

**MICROPROPAGATION OF TONGKAT ALI  
(*EURYCOMA LONGIFOLIA* JACK) VIA SOMATIC EMBRYOGENESIS  
AND DIRECT PLANT REGENERATION TECHNIQUES**

**By**

**SOBRI BIN HUSSEIN**

**Thesis Submitted to the School of Graduate Studies,  
Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree  
of Doctor of Philosophy**

**October 2004**

***Especially dedicated to:***

**My parents, Brothers, Sisters  
&  
Dr. Anna Ling**

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirements for the Degree of Doctor of Philosophy

**MICROPROPAGATION OF TONGKAT ALI  
(*EURYCOMA LONGIFOLIA* JACK) VIA SOMATIC EMBRYOGENESIS  
AND DIRECT PLANT REGENERATION TECHNIQUES**

By

**SOBRI BIN HUSSEIN**

October 2004

**Chairman: Professor Maziah Mahmood, Ph.D.**

**Faculty: Biotechnology and Biomolecular Sciences**

*Eurycoma longifolia* Jack or Tongkat Ali is well known among the local communities mainly for its aphrodisiac properties and its effectiveness as the cytotoxic, anti-malarial, anti-ulcer, anti-tumor promoting and anti-parasitic agent. In view of its potential commercial value as a plantation crop as well as to conserve its germplasm, the somatic embryogenesis and direct plant regeneration of *E. longifolia* were carried out as these *in vitro* micropropagation protocols had not been reported.

In attempts to establish the somatic embryos of *E. longifolia*, the potential of cotyledon, zygotic embryo, leaf, petiole, stem and taproot in forming embryogenic callus were examined in the basal Murashige and Skoog (MS) medium supplemented with different auxins at various concentrations. Only cotyledon explants were able to form embryogenic callus in the presence of 1.0 mg/L (w/v) of 2,4-dichlorophenoxyacetic (2,4-D) at 30%. A higher yield (60%) of

embryogenic callus was obtained when the Type 4 method dissected cotyledon explants were cultured in basal MS medium containing 0.5 mg/L (w/v) of kinetin and 1.0 mg/L (w/v) of 2,4-D. The highest number of somatic embryos ( $45 \pm 2$ ) was observed in the same medium formulation with the addition of 1.0 g/L (w/v) activated charcoal. Subsequent transfer of these mature somatic embryos in basal MS media supplemented with 1.0 mg/L (w/v) of kinetin produced a 90% of plantlet regeneration. Addition of activated charcoal, casein hydrolysate, abscisic acid, proline and polyethylene glycol (PEG) at various concentrations into the regeneration medium did not stimulate the conversion of *E. longifolia* somatic embryo into plantlet. The differences between the embryogenic and non-embryogenic callus were also determined based on histological studies.

Successful direct plant regeneration was obtained from the root, stem, shoot tip, axillary and adventitious bud explants. Each explant generally requires different combinations of media and plant growth regulators to produce the highest regeneration percentage. In the root explant, the best medium formulation determined was basal Juglans medium (DKW) supplemented with 1.0 mg/L (w/v) of kinetin + 1.0 mg/L (w/v) of zeatin whereas in the stem explants, basal woody plant medium (WPM) enriched with 2.0 mg/L (w/v) of BAP and 2.0 mg/L (w/v) of zeatin was found to be the best medium formulation in increasing the regeneration rate and healthy plantlets formation. Stem and root explants that were 2 cm distant from one another as has been identified as the most suitable position for attaining the maximum percentage of direct plant regeneration. Successful direct plant regeneration from *in vitro* and *in vivo* shoot tip explants of

*E. longifolia* was achieved in basal MS medium supplemented with 3.0 and 5.0 mg/L (w/v) of kinetin, respectively. As for the direct plant regeneration from axillary bud explant, explants produced the highest regeneration capability (90%) in basal Nitsch medium (NM) supplemented with 10.0 mg/L (w/v) of zeatin while basal NM supplied with 6.0 mg/L (w/v) of zeatin produced the highest regeneration percentage in adventitious bud explants. Rooting induction of *in vitro* plantlets of *E. longifolia* was also achieved and the highest induction rate was attained in basal MS medium supplemented with 0.5 mg/L (w/v) of Indole-3-butyric acid (IBA). Acclimatization of *in vitro* plantlets regenerated from somatic embryogenesis and direct plant regeneration survived well with no morphological differences from the parent plants after two months of transplantation to the soil.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Doktor Falsafah

