/v) PEG-20,000, demonstrated that the plants were moderately water-stressed. The stressed plants had high levels of proline and low levels of chlorophyll. At enzyme and metabolite levels, both the root and leaf NADH-ADH activities were increased 5.9 and 4.4 folds, respectively. For pyruvate decarboxylase (PDC), the activity was increased in the root

(1.2 folds) and in the leaf (4.4 folds). Ethanol, the end product of ethanol fermentation was accumulated in both the leaf (3 folds) and root (2 folds). The increase in the level of ethanol was parallel with the increase observed in the activities of NADH-ADH and PDC. At gene level, the majority of the *ADH* and *PDC* genes were up-regulated. Two of the *PDC* genes (*AT5G01320* and *AT4G33070*) genes and three of the *ADH* genes (*AT1G64710*, *AT1G77120* and *AT5G24760*) were up-regulated in the leaf and root. These evidences support the conclusion that the capacity of ethanolic fermentation was enhanced in response to drought.

When the individual *ADH* gene was defective, a severe reduction in the ADH activities and growth performance of the mutant plants were observed when exposed to drought. The T-DNA insertion *adh* knock-out mutant lines [*adh1*mutant (*AT1G77120*) and two *adh-like* mutants (*AT1G64710* and *AT5G24760*)] demonstrated reduced growth judging by a shorter root system and lower biomass content. The plants also failed to retain cellular water which subsequently affected their physiological process including photosynthesis.

In the transgenic *Arabidopsis* plant over-expressing the *ADH1* gene, the capacity of ethanolic fermentation was enhanced judging by the increase in the ADH enzyme activity (6 folds). Under drought stress, the transgenic plant exhibited the following phenotypic improvements i) improved ability to retain cellular water; ii) increased chlorophyll content; iii) increased proline level; iv) increased NADH-ADH activity; v) increased volume of root system and iv) increased biomass. All these features contributed to the overall improvement of the transgenic plants under drought.

As a conclusion, ethanolic fermentation is important for plants grown under drought condition. Enhancing the capacity of ethanolic fermentation improves plant ability to maintain cellular water; thus, supports the normal function of photosynthesis. To reduce the impacts of drought in plants, the capacity of plant ethanolic fermentation may be enhanced, and this strategy could be implemented in crop plants of economic importance.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

## **PENCIRIAN FUNGSI GEN ALKOHOL DEHIDROGENASE DALAM TUMBUHAN ARABIDOPSIS DI BAWAH KEADAAN KEMARAU**

Oleh

**THAWDA MYINT**

**Jun, 2013**

**Pengerusi: Profesor Madya Mohd Puad Abdullah, PhD**

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Sebagai tindak balas kepada kemarau, tumbuhan mengubah aktiviti metabolisme ke arah penjimatan penggunaan dan kehilangan air. Satu proses metabolisme yang dipengaruhi oleh stres ini adalah fermentasi etanol. Dalam tumbuhan, fermentasi etanol berlaku semasa keadaan kekurangan oksigen dan di bawah stres alam sekitar yang tertentu. Kesan fermentasi etanol ke atas pertumbuhan tumbuhan yang hidup dalam keadaan kemarau tidak diketahui dengan jelas. Di samping itu, kajian terdahulu mengenai fermentasi etanol dalam tumbuhan terbatas kepada aktiviti enzim dan gen alkohol dehidrogenase (EC.1.1.1.1). Hipotesis kajian ini adalah fermentasi etanol diperlukan untuk meningkatkan keupayaan tumbuhan untuk mengekalkan air sel dalam keadaan kemarau. Meningkatkan kapasiti fermentasi etanol akan meningkatkan keupayaan tumbuhan untuk mengekalkan air sel; oleh itu, mengekalkan kapasiti fotosintesis. Untuk menguji hipotesis tersebut, kajian ini dijalankan dengan objektif berikut: i) untuk mengenal pasti gen *ADH* tertentu yang bertindakbalas ke atas kemarau dalam tumbuhan *Arabidopsis*, ii) untuk menilai kesan kecacatan gen *ADH* kepada pertumbuhan dan parameter kemarau yang berkaitan, iii) menilai kesan peningkatan kapasiti fermentasi etanol ke atas pertumbuhan dan parameter kemarau yang berkaitan. Objektif berkenaan telah dicapai melalui pendekatan kehilangan-fungsi dan kedapatan-fungsi gen *ADH*. Bagi pendekatan kedapatan-fungsi, tumbuhan *Arabidopsis* yang mengekspreskan *ADH1* secara berlebihan telah dibangunkan menggunakan teknologi Gateway. Pokok homozigous yang telah dicirikan sepenuhnya telah digunakan untuk tujuan analisis. Bagi pendekatan kehilangan-fungsi, tumbuhan *Arabidopsis* mutan yang mempunyai selitan T-DNA dengan gen *ADH* yang cacat telah digunakan. Tumbuhan tersebut telah didedahkan kepada polietilena glikol untuk menjana kesan stres kemarau, dan tindak balas tumbuhan tersebut di peringkat fisiologi, biokimia dan molekul telah dianalisis bersama dengan prestasi pertumbuhan tersebut secara keseluruhan.

Dalam kajian ini, tahap kandungan air relatif (RWC) tumbuhan *Arabidopsis* menurun kepada 75% daripada tahap awal sebanyak 85% apabila dirawat dengan 5% (w / v) PEG-20, 000, menunjukkan bahawa tumbuhan tersebut berada di bawah stres kemarau yang sederhana. Tumbuhan tersebut mempunyai tahap prolina yang tinggi

dan paras klorofil yang rendah. Pada peringkat enzim dan metabolit, aktiviti enzim NADH-ADH pada akar dan daun telah meningkat sebanyak 5.9 dan 4.4 kali ganda, masing-masing. Manakala untuk enzim piruvat dekarboksilase (PDC), aktiviti enzim tersebut telah meningkat pada akar (1.2 kali ganda) dan daun (4.4 kali ganda). Etanol, produk akhir fermentasi etanol telah terkumpul di dalam daun (3 kali ganda) dan akar (2 kali ganda). Peningkatan paras etanol adalah selari dengan peningkatan yang diperhatikan dalam aktiviti enzim NADH-ADH dan PDC. Di peringkat gen, majoriti gen *ADH* dan *PDC* telah meningkat dengan ketara. Dua daripada gen *PDC* (AT5G01320 dan AT4G33070) dan tiga daripada gen *ADH* (AT1G64710, AT1G77120 dan AT5G24760) telah mengalami kenaikan dalam pengekspresan yang ketara pada daun dan akar. Kesemua bukti berkenaan menyokong peningkatan kapasiti fermentasi etanol sebagai tindak balas terhadap kemarau.

Apabila gen *ADH* mengalami kecacatan, pengurangan yang ketara dalam aktiviti enzim ADH dan prestasi pertumbuhan tanaman mutan telah diperhatikan apabila tumbuhan tersebut didedahkan kepada kemarau. Tumbuhan mutan Arabidopsis yang mempunyai selitan T-DNA dengan gen *ADH* yang cacat [mutan *adh1* (AT1G77120) dan dua mutan *adh*-setara (AT1G64710 dan AT5G24760)] telah menunjukkan penurunan dalam prestasi pertumbuhan berdasarkan kepada sistem akar yang pendek dan biomas yang rendah. Tumbuhan tersebut juga gagal untuk mengekalkan air sel dan seterusnya telah menjejaskan proses fisiologi termasuk fotosintesis.

Dalam tumbuhan Arabidopsis transgenik yang mengekspreskan gen *ADH1* secara berlebihan, kapasiti fermentasi etanol telah dipertingkatkan berdasarkan kepada peningkatan aktiviti enzim ADH (6 kali ganda). Di bawah stres kemarau, tumbuhan transgenik tersebut mempamerkan penambahbaikan fenotip seperti berikut: i) peningkatan keupayaan untuk mengekalkan air sel; ii) peningkatan kandungan klorofil; iii) peningkatan paras prolina; iv) peningkatan aktiviti enzim NADH-ADH; v) peningkatan jumlah akar; dan iv) peningkatan biomas. Ke semua ciri-ciri ini menyumbang kepada peningkatan prestasi keseluruhan tumbuhan transgenik tersebut di bawah keadaan kemarau.

Kesimpulannya, fermentasi etanol adalah penting untuk tumbuhan di bawah keadaan kemarau. Meningkatkan kapasiti fermentasi etanol telah meningkatkan keupayaan tumbuhan untuk mengekalkan air sel; oleh itu, menyokong fungsi normal fotosintesis. Untuk mengurangkan kesan kemarau pada tumbuhan, kapasiti fermentasi etanol dalam tumbuhan boleh dipertingkatkan dan strategi ini boleh dikembangkan kepada tanaman yang mempunyai kepentingan ekonomi.



## ACKNOWLEDGEMENTS

I would like to express my heartfelt gratitude beginning with my main supervisor Assoc. Professor Dr. Mohd Puad Abdullah for his patience and scientific advice during my dissertation process, which made an impetus for me to finish this doctoral thesis. It is not an easy task, providing guidance and at the same time reviewing my thesis and I am grateful for his thought and guidance.

Secondly, I am grateful to the Ministry of Agricultural and Irrigation, Myanmar Agriculture Service for providing an opportunity to pursue this Doctor of Philosophy programme. My gratitude is also extended to Dr. Khin Maung Thet for his constant encouragement, concerns and great support. I appreciate very much to assistance from Dr. Pa Pa Aung and all of lab colleagues at the Biotechnology laboratory, Shwe Nantha farm for their warm and cordial friendship.

My acknowledgment would be incomplete without appreciating my scholarship provider. I would not have contemplated this road if not for this generous financial support for my doctoral study from the oil crop development project in Myanmar, initiated by the MOAI, Myanmar and technical assistance by FAO (Food and Agriculture Organization).

My sincere appreciation goes to the supervisory committee, Associate professor Dr. Parameswari Namasivayam and Associate professor Dr. Suhaimi Napis from the Department of Cell and Molecular Biology, Faculty of Biotechnology and Bimolecular Sciences, Universiti Putra Malaysia for their advice and constructive feedbacks. Life at Proteomic Lab has always been a warm and inviting place to work with, where I have got very good working relationship and support from my lab mates with distinct personality of gentleness and amicability.

Of course no acknowledgements would be complete without appreciating the sacrifices made by my parents. The completion of this thesis would not be possible without the love and support from my family, who have blessed me to away from home to pursue my study. My parents have instilled many admirable qualities of life and taught me about hard work, self-respect and about being persistence. To my family, thank you.

