



UNIVERSITI PUTRA MALAYSIA

***IN VITRO PROPAGATION AND MOLECULAR CHARACTERIZATION
OF SOMACLONAL VARIATION IN PHALAENOPSIS GIGANTEA***

SAMIRA SAMARFARD

FP 2013 62



***IN VITRO PROPAGATION AND MOLECULAR
CHARACTERIZATION OF SOMACLONAL VARIATION
IN PHALAENOPSIS GIGANTEA***

By

SAMIRA SAMARFARD

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,
in Fulfilment of the Requirements for the Degree of Master of Science**

May 2013

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs, and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright© Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

**IN VITRO PROPAGATION AND MOLECULAR
CHARACTERIZATION OF SOMACLONAL VARIATION
IN *PHALAENOPSIS GIGANTEA***

By

SAMIRA SAMARFARD

May 2013

Supervisor: Associate Professor Mihdzar Abdul Kadir, PhD

Faculty: Agriculture

Phalaenopsis gigantea (Elephant's Ear orchid) is the largest species of *Phalaenopsis* genus originating from the lowland forests of Malaysia and Indonesia. Deforestation and over-collection have resulted in the extinction of this orchid. *P. gigantea* has the potential of producing beautiful hybrids and currently research on micropropagation using plant growth regulators of this orchid is ongoing. Chitosan is an environmentally friendly carbohydrate polymer and has been reported to stimulate growth of some plant species, including orchids. Multiplication was undertaken through *in vitro* inoculation of PLBs in liquid New Dogashima medium (NDM) and Vacin and Went (VW) medium supplemented with different concentrations of chitosan (0, 5, 10, 15, 20 and 25 mg/L) during 8 weeks of culture. The best response was established at 10 mg/L of chitosan supplementation in both media with the mean number of 177 and 147 PLBs formed on VW and NDM, respectively. After 6 weeks of culture in liquid media, some PLBs differentiated producing juvenile leaves and

the best response was obtained on NDM at 20 mg/L chitosan with mean number of 66 leaves. To establish an efficient treatment combination in semi solid culture for enhancing PLBs multiplication and subsequent shoot regeneration, solid NDM and VW medium supplemented with various concentrations of chitosan (0, 5, 10, 15, 20 and 25 mg/L) and thidiazuron (TDZ) (0, 0.1, 0.5 mg/L) were used. The optimum treatment for PLB multiplication in solid medium was NDM at 10 mg/L chitosan in combination with 0.1 mg/L TDZ with the mean number of 353 PLBs after 20 weeks of cultivation. NDM containing 10 mg/L chitosan and 0.1 mg/L TDZ showed a 19-fold increase in fresh weight. Whilst, the efficiency of shoot regeneration in semi solid VW was higher than NDM and the best response was observed on VW in addition with 10 mg/L chitosan and 0.5 mg/L TDZ (16), VW at 20 mg/L chitosan (15) and VW including 15 mg/L chitosan and 0.5 mg/L TDZ (13). In order to assess the genetic fidelity among initial PLBs and proliferated PLBs obtained at the end of each two week's sub-culture from the optimum treatment (10 mg/L chitosan). Eight inter-simple sequence repeat (ISSR) primers were finally selected from 10 used for initial screening. The ISSR primers generated 55 clear band classes with 0% polymorphism. The somaclonal variations among mother plant (MP) and PLBs from the sub-cultures of optimum treatment in PLB multiplication (solid NDM supplemented with 10 mg/L chitosan and 0.1 mg/L TDZ) have been estimated. The primers selected produced 67 bands with 11 of it being polymorphic. The highest number of polymorphic bands (3) was obtained using primers I65 and I2 with 27.3% polymorphism. It was found that no genetic changes occurred among mother plant and PLBs after 4, 8 and 12 weeks of culture. After 16 and 20 weeks of culture, PLBs were 95% and 80% similar to MP, respectively. In summary, the present report expressed that the addition of 10 mg/L chitosan in liquid medium could provide a

promising *in vitro* culture system to stimulate PLBs proliferation without any somaclonal variation up to 16 weeks of culture. The incorporation of 10 mg/L chitosan and 0.1 mg/L TDZ in solid NDM was also efficient for PLB proliferation. However, it resulted in 20% dissimilarity with the mother plant after 20 weeks of culture.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

PEMBIAKAN *IN VITRO* DAN PENCIRIAN MOLEKUL VARIASI SOMAKLONAL *PHALAEOPSIS GIGANTEA*

Oleh

SAMIRA SAMARFARD

Mei 2013

Pengerusi: Profesor Madya Mihdzar Abdul Kadir, PhD

Fakulti: Pertanian

Phalaenopsis gigantea (orkid telinga gajah) adalah spesies terbesar dalam genus *Phalaenopsis* yang berasal dari hutan tanah pamah di Malaysia dan Indonesia. Penebangan hutan dan pengumpulan orkid yang berleluasa telah menyebabkan spesies ini. *P. gigantea* pupus. Orkid ini berpotensi untuk menghasilkan hibrid yang cantik dan kajian pembiakan mikro keatas orkid ini sedang dijalankan dengan giat. Chitosan merupakan polimer karbohidrat mesra alam dan telah dilaporkan untuk merangsang pertumbuhan beberapa spesies tumbuhan, termasuk orkid. Peningkatan multiplikasi PLBs telah dijalankan secara *in vitro* melalui inokulasi PLBs dalam medium cecair New Dogashima (NDM) dan medium Vacin dan Went (VW) yang ditambah dengan chitosan (0, 5, 10, 15, 20 dan 25 mg/L) pada kepekatan yang berlainan sepanjang 8 minggu dikultur. Respons terbaik telah dicapai dengan penambahan chitosan sebanyak 10 mg/L dalam kedua-dua media dengan purata PLBs masing-masing sebanyak 177 dan 147 dalam VW dan NDM. Selepas 6 minggu pengkulturan dalam media cecair, beberapa PLBs telah membentuk daun juvana dan

respons terbaik telah diperolehi dalam NDM pada 20 mg/L chitosan dengan purata sebanyak 66 daun. Medium pepejal NDM dan VW yang ditambah dengan pelbagai kepekatan chitosan (0, 5, 10, 15, 20 dan 25 mg/L) dan thidiazuron (TDZ) (0, 0.1, 0.5 mg/L) telah digunakan untuk mendapatkan satu rawatan kombinasi yang cekap dalam kultur pepejal yang berupaya meningkatkan multiplikasi PLBs dan penjanaan semula pucuk. Rawatan optimum bagi multiplikasi PLBs dalam medium pepejal adalah NDM dengan kombinasi 10 mg/L chitosan dan 0.1 mg/L TDZ dengan purata 353 PLBs selepas 20 minggu pengkulturan. Medium NDM pepejal yang mengandungi 10 mg/L chitosan dan 0.1 mg/L TDZ telah menghasilkan peningkatan 19 kali ganda dalam berat segar PLB. Sementara itu, keberkesanan penjanaan semula pucuk dalam VW pepejal adalah lebih tinggi daripada NDM dan respons terbaik telah diperhatikan pada VW dengan penambahan 10 mg/L chitosan dan 0.5 mg/L TDZ (16), VW dengan penambahan 20 mg/L chitosan (15) dan VW dengan penambahan 15 mg/L chitosan dan 0.5 mg/L TDZ (13). Lapan inter-simple sequence repeat (ISSR) telah dipilih daripada sepuluh yang digunakan dalam penyaringan awal dan penentuan ketulenan genetik diantara PLBs induk berbanding dengan PLBs yang diperolehi pada akhir setiap dua minggu sub-kultur daripada rawatan optimum (10 mg/L chitosan) sepanjang pengkulturan. Primer ISSR telah menjana 55 jalur yang jelas dengan polimorfisma sebanyak 0%. Variasi somaklonal dalam kalangan tumbuhan induk (MP) dan PLBs sekunder daripada sub-kultur rawatan optimum multiplikasi PLB yang berbeza (NDM pepejal yang ditambah dengan 10 mg/L chitosan dan 0.1 mg/L TDZ) telah dianggarkan. Lapan primer ISSR menghasilkan 67 jalur dengan 11 jalur polimorfik. Bilangan jalur polimorfik terbanyak (3) telah diperolehi menggunakan primers I65 dan I2 dengan polimorfisma sebanyak 27.3%. Perubahan genetik tidak berlaku dalam kalangan tumbuhan induk dan PLBs sekunder selepas 4,

8 dan 12 minggu dikultur. Terdapat sebanyak 95% dan 80%, persamaan di antara PLBs yang dimultiplikasi dengan PLB induk selepas 16 dan 20 minggu dikultur, masing-masing. Secara kesimpulan, laporan ini menyatakan bahawa penambahan 10 mg/L chitosan dalam medium cecair boleh merangsang proliferasi PLBs kultur *in vitro* tanpa menyebabkan variasi somaklonal selama 16 minggu. Penggabungan 10 mg/L chitosan dan 0.1 mg/L TDZ dalam NDM pepejal juga berkesan untuk percambahan PLBs. Walau bagaimanapun, ia menyebabkan perbezaan sebanyak 20% dengan tumbuhan induk selepas 20 minggu dikultur.

ACKNOWLEDGEMENTS

First and foremost I would like to express my gratitude to THE ALMIGHTY ALLAH the most Benevolent, Merciful and Compassionate, for giving me the utmost strength, patience, HIS grace and mercy to have this work completed.