**MCROPROPAGASI TONGKAT ALI  
(*EURYCOMA LONGIFOLIA* JACK) MELALUI KAEDAH SOMATIK  
EMBRIOGENESIS DAN REGENERASI SECARA TERUS**

Oleh

**SOBRI BIN HUSSEIN**

**Oktober 2004**

**Pengerusi : Profesor Maziah Mahmood, Ph.D.**

**Fakulti : Bioteknologi dan Sains Biomolekul**

*Eurycoma longifolia* Jack atau Tongkat Ali lebih dikenali oleh penduduk tempatan kerana ianya mempunyai ciri afrodisiak dan keberkesanannya sebagai sitotoksik, anti malaria, anti ulser, anti tumor dan agen anti parasit. Memandangkan potensi nilai komersial sebagai tanaman estet dan juga untuk pemuliharaan germplasma, pembentukan embrio somatik dan regenerasi secara terus *E. longifolia* telahpun dijalankan kerana cara micropropagasi secara *in vitro* tidak pernah dilaporkan sebelum ini.

Dalam usaha untuk mendapatkan embrio somatik daripada *E. longifolia*, kalus yang berpotensi daripada eksplan kotilidon, embrio zigotik, daun, petiol, batang dan akar tunjang telah dianalisis dalam basal media Murashige and Skoog (MS) yang dibekalkan dengan pelbagai jenis auksin pada kepekatan yang berbeza. Hanya eksplan kotilidon sahaja yang mampu menghasilkan kalus embriogenik sebanyak 30% dengan kehadiran 1.0 mg/L (b/i) 2,4-dichlorophenoxyacetic (2,4-

D). Sebanyak 60% kalus embriogenik dihasilkan bila kaedah pemotongan jenis ke-4 dikulturkan di dalam basal media yang mengandungi 0.5 mg/L (b/i) kinetin dan 1.0 mg/L (b/i) 2,4-D. Sebanyak  $45 \pm 2$  embrio somatik dapat diperhatikan di dalam formulasi medium yang sama dan telah ditambah dengan 1.0 g/L (b/i) arang teraktif. Seterusnya, pemindahan embrio somatik yang matang ke dalam basal MS medium yang telah dibekalkan dengan 1.0 mg/L (b/i) kinetin boleh menghasilkan regenerasi anak pokok sebanyak 90%. Penambahan arang teraktif, kasein hidrolisat, asid absisik, prolin dan polietilene glicol (PEG) pada kepekatan yang berbeza ke dalam medium regenerasi tidak berjaya merangsang regenerasi embrio somatik kepada anak pokok. Perbezaan di antara kalus embriogenik dan kalus bukan embriogenik juga dapat dikenalpasti melalui analisis histologi.

Regenerasi secara terus telah berjaya diperolehi daripada eksplan akar, batang, tunas ketiak dan juga tunas sisi. Setiap eksplan memerlukan kombinasi medium dan hormon perangsang tumbuhan yang berbeza untuk menghasilkan peratus regenerasi yang tinggi. Bagi eksplan akar, formulasi medium terbaik telah dikenalpasti di dalam basal Juglans medium DKW yang dibekalkan dengan 1.0 mg/L (b/i) kinetin + 1.0 mg/L (b/i) zeatin, manakala bagi eksplan batang, basal medium WPM yang dibekalkan dengan 2.0 mg/L (b/i) BAP dan 2.0 mg/L (b/i) zeatin telah dikenalpasti sebagai formulasi terbaik kerana menghasilkan anak pokok yang sihat serta peratus regenerasi yang tinggi. Eksplan akar dan batang yang berada pada jarak 2 cm di antara satu sama lain telah dikenalpasti sebagai posisi yang paling sesuai untuk menghasilkan peratus regenerasi yang maksima. Regenerasi secara terus daripada eksplan pucuk *in vitro* dan *in vivo* *E. longifolia*

telah diperolehi di dalam basal medium MS yang dibekalkan masing-masing dengan 3.0 dan 5.0 mg/L (b/i) kinetin. Bagi regenerasi secara terus daripada eksplan tunas ketiak, sebanyak 90% kadar regenerasi diperolehi di dalam basal medium Nitsch (NM) yang dibekalkan dengan 10.0 mg/L (b/i) zeatin manakala basal medium NM yang dibekalkan dengan 6.0 mg/L (b/i) zeatin telah menghasilkan peratus regenerasi yang tinggi bagi eksplan tunas sisi. Pengakaran daripada anak pokok *in vitro* *E. longifolia* juga berjaya dihasilkan dan kadar penghasilan yang tertinggi telah diperolehi di dalam basal media MS yang dibekalkan dengan 0.5 mg/L (b/i) Indole-3-butyric acid (IBA). Anak pokok *in vitro* yang terbentuk daripada embrio somatik dan regenerasi secara terus tumbuh dengan baik dan tidak menunjukkan sebarang perbezaan dari segi morfologi dengan pokok induknya selepas dua bulan dipindahkan ke dalam tanah.

