

UNIVERSITI PUTRA MALAYSIA

ENHANCED ANTINOCICEPTIVE EFFECTS OF MITRAGYNINE IN COMBINATION WITH MORPHINE VIA OPIOID RECEPTORS ACTIVATION

SHAMIMA BINTI ABDUL RAHMAN

FPSK(p) 2014 14



ENHANCED ANTINOCICEPTIVE EFFECTS OF MITRAGYNINE IN COMBINATION WITH MORPHINE VIA OPIOID RECEPTORS ACTIVATION



By

SHAMIMA BINTI ABDUL RAHMAN

 \mathbb{C}

Thesis submitted to the School of Graduate Studies, Universiti Putra Malaysia in Fulfillment of the requirements for the degree of Doctor of Philosophy

June 2014

This thesis is specially dedicated to:

My husband: Abang, Yudi Kurniawan Budi for his patience and stay by my side through all the day

My children: Muhammad, Ibrahim, Maryam, Adam and Zulaikha for their love and understanding of ummi's doing

My parents:

Mama, Bedah Musooh and Abah, Abdul Rahman Shamsuddin for being with me throughout the up's and down's, through happiness and sorrow...for all the du'a for my success and easinest of my way Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Doctor of Philosophy

ENHANCED ANTINOCICEPTIVE EFFECTS OF MITRAGYNINE IN COMBINATION WITH MORPHINE VIA OPIOID RECEPTORS **ACTIVATION**

By

SHAMIMA ABDUL RAHMAN

June 2014

Chairman Faculty

:

:

Assoc Prof Datin Sharida Fakurazi, PhD **Medicine and Health Sciences**

The management of chronic pain is one of the greatest challenges in modern medicine. Opiates such as morphine have been used to treat pain for centuries. However, the long term use of morphine is limited due to its side-effects. To date, a number of natural compounds have been detected to possess analgesic effects. One of these natural compound is mitragynine (MG) which is isolated from Mitragyna speciosa Korth. Mitragyna speciosa is popularly known as 'ketum' in Malaysia and 'kratom' in Thailand. Over 25 alkaloids are found in *Mitragyna speciosa*, MG being a major one. In this study, we investigated the action of MG as antinociceptive agent and the receptor selectivity effect. The nociceptive effect was estimated in a hot plate test (Ugo Basile model 7280; 50.0 °C). The latency time was estimated until the mice showed pain responses such as shaking, licking or jumping and the duration of latency was measured for every 15 minutes until 120 minutes. Male ICR mice (n=8/group) were administered intraperitoneally with single dosage of MG (3, 10, 15, 30, and 35 mg/kg), 15 minutes prior to pain induction. The control groups were given appropriate dose of vehicle. For the receptor selectivity test, the treated groups were administered naloxone (non-selective opioid antagonist), naltrindole (δ-opioid antagonist), norbinaltorpimine (κ -opioid antagonist) and AM251 (cannabinoid 1 antagonist) respectively prior to MG injection at the dosage of 35 mg/kg. The groups administered with MG showed an increased in latency time as compared to the control groups in a dose-dependent manner. Meanwhile, 35 mg/kg of MG was found to significantly increase the latency time. The results also showed that naloxone and naltrindole fully blocked the antinociceptive effect of MG, whilst norbinaltropimine partially blocked the effect, but the antinociceptive effect of MG was not antagonized by AM251. These results demonstrated that MG acts through opioid receptor specifically on δ and κ receptor and not through the cannabinoid CB1 receptor. Later on, we investigated the enhancement of analgesic action of this compound when combined with morphine and the effect on the development of tolerance due to morphine acutely and chronically. Male ICR mice (n=7/group) were administered intraperitoneally with a single dose of MG either 15 mg/kg or 25 mg/kg combined with morphine (5 mg/kg) in the acute study, whilst the study was continued for 9 days for the chronic phase. The control groups were given the appropriate dose of a vehicle. The antinociceptive effect was estimated with a hot



plate test (Ugo Basile model 7280; 50.0 °C). The latency time was assessed until the mice showed a pain response such as shaking, licking or jumping. The expression of cAMP, cAMP response element binding (CREB) protein, ERK and c-fos were analyzed. Liver and kidney function test were also analyzed and compared between groups. In acute study, the administration of MG and morphine showed a significant latency period compared to the vehicle treated groups. The combination of MG and morphine has enhanced morphine-induced analgesia which shows synergism in analgesic action. In the chronic phase, the concurrent administration of MG and morphine showed a significant increase in the latency time when compared to morphine alone groups and the remarkable analgesic effects in the combination regimens were maintained from day 1 until day 9. The result was in contrast when compared to morphine alone groups, where the latency time were reduced from day 5 to day 9. For the protein expressions, there were a significant increment of the cAMP and CREB levels (p<0.001) in groups treated with 5 mg/kg morphine but there was no significant changes of cAMP and CREB expression for MG alone groups and groups combined with morphine. There were no significant changes in other proteins (ERK and c-fos) for all groups when compared with the control group. There was also no significant changes in the liver enzymes of the treated groups when compared to the control group except for the AST level. There were no significant changes in the excretion level of urea in all groups when compared to the control groups. Similar results were found for the excretion of creatinine. However, the creatinine excretion was significantly increased when the treatment was combined. This study indicates that MG has antinociceptive properties and act fully via the opioid system. It also indicates that concurrent administration of morphine and MG enhanced the analgesic effects. Following the inclusion of MG, tolerance due to repeated administration of morphine is delayed.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

PENINGKATAN KESAN ANTINOSISEPTIF GABUNGAN MITRAGYNINE DAN MORFIN MELALUI PENGAKTIFAN RESEPTOR OPIAT

Oleh

SHAMIMA ABDUL RAHMAN

Jun 2014

Pengerusi:Prof Madya Datin Sharida Fakurazi, PhDFakulti:Perubatan dan Sains Kesihatan

Pengurusan sakit kronik adalah salah satu cabaran terbesar dalam perubatan moden. Opiat seperti morfin telah lama digunakan dalam pengurusan sakit kronik. Walau bagaimanapun, penggunaan jangka panjang morfin adalah terhad disebabkan oleh kesan sampingan. Setakat ini, beberapa sebatian semulajadi telah dikenalpasti sebagai memiliki kesan analgesik. Salah satu sebatian semula jadi adalah mitragynine (MG) yang diekstrak daripada *Mitragyna speciosa* Korth. *Mitragyna* speciosa lebih dikenali sebagai ketum di Malaysia dan Kratom di Thailand. Lebih 25 alkaloid terdapat dalam *Mitragyna* speciosa dan MG merupakan komponen utama. Dalam kajian ini, kami mengkaji kesan MG sebagai ejen antinosiseptif dan kesan pemilihan reseptor. Kesan nosiseptif dikenalpasti menggunakan ujian plat panas (Ugo Basile model 7280; 50.0 °C). Masa laten dianggarkan sehingga tikus menunjukkan tindak balas kesakitan seperti menggigit, menjilat atau melompat dan tempoh laten diukur bagi setiap 15 minit sehingga 120 minit. Tikus jantan ICR (n = 8/group) telah disuntik secara intraperitoneal dengan dos tunggal MG (3, 10, 15, 30, dan 35 mg / kg), 15 minit sebelum induksi kesakitan. Kumpulan kawalan diberi dos kenderaan yang sesuai. Bagi ujian pemilihan reseptor, kumpulan yang dirawat telah diberikan naloxone (antagonis opioid tidak terpilih), naltrindole (antagonis δ opioid), norbinaltorpimine (κ - opioid antagonis) dan AM251 (cannabinoid 1 antagonis) masing-masing sebelum suntikan MG pada dos 35mg/kg. Kumpulan yang diberi MG menunjukkan peningkatan dalam masa laten berbanding dengan kumpulan kawalan dan kesannya adalah bergantung kepada dos. Sementara itu, 35 mg/kg MG didapati meningkatkan masa latennya secara signifikan. Keputusan juga menunjukkan bahawa naloxone dan naltrindole menyekat sepenuhnya kesan antinosiseptif MG, manakala norbinaltropimine menyekat sebahagian kesannya, tetapi kesan antinosiseptif MG tidak dihalang oleh AM251. Keputusan ini menunjukkan bahawa MG bertindak melalui reseptor opioid khusus pada δ dan κ reseptor dan bukan melalui reseptor cannabinoid CB1. Kemudian, kami menyiasat peningkatan tindakan analgesik sebatian ini apabila digabungkan dengan morfin dan kesan ke atas ketahanan terhadap morfin secara akut dan kronik. Tikus jantan ICR (n = 7/group) telah disuntik secara intraperitoneal dengan dos tunggal MG sama ada 15mg/kg atau 25 mg/kg digabungkan dengan morfin (5mg/kg) dalam kajian akut, manakala kajian diteruskan selama 9 hari untuk fasa kronik. Kumpulan kawalan diberi dos kenderaan yang sesuai. Kesan antinosiseptif dikenalpasti dengan ujian plat panas (Ugo Basile model 7280; 50.0 °C). Masa laten dikira sehingga tikus

v

menunjukkan tindak balas kesakitan seperti menggigit, menjilat atau melompat. Protein cAMP, unsur tindak balas CAMP mengikat (CREB) protein, ERK dan c-fos telah dianalisis. Ujian fungsi hati dan buah pinggang juga ditentukan dan dibandingkan diantara semua kumpulan. Dalam kajian akut, penggunaan MG dan morfin menunjukkan tempoh laten yang ketara berbanding dengan kumpulan kenderaan dirawat. Gabungan MG dan morfin telah meningkatkan kesan analgesik morfin yang menunjukkan sinergi dalam tindakan analgesik. Dalam fasa kronik, suntikan serentak MG dan morfin menunjukkan peningkatan yang ketara pada masa laten berbanding kumpulan morfin sahaja dan kesan analgesik yang luar biasa dalam rejimen gabungan dikekalkan dari hari 1 hingga hari 9. Hasilnya adalah berbeza apabila dibandingkan dengan kumpulan morfin sahaja, di mana masa laten dikurangkan dari hari ke 5 ke hari ke 9. Bagi analisis protein, terdapat kenaikan yang ketara daripada cAMP dan tahap CREB (p < 0.001) dalam kumpulan yang dirawat dengan 5 mg/kg morfin tetapi tidak terdapat sebarang perubahan signifikan terhadap cAMP dan CREB protein untuk kumpulan MG sahaja dan kumpulan digabungkan dengan morfin. Tidak ada perubahan ketara dalam protein lain (ERK dan c-fos) untuk semua kumpulan berbanding dengan kumpulan kawalan. Tiada sebarang perubahan penting dalam enzim hati kumpulan yang dirawat berbanding dengan kumpulan kawalan kecuali bacaan AST. Tiada perubahan ketara dalam tahap perkumuhan urea dalam semua kumpulan apabila dibandingkan dengan kumpulan kawalan. Keputusan yang sama diperolehi untuk perkumuhan kreatinin. Walau bagaimanapun, perkumuhan kreatinin telah meningkat dengan ketara apabila rawatan digabungkan. Kajian ini menunjukkan bahawa MG mempunyai ciri-ciri antinosiseptif dan bertindak sepenuhnya melalui sistem opioid. Ia juga menunjukkan bahawa suntikan serentak morfin dan MG meningkatkan kesan analgesik. Kombinasi MG bersama morfin melewatkan toleransi terhadap penggunaan morfin yang berlanjutan.



In the name of Allah the most Gracious and the most Merciful, peace upon Muhammad SAW the last prophet. Alhamdulillah, with Allah's will, I have finally completed my PhD study and thesis writing.

I would like to thank those who have supported me and contributed in the completion of my thesis. Firstly, I would like to express my deep appreciation to my main supervisor, Assoc Prof Dr Sharida Fakurazi for her guidance, patience, support and advice throughout my study. This was priceless, without her, this project will not be completed and successful. Special thanks are dedicated to all my co-supervisors Prof Dr Hairuszah Ithnin, Dr Mohamad Aris Mohd Moklas and Dr Mohamad Taufik Hidayat Baharuldin. Their suggestion, advice and outstanding attention have contributed much to the success of this project.