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## LIST OF ABBREVIATIONS

µg	microgram
µl	microliter
µM	micromolar
%	percentage
ACS	acetyl-CoA synthetase
ADH	alcohol dehydrogenase
ALDH	aldehyde dehydrogenase
ANOVA	analysis of variance
ATP	adenosine-5'-triphosphate
BLAST	basic local alignment search tool
bp	base pair
BSA	bovine serum albumin
CaMV	cauliflower mosaic virus
cDNA	complementary DNA
Chl	chlorophyll
CTAB	cetyltrimethylammonium bromide
C-terminal	carboxyl terminal
DEPC	Diethylpyrocarbonate
DNA	deoxyribonucleic acid
dNTPs	mixture of dATP, dTTP, dCTP and dGTP
DTT	dithiothreitol
DW	dry weight
EDTA	ethylenediaminetetraacetic acid
<i>gfp</i>	green fluorescent protein
g	gram
<i>gus</i>	β-glucuronidase
g/L	gram per liter
H <sub>2</sub> O <sub>2</sub>	hydrogen peroxide
H <sub>3</sub> PO <sub>4</sub>	Phosphoric acid
<i>hptII</i>	hygromycin phosphotransferase II
K <sub>3</sub> Fe(CN) <sub>6</sub>	Potassium ferricyanide
K <sub>4</sub> Fe(CN) <sub>6</sub>	Potassium ferrocyanide
kbp	kilo-base pair
KCl	Potassium Chloride
L	liter
LB	Lysogeny broth
LDH	lactate dehydrogenase
LEA	Late Embryogenesis abundant
M	molar
Mb	mega bases
MCS	multiple cloning sites
MgCl <sub>2</sub>	Magnesium Chloride
MgSO <sub>4</sub>	Magnesium Sulphate
min	minute
ml	milliliter
mm	millimeter
mM	millimolar
Mpa	Megapascal (water potential unit)

mRNA	messenger ribonucleic acid
MS	medium of Murashige and Skoog
Na <sub>3</sub> PO <sub>4</sub>	sodium phosphate
NAD	nicotinamide adenine dinucleotide
NADP	nicotinamide adenine dinucleotide phosphate
NCBI	national center for biotechnology information
ng	nanogram
NH <sub>4</sub>	ammonium
nm	nanometer
NO <sub>3</sub> <sup>-</sup>	nitrate
<i>npt-II</i>	neomycin phosphotransferaseII
OD	optical density
OH	hydroxide
ORF	open reading frames
P5CS	Δ 1 -pyrroline-5-carboxylate synthetase
PCR	polymerase chain reaction
PDC	pyruvate decarboxylase
PDH	pyruvate decarboxylase dehydrogenase
pg	pictogram
PEG	Polythethylene glycol
PVP-40	polyvinylpyrrolidone
RNA	ribonucleic acid
RNase	ribonuclease
ROS	reactive oxygen species
rpm	rotation per minute
RT	room temperature
RT-PCR	Reverse transcriptase polymerase chain reaction
RuBP	Ribulose-1,5-biphosphate
RWC	Relative water content
SDS	sodium dodecyl sulphate
SE	standard error
s	second
SOD	superoxide dismutases
TAE	Tris-acetate-EDTA
TCA	tri carboxylase acid
T <sub>m</sub>	melting temperature
TW	turgid weight
U	Unit
UTR	untranslated region
UV	Ultra violet
V	volt
v/v	volume per volume
w/v	weight per volume
X-Gluc	5-bromo-4-chloro-3-indolyl-β-D-glucuronide

## CHAPTER 1

### INTRODUCTION

The impacts of global warming and climate change are becoming important. Especially in prolonged drought and frequent flooding are common phenomenon in many parts of the world (Qiu, 2010; Schiermeier, 2011). Together with other biotic and abiotic stresses including salinity, low temperature, pest and disease, these could severely affect agricultural productivity as the stress could restrict the expression of the full genetic potential of a crop plant, and threaten the sustainability of agricultural industry (Shilpi and Narendra, 2005). One estimate puts a reduction of more than 50% in yield because of environmental stress (Bray, 2000).

Drought severely reduces plant productivity as a result of reduced photosynthetic capacity (Hummel *et al.*, 2010) through stomatal closure of CO<sub>2</sub> diffusion (Sharkey, 1990; Chaves, 1991; Ort *et al.*, 1994; Cornic and Massacci, 1996) or by metabolic impairment of carbon reduction cycle (Boyer, 1976; Lawlor, 1995; Allen and Ort, 2001). Evidence that impaired ATP synthesis is the main factor limiting photosynthesis even under mild drought (Boyer, 1976; Tezara *et al.*, 1999) has further stimulated debate (Cornic, 2000; Lawlor and Cornic, 2002). While some plants can withstand the adverse effects of prolonged drought, most are not able to hold their metabolic function long enough for survival before the rain fall again. The mechanism that governs these differential abilities of different plants to withstand different intensities of drought is not fully understood. Changes in the levels of certain metabolites such as chlorophyll content, sugar-alcohol and proline are commonly observed in the plants exposed to drought condition (Sperdouli and Moustakas, 2012; Silvente *et al.*, 2012); however, these biochemical changes are often overlapped with plant responses to other environmental stresses.

To overcome this potential threat to agriculture, scientists turn to biotechnology for long-term solution of intensifying research on various aspects of plant adaptative response and survival to various environmental stresses. One approach is to utilize genome-wide expression analysis where drought-related genes could be obtained from thousands of genes analysed (Seki *et al.*, 2002, Patrica *et al.*, 2011). The efforts were proven to be fruitful as scientists can identify important genes related to drought and carry out gene functional studies for more in-depth analysis of drought gene network.

One particular gene that responds to drought is alcohol dehydrogenase (*ADH*). In *Arabidopsis* plant, alcohol dehydrogenase enzyme (EC.1.1.1.1) has been encoded by *ADH* gene which is involved in mediating stress responses, mainly in anaerobic condition (Dolferus *et al.*, 1994; Peters and Frenkel, 2004). In addition, numerous stress-induced genes have been identified using microarray experiment in which *ADH* gene was up-regulated under drought condition (Seki *et al.*, 2002). This observation supports an earlier study on ethanol production under drought condition. Kimmerer and Kozolowski (1982) reported that high level of ethanol content was produced in dehydrated woody plants. These evidences of upregulation of *ADH* gene expression and production of ethanol under drought condition connect to induction of ethanolic

fermentation as ADH is the main enzyme of ethanolic fermentation. So far, little effort has been done in experiments to follow up these findings with functional studies of the *ADH* gene in plants exposed to drought stress condition.

Ethanolic fermentative pathway normally occurs in plants grown under anaerobic condition. This topic has been well researched in animals and yeasts but not so much in plants. Under hypoxic conditions where molecular oxygen becomes limited, fermentative enzymes in the ethanolic pathway are upregulated, causing increased production of ethanol and  $\text{NAD}^+$ . The cofactor  $\text{NAD}^+$  was generated as a by-product of this process is what makes ethanolic fermentation important for the survival of living systems under anaerobic condition. In the context of the fermentative enzyme in plants, the activities of the ADH enzymes are up-regulated not only in anaerobic conditions but also in other environmental stresses condition where oxygen was not completely depleted (Robert *et al.*, 1984; 1989; Tadege *et al.*, 1998; Mustroph and Albrecht, 2003; Geigenberger, 2003; Fukao and Bailey-Serres, 2004). Hence, some suggest that plant ADH (EC.1.1.1.1) is involved in stress adaptation mechanism for energy production (Tesniere *et al.*, 2006; Ismond *et al.*, 2003; Kato-Noguchi *et al.*, 2006).

Previous functional analyses of the *ADH* gene were mainly done on the effects of the over-expression on plant tolerance to low oxygen levels when the plant or cells are submerged in water (Shiao *et al.*, 2002, Ismond *et al.*, 2003). In the model plant, *Arabidopsis thaliana*, the ADH enzyme (EC 1.1.1.1) is encoded by the *ADH1* gene and other seven *ADH*-like genes (The Arabidopsis Genome Initiative, 2000). So far, only *ADH1* has been studied in the plant including its expression. The gene was reported to be associated with various environmental stresses. However, the mechanism of alcohol dehydrogenase genes (*ADH*) function under drought stress is still not clear.

In this study, it was hypothesized that ethanolic fermentation is required to enhance plant ability to retain cellular water under drought stress condition. Enhancing ethanolic fermentation in a plant would improve water retention in the plant; thus, improving the plant photosynthetic capacity. The hypothesis was tested in a combination of the gain- or loss-of-function approaches. For the gain-of-function approach, an *Arabidopsis* plant overexpressing the *ADH1* transgene was developed using the Gateway technology; fully characterized homozygous lines were used for the analysis. As for the loss-of-function approach, the T-DNA insertion mutant lines with impaired *ADH* genes were used. The plants were exposed to PEG-induced drought stress conditions, and their responses at the physiological, biochemical and molecular levels were analysed together with their overall growth performance.

To test the hypothesis, this study was carried out with the following objectives:

- i) to identify the specific *ADH* genes in *Arabidopsis thaliana* responding to drought stress condition
- ii) to evaluate the impacts of defective *ADH* on growth and drought-related parameters of plant
- iii) to evaluate the impacts of enhanced ethanolic fermentation on growth and drought-related parameters

## BIBLIOGRAPHY

- Ábrahám, E., Rigó, G., Székely, G., Nagy, R., Koncz, C., and Szabados, L. 2003. Light-dependent induction of proline biosynthesis by abscisic acid and salt stress is inhibited byassinosteroid in *Arabidopsis*. *Plant Mol Biol* 51: 363–372.
- Acevedo, E., Hsiao, T.C., and Henderson, D.W. 1971. Immediate and subsequent growth responses of maize leaves to changes in water status. *Plant Physiol* 48: 631-636.
- Acharya, B.R., and Assmann, S.M. 2009. Hormone interactions in stomatal function. *Plant Mol Biol* 69: 451-462.
- Agarwal, S., Kapoor, A., Lakshmi, O.S., and Grover, A. 2007. Production and phenotypic analysis rice transgenics with alter the levels of pyruvate decarboxylase and alcohol dehydrogenase proteins. *Plant Physiol Biochem* 45: 637-646.
- Alexieva, V., Sergiev, I., Mapelli, S., and Karanov, E. 2001. The effect of drought and ultraviolet radiation on growth and stress markers in pea and wheat. *Plant Cell Environ* 24: 1337-1344.
- Alonso, J.M., Stepanova, A.N., and Leisse, T.J., *et al.* 2003. Genome wide insertional mutagenesis of *Arabidopsis thaliana*. *Science* 301: 653–657.
- Alpert, P. 2005. The limits and frontiers of desiccation tolerant life. *Integr Comp Biol* 45:685–695.
- Al-Shehbaz, I.A., and O’Kane, S.L. 2002. Taxonomy and phylogeny of *Arabidopsis* (Brassicaceae). In Somerville, C. R. and Meyerowitz, E. M. (eds), *The Arabidopsis Book*. American Society of Plant Biologists, Rockville, Maryland. doi/10.1199/tab. 0001, [http:// www.aspb.org/publications/arabidopsis/](http://www.aspb.org/publications/arabidopsis/)
- Amal, H., Arjun, K., Madana, M.R., Ambavaram., and Andy, P. 2010. Molecular and physiological analysis of drought stress in *Arabidopsis* reveals early responses leading to acclimation in plant growth. *Plant Physiol* 154: 1254-1271.
- Anjum, S.A., Wang, L.C., Farooq, M., Hussain, M., Xue, L.L., and Zou, C.M. 2011. Brassinolide application improves the drought tolerance in maize through modulation of enzymatic antioxidants and leaf gas exchange. *J Agron Crop Sci* 197(3):177-185.
- Antonio, C., Pinheiro, C., Chaves, M.M., Ricardo, C.P., Ortuno, M.F., and Thomas-Oates, J. 2008. Analysis of carbohydrates in *Lupinus albus* stems on imposition of water deficit, using porous graphitic carbon liquid chromatography–electrospray ionization mass spectrometry. *J Chromatogr A* 1187:111-118.



- Arabidopsis Genome Initiative, 2000. Analysis of the genome sequence of the flowering plant *Arabidopsis thaliana*. *Nature* 408: 796-815.
- Arkhipova, T.N., Veselov, S.U., Melantiev, A.I., Marty, N.E.V., and Kudoyerova, G.R. 2005. Ability of bacterium *Bacillus* to produce cytokinins and to influence the growth and endogenous hormone content of lettuce plants. *Plant and Soil* 272: 201-209.
- Assmann, S.M., Snyder, J.A, Lee, Y.R.J. 2000. ABA-deficient (*aba1*) and ABA-insensitive (*abi1-1*, *abi2-1*) mutants of *Arabidopsis* have a wild-type stomatal response to humidity. *Plant Cell Environ* 23: 387-395.
- Atkin, O.K. and Macherel, D. 2009. The crucial role of plant mitochondria in orchestrating drought tolerance. *Ann Bot* 103: 581-597.
- Atteia, A., van Lis, R., Tielens, A.G.M., and Martin, W.F. 2012. Anaerobic energy metabolism in unicellular photosynthetic eukaryotes. *Biochim Biophys Acta*, 10: p. 1016.
- Banerjee, S., Schmidt, T., Fang, J., Stanley, C.A., and Smith, T.J. 2003. Structural Studies on ADP Activation of Mammalian Glutamate Dehydrogenase and the Evolution of Regulation. *Biochemistry* 42: 3446-3456.
- Bartels, D., and Sunkars, R. 2005. Drought and salt tolerance in plants. *Cri Rev Plant Sci* 24: 23-58.
- Bates, L.S., Waldren, R.P., and Teare, I.D. 1973. Rapid determination of free proline for water stress study. *Plant Soil* 39: 205-207.
- Baulcombe, D. 2004. RNA silencing in plants. *Nature* 431: 356-363.
- Bechtold, N., Ellis, J., and Pelletier, G. 1993. *In planta Agrobacterium* mediated gene transfer by infiltration of adult *Arabidopsis thaliana* plants. *C R Acad Sci Paris, Life Sciences*. 316:1194–1199.
- Bent, A. 2006. *Arabidopsis thaliana* floral dip transformation method. *Methods Mol Biol* 343: 87-103.
- Berger, J and Avery, G.S.1943. Dehydrogenase of the *Avena* coleoptiles. *Amer J Bot* 30:290-297.
- Bhatt, R.M., and Srinivasa Rao, N.K. 2005. Influence of pod load response of okra to water stress. *Indian J Plant Physiol* 10: 54–59.
- Biale, J.B. 1946. Effect of oxygen concentration on respiration of the 'Fuerte' avocado fruit. *Amer J Bot* 33:363-373.

