



UNIVERSITI PUTRA MALAYSIA

***CHARACTERIZATION OF STEMPHYLIUM SPECIES ASSOCIATED WITH
DISEASE OF VEGETABLE CROPS IN MALAYSIA***

ABBAS NASEHI

FP 2014 42



**CHARACTERIZATION OF *STEMPHYLIUM* SPECIES ASSOCIATED WITH
DISEASE OF VEGETABLE CROPS IN MALAYSIA**

By

ABBAS NASEHI

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfillment of the Requirements for the Degree of
Doctor of Philosophy**

June 2014

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



DEDICATION

This thesis is dedicated to:

All I love,

My beloved mother,

My beloved father,

Specially my beloved wife "Hajar" who have been a constant source of encouragement to continue my study and for their boundless love, understanding, patience and support throughout my study in Malaysia.

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfillment for the requirement for the degree of Doctor of Philosophy

CHARACTERIZATION OF *STEMPHYLIUM* SPECIES ASSOCIATED WITH DISEASE OF VEGETABLE CROPS IN MALAYSIA

By

ABBAS NASEHI

June 2014

Chairman: Associate Professor Jugah B Kadir, PhD
Faculty: Agriculture

Leaf spot caused by *Stemphylium* spp. is one of the important diseases of vegetable crops worldwide. Forty-six isolates of *Stemphylium* were collected from diseased tomato (*Lycopersicon esculentum* L.), eggplant (*Solanum melongena* L.), pepper (*Capsicum annuum* L.) and lettuce (*Lactuca sativa* L.) from major vegetable production regions of Malaysia, including the states of Pahang, Johor and Selangor during 2011 - 2012. The isolates were examined based on morphological, cultural and molecular characterizations and pathogenicity. Morphological characterization revealed that the isolates were variable in color, texture, shape and pigmentation of the colonies on potato dextrose agar (PDA) and divided into 5 groups. The isolates were also variable in terms of colony size, growth rate, and size of conidia and conidiophores. Little variations were observed in the number of transverse and longitudinal septa of conidia in the isolates. The variations observed among the isolates were not correlated either to host or geographical origins of the isolates. Conidia of the isolates were similar in length, width and length/width ratio to *Stemphylium solani* G. F. Weber, but the isolates were also similar to *S. lycopersici* (Enjoji) W. Yamam. with distinct constrictions at the main transverse septa of conidia and in cultural features, particularly secretion of a yellow to dark red pigment of majority of the isolates on PDA.

Various DNA regions of the isolates, including the internal transcribed spacer (ITS), glyceraldehyde-3-phosphate dehydrogenase (*gpd*), elongation factor 1-alpha protein (*EF-1 alpha*) and the noncoding region between the vacuolar membrane ATPase catalytic subunit A gene (*vmaA*) and a gene involved in vacuolar biogenesis (*vpsA*) regions, were compared to epitype species of the genus *Stemphylium* existing in the GenBank. No variation was observed among the isolates based on the four DNA regions examined and the isolates showed 100 % similarity to each other. Based on molecular analyses, the isolates associated with vegetable crops were confirmed to be *S. lycopersici*,

and were distinguished clearly from *S. solani* and other *Stemphylium* species available in the GenBank.

Pathogenicity tests revealed that all *S. lycopersici* isolates were pathogenic on the original hosts, namely tomato, eggplant, pepper and lettuce. However, virulence variability was observed among the isolates. This study confirmed that *S. lycopersici* isolates were the causal agent of leaf spot on the vegetable crops examined. In cross-inoculation assay, the representative *S. lycopersici* isolates selected from each crop were able to cause leaf spot on the inoculated leaves of host plants found in this study, but not on cabbage as a non-host plant. This indicated an apparent lack of host-specificity in *S. lycopersici* isolates on the four host crops.

Genetic diversity was also assessed among 46 *S. lycopersici* isolates using RAPD and ISSR markers. Both RAPD and ISSR markers used to investigate the genetic variability among the isolates from the vegetable crops were able to differentiate the isolates examined. The results between the markers were congruent, however the effective multilocus marker ratio and subsequent to that the number of private alleles was greater with ISSR markers. Analysis of the genetic variation among the isolates showed significant concordance with the host plants, and no apparent geographic effect was observed. The results of the present study will provide better insight into the biology and etiology of *S. lycopersici* and these achievements will be helpful in the development of better disease management strategies and in the breeding of resistant varieties for vegetable production in Malaysia.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia
Sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

PENCIRIAN SPESIS *STEMPHYLIUM* YANG TERLIBAT DALAM PENYAKIT TANAMAN SAYUR DI MALAYSIA

Oleh

ABBAS NASEHI
Jun 2014

Pengerusi: Profesor Madya Jugah B Kadir, PhD
Fakulti: Pertanian

Kesan bintik pada daun yang disebabkan oleh *Stemphylium* spp. merupakan salah satu penyakit yang utama dalam tanaman sayur-sayuran di seluruh dunia. Empat puluh enam isolat *Stemphylium* telah dikumpulkan daripada tomato berpenyakit (*Lycopersicon esculentum* L.), terung (*Solanum melongena* L.), lada (*Capsicum annuum* L.) dan salad (*Lactuca sativa* L.) dari kawasan pengeluaran sayur-sayuran utama di Malaysia termasuklah negeri Pahang, Johor dan Selangor pada 2011 - 2012. Isolat telah diperiksa berdasarkan ciri morfologi, kultur, ciri-ciri molekular dan patogenisiti. Pencirian morfologi mendedahkan bahawa isolat-isolat mempunyai kepelbagaian koloni dari segi warna, tekstur, bentuk dan pigmentasi di atas agar potato dextrose (PDA) dan telah dibahagikan kepada 5 kumpulan. Isolat juga berubah dari segi saiz koloni, kadar pertumbuhan, dan saiz konidia dan konidiofor. Sedikit variasi diperhatikan pada beberapa septa konidia menegak dan melintang dalam isolat tersebut. Perbezaan antara isolat-isolat tersebut tidak mempunyai korelasi, sama ada kepada perumah atau asal-usul geografi isolat tersebut. Konidia daripada isolat-isolat tersebut mempunyai persamaan dari segi panjang, lebar dan nisbah panjang/lebar *Stemphylium solani* GF Weber, tetapi isolat tersebut juga sama dengan *S. lycopersici* (Enjoji) W. Yamam dengan keadaan jelas lebih sempit pada septa utama melintang di konidia dan juga pada ciri-ciri kultur; terutamanya terdapat rembesan pigmen kuning sehingga warna merah gelap pada kebanyakan isolat atas PDA.

Pelbagai kawasan DNA pada isolat-isolat tersebut telah dikenalpasti termasuklah, spacer dalaman disalin (ITS), gliseraldehid-3-fosfat dehidrogenase (*gpd*), faktor pemanjangan 1-alfa protein (*EF-1 alfa*) dan kawasan tidak berkod antara membran vakuolar ATPase sub unit pemangkin gen A (*vmaA*) dan satu gen yang terlibat dalam kawasan vakuolar biogenesis (*vpsA*). Kesemua kawasan yang telah dikenal pasti itu telah dibandingkan dengan spesies epitype daripada genus *Stemphylium* yang sedia ada dalam

GenBank. Tiada perubahan yang diperhatikan di kalangan isolat-isolat berdasarkan empat kawasan DNA dikaji dan isolat tersebut menunjukkan 100% persamaan antara satu sama lain. Berdasarkan analisis molekular, isolat-isolat yang diperoleh dari tanaman sayur-sayuran telah disahkan merupakan *S. lycopersici*, dan telah dibezakan dengan jelas dari *S. solani* dan spesies *Stemphylium* lain yang terdapat di GenBank.

