

UNIVERSITI PUTRA MALAYSIA

IN VITRO PROPAGATION AND MUTATION INDUCTION OF TORCH GINGER (Etlingera elatior J.)

MUHAMAD FAHMI YUNUS

FP 2013 73



IN VITRO PROPAGATION AND MUTATION INDUCTION OF TORCH GINGER

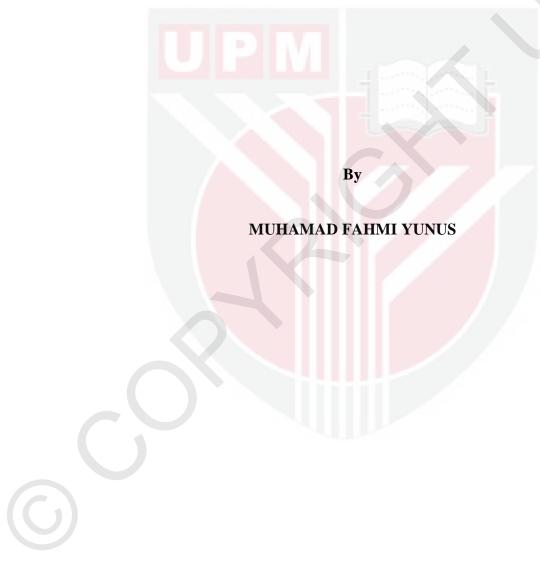
(Etlingera elatior J.)

MUHAMAD FAHMI YUNUS

MASTER OF SCIENCE UNIVERSITI PUTRA MALAYSIA 2013



IN VITRO PROPAGATION AND MUTATION INDUCTION OF TORCH GINGER (Etlingera elatior J.)



Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fullfilment of the Requirements for the Degree of Master of Science

October 2013

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia

Dedicated to:



My dearest parents

Yunus bin Jamaludin Zawiah binti Basnun

and

My Siblings

Muhamad Aqqat bin Yunus Muhammad Ehsan Sabri bin Yunus Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

IN VITRO PROPAGATION AND MUTATION INDUCTION OF TORCH GINGER (Etlingera elatior J.)

By

MUHAMAD FAHMI YUNUS

October 2013

Chairman : Associate Professor Maheran Abd Aziz, PhD Faculty : Faculty of Agriculture

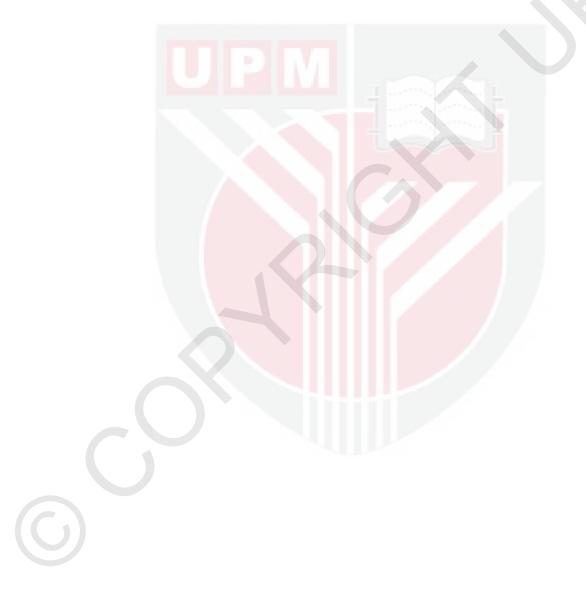
The aim of this study was to develop a protocol for *in vitro* propagation and mutation induction of *Etlingera elatior* by using gamma ray irradiation. The study included establishment an efficient *in vitro* plant propagation system in *E. elatior*, investigation of the optimum dose for radio sensitivity test, to determine the effects of various doses of gamma irradiation on multiple bud induction and also to determine the variation in genomic DNA of regenerated shoots by using random amplification of polymorphic DNA (RAPD) technique.

In this study, an efficient and systematic protocol for complete plant regeneration from suckers of *Etlingera elatior* (J.) has been developed. The addition of N6-benzyl amino-purine (BAP) (0, 3, 5, 7 and 10 mg L⁻¹) to the culture medium comprising of Murashige and Skoog (MS) basal salts, 3% sucrose, 0.4% gelrite did not show any significant effects on percentage of shoot induction and mean number of shoots produced. However, BAP at 3 mg L⁻¹ was chosen as the best medium for shoot induction due to economic feasibility and it gave the highest result in all four parameters recorded. Various concentrations of BAP, 6-furfurylaminopurine (kinetin) and N6-(2-isopentenyl) adenine (2-iP) alone at 0, 3, 5, 7 and 10 mg L⁻¹ were tested for shoot multiplication. BAP at all levels were found suitable for the multiplication of shoot. However, the low level of 3 mg L⁻¹ BAP was chosen as the best concentration of BAP due to economic feasibility. The best root proliferation was observed on MS medium without plant growth regulator (PGR). Assessment of various potting media for acclimatization showed medium containing soil: sand: peat moss (1:1:1) produced high survival of plantlets, number of leaves produced per plant and the plant height.

Mutation breeding techniques in combination with tissue culture and molecular marker methods provide a powerful tool for improvement of vegetatively propagated plants. The results of irradiation on *in vitro* buds of *E. elatior* showed that LD_{50} to be 10 Gy with the survival of explants being sharply reduced after this dosage. The gamma irradiated shoots were subcultured for three cycles (M_1V_1 to M_1V_3) to obtain potential mutant lines. This study showed that RAPD marker was efficient in differentiating the induced mutants from the untreated control of *E. elatior*. All eight selected gamma irradiated regenerants were differentiated from the untreated control based on the banding patterns obtained using 9 primers which generated 59 reproducible bands, whereby 35 (55.31%) were found to be polymorphic. The Jaccard's coefficient of similarity values ranging from 0.537 to 0.860 were indicative of the level of genetic variation among the mutants studied. For comparison between the potential lines (PL) and the control, a maximum similarity value (0.814) was observed in PL1 mutant while the minimum value (0.537) was observed in PL7. The

presence of polymorphic bands in 8 potential lines suggested that genetic variation occurred in all the treatments as compared to the control.

In summary, the combination of techniques of *in vitro* propagation, multiplication, gamma irradiation, and RAPD analysis for early screening of mutants can facilitate breeding programme of *E. elatior*.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk Ijazah Master Sains

PROPAGASI IN VITRO DAN ARUHAN MUTASI POKOK KANTAN (Etlingera elatior J.)

Oleh

MUHAMAD FAHMI YUNUS

Oktober 2013

Pengerusi : Professor Madya Maheran Abd Aziz, PhD Fakulti : Fakulti Pertanian

Tujuan penyelidikan yang dijalankan ialah untuk membangunkan teknik propagasi *in vitro* dan aruhan mutasi *Etlingera elatior* dengan menggunakan penyinaran sinar gamma. Kajian ini merangkumi penghasilan satu sistem propagasi tumbuhan secara *in vitro* bagi *E. elatior*, kajian mengenai dos optimum bagi ujian sensitiviti radio, penentuan kesan-kesan pelbagai dos sinaran gamma dan juga penentuan variasi pada genom DNA dari pucuk yang dihasilkan dengan menggunakan teknik penanda molekul amplikasi rawak DNA polimorfik (RAPD).