I also would like to convey my thanks to the laboratory staff of Department of Human Anatomy of Faculty of Medicine and Health Sciences, UPM especially to Puan Noridah Md Top, Encik Shahidan Sulaiman, Puan Farhatani Mahmud and others for their help and guidance throughout my research. I would like to acknowledge the support of my labmates and friends, Fatin Nadzirah, Muhammad Khairulasyraf Muhammad Yusuf, Syazana Akmal Sharifudin, Nurul Raudzah Adib Ridzuan, Farah Nasir and Noor Azuin Suliman who had always given their hand to help me. Also to my friends that encourage me in submitting my thesis, Semira Abdi Beshir and Waheedah Abdul Hakeem.

I also would like to thank the staff of Immunology Laboratory; Puan Norazren Ismail and Encik Zulkhairi Zainol and Pathology Laboratory, Department of Biomedical Sciences; Puan Normah Ibrahim and Puan Juita Chupri, for helping me and allowing me to use their equipments and services.

Last but not least, my deepest appreciation to my beloved husband, Yudi Kurniawan Budi and my children, Muhammad, Ibrahim, Maryam, Adam and Zulaikha for their support, understanding and unconditional love. Their love and support become my inspiration, and I am so blessed to have such a caring and supporting family.

Finally to my family that has been the source of my strength, always giving me encouragement and support to finished my study especially my mother, Bedah Musooh, my father, Abdul Rahman Shamsudin and my siblings. Thank you very much for being there for me and making my life full of love and care.

May Allah bless you all...ameen.

I certify that an Examination Committee met on 2012 to conduct the final examination of Shamima Abdul Rahman on her Doctor of Philosophy entitled 'Enhanced antinociceptive effects of mitragynine in combination with morphine via opioid receptors activation' in accordance with Universities and University College Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U. (A) 106] 15 March 1998. The committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

Sabrina Sukardi, Ph.D Associate Professor, Faculty of Medicine and Health Sciences, Universiti Putra Malaysia (Chairman)

Arifah Abdul Kadir, Ph.D Associate Professor, Faculty of Veterinary Medicine, Universiti Putra Malaysia (Member)

Hamidon Basri, Ph.D Professor, Faculty of Medicine and Health Sciences, Universiti Putra Malaysia (Member)

Mustafa Culha, Ph.D Professor, Faculty of Engineering and Architecture, Yeditepe University, Atasehir, Turkey (Member)

> Bujang B. K. Huat Professor and Dean, School of Graduate Studies Universiti Putra Malaysia

Date :

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree. The members of the Supervisory Committee are as follows:

Sharida Fakurazi, Ph.D

Associate Professor Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Chairperson)

Hairuszah Ithnin, MD, MPATH, FAMM

Professor Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Member)

Mohamad Aris Mohd Moklas, Ph.D

Associate Professor Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Member)

Mohamad Taufik Hidayat Baharuldin, Ph.D

Associate Professor Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean, School of Graduate Studies Universiti Putra Malaysia

Date :

DECLARATION

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/ fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature : ____

_ Date : _

Name and Matric No : Shamima Abdul Rahman (GS 19562)

DECLARATION

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Sharida Fakurazi,

Ph.D Associate Professor Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Chairperson)

Hairuszah Ithnin, MD, MPATH, FAMM Professor Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Member)

Mohamad Aris Mohd Moklas, Ph.D

Associate Professor Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Member)

Mohamad Taufik Hidayat Baharuldin, Ph.D

Associate Professor Faculty of Medicine and Health Sciences Universiti Putra Malaysia (Member)

TABLE OF CONTENT

	Page
DEDICATION	ii
ABSTRACT	iii
ABSTRAK	v
ACKNOWLEDGEMENTS	vii
APPROVAL	viii
DECLARATION	X
DECLARATION OF SUPERVISORY COMMITTEE	xi
LIST OF TABLES	xvi
LIST OF FIGURES	xvii
LIST OF ABBREVIATIONS	XX

CHAPTER

1

2

INTRO	DUCTION				
1.1		nd of Study	1		
1.2					
1.3	Significan	ce of Study	2 3		
1.4	0	al framework of the study	5		
1.5	Study hyp		6		
1.6	Objectives	s of the Study	7		
	1.6.1	General objectives	7		
	1.6.2	Specific objectives	7		
LITER	ATURE RI	FVIFW			
2.1	Introductio		8		
2.1	2.1.1	Definition of pain	8		
	2.1.2	Classification of pain	8		
2.2	Pain pathw	-	9		
2.2	2.2.1	Activation of nociceptors	9		
	2.2.2	Transmission of pain information	12		
	2.2.3	Descending pathways for pain modulation	13		
2.3	Manageme		14		
2.4	Opioid Systems 10				
	2.4.1	Opioid receptors	16		
	2.4.2	Endogenous opioids	17		
	2.4.3	Opioid analgesic drugs	18		
	2.4.4	Opioid antagonist	19		
2.5	Cannabino	oid Systems	20		
	2.5.1	Cannabinoid receptors	20		
	2.5.2	Endogenous cannabinoids	20		
	2.5.3	Cannabinoid antagonist	21		
	2.5.4	Role of cannabinoid system in pain	21		
2.6	Morphine		21		
	2.6.1	Mechanism of action of morphine	22		
2.7	Tolerance	-	24		
	2.7.1	Early adaptation in tolerance	26		
	2.7.2	Long term adaptation in tolerance	27		

			e expression	of opioid tolerance	27
		2.8.1	cAMP		29
		2.8.2	CREB pro	tein	30
		2.8.3	ERK1/2		31
		2.8.4	c-fos gene		32
	2.9	Mitragyna	speciosa K	Lorth	33
		2.9.1	Botanical	origin	34
		2.9.2	Mitragynii	ne and other analogues	34
		2.9.3	Medical us	ses of Mitragyna speciosa	35
	2.10	Combinati	ion treatment	nt	37
	2.11	Toxicity			37
		2.11.1	Liver		38
			2.11.1.1	Liver Function Tests	39
		2.11.2	Kidney		40
			2.11.2.1	Kidney Function Tests	41
				NINE FROM MITRAGYNA	
		SA KOR			
		Introductio			42
			and method	s	43
		3.2.1	Plants		43
		3.2.2		and reagents	44
		3.2.3	Equipmen		44
				ne from Mitragyna speciosa	44
		3.3.1		hanolic extraction	44
		3.3.2	Alkaloid e		44
		3.3.3		purification and identification of	45
	3.4	Results	minagymi	e compound	47
•		3.4.1	Crude met	hanolic extract and crude alkaloids	47
		3.4.1	extract	natione extract and crude alkaloids	4/
		3.4.2		purification and identification of	47
		3.4.2	mitragynir		4/
	3.5	Discussion			53
	5.5	Discussion	•		55
4	ANTIN	OCICEPT	IVE ACTI	VITY OF MITRAGYNINE AND	
	RECEP	TOR SEL	ECTIVITY	Y STUDY	
4	4.1	Introductio	on		55
4	4.2	Materials a	and method	S	57
		4.2.1	Animals		57
		4.2.2	Drugs		57
		4.2.3	Mitragyni	ne compound	57
		4.2.4	Equipmen	ts and materials	57
		4.2.5	Chemical	reagents	58
		4.2.6	Hot plate t	est	58
		4.2.7	-	ptive study and the determination of	58
			ED ₅₀	-	
		4.2.8	The deter	mination on the effect of MG	59
			following	the administration of opioid	
			antagonist	S	

xiii

		4.2.9	The determination on the effect of MG	60
			following the administration of cannabinoid	
		4 2 10	antagonists Statistical employie	(1
	1 2	4.2.10	Statistical analysis	61
	4.3	Results	Antino gigantive study and the determination of	61
		4.3.1	Antinociceptive study and the determination of ED50	61
		4.3.2	The effect of mitragynine following the	65
		4.3.2	administration of opioid antagonist	05
			4.3.2.1 The effect of MG in the presence of	65
			naloxone antagonist	0.5
			4.3.2.2 The effect of MG in the presence of	66
			naltrindole antagonist	00
			4.3.2.3 The effect of MG in the presence of	67
			naloxonazine antagonist	07
			4.3.2.4 The effect of MG in the presence of	68
			norBNI antagonist	00
		4.3.3	The effect of mitragynine following the	69
		11010	administration of cannabinoid CB_1 receptor	0,
			antagonist	
	4.4	Discussio		71
5	ANTIN	IOCICEP	IVE EFFECTS ON COMBINATION	
	TREA '	rment c	PF MITRAGYNINE AND MORPHINE IN	
	HOT F	PLATE TE	ST	
	5.1	Introducti	on	77
	5.2	Materials	and Methods	80
		5.2.1	Plant	80
		5.2.2	Isolation of mitragynine from Mitragyna	80
			speciosa	
		5.2.3	Animals	80
		5.2.4	Drugs	81
		5.2.5	HPT	81
		5.2.6	Experimental design	81
			5.2.6.1 Acute Study	81
		507	5.2.6.2 Chronic Study	82
	5.2	5.2.7	Statistical analysis	83
	5.3	Results	A sector Oter las	05
		5.3.1	Acute Study	85
	5.4	5.3.2 Discussio	Chronic Study	86 88
	5.4	Discussio		00
6	EFFE(CTS OF	COMBINATION TREATMENT OF	
U		AGYNINE	AND MORPHINE ON PROTEIN	
			ND TOXICOLOGY PROFILE	
	6.1	Introducti		91
	6.2		and methods	91
	0.2	6.2.1	Chemicals	95
		6.2.2	Antibody	95 95
		6.2.3	Equipments	95
			1 f ····	

		6.2.4	Experimen	0	96
		6.2.5	Brain samp		96
			6.2.5.1	Determination of protein	96
				concentration of whole brain	
			6.2.5.2	cAMP measurement from cortex	97
				and thalamus	
			6.2.5.3	Expression of CREB, ERK and c- fos	98
		6.2.6	Serum ana	lysis for LFT and KFT	103
			6.2.6.1	Liver function test	103
		6.2.7	Histopatho	logical examination	104
		6.2.8	Scoring	-	105
	6.3	Results	-		
		6.3.1	cAMP		107
		6.3.2	CREB		109
		6.3.3	ERK1/2		110
		6.3.4	c-fos		111
		6.3.5	The effect	of mitragynine and morphine on	111
			liver weigh	nt the second	
		6.3.6	The effect	of mitragynine and morphine on	112
			kidney wei		
		6.3.7	The effect	of mitragynine and morphine on	113
			liver enzyr		
		6.3.8		of mitragynine and morphine on	117
			kidney enz		
		6.3.9	U	al result of liver	119
		6.3.10	U	al result of kidney	124
	6.4	Discussion	1		129
7	GENER	RAL DISCU	USSION A	ND CONCLUSION	133
REFERI	ENCES				136
APPENDICES				154	
BIODAT	'A OF S'	TUDENT			160

6

LIST OF TABLES

Table		Page
2.1	Stimuli that activate nociceptors	10
2.2	Characteristic of primary afferent fibers	10
2.3	Important chemicals involved in nociception	11
2.4	Principle Endogenous Opioid peptides	18
2.5	Main adverse effects of opioid therapy in various systems	19
2.6	The characteristic of MG	35
3.1	Type of extracts, appearance and their weight	47
3.2	The weight of MG isolated samples, Rf value and TLC plate appearance	48
4.1	Illustration of each group and its respective treatment	59
4.2	Illustration of each group and its respective treatment with opioid antagonist	60
4.3	Illustration of each group and its respective treatment with cannabinoid antagonist	61
5.1	Examples of combination analgesics and their advantages	79
5.2	Illustrations of groups in acute study	82
5.3	Illustrations of groups in chronic study	83
6.1	Stacking and resolving gel for the CREB, ERK 1/2 and c-fos	98
6.2	Antibody concentration for the CREB, ERK 1/2 and c-fos	100
6.3	The scoring liver in histopathological study	105
6.4	The scoring kidney in histopathological study	105
6.5	Mean \pm SEM of liver for chronic combination study	119
6.6	Mean \pm SEM of kidney for chronic combination study	124