## ACKNOWLEDGEMENTS

First of all, I am grateful to Allah the most merciful for blessing me with good health, power and support throughout my studies.

I would like to express my appreciation to my supervisor, Prof. Dr. Maziah Mahmood, my co-supervisors, Assoc.Prof. Dr. Siti Khalijah Daud and Assoc. Prof. Dr. Nor'aini Mohd Fadzillah whose encouragement and support provided a strong foundation on which I was able to develop my thesis and continue my research. I would also like to extend my thanks to Assoc. Prof. Dr. Jafar Sidik for allowing me to use the facilities in his lab for my histology studies.

I appreciated the financial support from the Malaysia government under the Malaysia-MIT Biotechnology Partnership Program (MMBPP). Thanks to Universiti Putra Malaysia also for offering me with the Graduate Research Assistantship.

A heartfelt thank-you to all the members of lab 235 for their support, help and encouragement. Special thanks to Dato, Dr. Yunus and Thiyagu for being the true friends in my life. I am indebted to your never-ending kindness, generosity and friendship. To Pak Azis and Pak Shuib, thanks for providing me with the plant materials and the invaluable advices. I could not have done it without the support from both of you.

Finally and most importantly, I must thank my family. To my wonderful parents who give me the opportunity and provided unyielding support throughout my academic years. I would also like to thank my brothers and sisters who have not only been my source of inspiration, but also given themselves in the most unselfish way.

To the one who are not only close to myself, but also are close to my heart and mind, Dr. Anna. Thanks for all the affection, support and encouragement during the good and hard times it took for me to finish this work. Thanks also for spending the valuable time in reading and editing my thesis. Your patience and understanding were essential to the completion of this thesis. To Teresa, Joseph, Angela, Andrew, Catherine and Paul, thanks for treating me as part of the family. I am grateful to your listening ears, advices and encouragement.

I certify that an Examination Committee met on 4<sup>th</sup> October 2004 to conduct the final examination of Sobri Bin Hussein on his Doctor of Philosophy thesis entitled “Micropropagation of Tongkat Ali (*Eurycoma longifolia* Jack) via Somatic Embryogenesis and Direct Plant Regeneration Techniques” in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher degree) Regulations 1981. The Committee recommends that the candidate be awarded the relevant degree. Members of the Examination Committee are as follows:

**Radzali Muse, Ph.D.**

Associate Professor  
Faculty of Biotechnology and Biomolecular Sciences  
Universiti Putra Malaysia  
(Chairman)

**Janna Ong Abdullah, Ph.D.**

Lecturer  
Faculty of Biotechnology and Biomolecular Sciences  
Universiti Putra Malaysia  
(Member)

**Mohd Puad Abdullah, Ph.D.**

Lecturer  
Faculty of Biotechnology and Biomolecular Sciences  
Universiti Putra Malaysia  
(Member)

**Chan Lai Keng, Ph.D.**

Professor  
School of Biological Sciences  
Universiti Sains Malaysia  
(Independent Examiner)

---

**GULAM RUSUL RAHMAT ALI, Ph.D.**

Professor/Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date:

This thesis submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee are as follows:

**Maziah Mahmood, Ph.D.**

Professor  
Faculty of Biotechnology and Biomolecular Sciences  
Universiti Putra Malaysia  
(Chairman)

**Siti Khalijah Daud, Ph.D.**

Associate Professor  
Faculty of Science  
Universiti Putra Malaysia  
(Member)

**Nor'aini Mohd Fadzillah, Ph.D.**

Associate Professor  
Faculty of Science  
Universiti Putra Malaysia  
(Member)

---

**AINI IDERIS, Ph.D.**

Professor/Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date :

## **DECLARATION**

I hereby declare that the thesis is based on my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at UPM or other institutions.