Di dalam penyelidikan ini, protokol yang effisien dan sistematik untuk regenerasi tumbuhan dari sulur *E. elatior* (J.) telah dibangunkan. Penambahan N6-benzil aminopurin (BAP) pada kepekatan 0, 3, 5, 7 dan 10 mg L⁻¹ pada kultur medium yang mengandungi nutrien asas Murashige dan Skoog (MS), 3% sukrosa, 0.4% Gelrite tidak menunjukkan sebarang kesan yang signifikan terhadap peratusan penginduksian pucuk dan juga jumlah min penghasilan bilangan pucuk. Walaubagaimanapun, BAP pada kepekatan 3 mg L⁻¹ telah dipilih sebagai paras kepekatan terbaik disebabkan faktor ekonomi dan ia memberikan keputusan terbaik di dalam semua empat parameter yang direkodkan. Pelbagai kepekatan tunggal pengawalatur pertumbuhan BAP, 6-furfurilaminopurin (Kinetin) dan juga N6-(2isopentenil) adenin (2-iP) pada kepekatan 0, 3, 5, 7 dan 10 mg L⁻¹ telah diuji untuk penggandaan pucuk. BAP pada setiap kepekatan telah diuji berkesan untuk penggandaan pucuk. Sungguhpun demikian, BAP pada kepekatan 3 mg L⁻¹ telah dipilih sebagai paras kepekatan terbaik berdasarkan sifat ekonomi yang dimiliki. Medium penggandaan akar yang terbaik ialah medium MS tanpa sebarang penambahan pengawalatur pertumbuhan. Penilaian pelbagai media berpasu bagi tujuan aklimitasi menunjukkan bahawa medium yang mengandungi tanah: pasir: tanah gambut berlumut (1:1:1) memberikan kadar kemandirian yang tinggi kepada anak pokok, penghasilan daun per anak pokok dan juga tinggi anak pokok.

Teknik pembiakbaka mutasi dengan kombinasi teknik kultur tisu dan penanda molekul boleh menjadi teknik yang berkesan untuk penambahbaikan tumbuhan yang dibiakkan melalui kaedah tampang. Keputusan irradiasi tunas *in vitro E. elatior* menunjukkan bahawa LD_{50} ialah pada 10 Gy dengan kadar hidup berkurangan dengan drastik selepas dos ini. Pucuk yang telah disinari dengan sinar gamma telah disubkultur untuk tiga kitaran (M_1V_1 ke M_1V_3) untuk memperolehi titisan mutan yang berpotensi. Kajian ini menunjukkan RAPD adalah berkesan untuk membezakan antara mutan yang diaruh daripada kawalan *E. elatior* yang tidak diaruh. Kesemua lapan regenerasi yang diaruh dengan gamma telah dibezakan daripada kawalan yang tidak diaruh berdasarkan corak jalur yang diperolehi dengan menggunakan 9 primer yang menghasilkan 59 jalur reproduksi, di mana 35 (55.31%) adalah polimorfik. Nilai pekali pesamaan Jaccard berada di antara julat 0.537 hingga 0.860 menunjukkan paras kepelbagaian genetik antara mutan yang dikaji. Sebagai perbandingan, antara titisan berpotensi (PL) dan juga kawalan, nilai kesamaan maksimum (0.814) telah diperolehi pada mutan PL1 manakala nilai minimum (0.537) diperolehi pada PL7. Penghasilan jalur polimorfik pada 8 titisan berpotensi menyarankan bahawa variasi genetik berlaku pada semua rawatan berbanding dengan kawalan.

Sebagai kesimpulan, kombinasi antara teknik propagasi *in vitro*, penggandaan, penyinaran gamma, dan juga analisis RAPD untuk penyaringan awal mutan boleh membantu program pembiakbakaan *E.elatior*.

ACKNOWLEDGEMENTS

In the name of Allah, the Most Gracious and the Most Merciful. Foremost, I would like to express my deep and sincere gratitude to my supervisor, Associate Professor Dr. Maheran Abd Aziz for the continuous support of my MSc study and research. Her wide knowledge, understanding, encouraging and personal guidance have provided a good basis for the present thesis and have been of great value for me. Besides, I would like to thank the rest of my thesis committee: Associate Prof. Dr. Mihdzar Abdul Kadir and Associate Prof. Dr. Siti Khalijah Daud; Mr. Azmi Abdul Rashid, a lecturer in the Faculty of Agriculture for their encouragement, insightful comments and suggestions.

The financial support from the Ministry of Science, Technology and Innovation Malaysia for me to undertake this study is gratefully acknowledged. I would like to thank Nor Asiah binti Ismail, Research Officer from MARDI Jerangau, Terengganu for providing the seed materials. I am also grateful to the staff at Gamma Ray Laboratory, School of Applied Physics, Faculty of Science, Universiti Kebangsaan Malaysia, Selangor for providing the facilities to carry out the gamma irradiation of *E. elatior*.

I thank my fellow labmates in Agrobiotechnology Laboratory, Department of Agriculture Technology, Faculty of Agriculture for the stimulating discussions, and for all the fun we have had in the last two years. Last but not the least, I offer my regards and blessings to all of those who supported me in any respect during the completion of the project.



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fullfillment of the requirements for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Maheran Abd Aziz, PhD Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Mihdzar Abdul Kadir, PhD Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Member)

Siti Khalijah Daud, PhD

Associate Professor Faculty of Science Universiti Putra Malaysia (Member)

(BUJANG BIN KIM HUAT, PhD)

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

DECLARATION

Declaration by Graduate Student

I hereby confirm that:

- this thesis is my original work;
- quotations, illustrations and citations have been duly referenced;
- this thesis has not been submitted previously or concurrently for any other degree at any other institutions;
- intellectual property from the thesis and copyright of thesis are fully-owned by Universiti Putra Malaysia, as according to the Universiti Putra Malaysia (Research) Rules 2012;
- written permission must be obtained from supervisor and the office of Deputy Vice-Chancellor (Research and Innovation) before thesis is published (in the form of written, printed or in electronic form) including books, journals, modules, proceedings, popular writings, seminar papers, manuscripts, posters, reports, lecture notes, learning modules or any other materials as stated in the Universiti Putra Malaysia (Research) Rules 2012;
- there is no plagiarism or data falsification/fabrication in the thesis, and scholarly integrity is upheld as according to the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) and the Universiti Putra Malaysia (Research) Rules 2012. The thesis has undergone plagiarism detection software.

Signatu	re:				1 October	<u>2013</u>		
Name	and	Matric	No.:	Muhamad	Fahmi	Yunus	and	GS26314

Declaration by Members of Supervisory Committee

This is to confirm that:

- the research conducted and the writing of this thesis was under our supervision; Guide to Thesis Preparation
- supervision responsibilities as stated in the Universiti Putra Malaysia (Graduate Studies) Rules 2003 (Revision 2012-2013) are adhered to.