 \bigcirc

LIST OF FIGURES

Figure		Page
1.1	Market shares of major drug classes in pain market of US	3
1.2	Conceptual framework of the study	6
2.1	Summary of pain pathway and chemicals involved	14
2.2	The WHO analgesic ladder provides a guide to initiating for pain of different intensities	15
2.3	The chemical structure of morphine	22
2.4	Summary of mechanism of action of opiate (morphine) and NSAIDs as pain relief	24
2.5	Shift in a dose-response curve with tolerance	25
2.6	Gene expression in pain pathway	29
2.7	Mature leaves of Mitragyna speciosa	33
2.8	Compound found in MS and their percentage	34
3.1	Schematic diagram of indole alkaloid with nitrogen molecule	42
3.2	Flow chart of isolation process of MG from <i>Mitragyna speciosa</i>	46
3.3	H-NMR chromatogram of the isolated MG from <i>Mitragyna</i> speciosa leaves in deuterium chloroform	50
3.4	C-NMR chromatogram of the isolated MG from <i>Mitragyna</i> speciosa leaves in deuterium chloroform	51
3.5	Chemical structure of MG	52
4.3.1a	The effects of MG, morphine and controls on latency time in hot plate test (HPT) for 120 minutes	63
4.3.1b	The determination of effective dose 50 (ED_{50}) Of MG	64
4.3.2a	Effects of µ-opioid antagonist naloxone on MG-induced antinociception	65
4.3.2b	Effects of δ -opioid antagonist naloxone on MG-induced antinociception	66
4.3.2c	Effects of μ_1 -opioid antagonist naloxone on MG-induced antinociception	67
4.3.2d	Effects of κ-opioid antagonist naloxone on MG-induced antinociception	68
4.3.3	Effects of CB ₁ antagonist AM251 on MG-induced antinociception	70
5.1	Mechanism of action of morphine as analgesic agent	78
5.2	Flow chart of acute and chronic combination treatment of morphine and MG	84
5.3	The effects of control groups, MG, morphine and controls on	85
	latency time in hot plate test (HPT) of single dosage	
5.4	The effects of control groups, MG, morphine and controls on	87
	latency time in hot plate test (HPT) for 9 days of treatments	
6.1	Relationship of cAMP and CREB proteins. The activation of intracellular cAMP which leads to the activation of CREB	92
	proteins that later on promotes the synthesis of various genes	-
6.2	Brain mouse anatomy. Thalamus and cortex part was separated	97
	from the whole brain and used for CREB protein determination	
6.3	The flow chart in Western blotting procedure	102
6.4a	The expression of cAMP protein in thalamus of controls groups	107

	(Normal saline, 2% T80), mitragynine (15 mg/kg, 25 mg/kg), morphine (5 mg/kg) and combination groups (5 mg/kg Mor + 15 mg/kg MG; 5 mg/kg Mor + 25 mg/kg MG) in 9 days of treatment.	
6.4b	The expression of cAMP protein in tcortex of controls groups (Normal saline, 2% T80), mitragynine (15 mg/kg, 25 mg/kg), morphine (5 mg/kg) and combination groups (5 mg/kg Mor + 15 mg/kg MG; 5 mg/kg Mor + 25 mg/kg MG) in 9 days of treatment.	108
6.4c	The expression of CREB protein of control groups (Normal saline, 2% T80), mitragynine (15 mg/kg, 25 mg/kg), morphine (5 mg/kg) and combination groups (5 mg/kg Mor + 15 mg/kg MG; 5 mg/kg Mor + 25 mg/kg MG) in 9 days of treatment	109
6.4d	The expression of ERK 1/2 protein of control groups (Normal saline, 2% T80), mitragynine (15 mg/kg, 25 mg/kg), morphine (5 mg/kg) and combination groups (5 mg/kg Mor + 15 mg/kg MG; 5 mg/kg Mor + 25 mg/kg MG) in 9 days of treatment	110
6.4e	The expression of c-fos protein of control groups (Normal saline, 2% T80), mitragynine (15 mg/kg, 25 mg/kg), morphine (5 mg/kg) and combination groups (5 mg/kg Mor + 15 mg/kg MG; 5 mg/kg Mor + 25 mg/kg MG) in 9 days of treatment	111
6.5a	The liver weight in controls groups (Normal saline, 2% T80), mitragynine (15 mg/kg, 25 mg/kg), morphine (5 mg/kg) and combination groups (5 mg/kg Mor + 15 mg/kg; 5 mg/kg Mor + 25 mg/kg).	112
6.5b	The kidney weight in controls groups (Normal saline, 2% T80), mitragynine (15 mg/kg, 25 mg/kg), morphine (5 mg/kg) and combination groups (5 mg/kg Mor + 15 mg/kg; 5 mg/kg Mor + 25 mg/kg).	113
6.6а	The activities of alanine aminotransferase (ALT) in controls groups (Normal saline, 2% T80), mitragynine (15 mg/kg, 25 mg/kg), morphine (5 mg/kg) and combination groups (5 mg/kg Mor + 15 mg/kg; 5 mg/kg Mor + 25 mg/kg).	114
6.6b	The activities of aspartate aminotransferase (AST) in controls groups (Normal saline, 2% T80), mitragynine (15 mg/kg, 25 mg/kg), morphine (5 mg/kg) and combination groups (5 mg/kg Mor + 15 mg/kg; 5 mg/kg Mor + 25 mg/kg).	115
6.6c	The activities of gamma-glutamyltransferase (GGT) in controls groups (Normal saline, 2% T80), mitragynine (15 mg/kg, 25 mg/kg), morphine (5 mg/kg) and combination groups (5 mg/kg Mor + 15 mg/kg; 5 mg/kg Mor + 25 mg/kg).	116
6.7a	The levels of urea in control groups (Normal saline, 2% T80), mitragynine (15 mg/kg, 25 mg/kg), morphine (5 mg/kg) and combination groups (5 mg/kg Mor + 15 mg/kg; 5 mg/kg Mor + 25 mg/kg).	117
6.7b	The levels of creatinine in control groups (Normal saline, 2% T80), mitragynine (15 mg/kg, 25 mg/kg), morphine (5 mg/kg) and combination groups (5 mg/kg Mor + 15 mg/kg; 5 mg/kg Mor + 25 mg/kg).	118

- 6.8a-g The figure shows the images of liver for chronic combination 120 treatment
- 6.9a-g The figure shows the images of kidney for chronic combination 125 treatment

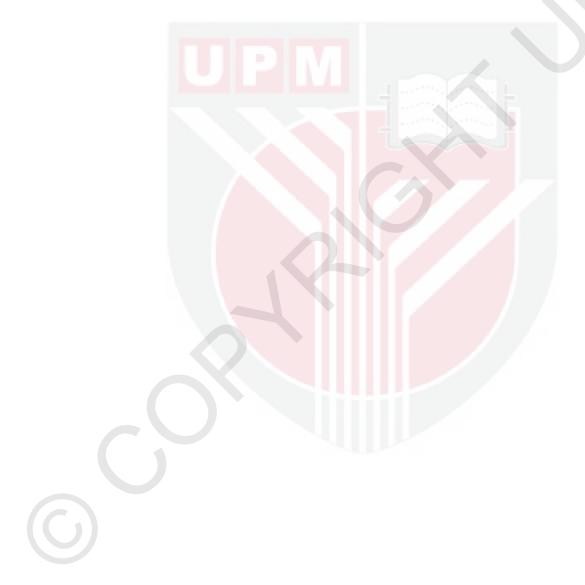


LIST OF ABBREVIATIONS

HPT MG MS min \$ h	Hot plate test mitragynine <i>Mitragyna speciosa</i> minute Dollar US hour
sec ° C	second
Rf	degree Celcius retention factor
ED ₅₀	effective dose at 50 percent
μ δ	mu delta
б К	kappa
MOR	mu opioid receptor
DOR	delta opioid receptor
KOR	kappa opioid receptor
CB_1	cannabinoid type 1 receptor
CB_2	cannabinoid type 2 receptor
norBNI	norbinaltorphimine
NS	normal saline
T80	Tween 80
AM251	1 - (2,4-diclorophenyl) - 5 - (4-iodophenyl) - 4 - methyl - N
CNIC	– 1 – piperidinyl – 1H – pyrazole 3 carboxamide
CNS	central nervous system
% MDI	percentage
MRI LFT	mean relative intensity Liver function test
KFT	Kidney function test
c-fos	gene
CREB	cAMP response element binding
PKA	cAMP-dependent protein kinase
МАРК	mitogen-activated protein kinase
ATF-1	activating transcription factor-1
cAMP	cyclic adenosine monophosphate
CREM	cAMP response element modulator
ERK	extracellular signal-regulated kinases
ALT	Alkaline phosphatase
AST	Aspartate transaminase
GGT	Gamma-glutamyltransferase
NSAIDs	non-steroidal anti-inflammatory drugs
WHO	World Health Organization
RVM	rostral ventromedial medulla
PAG	periaqueductal grey
GPCRs SEM	G-protein-coupled receptor standard error mean
SEM ELISA	enzyme-linked immunosrbant assay
BSA	bovine serum albumin
ANOVA	One-way analysis variance



i/p	intraperitoneal injection
kg	kilogram
mg	miligram
g	gram
Nal	Naloxone
NTI	Naltrindole
COX-2	cyclooxygenase 2 pathway
mL	milimeter
&	and
b.wt	body weight



CHAPTER 1

INTRODUCTION

1.1 Background of Study

Pain, both acute and chronic, remains a significant health problem despite tremendous progress in understanding its basic mechanism (Gregory et al., 2013). The International Association for the Study of Pain (IASP) defines pain as an unpleasant sensory and emotional experience associated with actual or potential tissue damage, or described in terms of such damage (American Pain Society, 2003). Relief from pain has been the paramount objectives of the medical profession throughout history (Dureja, 2010) and nowadays, the management of chronic pain is one of the greatest challenges in medicine. Pain as a whole, is a very active area for pharmaceutical research and development, not only as the cause of frequent mistreatment, but also of unacceptable side effects associated with older and still widely used compounds (Bruehl, 2013).

Plants or plant parts have been used as a source of medicine since prehistoric times (Smith-Hall, 2012). Until now, plants are an important source of chemical compounds that are developed into drugs. Between 1983 and 1994, of the 520 new prescription drugs approved, 39% were derived from plants or animals or natural sources, with 60% to 80% of those comprised of antimicrobials and anticancer drugs (Wecker, 2010). Drugs from plants continue to be a great source of revenue in the United States, with the annual sales of \$10 billion in the year 2000. More than 200 organizations worldwide are investigating new uses of plant-derived drugs, especially in the fight against AIDS, cancer, diabetes and cardiovascular diseases (Khan, 2011). Nowadays, drugs are also processed using a synthetic version of the active chemical found in the plant. Besides all these, plants have become the main component of the ever-growing alternative therapy development (Khan, 2011).

The source for opium is the opium poppy plant, *Papaver somniferum*. Morphine was isolated from crude opium in 1806 by Serturner, who named the substance after the Greek god of dreams, Morpheus (Wecker, 2010). Not long after its isolation, morphine was introduced into the medical practice. Subsequent to medicinal properties of opium poppy, many new plants were introduced and studied to increase the discovery of natural plant products as antinociceptive agents (Hajhashemi et al., 2011; Chen et al., 2011).

A number of natural compounds have been detected to have analgesic effect such as Papaver somniferum, Cannabis sativa, Clematis sanitora and Plantanus orientalis

(Hajhashemi et al., 2011; Chen et al., 2011). One of the most used compound is mitragynine (MG), which has been isolated from *Mitragyna speciosa* (MS) Korth. MS is a plant that is abundantly found in Thailand and Malaysia which is popularly known as 'kratom' in Thailand and 'ketum' in Malaysia. Over 25 alkaloids have been isolated from this plant (Houghton & Said, 1991), where MG was analysed as the major constituent (66.2%) together with its other analogues, paynantheine (8.6%), speciogynine (6.6%), 7-hydroxymitragynine (2.0%) and speciociliatine (0.8%) (Takayama, 2004). MG constitutes an indole structure, with its fourth position is substituted by the methoxy group. The moleculer structure is 9-methoxy-corynantheidine ($C_{23}H_{30}N_2O_4$) with molecular weight of 398.5 (Chee et al., 2008). Studies have indicated that MG plays a role as an antinociceptive agent and acts via opioid receptors (Yamamoto et al., 1999; Takayama et al., 2002; Takayama, 2004; Matsumoto et al., 2006).