- Bicsak, T.A., Kann, L.R., Reiter, A., and Chase Jr, T. 1982. Tomato alcohol dehydrogenase: purification and substrate specificity. *Arch Biochem Biophys* 216: 605-615.
- Bidel, L.P.R., Pages, L., Riviere, L.M. Pelloux, G., and Lorendeau, J.Y. 2000. MassFlowDyn I: A carbon transport and partitioning model for root system architecture. *Ann Bot* 85: 869-886.
- Blum, A., Sinmena, B., Mayer, J., Golan, G., and Shpiler, L. 1994. Stem reserve mobilization supports wheat-grain filling under heat stress. *Aust J Plant Physiol* 21: 771-781.
- Blum, A., Munns, R., Passioura, J.B., and Turner, N.C. 1996. Genetically engineered plants resistant soil drying and salt stress: How to interpret osmotic relation? *Plant Physiol* 110:1051.
- Bohnert, H. J., and Shen, B. 1999. Transformation and compatible solutes. *Sci Hortic* 78: 237-260.
- Bohnert, H. 2000. What makes desiccation tolerable? *Genome Biol* 1:1010.1011-1010.1014.
- Borsani, O., Diaz, P., Agius, M.F., Valpuesta, C., and Monza, J. 2001. Water stress generate and oxidative through the induction of a specifif Cu/Zn superoxidisedismutase I Lotus corniculates leaves. *Plant Sciences*. 161: 757-763.
- Bota, J., Flexas, J., and Medrano, H. 2004. Is photosynthesis limited by decreased Rubisco activity and RuBP content under progressive water stress? *New Phytol* 162: 671-681.
- Bray, E.A., Bailey-Serres, J., and Weretilnyk, E. 2000. Responses to abiotic stresses. Biochemistry and Molecular Biology of Plants, *American Society of Plant Biologists*, Rockville, MD, 2000, pp. 158-1249.
- Bray. 2002. Abscisis acid regulation of gene expression during water deficit in the era of the *Arabidopsis* genome. *Plant Cell Environ* 25: 153-161.
- Bucher, M., Bra"ndle, R., and Kuhlemeier, C. 1994. Ethanolic fermentation in transgenic tobacco expressing *Zymomonas mobilis* pyruvate decarboxylase. *EMBO J* 13: 2755-2763.
- Bucher, M., Brander, K.A., Sbicego, S., Mandel, T., and Kuhlemeier, C. 1995. Aerobic fermentation in tobacco pollen. *Plant Mol Biol* 28: 739-750.
- Buitink, J., and Leprince, O. 2004. Glass formation in plant anhydrobiotes: survival in the dry state. *Cryobiology* 48:215-228.

- Cameron, D.S., and Cossins, E.A. 1967. Studies of intermediary metabolism in germinating pea cotyledons. *Biochem J* 105: 323-331.
- Carpita, N., Sabularse, D., Montezinos, D., and Delmer, D.P. 1979. Determination of the pore size of cell walls of living plant cells. *Science* 205: 1144–1147.
- Cazares, B.X., Ortega, F.A.R., Elens, L.F. and Medrano, R.R. 2010. Drought tolerant in crop plant. *Amer J Plant Physiol* 5 (5): 241-256.
- Chandler, J., and Bartels, D. 1995. Plant Dessication. In: Plant responses to environmental stress: from Phytohormones to Genome Reorganization, Lerner, H.R. (Ed) Marcel Dekker, New York, USA. Pp 575-590.
- Chaves, M.M. 1991. Effects of water deficits on carbon assimilation. *J Exp Bot* 42: 1-16.
- Chaves, M.M., Pereira, J.S., Maroco, J.P., Rodrigues, M.L., Ricardo, C.P.P., Oso´rio, M.L., Carvalho, I., Faria, T., and Pinheiro, C. 2002. How plants cope with water stress in the field: photosynthesis and growth. *Ann Bot* 89: 907-916.
- Chaves, M.M., Pereira, J.S., and Maroco, J. 2003. Understanding plant response to drought from genes to the whole plant. *Funct Plant Biol* 30: 239-264.
- Chaves, M.M., and Oliveira, M. M. 2004. Mechanisms underlying plant resilience to water deficits: prospects for water-saving agriculture. *J Exp Bot* 55:2365-2384.
- Chaves, M.M., Flexas, J., and Pinheiro, C. 2009. Photosynthesis under drought and salt stress: regulation mechanisms from whole plant to cell. *Ann Bot* 103: 551-560.
- Chen, T.H.H., and Murata, N. 2002. Enhancement of tolerance of abiotic stress by metabolic engineering of betaines and other compatible solutes. *Curr Opin Plant Biol* 5: 250-257.
- Chen, J.Z, Lin, L.R., and Lu, G.A. 2010. An index of soil drought intensity and degree: an application on corn and a comparison with CWSI. *Agri Water Manag* 97: 865-871.
- Cheng, C., and Shuman, S. 2000. Recombinogenic Flap Ligation Pathway for Intrinsic Repair of Topoisomerase IB-Induced Double-Strand Breaks. *Mol Cell Biol* 20: 8059-8068.
- Cheng, J. S., Qiao, B., and Yuan, Y. J. 2008. Comparative proteome analysis of robust *Saccharomyces cerevisiae* insights into industrial continuous and batch fermentation. *Appl Microbiol Biotechnol* 81(2): 327-338.
- Choudhary, N.L., Sairam, A.K., and Tyagi, A. 2005. Expression of delta1-pyrroline-5-carboxylate synthetase gene during drought in rice (*Oryza sativa L.*). *Ind J Biochem Biophys* 42: 366–370.

- Christie, P.J., Hahn, M., and Walbot, V. 1991. Low-temperature of *alcohol dehydrogenase-1* mRNA and protein activity in maize and rice seedlings. *Plant Physiol* 95: 699-706.
- Chung, H.J., and Ferl, R.J. 1999. *Arabidopsis* alcohol dehydrogenase expression in both shoots and roots is conditioned by root growth environment. *Plant Physiol* 121: 429-436.
- Clough, S.J., and Bent, A.F. 1998. Floral dip: a simplified method for *Agrobacterium*-mediated transformation of *Arabidopsis*. *Plant J* 16: 735-743.
- Conley, T.R., Peng, H.P., and Shih, M.C. 1999. Mutations affecting induction of glycolytic and fermentative genes during germination and environmental stresses in *Arabidopsis*. *Plant Physiol* 119: 599-608.
- Cossins, E.A. 1978. Ethanol metabolism in plant. In: Hook DD, Crawford RMM, eds. *Plant life in anaerobic environments*. Ann Arbor Science Publishers, 169-202.
- Cough, J.S. 2005. Floral dip: *Agrobacterium*-mediated germ line transformation. *Methods Mol Biol* 286.
- Crawford, R.M.M. 1982. Physiological responses to flooding. In J H Milburn, M H Zimmermann, eds, *Encyclo Plant Physiol New Series*, Vol: IV C. Springer-Verlag, Berlin, pp 453-477.
- Crowe, J. H., Carpenter, J. F., and Crowe, L.M. 1998. The role of verification in anhydrobiosis. *Ann Rev Physiol* 60: 73-103.
- Cornic, G. 2000. Drought stress inhibits photosynthesis by decreasing stomatal aperture – not by affecting ATP synthesis. *Trends Plant Sci* 5: 187–188.
- Cummings, M. P., and Clegg, M. T. 1998. Nucleotide sequence diversity at the alcohol dehydrogenase 1 locus in wild barley (*Hordeum vulgare* ssp. *spontaneum*): an evaluation of the background selection hypothesis. *Proc Natl. Acad Sci USA* 95:5637–5642.
- Dai, A. 2010. Drought under global warming: a review. *Wiley Interdisc. Rev Clim Change* 2: 45–65.
- Daniles, M.J., Mirkov, T.E., and Chrispeels, M.J. 1994. The plasma membrane of *Arabidopsis thaliana* contains a mercury-insensitive aquaporin that is a homolog of the tonoplast water channel protein TIP. *Plant Physiol* 106: 1325-1333.
- Daniel, M.J., Chaumont, F., Mirkov, T.E., and Chrispeels, M.J. 1996. Characterization of a new vacuolar membrane aquaporin sensitive to mercury at a unique site. *Plant Cell* 8: 587-599.

- Das, P., and Joshi, N.C. 2011. Minor modifications in obtainable *Arabidopsis* floral dip method enhance transformation efficiency and production of homozygous transgenic lines harboring a single copy of transgene. *Adv Biosci Biotechnol* 2: 59-67
- Dat. J., Vandenabeele, S., Vrabova, E., Van, M., Inze, D., and Van, B.F. 2000. Dual action of the active oxygen species during plant stress responses. *Cell Mol Lief Sci* 57: 779-795.
- Davies, D.D., Grego, S., and Kenworthy, P. 1974. The control of the production of lactate and ethanol by higher plants. *Planta* 118: 297-310.
- Davies, D.D. 1980. Anaerobic metabolism and production of organic acids. In PK Stumpf, EE Conn, eds, *The Biochemistry of Plants: A Comprehensive Treatise*, vol 2. Academic Press, New York, pp 581-611
- de Bruxelles, G.L, Peacock, W.J., Dennis, E.S., and Dolferus, R. 1996. Abscisic acid induces the *alcohol dehydrogenase* gene in *Arabidopsis*. *Plant Physiol* 111: 381-391.
- Delauney, A.J. and Verma, D.P.S. 1993. Proline biosynthesis and osmo-regulation in planr. *Plant J* 4: 215-223.
- Demmig-Adams, B., and Adams, W.W. 1996. The role of the xanthophyll cycle carotenoids in the protection of photosynthesis. *Trends Plant Sci* 1: 21-26.
- De Ronde, J. A., Cress, W. A., Kruger, G. H. J., Strasser, R. J., and Van Staden, J. 2004. Photosynthetic response of transgenic soybean plants, containing an *Arabidopsis P5VR* gene, during heat and drought stress. *J Plant Physiol* 161:1211-1224.
- Desikan, R., Machkerness, S.A.H., Hancock, J.T., and Neill, S.J. 2001. Regulation of the *Arabidopsis* transcripts by oxidative stress. *Plant Physiol* 127: 159-172
- Dohmann, E.M.N., Kuhnle, C., Schwechheimer, C. 2005. Loss of the CONSTITUTIVE PHOTOMORPHOGENIC 9 Signalosome Subunit 5 Is Sufficient to Cause the *cop/det/fus* Mutant Phenotype in *Arabidopsis*. *Plant Cell* 17:1967-1978.
- Dodd, I.C. 2003. Hormonal interactions and stomatal responses. *J Plant Growth Regul* 22: 32-46.
- Dolferus, R., Bruxelles, G.D., Dennis, E.S., and Peacock, W.J. 1994a. Regulation of the *Arabidopsis Adh* gene by anaerobic and other environmental stress. *Ann Bot* 74:301-308.
- Dolferus, R., Bruxelles, G.D., Dennis, E.S., and Peacock, W.J. 1994a. Regulation of the *Arabidopsis Adh* gene by anaerobic and other environmental stress. *Ann Bot* 74:301-308.