I am very grateful to my supervisor Associate Professor Dr. Mihdzar Abdul Kadir from Department of Agriculture Technology, Faculty of Agriculture and member of my supervisory committee, Associate Professor Dr. Saleh Kadzimin from Department of Crop Science, Faculty of Agriculture for valuable suggestions, guidance and for the support given throughout this research. I also would like to extend my gratitude to the internal and external examiners who evaluated my thesis thoroughly and gave constructive comments to improve the quality of my thesis.

I would like to extend my heartiest appreciation and my deepest thanks to my dear father Hossein Samarfard and my beloved mother Parvaneh Ziaratian for their encouragement, kindness, moral as well as financial support that sustained me throughout my study and life.

Profound gratitude is also extended to Universiti Putra Malaysia (UPM), especially Faculty of Agriculture and Department of Agriculture Technology for the facilities available to conduct experiments and to acquire knowledge.

I would like to convey my appreciation to all my labmates from Agrotechnology Laboratory especially to Sang Mi Eum for sharing her knowledge, friendship throughout this research. Finally, my special thanks to my dear friends Dr Hossein

Kamaladini, Mahmoud Danaie, Fatemeh Haddadi and Naghmeh Nejat for sharing their knowledge and moral support.



This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of requirement for degree of Master of Science. The members of the Supervisory Committee were as follows:

Mihdzar Abdul Kadir, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Chairman)

Saleh Kadzimin, PhD

Associate Professor
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

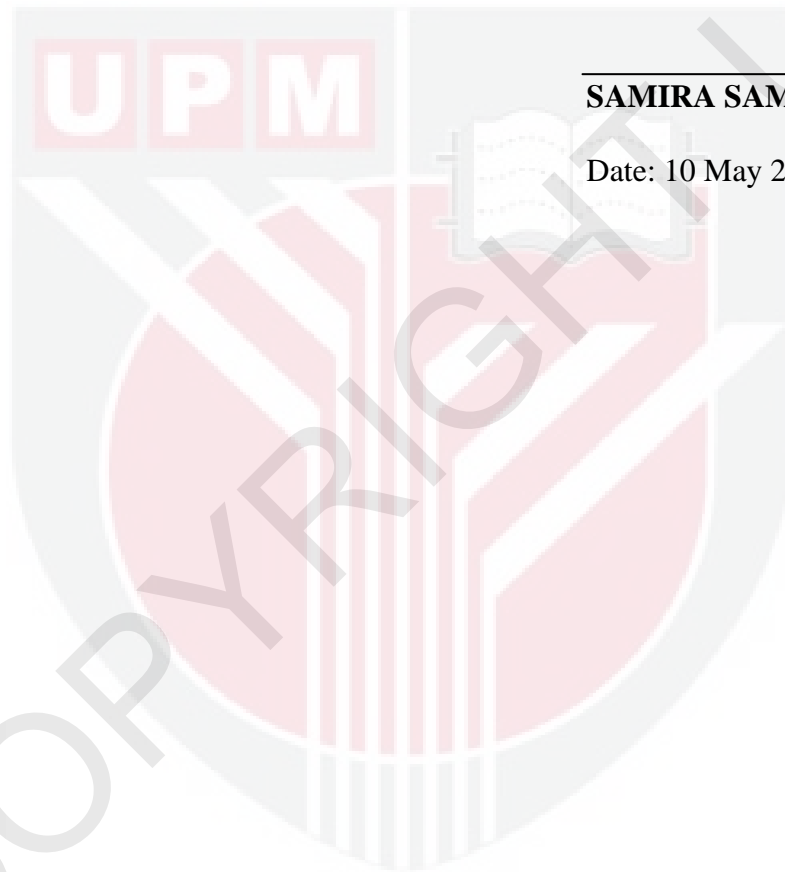
BUJANG BIN KIM HUAT, PhD

Professor and Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

DECLARATION

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously, and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institution.



SAMIRA SAMARFARD

Date: 10 May 2013

TABLE OF CONTENTS

| | Page |
|--|----------|
| ABSTRACT | ii |
| ABSTRAK | v |
| ACKNOWLEDGEMENTS | viii |
| APPROVAL | x |
| DECLARATION | xii |
| LIST OF TABLES | xvi |
| LIST OF FIGURES | xix |
| LIST OF ABBREVIATIONS | xix |
| CHAPTER | |
| 1 INTRODUCTION | 1 |
| 2 LITERATURE REVIEW | 5 |
| 2.1 <i>Orchidaceae</i> | 5 |
| 2.2 Genus <i>Phalaenopsis</i> | 6 |
| 2.3 <i>Phalaenopsis gigantea</i> (Elephant's Ear orchid) | 7 |
| 2.4 Orchid Propagation | 8 |
| 2.4.1 Sexual | 8 |
| 2.4.2 Asexual | 9 |
| 2.5 <i>In Vitro</i> Propagation of <i>Phalaenopsis</i> | 9 |
| 2.6 Multiplication of Protocorm Like-Bodies (PLBs) | 11 |
| 2.7 Culture Medium | 12 |
| 2.8 Plant Growth Regulators | 13 |
| 2.9 Chitosan as Plant Growth Stimulator | 14 |
| 2.9.1 Chitosan Composition | 14 |
| 2.9.2 Properties and Solubility of Chitosan | 15 |
| 2.9.3 The Application of Chitosan in Agriculture | 15 |
| 2.9.4 The Effects of Chitosan on Orchids | 17 |
| 2.10 Somaclonal variation | 18 |
| 2.10.1 Definition and Importance of Somaclonal Variation | 18 |
| 2.10.2 The Sources of Somaclonal Variation | 19 |
| 2.10.3 Somaclonal Variation in <i>Phalaenopsis</i> Orchids | 21 |
| 2.11 Detection of Somaclonal Variation | 22 |
| 2.11.1 Morphological Detection | 22 |
| 2.11.2 Physiological and Biochemical detection | 23 |
| 2.11.3 Cytological Methods | 24 |
| 2.11.4 Proteins and Isozymes | 25 |
| 2.11.5 Molecular Detection | 26 |
| 2.12 Advantages of Inter Simple Sequence Repeat (ISSR) | 27 |
| 2.13 ISSR Studies in Orchids | 28 |

| | | |
|----------|--|-----------|
| 3 | <i>IN-VITRO</i> PROLIFERATION OF <i>PHALAENOPSIS GIGANTEA</i> PROTOCOL-LIKE BODIES (PLBs) BY CHITOSAN AND THIDIAZURON SUPPLEMENTATION | 29 |
| 3.1 | Introduction | 29 |
| 3.2 | Materials and Methods | 31 |
| 3.2.1 | PLB Proliferation in Liquid Media | 31 |
| 3.2.1.1 | PLB Induction from Leaf Segment | 31 |
| 3.2.1.2 | Chitosan Preparation | 32 |
| 3.2.1.3 | Media Preparation | 32 |
| 3.2.1.4 | Proliferation Condition | 33 |
| 3.2.1.5 | Parameters Recorded | 33 |
| 3.2.1.6 | Experimental Design and Statistical Analysis | 33 |
| 3.2.2 | PLB Proliferation and Shoot Formation on Solid Media | 34 |
| 3.2.2.1 | Explants and Media Preparation | 34 |
| 3.2.2.2 | Treatment Combination | 34 |
| 3.2.2.3 | Culture Conditions | 36 |
| 3.2.2.4 | Parameters Observed | 36 |
| 3.2.2.5 | Experimental Design and Statistical Analysis | 36 |
| 3.3 | Results and Discussion | 37 |
| 3.3.1 | Effects of Chitosan and Liquid Media on PLBs Proliferation | 37 |
| 3.3.2 | Effect of Chitosan on Differentiation | 42 |
| 3.3.3 | Effect of Chitosan on the Fresh Weight of PLBs | 44 |
| 3.3.4 | Effect of Chitosan and TDZ Combination on PLB Proliferation | 46 |
| 3.3.5 | Effect of Chitosan and TDZ Combination on Fresh Weight of PLBs | 51 |
| 3.3.6 | Effect of Chitosan and TDZ Combination on shoot Formation | 54 |
| 4 | DETECTION OF SOMACLONAL VARIATION BY INTER SIMPLE SEQUENCE REPEAT DURING PROLIFERATION OF <i>PHALAENOPSIS GIGANTEA</i> | 59 |
| 4.1 | Introduction | 59 |
| 4.2 | Materials and Methods | 61 |
| 4.2.1 | Detection of Somaclonal Variation | 61 |
| 4.2.2 | Extraction of Genomic DNA | 62 |
| 4.2.3 | Screening of ISSR Primers | 63 |
| 4.2.4 | PCR Amplification and Gel Scoring | 64 |
| 4.2.5 | Analysis of ISSR Data | 65 |
| 4.3 | Results and Discussion | 66 |
| 4.3.1 | Assessment of Genetic Stability of PLBs from Optimum Liquid Culture | 66 |

| | | |
|----------|--|------------|
| 4.3.2 | Assessment of Genetic Stability of PLBs from Optimum Solid Culture | 70 |
| 4.3.3 | Genetic Similarity and Multivariate Analysis | 73 |
| 5 | GENERAL CONCLUSION AND RECOMMENDATION FOR FUTURE RESEARCH | 78 |
| | REFERENCES | 81 |
| | APPENDICES | 98 |
| | BIODATA OF STUDENT | 105 |