Opioids analgesic drugs such as morphine continue to be the mainstream therapy available for the management of acute and chronic pain (Bruehl, 2013). Up till now, morphine is the most important and powerful analgesic. It has long and widely been used to alleviate various types of severe pain, including acute postoperative and chronic cancer pain.

1.2 Problem Statement

No single analgesic is perfect and no single analgesic can treat all types of pain. Each agent has distinct advantages and disadvantages compared to others. A combination is most effective when the individual agents act through different analgesic mechanisms and act synergistically. Combination analgesic can provide more effective pain relief for a broader spectrum of pain, and might also reduce adverse drug reactions (Raffa, 2001). Many combinations analgesic are available and are commonly prescribed for pain. Combination of acetaminophen and codeine, codeine and ibuprofen, and acetaminophen and oxycodone was found to be a safe and effective analgesic (Palangio et al., 2000).

Opiates such as morphine have been used to treat pain for centuries. However, the long term use of morphine is limited due to its side-effects, which include nausea, vomiting, being in a state of euphoria and mental detachment (Macadante *et al*, 2006). Among other side-effects of morphine, development of tolerance and dependence are the most difficult to overcome.

Active compounds such as MG have been shown to have analgesic properties (Matsumoto et al., 1996, 1998; Takayama et al., 2002; Takayama et al., 2004; Horie et al., 2005; Matsumoto et al., 2006). Many studies have been conducted and revealed that MG can give antinociceptive activity without developing toxicity effects (Macko et al.,

1972; Janchawee et al., 2007; Reanmongkol et al., 2007). Besides, MG is reported to be comparable to codeine as an analgesic (Macko et al., 1972; Jansen & Prast, 1988). Eventhough MS have been regarded as an unsafe plant under the Dangerous Drug Act 1953 if it is used repeatedly until the development of addiction by Malaysian government, perhaps the combination of potent morphine and MG will reduce the side effect of morphine (Raffa, 2001).

1.3 Significance of Study

According to the American Pain Society, prevalence of chronic pain in the United States is estimated to be 35.5% or equivalent to 105 million people (Datamonitor, 2009). This costs more than US\$100 billion per year in direct health care expenditure and the loss of work productivity time. Current pain management relies heavily on agents that have analgesic properties. Non-narcotic analgesics (acetominophen and aspirin), narcotic analgesics (opioids), non-steroidal anti-inflammatory drugs (NSAIDs), and thermal agents continue to be the mainstays of pain management. More recently, other medicines have been added, such as antidepressants, anticonvulsants and selective cyclooxygenase 2 (COX-2) inhibitors (Katzung, 2010).

The global pain market in 2009 was valued at over US\$50 billion in seven major countries namely United States (US), Japan, France, Germany, Italy, Spain and United Kingdom. In US alone, the expenditure is US\$27 billion out of the US\$50 billion. Figure 1.1 shows the US market share of major drug classes of pain agents. Strong opioid such as morphine become the biggest contributor in the shares (US\$7.83). From this chart, it can be deduced that pain killer was the most costly among major drug classes in pain market.

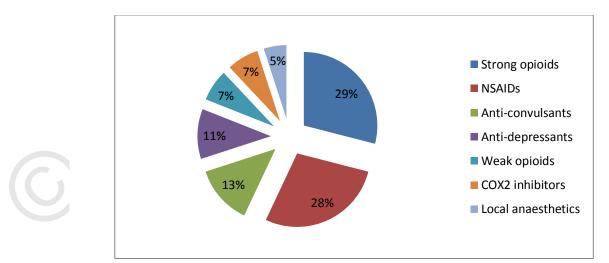


Figure 1.1: Market shares of major drug classes in pain market of US (Sources: Datamonitor, IMS Health, Decision Resources, 2009)

In Malaysia, the Malaysia Statistic of Medicine (2007) reported that strong opioids such as morphine have been used tremendously even though it is costly and induced many side effects. The total opioid consumption in Malaysia in 2007 was 0.4184 defined daily dose (DDD)/1000 population/day. Strong opioids have been widely consumed compared to weak opioid such as tramadol. In reducing the use of morphine, the medical cost in Malaysia will be much reduced as well. Thus, in finding an alternative source to pain treatment from natural products, the side effects and high cost of morphine may be reduced as well.

Previous findings have suggested that the combinations of opioid analgesics and other analgesics can be used to control pain (Lauretti *et al*, 2003; Miranda et al., 2006; Smith et al., 2007). The use of several combinations of potent opioids were suggested to reduce the toxic effects of opioid treatment, to improve analgesia and to reduce opioid tolerance (Lauretti *et al*, 2003; Morita *et al*, 2003; Mercandante *et al*., 2004). Combination of opioids with other classes of analgesics can also help to reduce sensitization processes and optimize pain therapy, as opioids such as morphine will keep their central role in postoperative, traumatic or tumor pain therapy (Wolfang, 2007). Thus, the combinations of medications that offer analgesic synergism should allow a reduction in required dosage which gives the maximum analgesic effects and a decrease in the incidence of adverse effects.

The leaf of MS has been used in Thailand and Malaysia for its opium-like effect (Burkill, 1935). It is also commonly abused due to its stimulant ability to combat fatigue (Grewal, 1932; Suwanlert, 1975). Besides, the Thailand people use the leaf to alleviate pain, coughing and diarrhea (Suwanlert, 1975). Mitragynine is the major indole alkaloid in MS (Takayama, 2004). A study by Matsumoto et al. in 2006 has found that this compound has shown some opioid activities.

Since it has been proven to have antinociceptive effects (Yamamoto et al., 1999; Takayama et al., 2002; Takayama, 2004; Matsumoto et al., 2006) MG could be a potential pain relief alternative to morphine. Thus, a combination of this compound and morphine is predicted to minimize tolerance by reducing the dosage requirement of morphine. Furthermore, the combination might have synergistic effect probably by acting at the same site of action.

Apart from that, the existence of an endogenous cannabinoid system, comprising of cannabinoid CB_1 and CB_2 receptor subtypes together with their signaling pathways and endogenous ligands, is now well recognized (Martin et al., 2004). Cannabinoids have been shown to exert a broad pharmacological action, including the central and peripheral effects through receptor-mediated mechanisms (Howlett et al., 2002). Pharmacological and molecular biological studies have identified at least two types of cannabinoid receptors, cannabinoid type 1 (CB₁) receptor and cannabinoid type 2 (CB₂)

receptor both coupled to the G-protein (Takayuki et al., 2006). The cannabinoid CB_1 receptor is predominantly found in the central nervous system. To date, cannabinoid CB_1 receptor has been shown to play a role in managing pain. However, the study on the effects of MG on opioid receptors especially CB_1 which are involved in pain management has still not been explored.

1.4 Conceptual Framework of the Study

This study consists of several phases of experiment related to one another. Figure 1.2 shows the conceptual framework of this study. Briefly, the first phase was the isolation of pure compounds which is MG from MS Korth (CHAPTER 3). The second phase was to determine the antinociceptive action of MG together with the receptor selectivity (CHAPTER 4). In this phase, opioid receptors as well as the CB₁ receptor have been selected for the determination of action. In Chapter 4, morphine was used as a positive control drug.

The third phase was to determine the effects of MG counteracting the tolerance effect of analgesic morphine. For chapter 5, the analgesic used was morphine. In **CHAPTER 5**, acute and chronic study has been conducted. In acute study, the combination regimen (MG + morphine) was given once whilst in chronic study, the combination regimen (MG + morphine) was carried out for 9 days. This was to confirm and to evaluate the development of tolerance to morphine, the analgesic of choice throughout the study.

The changes of protein expression in relation to tolerance was analysed and carried out for the groups that received 9 days of combination treatment (CHAPTER 6). Finally, in the final chapter (CHAPTER 7), the liver and kidneys were analysed for determination of toxicological changes to the metabolic and excretory organs respectively.

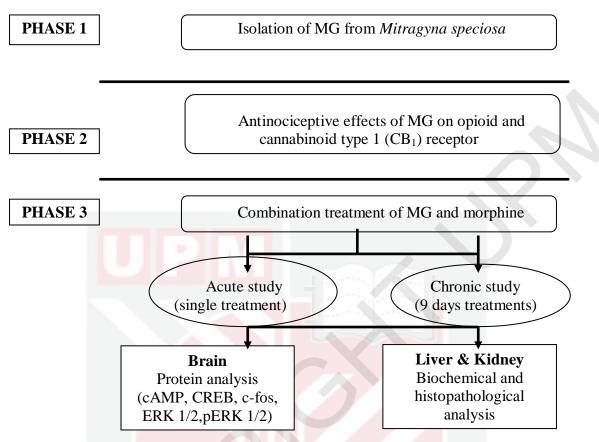


Figure 1.2: Conceptual framework of the study

1.5 Study Hypotheses

- 1. Mitragynine (MG) isolated from *Mitragyna speciosa* (MS) will have antinociceptive effect on opioids and cannabinoid receptor.
- 2. The combination treatment of morphine and MG will give synergistic antinociceptive effect.
- 3. There are no pathological changes in liver and kidney following the combination treatment.

1.6 Objectives of the Study

1.6.1 General objectives

- 1. To investigate the antinociceptive activities of MG isolated from local MS on opioid and cannabinoid receptors.
- 2. To determine the antinociceptive activity of MG and morphine as a potential combination to reduce tolerance.
- 3. To investigate any pathological changes in liver and kidney following combination treatment of MG and morphine.

1.6.2 Specific Objectives

- 1. To isolate MG from MS leaves obtained from North Peninsular Malaysia.
- 2. To investigate the effective dose at 50 percent (ED_{50}) of MG with the hot plate test.
- 3. To determine the antinociceptive effects of MG on opioids (μ,κ,δ) and cannabinoid (CB₁) receptor.
- 4. To evaluate the antinociceptive effects of MG in combination with morphine in acute and chronic study by using hot plate test.
- 5. To assess changes on cAMP, CREB, c-fos and ERK 1/2 protein expression following combination treatments.
- 6. To investigate the effect of the combination therapy on liver function test (LFT) and kidney function test (KFT).
- 7. To conduct histopathology analysis of liver and kidney following combination treatments.

REFERENCES

- A. Mestek, J.H Hurley, L.S Bye, A.D Campbell, Y. Chen, M Tian. 1995. The human mu opioid receptor: modulation of functional desensitization by calcium/ calmodulin-dependent protein kinase and protein kinase C. Journal of Neuroscience 15: 2396-2406.
- Angeles-López, G.; Pérez-Vásquez, A.; Hernández-Luis, F.; Déciga-Campos, M.; Bye, R.; Linares, E.; Mata, R. 2010. Antinociceptive effect of extracts and compounds from Hofmeisteria schaffneri. J. Ethnopharmacol.131: 425-432.
- Angst MS, Clark JD. 2006. Opioid-induced hyperalgesia: a qualitative systematic review. Anesthesiology 104(3): 570–587
- Anna DuPen MN, Danny Shen, Mary Ersek. 2007. Mechanism of opioid induced tolerance and hyperalgesia. Pain Management Nursing. Vol 8, No 3: 113-121.
- Araujo, A.J., Studzinski, M.C., Milgram, W.N. 2004. The effects of age and scopolamine on skilled motor-function in dogs : further evidence of age-dependent cholinergic dysfunction. Neurobiology of Aging. Vol.2 Supp. 2. 230
- Arden JR, Segredo V, Wang Z, Lameh J, Sadee W. 1995. Phosphorylation and agonistspecific intracellular trafficking of an epitop-tagged mu-opioid receptor expressed in HEK 293 cells. Journal of Neurochemistry. 65: 1636-1645.
- Barker, R.I.G., Bird, F., Alexander, V., Warburton, C. 2007. Recognition memory for objects, place and temporal order : a disconnection analysis of the role of the medial prefrontal cortex and perirhinal cortex. The Journal of Neuroscience. Vol. 27 (11): 2948-2957.
- Beggs, S.; Micheal, WS. 2010. A straightjacket for pain? Cell 143: 505–507.
- Bilecki W, Przewlocki R. 2000. Effect of opioids on Ca2+/cAMP responsive element binding protein. Acta Neurobiology. Exp. Wars 60 (4): 557-567.
- Bilecki, W.; Wawrzczak-Bargiela, A.; Przewlocki, R. 2004. Activation of AP-1 and CRE dependent gene expression via mu-opioid receptor. J. Neurochem. 90: 874–882.
- Berrocal, Y.A., Pearse, D.D., Andrade, C.M., Hechtmana, J.F., Puentes, R. and Eato, M.J. 2007. Increased spinal c-Fos expression with noxious and non-noxious peripheral stimulation after severe spinal contusion. *Neuroscience Letters*. 413: 58– 62.
- Bilecki, W.; Wawrzczak-Bargiela, A.; Przewlocki, R. 2004. Activation of AP-1 and CRE dependent gene expression via mu-opioid receptor. J. Neurochem. 90: 874– 882.