- Dolferus, R., Jacob, M., Peacock, W.J and Dennis, E.S. 1994b. Differential interactions of promoter elements in stress responses of the *Arabidopsis Adh* gene. *Plant Physiol* 105:1075-1087.
- Dolferus, R., Ellis, M., de Bruxelles, G., Trevaskis, B., Hoeren, F., Dennis, E.S., and Peacock, W.J. 1997a. Strategies of gene action in *Arabidopsis* during anoxia. *Ann Bot* 79: 21–31.
- Dolferus, R., Osterman, J., Peacock, W.J. and Dennis, E.S. 1997. Cloning of the *Arabidopsis* and rice class III Adh genes: implications for the origin of plant ADH enzymes. *Genetics* 146: 1131–1141.
- Edward, K., Johnston, C., and Thompson, C. 1991. A simple and rapid method for the presentation of plant genomic DNA for PCR analysis. *Nucl Acids Res* 19: 1349-1349.
- Else, M.A., and Jackson, M.B. 1998. Transport of 1-aminocyclopropane-1-carboxylic acid (ACC) in the transpiration stream of tomato (*Lycopersicon esculentum*) in relation to foliar ethylene production and petiole epinasty. *Aust J Plant Physiol* 25(4): 453-458.
- Eyre-Walker Adam., Rebecca, G., Holly, H., Dan, F., and Brandon, G. 1998. Investigation of the bottle neck leading to the Domestication of maize. *Proc natl Acad Sci USA* 95:4441-4446.
- Fan, L., and Neumann, P.M. 2004. The spatially variable inhibition by water deficit of maize root growth correlates with altered profiles of proton flux and cell wall pH. *Plant Physiol* 135: 2291-2300.
- Farrant, J.M. 2000. A Comparison of mechanisms of Desiccation Tolerance among three-angiosperm resurrection plant species. *Plant Ecol* 151: 29-39.
- Farooq, M., Wahid, A., Kobayashi, N., Fujita, D., and Basra, S.M.A. 2009. Plant drought stress: effect, mechanisms, and management. *Agron Sustain Dev* 29: 185-212.
- Feldmann, K.A., and Marks, M.D. 1987. Agrobacterium-related transformation of germinating seeds of *Arabidopsis thaliana*: A non-tissue culture approach. *Mol Gen Genet* 208: 1-9.
- Feldmann, K.A. 1991. T-DNA insertion mutagenesis in *Arabidopsis*: Mutational spectrum. *Plant J* 1: 71-82.
- Feng, A.L., Tang, X., and Wang, X. 2000. Changes of microsomal membrane properties in spring wheat leaves (*Triticum aestivum* L.) exposed to enhanced and ultraviolet-B radiation. *J Photochem Photobio B* 57: 60–65.

- Finnegan, J., and McElroy, D. 1994. Transgene inactivation: Plant fight back! *Nat Biotechnol* 12: 883-888.
- Flexas, J., Bota, J., Loreto, F., Cornic, G., and Sharkey, T.D. 2004. Diffusive and metabolic limitations to photosynthesis under drought and salinity in C3 plants. *Plant Biol* 6: 269-279.
- Flexas, J., Ribas-Carbo, M., Hanson, D.T., Bota, J., Otto, B., Cifre, J., McDowell, N., Medrano, H. and Kaldenhoff, R. 2006. Tobacco aquaporin NtAQP1 is involved in mesophyll conductance to CO<sub>2</sub> in vivo. *Plant J* 48: 427-439
- Flexas, J., Diaz-Espejo, A., Galmes, J., Kaldenhoff, R., Medrano, H., and Ribas-Carbo, M. 2007. Rapid variations of mesophyll conductance in response to changes in CO<sub>2</sub> concentration around leaves. *Plant Cell Environ* 30:1284-1298.
- Fotovat, R., Valizadeh, M., and Toorehi, M. 2007. Association between water-use-efficiency components and total chlorophyll content (SPAD) in wheat (*Triticum aestivum* L.) under well-watered and drought stress conditions. *J Food Agric Environ* 5: 225-227.
- Fu, J., and Ristic, Z. 2010. Analysis of transgenic wheat (*Triticum aestivum* L.) harboring a maize (*Zea mays* L.) gene for plastid EF-Tu: Segregation pattern, expression and effects of the transgene. *Plant Mol Biol* 73: 339-347.
- Foyer, C.H., Valadier, M.H., Migge, A., and Becker, T.W. 1998. Drought-induced effect on nitrate reductase activity and mRNA and on the coordination of nitrogen and carbon metabolism in maize leaves. *Plant Physiol* 117:283-292.
- Frenkel, C., and Erez, A. 1996. Induction of chilling tolerance in cucumber (*Cucumis sativus*) seedlings by endogenous and applied ethanol. *Physiol Plant* 96: 593-600.
- Fukao, T., and Bailey-Serres, J. 2004. Plant response to hypoxia- is survival a balancing act? *Trands plants sci* 9:(9) 449-456.
- Galbiati, F., Volonte, D., Brown, A. M., Weinstein, D. E., Ben-Ze'ev, A., Pestell, R. G., and Lisanti, M. P. 2000. Caveolin-1 expression inhibits Wnt/beta-catenin/Lef-1 signaling by recruiting beta-catenin to caveolae membrane domains. *J Biol Chem* 275: 23368-23377.
- Garabagi, F., and Strommer, J. 2004. Distinct genes produce the alcohol dehydrogenases of pollen and maternal tissues in *Petunia hybrida*. *Biochem Genet* 42: 199-207.
- Garabagi, F., Duns, G., and Strommer, J. 2005. Selective recruitment of *Adh* genes for distinct enzymatic functions in *Petunia hybrida*. *Plant Mol Biol* 58: 283-294.

- García-Plazaola, J.I., Hernández, A., Olano, J.M., Becerril, J.M. 2003. The operation of the lutein epoxide cycle correlates with energy dissipation. *Func Plant Bio* 30: 319-324.
- Garrido, J.J., Dorado, G., and Barbancho, M. 1988. Participation of *Drosophila melanogaster* alcohol dehydrogenase (ADH) in the detoxification of 1-pentene-3-ol and 1-pentene-3-one. *Heredity* 61: 85-92.
- Gass, N., Glagotskaia, T., Mellema, S., Stuurman, J., Barone, M., Mandel, T., Roessner-Tunali, U., and Kuhlemeier, C. 2005. Pyruvate decarboxylase provides growing pollen tubes with a competitive advantage in petunia. *Plant Cell* 17: 2355–2368.
- Gaston, S., Zabalza, A., González, E.M., Arrese-Igor, C., Aparicio-Tejo, P.M., Royuela, M. 2002. Imazethapyr, an inhibitor of the branched-chain amino acid biosynthesis, induces aerobic fermentation in pea plants. *Physiol Plant* 114: 524–53.
- Gaut, B. S., Morton, B. R., Mccaig, B. M., and Clegg, M. T. 1996. Substitution rate comparisons between grasses and palms: synonymous rate differences at the nuclear gene *Adh* parallel rate differences at the plastid gene. *Acad Sci USA* 93: 10274-10279.
- Gaut, B.S., Peek, A.S., Morton, V.E., and Clegg, M.T. 1999. Patterns of genetic diversification within the *Adh* gene family in the grasses (Poaceae). *Mol Biol Evol* 16: 1086-1097.
- Geigenberger, P. 2003. Response of plant metabolism to too little oxygen. *Curr Opin Plant Bio* 6: 247-256.
- Genty, B., Briantais, J.M., and Baker, J.M. 1989. The relationship between the quantum yield of photosynthetic electron transport and quenching of chlorophyll fluorescence. *Biochimica et Biophysica Acta* 990: 87-92.
- Gibson, S.I. 2000. Plant sugar-response pathways. Part of a complex regulatory web. *Plant Physiol* 124: 1532-1539.
- Gigon, A., Matos, A., Laffray, D., Fodil, Y.Z., and Pham-Thi, A. 2004. Effect of drought stress on lipid metabolism in leaves of *Arabidopsis thaliana* (Ecotype Columbia). *Ann Bot* 94: 345-351.
- Gómez-Cadenas, A., Tadeo, F.R., Talon, M., and Primo-Millo, E. 1996. Leaf abscission induced by ethylene in water-stressed intact seedlings of *Cleopatra mandarin* requires previous abscisic acid accumulation in roots. *Plant Physiol* 112: 401-408.
- González-Aguero, M., Troncoso, S., Gudenschwager, O., Campos-Vargas, R., Moya-León, A., and Defilippi, B.G. 2009. Differential expression levels of aroma-



- related genes during ripening of apricot (*Prunus armeniaca* L.). *Plant Physiol Biochem* 47: 435-440.
- Grassi, G., and Magnani, F. 2005. Stomatal, mesophyll conductance and biochemical limitations to photosynthesis as affected by drought and leaf ontogeny in ash and oak trees. *Plant Cell Environ* 28: 834-849.
- Gu, Z., Steinmetz, L.M., Gu, X., Scharfe, C., Davis, R.W., and Li, W.H. 2003. Role of duplicate genes in genetic robustness against null mutations. *Nature* 421: 63-66.
- Hageman, R.H., and Flesher, D. 1960. The effect of an anaerobic environment on the activity of alcohol dehydrogenase and other enzymes of corn seedlings. *Arch Biochem Biophys* 87: 203-209.
- Hanson, A.D., and Hitz, W.D. 1982. Metabolic response of mesophytes to plant water deficit. *Ann Rev Plant Physiol* 33: 163-203.
- Hanson, A.D., Jacobsen, J., and Zwar, J.A. 1984. Regulated expression of three alcohol dehydrogenase genes in barley aleurone layers. *Plant Physiol* 75:573-81.
- Hanson, J., and Smeekens, S. 2009. Sugar perception and signalling-an update. *Curr Opin Plant Bio* 12: 562-567.
- Harberd, N.P., and Edwards, K.J.R. 1982. The effect of a mutation causing alcohol dehydrogenase deficiency of flooding tolerance in barley. *New Phytol* 90: 631-644.
- Hare, P.D., and Cress, W.A. 1997. Metabolic implications of stress-induced proline accumulation in plants. *Plant Growth Regul* 21: 79-102
- Hare, P.D., Cress, W.A., and Van Staden, J. 1999 Proline synthesis and degradation: a model system for elucidating stress-related signal transduction. *J Exp Bot* 50: 413-434
- Harry, D.E., and Kimmerer, T.W. 1990. Molecular genetics and physiology of alcohol dehydrogenase in woody plants. *Forest Ecol Manag* 43: 252-272.
- Hsiao, T.C. 1973. Plant responses to water stress. *Ann Rev Water Physiol* 24:519-570.
- Herzog, H. 1986. Source and sink during the reproductive period of wheat. *Scientific Publication, Berlin and Hamburg* 147-148.
- Hoffmann, A. A., Turelli, M., and Simmons, G. M. 1986. Unidirectional incompatibility between populations of *Drosophila simulans*. *Evolution* 40: 692-701.