---

**SOBRI BIN HUSSEIN**

**Date:**

## TABLE OF CONTENTS

	<b>Page</b>
<b>DEDICATION</b>	<b>ii</b>
<b>ABSTRACT</b>	<b>iii</b>
<b>ABSTRAK</b>	<b>vi</b>
<b>ACKNOWLEDGEMENTS</b>	<b>ix</b>
<b>APPROVAL</b>	<b>xi</b>
<b>DECLARATION</b>	<b>xiii</b>
<b>LIST OF TABLES</b>	<b>xvii</b>
<b>LIST OF FIGURES</b>	<b>xx</b>
<b>LIST OF PLATES</b>	<b>xxii</b>
<b>LIST OF ABBREVIATIONS</b>	<b>xxv</b>
<b>CHAPTER</b>	
1 INTRODUCTION	1
1.1 Plant Tissue Culture	2
1.2 <i>Eurycoma</i> species	4
2 LITERATURE REVIEW	7
2.1 <i>Eurycoma longifolia</i> Jack	7
2.1.1 Medicinal importance	8
2.1.2 Chemical constituents	10
2.2 Conventional plant propagation	14
2.3 Micropropagation through somatic embryogenesis	16
2.3.1 Stages of somatic embryogenesis	19
2.3.2 Regeneration of cell suspension	23
2.4 Micropropagation through direct plant regeneration	25
2.5 Micropropagation through organogenesis	28
2.6 Factors influencing micropropagation	30
2.6.1 Genotypes	30
2.6.2 Types of explants	31
2.6.3 Basal media	35
2.6.4 Plant growth regulators (PGRs)	37
2.6.5 pH of Medium	40
2.6.6 Nitrogen sources	41
2.6.7 Carbon sources	43
2.6.8 Abscisic acid and osmoticum	44
2.6.9 Incubation conditions	46

2.7	Problems associated with micropropagation of woody species	47
2.7.1	Genotypic variation	48
2.7.2	Restricted availability of explant sources	48
2.7.3	Age of donor plant	49
2.7.4	Endogenous contamination and browning effects	49
2.7.5	Instability and low multiplication rate	50
2.7.6	Rooting	51
2.7.7	Somaclonal variation	52
2.8	Biochemical markers for somatic embryogenesis and plant regeneration	52
3	SOMATIC EMBRYOGENESIS OF <i>EURYCOMA LONGIFOLIA</i> JACK	54
3.1	Introduction	54
3.2	Materials and Methods	56
3.2.1	Plant materials	56
3.2.2	Selection of potential explants using different auxin at various concentrations	57
3.2.3	Effects of different dissection methods using cotyledons	59
3.2.4	Embryogenic callus induction from potential explant	61
3.2.5	Multiplication of somatic embryos	62
3.2.6	Plantlet regeneration from mature somatic embryos	63
3.2.7	Hardening and acclimatization	65
3.2.8	Histological Studies	66
3.2.9	Statistical analysis	68
3.3	Results and Discussions	69
3.3.1	Selection of potential explants using different auxins at various concentrations	69
3.3.2	Effects of different dissection methods using cotyledons	90
3.3.3	Embryogenic callus induction from potential explant	92
3.3.4	Multiplication of somatic embryos	97
3.3.5	Plantlet regeneration from mature somatic embryos	107
3.3.6	Hardening and acclimatization	126
3.3.7	Histological Studies	126
3.4	Conclusions	131

4	DIRECT PLANT REGENERATION FROM DIFFERENT EXPLANTS OF <i>EURYCOMA LONGIFOLIA</i> JACK	134
4.1	Introduction	134
4.2	Materials and Methods	135
4.2.1	Plant materials	135
4.2.2	Direct plant regeneration from root explants	135
4.2.3	Direct plant regeneration from stem explants	137
4.2.4	Direct plant regeneration from shoot tip explants	139
4.2.5	Direct plant regeneration from axillary and adventitious buds explants	140
4.2.6	Rooting	141
4.2.7	Hardening and acclimatization	141
4.2.8	Statistical analysis	142
4.3.	Results and Discussions	142
4.3.1	Direct plant regeneration from root explants	142
4.3.2	Direct plant regeneration from stem explants	156
4.3.3	Direct plant regeneration from <i>in vitro</i> shoot tip explants	168
4.3.4	Direct plant regeneration from <i>in vivo</i> shoot tip explants	179
4.3.5	Direct plant regeneration from axillary bud explants	191
4.3.6	Direct plant regeneration from adventitious bud explants	195
4.3.7	Induction of rootlets from <i>in vitro</i> plantlets	198
4.3.8	Hardening and acclimatization	208
4.4	Conclusions	208
5	GENERAL DISCUSSIONS AND CONCLUSION	211
	<b>REFERENCES</b>	<b>217</b>
	<b>APPENDICES</b>	<b>249</b>
	<b>BIODATA OF THE AUTHOR</b>	<b>293</b>