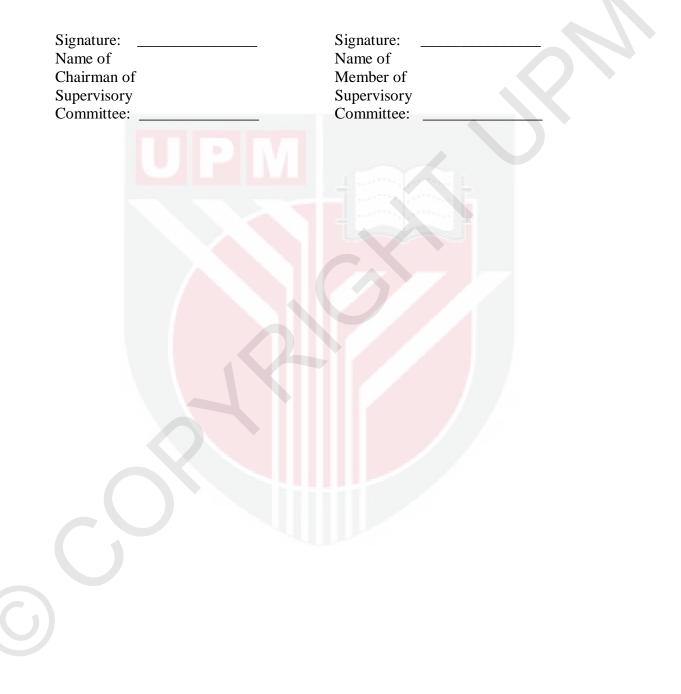


TABLE OF CONTENTS

DF	EDICATI	ON	Page ii		
	BSTRAC'		iii		
	ABSTRAK				
		LEDGEMENTS	xivi ix		
	PROVA		X		
	ECLARA		xii		
	ST OF T		xvii		
	ST OF F		xviii		
		BBREVIATIONS	XX		
,					
CH	IAPTER				
1		RODUCTION			
	1.1	Background	1		
	1.2	Problem Statement	2		
	1.3	Significance of the Study	2 3		
	1.4	Objective	5		
2	LITI	ERATURE REVIEW			
	2.1	Taxonomy	6		
	2.2	Botany of Torch Ginger	8		
	2.3	Comercial Potential of Torch Ginger	11		
	2.4	Plant tissue culture	12		
		2.4.1 Direct Regeneration	12		
		2.4.2 Indirect Regeneration	14		
		2.4.3 Plant Growth Regulator	14		
		2.4.4 Interaction between Auxin and Cytokinin	17		
	2.5	Conventional Breeding of Torch Ginger	19		
	2.6	Mutation Breeding	19		
		2.6.1 Effect of Mutagen on Plants	20		
		2.6.2 Conventional Mutagenesis	21		
		2.6.3 In vitro Mutagenesis	21		
		2.6.4 Advantages of <i>In Vitro</i> Mutagenesis	22		
	2.7	Radiation Process	24		
	2.8	Gamma Rays	25		
	2.9	Radiosensitivity Test	25		
	2.10	Lethal Dose 50	26		
	2.11	Chimera	27		
	2.12	Molecular Marker	29		
		2.12.1 Random Amplification of Polymorphic DNA	29		

3 MATERIALS AND METHODS

4

 \bigcirc

MAI	ERIALS AND METHODS		
Part I:	In vitro propagation	31	
3.1	1 Plant Materials		
3.2	Surface Sterilization Techniques	32	
3.3	Effects of Different Concentration of BAP on Shoot Induction	33	
3.4	Effects of Different Types and Concentrations of	34	
2.5	Cytokinin on Shoot Multiplication	25	
3.5	Effects of Different Types and Concentrations of Auxin on Root Induction	35	
3.6	Effects of Different Types of Growth Media on Acclimatization	35	
3.7	Effects of 2,4-D and BAP Concentration in ¹ / ₂ MS Medium	36	
20	on Percentage and Intensity of Callus Formation Callus Proliferation	20	
3.8		38	
3.9	Experimental Design and Statistical Analysis	38	
Part II:	: In vitro Mutagenesis and RAPD Analysis	39	
3.10	Plant Materials	39	
3.11	Effect of Gamma Irradiation on Regeneration of Shoots from In Vitro Buds	39	
3.12	Post Mutagenesis Handling	40	
3.13	Experimental Design and Statistical Analysis	42	
3.14	Plant Materials and Genomic DNA Extraction	42	
3.15	DNA Quantification	43	
3.16	Primer Selection	44	
3.17	Polymerase Chain Reaction and Primer Screening	45	
3.18	Statistical Analysis	45	
RESU	LTS AND DISCUSSION		
Part 1:	In vitro Propagation	47	
4.1	Effects of Different Concentration of BAP on Shoot Induction	47	
1.2		52	
4.2	Effects of Different Types and Concentration of Cytokinin on Shoot Multiplication	53	
4.3	Effects of Different Types and Concentration of Auxin on Root Induction	60	
4.4	Effects of Different Types of Growth Media on	64	
	Acclimatization		
4.5	Effects of 2,4-D and BAP Concentration in ¹ / ₂ MS	68	
	Medium on Percentage of Callus Formation and		
	Intensity of Callus Formation		

Part II	: In vitro Mutagenesis and Molecular Marker	73
4.6	Effect Of Gamma Irradiation On Regeneration Of Shoots	73
	From In Vitro Buds	
4.7	Management of Chimera and Biological Effect Of	79
	Gamma Rays	
4.8	Post Mutagenesis Handling and Modification of the	81
	Culture Conditions	
4.9	Evaluation of Genetic Variation of Gamma Rays	82
	Treated Regenerants with RAPD Markers	

5 CONCLUSION

G

REFERENCES APPENDICES BIODATA OF STUDENT LIST OF PUBLICATIONS 89 91

100 112

113

LIST OF TABLES

Table		Page
1	Common species from Zingiberaceae family that can be found in Malaysia.	7
2	Variability among six accessions of E. elatior maintained in MARDI Gene Bank, Jerangau, Terengganu	10
3	Different combinations and concentrations of BAP and 2,4-D in half strength MS medium for callus induction	36
4	Scores for callus intensity	37
5	Treatments dose and exposure time provided by UKM. Dose rate: 2.8741 Kgy/ hour	40
6	A total of 14 RAPD primers used in preliminary primer screening	44
7	Effects of different concentrations of BAP on shoot induction from <i>E. elatior</i> suckers after 12 weeks of culture	49
8	Effects of cytokinin type and concentration on shoot multiplication of <i>E. elatior</i> after 12 weeks of culture	54
9	Rooting of <i>E. elatior</i> shoots on MS media with different types and concentrations of auxin after 8 weeks of culture	61
10	Effect of potting media on plantlet performance of <i>E. elatior</i> after six weeks of acclimatization	65
11	Effects of 2,4-D and BAP concentration in half MS medium on percentage of callus formation and intensity of callus formation after 12 weeks of culture	70
12	Effects of different doses of gamma irradiation on shoot regeneration from buds of <i>E elatior</i> after 8 weeks of culture	77
13	List of primers, number of amplified products, polymorphic bands and polymorphism percentage	83
14	Jaccard's coefficient of similarity matrix for control and potential mutant lines of <i>E. elatior</i> determined from RAPD analysis using 9 different primers and analyzed by UPGMA programme	85