Ujian pathogenisiti telah menunjukkan bahawa semua isolat-isolat *S. lycopersici* adalah bersifat patogenik pada perumah asal, iaitu tomato, terung, lada dan salad. Walau bagaimanapun, tahap kepelbagaian yang agak tinggi dalam sifat pathogenisiti telah diperhatikan antara isolat-isolat tersebut terutamanya perbezaan dari segi keagresifannya. Kajian ini telah mengesahkan bahawa isolat-isolat *S. lycopersici* merupakan ejen penyebab penyakit daun berbintik pada tanaman sayur-sayuran yang telah diperiksa. Dalam kajian silang inokulasi, isolat *S. lycopersici* yang dipilih dari setiap tanaman telah menunjukkan simptom berbintik pada daun tumbuh-tumbuhan perumah yang telah diinokulasi dalam kajian ini, tetapi tidak pada kobis sebagai tumbuhan bukan perumah. Ini menunjukkan bahawa, kekurangan kekhususan isolat *S. lycopersici* terhadap perumah di dalam empat tanaman tersebut.

Kepelbagaian genetik turut dinilai di antara 46 isolat *S. lycopersici* dengan menggunakan penanda RAPD dan ISSR. Kedua-dua penanda RAPD dan ISSR digunakan untuk mengkaji kepelbagaian genetik di kalangan isolat-isolat yang diambil daripada tanaman sayur-sayuran dan ia dapat membuat perbezaan antara isolat yang diperiksa. Keputusan antara penanda adalah kongruen, tetapi nisbah penanda berkesan dan seterusnya bilangan alel yang lebih tinggi diperolehi dengan menggunakan penanda ISSR. Analisis perubahan genetik antara isolat menunjukkan konkordans yang signifikan dengan tumbuhan perumah dan tiada kesan geografi yang jelas diperhatikan. Hasil kajian ini akan memberikan gambaran yang lebih baik ke dalam biologi dan etiologi *S. lycopersici* dan pencapaian ini akan membantu dalam pembangunan strategi pengurusan penyakit yang lebih baik dan di dalam pembiakbakaan varieti yang lebih resistan terutama bagi pengeluaran sayur-sayuran di Malaysia.

ACKNOWLEDGEMENTS

My special thanks go to my supervisor Assoc. Prof. Dr. Jugah Kadir for his guidance, patience and support throughout this study. This dissertation would not have been possible without his support and guidance. Much thanks to all of my committee members; Assoc. Prof. Dr. Zainal Abidin b Mior Ahmad and Assoc. Prof. Dr. Wong Mui Yun. They have been an excellent source of advice throughout the source of my study. My gratitude also goes to former and current lab assistants. Department of plant protection and agriculture program faculty and staff members were also very helpful. I want to express deepest gratitude to the people I have had the pleasure to work within the Department of Plant Protection, UPM. I would like to acknowledge the support and encouragement of my family. Thank you especially to my wife, Hajar, for her love, endless support and being there for me throughout it all. I just want to say thank you. Last but not least, I want to express my gratitude to my family members especially my mom, my father, my brothers and sisters, who have always been encouraging me, giving me hope for my life from beginning until now. Above all, I am grateful to God for health and the many blessings in my life.

I certify that a Thesis Examination Committee has met on 6 June 2014 to conduct the final examination of Abbas Nasehi on his thesis entitled “Characterization of *Stemphylium* Species Associated with Disease of Vegetable Crops in Malaysia” in accordance with the Universities and University College Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A)106] 15 March 1998. The committee recommends that the student be awarded the degree of Doctor of Philosophy.

Members of the thesis Examination Committee were as follows:

Lau Wei Hong, PhD

Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Kamaruzaman Sijam, PhD

Faculty of Agriculture
Universiti Putra Malaysia
(Internal Examiner)

Ganesan Vadamalai, PhD

Faculty of Agriculture
Universiti Putra Malaysia
(Internal Examiner)

Robert Chester Kemerait, Jr., PhD

College of Agricultural and Environmental Sciences
University of Georgia
USA
(External Examiner)

NORITAH OMAR, PhD

Associate Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date: 23 June 2014

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows:

Jugah bin Kadir, PhD
Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Zainal Abidin b Mior Ahmad, PhD
Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

Wong Mui Yun, PhD
Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

BUJANG BIN KIM HUAT, PhD
Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Declaration by graduate student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signature: _____ Date: _____

Name and Matric No.: Abbas Nasehi / GS29711

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision;
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

Signature: _____

Name of
Chairman of
Supervisory
Committee: **Associate Professor
Jugah bin Kadir**

Signature: _____

Name of
Member of
Supervisory
Committee: **Associate Professor
Zainal Abidin b Mior Ahmad**

Signature: _____

Name of
Member of
Supervisory
Committee: **Associate Professor
Wong Mui Yun**

TABLE OF CONTENTS

	Page
DEDICATION	ii
ABSTRACT	iii
ABSTRAK	v
ACKNOWLEDGEMENTS	vii
DECLARATION	x
LIST OF TABLES	xv
LIST OF FIGURES	xvii
LIST OF ABBREVIATIONS	xx
CHAPTER	
1 INTRODUCTION	1
1.1 Importance and constraints of vegetable production	1
1.2 Objectives of study	2
2 LITERATURE REVIEW	3
2.1 <i>Stemphylium</i> leaf spot	3
2.2 The genus <i>Stemphylium</i>	3
2.3 Symptoms caused by <i>Stemphylium</i> species	4
2.3.1 Symptoms caused by <i>Stemphylium</i> species on solanaceae crops	4
2.3.2 Symptoms caused by <i>Stemphylium</i> species on lettuce	5
2.3.3 Symptoms caused by <i>Stemphylium</i> species on onion and garlic	5
2.3.4 Symptoms caused by <i>Stemphylium</i> species on spinach and asparagus	5
2.4 Morphological Characterization	5
2.5 Phylogeny of <i>Stemphylium</i> species	11
2.6 Genetic variability	13
3 MORPHOLOGICAL, MOLECULAR AND PHYSIOLOGICAL CHARACTERIZATION OF <i>STEMPHYLIUM</i> SPECIES ASSOCIATED WITH VEGETABLE CROPS	17
3.1 Introduction	17
3.2 Materials and Methods	19
3.2.1 Sample collection	19
3.2.2 Isolation	19
3.2.3 Morphological characteristics	19

3.2.3.1 Colony morphology	19
3.2.3.2 Conidial morphology	20
3.2.3.3 Statistical analysis	20
3.2.4 Molecular characterization	20
3.2.4.1 DNA extraction	20
3.2.4.2 PCR amplification	21
3.2.4.3 Gel electrophoresis and staining	23
3.2.4.4 Purification and sequence analysis of multigene	23
3.2.4.5 Phylogenetic analyses	23
3.2.5 Cultural and physiological studies	26
3.2.5.1 Cultural characteristics on different solid media	26
3.2.5.2 Physiological characteristics	26
3.3 Results	27
3.3.1 Symptoms	27
3.3.2 Isolation	29
3.3.3 Morphological characteristics	31
3.3.3.1 Colony morphology	31
3.3.3.2 Conidial morphology	34
3.3.4 Molecular characterization	40
3.3.4.1 Sequence analysis of <i>Stemphyllum</i> isolates based on the ITS region	40
3.3.4.2 Phylogenetic analysis of <i>Stemphyllum</i> isolates based on the ITS region	41
3.3.4.3 Sequence analysis of <i>Stemphyllum</i> isolates based on the <i>gpd</i> gene	45
3.3.4.4 Phylogenetic analysis of <i>Stemphyllum</i> isolates based on the <i>gpd</i> gene	46
3.3.4.5 Sequence analysis of <i>Stemphyllum</i> isolates based on the <i>EF-1 alpha</i> gene	50
3.3.4.6 Phylogenetic analysis of <i>Stemphyllum</i> isolates based on the <i>EF-1 alpha</i> gene	51
3.3.4.7 Sequence analysis of <i>Stemphyllum</i> isolates based on the <i>vmaA-vpsA</i> region	55
3.3.4.8 Phylogenetic analysis of <i>Stemphyllum</i> isolates based on the <i>vmaA-vpsA</i> region	56
3.3.4.9 Phylogenetic analysis of <i>Stemphyllum</i> isolates based on multigene sequences	60
3.3.5 Cultural and physiological studies	63
3.3.5.1 Cultural characteristics on different solid media	63
3.3.5.2 Effect of temperature on mycelial growth	66
3.3.5.3 Effect of temperature on conidial germination	67
3.3.5.4 Effect of media and light on sporulation	68
3.4 Discussion	69