LIST OF TABLES

| Table | | Page |
|-------|--|------|
| 3.1 | Different concentrations and combinations of chitosan and TDZ | 35 |
| 3.2 | Mean number of leaves after 8 weeks of culture | 43 |
| 3.3 | Effect of chitosan and TDZ combination (mg/L) on total fresh weight of PLBs after 20 weeks of cultivation in semi solid NDM and VW | 52 |
| 4.1 | List of 10 ISSR primers used for initial screening and amplification of DNA | 63 |
| 4.2 | Sequences of screened primers used in ISSR analysis for genetic stability of the PLBs | 68 |
| 4.3 | Total number and size range of amplified ISSR bands and number of polymorphic fragments induced by MP and PLBs achieved from different sub-cultures of optimum treatment | 71 |
| 4.4 | Summary of ISSR amplified products from six samples of <i>P. gigantea</i> PLBs | 72 |
| 4.5 | Similarity matrix of the somaclonal variation between mother plant and PLBs from different subcultures of optimum treatment | 74 |

LIST OF FIGURES

| Figure | | Page |
|--------|--|------|
| 3.1 | PLB Induction from leaf segment of donor plant after 4 weeks of culture (bar=2mm) | 38 |
| 3.2 | (a) Swollen protocorm after 2 weeks of culture (bar = 4mm), (b) PLB multiplication after 3 weeks of culture (bar = 20 mm) | 38 |
| 3.3 | Effect of media type and chitosan concentrations (mg/L) on mean number of PLBs obtained after eight weeks of culture | 40 |
| 3.4 | PLB and leaf formation (a) PLB multiplication after 5 weeks of culture in NDM at 20 mg/L chitosan (bar = 20 mm). (b) The leaf formation from mature PLBs after 8 weeks of culture (bar = 20 mm) | 43 |
| 3.5 | Effect of chitosan concentrations (mg/L) on total fresh weight of PLB's after eight weeks of culture in liquid NDM and VW medium | 45 |
| 3.6 | PLBs obtained after 20 weeks of in vitro culture in solid NDM supplemented with 10 mg/L chitosan and 0.1 mg/L of TDZ (bar = 20mm) | 47 |
| 3.7 | Effect of chitosan and TDZ combination (mg/L) on mean number of PLBs after 20 weeks of culture in solid NDM and VW medium | 48 |
| 3.8 | Shoot formation after 20 weeks of culture in VW medium with (a) 10 mg/L chitosan and 0.5 mg/L TDZ and (b) 20 mg/L chitosan (bar = 20mm). | 55 |
| 3.9 | Effect of chitosan and TDZ combination (mg/L) on mean number of shoots after 20 weeks of culture in solid NDM and VW medium | 56 |
| 4.1 | ISSR banding pattern in both multiplied PLBs obtained with optimal chitosan and mother plants of <i>Phalaenopsis gigantea</i> (lane MP is mother plant and lanes 1-4 are multiplied PLBs obtained after subcultures 1-4 with optimal chitosan) | 68 |
| 4.2 | ISSR banding pattern in mother plants and multiplied PLBs in NDM at 10 mg/L chitosan and 0.1 mg/L TDZ (lane MP is mother plant, lanes 1-5 are multiplied PLBs obtained after subcultures 1-5 with optimal chitosan) | 72 |

- 4.3 Dendrogram exhibiting coefficient similarities among 6 samples from different subcultures of *P. gigantea* PLBs by the UPGMA cluster analysis (NTSYS) of the ISSR profiles derived from 8 primers using Jaccard's similarity coefficient (MP corresponding to Mother Plant and S corresponding to subculture 1-5) 75



LIST OF ABBREVIATIONS

| | |
|---------------|---|
| ABA | Abscissic Acid |
| ANOVA | Analysis of Variance |
| BAP | 6-Benzylaminopurine |
| Bp | Base pair |
| CTAB | Cetylmethylammonium bromide |
| CW | Coconut Water |
| DNA | Deoxyribonucleic Acid |
| DNMRT | Duncan New Multiple Range Test |
| EDTA | Ethylenediaminetetra-acetic acid |
| ISSR | Inter Simple Sequence Repeat |
| M | Molar |
| μM | Micromolar |
| Mm | mili Molar |
| NAA | Naphthalene Acetic Acid |
| NaOH | Sodium Hydroxide |
| NDM | New Dogashima Medium |
| NTSYS | Numerical Taxonomy and Multivariate Analysis system |
| PAL | Phenylalanine Ammenio-Lyase |
| PCR | Polymerase Chain Reaction |
| PLBs | Protocorom-like-bodies |
| RAPD | Random Amplified Polymorphic DNA |
| RCBD | Randomized Complete Block Design |

| | |
|-------|--|
| RFLP | Restriction Fragment Length Polymorphism |
| RNA | Ribonucleic Acid |
| RNase | Ribonuclease |
| SAMPL | Selective Amplified Microsatellite Polymorphism Length |
| SSR | Simple Sequence Repeat |
| TDZ | Thidiazuron |
| UPGMA | Unweighted Pair Group Method with Arithmetic Mean |
| VW | Vacin and Went (tissue culture medium) |

CHAPTER 1

INTRODUCTION

The family *Orchidaceae* includes about 7% of all angiosperms and is regarded as one of the largest families of this group. It constitutes more than 25,000 recognized species, propagated around the world exhibiting highest genetic range in tropical areas (Thammasiri, 2002). Most orchid species are dispersed mainly in the Neotropical zone and the majority are epiphytes with many being rupicolous, terrestrial and propagated in marshy regions (Pansarin and Pansarin, 2011). Orchids are esteemed as ornamentals and cut flowers because of their marvelous beauty and long lasting flowers. Currently their production stand as multi-million dollar floral industries in several countries like Singapore, Australia, Thailand, Malaysia and several others (Chugh *et al.*, 2009).

The genus *Phalaenopsis* is one of the most important epiphytic monopodial orchid, known as the moth orchid due to similarity of the flowers to night active moth butterflies (Nash, 2003), valued for its attractive cut flowers and potted plants (Chai *et al.*, 2002), a long shelf-life and large diversities (Zheng *et al.*, 2008). Commercial production of potted *Phalaenopsis* exists in Taiwan, China, Netherlands, Germany, United States, and Japan (Griesbach, 2002). About 85–90% of orchid sales among all commercial orchids in the USA are *Phalaenopsis* because of their easiness of arrangement to meet specific market dates, high wholesale value, and long shelf life (Nash, 2003).

Phalaenopsis gigantea (Elephant's Ear orchid) is the largest species in the genus, occurring in the lowland forests of Sabah in Malaysia; but deforestation and over-collection have resulted in near extinction of this species (Rodrigues and Kumar, 2009). The species has the capability of producing attractive hybrids (Niknejad *et al.*, 2011). The species name comes from its vast leaf size and propagation is typically performed through the configuration of new buds induced at the bases of mature plants. However, the procedure is much unfrequented due to low number of new buds initiated by a mature plant (Shu-guo, 2008).

Tissue culture techniques have been used not only for rapid propagation on a large scale for orchids, but also for conservation purposes (Murdad *et al.*, 2006). Various explants and combination and concentrations of plant growth regulators have been significant factors for *in vitro* propagation of orchids. However, enhancement of multiplication, total yield and successive maturity into plantlets without mutation rates are most preferred (Pornpienpakdee *et al.*, 2010). Application of some growth stimulators enhances the rate of growth during *in vitro* multiplication. Chitosan is a cationic polymer and N-deacetylated product derivative of chitin which is present in shells of crustaceans and cell wall of fungi. This component is an environmentally friendly carbohydrate polymer and has been reported to stimulate the growth of some plant species, including orchids (Nge *et al.*, 2006). Similar to some other orchids, *Phalaenopsis gigantea* is inherently difficult to propagate and the supplementation of growth stimulants like chitosan and thidiazuron (TDZ) in tissue culture medium can provide an alternative means for multiplication of protocorm-like bodies (PLBs). Propagation through PLB formation is preferred by commercial growers of orchids due to the large number of PLBs that can be obtained within a relatively short period

of time. There are two rate-limiting steps in mass production of *Phalaenopsis* PLBs: growth and multiplication. These steps require precise cultural conditions including medium composition and growth regulation, and it is necessary to reduce the time for growth and multiplication of PLBs (Park *et al.*, 1996).

The process of sub-culturing is an important stage in order to produce more PLBs and subsequent plantlet regeneration. However, excessive sub-culturing can result in unpredictable mutations. Somaclonal variations can occur due to several reasons such as types of tissue culture media, plant growth regulators and their concentrations, explant types and number of sub-culturing cycles (Reuveni *et al.*, 1986). The use of some synthetic PGRs at sub and supra-optimal concentrations was reported to be effective in inducing somaclonal variations in some crops (Martins *et al.*, 2004). The occurrence of somaclonal variation is an important issue in plant tissue culture especially in ornamental plants because it can result in the production of undesirable off-types. The assessment of somaclonal variation by using molecular markers like Inter Simple Sequence Repeat (ISSR) will give indication on the limit to the number of sub cultures and duration of cultures.

The present study was conducted to optimize PLB multiplication of *Phalaenopsis gigantea* with chitosan and TDZ supplementation in different growth media. The study was also conducted to detect the genetic stability between mother plant and PLBs obtained from different sub-cultures of the optimal treatments. Inter Simple Sequence Repeats (ISSR) molecular technique has been applied in order to verify the genetic fidelity of PLBs produced during the process of multiplication in *Phalaenopsis gigantea*. Therefore, the study focuses on the following objectives:

- (1) To determine the optimum chitosan concentration for promoting PLB proliferation in two types of liquid medium.
- (2) To determine the optimum chitosan and TDZ combination for enhancing PLBs proliferation on two types of solid medium.
- (3) To assess genetic stability of PLBs produced from different subcultures of optimum treatment by the Inter Simple Sequence Repeats (ISSR).