LIST OF FIGURES

Figure		Page
1	Suckers (arrow) of E. elatior used in this study.	32
2	Explants used in shoot induction study. The outer scales and dead surface of the sterilized suckers were trimmed off before placing on the culture medium	33
3	Effects of different concentrations of BAP on shoot induction from <i>E. elatior</i> suckers after 12 weeks of culture on (A) BAP 3 mg L^{-1} (B) BAP 5 mg L^{-1} (C) BAP 7 mg L^{-1} and (D) BAP 10 mg L^{-1}	51
4	Development of shoots from sucker's of <i>E.elatior</i> on MS medium containing 3 mg L^{-1} BAP after (A) 5 weeks (B) 6 weeks (C) 8 weeks (D) 9 weeks (E) 11 weeks and (F) 12 weeks of culture	52
5	Effects of cytokinin type and concentration on shoot multiplication of <i>E. elatior</i> after 12 weeks of culture on (A) MSO	55
5	Effects of cytokinin type and concentration on shoot multiplication of <i>E. elatior</i> after 12 weeks of culture on (B) BAP 3 mg L^{-1} (C) BAP 5 mg L^{-1} (D) BAP 7 mg L^{-1} and (E) BAP 10 mg L^{-1}	56
5	Effects of cytokinin type and concentration on shoot multiplication of <i>E. elatior</i> after 12 weeks of culture on (F) KIN 3 mg L^{-1} (G) KIN 5 mg L^{-1} (H) KIN 7 mg L^{-1} and (I) KIN 10 mg L^{-1}	57
5	Effects of cytokinin type and concentration on shoot multiplication of <i>E. elatior</i> after 12 weeks of culture on (J) 2-iP 3 mg L^{-1} (K) 2-iP 5 mg L^{-1} (L) 2-iP 7 mg L^{-1} and (M) 2-iP 10 mg L^{-1}	58
6	Rooting of <i>E. elatior</i> shoots on MS media with different types and concentrations of auxin after 8 weeks of culture on (A) MSO	62
6	Rooting of <i>E. elatior</i> shoots on MS media with different types and concentrations of auxin after 8 weeks of culture on (B) IBA 1 mg L ⁻¹ (C) IBA 2 mg L ⁻¹ (D) IBA 3 mg L ⁻¹ (E) NAA 1 mg L ⁻¹ (F) NAA 2 mg L ⁻¹ and (G) NAA 3 mg L ⁻¹	63

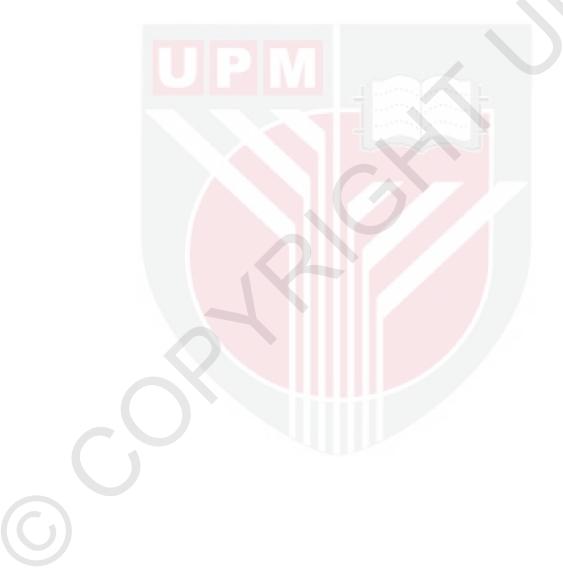
- 7 Effect of potting media on plantlet performance of *E. elatior* 67 after six weeks of acclimatization on (A) Sand (B) Soil (C) Peat moss and (D) Sand + soil + peatmoss (1:1:1).
- 8 Effects of 2,4-D and BAP in half MS medium on callus 72 formation from leaf bases of of *E. elatior* after 12 weeks of culture on (A) C9 (1.0 mg L⁻¹ 2,4-D and 0.1 mg L⁻¹ BAP) (B) C12 (6.0 mg L⁻¹ 2,4-D and 0.1 mg L⁻¹ BAP) (C) C18 (6.0 mg L⁻¹ 2,4-D and 0.5 mg L⁻¹ BAP) and (D) C24 (6.0 mg L⁻¹ 2,4-D and 1.0 mg L⁻¹ BAP) media
- 9 LD_{50} determination on survival rate of irradiated buds of *E*. 75 *elatior* after 8 weeks of incubation
- 10 Effect of different levels of gamma irradiation on *in vitro* buds 76 of *E. elatior*. A) Survival of explants exposed to 10 Gy after 8 weeks of culture. B) Morphological abnormality observed on irradiated explant which later died after M_1V_1 stage. C) Explant irradiated with 20 Gy failed to survive after 12 weeks of culture. D) M_1V_1 shoots produced from 10 Gy irradiated explant after 12 weeks of culture.
- 11 Amplification of genomic DNA from *in vitro* shoots (M_1V_3) 84 treated with 10 Gy of gamma rays using various RAPD primers, A) primer OPAW11, B) primer OPU16, and C) primer OPU13. In each of the three panels, lane M corresponds to 10 kb DNA ladder; lane C corresponds to the control (non irradiated plant), lanes 2 through 10 correspond to DNA from 9 potential regenerant lines, lane NC on the third panel corresponds to the negative control. Note: solid arrows point to the polymorphic bands, while the blank arrows point to the monomorphic bands
- 12 Dendogram constructed from Jaccard's similarity coefficients 86 from RAPD data, showing the clustering of the 9 regenerants. (C: control, PL1 - PL8: Potential lines)

xix

LIST OF ABBREVIATIONS

	%	Percentage
	$\mu mol \ m^{-2} \ s^{-1}$	Micromole per square meter per second
	2, 4-D	2,4-Dichlorophenoxyacetic acid
	⁶⁰ Co	Cobalt-60
	2-iP	N6-(2-isopentenyl) adenine
	ANOVA	Analysis of variance
	BAP	N6-benzyl amino-purine
	cm	Centimetre
	СТАВ	Cetyl trimethylammonium bromide
	DNA	Deoxyribonucleic acid
	DMRT	Duncan's Multiple Range Test
	EDTA	Ethylenediaminetetraacetic acid
	g	Gram
	Gy	Gray
	IAA	indole-acetic acid
	IBA	indole-3-butyric acid
	KIN	6-furfurylaminopurine
	LD	Lethal Dose
	MARDI	Malaysian Agricultural Research and Development Institute
	MS	Murashige and Skoog
	NAA	1-naphthaleneacetic acid
	PCR	Polymerase Chain Reaction
	PPM	Plant Preservative Mixture
	RAPD	Random Amplification of Polymorphic DNA

RCBD	Randomized Complete Block Design
SAS	Statistical Analysis System
SE	Standard Error
UKM	Universiti Kebangsaan Malaysia
UPM	Universiti Putra Malaysia



CHAPTER 1

INTRODUCTION

1.1 Background

Zingiberaceae is one of the largest families of the plant kingdom. It is an important family which provides many useful products for food, spices, medicines, perfume, dyes essentials oil and aesthetics to man (Jaafar et al., 2007; Poulsen, 2006). It provides plants of economic value mainly for its beautiful flowers, vegetable and also ingredient in a dish. These species are represented throughout the tropical and subtropical regions where the Indo Malayan (Indonesia, Malaysia, Brunei, Singapore, Papua New Guinea and the Southern Philippines) region is the centre of diversity for the Zingiberaceae. Of the 52 genera and 1500 species known in the world, at least 25 genera and 650 species can be found in Malaysia (Sirirugsa, 1999).