3.5 Conclusion	73
4 PATHOGENICITY AND CROSS-INOCULATION OF <i>STEMPHYLIUM LYCOPERSICI</i> ON VEGETABLE CROPS	75
4.1 Introduction	75
4.2 Materials and Methods	75
4.2.1 Fungal isolates	75
4.2.2 Pathogenicity and cross-inoculation assays	75
4.2.3 Statistical analyses	76
4.3 Results	77
4.3.1 Pathogenicity assays	77
4.3.2 Cross-inoculation assays	78
4.4 Discussion	81
4.5 Conclusion	81
5 GENETIC DIVERSITY AMONG <i>STEMPHYLIUM LYCOPERSICI</i> ISOLATES USING RAPD AND ISSR MARKERS	83
5.1 Introduction	83
5.2 Materials and Methods	84
5.2.1 Fungal isolates	84
5.2.2 DNA extraction	84
5.2.3 Random amplified polymorphic DNA (RAPD) analysis	84
5.2.4 Inter Simple Sequence Repeat (ISSR) Analysis	85
5.2.5 Gel electrophoresis and staining	86
5.2.6 Data analysis	87
5.3 Results	87
5.3.1 Cluster analysis of <i>Stemphylium lycopersici</i> isolates based on RAPD markers	87
5.3.2 Cluster analysis of <i>Stemphylium lycopersici</i> isolates based on ISSR markers	90
5.4 Discussion	93
5.5 Conclusion	94
6 SUMMARY, CONCLUSIONS AND RECOMMENDATIONS FOR FUTURE RESEARCH	95
6.1 Summary and conclusions	95
6.2 Recommendations for future research	98
REFERENCES	101
APPENDIX	115
BIODATA OF STUDENT	147
LIST OF PUBLICATIONS	149

LIST OF TABLES

Table	Page
2. 1 A synopsis of morphological measurements of conidia and conidiophores of some <i>Stemphylium</i> species.	8
2. 2 The key for identification of some plant pathogenic <i>Stemphylium</i> species.	10
3. 1 The four primer pairs used for PCR amplification and DNA sequencing.	22
3. 2 GenBank accession number for species and isolates included in DNA sequence comparisons.	25
3. 3 Sources of <i>Stemphylium</i> isolates used in the study.	30
3. 4 Visual characteristics of <i>Stemphylium</i> cultures.	32
3. 5 Conidial morphology of <i>Stemphylium</i> isolates on potato carrot agar (PCA).	36
3. 6 Morphology of conidiophores of <i>Stemphylium</i> isolates on potato carrot agar (PCA).	37
3. 7 Morphology of conidia and conidiophores of representative <i>Stemphylium</i> isolates (SSN-T03, SSN-E12, SSN-P05, SSN-L05) on naturally infected leaves.	38
3. 8 Comparison of the ITS region of all 46 <i>Stemphylium</i> isolates to the two close species <i>Stemphylium xanthosomatis</i> and <i>Stemphylium lycopersici</i> available in the GenBank.	41
3. 9 Comparison of the <i>gpd</i> gene of all 46 <i>Stemphylium</i> isolates to the two close species <i>Stemphylium xanthosomatis</i> and <i>Stemphylium lycopersici</i> available in the GenBank.	46
3. 10 Comparison of the <i>EF-1 alpha</i> gene of all 46 <i>Stemphylium</i> isolates to the two close species <i>Stemphylium xanthosomatis</i> and <i>Stemphylium lycopersici</i> available in the GenBank.	51

3. 11 Comparison of the *vmaA-vpsA* region of all 46 *Stemphylium* isolates to the two close species *Stemphylium xanthosomatis* and *Stemphylium lycopersici* available in the GenBank. 55
3. 12 Descriptive tree statistics of the ITS, *gpd*, *EF-1 alpha* and *vmaA-vpsA* and the combined regions. 61
3. 13 Cultural characteristics of four representative *Stemphylium lycopersici* isolates (SSN-T03, SSN-E12, SSN-P05 and SSN-L05) on different media. 65
3. 14 Conidial germination and length of germ tube of four representative *Stemphylium lycopersici* isolates (SSN-T03, SSN-E12, SSN-P05 and SSN-L05) on water agar (WA) as influenced by temperature. 68
3. 15 Sporulation of four representative *Stemphylium lycopersici* isolates (SSN-T03, SSN-E12, SSN-P05 and SSN-L05) produced on three culture media at 25 °C, after 7 days. 69
4. 1 Virulence variability of all 46 *Stemphylium lycopersici* isolates on their own host plants, including tomato, eggplant, pepper and lettuce using detached leaf assay. 78
4. 2 Cross-inoculation assays and percent disease severity of representative *Stemphylium lycopersici* isolates (SSN-T12, SSN-E06, SSN-P07 and SSN-L07) on different vegetable crops by artificial inoculation. 79
5. 1 RAPD and ISSR primers utilized to identify and assess interspecific genetic diversity among *Stemphylium lycopersici* isolates. 86

LIST OF FIGURES

Figure	Page
2. 1 Variability of conidia in <i>Stemphylium</i> species.	7
2. 2 Five (A-E) major groups of <i>Stemphylium</i> spp. according to an evolutionary relationship based on the combined dataset (glyceraldehyde-3-phosphate dehydrogenase and ITS1-5.8s-ITS2) from <i>Stemphylium</i> species.	12
3. 1 Positions of primers used for PCR amplification and DNA sequencing to generate <i>Stemphylium</i> phylogeny.	22
3. 2 Symptoms of <i>Stemphylium</i> leaf spot on tomato in an infected greenhouse, Tanah Rata, Pahang (A1, A2); eggplant in an infected field, Tangkak, Johor (B1, B2); pepper in an infected field, Tangkak, Johor (C); and lettuce in an infected greenhouse, Keama Farms, Pahang (D).	28
3. 3 Variability in colony morphology among <i>Stemphylium</i> isolates 10 days after incubation on potato dextrose agar (PDA) observed from top and bottom of the Petri dishes.	33
3. 4 Conidia of <i>Stemphylium</i> isolates obtained from A) tomato (SSN-T03), B) eggplant (SSN-E09), C) pepper (SSN-P01) and D) lettuce (SSN-L04). E) Conspicuously punctate on the conidia. F) Sporulation on top of vesicular tip of long conidiophores.	39
3. 5 Gel electrophoresis of PCR products from the ITS region (567 bp) of <i>Stemphylium</i> isolates.	40
3. 6 Nucleotide differences and their positions between four representative <i>Stemphylium</i> isolates (SSN-T03, SSN-E12, SSN-P05 and SSN-L05) and the two close species <i>Stemphylium lycopersici</i> and <i>Stemphylium xanthosomatis</i> in the ITS region.	43
3. 7 Phylogenetic tree based on sequence analysis of the ITS region of four representative <i>Stemphylium</i> isolates (SSN-T03, SSN-E12, SSN-P05 and SSN-L05) and compared to <i>Stemphylium</i> species available in the GenBank using the maximum parsimony method.	44
3. 8 Gel electrophoresis of PCR products from the <i>gpd</i> gene (591 bp) of <i>Stemphylium</i> isolates.	45