- Bonavita, C., Ferrero, A., Cereseto, M., Velardez, M., Rubio, M., Wikinski, S. 2003. Adaptive changes in the rat hippocampal glutamatergic neurotransmission are observed during long-term treatment with lorazepam. Psychopharmacology. 166:163–167.
- Boshart, M.; Weih, F.; Schmidt, A.; Fournier, R.E.; Schütz, G.A. 1990. Cyclic AMP response element mediates repression of tyrosine aminotransferase gene transcription by the tissue-specific extinguisher locus Tse-1. Cell 61: 905–916.
- Brandt M, Fisher K, Moroder L, Wunsch E, Hamprecht, B. 1976. Enkephalin evokes biochemical correlates of opiate tolerance and dependence in neuroblastomaXglioma hybrid cells. FEBS Lett 68 (1): 38-40.
- Burkill, I.H., 1935. A Dictionary of the Economic Products of th Malay Peninsula, vol.II. Crown Agents for the Colonies, London, pp 1480-1483
- C. Sternini, M. Spann, B Anton, D.E Keith, N.W Burnett, M Von Zastrow. 1996. Agonist-selective endocytosis of mu opioid receptor by neurons in vivo. Proceeding National Academy Science USA 93. 9241-9246.
- Caarsten Smith-Hall, Helle Overgaard and Marieve Poulict. 2012. Journal of Ethnobiology and Ethnomedicine. 1-11.
- Carol Mattson Porth, 2004. Essentials of Pathophysiology, Concepts of Altered Health States. Lippincott Willims and Milkins. 731-737.
- Catherine F. Stannard and Sara Booth. 2004. Pain. Churchill Livingstone.
- Celik, E., Uzbay, T., and Karakas, S. 2006. Caffeine and amphetamine produce crosssensitization to nicotine-induced locomotor activity in mice. Progress in Neuro-Psychopharmacology and Biological Psychiatry Vol. 30. P. 50-55.
- Chakrabarti, S., Wang, L., Tang, W.-J., Ginzler, A.R. 1998. Chronic morphine augments adenylyl cyclase phosphorylation: Relevance to altered signaling during tolerance/dependence. Molecular Pharmacology 54, 949–953.
- Chan, B.K., Pakiam, C., Rahim, A.R., 2005. Psychoactive plant abuse: the identification of mitragynine in ketum and in ketum preparations. Bulletin on Narcotics, vol. LVII, Nos. 1 and 2
- Chang, P.H., Lindberg, P.F., Wang, L.H., Huang, A.M., Lee, E.H.Y. 1999. Impaired Memory Retention and Decreased Long-Term Potentiation in Integrin-Associated Protein-Deficient Mice. Learning and Memory.Vol. 6.P :448–457
- Chartoff EH, Papadopoulou M, Konradi C, Carlezon Jr WA. 2003. Dopaminedependent increases in phosphorylation of cAMP response element binding protein

(CREB) during precipitated morphine withdrawal in primary cultures of rat striatum. J. Neurochem 87 (1): 107-118.

- Chee, J-W., Amirul, A.A., Majid, M.I.A and Mansor, S.M. 2008. Factors influencing the release of Mitragyna speciosa crude extracts from biodegradable P(3HB-co4HB). International Journal of Pharmaceutics 361: 1-6.
- Chen J, Marmur R, Pulles A, Paredes W, Gardner EL. 1993. Ventral tegmental microinjection of delta 9-tetrahydrocannabinol enhances ventral tegmental somatodendritic dopamine levels but not forebrain dopamine levels: evidence for local neural action by marijuana's psychoactive ingredient. Brain Res 621:65–70
- Chittrakarn, S.; Keawpradub, N.; Sawangjaroen, K.; Kansenalak, S.; Janchawee, B. 2010. The neuromuscular blockade produced by pure alkaloid, mitragynine and methanol extract of kratom leaves (Mitragyna speciosa Korth.). J. Ethnopharmacol. 129, 344-349.
- Cichewicz DL., Martin ZL, Smith FL, Welch SP. 1999. Enhancement mu opioid antinociception by oral delta 9-tetrahydrocannabinol: dose-response analysis and receptor identification. J. Pharmacol Exp Ther. 289:859-867
- Collier, H.O.; Francis, D.L. 1975. Morphine abstinence is associated with increased brain cyclic AMP. Nature. 255: 159–162.
- Corbett A.D., Henderson, G., McKnight, A.T., and Paterson, S. 2006. 75 years of opioid research : the exciting but vain quest for the holy grail. British Journal of Pharmacology. vol. 147: 153-126.
- Coven E, Ni Y, Widnell KL, Chen J, Walker WH, Habener JF, Nestler EJ. 1998. Cell type-specific regulation of CREB gene expression: mutational analysis of CREB promoter activity. J. Neurochem. 71 (5): 1865-1874.
- D.E. Keith, S.R Murray, P.A Zaki, P.C Chu, D.V Lissin, L. Kang. 1996. Morphine activates opioid receptors without causing their rapid internalization. Journal of Biological Chemistry 271. 19021-19024.
- David Borenstein. 2010. The role of the rheumatologist in managing pain therapy. Nature Review Rheumatology. 6: 227-231.
- David M. Simpson, Justin C McArthur, Robert H. Dworkin. 2012. Neuropathic pain: Mechanisms, diagnosis and treatment.
- Devane WA, Hanus L, Breuer A, Pertwee RG, Stevenson LA, Griffin G, Gibson D, Mandelbaum A, Etinger A, Mechoulam R. 1992. Isolation and structure of a brain constituent that binds to the cannabinoid receptor. Science 258:1946–1949.

- DG Lambert. 2004. Drugs and receptors. Continuing Education in Anaesthesia, Critical Care and Pain. Volume 4.
- Dhawan BN, Cesselin F, Raghubir R, Reisine T, Bradley PB, Portoghese PS and Hamon M. 1996. International Union of Pharmacology XII classification of opioid receptors. Pharmacol Rev 48: 567-592.
- Drew, G.M.. Siddall, P.J. and Duggan, A.W. 2004. Mechanical allodynia following contusion injury of the rat spinal cord is associated with loss of GABAergic inhibition in the dorsal horn. *Pain*. 109 (3): 379–388.
- Drolet G, Dumont EC, Gosselin I, Kinkead R, Laforest S, and Trottier JF. 2001. Role of endogenous opioid system in the regulation of the stress response. Prog Neuropsychopharmacol Biol Psychiatry 25: 729-741.
- Du Pen, A.; Shen, D.; Ersek, M. 2007. Mechanisms of opioid-induced tolerance and hyperalgesia.Pain Manag. Nurs. 8: 113–121.
- E. Johnson, S. Oldfield, E. Braksator, A. Gonzales-Cuello, D. Couch, K.J Hall. 2006. Agonist-selective mechanisms of mu opioid receptor desensitization in human embryonic kidney 293 cells. Molecular Pharmacology 70. 676-685.
- Eddy, N.B., and E.L May. 1973. "The search for better analgesic," Science 181: 407-414.
- Elizabeth, A.B., Tallarida, R.J., Pasternak G.W. 2002. Synergy between μ opioid ligands : Evidence for functional interactions among μ opioid receptor subtypes. Pharmacology and Experimental Therapeutice. Vol. 303, Issues 2: 557-562.
- Ferrante FM, Paggioli J, Cherukuri S, Arthur GR. 1996. The analgesic response to intravenous lidocaine in the treatment of neuropathic pain. Anesth Analg 82: 91-117.
- Ferrante FM. 1998. Patient-controlled analgesia: a conceptual framework for analgesic administmiceion. In: Ferrante FM, Vadeboncouer TR, eds. Postopemiceive pain management. New York: Churchill Livingston. 255-277.
- Finkel, M.P., Biskis, B.O. and Jinkins, P.B. 1966. Virus induction of osteosarcomas in mice. Science. 151(3711): 698-701.
- G.P Dureja, H. Usmani, M. Khan, M. Tahseen, A. Jamal. 2010. Efficacy of intrathecal midazolam with or without epidural methylprednisolone for management of post herpetic neuralgia involving lumbosacral dermatomes. Pain Physician, 13: 2013-221.
- Gaoni, Y.; Mechoulam, R. 1964. Isolation, structure and partial synthesis of an active constituent of hashish. J. Am. Chem. Soc. 86: 1646-1647.

- Gaw, A., Cowan, R.A. O'Reilly, D.S.T., Stewart M.J., Shephed, J.S. 1998. Clinical Biochemistry. Churchill Livingstone, Edinburg.
- Gerald J Mulder, Lennart Dencker. 2006. Pharmaceutical Toxicology: Safety sciences of drugs. RPS Publishing. 137-160.
- Grewal, K.S., 1932. Observations on the pharmacology of mitragynine. Journal of Pharmacology and Experimental Therapeutics 46, 251-271
- Guitart, X.; Thompson, M.A.; Mirante, C.K.; Greenberg, M.E.; Nestler, E.J. 1992. Regulation of cyclic AMP response element-binding protein (CREB) phosphorylation by acute and chronic morphine in the rat locus coeruleus. J. Neurochem. 58: 1168–1171.
- Gurdeep, R.; Chatwal, S.; Anand, K. 1998. Instrumental methods of chemical analysis. Himalaya Publishing House. 2: 185 - 234.
- Guy T. Carter. 2010. Natural Products in Drug Discovery, Textbook of Drug Design and Discovery. CRC Press. 89-92
- H.B. Deng, Y. Yu, Y. Pak, B.F. O'Dowd. S.R George. C.K. Surratt. 2000. Role for the C-terminus in agonist-induced mu opioid receptor phosphorylation and desensitization. Biochemistry 39: 5492-5499.
- H.P Rang, M.M Dale, J.M. Ritter, P.K Moore. 2003. Pharmacology, Fifth Edition. Churchill Livingstone, 562-584.
- Harry J. Gould. 2007. Understanding Pain. What it is, Why it happens and how it's managed. American Academy of Neurology Press: 40 60.
- Herkenham, H. 1995. Localization of cannabinoid receptors in brain and periphery. In: Pertwee RG, editor. Cannabinoid receptors. London: Academic Press. 145–166.
- Herkenham, M.; Lynn, A.B.; Little, M.D.; Johnson, M.R.; Melvin, L.S.; de Costa, B.R.; Rice, K.C. 1990. Cannabinoid receptor localization in brain. Proc. Natl. Acad. Sci. USA. 87: 1932-1936.
- Herzberg, U.; Eliav, E.; Bennett, G.J.; Kopin, I.J. 1997. The analgesic effects of R(+)-WIN 55,212-2 mesylate, a high affinity cannabinoid agonist, in a rat model of neuropathic pain. Neurosci. Lett. 221: 157-160.
- Holaday AS, Salvucci ME, Bowes G. 1983. Variable photosynthesis/photorespimiceion miceios in Hydrilla and other submersed aquatic macrophyte species. Can J Bot. 61:229–236.
- Hoon Oh, J., Jae Choi, B., Seog Chang, M., Kyu Park, S. 2009. Nelumbo nucifera semen extract improves memory in rats with scopolamine-induced amnesia through

the induction of choline acetyltransferase expression. Neuroscience Letters 461: 41–44.