- Hohl, M., and Schopfer, P. 1991. Water relations of growing maize coleoptiles. Comparison between manitol and polyethylene glycol 6000 as external osmotica for adjusting turgor pressure. *Plant Physiol* 95: 716-722.
- Hong, Z., Lakkineni, K., Zhang, Z., and Verma, D. P. S. 2000. Removal of feedback inhibition of pyrroline-5-carboxylate synthetase results in increased proline accumulation and protection of plants from osmotic stress. *Plant Physiol* 122: 1129-1136.
- Hu, C.A.A., Delauney, A.J., and Verma, D.P.S. 1992. A bifunctional enzyme  $\Delta^1$ -pyrroline-5-carboxylate synthetase catalyzes the first two steps in proline biosynthesis in plants. *Proc Natl Acad Sci USA* 89: 9354-9358.
- Gottlieb, L.D. 1982. Evolution of alcohol dehydrogenase genes in the palm and grass families. *Science* 216: 373-380.
- Hummel, I., Pantin, F., Sulpice, R., Piques, M., Rolland, G., Dauzat, M., Christophe, A., Pervent, M., Bouteillé, M., Stitt, M. 2010. *Arabidopsis* plants acclimate to water deficit at low cost through changes of carbon usage: an integrated perspective using growth, metabolite, enzyme, and gene expression analysis. *Plant Physiol* 154: 357-372.
- Ingersoll, J.C., Rothenberg, M., Liedl, B.E., Folkerts, K., Garvin, D., Hansom, M.R., Doyle, J.J., and Mutschler, M. 1994. A novel anther-expressed *adh*-homologous gene in *Lycopersicon*. *Plant Mol Biol* 26:6 1875-1891.
- Ingram, J., and Bartels, D. 1996. The molecular basis of dehydration tolerance in plants. *Ann Rev Plant Mol Biol* 47: 377-403.
- Ismond, K.P., Dolferus, R., De Pauw, M., Dennis, E.S., and Good, A.G. 2003. Enhanced low oxygen survival in *Arabidopsis* through increased metabolic flux in the fermentative pathway. *Plant Physiol* 132: 1292-1302.
- IturbeOrmaetxe, I., Escuredo, P.R., Arrese-Igor, C., and Becana, M. 1998. Oxidative damage in pea plant expose to water deficit or paraquat. *Plant Physiol* 116: 173-181.
- Iturriaga, G., Suarez, R., and Nova-Franco, B. 2009. Trehalose metabolism: from osmoprotection to signaling. *Int J Sci* 10: 3793-3810.
- Jacobs, M., Dolferus, R., and Van Den Bossche, D. 1988. Isolation and biochemical analysis of ethyl methanesulfonate-induced alcohol dehydrogenase mutants of *Arabidopsis thaliana* (L.) Heynh *Biochem Genet* 26: 105-122.
- Jager, H.J., and Meyer, H.R. 1977. Effect of water stress on growth and proline metabolism of *Phaseolus vulgaris* L. *Oecologia* 30: 83-96.

- Jaleel, C.A., Manivannan, P., Sankar, B., Kishorekumar, A., Gopi, R., Somasundaram, R., and Panneerselvam, R. 2007. Induction of drought stress tolerance by ketoconazole in *Catharanthus roseus* is mediated by enhanced antioxidant potentials and secondary metabolite accumulation. *J Colloids Surf B Biointerfaces* 60: 201-206.
- Jaleel, C.A., Sankar, B., Murali, P.V., Gomathinayagam, M., Lakshmanan, G.M.A., and Panneerselvam, R. 2008. Water deficit stress effects on reactive oxygen metabolism in *Catharanthus roseus*; impacts on ajmalicine accumulation. *J Colloids Surf B Biointerfaces* 62: 105-111.
- James, R.A., von Caemmere, S., Condon, A.G., Zwart, A.B., and Munns, R. 2008. Genetic variation in tolerant to the osmotic stress component of salinity stress in durum wheat. *Func Plant Bio* 35: 111-123.
- Jennings, P., and Saltveit, M.E. 1994. Temperature and chemical shocks induce chilling tolerance in germinating *Cucumis sativus* (cv. Poinsett 76) seeds. *Physiol Plant* 91: 703-7.
- Jia, W.S., and Davies, W.J. 2007. Modification of leaf apoplastic pH in relation to stomatal sensitivity to root-sourced ABA signals. *Plant Physiol* 143: 68-77.
- Kalefetoglu, T., and Ekmekci, Y. 2005. The effect of drought on plants and tolerance mechanisms. *Gazi Univ J Sci* 18: 723-740.
- Kao, C.H. 1981. Senescence of rice leaves. Comparative study of the metabolic changes of senescing turgid and water stressed excised leaves. *Plant Cell Physiol* 22:683-685.
- Karamanos, A.S. 1980. Water stress and leaf growth of field beans (*Vicia faba*) in the field: Leaf number and total area. *Ann Bot* 42:1393-1402.
- Karimi, M., Inze, D., and Depicker, A. 2002. GATEWAY™ vectors of Agrobacterium-mediated plant transformation. *Trends Plant Sci* 7: 193-195.
- Kato-Noguchi, H. 2000. Evaluation of the importance of lactate for the activation of ethanolic fermentation in lettuce roots in anoxia. *Physiol Plant* 109:28-33.
- Kato-Noguchi, H. 2001. Wounding stress induces alcohol dehydrogenase in maize and lettuce seedlings. *Plant Growth Regul* 35: 285-288.
- Kato-Noguchi, H. 2002. Ethanol sensitivity of rice and oat coleoptiles. *Physiol Plant* 115:119-24.
- Kato-Noguchi, H. 2006. Pyruvate metabolism in rice coleoptiles under anaerobiosis. *Plant Growth Regul* 50: 41-46.

- Kato-Noguchi, H. 2007. Low temperature acclimation to chilling tolerance in rice roots. *Plant growth regul.* 51: (2)171-175.
- Kato-Noguchi, H., Yasuda, Y., and Sasaki, R. 2010. Soluble sugar availability of aerobically germinated barley and oat and rice coleoptiles in anoxia. *J Plant Physiol* 167: 1571- 1576.
- Katzen, F. 2007. Gateway recombinational cloning: a biological operating system. *Expert Opin Drug Discov* 2: 571-589.
- Kavi Kishor, P.B., Hong, Z., Miao, G.H., Hu, C.A.A., and Verma, D.P.S. 1995. Overexpression of  $\Delta^1$  pyrroline-5-carboxylate synthetase increases proline production and confers osmotolerance in transgenic plants. *Plant Physiol* 108: 1387-1394.
- Kavi Kishor, P.B., Sangam, S., Amruth, R.N., Sri Laxmi, P., Naidu, K.R., Rao, K.R.S.S., Sreenath Rao Reddy, K.J., Theriappan, P., and Sreenivasulu, N. 2005. Regulation of proline biosynthesis, degradation, uptake and transport in higher plants: its implications in plant growth and abiotic stress tolerance. *Curr Sci* 88: 424-438.
- Kawamitsu, Y., Driscoll, T., and Boyer, S.J. 2000. Photosynthesis during desiccation in an Intertidal Algae and Land Plant. *Plant Cell Physiol* 41(3): 344-353.
- Kelly, M.O., and Saltveit, M.E. 1988. Effect of endogenously synthesized and exogenously applied ethanol on tomato fruit ripening. *Plant Physiol* 88: 143-147.
- Kennedy, R.A., Rumpho, M.E., and Fox, T.C. 1992. Anaerobic metabolism in plants. *Plant Physiol* 100:1-6.
- Kimmerer, T.W., and Kozolowski, T. 1982. Ethylene, Ethane, Acetaldehyde, and Ethanol production by plants under Stress. *Plant Physiol* 69: 840-847.
- Kimmerer, T.W., and MacDoland, R.C. 1987. Acetaldehyde and ethanol biosynthesis in leaves of plants. *Plant Physiol* 84: 1204-1209
- Kimmerer, T.W., and Stringer, M.A. 1988. Alcohol dehydrogenase and ethanol in the stems of trees. *Plant Physiol* 87:693-697.
- Kimmerer, T.W. 1990. Structure and function of forest tree. Young, R.A., and Giese, R.L. eds; Introduction to forest science. John Wiley & sons, New York.
- Krieger, F., Spinka, M., Golbik, R., Hübner, G., and König, S. 2002. Pyruvate decarboxylase from *Kluyveromyces lactis*. An enzyme with extraordinary substrate activation behaviour. *Eur J Biochem* 269: 3256–3263.

- Knapp, A.K., Briggs, J.M., and Koelliker, J.K. 2001. Frequency and extent of water limitation to primary production in a mesic temperate grassland. *Ecosystems* 4: 19-28.
- Knee, M., and Hatfield, S.G.S., 1981. The metabolism of alcohols by apple fruit tissue. *J Sci Food Agric* 32: 593–600.
- Koch, K.E. 1996. Carbohydrate-modulated gene expression in plants. *Ann Rev Plant Physiol Plant Mol Biol* 47: 509-540.
- Kramer, P.J., and Boyer, J.S. 1995. Water Relations of Plants. *Acad press* 1995.
- Krapp, A., Hofmann, B., Schäfer, C., and Stitt, M. 1993. Regulation of the expression of rbcS and other photosynthetic genes by carbohydrates: a mechanism for the “sink regulation” of photosynthesis? *Plant J* 3:817-828.
- Krysan, P.J., Young, J.C., and Susman, M.R. 1999. T-DNA as an insertional mutagen in Arabidopsis. *Plant Cell* 11: 2283-2290.
- Kumutha, D., Sairam, R.K., and Meena, R. C. 2008. Role of root carbohydrate reserves and their mobilization in imparting water logging tolerance in green gram (*Vigna radiata* (L.) Wilczek) genotypes. *Ind J Plant Physiol* 13: 339-346.
- Kusaka, M., Ohta, M., and Fujimura, T. 2005. Contribution of inorganic components to osmotic adjustment and leaf folding for drought tolerance in pearl millet. *Physiol Plant* 125: 474-489
- Kypouris, A., Petropoulou, Y., and Manetas, Y. 1995. Summer survival of leaves in a soft-leaved shrub (*Phlomis fruticosa* L., Labiatae) under Mediterranean field conditions: avoidance of photoinhibitory damage through decreased chlorophyll contents. *J Exp Bot* 46: 1825-1831.
- Lambers, H., Chapin III, F.S., and Pons, T.L. 1998. Plant physiological ecology. *Springer, Berlin*, pp 540.
- Laszlo, A., and St Lawrence, P. 1983. Parallel induction of PDC and ADH in anoxic maize roots. *Mol Gen Genet* 192: 110-117.
- Lawlor, D.W., and Cornic, G. 2002. Photosynthetic carbon assimilation and associated metabolism in relation to water deficits in higher plants. *Plant Cell Environ* 25: 275-294.
- Lawlor, D.W. 2009. Musings about the effects of environment on photosynthesis. *Ann Bot* 103: 543-549.

- Lawlor, D.W., and Tezara, W. 2009. Causes of decreased photosynthetic rate and metabolic capacity in water-deficient leaf cells: a critical evaluation of mechanisms and integration of processes. *Ann Bot* 103: 561-579.
- Le, T.N., and McQueen-Mason, S. J. 2006. Desiccation-tolerant plants in dry environments. *Rev Environ Sci Biotechnol* 5:269-279.
- Leblova, S., Sinecka, E., and Vanickova, V. 1974. Pyruvate metabolism in germinating seeds during natural anaerobiosis. *Biol Plant* 16: 406-411.
- Lechtenberg, B., Schubert, D., Forsbach, A., Gils, M., and Schmidt, R. 2003. Neither inverted repeat T-DNA configurations nor arrangements of tandemly repeated transgenes are sufficient to trigger transgene silencing. *Plant J.* 34: 507-517
- Lee, S., Lee, E.J., Yang, E.J., Lee, J.E., Park, A.R., Song, W.H., and Park, O.K. 2004. Proteomic identification of annexins, calcium-dependent membrane binding proteins that mediate osmotic stress and abscisic acid signal transduction in *Arabidopsis*. *Plant Cell* 16:1378-1391.
- Lemke-Keyes, C.A., and Sach, M.M. 1989. Anaerobic tolerant null: a mutant that allows *Adh1* nulls to survive anaerobic treatment. *J Heredity* 80: 316-319.
- Leung, J., and Giraudat, J. 1998. Abscisic acid signal transduction. *Ann Rev Plant Physiol Plant Mol Biol* 49:199-222.
- Li, Y., Ye, W., Wang, M., and Yan, X. 2009. Climate change and drought: a risk assessment of crop-yield impacts. *Clim Res* 39: 31-46.
- Lin, M., and Oliver, D.J. 2008. The role of acetyl-coenzyme a synthetase in *Arabidopsis*. *Plant Physiol* 147: 1822-1829.
- Liu, F., Andersen, M.N., and Jensen, C.R. 2004. Root signal controls pod growth in drought-stressed soybean during the critical, abortion-sensitive phase of pod development. *Field Crops Research* 85: 159-166.
- Longhurst, T.J., Tung, H.F., and Brady, C.J. 1990. Developmental regulation of the expression of alcohol dehydrogenase in ripening tomato fruit. *J Food Biochem* 14: 421-433.
- MacDonald, R.C., Kimmerer, T.W., and Stringer, J.W. 1989. Remetabolism of transpirational ethanol by the leaves of eastern cottonwood (*Populus deltoids* Bartr.) In Proc. IUFRO-EcoPhys Workshop, Rhinelander, WI, p.47.
- MacDonald, R.C., and Kimmerer, T.W. 1990. Remetabolism of transpired ethanol by *Populus deltoids*. *Plant Physiol* 93: s-112.