3. 9 Nucleotide differences and their positions between four representative *Stemphylium* isolates (SSN-T03, SSN-E12, SSN-P05 and SSN-L05) and the two close species *Stemphylium lycopersici* and *Stemphylium xanthosomatis* in the *gpd* gene. 48
3. 10 Phylogenetic tree based on sequence analysis of the *gpd* gene of four representative *Stemphylium* isolates (SSN-T03, SSN-E12, SSN-P05 and SSN-L05) and compared to *Stemphylium* species available in the GenBank using the maximum parsimony method. 49
3. 11 Gel electrophoresis of PCR products from the *EF-1 alpha* genes (572 bp) of *Stemphylium* isolates. 50
3. 12 Nucleotide differences and their positions between four representative *Stemphylium* isolates (SSN-T03, SSN-E12, SSN-P05 and SSN-L05) and the two close species *Stemphylium lycopersici* and *Stemphylium xanthosomatis* in the *EF-1 alpha* gene. 53
3. 13 Phylogenetic tree based on sequence analysis of the *EF-1 alpha* gene of four representative *Stemphylium* isolates (SSN-T03, SSN-E12, SSN-P05 and SSN-L05) and compared to *Stemphylium* species available in the GenBank using the maximum parsimony method. 54
3. 14 Gel electrophoresis of PCR products from the *vmaA-vpsA* region (685 bp) of *Stemphylium* isolates. 55
3. 15 Nucleotide differences and their positions between four representative *Stemphylium* isolates (SSN-T03, SSN-E12, SSN-P05 and SSN-L05) and the two close species *Stemphylium lycopersici* and *Stemphylium xanthosomatis* in the *vmaA-vpsA* region. 58
3. 16 Phylogenetic tree based on sequence analysis of the *vmaA-vpsA* region of four representative *Stemphylium* isolates (SSN-T03, SSN-E12, SSN-P05 and SSN-L05) and compared to *Stemphylium* species available in the GenBank using the maximum parsimony method. 59
3. 17 Phylogenetic tree based on combined dataset of the ITS, *gpd*, *EF-1 alpha* and *vmaA-vpsA* regions of representative *Stemphylium* isolates (SSN-T03, SSN-E12, SSN-P05 and SSN-L05) and compared to *Stemphylium* species available in the GenBank using the maximum parsimony method. 62

3. 18 Colony diameters of four representative *Stemphylium lycopersici* isolates (SSN-T03, SSN-E12, SSN-P05 and SSN-L05) on different media, 7 days after seeding with mycelial plug (5 mm). 64
3. 19 Growth rates of four representative *Stemphylium lycopersici* isolates (SSN-T03, SSN-E12, SSN-P05 and SSN-L05) on different media, 7 days after seeding with mycelial plug (5 mm). 64
3. 20 Colony morphology of representative *Stemphylium lycopersici* isolate SSN-L05 on different media. 66
3. 21 Mycelial growth of four representative *Stemphylium lycopersici* isolates (SSN-T03, SSN-E12, SSN-P05 and SSN-L05) as influenced by temperature, 7 days after seeding with mycelial plug (5 mm) on potato dextrose agar (PDA). 67
4. 1 Pathogenicity assay using *Stemphylium lycopersici* isolates in vitro conditions. Disease symptoms on A) tomato (SSN-T12), B) eggplant (SSN-E06), C) pepper (SSN-P06) and D) lettuce (SSN-L06). E) No symptom of the disease was observed on cabbage leaf as the non-host plant inoculated with representative isolate SSN-T12. 80
5. 1 Examples of polymorphisms with primers OPJ20 and OPY2 on 46 *Stemphylium lycopersici* isolates. 88
5. 2 UPGMA dendrogram generated by RAPD fingerprint analysis of *Stemphylium lycopersici* from different vegetable crops using a combination of seven primer sets (OPA-03, OPF-20, OPG-05, OPJ-20, OPJ-21, OPX-04 and OPY-02). 89
5. 3 Examples of polymorphisms with primers T3B and (ACA)₅ on 46 *Stemphylium lycopersici* isolates. 91
5. 4 UPGMA dendrogram generated by ISSR fingerprint analysis of *Stemphylium lycopersici* from different vegetable crops using a combination of four primer sets (M13, T3B, [ACA]₅ and [CCA]₅). 92

LIST OF ABBREVIATIONS

CRD	Complete Randomized Design
LSD	Least Significant difference
PDA	Potato Dextrose Agar
PCA	Potato Carrot Agar
WA	Water Agar
MEA	Malt Extract Agar
PSA	Potato Sugar Agar
V-8	V-8 juice agar
SDS	Sodium Dodecyl Sulfate
ddH ₂ O	Double Distilled Water
NUV	Near Ultraviolet
EB	Ethidium Bromide
v	Volt
EDTA	Ethylene Diamine Tetraacetic Acid
TAE	Tris Acetic Acid EDTA
TE	Tris EDTA
ng	Nanogram
pg	Picogram
fg	Festogram
rpm	Revolutions Per Minute
PCR	Polymerase Chain Reaction
rDNA	Ribosomal Deoxyribonucleic Acid
rRNA	Ribosomal Ribonucleic Acid
NCBI	National Center Biotechnology Information
ISSR	Inter Simple Sequence Repeat
RAPD	Random amplification of polymorphic DNA
ERIC	Enterobacterial Repetitive Intergenic Consensus
REP-PCR	Repetitive Extragenic Palindromes
ITS	Internal Transcribed Spacer
<i>gpd</i>	Glyceraldehyde-3-Phosphate Dehydrogenase
<i>EF-1 alpha</i>	Elongation Factor 1-Alpha Protein
BLASTn	Basic Local Alignment Search Tool Nucleotide



CHAPTER 1

INTRODUCTION

1.1 Importance and constraints of vegetable production

More than 10,000 vegetable species have been reported around the world (Decoteau, 2000). Vegetable crops have been always considered an important part of a healthy diet. Vegetable crops contain rich sources of vitamins and minerals (Prior and Guohua, 2000), and also significant amounts of bioactive compounds such as polyphenols, which provide a wide range of health benefits beyond basic nutrition. An association between high consumption of vegetable crops and a lower risk of chronic diseases such as certain types of cancer (Kwon *et al.*, 2006; Reddivari *et al.*, 2007; Rose *et al.*, 2005; Smiechowska *et al.*, 2008), type II diabetes (Liu *et al.*, 2004; Villegas *et al.*, 2008), and cardiovascular diseases (Radhika *et al.*, 2008; Terao *et al.*, 2008; Dauchet *et al.*, 2009), have consistently been indicated by epidemiological studies. Additionally, low consumption of vegetables and fruits were identified as an important risk factor for chronic diseases as reported by the World Health Organization World Health Report 2005 (Lock *et al.*, 2005). In Malaysia, a wide range of more than 50 types of vegetables grown by smallholders in both Highland and Lowland areas, have formed the important daily diet for the nation (Ding *et al.*, 1981). In 2006, the total area under vegetable cultivation in Malaysia was reported to be 39,660 ha, of which 11,415 and 10,498 ha was located in Johor (28.8 %) and Pahang (26.8 %), respectively (Anonymous, 2008).

Leaf spot caused by *Stemphylium* spp. is one of the important diseases of vegetable crops that attack the fruit, stem and foliage (Ellis, 1971). This disease is most common and severe on vegetables such as tomato, pepper, eggplant, lettuce, onion, garlic, spinach and asparagus (Farr and Rossman, 2014). Yield loss due to *Stemphylium* leaf spot can be high and in some cases is 100 % when proper disease control measures are not adopted (Cedeño and Carrero, 1997). The genus *Stemphylium*, anamorphic *Pleospora* (Dothideomycetes), was proposed by Wallroth (1833) with *S. botryosum* Wallr. as the type species. More than 33 species have been recognized in this genus (Câmara *et al.*, 2002). Most of *Stemphylium* species are saprophytes growing on dead plants and cellulose materials (Ellis, 1971, Simmons, 1969), but various *Stemphylium* species such as *S. botryosum*, *S. solani*, *S. lycopersici* and *S. vesicarium* have been reported as common pathogens on vegetables causing severe disease in almost all regions where these crops are grown (Ellis, 1971).