- Houghton, P.J & 1kram M. Said. 1986. 3- Dehydromitragynine: an alkaloid from Mitragyna speciosa. Phytochemistry. 25: 2910-2912.
- Howlett, A.C.; Barth, F.; Bonner, T.I.; Cabral, G.; Casellas, P.; Devane, W.A.; Felder, C.C.; Herkenham, M.; Mackie, K.; Martin, B.R.; Mechoulam, R.; Pertwee, R.G. . 2002. International Union of Pharmacology. XXVII. Classification of cannabinoid receptors. Pharmacol. Rev. 54: 161-202.
- Hughes J., Smith T.W., Kolsterlitz H.W., Fothergill L.A., Morgan B.A., and Morris H.R. 1975. Identification of two related pentapeptides from the brain with potent opiate agonist activity. Nature 258: 577-579.
- Hugo F. Miranda, Margarita M. Puig, Juan Carlos Prieto, Gianni Pinardi. 2006. Synergism between paracetamol and nonsteroidal anti-inflammatory drugs in experimental acute pain, Pain 121: 22-28.
- Hunt, S.P., Pini, A. and Evan, G. 1987. Induction of *c-fos*-like protein in spinal cord neurons following sensory stimulation. *Nature*. 328: 632-634.
- Idid, S.Z.,, K. Norehan and A. Roslan. 1992. The involvement of the noradrenergic system in analgesia induced by the alkaloidal extract of Mitragyna speciosa in the rat. Proc. 3rdMedical Colloquim, UKM. pp 337-340.
- Ikram, M.S. 1985. Studies on the components of fresh leaves of Mitragyna speciosa. In: M.S.Ikram & Z.Zakaria (Eds.) Proceedings of 2nd Meeting of the Natural Products Research Group, Chemistry Dept., UKM Malaysia. pp 123-127.
- Isomae, K., Morimoto, S., Hasegawa, H., Morita, K., Kamei, J. 2003. Effects of T-82, a novel acetylcholinesterase inhibitor, on impaired learning and memory in passive avoidance task in rats. European Journal of Pharmacology 465 (2003) 97–103.
- J K Limdi, G M Hyde. 2003. Evaluation of abnormal liver function tests. Pmj. Bmj.com.
- James., L.P Mayeux P.R., Hinson J.A, Acetaminophen-induced hepatotoxicity, Drug Metabolism Dispos.,2003; 31(12), 1499-1506.
- Janchawee, B., Keawpradub, N., Chittrakarn, S., Prasettho, S., Wararatananurak, P., Sawangjareon, K., 2007. A high-performance liquid chromatographic method for determination of mitragynine in serum and its application to a pharmacokinetic study in rats. Biomedical Chromatography 21, 176–183.
- Jansen K.L.R. and C.J.Prast, 1988. Ethnopharmacology of Kratom and the Mitragyna alkaloids. J Ethnopharmacology 23: 115-119.

- Jeffrey A. Grass, MD, M.M.M (2005). Patient controlled analgesia. Anesth Analg 2005;101:S44-S61.
- Jordan BA and Devi LA (1999) G-protein-coupled receptor heterodimerization modulates receptor function. Nature (Lond) 399: 697-700.
- Jordan BA, Cvejic S, and Devi LA (2000) Opioids and their complicated receptor complexes. Neuropsychopharmacology 23: S5-S18.
- Jurgen, S. Understanding LTP in Pain Pathway. Mol. Pain 2007, 3, 100–106. Molecules 2013, 18 681
- Kalivas, P.W., Jackson, D., Romanidies, A., Wyndham, L., Duffy, P. 2001. Involvement of pallidothalamic circuitry in working memory. Neuroscience Vol. 104: 129-136.
- Kathryn L. McCance and Sue E. Huether, 2002. Pathophysiology: The Biologic Basis for Disease in adults and children, Fourth Edition. Mosby Inc : 402-408.
- Kikura-Hanajiri, R., Maruyama, T., Kawamura, M., Takayama, H., Goda, Y. 2009. The botanical origin of kratom (Mitragyna speciosa; Rubiaceae) available as abused drugs in the Japanese markets. Nat Med (Tokyo): 63 (3): 340-4.
- Kjelstrup KG, Tuvnes FA, Steffenach HA, Murison R, Moser EI, and Moser MB. . 2002. Reduced fear expression after lesions of the ventral hippocampus. Proc Natl Acad Sci USA 99: 10825-10830.
- Kumar G, Banu GS, Pappa PV, Sundararajan M, Pandian MR. 2004. Hepatoprotective activity of Trianthema portulacastrum L. against paracetamol and thioacetamide intoxication in albino rats. Journal Ethnopharmacol. 92: 37-40.
- Kumarnsit, E., Vongvatcharanon, U., Keawpradub, N., Intasaro, P. 2007. Fos-like immunoreactivity in rat dorsal raphe nuclei induced by alkaloid extract of Mitragyna speciosa. Neuroscience letters. Vol. 416, Issue 2. P. 128-132.
- Kumarnsit, E.; Keawpradub, N.; Nuankaew, W. 2007. Effect of Mitragyna speciosa aqueous extract on ethanol withdrawal symptoms in mice. Fitoterapia 78: 182-185.
- L.C. Saland, C.M. Hastings, A. Abeyta, J.B. Chavez. 2005. Chronic ethanol modulates delta and mu-opioid receptor expression in rat CNS: immunohistochemical analysis with quantitative confocal microscopy. Neuroscience Letters 381: 163-168.
- Lauretti GR, Lima IC, Reis MP, Prado WA, Pereira NL. 1999. Oral ketamine and transdermal nito-glycerin as analgesic adjuvants to oral morphine therapy for cancer pain management. Anaesthesiology 90: 1528-1533.
- Law PY, Wong YH, Loh HH. 2000. Molecular mechanisms and regulation of opioid receptor signaling. Annu Rev Pharmacol Toxicol. ;40:389–430.

Li, J.; Daughters, R.S.; Bullis, C.; Bengiamin, R.; Stucky, M.W.; Brennan, J.; Simone, D.A. 1999. The cannabinoid receptor agonist WIN 55,212-2 mesylate blocks the development of hyperalgesia produced by capsaicin in rats. Pain 81: 25-33.

Loeser R. 2001. The pain cycle. 107.

- Loewe S. Die quantitativen probleme der pharmakologie. 1928. Ergebn Physiolo 27: 47-187
- Lynn, W.; Crespo, L.M.; George, D.; Carl, F.G.; Stephanie, W. Brody's Human Pharmacology Molecular to Clinical. Mosby Elsevier, Philadelpia, PA, USA, 2010; pp: 74-78.
- M. Hanks-Bell, K. Halvey, J.A. Paice. 2004. Pain assessment and management in aging. Online J. Issues Nursing. 9: 8.
- M.A.A Khan. 2011. Indian medicinal plants, Vol 10. Indian Journal of Experimental Biology, Vol 49 : 797-798.
- Macko, E.; Weisbach, J.A.; Douglas, B. 1972. Some observations on the pharmacology of mitragynine. Arch. Int. Pharmacodyn Ther. 198 : 145-161.
- Mansour A, Fox CA, Akil H, et al. Opioid-receptor mRNA expression in the mice CNS: anatomical and functional implications. Trends Neurosci. 1995;18:22–9.
- Mao, J., Sung, B., Ji, R.-R., Lim, G., 2002. Neuronal apoptosis associated with morphine tolerance: Evidence for an opioid induced neurotoxic mechanism. Journal of Neuroscience 22, 7650–7661.
- Martin, B.R.; Compton, D.R.; Thomas, B.F.; Prescott, W.R.; Little, P.J.; Razdan, R.K.; Johnson, M.R.; Melvin, L.S.; Mechoulam, R.; Ward, S.J. Behavioral, biochemical, and molecular modeling evaluations of cannabinoid analogs. Pharmacol. Biochem. Behav. 1991, 40, 471-478.
- Mary Cardosa and Phoon Ping Chen. 2010. Epidemiology in chronic pain, gender and cultural aspects: 49-59.
- Matsuda, L.A.; Lolait, S.J.; Brownstein, M.J.; Young, A.C.; Bonner, T.I.1990. Structure of a cannabinoid receptor and functional expression of the cloned cDNA. Nature 346 : 561-564.
- Matsumoto K, Mizowaki M, Suchitra T, Takayama H, Sakai S, Aimi N, Watanabe H: 1996a. Antinociceptive action of mitragynine in mice: evidence for the involvement of supraspinal opioid receptors. Life Sciences 59 (14): 1149-1155.

- Matsumoto K, Mizowaki M, Suchitra T, Takayama H, Sakai S, Aimi N, Watanabe H: 1996b. Central antinociceptive effects of mitragynine in mice: contribution of descending noradrenergic and serotogenic systems. European Journal of Pharmacology 317 (1): 75-81.
- Matsumoto K, Takayama H, Ishikawa H, Aimi N, Ponglux D, Watanabe K, Horie S: 2006. Partial agonistic effect of 9-hydroxycorynantheidine on mu-opioid receptor in the guinea-pig ileum. Life Science 78: 2265-2271.
- Matsumoto, K., Yamamoto, L.T., Watanabe, K., Yano, S.,Shan, J., Pang, P.K., Ponglux, D., Takayama, H., Horie, S., 2005. Inhibitory effect of mitragynine, an analgesic alkaloid from Thai herbal medicine, on neurogenic contraction of the vas deferens. Life Sci. 78, 187-194.
- Matsumoto, K.; Horie, S.; Ishikawa, H.; Takayama, H.; Aimi, N.; Ponglux, D.; Watanabe, K. 2004. Antinociceptive effect of 7-hydroxymitragynine in mice: Discovery of an orally active opioid analgesic from the Thai medicinal herb Mitragyna speciosa. Life Sci. 74 : 2143–2155.
- Matsumoto, K.; Mizowaki, M.; Suchitra, T.; Murakami, Y.; Takayama, H.; Sakai, S.; Aimi, N.;Watanabe, H. 1996. Central antinociceptive effects of mitragynine in mice: Contribution of descending noradrenergic and serotonergic systems. Eur. J. Pharmacol. 317 : 75–81.
- Matsumoto, K.; Takayama, H.; Ishikawa, H.; Aimi, N.; Ponglux, D.; Watanabe, K.; Horie, S. 2006. Partial agonistic effect of 9-hydroxycorynantheidine on mu-opioid receptor in the guinea-pig ileum. Life Sci. 78 : 2265-2271.
- McDonald, J.; Lambert, D.G. 2008. Opioid mechanisms and opioid drugs. Anaesth. Intens. Care Med 9: 33–37.
- McMillen, A.B. 1983. CNS stimulants: two distinct mechanisms of action for amphetamine-like drugs. Elsevier Science Publishers, TIPS.
- Mercadante S. 2004. Opioid rotation for cancer pain: rationale and clinical aspects. Cancer 86: 1856-1866.
- Menetrey, D., Gannon, A., Levine, J.D. and Basbaum, A.I. 1989. Expression of c-fos protein in interneurons and projection neurons of the rat spinal cord in response to noxious somatic, articular, and visceral stimulation. *Journal of Comparative Neurology*. 285 (2): 177–195.
- Meyer TE, Habener JF. 1993. Cyclic adenosine 3'5'-monophosphate response element binding protein (CREB) and related transcription-activating deoxyribonucleic acid-binding proteins. Endocr. Rev 14 (3): 269-290.