REFERENCES

- Aktar, S., Nasiruddin, K. M. and Hossain, K. (2008). Effects of Different Media and Organic Additives Interaction on *In Vitro* Regeneration of *Dendrobium* Orchid. *Agriculture and Rural Development* 6, 69-74.
- Al-Zahim, M.A., Ford-Lloyd, B.V. and Newbury, H.J. (1999). Detection of somaclonal variation in garlic (*Allium sativum* L.) using RAPD and cytological analysis. *Plant Cell Reports* 18: 473-477.
- Albani, M.C. and Wilkinson, M. J. (1998). Inter simple sequence repeat polymerase chain reaction for the detection of somaclonal variation. *Plant Breeding* 117: 573-575.
- Araditti, J. and Ernest, R. (1993). *Micropropagation of Orchid*. Wiley Publisher, New York. JohnWiley and Son, pp. 682.
- Araújo, L.G., Prabhu, A.S., Filippi, M.C. and Chaves, L.J. (2001). RAPD analysis of blast resistant somaclones from upland rice cultivar IAC 47 for genetic divergence. *Plant Cell Tissue and Organ Culture*, 67: 165-172.
- Arditti, J. (2008). *Micropropagation of Orchids*. 2nd edn. Blackwell Publishing Ltd, Maiden, MA, USA.
- Arditti, J. and Robert, E. (1993). *Micropropagation of Orchids*. John Wiley and Sons, New York, pp. 467-520.
- Azian, E., Zaki, A.R.M., Mohamed, M.T.M. and Kamuruzaman, S. (2004). The use of chitosan on vase life of cut chrysanthemum (*Dendranthema morifolium* Ramat). *Proceedings of APEC Symposium on quality management in postharvest system*, Bangkok, Thailand, Aug 3-5, 2004. p 403.
- Bairu, M. W., Fennell, C. W. and van Staden, J. (2006). The effect of plant growth regulators on somaclonal variation in Cavendish banana (*Musa* AAA cv. 'Zelig'). *Scientia Horticulturae* 108:347–351.
- Baker, K.M., Mathes, M.L. and Wallace, B.J. (1987). Germination of *Pantheva* and *Cattleya* seeds and development of *Phalaenopsis* protocorms *Lindleyana* 2(2) 7783
- Batchelor, S.R. (1982). *Phalaenopsis*. Part 1. *American Orchid Society Bulletin* 51:1267-1275.

- Ben-Shalom, N., Ardi, R., Pinto, R., Aki, C. and Fallik, E. (2003). Controlling gray mould caused by *Botrytis cinerea* in cucumber plants by means of chitosan. *Crop Protection* 22 285-290
- Bhatia, R., Singh, K.P., Jhang, T. and Sharma, T.R. (2009). Assessment of clonal fidelity of micropropagated gerbera plants by ISSR markers. *Scientia Horticulturae* 119(2): 208-211.
- Bhattacharya, S., Dey, T., Bandopadhyay, T.K. and Ghosh, P.D. (2008). Genetic polymorphism analysis of somatic embryo-derived plantlets of *Cymbopogon flexuosus* through RAPD assay. *Plant Biotechnology Reports* 2: 245-252.
- Bittelli, M., Flury, M., Campbell, G. and Nichils, E. J. (2001). Reduction of transpiration through foliar application of chitosan. *Agricultural and Forest Meteorology* 107: 167-175.
- Bottcher, I., Zoglauer, K. and Goring, H. (1988). Induction and reversion of vitrification of plants cultured *in vitro*. *Physiologia Plantarum* 72: 560-654.
- Bouharmont, J. (1994). Application of somaclonal variation and *in vitro* selection to plant improvement. *Acta Horticulturae* 355: 213-218.
- Brar, D.S. and Jain, S.M. (1998). Somaclonal variation: mechanism and applications in crop improvement. In B.S. Ahloowalia (ed) *Somaclonal variation and induced mutations in crop improvement* (pp. 15-37). Dordrecht: Kluwer Academic Publishers.
- Chai, M.L., Xu, C.J., Senthil, K.K., Kim, J.Y. and Kim, D.H. (2002). Stable transformation of protocorm-like bodies in *Phalaenopsis* orchid mediated by *Agrobacterium tumefaciens*. *Scientia Horticulturae*, 96(1-4): 213-224.
- Chen, I. (2004). Management and utilization of intelligent right of orchid *Phalaenopsis*. *Monthly Report of Taiwan Floriculture*, 202: 46-48.
- Chen, J. T. and Chang, W. C. (2006). Direct somatic embryogenesis and plant regeneration from leaf explants of *Phalaenopsis amabilis*. *Biologia Plantarum*, 50: 169-173.
- Chen, J.T. and Chang, W.C. (2004). Induction of repetitive embryogenesis from seed-derived protocorms of *Phalaenopsis amabilis* var. formosa shimadzu. *In Vitro Cellular & Developmental Biology Plant* 40: 290-293.
- Chen, L.R., Chen, J.T. and Chang, W.C. (2002). Efficient production of protocorm-like bodies and plant regeneration from flower stalk explants of the sympodial orchid *Epidendrum radicans*. *In Vitro Cellular & Developmental Biology Plant* 38: 441-445.

- Chen, W.H., Chen, T.M., Fu, Y.M., Hsieh, R.M. and Chen, W.S. (1998). Studies on somaclonal variation in *Phalaenopsis*. *Plant Cell Reports* 18:7-13.
- Chen, J.T., Chang, C. and Chang, W.C. (1999). Direct somatic embryogenesis on leaf explants of *Oncidium Gower Ramsey* and subsequent plant regeneration. *Plant Cell Reports*, 19: 143-149.
- Chen, Y.C., Chang, C. and Chang, W.C. (2000). A reliable protocol for plant regeneration from callus culture of *Phalaenopsis*. *In Vitro Cellular & Developmental Biology Plant*, 36: 420-423.
- Chen, Y. and Piluek, C. (1995). Effects of thidiazuron and N6- benzylaminopurine on shoot regeneration of *Phalaenopsis*. *Plant Growth Regulation*, 16: 99-101.
- Chugh, S., Guha, S., and Rao, I. U. (2009). Micropropagation of orchids: A review on the potential of different explants. *Scientia Horticulturae*, 122(4): 507-520.
- Clements, M. A. (1988). Orchid mycorrhizal associations. *Lindeleyana* 3, 73-86.
- Cloutier, S. and Landry, B. (1994). Molecular markers applied to plant tissue culture. *In Vitro Cellular & Developmental Biology plant* 30: 32-39.
- Cooper, C., Crowther, T., Smith, B.M., Isaac, S. and Collin, H.A. (2006). Assessment of the response of carrot somaclones to *Pythium violae*, causal agent of cavity spot. *Plant Pathology* 55:427-432.
- Cote, F. X., Sandoval, J. A., Marie, P. and Auboiron, E. (1993). Variations in micropropagated bananas and plantains. *Literature survey. Fruits* 48:15-22.
- D'Amato, F. (1977). Cytogenetics of differentiation in tissue and cell culture. In: Reinert J, Bajaj YPS (eds) *Applied and fundamental aspects of plant cell, tissue and organ culture* (pp 343-464). New York: Springer.
- Damasco, O.P., Smith, M.K., Godwin, I.D., Adkins, S.W., Smillie, R.M. and Hetherington, S.E. (1997). Micropropagated dwarf off-type Cavendish bananas (*Musa* spp., AAA) show improved tolerance to suboptimal temperatures. *Australian Journal of Agricultural Research* 48: 377-384.
- Decruse, S.W., Gangaprasad, A., Seeni, S., and Menon, VS. (2003). Micropropagation and ecorestoration of *Vanda spathulata*, an exquisite orchid. *Plant Cell, Tissue and Organ Culture*. 72:199-202
- Devarumath, R.M., Nandy, S., Rani, V., Marimuthu, S., Muraleedharan, N. and Raina, S.N. (2002). RAPD, ISSR and RFLP fingerprints as useful markers to evaluate genetic integrity of micropropagated plants of three diploid and triploid elite tea clones representing *Camellia sinensis* (China type) and *C. assamica* ssp. *Assamica* (Assam-India type). *Plant Cell Reports* 21: 166-173.