Unfortunately to date, there is no study on the causal agent of *Stemphylium* leaf spot of vegetable crops in Malaysia. Therefore, studies on *Stemphylium* species associated with leaf spot of vegetable crops in Malaysia are addressed in the present study. Additionally, genetic variability of *Stemphylium* populations obtained from various vegetables and different geographical regions was investigated. The results of this study will provide better insight into the biology and etiology of the pathogen and these achievements will be helpful in the development of better disease management strategies and in the breeding of resistant varieties for vegetable production in Malaysia.

1.2 Objectives of study

The present study was aimed at identifying and characterizing the biology and diversity of *Stemphylium* isolates from various vegetable crops collected from different geographical regions of Malaysia. The objectives of this study were to: (1) identify the causal agent of leaf spot of vegetable crops, and assess phenotype variability among monoconidial isolates using morphological and cultural characteristics; (2) evaluate the pathogenicity and cross-inoculation of *Stemphylium* isolates on host plants; (3) confirm the morphological studies, and provide a taxonomic position of *Stemphylium* isolates at the species level using the ITS, *gpd*, *EF-1 alpha* and *vmaA-vpsA* sequence comparisons; and (4) estimate the genetic variability among *Stemphylium* isolates associated with the various vegetable crops using randomly amplified polymorphic DNA (RAPD) and Internal simple sequence repeats (ISSR) molecular markers.

REFERENCES

- Achenbach, L. A., Patrick, J. A., & Gray, L. E. (1997). Genetic homogeneity among isolates of *Fusarium solani* that cause soybean sudden death syndrome. *Theoretical and Applied Genetics*, 95, 474-478.
- Altschul, S. F., Madden, T. L., Schäffer, A. A., Zhang, J., Zhang, Z., Miller, W., & Lipman, D. J. (1997) Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acids Research*, 25, 3389-3402.
- Alves, A., Crous, P. W., Correia, A., & Phillips, A. J. (2008). Morphological and molecular data reveal cryptic speciation in *Lasiodiplodia theobromae*. *Fungal Diversity*, 28, 1-13.
- Andersen, B., Solfrizzo, M., & Visconti, A. (1995). Metabolite profiles of common *Stemphylium* species. *Mycological Research*, 99(6), 672-676.
- Anonymous. (2008). Data keluasaan bertanam dan pengeluaran sayur, tanaman ladang dan rempah ratus mengikut jenis atau negeri. DOA statistic 2008.
- Antonio, A. F., Garcia, L. L., Benchimol, A. M., Barbosa, I. O., Geraldi, C. L., Souza Jr, C. L. & Souza, A. P. (2004). Comparison of RAPD, RFLP, AFLP and SSR markers for diversity studies in tropical maize inbred lines. *Genetic and Molecular Biology*, 27, 579-588.
- Atan, S., & Hamid, N.H. (2003). Differentiating races of *Corynespora cassicola* using RAPD and internal transcribed spacer markers. *Journal of Rubber Research*, 6(1), 58-64.
- Aveling, T. A. S. (1992). First report of *Stemphylium vesicarium* on garlic in South Africa. *Plant Disease*, 76(1), 426.
- Aveling, T. A. S., & Snyman, H. G. (1993). Evaluation of seed treatments for reducing *Alternaria porri* and *Stemphylium vesicarium* on onion seed. *Plant Disease*, 77(2), 1009-1011.
- Banke, S., Peschon, A., & McDonald, B. A. (2004). Phylogenetic analysis of globally distributed *Mycosphaerella graminicola* populations based on three DNA sequence loci. *Fungal Genetics and Biology*, 41, 226-238.

- Barash, I., Karr, A. L. J., & Strobel, G. A. (1975). Isolation and characterization of stemphylin, a chromone glucoside from *Stemphylium botryosum*. *Plant Physiology*, 55(4), 646-651.
- Barondes, S. H. (2000). Basic concepts and techniques of molecular genetics. <http://www.acnp.org>.
- Basset, I. J., Crompton, C.W., & Parmelee, J. A. (1978). Common airborne fungus spores. In: Basset, I. J., Crompton, C. W., Parmelee, J. A. (Eds.), *An atlas of airborne pollen grains and common fungus spores of Canada*, Quebec, Agriculture Canada, pp. 269-315.
- Batista, D. C., Lima, M. A., Haddad, F., Maffia, L. A., & Mizubuti, E. S. G. (2006). Validation of decision support systems for tomato early blight and potato late blight, under Brazilian conditions. *Crop Protection*, 25, 664-670.
- Berbee, M. L., Pirseyedi, M., & Hubbard, S. (1999). *Cochliobolus* phylogenetics and the origin of known, highly virulent pathogens, inferred from ITS and glyceraldehyde-3-phosphate dehydrogenase gene sequences. *Mycologia*, 91, 964-977.
- Blancard, D. (1992). A colour Atlas of tomato diseases. Wolfe Publishing Ltd., Montfavet, France, 210 p.
- Bradley, D. J., Gregory, S. G., & Parker, I. M. (2003). Susceptibility of clover species to fungal infection: the interaction of leaf surface traits and environment. *American Journal of Botany*, 6(90): 857-864.
- Callac, P., & Guinberteau, J. (2005). Morphological and molecular characterization of two novel species of *Agaricus* section *Xanthodermatei*. *Mycologia*, 97, 416-424.
- Câmara, M. P. S, O'Neill, N. R., & van Berkum, P. (2002). Phylogeny of *Stemphylium* spp. based on ITS and glyceraldehyde-3-phosphate dehydrogenase gene sequences. *Mycologia*, 94, 660-672. doi:10.2307/3761717.
- Carter, J., Rezanoor, H., Holden, D., Desjardins, A., Plattner, R., & Nicholson, P. (2002). Variation in pathogenicity associated with the genetic diversity of *Fusarium graminearum*. *European Journal of Plant Pathology*, 108(6), 573-583.

- Cedeño L., & Carrero C. (1997). First report of tomato gray leaf spot caused by *Stemphylium solani* in the Andes region of Venezuela. *Plant Disease*, 81(11), 1332-1332.
- Chaisrisook, C., Skinner, D. Z., & Stuteville, D. L. (1995). Molecular genetic relationships of five *Stemphylium* species pathogenic to alfalfa. *Sydowia*, 47, 1-9.
- Cho, H. J., Kim, B. S. & Hwang, H. S. (2001). Resistance to gray leaf spot in Capsicum pepper. *HortScience*, 36, 753-754.
- Coles, R. B., & Wicks, T. J. (2001). New fungal pathogens of carrots in South Australia. Proceedings of 13th Biennial-Plant Pathology Conference. Queensland Government Department of Primary Industries, Australia, pp. 103.
- Darmono, T. W., Darussamin, A., & Pawirosoemardjo, S. (1996). Variation among isolates of *Corynespora cassicola* associated with Hevea brasiliensis in Indonesia. In: Proceeding workshop on Corynespora leaf fall disease of Hevea rubber Medan, Indonesia, pp. 79-91.
- Dauchet, L., Amouyel, P., & Dallongeville, J. (2009). Fruits, vegetables and coronary heart disease. *Nature Reviews Cardiology*, 6(9), 599-608.
- Decoteau, D. R. (2000). Vegetable Crops. Charles E. Stewart, Jr., Upper Saddle River, pp. 32-38.
- Deng, J. X., Paul, N. Ch., Park, M. S., & Yu, S. H. (2013). Molecular characterization, morphology, and pathogenicity of *Alternaria panax* from araliaceous plants in Korea. *Mycological Progress*, (In press) doi: 10.1007/s11557-012-0844-8.
- Ding, T. H., Vimala, P., & Yusof, S. (1981). A special report on an agro-economic survey of vegetable farming in Peninsular Malaysia. Serdang: Malaysia Agricultural Research and Development institute.
- DOA, (2011). Annual Report. <http://www.doa.gov.my/>.
- du Toit, L. J., & Derie, M. L. (2002). Leaf spot of spinach seed crops in Washington State. *Phytopathology*, 92, S21.