- Mafakheri, A., Siosemardeh, A., Bahramnejad, B., Struik, P.C., and Sohrabi, Y. 2010. Effect of drought stress on yield, proline and chlorophyll contents in three chickpea cultivars. *Aust J Crop Sci* 4(8):580-585.
- Magdaleno, A., Ahn, I-Y., Paes, L.S., Silber, A.M. 2009. Actions of a Proline Analogue, L-Thiazolidine-4-Carboxylic Acid (T4C), on *Trypanosoma cruzi*. PLoS ONE 4(2): e4534. doi:10.1371/journal.pone.0004534.
- Maggio, A., Miyazaki, S., Veronese, P., Fujita, T., Ibeas, J.I., Damsz, B., Narasimhan, M.L., Hasegawa, P.M., Joly, R.J., and Bressan, R.A. 2002. Dose proline accumulation play an active role in stress-induced growth induction. *Plant J* 31:699-712.
- Margo, M.J.C., John, V., Linus, H.W.van der Plas., Alexander, R.van der Krol and Dick, V. 2005. Ethanol breaks dormancy of the potato tuber apical bud. *J Exp Bot* 56; 2515-2525.
- Maruyama, H., Koyama, R., Oi, T., Yagi, M., Takeda, M., Kanechi, M., Inagaki, N., and Uno, Y. 2008. In vitro evaluation of osmotic stress tolerance using a novel root recovery assay. *Plant Cell Tiss Organ Cult* 95:101–106
- Matsumura, H., Takano, T., Takeda, G., and Uchimiya, H. 1998. *Adh1* is transcriptionally active but its translational product is reduced in a rad mutant of rice (*Oryza sativa* L.), which is vulnerable to submergence stress. *Theor Appl Genet* 97:1197-1203.
- Matton, D.P., Constabel, P., and Brisson, N. 1990. Alcohol dehydrogenase gene expression in potato following elicitor and stress treatment. *Plant Mol Biol* 14: 775-783.
- Matzke, M.A., Mette, M.F., and Matzke, A.J. 2000. Transgene silencing by the host genome defense: implications for the evolution of epigenetic control mechanisms in plants and vertebrates. *Plant Mol Biol* 43: (2-3) 401-15.
- McMichael, A.J. 2011. Climate Change and Health: Policy Priorities and Perspectives. Briefing paper. www.chathamhouse.org.
- Meinshausen, M., Meinshausen, N., Hare, W., Raper, S.C.B., Frieler, K., Knutti, R., Frame, D.J., and Allen, M.R. 2009. 'Greenhouse-gas emission targets for limiting global warming to 2 °C. *Nature* 458: 1158–63.
- Melmma, S., Eichenberger, W., Rawlter, A., Suter, M., Tadege, M., and Kuhlemeier, C. 2002. The ethanolic fermentative pathway supports respiration and lipid biosynthesis in tobacco pollen. *Plant J* 30: (3) 329-336.

- Miyashita, R., and Good A.G. 2007. Contribution of the GABA shunt to hypoxic-induced alanine accumulation in roots of *Arabidopsis thaliana*. *Plant Cell physiol* 49: 92-102.
- Molina, I., Nicolas, M., and Crouzet, J. 1986. Grape alcohol dehydrogenase. I. Isolation and characterization. *Am J Ecol Viticulture* 37: 169-173.
- Mott, K.A., and Parkhurst, D. F. 1991. Stomatal responses to humidity in air and helox. *Plant Cell Environ* 14: 509-515.
- Müller, M., Mentel, M., van Hellemond, J., Henze, K., Wöhle, C., Gould, S.B., Yu, R.Y., van der Giezen, M., Tielens, A.G.M., and Martin, W.F. 2012. Biochemistry and evolution of anaerobic energy metabolism in eukaryotes. *Microbiol Mol Biol Rev* 76: 444-495.
- Munns, R., James, R.A., Sirault, X.R.R., Furbank, R.T., and Jones, H.G. 2010. New phenotyping methods for screening wheat and barley for beneficial responses to water deficit. *J Exp Bot* 61:(13) 3499-3507.
- Mustroph, A., and Albrecht, G. 2003. Tolerance of crop plants to oxygen deficiency stress: fermentative activity and photosynthetic capacity of entire seedlings under hypoxia and anoxia. *Physiol Planta* 117: 508-520.
- Nakajima, K., Furutani, I., Tachimoto, H., Matsubara, H., and Hashimoto, T. 2004. *SPIRAL1* encodes a plant-specific microtubule-localized protein required for directional control of rapidly expanding *Arabidopsis* cells. *Plant Cell* 16:1178-1190.
- Nelson, S.E., and Assmann, S.M. 2007. The control of transpiration: Insights from *Arabidopsis*. *Plant physiol* 143: 19-27.
- Nayyer, H., and Halia, D.P. 2003. Water stress induced proline accumulation in contrasting wheat genotypes as affected by calcium and abscisic acid. *Biol Plant* 46: 275-279.
- Nayyar, H., Kaur, S., Singh, S., and Upadhyaya, H.D. 2006. Differential sensitivity of Desi (small-seeded) and Kabuli (large-seeded) chickpea genotypes to water stress during seed filling: effects on accumulation of seed reserves and yield. *J Sci Food Agric* 86: 2076-2082.
- Nageswara Rao, R.C., Talwar, H.S., and Wright, G.C. 2001. Rapid assessment of specific leaf area and leaf nitrogen in peanut (*Arachis hypogaea* L.) using a chlorophyll meter. *J Agron Crop Sci* 189: 175-182.
- Nakashima, K., Satoh, R., Kiyosue, T., Yamaguchi-Shinozaki, K., and Shinozaki, K. 1998. A gene encoding proline dehydrogenase is not only induced by proline and



hypoosmolarity, but is also developmentally regulated in the reproductive organs of *Arabidopsis*. *Plant Physiol* 118: 1233–1241.

- Noctor, G., Veljovic-Jovanovic, S., Driscoll, S., Novitskaya, L., and Foyer, C.H. 2002. Drought and oxidative load in leaves of C<sub>3</sub> plants: a predominant role for photorespiration? *Ann Bot* 89: 841–850.
- Nonami, H. 1998. Plant water relations and control of cell elongation at low water potentials. *J Plant Res* 111: 373-382.
- Norwood, M., Toldi, O., Richter, A., and Scott, P. 2003. Investigation into the ability of roots of the poikilohydric plant *Craterostigma plantagineum* to survive dehydration stress. *J Exp Bot* 54: 2313-2321.
- Novillo, F., Alonso, J.M., Ecker, J.R., and Salinas, J. 2004. CBF2/DREB1C is a negative regulator of CBF1/DREB1B and CBF3/DREB1A expression and plays a central role in stress tolerance in *Arabidopsis*. *Proc Natl Acad Sci USA* 101:3985-3990.
- Oertli, J.J. 1985. The response of plant cells to different forms of moisture stress. *J Plant Physiol* 121: 295–300.
- Okimoto, R., Sachs, M.M., Porter, E.K., and Freeling, M. 1980. Patterns of polypeptide synthesis in various maize organs under anaerobiosis. *Planta* 150: 89-94.
- Oliver, M.J., Tuba, Z., and Mishler, B.D. 2000. The evolution of vegetative desiccation tolerance in land plants. *Plant Ecol* 151: 85-100.
- Oliver, J.M., Cushman, J.C., and Koster, K.L. 2010. Dehydration tolerance in plant. In: *Sunkar (Ed.), Plant Stress Tolerance, Methods Mol Bio*. Humana Press, New York. 639:3-24.
- Ommen, O.E., Donnelly, A., Vanhoutvin, S., van Oijen M., and Manderscheid, R. 1999. Chlorophyll content of spring wheat flag leaves grown under elevated CO<sub>2</sub> concentrations and other environmental stresses within the ESPACE-wheat project. *Eur J Agron* 10: 197-203.
- Parkin, K.L., and Kuo, S.J. 1989. Chilling-induced lipid degradation in cucumber (*Cucumis sativus* L., cv. Hybrid C) fruit. *Plant Physiol* 90: 1049–56.
- Passioura, J.B. 2006. The perils of pot experiment. *Func Plant Bio* 33: 1075-1079.
- Pathan, M.S., Subudhi, P.K., Courtois, B., and Nguyen, H.T. 2004. Molecular dissection of abiotic stress tolerance in sorghum and rice. In *Physiology and Biotechnology Integration for Plant Breeding*. Edited by Nguyen HT, Blum A. Marcel Dekker, Inc.; 525-569.

- Pego, J.V., Kortstee, A.J., Huijser, C. and Smeekens, S.C.M. 2000. Photosynthesis, sugars and the regulation of gene expression. *J Exp Bot* 51: 407-416.
- Peng, Z., Lu, Q., and Verma, D.P. 1996. Reciprocal regulation of  $\Delta^1$ -pyrroline-5-carboxylate synthetase and proline dehydrogenase genes controls proline levels during and after osmotic stress in plants. *Mol Gen Genet* 253: 334-341.
- Peng, H.P., Chan, C.S., Shih, M.C., and Yang, S.F. 2001. Signalling events in the hypoxic induction of alcohol dehydrogenase gene in *Arabidopsis*. *Plant Physiol* 126: 742-749.
- Pessarakli, M. 1999. Response of green beans (*Phaseolus vulgaris* L.) to salt stress. In Pessarakli, M (Ed). *Handbook of Plant and Crop Stress 2ed*. MARCED LEKKERIN, C. E.E.U.U., pp 827-842.
- Pereira, J.S., and Chaves, M.M. 1993. Plant water deficits in Mediterranean ecosystems. In: Smith JAC, Griffiths H, eds. *Plant responses to water deficits from cell to community*. Oxford: *BIOS Scientific*, 237-251.
- Petel, G., Candelier, P., and Gendraud, M. 1993. Effect of ethanol on filiate tubers of *Jerusalem artichoke*: a new tool to study tuber dormancy. *Plant Physiol Biochem* 31: 67-71.
- Peters, J.S., and Frenkel, C. 2004. Relationship between alcohol dehydrogenase activity and low-temperature in two maize genotypes, Silverado F1 and *Adh1\_Adh2* doubly null. *Plant Physiol Biochem* 42:841-846.
- Pinheiro, C., Chaves, M.M., and Ricardo, C.P. 2001. Alterations in carbon and nitrogen metabolism induced by water deficit in stem and leaves of *Lupinus albus* (L.). *J Exp Bot* 52: 1063-1070.
- Pinheiro, C., and Chaves, M.M. 2011. Photosynthesis and drought: can we make metabolic connections from available data? *J Exp Bot* 62: ( 3) 869-882.
- Porra, R.J. 2002. The chequered history of the development and use of simultaneous equations for the accurate determination of chlorophylls a and b. *Photosynth Res* 73: 149-156.
- Podd, L.A., and vanStaden, J. 1999. Is acetaldehyde the causal agent in the retardation of carnation flower senescence by ethanol? *J Plant Physiol* 154: 351–354.
- Preiszner, J., VanToai, T., Huynh. L., Bolla, R., and Yen, H. 2001. Structure and activity of a soybean *Adh* promoter in transgenic hairy roots. *Plant Cell Rep* 20: (8) 763-769).
- Pronk, J.T., Wenzel, T.J., Luttik, M.A.H., Klaassen, C.C.M., Scheffer, W.A., Steensma, H.Y., and Dijken, J.P.van. 1994. Energetic aspects of glucose metabolism in a