- Miles Herkenham, Allison B Lynn, Mark D. Litrle, M Ross Johnson, Lawrence S. Melvin, Brian R. De Costa and Kenner C. Riceo. 1990. Nati. Acad. Sci. USA, Neurobiology Vol. 87: 1932-1936.
- Moklas, M.A.M, Nurul Raudzah. A.R., Taufik Hidayat M., Sharida F., Farah Idayu N., Zulkhairi A., Shamima A.R. 2008. A preliminary toxicity study of mitragynine, an alkaloid from Mitragyna speciosa korth and its effects on locomotor activity in mice. Advances in Medical and Dental Sciences, 2(3): 56-60.
- Monje, P., Hernández-Losa, J., Lyons, R.J., Castellone, M.D. and Gutkind, J.S. 2005. Regulation of the transcriptional activity of c-Fos by ERK. A novel role for the prolyl isomerase PIN1. *Journal of Biological Chemistry*. 280 (42): 35081–35084.
- Morgan, James, I. and Curran, Tom. 1991. Stimulus-transcription coupling in the nervous system: involvement of the inducible proto-oncogenes fos and jun *Annual Review of Neuroscience*. 14: 421-51.
- Moritta McCleane. 2003. The cholecystokinin antagonist proglumide enhances the analgesic effect of dihydrocodeine. Clinical Journal of Pain 19: 200-201.
- Mossadeq,W.M.,Sulaiman,M.R.,Mohamad,T.A.T.,Chion,H.S.,Zakaria,Z.A.,Jabit, M.L., Baharuldin,M.T.H.,Israf,D.A.,2009.Anti-inflammatory and antinociceptive effects of Mitragyna speciosa methanolic extract.Medical Principles and Practice 18,378– 384.
- Mulder, G.J. 2000. Pharmaceutical Toxicology: Safety Sciences of Drugs; Dencker, L., Ed.; RPS Publishing: London, UK. : 137–160
- Mumby, G.D., Gaskin, S., and Glenn, J.M., 2002. Hippocampal damage and explomiceory preferences in mice: memory for objects, places and contexts. Learn. Mem. 2002. 9: 49-57.
- Narita, M.; Funada, M.; Suzuki, T. 2001. Regulations of opioid dependence by opioid receptor types. Pharmacol. Ther. 89 : 1–15.
- Nestler E.J, 1993. Cellular responses to chronic treatment of drugs of abuse. Crit. Rev. Neurobiology 7 (1), 23-29.

Nestler EJ. 2000. Genes and addiction. Nat. Genet 26 (3): 277-281.

- Nicholas S. Gregory, Amber L. Harris, Caleb R. Robinson, Patrick M. Dougherty, Perry N. Fuschs and Kathleen A. Sluka. 2013. An Overview of Animal Models of Pain: Disease models and outcome measures. The journal of Pain, Vol 14, No 11: 1255-1269.
- Nicholson, G.P. 2003. Treatment of anterior superior shoulder instability with a reverse ball and socket prosthesis. Oper. Technol. Orthop. 13, 235–241.

- Noble F, Cox BM: Differential desensitization of mu and delta opioid receptors in selected neural pathways following chronic morphine treatment. British Journal of Pharmacology, 117: 161-169.
- Noguchi, K., Kowalski, K., Traub, R., Solodkin, A., Iadarola, M.J. and Ruda, M.A. 1991. Dynorphin expression and Fos-like immunoreactivity following inflammation induced hyperalgesia are colocalized in spinal cord neurons. *Molecular Brain Research.* 10 (3): 227–233.
- Okuda K., Hepatocellular carcinoma: clinicopathological aspects, Journal Gastroenterol. Hepatology. 2(9-10), S314-S318, 1997.
- Onoda, N. 1992. Odor-induced fos-like iminunoreactivity in the rat olfactory bulb. Neuroscience Letters. 137: 147 160.
- Ozsoy-Sacan, O.; Yanardag, R.; Orak, H.; Ozgey, Y.; Yarat, A.; Tunali, T. Effects of parsley activity. J. Neurochem. 1998, 71, 1865–1874.
- Pamela E. Macintyre, Stephen A. Schung (2007). Acute pain management A Practical guide. Saunders Elsevier. Pg 6-74
- Panteghini M., Aspartate aminotransferase isoenzymes. Clinical Biochemistry, 1990. 23 (4) page 311 319.
- Pasternak GW and Standifer KM (1995). Mapping of opioid receptors using antisense oligodeoxynucleotides : correlating their molecular biology and pharmacology. Trends Pharmacol Sci 16: 344-350.
- Pasternak, G.W.: Pharmacological mechanism of opioid analgesics. Clin. Neuropharmacol. 16, 1–18 (1993). The brain. J. Neurosci. 2000, 20, 4555–4562
- Paul A. Smith, Dana E. Selley, Laura J. Sim-Selley and Sandra P. Welch. Low dose combination of morphine and Δ9-tetrahydrocannabinol circumvents antinociceptive tolerance and apparent desensitization of receptors, Eur J Pharmacology; 2007 1; 571 (2-3): 129-137

Penny North-Lewis (2008). Drugs and the liver. Pharmaceutical Press UK. 70-90.

- Pelto-Huikko, M., Schultz, R., Koistinaho, J. and Hökfelt, T. 1991. Immunocytochemical demonstration of c-fos protein in sertoli cells and germ cells in rat testis. *Acta Physiologica Scandinavica*. 141(2): 283-4.
- Pertwee RG (1999) Pharmacology of cannabinoid receptor ligands. Curr Med Chem 6:635–664
- Pertwee RG, Stevenson LA, Griffin G (1993) Cross-tolerance between delta-9tetrahydrocannabinol and the cannabimimetic agents, CP 55,940, WIN 55,212–2 and anandamide. Br J Pharmacol 110:1483–1490

- Peter, J.H.; Ikram, M.S. 3-dehydromitragynine: An alkaloid from Mitragyna speciosa. Phytochemistry 1986, 25, 2910–2912.
- Ploghaus A, Narain C, Beckmann CF, Clare S, Bantick S, Wise R, Matthews PM, Rawlins JN, and Tracey I. Exacerbation of pain by anxiety is associated with activity in a hippocampal network. J Neurosci 21: 9896-98903, 2001.
- Ponglux, D.; Wongseripipatana, S.; Takayama, H.; Kikuchi, M.; Kurihara, M.; Kitajima, M.; Aimi, N.; Sakai, S. A New Indole Alkaloid, 7 alpha-Hydroxy-7Hmitragynine, from Mitragyna speciosa in Thailand. Planta Med. 1994, 60, 580-581.
- Przewlocki R, Przewlocka B (2005).Opioids in neuropathic pain. Curr Pharm Des 11(23):3013–3025
- R.D. Polakiewicz, S. M. Schieferl, L.F. Dorner, V. Kansra, M.J. Comb. 1998. A mitogen-activated protein kinase pathway is required for mu-opioid receptor desensitization. Journal of Biological Chemistry 273.12402-12406.
- Raffa, R.B. Pharmacology of oral combination analgesics: Rational therapy for pain. J. Clin.Pharm. Ther. 2001, 26, 257–264.
- Reanmongkol, W., Keawpradub, N. and Sawangjaroen, K. 2007. Effects of the extracts from Mitragyna speciosa Korth. leaves on analgesic and behavioral activities in experimental animals. Songklanakarin J. Sci. Technol., 29 (Suppl. 1): 39-48.
- Rios, C.; Gomes, I.; Devi, L.A. mu opioid and CB1 cannabinoid receptor interactions: reciprocal inhibition of receptor signaling and neuritogenesis. Br. J. Pharmacol. 2006, 148, 387-395.
- Rodney Croteau, Toni M. Kutchan, Norman G. Lewis, 2000. Natural Products (Secondary metabolites), page 1250-1300.
- S. Chakrabarti, M Rivera, S. Z. Yan, W. J. Tang & A. R. Gintzler. 1998. Chronic morphine augments $G\beta\gamma/G\alpha$ stimulation of adenylyl cyclase: relevance to opioid tolerance. Molecular Pharmacology 54. 655-662
- S. Cvejic & L.A Devi. 1997. Dimerization of the delta opioid receptor: implication for a role in receptor internalization. Journal Biological Chemistry 272. 26959-26964.
- S. Z. Idid, L.B. Saad, H. Yaacob and M.M. Shahimi, ASEAN Review of Biodiversity and Environmental Conservation (ARBEC)
- Saraf, K.M., Anand, A., Prabhakar, S. 2010. Scopolamine Induced Amnesia is Reversed by Bacopa monniera Through Participation of Kinase-CREB Pathway. Neurochem Res (2010) 35:279–287.

- Schmidt, S. Schulz, M. Klutzny, T. Koch, M. Handel and V Hollt. 2000. Involvement of mitogen-activated protein kinase in agonist-induced phosphorylation of the mu opioid receptor desensitization by sustained phosphorylation of serine-375. EMBO Journal 23. 3282-3289.
- Schubert, M. and Albrecht, D. 2008. Activation of Kainate GLUK5 Transmission Rescues Kindling-Induced Impairment of LTP in the Rat Lateral Amygdala. Neuropsychopharmacology (2008) 33, 2524–2535
- Shaik Mossadeq, W.M.; Sulaiman, M.R.; Tengku Mohamad, T.A.; Chiong, H.S.; Zakaria, Z.A.; Jabit, M.L.; Baharuldin, M.T.; Israf, D.A. Anti-inflammatory and antinociceptive effects of Mitragyna speciosa Korth methanolic extract. Med. Princ. Pract. 2009, 18, 378-384.
- Shamima, A.R.; Fakurazi, S.; Hidayat, M.T.; Hairuszah, I.; Moklas, M.A.M.; Arulselvan, P. Antinociceptive Action of Isolated Mitragynine from Mitragyna Speciosa through Activation of Opioid Receptor System. Int. J. Mol. Sci. 2012, 13, 11427–11442.
- Sharma SK, Nirenberg M, Klee WA: Morphine receptors as regulators of adenylate cyclase activity. Proc. Natl. Acad. Sci. USA, 1975, 72 (2): 590-594.
- Shaw-Lutchman TZ, Barrot M, Wallace T, Gilden L, Zachariou V, Impey S, Duman RS, Storm D, Nestler EJ: Regional and cellular mapping of cAMP response element-mediated transcription during naltrexone-precipitated morphine withdrawal. J. Neuroscience 2002, 22 (9): 3663-3672.
- Sheng M, Thompson MA, Greenberg ME: CREB: a Ca(2+)-regulated transcription factor phosphorylated by calmodulin-dependent kinases. Science, 1991, 252 (5011): 1427-1430.
- Sheng, M.; Thompson. M.A.; Greenberg, M.E. CREB: A Ca(2+)-regulated transcription factor phosphorylated by calmodulin-dependent kinases. Science 1991, 252, 1427– 1430.
- Shyamal S., Latha, P.G Shine, V.J Suja, S.R Rajasekharan S., Devi, T.G. Hepatoprotective effects of Pittosporum neelgherrense Wight&Arn., a popular India ethnomedicine, Journal Ethnopharmacology, 107, 151-155.
- Sim LJ, Selley DE, Dworkin SI, Childers SR: Effects of chronic morphine administration on mu-opioid receptor-stimulated [35S]GTPgammaS autoradiography in rat brain. Journal of Neuroscience, 1996, 16, 2684-2692.
- Simone, C.; Bosshard, C.B.; Matthias, T.; Wyss, T.M.; Bruno, W.; Markus, R. Assessment of Pain responses to innocuous and noxious electrical forepaw stimulation in mice using fMRI. Pain 2010, 655–663.

- Sim-Selley LJ, Selley DE, Vogt LJ, Childers SR, Martin TJ: Chronic heroin selfadministration desensitizes mu opioid receptor-stimulated 35SGTPgammaS autoradiography in rat brain. Journal of neuroscience, 2000, 20: 4555-4562.
- Singh, S.N., Vats, P.; Suri, S.; Shyam, R.; Kumria, M.M.; Ranganathan, S.; Sridharan, K. Effect of an antidiabetic extract of Catharanthus roseus on enzymic activities in streptozotocin induced diabetic rats. J. Ethnopharmacol. 2001, 76, 269–277.
- Smeyne, R.J., Schilling, K., Robertson, L., Luk, D., Oberdick, J., Curran, T. and Morgan, J.I. 1992. Fos-lacZ transgenic mice: mapping sites of gene induction in the central nervous system. *Neuron*. 8(1): 13-23.
- Smith et al (1998) The enhancement of morphine antinocicpetion in mice by r9tetrahydrocannabinol, Pharm Biochem and Behaviour, 60(2), 559-566
- Smith, S., Dringenberg, C.H., Bennet, M.B., Thatcher, R.J.G., Reynolds, N.J. 2000. A novel nitrate ester reverses the cognitive impairment caused by scopolamine in the Morris water maze. Neuro Report. Lippincott Williams & Wilkins. Vol. II. No.17.
- Spinella, M. 2001. The Psychopharmacology of Herbal Medicine : Plant Drugs That Alter Mind, Brain, and Behavior. MIT Press. Cambridge, England. P.1-3.
- Suwanlert, 1974. A study of kratom eaters in Thailand. Thai Bull. on Narcotics. 26:21-27.
- Svensson, L.A., Bucht, N., Hallberg, M., Nyberg, F. 2008. Reversal of opiate-induced apoptosis by human recombinant growth hormone in murine foetus pri mary hippocampal neuronal cell cultures. PNAS. Vol. 105. No. 20. p.7304-7308.
- Sylvia Prosser, Barbara Morster, Janet MacGregor, Kate Dewar, Pauline Runyard, Julie Fagan (2000). Applied Pharmacology. Mosby.344-360
- T. Koch, T. Kroslak, P. Mayer, E. Raulf, V. Hollt. 1997. Site mutation in the rat mu opioid receptor dmonstrates th involvement of calcium/ calmodulin-dependent protein kinase II in agonist-mediated desensitization. Journal of Neurochemistry 69. 1767-1770.
- Takayama H, Ishikawa H, Kurihara M, Kitajima M, Aimi N, Ponglux D, Koyama F, Matsumoto K, Moriyama T, Yamamoto LT, Watanabe K, Murayam T, Horie S: Studies on the synthesis and opioid agonistic activities of mitragynine-related indole alkaloids: discovery of opioid agonists structurally different from other opioid ligands. J. Med. Chem. 2002, 45: 1949-1956.
- Takayama, H., 2004. Chemistry and pharmacology of analgesic indole alkaloids from the rubiaceous plant, Mitragyna speciosa. Chem. Pharm. Bull. 52, 916-928.