- Devlieghere, F., Vermeulen, A. and Debevere, J. (2004). Chitosan: antimicrobial activity, interactions with food components and applicability as a coating on fruit and vegetables. *Food Microbiology* 26(6): 703-714.
- Doares, S.H., Syrovets, T., Weiler, E.W. and Ryan, C.A. (1995). Oligogalacturonides and chitosan activate plant defensive genes through the octadecanoid pathway. *Proceedings of the National Academy of Science USA* 92: 4095-4098.
- Dolezel, J. (1997). Application of flow cytometry for the study of plant genomes. *Journal of Applied Genetic* 38: 285-302.
- Dolezel, J., Valarik, M., Vrana, J., Lysak, M. A., Hr̃ibova, E., Bartos, J., Gasmanova, N., Dolezelova, M., S afar, J. and S imkova, H. (2004). Molecular cytogenetics and cytometry of bananas (*Musa* spp.). In S.M. Jain and R. Swennen (eds) *Banana improvement: cellular, molecular biology, and induced mutations* (pp 229-244). Enfield: Science Publishers, Inc.
- Duncan, D.B. (1995). Multiple range and multiple F test-Biometrics 11:1-42
- El Ghaouth, A. (1994). Manipulation of defense systems with elicitors to control postharvest diseases. In C.L. Wilson and M.E. Wisniewski (eds) *Biological Control of Postharvest Diseases: Theory and Practice* (pp 153-167). Boca Raton: CRC.
- Ernst, R. (1994). Effect of thidiazuron on *in vitro* propagation of *Phalaenopsis* and *Doritaenopsis* (*Orchidaceae*). *Plant Cell, Tissue and Organ Culture*. 39, 273-275
- Goh, C.J. and Yang, A.L. (1978). Effects of growth regulators and decapitation on flowering of *Dendrobium* orchid hybrids. *Plant Science Letters*. 12: 287-292
- Gostimsky, S.A., Kokaeva, Z.G. and Konovalov, F.A. (2005). Studying plant genome variation using molecular markers. *Russian Journal of Genetics* 41:378-388.
- Graebe, J. E. (2003). Gibberellin biosynthesis and control. *Plant Physiology*, 38: 419-465.
- Griesbach, R. J. (2002). Development of *Phalaenopsis* Orchids for the Mass-Market. In: J. Janick and A. Whipkey (eds.). *Trends in New Crops and New Uses*.ASHS press, Alexandria, VA.
- Hadley, G. (1982). Orchid Biology. In J. Araditti (ed) *Reviews and prespectives II"* (pp. 82-118). New York: Cornell University Press.
- Hadwiger, L.A., Klosterman, S.J. and Choi, J.J. (2002). The mode of action of chitosan and its oligomers in inducing pant promoters and developing disease resistance in plants. *Advances in Chitin Science* 5: 452-457.

- Hartmann, C., Henry, Y., Buyser, J., Aubry, C. and Rode, A. (1989). Identification of new mitochondrial genome organizations in wheat plants regenerated from somatic tissue cultures. *Theoretical and Applied Genetics* 77:169-175.
- Hong, P.I., Chen, J.T. and Chang, W.C. (2009). Shoot development and plant regeneration from protocorm-like bodies of *Zygopetalum mackayi*. *In Vitro Cellular & Developmental Biology Plant* 46(3): 306-311.
- Hong, P.I., Chen, J.T. and Chang, W.C. (2010). Shoot development and plant regeneration from protocorm-like bodies of *Zygopetalum mackayi*. *In Vitro Cellular & Developmental Biology Plant*, 46, 306-311.
- Hossain, M.M., M, S., Teixeira da Silva, J.A. and P, P. (2010). Seed germination and tissue culture of *Cymbidium giganteum* Wall. ex Lindl. *Scientia Horticulture* 123: 479-487.
- Huan, L.V.T., Takamura, T. and Tanaka, M. (2004). Callus formation and plant regeneration from callus through somatic embryo structures in *Cymbidium* orchid. *Plant Science* 166:1443-1449.
- Hunter, R.L. and Merkert, C.L. (1957). Histochemical demonstration of enzymes separated by zone electrophoresis in starch gels. *Science* 125: 1294-1295.
- Ishii, Y., Takamura, T., Goi, M. and Tanaka, M. (1998). Callus induction and somatic embryogenesis of *Phalaenopsis*. *Plant Cell Reports*, 17: 446-450.
- Islam, M.O., Rahman, A.R.M.M., Matsui, S. and Prodhan, A.K.M.A. (2003). Effects of complex organic extracts on callus growth and PLB regeneration through embryogenesis in the *Doritaenopsis* orchid. *Japan Agricultural Research Quarterly* 37: 229-235.
- Israeli, Y., Lahav, E. and Reuveni, O. (1995). *In vitro* culture of bananas. In S. Gowen (ed) *Bananas and plantians* (pp. 147-178). London: Chapman and Hall.
- Israeli, Y., Reuveni, O. and Lahav, E. (1991). Qualitative aspects of somaclonal variations in banana propagated by *in vitro* techniques. *Scientia Horticulturae* 48: 71-88
- Jain R.K. (1997). Effect of some factors on plant regeneration from indica rice cells and protoplasts. *Indian journal of experimental biology*, 35:323-331
- Jain, S. (2001). Tissue culture-derived variation in crop improvement. *Euphytica* 118: 153-166.
- Jarret, R.L. and Gawel, N. (1995). Molecular markers, genetic diversity and systematics in *Musa*. In Gowen S (ed) *Bananas and plantians* (pp. 66-83). London: Chapman and Hall.

- Jarret, R.L. and Litz, R.E. (1986). Isozymes as genetic markers in bananas and plantains. *Euphytica*, 35: 539-549.
- Jiang, Y.M. and Li, Y.B. (2001). Effects of chitosan coating on storage life and quality of longan fruit. *Food Chemistry* 73: 139-143.
- Joshi, P. and Dhawan, V. (2007). Assessment of genetic fidelity of micropropagated *Swertia chirayita* plantlets by ISSR marker assay. *Biologia Plantarum*, 51, 22-26.
- Kaeppeler, S.M., Kaeppeler, H.F. and Rhee, Y. (2000). Epigenetic aspects of somaclonal variation in plants. *Plant Molecular Biology* 43:179-188.
- Kalpona, S., Sathyanarayana, B.N. and Sachdev, K. (2000). Effect of coconut water and banana pulp on *in vitro* culture of *Dendrobium*. *Journal of Plant Biology* 29(2): 209- 210.
- Karp, A. (1995). Somaclonal variation as a tool for crop improvement. *Euphytica* 85: 295-302.
- Khan, W. (2003). Signal compounds involved with plant perception and response to microbes alter plant physiological activities and growth of crop plants. PhD Thesis, McGill University, Canada.
- Khanna, H. K. and Raina, S. K. (1998). Genotype X culture media interaction effects on regeneration response of three indica rice cultivars *Plant Cell, Tissue and Organ Culture* 52(3) 145-153
- Khoddamzadeh, A., Sinniah, U., Kadir, M., Kadzimin, S., Mahmood, M. and Sreeramanan, S. (2011). *In vitro* induction and proliferation of protocorm-like bodies (PLBs) from leaf segments of *Phalaenopsis bellina* (Rchb.f.) Christenson. *Plant Growth Regulation* 65(2): 381-387.
- Kiefer, E., Heller, W. and Ernest, D. (2000). A simple and efficient protocol for isolation of functional rna from plant tissues rich in secondary metabolites. *Plant Molecular Biology Reporter* 18: 33-39.
- Kishor, R. and Devi, H. (2009). Induction of multiple shoots in a monopodial orchid hybrid (*Aerides vandarum* Reichb.f × *Vanda stangeana* Reichb.f) using thidiazuron and analysis of their genetic stability. *Plant Cell, Tissue and Organ Culture* 97(2): 121-129.
- Knudson, L. (1922). Nonsymbiotic Germination of Orchid Seeds. *Botanical Gazette*, 73(1): 1-25.
- Kunitake, H., Koreeda, K. and Mii, M. (1995). Morphological and cytological characteristics of protoplast-derived plants of statice (*Limonium perezii* Hubbard). *Scientea Horticulturae* 60: 305-312.

- Lakshmanan, V., Sreedhar, R.V. and Bhagyalakshmi, N. (2007). Molecular analysis of genetic stability in long term micropropagated shoots of banana using RAPD and ISSR markers. *Electronic Journal of Biotechnology* 10: 1-8.
- Larkin, P. and Scowcroft, W. (1981). Somaclonal variation: a novel source of variability from cell cultures for plant improvement. *Theoretical and Applied Genetics* 60: 197-214.
- Larkin, P.J. (1998). Introduction. In S.M. Jain, D.S. Brar and B.S. Ahloowalia (eds) *Somaclonal variation and induced mutations in crop improvement* (pp. 3-13). Dordrecht: Kluwer Academic Publishers.
- Latip, M., Murdad, R., Aziz, Z.A., Ting, L.H., Govindasamy, L.M. and Ripin, R. (2007). Effects of N6-Benzyladenine and Thidiazuron on Proliferation of *Phalaenopsis gigantea* Protocorms. *Proceedings Asia Pacific Conference on Plant Tissue and Agribiotechnology*, 18 (1): 217-220.
- Lee, Y.S., Kim, Y.H. and Kim, S.B. (2005). Changes in the respiration, growth, and vitamin C content of soybean sprouts in response to chitosan of different molecular weights. *HortScience* 40: 1333-1335.
- Leroy, X.J., Leon, K., Hily, J.M., Chaumeil, P. and Branchard, M. (2001). Detection of *in vitro* culture-induced instability through inter-simple sequence repeat analysis. *Theoretical and Applied Genetics* 102: 885-891.
- Li, Q., Dunn, E.T., Grandmaison, E.W. and Goosen, M.F.A. (1992). Applications and properties of chitosan. *Journal of Bioactive and Compatible Polymers* 7: 370-397.
- Liau, C.H., You, S.J., Prasad, V., Hsiao, H.H., Lu, J.C., Yang, N.S. and Chan, M.T. (2003). *Agrobacterium tumefaciens*-mediated transformation of an *Oncidium* orchid. *Plant Cell Reports* 21: 993-998.
- Limpanavech, P., Pichyangkura, R., Khunwasi, C., Chadchawan, S., Lotrakul, P., Bunjongrat, P., Chaidee, A. and Akaraekpanya, T. (2003). The effects of polymer type, concentration and % DD of bicatalyte modigied chitosan on flora production of *Dendrobium* 'Eiskul'. *National chitin-chitosan conference*, Chulalongkorn University, Bangkok, Thailand, July 17-18, 2003. pp 60-64.
- Liu, T.A., Lin, J.J. and Wu, R.Y. (2006). The effects of using trehalose as a carbon source on the proliferation of *Phalaenopsis* and *Doritaenopsis* protocorm-like bodies. *Plant Cell, Tissue and Organ Culture* 86(1): 125-129.
- Long, S.P., Humphries, S. and Falkowski, P.G. (1994). Photoinhibition of photosynthesis in nature. *Annual Review of Plant Physiology and Plant Molecular Biology* 45: 633-662.
- LoSchiavo, F., Pitto, L., Giuliano, G., Torti, G., Nuti-Ronchi, V., Marazziti, D., Vergara, R., Orselli, S. and Terzi, M. (1989). DNA methylation of

embryogenic carrot cell cultures and its variations as caused by mutation, differentiation, hormones and hypomethylating drugs. *Theoretical and Applied Genetics* 77: 325-331.