- pyruvate dehydrogenase-negative mutant of *Saccharomyces cerevisiae*. *Microbiology* 140: 601-610.
- Purvis, A.C., and Grierson, W., 1982. Accumulation of reducing sugar and resistance of grapefruit peel to chilling injury as related to winter temperatures. *J Am Soc Hortic Sci* 107, 139±142.
- Qiu, J. 2010. China drought highlights future climate threats. *Nature* 465: (7295) 142-143.
- Raghavendra, A.S., Gonugunta, A., Christmann., and Grill, E. 2010. ABA perception and signaling. *Trends Plant Sci.* 15:395-401.
- Rahnama, A., Poustini, K., Munns, R., and James, R.A. 2010. Stomata conductance as a screen for osmotic stress tolerance in durum wheat growing in saline soil. *Func Plant Biolo* 37: 255-265.
- Rai, M.K, Kalia, R.K, Singh, R, Gangola, M.P., and Dhawana, A.K. 2010. Developing stress tolerant plants through in vitro selection—An overview of the recent progress. *Enviro Exp Bot* 71: 89-98.
- Ramanjulu, S., and Bartels, D. 2002. Drought- and desiccation-induced modulation of gene expression in plants. *Plant Cell Environ* 25(2): 141-151.
- Ramel, F., Sulmon, C., Gouesbet, G., and Couee, I. 2009. Natural variation reveals relationships between pre-stress carbohydrate nutritional status and subsequent responses to xenobiotic and oxidative stress in *Arabidopsis thaliana*. *Ann Bot* 104: 1323-1337.
- Ricard, B., and Pradet, A. 1989. Anaerobic protein synthesis in different organs of germinating rice seeds. *Plant Physiol Biochem* 27: 761-768.
- Ricard, B., VanToai, T., Chourey, P., and Saglio, P. 1998. Evidence for the critical role of sucrose synthase for anoxic tolerance of maize roots using a double mutant. *Plant Physiol* 116:1323-1331.
- Riechmann, J.L., and Ratcliffe, O.J. 2000. A genomic perspective on plant transcription factors. *Curr Opin Plant Biol* 3: 423-434.
- Riveras-Rosas, H., Julian-Sanchez, A., and Pina, E. 1997. Enzymology of ethanol and acetaldehyde metabolism in mammals. 28 (4); 453-471.
- Ritchie, S.W., Nguyen, H.T., and Holaday, A.S. 1990. Leaf water content and gas exchanges parameters of two wheat genotypes differing in drought resistant. *Crop Sci* 30: 105-111.

- Rizal, G., and Karki, S. 2012. Alcohol dehydrogenase (ADH) activity in soybean *Glycine max* under flooding stress. *J plant breed* 2: (1) 50-57.
- Roberts, J. K. M., Callis, J., Wemmer, D., Walbot, V., and Jardetzky, O. 1984. Mechanisms of cytoplasmic pH regulation in hypoxic maize root tips and its role in survival under anoxia. *Proc Natl Acad Sci USA* 81: 3368-3372.
- Roberts, J. K. M., Callis, J., Jardetzky, O., Walbot, V., and Freeling, M. 1985. Cytoplasmic acidosis as a determinant of flooding intolerance in plants. *Proc Natl Acad Sci USA* 81: 6029-6033.
- Rolland, F., Baena-Gonzalez, E., and Sheen, J. 2006. Sugar sensing and signaling in plants: conserved and novel mechanisms. *Ann Rev Plant Biol* 57: 675-709.
- Rosa, M., Prado, C., Podazza, G., Interdonato, R., González, J.A., Hilal, M., and Prado, F.E. 2009. Soluble sugars: metabolism, sensing and abiotic stress. A complex network in the life of plants. *Plant Signal Behav* 4: 388-393.
- Rosso, M.G., Li, Y., Strizhov, N., Reis, B., Dekker, K., and Weisshaar, B. 2003. An *Arabidopsis thaliana* T-DNA mutagenized population (GABI-Kat) for flanking sequence tag-based reverse genetics. *Plant Mol Biol* 53:247-259.
- Routley, D.G. 1996. Proline accumulation in wilted Ladino clover leaves. *Crop Sci.* 6: 358-361.
- Ruhland and Ramshorn. 1938. Aerobe Garung in aktiven pflanzlichen Meristemen. *Planta* 28:471-514.
- Russell, D.A., and Sachs, M.M. 1989. Differential expression and sequence analysis of the maize glyceraldehyde-3-phosphate dehydrogenase gene family. *Plant Cell* 1: 793-803.
- Russell, D.A, Wong, D.M.L., and Sachs, M.M. 1990. The anaerobic response of soybean. *Plant Physiol* 92: 401-407.
- Russell, R (16 May 2007). "The Greenhouse Effect & Greenhouse Gases". University Corporation for Atmospheric Research Windows to the Universe.
- Sachs, M.M., Freeling, M., and Okimoto, R. 1980. The anaerobic proteins of maize. *Cell* 20: 761-767.
- Sade, B., Soylu, S., and Yetim, E. 2011. Drought and oxidative stress. *Afri J Biotechno* 10(54): 11102-11109.
- Sairam, R. K., and Saxena, D. C. 2000. Oxidative stress and antioxidant wheat genotypes: possible mechanism of water stress tolerance. *J Agro Crop Sci* 184: 55-61.

- Sairam, R.K., Kumutha, D., Ezhilmathi, K., Deshmukh, P.S., and Srivastava, G.C. 2008. Physiology and biochemistry of waterlogging tolerance in plants. *Biol Plant* 52: 401- 412.
- Saltveit, M.E. 1994. Exposure to alcohol vapours reduces chilling-induced injury of excised cucumber cotyledons, but not of seedlings or excised hypocotyls segments. *J Exp Bot* 45:813-21.
- Saltveit, M.E., and Hepler, P.K. 2004. Effect of heat shock on the chilling sensitivity of trichomes and petioles of African violet (*Saintpaulia ionantha*). *Physiol Plant* 121: 35–43.
- Sambrook, J., Fritsch, E. F., and Maniatis, T. 1989. Molecular Cloning: A Laboratory Manual, Ed 2. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY.
- Sanchez, F.J., Manzanares, M., de Andres, E.F., Tenorio, J. L., and Ayerbe, L. 1998. Turgor maintenance, osmotic adjustment and soluble sugar and proline accumulation in pea cultivars in response to water stress. *Field Crops Res* 59: 225-235.
- Schiermeier, Q. 2011. Increased flood risk link to global warming. *Nature* 470: (7334) 316.
- Schmutz, J., Cannon, S. B., Schlueter, J. A., Ma, J., and Mitros, T. 2010. Genome sequence of the paleopolyploid soybean. *Nature* 463: 178–183.
- Schonfeld, M.A., Johnson, R.C., Carver, B.F., and Mornhinweg, D.W. 1988. Water relations in winter wheat as drought resistance indicator. *Crop Sci.* 28: 526-531.
- Schwartz, D., and Endo, T. 1966. Alcohol dehydrogenase polymorphism in maize simple and compound loci. *Genetics.* 53: 709-715.
- Schwartz, D. 1971. Genetic control of alcohol dehydrogenase – a competition model for regulation of gene action. *Genetics* 67: 411–425.
- Seki, M., Narusaka, M., Abe, H., Kasuga, M., Yamaguchi-Shinozaki, K., Caminci, P., Hayashizaki, Y., and Shinozaki, K. 2001. Monitoring the expression pattern of 1300 Arabidopsis genes under drought and cold stresses using a full-length cDNA microarray. *Plant Cell* 13, 61-72.
- Seki, M., Narusaka, M., Ishida, J., Nanjo, T., Fujita, M., Oono, Y., Kamiya, A., Nakajima, M., Enju, A., and Sakurai, T. 2002. Monitoring the expression profiles of 7000 Arabidopsis genes under drought, cold and high-salinity stresses using a full-length cDNA microarray. *Plant J* 31: 279-292.
- Senthil-Kumar, M., Hema, R., Suryachandra, T.R., Ramegowda, H.V., Gopalakrishna, R., Rama, N., Udayakumar, M., and Mysore, K.S. 2010. Functional

characterization of three water deficit stress-induced genes in tobacco and Arabidopsis: An approach based on gene down regulation. *Plant Physiol Biochem* 48: 35-44.

Sessions, A., Burke, E., Presting, G., Aux, G., McElver, J., Patton, D., Dietrich, B., and Ho, P. 2002. A high-throughput *Arabidopsis* reverse genetic system. *Plant Cell* 14: (12) 2985-2994.

Shafqat, J., El-Ahmad, M., Danielsson, O., Martinez, M.C., Persson, B., Parés, X., and Jörnvall, H. 1996. Pea formaldehyde-active class III alcohol dehydrogenase: common derivation of the plant and animal forms but not of the corresponding ethanol-active forms (classes I and P). *Proc Natl Acad Sci USA* 93: 5595–5599.

Sharp, R. E. 2002. Interaction with ethylene: changing views on the role of abscisic acid in root and shoot growth responses to water stress. *Plant Cell Environ* 25: 211-222.

Shavrukov, Y., Genc, Y., and Hayes, J. 2012. The use of hydroponics in abiotic stress tolerance research. In: Asao T, (Ed), *Hydroponics – a standard methodology for plant biological researches* InTech Open Access Publisher pp 39-66.

Shevyakova, N. I. 1984. Metabolism and the physiological role of proline in plants under conditions of water and salt stress. *Soviet Plant Physiol* 30:597-608.

Shiao, T, Ellis, M.H., Dolferus, R., Dennis, E.S., and Doran, P.M. 2002. Overexpression of alcohol dehydrogenase or pyruvate decarboxylase improves the growth of hairy roots under hypoxia. *Biotechno Bioeng* 77: 455-461.

Shilpi, M., and Narendra, T. 2005. Cold, salinity and drought stresses: *An overview*. *Arch Biochem Biophys* 444: 139-158.

Shimokawa, K., and Kasai, Z. 1966. Biogenesis of ethylene in apple tissue. Formation of ethylene from glucose, acetate, pyruvate and acetaldehyde in apple tissue. *Plant Cell Physiol* 7: 1-9.

Shinozaki, K., and Yamaguchi-Shinozaki, K. 1996. Molecular responses to drought and cold stress. *Curr Opin Biotechnol* 7: 161-167.

Shinozaki, K., and Yamaguchi-Shinozaki, K. 1997. Gene expression and signal transduction in water-stress response. *Plant Physiol* 115: 327-334.

Shinozaki, K., Yamaguchi-Shinozaki, K., and Seki, M. 2003. Regulatory network of gene expression in the drought and cold stress responses. *Curr Opin Plant Biol* 6:410-417.