Etlingera elatior (Jack) R.M.Sm which belongs to the Zingiberaceae family is one of the most commonly known species of *Etlingera*. This species is also known as torch ginger or wax flower due to the striking resemblance of the inflorescence to a flaming torch. Torch ginger is widely cultivated in the tropical country and possibly native to Indonesia and Malaysia. It is known as *kantan* in Malaysia and *kecombrang* in Indonesia.

1.2 Problem Statement

E. elatior is one of the neglected plants in the Zingiberaceae family and scientific research on their propagation technique, production, biotechnology and ecology is limited. However, this ginger species offers great scope for the development of a large range of ornamental and cut flower types (Poulsen, 2006). Ismail (2009) reported that *E. elatior* is one of the 30 popular herbs or new industrial crops that have high demand in Malaysia. It is now cultivated on a commercial scale in places like Australia, Thailand and Costa-Rica for cut flower production (Ismail, 2009; Segalen, 2010). The plant itself makes a great garden landscape, their flowers having an immense ornamental value and also has a place in an eco-garden. To sustain in the ornamental and cut flower industry, torch ginger requires continuous improvement in certain characters like flower colour, morphology, longevity, size, odour and decreased time to flower formation. Unfortunately, conventional breeding of *E.elatior* is handicapped by the cross incompatibility and poor fruit set and also low seed production (Marcsik and Hoult, 2010). Due to these factors, alternative approaches for crop improvement of E. elatior such as through mutation induction could be explored.

In vitro culture techniques provide an alternative means of propagation and a tool for crop improvement (Zheng et al., 2008). Unfortunately, this medicinal plant has not received much attention from tissue culturists and to date, there is limited information on plant regeneration of this medicinal plant. Hence, with the increasing importance of torch ginger as an ornamental species, there is a need for an efficient

protocol for plant regeneration using tissue culture techniques. The protocol developed here will be helpful for regenerating plants at much higher rates than any other conventional breeding methods and also may serve as a potential source of new variants and further genetic improvement in this species (Xu et al., 2009). In addition, development of new torch ginger cultivars with improved characters is another approach that can be explored. Possible approaches to create genetic variability for the selection of useful plants are through conventional plant breeding, mutagenesis, somaclonal variation and genetic transformation.

1.3 Significance of the Study

In a modern and industrialized horticulture, there is always a demand and necessity for new cultivars. Modern day plant breeding is based on creating variations followed by selection, evaluation and multiplication of the desired genotypes. Induced mutations have played an important role in the improvement of plants and more than 2500 mutant cultivars have been developed through mutation breeding (Patade et al., 2008). Mutation breeding is an established method for plant improvement, thus encouraging the plant breeders to use induced mutagenesis. Induction of mutation can increase the possibility a thousandfold compared with spontaneous mutation under natural condition (Broertjes and Van Harten, 1988).

In a crop improvement programme, plant breeders often combine several techniques in order to increase efficiency and reduce the time needed for the development of a new cultivar. Such combination has been exploited for the creation of new and novel plant cultivars, particularly in vegetatively propagated species (Pinet-Leblay et al., 1992; Broertjes and Van Harten, 1988). Successful outcome of a mutation depends on an efficient induction of mutation as well as an effective recognition and recovery of the desired mutant plants through repeated subculture (Puchooa, 2005).

The present study is divided into two parts: a) *In vitro* propagation and b) *In vitro* mutagenesis and RAPD analysis. A new technique is needed to standardize the management of chimeric tissues through multiple bud regeneration. Molecular techniques can provide an understanding of plant cell responses to mutation induction. It also facilitates a better understanding of the potential and limitations of mutation breeding, which can lead to early identification of useful variants. Major advantages of random amplification of polymorphic DNA (RAPD) are the low cost and effort required for its application (Dhakshanamoorthy, et al., 2011; Atienzar and Jha, 2006).

1.4 Objective

The objectives of this study are:

- 1) To establish an efficient *in vitro* plant regeneration system in torch ginger.
- 2) To investigate the optimum dose for radio sensitivity test and to determine
- the effects of various doses of gamma irradiation on multiple bud induction.
- To determine the variation in genomic DNA of regenerated shoots by using RAPD technique.

REFERENCES

- Abdullah, T.L., Endan, J. and Mohd Nazir, B. (2009). Changes in flower development, chlorophyll mutation and alteration in plant morphology of *Curcuma alismatofolia* by gamma irradiation. *American Journal of Applied Sciences*. 6(7): 1436-1439.
- Ahloowalia, B.S. and Maluszynski, M. (2001). Induced mutations a new paradigm in plant breeding. *Euphytica*. 119: 167-173.
- Ahmed Khan, I., Dahot, M.U., Seema, N., Yasmin, S., Bibi, S., Raza, S. and Khatri, A. (2009). Genetic variability in sugarcane plantlets developed through *in vitro* mutagenesis. *Pakistan Journal of Botany*. 41(1): 153-166.
- Al-Safadi, B. and Elias, R. (2011). Improvement of caper (*Capparis spinosa* L.) propagation using *in vitro* culture and gamma irradiation. *Scientia Horticulturae*. 127: 290–297.
- Andarwulan, N., Batari, R., Sandrasari, D.A., Bolling, B. and Wijaya, H. (2010). Flavonoid content and antioxidant activity of vegetables from Indonesia. *Food Chemistry*. 121: 1231-1235.
- Anon. (2010). Grower information Cut-flowers. http://www.nt.gov.au/d/Primary_Industry/index.cfm?header=Cut-flowers (accessed 19 Feb. 2010).
- Atak, C., Celik, O. and Acik, L. (2011). Genetic analysis of *Rhododendron* mutants using random amplified polymorphic DNA (RAPD). *Pakistan Journal of Botany*. 43(2): 1173-1182.
- Atak, C., Alikamanoglu, S., Acik, L. and Canbolat, Y. (2004). Induced of plastid mutations in soybean plant (*Glycine max L. Merrill*) with gamma radiation and determination with RAPD. *Mutation Research*: 556: 35-44.
- Atienzar, F.A and Jha, A.N. (2006). The random amplified polymorphic DNA (RAPD) assay and related techniques applied to genotoxicity and carcinogenesis studies: A critical review. *Mutation Research*. 613: 76-102.
- Barakat, M.N, Abdel Fattah, R.S., Badr, M. and El-Torky, M.G. (2010). In vitro mutagenesis and identification of new variants via RAPD markers for improving Chrysanthemum morifolium. African Journal of Agriculture Research. 5: 748-757.
- Barakat, M.N. and El-Sammak, H. (2011). *In vitro* mutagenesis, plant regeneration and characterization of mutants via RAPD analysis in Baby's breath *Gypsophila paniculata* L. *American Journal of Crop Science*. 5(2): 214-222.