Lu, R (2004). Multispectral imaging for predicting firmness and soluble solids content of apple fruit. *Postharvest Biology and Technology*, 31(2), 147–157

Lu, J.J., Zhao, H.Y., Suo, N.N., Wang, S., Shen, B., Wang, H.Z. and Liu, J.J. (2012). Genetic linkage maps of *Dendrobium moniliforme* and *D. officinale* based on EST-SSR, SRAP, ISSR and RAPD markers. *Scientia Horticulturae*, 137: 1-10.

Luo, J.P., Wang, Y., Zha, X.Q. and Haung, L. (2008). Micropropagation of *Dendrobium densiflorum* Lindl. ex Wall. through protocorm-like bodies: effects of plant growth regulators and lanthanoid. *Plant Cell, Tissue and Organ Culture* 93: 333-340.

Luque, C., Legal, L., Staudter, H., Gers, C. and Wink, M. (2002). ISSR (inter simple sequence repeats) as genetic markers in noctuids (Lepidoptera). *Hereditas* 136: 251-253.

Malabadi, R.B., Mulgund, G.S. and Nataraja, K. (2004). Efficient regeneration of *Vanda coerulea*, an endangered orchid using thidiazuron. *Plant Cell Tissue and Organ Culture* 76: 289-293.

Mandal, A., Maiti, A., Chowdhury, B. and Elanchezhian, R. (2001). Isoenzyme markers in varietal identification of banana. *In Vitro Cellular & Developmental Biology* 37: 599-604.

Martins, M., Sarmiento, D. and Oliveira, M.M. (2004). Genetic stability of micropropagated almond plantlets as assessed by RAPD and ISSR markers. *Plant Cell Reports* 23: 492-496.

Mason, M.E. and Davis, J.M. (1997). Defense response in slash pine: chitosan treatment alters the abundance of specific mRNAs. *Molecular Plant-Microbe Interactions* 10:135-137.

Mayr (1998). *Orchid names and their meaning*. Koenigstein: Koeltz Scientific Books, Germany.

Mehta, Y.R. and Angra, D.C. (2000). Somaclonal variation for disease resistance in wheat and production of dihaploids through wheat × maize hybrids. *Genetics and Molecular Biology* 23: 617-622.

Miles, K. (1982). Growing equitant *Oncidium*s. *Amer. Orchid SCO. Bull.* 51: 155-169.

Mishiba, K. and Mii, M. (2001). Increasing ploidy level in cell suspension cultures of *Doritaenopsis* by exogenous application of 2,4-dichlorophenoxyacetic acid. *Physiologia Plantarum* 112: 142-148.

- Mujib, A. (2005). Colchicine induced morphological variants in pineapple. *Plant Tissue Culture & Biotechnology* 15: 127-133.
- Mujib, A., Banerjee, S. and Dev Ghosh, P. (2007). Callus induction, somatic embryogenesis and chromosomal instability in tissue culture raised hippeastrum (*Hippeastrum hybridum* cv. *United Nations*). *Propagation of Ornamental Plants* 7: 169-174.
- Murdad, R., Hwa, K.S., Seng, C.K., Latip, M.A., Aziz, Z.A. and Ripin, R. (2006). High frequency multiplication of *Phalaenopsis gigantea* using trimmed bases protocorms technique. *Scientia Horticulturae*, 111(1): 73-79.
- Nagaoka, T. and Ogihara, Y. Applicability of inter-simple sequence repeat polymorphisms in wheat for use as DNA markers in comparison to RFLP and RAPD markers. *Theoretical and Applied Genetics* 97(5): 597-602.
- Nagaraju, J., Reddy, K., Nagaraja, G. and Sethuraman, B. (2001). Comparison of multilocus RFLPs and PCR-based marker systems for genetic analysis of the silk-worm, *Bombix mori*. *Heredity* 86: 588-597.
- Naing, A., Chung, J., Park, I. and Lim, K. (2010). Efficient plant regeneration of the endangered medicinal orchid, *Coelogyne cristata* using protocorm-like bodies. *Acta Physiologiae Plantarum* 33(3): 659-666.
- Nash, N., (2003). *Phalaenopsis* primer :a beginner's guide to growing moth orchids. *Orchids*, 72: 906-913.
- Nayak, N.R., Rath, S.P. and Patnaik, S. (1997). *In vitro* propagation of three epiphytic orchids, *Cymbidium aloifolium* (L.) Sw., *Dendrobium aphyllum* (Roxb.) Fisch. and *Dendrobium moschatum* (Buch-Ham) Sw. through thidiazuron-induced high frequency shoot proliferation. *Scientia Horticulturae* 71: 243-250.
- Nayak, N.R., Sahoo, S., Patnaik, S. and Rath, S.P. (2002). Establishment of thin cross section (TCS) culture method for rapid micropropagation of *Cymbidium aloifolium* (L.) Sw. and *Dendrobium nobile* Lindl. (*Orchidaceae*). *Scientia Horticulturae* 94: 107-116.
- Ng, C.Y. and Saleh, N. (2010). *In vitro* propagation of *Paphiopedilum* orchid through formation of protocorm-like bodies. *Plant Cell, Tissue and Organ Culture* 105(2): 193-202.
- Nge, K.L., Nwe, N., Chandkrachang, S. and Stevens, W.F. (2006). Chitosan as a growth stimulator in orchid tissue culture. *Plant Science* 170(6): 1185-1190.
- Niknejad, A. (2009). *Determination of Genetic Relationships among Phalaenopsis Spp. Using Random Amplified Polymorphic DNA and In Vitro Propagation of Phalaenopsis Gigantea*. Masters thesis, Universiti Putra Malaysia.

- Niknejad, A., Kadir, M.A. and Kadzimin, S.B. (2011). *In vitro* plant regeneration from protocorms-like bodies (PLBs) and callus of *Phalaenopsis gigantea* (Epidendroideae:Orchidaceae). *African Journal of Biotechnology* 10(56): 11808-11816.
- No, H.K. and Lee, M.Y. (1995). Isolation of chitin from crab shell waste. *Journal Korean Soc. Food Nutrition* 24(1):105-113.
- No, H. K. and Meyers, S. P. (1995). Preparation and characterization of chitin and chitosan- a review. *Journal of Aquatic Food Product Technology* 4(2): 27-52.
- Notsu, S., Saito, N., Kosaki, H., Inui, H. and Hirano, S. (1994). Stimulation of phenylalanine ammonia-lyase activity and lignification in rice callus treated with chitin, chitosan and their derivatives. *Bioscience, Biotechnology, and Biochemistry* 58: 552-553.
- Orbovic', V., Calovic, M., Vilorija, Z., Nielsen, B., Gmitter, F., Castle, W. and Grosser, J. (2008). Analysis of genetic variability in various tissue culture-derived lemon plant populations using RAPD and flow cytometry. *Euphytica* 161: 329-335.
- Pansarin, E.R. and Pansarin, L.M. (2011). *The family Orchidaceae in the Serra do Japi, São Paulo State, Brazil* (pp. 6-11). Vienna: Springer.
- Park, S.Y., Murphy, H.N. and Paek, K.Y. (2000). Mass multiplication of protocorm-like bodies using bioreactor system and subsequent plant regeneration in *Phalaenopsis*. *Plant Cell, Tissue and Organ Culture* 63: 67-72.
- Park, S.Y., Murthy, H.N. and Paek, K.Y. (2002). Rapid propagation of *Phalaenopsis* from floral stalk-derived leaves. *In Vitro Cellular & Developmental Biology - Plant*, 38: 168-172.
- Park, S.Y., Kakuta, S., Kano, A. and Okabe, M. (1996). Efficient propagation of protocorm-like bodies of *Phalaenopsis* in liquid medium. *Plant Cell, Tissue. and Organ Culture* 45: 79-85.
- Peng, X., Liu, J.J., Xiang, Y. and Huang, S. (2006). A practical handbook of plant molecular biotechnology. *Beijing: Chemical Industry Press*.
- Peredo, E.L., Arroyo-Garcia, R. and Revilla, M.A. (2009). Epigenetic changes detected in micropropagated hop plants. *Journal of Plant Physiology* 166(10): 1101-1111.
- Petolino, J.F., Roberts, J.L. and Jayakumar, P. (2003). Plant cell culture: a critical tool for agricultural biotechnology. In V.A. Vinci and S.R. Parekh (eds) *Handbook of industrial cell culture: mammalian, microbial and plant cells*. (pp 243–258). New Jersey: Humana Press.