- du Toit, L. J. & Derie, M. L. (2003). Leaf spot of spinach: Research results for 2002. In: Proc. 92nd Ann. Mtg. Western WA Hort. Assoc., Sea Tac, WA, pp. 61-64
- Ellis, M. B. & Gibson, I. A. S. (1975a). *Stemphylium lycopersici*. CMI Descriptions of Pathogenic Fungi and Bacteria, No. 471.
- Ellis, M. B. & Gibson, I. A. S. (1975b). *Stemphylium solani*. CMI Descriptions of Pathogenic Fungi and Bacteria, No. 472.
- Ellis, M. B. (1971). Dematiaceous hyphomycetes. Commonwealth Mycological Institute, Kew, Surrey, UK, 608 p.
- Elmer, W. H. 2001. The economically important diseases of asparagus in the U.S. Online. Plant Health Progress. doi:10.1094/PHP-2001-0521-01-RV.
- Emelyanov, V. V. (2003). Mitochondrial connection to the origin of the eukaryotic cell. *European Journal of Biochemistry*, 270, 1599-1618.
- Farr, D. F., & Rossman, A. Y. (2014). Fungal databases, systematic mycology and microbiology laboratory, ARS, USDA. Retrieved January 12, from <http://nt.ars-grin.gov/fungaldatabases/>.
- Goldwin, I. D., Aitken, A. B., & Smith, L. W. (1997). Application of Inter simple sequence repeats (ISSR) markers to plant genetics. *Electrophoresis*, 18, 1524-8. doi:10.1002/elps.1150180906.
- Gonzalez-Mendoza, D., Argumedo-Delira, R., Morales-Trejo, A., Pulido-Herrera, A., Cervantes-Diaz, L., Grimaldo-Juarez, O., & Alarcon, A. (2010). A rapid method for isolation of total DNA from pathogenic filamentous plant fungi. *Genetics and Molecular Research*, 9, 162-6. doi: 10.4238/vol9-1gmr680.
- Goyal, P., Chahar, M., Mathur, A. P., Kumar, A., & Chattopadhyay, C. (2011). Morphological and cultural variation in different oilseed *Brassica* isolates of *Alternaria brassicae* from different geographical regions of India. *Indian Journal of Agricultural Sciences*, 81 (11), 1052-8.
- Hall, T. A. (1999). BioEdit: A user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symposium Series*, 41, 95-98.

- Hausbeck, M. K. (2011). Purple spot disease of asparagus: Cool, wet growing conditions favor purple spot on asparagus spears. <http://www.msue.msu.edu>.
- Hosen, M. I., Ahmed, A. U., Zaman, J. J., Ghosh, S., & Hossain, K. M. K. (2009). Cultural and physiological variation between isolates of *Stemphylium botryosum* the causal of Stemphylium blight disease of lentil (*Lens culinaris*). *World Journal of Agricultural Sciences*, 5, 94-98.
- Howard, R. J., Garland, J. A., & Seaman, W. L. (1994). Pea and bean. In: Howard, R. J., Garland, J. A., & Seaman, W. L. (Ed.). Diseases and pests of vegetable crops in Canada: an illustrated compendium. Ottawa, Canadian Phytopathological Society, pp. 202-210.
- Hughes, S. J. (1953). Conidiophores, conidia and classification. *Canadian Journal of Botany*, 31, 577-659.
- Huguenin, B. (1965). Dematiées de Nouvelle-Calédonie. *Bulletin trimestriel de la Société mycologique de France*, 81, 696-697.
- Inderbitzin, P., Harkness, J., Turgeon, B. G., & Berbee, M. L. (2005). Lateral transfer of mating system in *Stemphylium*. *Proceedings of the National Academy of Sciences, USA*, 102, 11390-11395.
- Inderbitzin, P., Mehta, Y. R., & Berbee, M. L. (2009). *Pleospora* species with *Stemphylium* anamorphs: a four locus phylogeny resolves new lineages yet does not distinguish among species in the *Pleospora herbarum* clade. *Mycologia*, 101, 329-339.
- Innis, M. A., Gelfand, D. H., Snisky, J. J., & White T. J. (Ed.) (1990). PCR protocols In: A guide to methods and applications. Academic Press. San Diego.
- Kim, B. S., Cho, H. J., Hwang, H. S. & Cha, Y. S. (1999). Gray leaf spot of tomato caused by *Stemphylium solani*. *The Plant Pathology Journal*, 15, 348-350.
- Kim, B. S., Yu, S. H., Cho, H. J., & Hwang, H. S. (2004). Gray leaf spot in peppers caused by *Stemphylium solani* and *Stemphylium lycopersici*. *The Plant Pathology Journal*, 20, 85-91.
- Kistler, H. (1997). Genetic diversity in the plant-pathogenic fungus *Fusarium oxysporum*. *Phytopathology*, 87(4), 474-479.

- Köhl, J., Groenenboom-de Haas, B., Goossen-van de Geijn, H., Speksnijder, A., Kastelein, P., de Hoog, S., & van den Ende, B. G. (2009). Pathogenicity of *Stemphylium vesicarium* from different hosts causing brown spot in pear. *European Journal of Plant Pathology*, 124(1), 151-162.
- Koike, S. T., Henderson, D., & Butler, E. (2001). Host specific strain of *Stemphylium* cause leaf spot disease of California spinach. *California Agriculture*, 55(5), 31-34.
- Kulik, M. M. (1995). Seed quality and microorganisms. In: Basra AS ed. Seed quality- basic mechanisms and agricultural implications. New York, Food Products Press, pp. 153-173.
- Kumar, M., & Shukla, P. K. (2005). Use of PCR targeting of internal transcribed spacer regions and single-stranded conformation polymorphism analysis of sequence variation in different regions of rRNA genes in fungi for rapid diagnosis of mycotic keratitis. *Journal of Clinical Microbiology*, 43, 662-668.
- Kwon, D., Yoon, S., Carter, O., Bailey, G. S., & Dashwood, R. H. (2006). Antioxidant and antigenotoxic activities of *Angelica keiskei*, *Oenanthe javanica* and *Brassica oleracea* in the Salmonella mutagenicity assay and in HCT116 human colon cancer cells. *BioFactors*, 26(4):231-244.
- Lamprecht, S. C., & Knox-Davies, P. S. (1984). *Stemphylium vesicarium* on lucerne (*Medicago sativa*) seeds in South Africa. *Phytophylactica*, 16, 189-193.
- Leach, C. M. (1962). Sporulation of diverse species of fungi under near-ultraviolet radiation. *Canadian Journal of Botany*, 40, 151-161.
- Leach, C. M., & Aragaki, M. (1970). Effect of temperature on conidium characteristics of *Ulocladium chartarum* and *Stemphylium floridanum*. *Mycologia*, 62,1071-1076.
- Ligero, A. M. P, Andujar, J. L. G, Vara, J. M. M., & Ureba, M. J. U. (1998). Development of *Pleospora allii* on garlic debris infected by *Stemphylium vesicarium*. *European Journal of Plant Pathology*, 104, 861-870.
- Liu, S., Serdula, M., Janket, S. J., Cook, N. R., Sesso, H. D., Willett, W. C., Manson, J. E., & Buring, J. E. (2004). A prospective study of fruit and

vegetable intake and the risk of type 2 diabetes in women. *Diabetes Care*, 27(12), 2993-2996.

Lock, K., Pomerleau, J., Causer, L., Altmann, D. R., & McKee, M. (2005). The global burden of disease attributable to low consumption of fruit and vegetables: implications for the global strategy on diet. *Bulletin of the World Health Organization*, 83, 100-108.

Lourenço Jr, V., Rodrigues, T. T., Campos, A., Bragança, C. A., Scheuermann, K. K., Reis, A., & Mizubuti, E. S. (2011). Genetic structure of the population of *Alternaria solani* in Brazil. *Journal of Phytopathology*, 159(4), 233-240.

Malloch, D. (1981). *Moulds: their isolation, cultivation and identification*. Toronto: University of Toronto Press, 97 p.