- Bhojwani, S.S and Razdan, M.K. (1983). Plant tissue culture: theory and practice. Amsterdam: Elsevier Science Publisher.
- Bibi, S., Ahmed Khan, I., Khatri, A., Yasmin, S., Seema, N., Afghan, S and Arain, M.A. (2010). Screening of mutated population of sugarcane through RAPD. *Pakistan Journal of Botany*. 42(6): 3765-3773.
- Borthakur, M., Hazarika, J. and Singh, R.S. (1999). A protocol for micropropagation of *Alpinia galanga*. *Plant Cell, Tissue and Organ Culture*. 55: 231–233.
- Broertjes, C. and Van Harten, A.M. (1988). Applied mutation breeding for vegetatively propagated crops. Amsterdam: Elsevier.
- Chan, E.W.C., Lim, Y.Y., Ling, S.K., Tan, S.P., Lim, K.K. and Khoo, M.G.H. (2009a). Caffeoylquinic acids from leaves of *Etlingera* species (Zingiberaceae). *Food Chemistry*. 104: 1586–1593.
- Chan, E.W.C., Lim, Y.Y., Wong, S.K., Lim, K.K., Tan, S.P., Lianto, F.S. and Yong, M.Y. (2009b). Effects of different drying methods on the antioxidant properties of leaves and tea of ginger species. *Food Chemistry*. 113: 166–172.
- Chan, E.W.C., Lim, Y.Y. and Omar, M. (2007). Antioxidant and antibacterial activity of leaves of *Etlingera* species (Zingiberaceae) in Peninsular Malaysia. *Food Chemistry*. 104: 1586–1593.
- Chin, H.F. (1999). Malaysian Vegetables in Colour. A complete Guide. Kuala Lumpur: Tropical Press.
- Christianson, M.L. and Warnick, D.A. (1983). Competence and determination in the process of in vitro shoot organogenesis. *Developmental Biology*. 95(2): 288–293.
- Das, A., Gosal, S.S, Sidhu, J.S. and Dhaliwal, H.S. (2000). Induction of mutations for heat tolerance in potato by using *in vitro* culture and radiation. *Euphytica*. 114: 205–209.
- Das, A., Kesari, V. and Rangan, L. (2010). Plant regeneration in *Curcuma* species and assessment of genetic stability of regenerated plants. *Biologia Plantarum*. 54 (3): 423-429.
- Datta, S.K, Misra, P. and Mandal, A.K.A. (2005). *In vitro* mutagenesis a quick method for establishment of solid mutant in Chrysanthemum. *Current Science*. 88(1): 155-158.
- De Klerk, G. J, Krieken, W.V.D. and Jong, J.C.D. (1999). Review the formation of adventitious roots: new concepts, new possibilities. *In Vitro Cellular and Development Biology-Plant*. 35:189-199.

- Dhakshanamoorthy, D., Selvaraj, R., Chidambaram, A.L.A. (2011). Induced mutagenesis in *Jatropha curcas* L. using gamma rays and detection of DNA polymorphism through RAPD marker. *Comptes Rendus Biologies*: 334: 24-30.
- Dodds, J.H. and Roberts, L.W. (1985). Experiments in Plant Tissue Culture Second edition. Cambridge: Cambridge University Press.
- Faridah, H. and Shamsul, K. (2004). A guide to the common plants Ayer Hitam Forests, Selangor. Serdang: Universiti Putra Malaysia Press.
- Ficker, C.E., Smith, M.L, Siti, S., Leamanb, D.J., Irawati. C. and Arnason, J.T. (2003). Inhibition of human pathogenic fungi by members of Zingiberaceae used by the Kenyah (Indonesian Borneo). *Journal of Ethnopharmacology*. 85: 289–293.
- Fuller, J.H. and Carothers, Z.B. (1963). The Plant World: A Text in College Botany (Fourth Edition). University of Illinois: Holt, Rinehart and Winston, Inc.
- Gaspar, T., Kevers, C., Penel, C., Greppin, H., Reid, D.M. and Thorpe, T.A. (1996). Plant hormones and plant growth regulators in plant tissue culture. *In Vitro Cellular and Developmental Biology – Plant.* 32:272-289.
- Gavidia, I. and Perez-Bermudez, P. (1999). Variants of *Digitalis obscura* from irradiated shoot tips. *Euphytica*. 110: 153-159.
- Goh, M.W.K., Kumar, P.P., Lim, S.H., and Tan, H.T.W. (2005). Random amplified polymorphic DNA analysis of the moth orchids, *Phalaenopsis* (Epidendroideae: Orchidaceae). *Euphytica*. 141: 11-22.
- Habsah, M., Ali, A.M., Lajis, N.H., Sukari, M.A., Yap, Y.H., Kikuzaki, H. and Nakatani, N. (2005). Antitumor - promoting and cytotoxic constituents of *Etlingera elatior. Malaysian Journal of Medical Sciences.* 12(1): 6-12.
- Haleagrahara, N., Jackie, T., Chakravarthi, S., Rao, M., and Pasupathi, T. (2010).
 Protective effects of *Etlingera elatior* extract on lead acetate induced changes in oxidative biomarkers in bone marrow of rats. *Food and Chemical Toxicology*. 48: 2688-2694.
- Hamirah, M.N., Sani, H.B., Boyce, P.C. and Sim, S.L. (2010). Micropropagation of red ginger (*Zingiber montanum* Koenig), a medicinal plant. *Asia Pacific Journal of Molecular Biology and Biotechnology*.18(1): 127-130.
- Hazarika, B.N. (2003). Acclimatization of tissue-cultured plants. *Current Science*. 85(12): 1704-1712.
- Henderson, M.R. (1954). Malayan Wild Flowers Monocotyledons. Kuala Lumpur: The Malayan Nature Society.