- Peyvandi, M., Noormohammadi, Z., Banihashemi, O., Farahani, F., Majd, A., Hosseini-Mazinani, M. and Sheidai, M. (2009). Molecular analysis of genetic stability in long-term micropropagated shoots of *Olea europaea* L. (cv. Dezful). *Asian Journal of Plant Science* 8: 146-152.
- Phinney, B. (1985). Gibberellin A1 dwarfism and shoot elongation in higher plants. *Biologia plantarum* 27: 172-179.
- Photchanachai, S., Singkaew, J. and Thamthong, J. (2006). Effects of chitosan seed treatment on *Colletotrichum* sp. and seedling growth of chili cv. 'Jinda'. *Acta Horticulturae* 712: 585-590.
- Pierik, R.L.M. (1987). *In vitro culture of higher plants*. Dordrecht: Kluwer Academic Publishers.
- Polonca, K. Susan, S and Zalata, L. (2004). Direct shoot regeneration from nodes of *Phalaenopsis* orchid. *Acta agriculturae Slovenica*, 83:233-242
- Pornpienpakdee, P., Singhasurasak, R., Chaiyasap, P., Pichyangkura, R., Bunjongrat, R., Chadchawan, S. and Limpanavech, P. (2010). Improving the micropropagation efficiency of hybrid *Dendrobium* orchids with chitosan. *Scientia Horticulturae*, 124(4): 490-499.
- Predieri, S. (2001). Mutation induction and tissue culture in improving fruits. *Plant Cell, Tissue and Organ Culture*, 64 185–210.
- Raimondi, J.P., Masuelli, R.W. and Camadro, E.L. (2001). Assessment of somaclonal variation in asparagus by RAPD fingerprinting and cytogenetic analyses. *Scientia Horticulturae* 90: 19-29.
- Raina, S.N., Rani, V., Kojima, T., Ogihara, Y., Singh, K.P. and Devarumath, R.M. (2001). RAPD and ISSR fingerprints as useful genetic markers for analysis of genetic diversity varieties identification and phylogenetic relationships in peanut (*Arachis hypogaea*) cultivars and wild species. *Genome* 44:763-772.
- Rakwal, R., Tamogami, S., Agrawal, G.K. and Iwahashi, H. (2002). Octadecanoid signaling components "burst" in rice (*Oryza sativa* L.) seedling leaves upon wounding by cut and treatment with fungal elicitor chitosan. *Biochemical and Biophysical Research Communications* 295:1041-1045.
- Rani, V. and Raina, S. (2000). Genetic fidelity of organized meristem-derived micropropagated plants: a critical reappraisal. *In Vitro Cellular & Developmental Biology - Plant* 36: 319-330.
- Rasheed, S., Tahira, F., Khurram, B., Tayyab, H. and Shiekh, R. (2003). Agronomical and physiochemical characterization of somaclonal variants in Indica basmati rice. *Pakistan Journal of Biological Sciences* 6:844-848.

- Ravindra, M., Gangadhar, M. and Nataraja, K. (2004). Efficient regeneration of *Vanda coerulea*, an endangered orchid using thidiazuron. *Plant Cell, Tissue and Organ Culture*. 289-293.
- Reuter, E. (1983). The importance of propagating *Phalaenopsis* by tissue culture. *Orchid Review*, 91: 199-201.
- Reuveni, O. and Israeli, Y. (1990). Measures to reduce somaclonal variation in *in vitro* propagated bananas. *Acta Horticulturae* 275: 307-313.
- Reuveni, O., Israeli, Y., Degani, H. and Eshdat, Y. (1986). Genetic variability in banana multiplied via *in vitro* technique. *International Board for Plant Genetic Resource meeting*, Rome. Resumos. Rome: IBPGR.
- Roberts, D.L. and Dixon, K.W. (2008). Orchids. *Current Biology* 18(8): 325-329.
- Rodrigues, K. and Kumar, S. (2009). Isolation and characterization of microsatellite loci in *Phalaenopsis gigantea*. *Conservation Genetics* 10(3): 559-562.
- Rodrigues, P.H.V., Neto, A.T., Neto, C.P. and Mendes, B.M.J. (1998). Influence of the number of subcultures on somoclonal variation in micropropagated Nanico (*Musa* spp., AAA group). *Acta Horticulturae* 490: 469-473.
- Rohlf, F. J. (1993). NTSYS-pc, Numerical Taxonomy and Multivariate Analysis System. Version 1.80. Exeter Software: Setauket, NY.
- Roy, J. and Banerjee, N. (2003). Induction of callus and plant regeneration from shoottip explants of *Dendrobium fimbriatum* Lindl var. *oculatum* Hk. F. *Scientia Horticulturae* 97: 333-340.
- Roy, J., Naha, S., Majumdar, M. and Banerjee, N. (2007). Direct and callus mediated protocorm-like body induction from shoot-tips of *Dendrobium chrysotoxum* Lindl. (*Orchidaceae*). *Plant Cell, Tissue and Organ Culture* 90: 31-33.
- Sagawa, Y. and Kunisaki, J. (1982). Clonal propagation of orchids by tissue culture. *Proceedings of the 5th plant tissue and cell culture*. 683-684.
- Sahijram, L., Soneji, J. and Bollamma, K. (2003). Analysing somaclonal variation in micropropagated bananas (*Musa* spp.). *In Vitro Cellular & Developmental Biology - Plant* 39: 551-556.
- Saiprasad, G.V.S., Raghuveer, P., Khetarpal, S. and Chandra, R. (2004). Effect of various polyamines on production of protocorm-like bodies in orchid *Dendrobium* 'Sonia'. *Scientia Horticulturae*. 100(14): 161-168.
- Saker, M.M., Bekheer, S.A., Taha, H.S., Fahmy, A.S. and Moursy, H.A. (2000). Detection of somaclonal variations in tissue cultured-derived date palm plants using iso-enzyme analysis and RAPD fingerprints. *Biologia Plantarum* 43: 347-351.

- Sala, F., Arencibia, A., Castiglione, S., Christou, P., Zheng, Y. and Han, Y. (1999). Molecular and field analysis of somaclonal variation in transgenic plants. In A. Altman, M. Ziv and S. Izhar (eds) *Plant biotechnology and in vitro biology in the 21st century* (pp. 259–262). Dordrecht: Kluwer.
- Sambrook, J., Fritsch, E.F. and Maniatis, T. (1989). *Molecular cloning: A laboratory manual*, 2nd ed : Cold Spring Harbor Laboratory Press.
- Sandal, I., Bhattacharya, A. and Ahuja, P.S. (2001). An efficient liquid culture system for tea shoot proliferation. *Plant Cell, Tissue and Organ Culture* 65: 75-80.
- Sandoval, J., Kerbellec, F., Co[^]te, F. and Doumas, P. (1995). Distribution of endogenous gibberellins in dwarf and giant off-types banana (*Musa* AAA cv. Grand nain) plants from *in vitro* propagation. *Plant Growth Regulation* 17: 219-224.
- Shah, S.H., Wainwright, S.J. and Merret, M.J. (2003). Regeneration and somaclonal variation in *Medicago sativa* and *Medicago media*. *Pakistan Journal of Biological Sciences* 6:816-820.
- Sharma, S., Bryan, G., Winfield, M. and Millam, S. (2007). Stability of potato (*Solanum tuberosum* L.) plants regenerated via somatic embryos, axillary bud proliferated shoots, microtubers and true potato seeds: a comparative phenotypic, cytogenetic and molecular assessment. *Planta* 226:1449-1458.
- Sheelavantmath, S.S., Murthy, H.N., Hema, B.P., Hahn, E.J. and Paek, K.Y. (2005). High frequency of protocorm like bodies (PLBs) induction and plant regeneration from protocorm and leaf sections of *Aerides crispum*. *Scientia Horticulturae* 106: 395-401.
- Shimasaki, K., Tanibuchi, Y. and Fukumoto, Y. (2003). The effects of chitosan on organogenesis in protocorm-like body (PLB) of *Cymbidium finlaysonianum* Lindl. *Journal of Society of High Technology in Agriculture* 15: 90-93.
- Shimura, H. and Koda, Y. (2004). Micropropagation of *Cypripedium macranthos* var. *rebunense* through protocorm-like bodies derived from mature seeds. *Plant Cell, Tissue and Organ Culture* 78(3): 273-276.
- Shu-guo, F. (2008). Tissue culture system of *Phalaenopsis* in genetic transformation. *Journal of Biotechnology* 136:164-165.
- Sinha, P., Hakim, M.L. and Alam, M.F. (2007). Efficient micropropagation of *Phalaenopsis amabilis* (L.) BL. cv. 'Cool Breeze' using inflorescence axis thin sections as explants. *Propagation of Ornamental Plants* 7(1): 9-15.