Mathur, S. B., & Kongsdal, O. (2002). *Common laboratory seed health testing methods for detecting fungi*. Frederiksberg: Danish Government Institute of Seed Pathology for Developing Countries (DGSISP), 427 p.

McDermott, J. M., & McDonald. B. A. (1993). Gene flow in plant pathosystems. *Annual Review of Phytopathology*, 31, 353-373.

McDonald, B. A. & Linde, C. (2002). Pathogen population genetics, evolutionary potential, and durable resistance. *Annual Review of Phytopathology*, 40, 349-379.

McDonald, B. A. (2004). *Population genetics of plant pathogens. The plant health instructor*. American Phytopathological Society, St Paul, Minnesota, USA. doi:10.1094/PHI-A-2004-0524-01.

Mehta, Y. R. (2001). Genetic diversity among isolates of *Stemphylium solani* from cotton. *Fitopatologia Brasileira*, 26(4), 703-709.

Mehta, Y. R., & Mehta, A. (2010). Genetic variability among the isolates of *Colletotrichum gossypii* of cotton. *Summa Phytopathologica*, 36(1), 40-44.

Mehta, Y. R., Mehta, A., & Rosato, Y. B. (2002). ERIC and REP-PCR banding patterns and sequence analysis of the internal transcribed spacer of rDNA of *Stemphylium solani* isolates from cotton. *Current microbiology*, 44(5), 323-328. doi: 10.1007/s00284-001-0026-4.

- Milgroom, M. G., & Peever, T. L. (2003). Population biology of plant pathogens. The synthesis of plant disease epidemiology and population genetics. *Plant Disease*, 87, 608-617.
- Min, J. Y., Kim, B. S., Cho, K. W., & Yu, S. H. (1995). Grey leaf spot caused by *Stemphylium lycopersici* on tomato plants. *Korean Journal of Plant Pathology* 11, 282-284.
- Mitchell, J. L., Roberts, P. J., & Moss, S. T. (1995). Sequence or structure? A short review on the application of Nucleic acid sequence information to fungal taxonomy. *Mycologist* 9, 67-75.
- Mogri, M. (2000). BioEditor. <http://bioeditor.sdsc.edu>.
- Moore, D., & Frazer, L. A. N. (2002). Systematics, phylogeny and evolution. In: Moore D Frazer LAN eds. Essential fungal genetics. New York, Springer, pp. 245-281.
- Moralejo, E., Clemente, A., Descals, E., Belbahri, L., Calmin, G., Lefort, F., Spies, C. F. J., & McLeod, A. (2008). *Pythium recalcitrans* sp. nov. revealed by multigene phylogenetic analysis. *Mycologia*, 100, 310-319.
- Mueller, U. G., Lipari, S. E., & Milgroom, M. G. (1996). Amplified fragment length polymorphism (AFLP) fingerprinting of symbiotic fungi cultured by the fungus-growing ant *Cyphomyrmex* minutes. *Molecular Ecology*, 5, 119-122.
- Neergaard, P. (1945). Danish species of *Alternaria* and *Stemphylium* - taxonomy, parasitism and economical significance. London, UK: Oxford University Press, 560 p.
- Nghia, N. A., Kadir, J., Sunderasan, E., Abdullah, M. P., Malik, A., & Napis, S. (2008). Morphological and inter simple sequence repeat (ISSR) markers analyses of *Corynespora cassiicola* isolates from rubber plantations in Malaysia. *Mycopathologia*, 166(4), 189-201.
- Okori, P., Rubaihayo, P. R., Adipala, E., Fahleson, J., & Dixelius, C. (2004). Population studies of fungal plant pathogens: Perspectives for control with specific reference to grey leaf spot. *African Crop Science Journal*, 12(4): 327-342.

- Oliver, R. (1993). Nucleic acid-based methods for detection and identification. In: Fox RTV ed. Principles of diagnostic techniques in plant pathology. Warlingford, CAB International, pp. 153-170.
- Park, M. S., Romanoski, C. E., & Pryor, B. M. (2008). A re-examination of the phylogenetic relationship between the causal agents of carrot black rot, *Alternaria radicina* and *A. carotiincultae*. *Mycologia*, 100(3), 511-527.
- Pei, Y. F., Geng, Y., Wang, Y., & Zhang, X. G. (2009). Two new species of *Stemphylium* from Sinkiang, China. *Mycotaxon*, 109, 493-497.
- Prior, R., & Guohua, C. (2000). Analysis of botanical and dietary supplements for antioxidant capacity: A review. *Journal of AOAC International*, 83, 950-956.
- Pryor, B. M., & Gilbertson, R. L. (2000). Molecular phylogenetic relationships amongst *Alternaria* species and related fungi based upon analysis of nuclear ITS and mtSSU-rDNA sequences. *Mycological Research*, 104, 1312-1321.
- Pryor, B. M., & Michailides, T. J. (2002). Morphological, pathogenic, and molecular characterization of *Alternaria* isolates associated with *Alternaria* late blight of pistachio. *Phytopathology*, 92, 406-416.
- Radhika, G., Sudha, V., Mohan Sathya, R., Ganesan, A., & Mohan, V. (2008). Association of fruit and vegetable intake with cardiovascular risk factors in urban south Indians. *British Journal of Nutrition*, 99(2):398-405.
- Raid, R., & Kucharek, T. (2003). Florida Plant Management Guide: Spinach. <http://edis.ifas.ufl.edu>.
- Reddivari, L., Hale, A. L., & Miller Jr, J. C. (2007). Genotype, location, and year influence antioxidant activity, carotenoid content, phenolic content, and composition in specialty potatoes. *Journal of Agricultural and Food Chemistry*, 55(20), 8073-8079.
- Rohlf, E. J. (1993). NTSYS-pc: Numerical taxonomy and multivariate analysis system, version 1.80. Applied Biostatistics Inc., Setauket, New York.
- Romruensukharom, P., Tragoonrung, S., Vanavichit, A., & Toojinda, T. (2005). Genetic variability of *Corynespora cassiicola* population in Thailand. *Journal of Rubber Research*, 8(1), 38-49.

- Rose, P., Huang, Q., Ong, C. N., & Whiteman, M. (2005). Broccoli and watercress suppress matrix metalloproteinase-9 activity and invasiveness of human MDAMB-231 breast cancer cells. *Toxicology and Applied Pharmacology*, 209(2), 105-113.
- RuiQian, L., Rui, H., YueBing, Z., YuMei, X., & JianMing, W. (2009). Establishment of ISSR reaction system of *Fusarium* and its analysis of genetic diversity. *Scientia Agricultura Sinica*, 42(9), 3139-3146.
- Schlotterer, C. (2004). The evolution of molecular markers - just a matter of fashion? *Nature*, 5, 63-69.
- Sharma, P., Deep, S., Sharma, M., & Bhati, D. S. (2013). Genetic variation of *Alternaria brassicae* (Berk.) Sacc., causal agent of dark leaf spot of cauliflower and mustard in India. *Journal of General Plant Pathology*, 79(1), 41-45.
- Shivji, M. (1997). Marine species molecular database. <http://www.nova.edu>.
- Silva, W. P. K., Deverall, B. J., & Lyon, B. R. (1998). Molecular, physiological and pathological characterization of *Corynespora* leaf spot fungi from rubber plantations in Sri Lanka. *Plant Pathology*, 47(3), 267-77. doi:10.1046/j.1365-3059.1998.00245.x.
- Silva, W. P. K., Karunanayake, E. H., Wijesundera, R. L. C., & Priyanka, U. M. S (2003). Genetic variation in *Corynespora cassiicola*: a possible relationship between host origin and virulence. *Mycological Research*, 107(5), 567-71. doi:10.1017/S0953756203007755.
- Simmons, E. G. (1967). Typification of *Alternaria*, *Stemphylium* and *Ulocladium*. *Mycologia*, 59, 67-92.
- Simmons, E. G. (1969). Perfect states of *Stemphylium*. *Mycologia*, 61, 1-26.
- Simmons, E. G. (1985). Perfect states of *Stemphylium* II. *Sydowia*, 38, 284-293.
- Simmons, E. G. (1989). Perfect states of *Stemphylium* III. *Memoirs of the New York Botanical Garden*, 49, 305-307.
- Simmons, E. G. (1990). *Alternaria* themes and variations (27-53). *Mycotaxon*, 37, 79-119.