- Ibrahim, R., Mondelaers, W. and Debergh, P.C. (1998). Effects of X-irradiation on adventitious bud regeneration from *in vitro* leaf explants of *Rosa hybrida*. *Plant Cell, Tissue and Organ Culture*. 54: 37–44.
- Ismail, N. A. (2009). Promising accessions of kantan (*Etlingera elatior*) with high antioxidant activity. In: Poster presentation, National Conference on New Crops and Bioresources, Seremban, Malaysia, Dec 15–17, 2009.
- Jaafar, M.F., Osman, C.P., Ismail, N.H. and Awang, K. (2007). Analysis of essential oils of leaves, stems, flowers and rhizomes of *Etlingera elatior* (Jack) R.M. *Malaysian Journal of Analytical Science*. 10: 269-273.
- Jaccard, P. (1908). Nouvelles recherché sur la distribution florale. *Bulletin de la Société vaudoise des sciences naturelles*. 44: 223-270.
- Jain, S.M. (2010). In vitro mutagenesis in banana (Musa spp.) improvement. Acta Horticulturae. 879: 605-614.
- Jain, S.M., Ahloowalia, B.S., Veilleux, R.E. (1998). Somaclonal variation in crop improvement. In: Somaclonal variation and induced mutation in crop improvement. Jain, S.M, Brar, D.S., Ahloowalia, B.S. (Ed). Kluwer Academic Publishers, Great Britain, pp. 203-218.
- Jatoi, S.A., Kikuchi, A., Yi, S.S., Naing, K.W., Yamanaka, S., Watanabe, J.A. and Watanabe, K.N. (2006). Use of rice SSR markers as RAPD markers for genetic diversity analysis in Zingiberaceae. *Breeding Science*. 56: 107-111.
- Jiang, K. and Feldman, L.J. (2003). Root meristem establishment and maintenance: The role of auxin. *Plant Growth Regulation*. 21: 432–44.
- Jimenez, M.V. (2005). Review paper: Involvement of plant hormones and plant growth regulators on in vitro somatic embryogenesis. *Plant Growth Regulation*. 47: 91–110.
- Kambaska, K.B. and Santilata, S. (2009). Effect of plant growth regulator on micropropagtion of ginger (*Zingiber officinale* Rosc.) cv Suprava and Suruchi. *Journal of Agricultural Technology*. 5(2): 271-280.
- Kavyashree, R. (2009). An efficient *in vitro* protocol for clonal multiplication of Ginger var. Varada. *Indian Journal Biotechnology*. 8: 328-331.
- Kho, P.E., Sani, H.B., Boyce, P.C. and Sim, S.L. (2007). In vitro propagation of Globba brachyanthera K.Schum. Asia Pacific Journal of Molecular Biology and Biotechnology. 18(1): 119-122.
- Kiefer, E., Heller, W. and Ernst, D. (2000). A simple and efficient protocol for isolation of functional RNA from plant tissues rich in secondary metabolites. *Plant Molecular Biology Reporter.* 18: 33-39.

- Kizhakkayil, J. and Sasikumar, B. 2010. Genetic diversity analysis of ginger (*Zingiber officinale* Rosc.) germplasm based on RAPD and ISSR markers. *Scientia Horticulturae*. 125: 73–76.
- Kovacs, E. and Keresztes, A. (2002). Effects of gamma and UV-B/C radiation on plant cells. *Micron*. 33: 199-210.
- Kuksova, V.B., Piven, N.M. and Gleba, Y.Y. (1997). Somaclonal variation and in vitro induced mutagenesis in grapevine. *Plant Cell, Tissue and Organ Culture*. 49: 17–27.
- Kume, T. (2006). In *Application of Radiation in Agriculture*, Proceedings of the International Workshop on Biotechnology in Agriculture, Nong Lam University, Ho Chi Minh City, Vietnam, Oct. 20-21, 2006.
- Larsen. K, Ibrahim H, Khaw S.H and Saw L.G. 1999. Gingers of Peninsular Malaysia and Singapore. Kota Kinabalu: Natural History Publications (Borneo).
- Leopold, A.C and Kriedemann, P.E. (1975). Plant Growth and Development Second Edition. New York.: McGraw-Hill.
- Loc, N.H., Duc, D.T., Kwon, T.H. and Yang, M.S. (2005). Micropropagation of zedoary (*Curcuma zedoaria* Roscoe) – a valuable medicinal plant. *Plant Cell, Tissue and Organ Culture*. 81: 119–122.
- Lu, G., Zhang, X., Zou, Y, Zou, Q., Xiang, X. and Cao, J. (2007). Effect of radiation on regeneration of Chinese narcissus and analysis of genetic variation with AFLP and RAPD markers. *Plant Cell, Tissue and Organ Culture*. 88: 319-327.
- Maluszynski, M., Ahloowalia, B.S. and Sigurbjrnsson, B. (1995). Application of in vivo and *in vitro* mutation techniques for crop improvement. *Euphytica*, 85: 303-315.
- Mandal, A.K.A., Chakrabarty, D. and Datta, S.K. (2000). Application of *in vitro* techniques in mutation breeding of chrysanthemum. *Plant Cell, Tissue and Organ Culture*. 60: 33-38.
- Marcsik, D., Hoult, M. Connelly, M. Lasker, E. and Ford, C. (2010). Commercialising new ornamentals. http://www.nt.gov.au/d/Primary_Industry/Content/File/horticulture/cut_flowe r/P AGES+FROM+TB311-ORNAMNETALS.pdf (accessed 19 May 2010).
- Marcsik, D and Hoult, M. (2010). Tissue culture of tropical ornamentals. http://www.nt.gov.au/d/Primary_Industry/Content/File/horticulture/cut_flowe r/P AGES+FROM+TB280-ORNAMENTALS+TC.pdf (accessed 19 May 2010).

- Martin, C., Uberhuaga, E and Perez, C. (2002). Application of RAPD markers in the characterisation of *Chrysanthemum* varieties and the assessment of somaclonal variation. *Euphytica*. 127: 247-253.
- Mat Taha, R. (2004). Kultur Tisu Tumbuhan Berbunga. Kuala Lumpur. Universiti Malaya Press.
- Mba, C., Afza, R., Jain, S.M., Gregorio, G.B and Zapata-arias, F.J. (2007). Induced mutations for enhancing salinity tolerance in rice. In: *Advances in Molecular Breeding Toward Drought and Salt Tolerant Crops*. Jenks, M.A. et al. (Ed). 413–454.
- Mineo, L. (1990). Plant tissue culture techniques. In: *Tested studies for laboratory teaching*. Proceedings of the Eleventh Workshop/Conference of the Association for Biology Laboratory Education (ABLE), Goldman, C.A. (Ed). 151-174.
- Mineo, L. (2010). Plant tissue culture techniques. http://www.ableweb.org/volumes/vol-Dis. 2010). 11/9- mineo.pdf (accesssed 19
- Murashige, T. and Skoog, F. (1962). A revised medium for rapid growth and bioassays with tobacco tissue cultures. *Physiologia Plantarum*.15: 473–497.
- Naz, S., Ilyas, S., Javad, S. and Ali, A. (2009). In vitro clonal multiplication and acclimatization of different varieties of turmeric (*Curcuma longa* L.). *Pakistan Journal of Botany*. 41(6): 2807-2816.
- Nielsen, D. and Hoult, M. (2010). Spectabilis and torch ginger breeding program. http://www.nt.gov.au/d/Primary_Industry/Content/File/horticulture/cut_flowe r/P AGES+FROM+TB268-ORNAMENTALS+PLANTS.pdf (accesssed 20 Feb. 2010).
- Nirmal Babu, K., Samsudeen, K. and Ratnambal, M.J. (1992). In vitro plant regeneration from leaf-derived callus in ginger (Zingiber officinale Rosc.) Plant Cell, Tissue and Organ Culture. 29: 71-74
- Patade, V.Y, Suprasanna, P. and Bapat, V.A. (2008). Gamma irradiation of embryogenic callus cultures and *in vitro* selection for salt tolerance in sugarcane (*Saccharum officinarum* L.). Agricultural Sciences in China. 7(9): 1147-1152.
- Patade, V.Y. and Suprasanna, P. (2008). Radiation induced *in vitro* mutagenesis for sugarcane improvement. *Sugar Technology*. 10(1): 14-19.
- Pinet-Leblay, C., Turpin, F.X. and Chevreau, E. (1992). Effect of gamma and ultraviolet irradiation on adventitious regeneration from *in vitro* cultured pear leaves. *Euphytica*. 62: 225-233.