- Shinozaki, K., and Yamaguchi-Shinozaki, K. 2006. Transcriptional regulatory networks in cellular responses and tolerance to dehydration and cold stresses. *Annu Rev Plant Biol* 57:781-803.
- Shinozaki, K., and Yamaguchi-Shinozaki, K. 2007. Gene networks involved in drought stress response and tolerance. *J Exp Bot* 58: 221-227.
- Small, R.L., and Wendel, J.D. 2000. Copy number lability and evolutionary dynamics of the *Adh* gene family in diploid and tetraploid cotton (*Gossypium*). *Genetics* 155: 1913-1926.
- Smeets, K., Ruytinx, J., Van Bellegheem, F.V., Semane, B., Lin, D., Vangronsveld, J., and Cuypers, A. 2008. Critical evaluation and statistical validation of a hydroponic culture system for *Arabidopsis thaliana*. *Plant Physiol Biochem* 46(2):212-218.
- Smirnoff, N., and Cumbes, Q.J. 1989. Hydroxyl radical scavenging activity of compatible solute. *Phytochemistry* 28:1057-1060.
- Silvente, S., Sobolev, A.P., and Lara, M. 2012. Metabolite Adjustments in Drought Tolerant and Sensitive Soybean Genotypes in Response to Water Stress. *PLoS ONE* 7(6): e38554. doi:10.1371/journal.pone.0038554.
- Speirs, J., Lee, E., Holt, K., Yong-Duk, K., Steele Scott, N., Loveys, B., and Schuch, W. 1998. Genetic manipulation of alcohol dehydrogenase levels in ripening tomato fruit affects the balance of some flavour aldehydes and alcohols. *Plant Physiol* 117: 1047-1058.
- Spollen, W.G., LeNoble, M.E., Samuels, T.D., Bernstein, N., and Sharp, R.E. 2000. Abscisic acid accumulation maintains maize primary root elongation at low water potentials by restricting ethylene production. *Plant Physiol* 122: 967-976.
- Shao, H.B., Chu, L.Y., Shao, M.A., Abdul Jaleel, C., and Hong-Mei, M. 2008. Higher plant antioxidants and redox signaling under environmental stresses. *Comp Rend Biol* 331: 433-441.
- Singh, R.K., Sane, V.A., Misra, A., Ali, S.A., and Nath, P. 2010. Differential expression of the mango alcohol dehydrogenase gene family during ripening. *Phytochemistry* 71:1485-94.
- Stewart, C.R. 1981. Proline accumulation: Biochemical aspects. In: Paleg LG, Aspinall D (Eds), *Physiology and Biochemistry of drought resistance in plants*. pp, 243-251.
- Stitt, M. 1991. Rising CO<sub>2</sub> levels and their potential significance for carbon flow in photosynthetic cells. *Plant Cell Environ* 14: 7431-762.

- Stitt, M., Gibon, Y., Lunn, J.E., and Piques, M. 2007. Multilevel genomics analysis of carbon signalling during low carbon availability: coordinating the supply and utilisation of carbon in a fluctuating environment. *Funct Plant Biol* 34: 526-549.
- Strommer, J. 2011. The plant genome: An evolutionary view on structure and function; The plant ADH gene family. *Plant J* 66: 128-142.
- Sylvester, D., and Krassner, S.M. 1976. Proline metabolism in *Trypanosoma cruzi* epimastigotes. *Comp Biochem Physiol B* 55: 443-447.
- Szabados, L., and Savoure, A. 2009. Proline: a multifunctional amino acid. *Trends in plant science* 15; 89-97.
- Szira, F., Balint, A.F., Borner, A., and Galiba, G. 2008. Evaluation of drought-related traits and screening methods at different developmental stages in Spring barley. *J Agro Crop Sci* 195 (5): 334-342.
- Tadege, M., and Kuhlemeier, C. 1997. Aerobic fermentation during tobacco pollen development. *Plant Mol Biol* 35: 343-354.
- Tadege, M., Bucher, M., Staehli, W., Suter, M., Dupuis, I., and Kuhlemeier, C. 1998. Activation of plant defense responses and sugar efflux by expression of pyruvate decarboxylase in potato leaves. *Plant J* 16: 661-671.
- Tadege, M., Dupuis, I., and Kuhlemeier, C. 1999. Ethanolic fermentation: new functions for an old pathway. *Trends Plant Sci* 4: 320-325.
- Tanksley, S.D., and Jones, R.A. 1981. Effects of O<sub>2</sub> stress on tomato alcohol dehydrogenase activity: description of a second ADH coding gene. *Biochem Genet* 19: 397-409
- Tatematsu, K., Kumagai, S., Muto, H., Sato, A., Watahiki, M.K., Harper, R.M., Liscum, E., and Yamamoto, K.T. 2004. *MASSUGU2* encodes Aux/IAA19, an auxin-regulated protein that functions together with the transcriptional activator NPH4/ARF7 to regulate differential growth responses of hypocotyl and formation of lateral roots in *Arabidopsis thaliana*. *Plant Cell* 16:379-393
- Tesniere, C., and Verries, C. 2000. Molecular cloning and expression of cDNAs encoding alcohol dehydrogenases from *Vitis vinifera* L. during berry development. *Plant Sci* 157: 77-88.
- Tesniere, C., Torregrosa, L., Pradal, M., Souquet, J. M, Gilles, K., Dos Santos., Chatelet, P., and Gunata, Z. 2006. Effect of genetic manipulation of alcohol dehydrogenase levels on the response to stress and synthesis of secondary metabolites in grapevine leaves. *J Exp Bot* 57: 91-99.



- Tezara, W., Mitchell, V.J, Driscoll, S.D., and Lawlor, D.W. 1999. Water stress inhibits plant photosynthesis by decreasing coupling factor and ATP. *Nature* 401: 914–917.
- The *Arabidopsis* Genome Initiative. 2000. Analysis of the genome sequence of the flowering plants *Arabidopsis thaliana*. *Nature* 408: 796-815.
- Theodore Chase, J.R. 1999. Alcohol Dehydrogenases: Identification and Names for Gene Families. *Plant Mol Biol Reporter* 17: 333-350.
- Todd, G.W. 1972. Water deficits and enzymatic activity. In: Kozlowski TT, eds. Water deficit and plant growth. New York: *Acad Press* 177-216.
- Uemura, M., and Yoshida, S. 1986. Studies on freezing injury in plant cells. II. Protein and lipid changes in the plasma membranes of *Jerusalem artichoke* tubers during a lethal freezing in vivo. *Plant Physiol* 80: 187-95.
- Umezawa, T., Fujita, M., Fujita, Y., Yamaguchi-Shinozaki, K., and Shinozaki, K. 2006. Engineering drought tolerance in plants: discovering and tailoring genes unlock the future. *Curr Opin Biotechnol* 17:113-122.
- Van der weele, C.M., Spollen, W.G., Sharp, R.E., and Baskin, T.I. 2000. Growth of *Arabidopsis thaliana* seedlings under water deficit studied by control of water potential in nutrient-agar media. *J Exp Bot* 51(350): 1555-1562.
- Vanrensburg, L., Kruger, G. H. J., and Kruger, R. H.1993. Proline accumulation as drought tolerance selection criterion: Its relationship to membrane integrity and chloroplast ultra structure in *Nicotiana tabacum* L. *J Plant Physiol* 141: 188-194.
- Van Waarde, A. 1991. Alcoholic fermentation in multicellular organisms. *Physiol Zool* 64: 895-920.
- Verslues, P.E., Ober, E.S., and Sharp, R.E. 1998. Root growth and oxygen relations at low water potentials. Impact of oxygen availability in polyethylene glycol solutions. *Plant Physiol* 116: 1403-1412.
- Verslues, P.E., Agarwal, M., Katiyar-Agarwal, S., Zhu, J., and Zhu, J.K. 2006. Methods and concepts in quantifying resistance to drought, salt and freezing, abiotic stresses that affect plant water status. *Plant J.* 45: 523-539.
- Vicre, M., Farrant, J.M., and Driouich, A. 2004. Insights into the cellular mechanisms of desiccation tolerance among angiosperm resurrection plant species. *Plant Cell Environ* 27: 1329 -1340.
- von Schaewen, A., Stitt, M., Schmidt, R., Sonnewald, U., and Willmitzer, L. 1990. expression of a yeast-derived invertase in the cell wall of tobacco and *Arabidopsis* plants leads to accumulation of carbohydrate and inhibition of

photosynthesis and strongly influences growth and phenotype of transgenic tobacco plants. *EMBO J* 9: 3033-3034.

- Weatherley, P.E. 1950. Studies in water relations of cotton plants. The field measurement of water deficit in leaves. *New Phytol* 49: 81-7.
- Wei, Y., Lin, M., Oliver, D.J., and Schnable, P.S. 2009. The roles of aldehyde dehydrogenases (ALDHs) in the PDH bypass of *Arabidopsis*. *BMC Biochem* 10: 7.
- Weigel, D. D., and Glazebrook, J. 2002. *Arabidopsis: A Laboratory Manual*. Cold Spring Harbor Laboratory Press.
- Wilkinson, S., and Davies, W.J. 1997. Xylem Sap pH Increase: A drought signal received at the Apoplastic face of the guard cell that involves the suppression of saturable abscisic acid uptake by the epidermal symplast. *Plant Physiol* 113(2):559-573.
- Wilkinson, S., and Davies, W.J. 2002. ABA-based chemical signaling: the co-ordination of responses to stress in plants. *Plant Cell Environ* 25: 195-210.
- Wilkinson, S., and Davies, W.J. 2010. Drought, ozone, ABA and ethylene: new insights from cell to plant to community. *Plant Cell Environ* 33: 510-525.
- Wignarajgh, K., Greenway, H., and John, C.D. 2006. Effect of waterlogging on growth and activity of alcohol dehydrogenase in barley and rice. *New Phytol* 77 (3); 585-592.
- Wise, R.R., Ortiz-Lopez, A., and Ort, D.R. 1992. Spatial distribution of photosynthesis during drought in field-grown and chamber grown acclimated and nonacclimated cotton. *Plant Physiol* 100:26-36.
- Wu, S.J., Lee, D., and Zhu, J.K. 1996. SOS1, a genetic locus essential for salt tolerance and potassium acquisition, *Plant Cell* 8: 617-627.
- Wu, Y., and Cosgrove, D.J. 2000. Adaptation of roots to low water potentials by changes in cell wall extensibility and cell wall proteins. *J Exp Bot* 51:1543-1553.
- Wu, Y.J., Sharp, R.E., Durachko, D.M., and Cosgrove, D.J. 1996. Growth maintenance of the maize primary root at low water potentials involves increases in cell-wall extension properties, expansin activity, and wall susceptibility to expansins. *Plant Physiol* 111: 765-772.
- Xian-He, J., Wang, J., and Guo Liang, H. 1995. Effects of water stress on photochemical function and protein metabolism of photosystem II in wheat leaves. *Physiol Plant* 93: 771-777.

- Xu, R., and Li, Q.Q. 2008. Protocol: StreamLine cloning of genes into binary vectors in *Agrobacterium* via the Gateway<sup>®</sup> TOPO vector system. *Plant methods*.4: 4.
- Yamaguchi-Shinozaki, K., and Shinozaki, K. 2005. Organization of cisacting regulatory elements in osmotic- and cold-stress-responsive promoters. *Trends in Plant Science* 10: 88-94.
- Yancey, P.H., Clark, M.E., Hand, S.C., Bowlus, R. D., and Somero, G.C. 1982. Living with water stress: Evolution of osmolyte systems. *Science* 217: 1214-1222.
- Yokota, A., Takahara, K., and Akashi, K. 2006. Water stress. In: Madhava Rao KV, Raghavendra AS, Janardhan Reddy K (Eds) *Physiology and Molecular Biology of Stress Tolerance in Plants*. Springer, The Netherlands, pp 15-39.
- Yordanov, I., Velikova, V., and Tsonev, T. 2003. Plant responses to drought and stress tolerance. *Bulg J Plant Physiol Special issue* 187-206.
- Yoshida, S. 1994. Low temperature-induced cytoplasmic acidosis in cultured mung bean (*Vigna radiate* (L.) Wilczek) cells. *Plant Physiol* 104: 1131-8.
- Zabalza, A., van Dongen, J.T., Froehlich, A., Oliver, S.N., Faix, B., Gupta, K.J., Schmäzlin, E., Igal, M., Orcaray, L., and Peter. 2009. Regulation of Respiration and Fermentation to Control the Plant Internal Oxygen Concentration. *Plant Physiol* 149: (2) 1087-1098.
- Zhang, J.Z., Creelman, R.A., and Zhu, J.K. 2004. From laboratory to field. Using information from *Arabidopsis* to engineer salt, cold, and drought tolerance in crops. *Plant Physiol* 135: 615-621.
- Zhang, X., Henriques, R., Lin, S-S., Niu, Q-W., and Chua, N.H. 2006. *Agrobacterium*-mediated transformation of *Arabidopsis thaliana* using the floral dip method. *Nature Proto* 1: (2) 87-103.
- Zhang, B., Shen, J.Y., Wei, W.W., Xi, W.P., Xu, C. J., Ferguson, I., and Chen, K. 2010. Expression of genes associated with aroma formation derived from the fatty acid pathway during peach fruit ripening. *J Agric Food Chem* 58: 6157-6165.