- Simmons, E. G. (2001). Perfect states of *Stemphylium* IV. *Harvard Pap. Bot.*, 6, 199-208.
- Simmons, E. G. (2002). *Alternaria* themes and variations (287-304)- species on Caryophyllaceae. *Mycotaxon*, 82, 1-40.
- Simmons, E. G. (2004). Novel dematiaceous hyphomycetes. *Studies in Mycology*, 50, 109-118.
- Simmons, E. G., & Roberts, R. G. (1993). *Alternaria* themes and variations (73). *Mycotaxon*, 48, 109-140.
- Smiechowska, A., Bartoszek, A., & Namiesnik, J. (2008). Cancer chemopreventive agents: glucosinolates and their decomposition products in white cabbage (*Brassica oleracea* var. capitata). *Postępy Higieny i Medycyny Doświadczalnej*, 62, 125-140.
- Smith, T. L. (1989). Disparate evolution of yeasts and filamentous fungi indicated by phylogenetic analysis of glyceraldehyde-3-phosphate dehydrogenase genes. *Proceedings of the National Academy of Sciences of the United States of America*, 86, 7063-7066.
- Sofi, T., Beig, M. A., Dar, G. H., Ahmad, M., Hamid, A., Ahangar, F., Padder, B. A., & Shah, M. D. (2013). Cultural, morphological, pathogenic and molecular characterization of *Alternaria mali* associated with *Alternaria* leaf blotch of apple. *African Journal of Biotechnology*, 12 (4), 370-381.
- Sotome, K., Hattori, T., Ota, Y., Lee, S.S., Vikineswary, S., Abdullah, N., & Kakishima, M. (2009). Taxonomic study of Asian species of Echinochaete (Polyporaceae, Basidiomycota) and description of *E. maximipora* sp. nov. *Mycological Progress*, 8, 123-132.
- Stuville, D. L., & Erwin, D. C. (1990). Compendium of alfalfa diseases. Minnesota: APS Press, 84 p.
- Sujatha, M., Prabakaran, A. J., & Chattopadhyay, C. (1997). Reaction of wild sunflowers and certain interspecific hybrids to *Alternaria helianthi*. *Helia*, 20, 15-24.
- Syndir, M., & Lacoste, L. (1994). Morphological and physiological aspects of *Stemphylium* leaf spot of the scarlet eggplant (*Solanum aethiopicum* L.) from Senegal [Abstract]. *Bulletin de l'Institut Fondamental d'Afrique Noire*, 47(33), 33-41.

- Tamura, K., Peterson, D., Peterson, N., Stecher, G., Nei, M., & Kumar S. (2011). MEGA5: Molecular Evolutionary Genetics Analysis Using Maximum Likelihood, Evolutionary Distance, and Maximum Parsimony Methods. *Molecular Biology and Evolution*, 28, 2731-2739. doi:10.1093/molbev/msr121.
- Terao, J., Kawai, Y. & Murota, K. (2008). Vegetable flavonoids and cardiovascular disease. *Asia Pac. J. Clin. Nutr.* 17 Suppl. 1, 291-293.
- Thompson, J. D., Higgins, D. G., & Gibson, T. J. (1994). Clustal W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Research*, 22, 4673-4680.
- van der Waals, J. E., Korsten, L., & Slippers, B. (2004). Genetic diversity among *Alternaria solani* isolates from potatoes in South Africa. *Plant Disease*, 88(9), 959-964.
- Villegas, R., Shu, X. O., Gao, Y. T., Yang, G., Elasy, T., Li, H., & Zheng, W. (2008). Vegetable but not fruit consumption reduces the risk of type 2 diabetes in Chinese women. *Journal of Nutrition*, 138(3), 574-580.
- Wallroth, F. G. (1833). *Flora Cryptogamica Germaniae*, pars. post. Schrag, Nürnberg.
- Wang, Y., & Zhang, X. G. (2006). Three new species of *Stemphylium* from China. *Mycotaxon*, 96, 77-8.
- Wang, Y., & Zhang, X. G. (2009). Two new species of *Stemphylium* from Shandong, China. *Nova Hedwig*, 88, 199-203.
- Wang, Y., Fu, H. B., O'Neill, N. R., & Zhang, X. G. (2009). Two new species of *Stemphylium* from Northwest China. *Mycological Progress*, 8, 301-304.
- Wang, Y., Geng, Y., Pei, Y. F., & Zhang, X. G. (2010). Molecular and morphological description of two new species of *Stemphylium* from China and France. *Mycologia*, 102, 708-717.
- Weber, B., & Halterman, D. A. (2012). Analysis of genetic and pathogenic variation of *Alternaria solani* from a potato production region. *European Journal of Plant Pathology*, 134(4), 847-858.

- Weber, G. F. (1930). Gray leaf spot of tomato caused by *Stemphylium solani* sp. nov. *Phytopathology*, 20, 513-518.
- White, T. J., Bruns, T., Lee, S., & Taylor, J. (1990). Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis, M. A., Gelfand, D. H., Sninsky, J. J., White, T. J., editors; Innis, M. A., Gelfand, D. H., Sninsky, J. J., & White, T. J., editors. PCR protocols: a guide to methods and applications. New York, N.Y: Academic Press, Inc. pp. 315-322.
- Wyk, M. V., Wingfield, B. D., Clegg, P. A., & Wingfield, M. J. (2009). *Ceratocystis larium* sp. nov., a new species from *Styrax benzoin* wounds associated with incense harvesting in Indonesia. *Persoonia*, 22, 75-82.
- Xue, F., Zhang, X. G., Wang, Y., & Wang, H. Z. (2005). Taxonomic studies of *Stemphylium* from China II. *Stemphylium subglobuliferum* sp. nov., and four new records. *Mycosystema*, 24, 322-329.
- Yamamoto, W. (1960). Synonymous species of *Alternaria* and *Stemphylium* in Japan. *Transactions of the Mycological Society of Japan*, 2, 88-93.
- Yu, S. H. (2001). Korean species of *Alternaria* and *Stemphylium*. Suwon: National Institute of Agricultural Science and Technology, pp. 159-166.
- Zhan, J., Kema, G. H. J. & McDonald, B. A. (2004). Evidence for natural selection in the mitochondria genome. *Phytopathology*, 94, 261-267.
- Zhang, X. G., & Zhang, T. Y. (2002). Taxonomic studies of *Stemphylium* from China I. *Mycosystema*, 21, 324-326.
- Zhang, X. G., & Zhang, T. Y. (2007). Taxonomic studies of *Stemphylium* from China III: four new species on plants of several families. *Mycosystema*, 26, 477-483.
- Zhang, X. G., Wu, Y. M., & Zhang, T. Y. (2003). Taxonomic studies of *Stemphylium* from China. *Mycotaxon*, 85, 247-252.
- Zheng, L., Huang, J., & Hsiang, T. (2008). First report of leaf blight of garlic (*Allium sativum*) caused by *Stemphylium solani* in China. *Plant Pathology*, 57, 380. doi: 10.1111/j.1365-3059.2007.01724.x.

Zheng, L., Lv, R.J., Hsiang, T., & Huang, J.B. (2009). Host range and phytotoxicity of *Stemphylium solani*, causing leaf blight of garlic (*Allium sativum*) in China. *European Journal of Plant Pathology*, 124, 21-30.

Zhou, S., Smith, D. R., & Stanosz, G. R. (2001). Differentiation of *Botryosphaeria* species and related anamorphic fungi using Inter Simple or Short Sequence Repeat (ISSR) fingerprinting. *Mycological Research*, 105(8), 919-926.