- Poulsen. A.D. (2006). Gingers of Sarawak. Kota Kinabalu: Natural History Publications (Borneo).
- Prathanturarug, S., Angsumalee, S., Pongsiri, N., Suwacharangoon, S. and Jenjittikul, T. (2004). *In vitro* propagation of *Zingiber petiolatum* (Holttum) I. Theilade, a rare zingiberaceous plant from Thailand. *In Vitro Cellular and Developmental Biology - Plant*. 40: 317–320.
- Predieri, S and Zimmerman, R.H. (2001). Pear mutagenesis: *In vitro* treatment with gamma-rays and field selection for productivity and fruit traits. *Euphytica*. 117: 217-227.
- Puchooa, D. (2005). *In vitro* mutation breeding of Anthurium by gamma radiation. *International Journal of Agriculture and Biology*. 7 (1): 11-20.
- Rohlf, F.J. (2000). NTSYSpc 21 Numerical Taxonomy and Multivariate Analysis System. Exeter Software, Setauket, New York.
- Roopadarshini, V. (2010). High frequency shoot multiplication and callus regeneration of turmeric. *International Journal of Biotechnology and Biochemistry*. 6(5): 723–733.
- Rose, R.J., Wang, X.D., Nolan, K.E. and Rolfe, B.G. (2006). Root meristems in *Medicago truncatula* tissue culture arise from vascular-derived procambiallike cells in a process regulated by ethylene. *Journal of Experimental Botany*. 57(10): 2227–2235.
- Sahavachari, O. (2010). Cut flower production in Thailand. www.fao.org/DOCREP/005/AC452E/ac452e09.htm (accessed 19 Feb. 2011).
- Saidin, I. (2000). Sayuran tradisional ulam dan penyedap rasa. Bangi: Penerbit Universiti Kebangsaan Malaysia.
- Saingproa, B. and Kanchanapoom, K. (1997). Clonal propagation through multiple shoot formation from ginger (*Zingiber officinale* Roscoe) callus and buds. *Suranaree Journal of Science and Technology*. 4: 1-5.
- Segalen, J. (2010). The torch ginger. http://www.rosemarybasil.net/html/modules.php?op=modloadandname=New san dfile=articleandsid=829 (accessed 22 Feb. 2010).
- Sharma, T.R. and Singh, B.M. (1997). High-frequency *in vitro* multiplication of disease- free *Zingiber officinale* Rosc. *Plant Cell Reports*. 17: 68–72.
- Shen, X.S., Wan, J.Z., Luol, W.Y. and Ding, X.L. (1990). Preliminary results of using *in vitro* axillary and adventitious buds in mutation breeding of Chinese gooseberry. *Euphytica*. 49: 77-82.

- Shukla. S. K., Shukla. S., Koche. V. and Mirsha, S.K. (2007). *In vitro* propagation of tikhur (*Curcuma angustifolia* Roxb.): A starch yielding plant. *Indian Journal* of Biotecnology. 6: 274:276.
- Sirirugsa, P. (1999). Thai Zingiberaceae: Species diversity and their uses. http://www.iupac.org/symposia/proceedings/phuket97/sirirugsa.html (accessed 22 Feb.2010).
- Sivasothy, Y. (2008) Phytochemical investigation on some species from the genera *Elettariopsis* and *Etlingera*. http://eprints.usm.my/10343/1/PHYTOCHEMICAL_INVESTIGATION_ON _SOE_SPECIES_FROM_THE_GENERA_ELETTARIOPSIS_AND_ETLIN GERA.pdf (accesssed 22 Feb. 2010).
- Stanly, C. and Chan, L.K. (2007). Micropropagation of *Curcuma zedoaria* Roscoe and *Zingiber zerumbet* Smith. *Biotechnology*. 6 (4): 555-560.
- Sultana, A., Hassan, L., Ahmad, S.D., Shah, A.H., Batool, F., Islam, M.A., Rahman, R. and Moonmoon, S. (2009). *In vitro* regeneration of ginger using leaf, shoot tip and root explants. *Pakistan Journal of Botany*. 41(4): 1667-1676.
- Suprasanna, P., Rupali, C., Desai, N.S. and Bapat, V.A. (2008). Partial desiccation augments plant regeneration from irradiated embryogenic cultures of sugarcane. *Plant Cell, Tissue and Organ Culture*. 92:101–105.
- Tassi, E., Pouget, J., Petruzzelli, G. and Barbafieri, M. (2008). The effects of exogenous plant growth regulators in the phytoextraction of heavy metals. *Chemosphere*. 71: 66–73.
- Tyagi, R.K, Agrawal, A. and Yusuf, A. (2006). Conservation of Zingiber germplasm through *in vitro* rhizome formation. *Scientia Horticulturae*. 108: 210–219.
- Weaver, R. J.(1981). Plant Growth Substances In Agriculture. New Delhi: W.H. Freeman and Company.
- Wong, W. (2008). Light up your garden with a torch ginger. www.greenculturesg.com/articles/may08/may08_torchginger.pdf (2010).
- Xu, L., Najeeb, U., Raziuddin, R., Shen, W.Q., Shou, J.Y., Tang, G.X. and Zhou,
 W.J. (2009). Development of an efficient tissue culture protocol for callus formation and plant regeneration of wetland species *Juncus effusus* L. *In Vitro Cellular and Developmental Biology Plant*. 45:610–618.
- Yang, H. and Schmidt, H. (1994). Selection of a mutant from adventitious shoots formed in X ray treated cherry leaves and differentiation of standard and mutant with RAPDs. *Euphytica*. 77: 89-92.
- Yusuf, N.A., Suffian Annuar, M.M. and Khalid, N. (2011). Rapid micropropagation of *Boesenbergia rotunda* (L.) Mansf. Kulturpfl (a valuable medicinal

plant) from shoot bud explants. *African Journal of Biotechnology*. 10(7): 1194-1199.

- Zapata, E.V., Morales, G.S., Lauzardo, A.N.H., Bonfil, B.M., Tapia, G.T., Sánchez, A.D.J., Del Valle, M.V. and Aparicio, A.J. (2003). *In vitro* regeneration and acclimatization of plants of turmeric (*Curcuma longa L.*) in a hydroponic system. *Biotecnología Aplicada*. 20:25-31.
- Zheng, Y., Liu, Y., Ma, M. and Xu, K. (2008). Increasing *in vitro* microrhizome production of ginger (*Zingiber officinale* Roscoe). Acta Physiologiae Plantarum. 30: 513–519.
- Zhou, L.B., Li, W.J., Ma, S., Dong, X.C., Yu, L.X., Li, Q., Zhou, G.M. and Gao, Q.X. (2006). Effects of ion beam irradiation on adventitious shoot regeneration from *in vitro* leaf explants of *Saintpaulia ionahta*. *Nuclear Instruments and Methods in Physics Research B*. 244: 349–353.