- Takayama, H.; Aimi, N.; Sakai, S.: Chemical studies on the analgesic indole alkaloids from the traditional medicine (Miragyna speciosa) used for opium substitute. Yakugaku Zasshi 120, 959–967 (2000).
- Takayama, H.; Ishikawa, H.; Kurihara, M.; Kitajima, M.; Aimi, N.; Ponglux, D.; Koyama, F.;Matsumoto, K.; Moriyama, T.; Yamamoto, L.T.; et al. Studies on the synthesis and opioid agonistic activities of mitragynine-related indole alkaloids: Discovery of opioid agonists structurally different from other opioid ligands. J. Med. Chem. 2002, 45, 1949–1956.
- Takayama, H: Chemistry and pharmacology of analgesic indole alkaloids from the rubiaceous plant, Mitragyna speciosa. Chem. Pharm. Bull, 2004, 52: 916-928.
- Taufik Hidayat, M., Apryani, E., Nabishah, B.M., Miklas, M.A.A., Sharida, F., Farhan, M.A., 2010. Determination of mitragynine bound opioid receptors. Advances in Medical and Dental Sciences 3 (3), 65–70.

Tham et al (2005) British J Pharmacol 144(6) 875-884

- Thomas Koch and Volker Hollt, 2008. Role of receptor internalization in opioid tolerance and dependence. Pharmacology and Therapeutics. 199-206.
- Thongpradichote S, Matsumoto K, Tohda M, Takayama H, Aimi N, Sakai S, Watanabe H: Identification of opioid receptor subtypes in antinociceptive actions of supraspinally administered mitragynine in mice. Life Sciences, 1998, 62 (16): 1371-1378.
- Thongpradichote, S.; Matsumoto, K.; Thoda, M.; Takayama, H.; Aimi, N.; Sakai, S.; Watanabe, H.: Identification of opioid receptor subtypes in antinociceptive actions of supraspinally administered mitragynine in mice. Life Sci. 62, 1371 –1378 (1998)
- Thongpradichote, S.; Matsumoto, K.; Tohda, M.; Takayama, H.; Aimi, N.; Sakai, S.; Watanabe, H. Identification of opioid receptor subtypes in antinociceptive actions of supraspinally-administered mitragynine in mice. Life Sci. 1998, 62, 1371-1378.
- Tirault, M.MD, Derrode, N. MD., Rolland, D. MD., Fletcher, D.MD., and Bertrand, D.MD., (2006). The effect of nefopam on morphine overconsumption induced by large-dose remifentanil during propofol anesthesia for major abdominal surgery. Anest Analg 2006; 102: 110-7.
- Todd, A.J., Spike, R.C., Young, S. and Puskar, Z. 2005. Fos induction in lamina I projection neurons in response to noxious thermal stimuli. *Neuroscience* 131 (1): 209–217.
- Tohda, M., Thongpradichote, S., Matsumoto, K., Murakami, Y., Sakai, S., Aimi, N., Takayama, H., Thongroach, P., Watanabe, H., 1997. Effects of mitragynine on

cAMP ormation mediated by δ -opiate receptors in NG108-15 cells. Biological Pharmaceutical Bulletin 20 (4), 338-340.

- Tsuchiya, J., Tsuchiya, T., Tsuneyuki, S., and Yamanaka, T. (2002) First principles calculation of a high pressure hydrous phase δ -AlOOH. Geophysical Research Letters, 29, 1909.
- Turk, D.C., Melzack R. 1992. Handbook of Pain Assessment. New York; Guilford Press.
- Valle L, Puig MM, Pol O. Effects of mu-opioid receptor agonists on intestinal secretion and permeability during acute intestinal inflammation in mice. Eur. J. Pharmacol. 2000;389:235–242.
- Victoria L. Haller, Diana L. Cicheewicz, Sandra P. Welch., 2006. Non-cannabinoid CB1, non-cannabinoid CB2 antinociceptive effects of several novel compounds in the PPQ stretch test in mice. European Journal of Pharmacology 546, 60-68.
- W. S. Puttfarcken, L.L Werling and B.M Cox. 1988. Effects of chronic morphine exposure on opioid inhibition of adnylyl cyclase in 7315 cell membranes: a useful model for the study of tolerance and mu opioid receptors. Molecular Pharmacology 33. 520-527
- W.M. Walwyn, D.E. Keith, W. Wei, A. Tan, A.M Xie, C.J Evans et al. 2004. Functional coupling, desensitization and internalization of virally expressed mu opioid receptors in cultured dorsal root ganglion neurons from mu opioid receptor knockout mice. Neuroscience 123. 111-121
- Walker, J.M.; Hohmann, A.G.; Martin, W.J.; Strangman, N.M.; Huang, S.M.; Tsou, K. The neurobiology of cannabinoid analgesia. Life Sci. 1999, 65, 665-673.
- Walczak, J.S., Pichette, V., LeBlond, F., Desbiens, K. and Beaulieu, P. 2006. Characterization of chronic constriction of the saphenous nerve, a model of neuropathic pain in mice showing rapid molecular and electrophysiological changes. *Journal of Neuroscience Research.* 83 (7): 1310–1322.
- Wantana, R., K. Niwat and S. Kitja, 2007. Effects of the extracts from Mitragyna speciosa Korth. leaves on analgesic and behavioral activities in experimental animals. Songklanakarin J. Sci. Technol., 29(Suppl.1): 39-48.
- Watanabe, H.; Shibuya, T.; Farnsworth, N.R.: Chapter 11: Harwood Academic Press, Tokyo, pp. 163–177 (1999)
- Watanabe, K., Yano, S., Horie, S., Yamamoto, L.T., Sakai, S., Takayama, H., Ponglux, D., Wongseripipatana, S., 1992,. Pharmacological profiles of "Kratom" (Mitragyna speciosa), a Thai medical plant with special reference to its analgesic activity. In: Tongroach, P., Watanabe, H., Ponglux, D., Suvanakoot, U., Ruangrungsi, N. (Eds.), Advances in Research on Chiang Mai University, Chiang Mai, Thailand, pp 125-132.

- Watanabe, K.; Yano, S.; Horie, S.; Yamamoto, L.T.: Inhibitory effect of mitragynine, an alkaloid with analgesic effect from Thai medicinal plant Mitragyna speciosa, on electrically stimulated contraction of isolated guinea-pig ileum through the opioid receptor. Life Sci. 60, 933–942 (1997).
- Watanabe, K.; Yano, S.; Horie, S.; Yamamoto, L.T.; Takayama, H.; Aimi, N.; Sakai, S.; Ponglux, D.; Tongroach, P.; Shan, J.; Pang, P.K.T.: Pharmacological properties of some structurally related indole alkaloids contained in the Asian herbal medicines, hirsutine and mitragynine, with special reference to their Ca2+ antagonistic and opioid-like effects. In Pharmacological Research on Traditional Herbal Medicines. ed.
- Wei, H., Chen, Y. and Hong, Y. 2005. The contribution of peripheral 5hydroxytryptamine2A receptor to carrageenan-evoked hyperalgesia, inflammation and spinal Fos protein expression in the rat. *Neuroscience*. 132(4): 1073–1082.
- White, P.F. The changing role of non-opioid analgesic techniques in the management of postoperative pain. Anesth. Analg. 2005, 101, S5–S22.
- Widnell, K.L.; Russell, D.S.; Nestler, E.J. Regulation of expression of cAMP response element binding protein in the locus coeruleus in vivo and in a locus coeruleus-like cell line in vitro. Proc. Natl. Acad. Sci. USA 1994, 91, 10947–10951.
- William A. Carlezon Jr., Johannes Thome, Valerie G. Olson, Sarah B. Lane-Ladd, Edward S. Brodkin, Noboru Hiroi, Ronald S. Duman, Rachael L. Neve, Eric J. Nestler. (1998). Science 282, 2272 (1998); DOI: 10.1126/science.282.5397.2272
- Willis, W.d., and Westlund KN. 1997. Neuroanatomy of Pain System. Journal of Clinical Neurophysiology 14: 2-31
- Wolfang PF: The changing role of nonopioid analgesic techniques in the management of postoperative pain. Anaesthesia and analgesia, 2007, 101(5 suppl): 5-22.
- Yabaluri, N.; Medzihradsky, F. Down-regulation of mu-opioid receptor by full but not partialagonists is independent of G protein coupling. Mol. Pharmacol. 1997, 52, 896–902.
- Yaksh, T.L.; Malmberg, A.B. Spinal actions of NSAIDS in blocking spinally mediated hyperalgesia: the role of cyclooxygenase products. Agents Actions Suppl. 1993, 41, 89-100.
- Yamamoto, L.T.; Horie, S.; Takayama, H.; Aimi, N.; Sakai, S.; Yano, S.; Shan, J.; Pang, P.K.T.; Ponglux, D.; Watanabe, K.: Opioid receptor agonistic characteristics of mitragynine pseudoindoxyl in comparison with mitragynine derived from Thai medicinal plant Mitragyna speciosa. Gen. Pharmacol. 33, 73–81 (1999).

- Yen FL, Wu TH, Lin LT, Lin CC: Hepatoprotective and antioxidant effects of Cuscuta chinensis against acetaminophen-induced hepatotoxicity in rats. J. Ethnopharmacol, 2007, 111: 123-128.
- Zhang L, Yu Y, Mackin S, Weight F.F, Uhl G.R and Wang J.B. 1996. Differential mu opiate receptor phosphorylation and desensitization induced by agonists and phorbol esters. Journal of Biological Chemistry 271. 11449-11454.
- Zhang, G., Franklin, P.H. and Murray, T.F. 1993. Manipulation of endogenous adenosine in the rat prepiriform cortex modulates seizure susceptibility. *Journal of Pharmacology and Experimental Therapeutics*. 264: 1415–1424.
- Zhao, X.; Xu, Y.; Zhao, Q.; Chen, CR.; Liu, A.M.; Huang, Z.L. Curcumin exerts antinociceptive effects in a mouse model of neuropathic pain: descending monoamine system and opioid receptors are differentially involved. Neuropharmacology 2012, 62, 843–854.
- Zhao-Qi, W., Catherine, O., Agamemnon, E., Grigoriadis, U.M., Ulrich, R. and Erwin, F.W. 1992. Bone and haematopoietic defects in mice lacking c-fos. *Nature*. 360: 741–745.
- Zurina Hassan, Mustapha Muzaimi, Visweswaran Navaratnam, Nurul H.M. Yusoff, Farah W. Suhaimi, Rajakumar Vadivelu, Balasingam K. Vicknasingam, Davide Amato, Stephan von Hörsten, Nurul I.W. Ismail, Nanthini Jayabalan, Ammar I. Hazim, Sharif M. Mansor, Christian P. Müller, Neuroscience and behavioural reviews, 37 (2013) 138–151.