- Skirvin, R. (1978). Natural and induced variation in tissue culture. *Euphytica* 27: 241-266.
- Skirvin, R.M., Norton, M. and McPheeters, K.D. (1993). Somaclonal variation: has it proved useful for plant improvement. *Acta Horticulturae* 336: 333-340.
- Smith, D.L. and Krikorian, A.D. (1990). Low external pH replaces 2,4-D in maintaining and multiplying 2,4-D initiated embryogenic cells of carrot. *Plant Physiology* 72: 329-336.
- So Young, P., Murthy, H.N. and Kee Yoeup, P. (2000). Mass multiplication of protocorm-like bodies using bioreactor system and subsequent plant regeneration in *Phalaenopsis*. *Plant Cell, Tissue and Organ Culture* 63(1): 67-72.
- Sopalun, K., Thammasiri, K. and Ishikawa, K. (2010). Micropropagation of the Thai orchid *Grammatophyllum speciosum* blume. *Plant Cell, Tissue and Organ Culture* 101(2): 143-150.
- Sreeramanan, S., Vinod, B., Sashi, S. and Xavier, R. (2008). Optimization of the transient Gusa gene transfer of *Phalaenopsis violacea* orchid via *Agrobacterium tumefaciens*: an assessment of factors influencing the efficiency of gene transfer Mechanisms. *Advances in Natural and Applied Sciences* 2: 77-88.
- Sukwattanasinitt, M., Klaikherd, A., Skulnee, K. and Aiba, S. (2001). Chitosan as a releasing device for 2,4-D herbicide. *Chitin and Chitosan in Life Science*. 198-201.
- Swartz, H.J. (1991). Post culture behaviour, genetic and epigenetic effects and related problems. In P.C. Debergh and R.H. Zimmerman (eds) *Micropropagation: technology and application*. (pp 95-122). Dodrecht :Kluwer Academic Publishers.
- Sweet, H.R. (1980). The genus *Phalaenopsis*. California: The Orchid Digest Inc. Tagged-site-facilitated PCR products as molecular markers in wheat. *Theoretical and Applied Genetics* 87: 789-794.
- Tanaka, M., Hasegawa, A. and Goi, M. (1975). Studies on the clonal propagation of monopodial orchid by tissue culture. I. formation of protocorm-like bodies from leaf tissues in *Phalaenopsis* and *Vanda*. *Journal of the Japanese Society for Horticultural Science*, 44: 47-57.
- Tang, C.W., HC (2007). Breeding and development of new varieties in *Phalaenopsis*. In W.H. Chen and H.H. Chen (eds) *Orchid Biotechnology* (pp. 1-20): World Scientific Publishing Co. Pte. Ltd.
- Tautz, D. (1989). Hypervariability of simple sequences as a general source for polymorphic DNA markers. *Nucleic Acids Research* 17: 5453-6471.

- Teixera da Silva, J.A. (2003). Thin cell layer technology for induced response and control of rhizogenesis in chrysanthemum. *Plant Growth Regulation* 39: 67-76.
- Teixera da Silva, J.A., Chan, M.T., Sanjaya, Chai, M.L. and Tanaka, M. (2006). Priming abiotic factor for optimal hybrid *Cymbidium* (*Orchidaceae*) PLB and callus induction, plantlet formation, and their subsequent cytogenetic stability analysis. *Scientia Horticulturae* 109: 368-378.
- Teixera da Silva, J.A. and Tanaka, M. (2006). Multiple regeneration pathway via thin cell layers in hybrid *Cymbidium* (*Orchidaceae*). *Journal of Plant Growth Regulation* 25: 203-210.
- Thammasiri, K (2002) Preservation of seeds of some Thai orchidspecies by vitrification. In: Proceedings of the 16th world orchi conference, pp 248–251
- Tian, C., Chen, Y., Zhao, X. and Zhao, L. (2008). Plant regeneration through protocorm like-bodies induced from rhizoids using leaf explants of *Rosa* spp. *Plant Cell Reports* 27: 823-831.
- Tokuhara, K. (1992). Somaclonal variation of flowers in micropropagated plant through flower stalk bud culture. In: Ichihashi S, Nagata H (eds). Proc Nagoya Int Orchid Show 92, Nagoya, Japan, pp. 317-319
- Tokuhara, K. and Mii, M. (1993). Micropropagation of *Phalaenopsis* and *Doritaenopsis* by culturing shoot tips of flower stalk buds. *Plant Cell Reports* 13: 7-11.
- Tokuhara, K.; Mii, M. (1998). Somaclonal variation in flower and inflorescence axis in micropropagated plants through flower stalk bud culture of *Phalaenopsis* and *Doritaenopsis*. *Plant Biotechnology*. 15(1):23-28
- Unai, E., Iselen, T. and de Garcia, E. (2004). Comparison of characteristics of bananas (*Musa* sp.) from the somaclone CIEN BTA-03 and its parental clone Williams. *Fruits* 59: 257-263.
- Uthairatanakij, A., Jitareerat, P., Kanlayanarat, S., Piluek, C. and Obsuwan, K. (2006). Efficacy of chitosan spraying on quality of *Dendrobium* Sonia 'NO. 17' inflorescence. 27th *International Horticultural Congress & Exhibition*, Korea. p 150.
- Uthairatanakij, A., Teixeira da Silva, J.A. and Obi Wan, K. (2007). Chitosan for Improving Orchid Production and Quality. *Orchid Science and Biotechnology* 1: 1-5.
- Vander, P., Kjell, M.V., Domard, A., El-Gueddari, N.E. and Moerschbacher, B.M. (1998). Comparison of the ability of partially *N*-acetylated chitosans and oligosaccharides to elicit resistance in wheat leaves. *Plant Physiology* 118: 1353-1359.

- Vasyukova, N.I., Zinoveva, L.I., Ilinskaya, E.A., Perekhod, G.I., Chalenko, N.G., Ilina, A.V., Varlamov, V.P. and Ozeretskoykaya, O.L. (2001). Modulation of plant resistance to diseases by water-soluble chitosan. *Applied Biochemistry and Microbiology* 37: 103-109.
- Venkatachalam, L., Sreedhar, R.V. and Bhagyalakshmi, N. (2007). Molecular analysis of genetic stability in long-term micropropagated shoots of bananas using RAPD and ISSR markers. *Electronic Journal of Biotechnology* 10: 1-8.
- Verma, P.C., Chakrabarty, D., Jena, S.N., Mishra, D.K., Singh, P.K., Sawant, S.V. and Tuli, R. (2009). The extent of genetic diversity among *Vanilla* species: Comparative results for RAPD and ISSR. *Industrial Crops and Products* 29 (2-3): 581-589.
- Vos, P., Hogers, R., Bleeker, M., Reijans, M., Van de Lee, T., Hornes, M., Frijters, A., Pot, J., Peleman, J., Kuiper, M. and Zabeau, M. (1995). AFLP: a new technique for DNA fingerprinting. *Nucleic Acids Research* 23: 4407-4414.
- Wang, H.Z., Feng, S.G., Lu, J.J., Shi, N.N. and Liu, J.J. (2009). Phylogenetic study and molecular identification of 31 *Dendrobium* species using inter-simple sequence repeat (ISSR) markers. *Scientia Horticulturae* 122(3): 440-447.
- Wang, Y., Wang, F., Zhai, H. and Liu, Q. (2007). Production of a useful mutant by chronic irradiation in sweet potato. *Scientia Horticulture* 111:173-178.
- Weising, K., Nybom, H., Wolff, K. and Kahl, G. (2005). DNA fingerprinting in plants: principles, methods, and applications. New York: CRC Press.
- William Decruse, S., Gangaprasad, A., Seeni, S. and Sarojini Menon, V. (2003). Micropropagation and ecorestoration of *Vanda spathulata*, an exquisite orchid. *Plant Cell, Tissue and Organ Culture* 72(2): 199-202.
- Williams, J.G., Kubelik, A.R., Livak, K.J., Rafalski, A. and Tingey, S.V. (1990). DNA polymorphisms amplified by arbitrary primers are useful as genetic markers. *Nucleic Acids Research* 18: 6531-6535.
- Willmer, C.M. and Fricker, M. (1996). *Stomata* (2nd Edn). London: Chapman and Hall. p. 375.
- Win, N.K.K., Jitareerat, P. and Kanlayanarat, S. (2005). Pre-harvest chitosan spraying on leaf spot disease and growth of orchid (*Dendrobium* Missteen). *Proceedings of APEC Symposium on assuring quality and safety of fresh product*, Bangkok, Thailand, Aug 1-3, 2005. pp 457-461.
- Witsenboer, H., Vogel, J. and Michelmore, R. W. (1997). Identification, genetic localization, and allelic diversity of selectively amplified microsatellite polymorphic loci in lettuce and wild relatives (*Lactuca* spp.). *Genome* 40: 923-936.

- Yang, W., De Oliveira, A.C., Godwin, I., Schertz, K., Bennetzen, J.L. (1996). Comparison of DNA marker technologies in characterizing plant genome diversity: Variability in Chinese sorghums. *Crop Science*, 36 (6), pp. 1669-1676.
- Yin, M. and Hong, S. (2009). Cryopreservation of *Dendrobium candidum* Wall. ex Lindl. protocorm-like bodies by encapsulation-vitrification. *Plant Cell, Tissue and Organ Culturae* 98:179-185.
- Yong, J.W., Ge, L., Ng, Y.F. and Tan, S.N. (2009). The chemical composition and biological properties of coconut (*Cocos nucifera* L.) Water. *Molecules*, 14(12): 5144-5164.
- Zhang, D. and Quantick, P.C. (1998). Antifungal effects of chitosan coating on fresh strawberries and raspberries during storage. *The Journal of Horticultural Science and Biotechnology* 73: 763-767.
- Zheng, Y.X., Chen, C.C., Chen, Y.K. and Jan, F.J. (2008). Identification and characterization of a potyvirus causing chlorotic spots on *Phalaenopsis* orchids. *European Journal of Plant Pathology* 121(1): 87-95.
- Zietkiewicz, E., Rafalski, A. and Labuda, D. (1994). Genome fingerprinting by simple sequence repeat (SSR)-anchored polymerase chain reaction amplification. *Genomics* 20: 176-183.