



**UNIVERSITI PUTRA MALAYSIA**

***INFLUENCE OF ALLELOPATIC MULCH ON WEED GROWTH AND  
POSTHARVEST QUALITY OF LEAF LETTUCE (*Lactuca sativa* L.)***

**BUNGA RAYA KETAREN**

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**MASTER OF SCIENCE  
UNIVERSITI PUTRA MALAYSIA**

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**By**

**BUNGA RAYA KETAREN**

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**January 2013**

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POSTHARVEST QUALITY OF LEAF LETTUCE (*Lactuca sativa* L.)**

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**January 2013**

**Chairman : Siti Hajar Ahmad, PhD**

**Faculty : Agriculture**

Today, there is a high demand for our food supply to be free of potentially toxic substances including those used in the production of raw products. During the production and processing of agricultural commodities, various chemicals and foreign substances such as pesticide may enter food commodities. Most pesticides are acutely toxic and can produce adverse health effects in humans and animals when ingested at low levels over long periods of time. A study was conducted to identify allelopathic mulch for sustainable weed control of leaf lettuce and determine its effect on postharvest quality characteristics. A quantitative survey was done to investigate major weeds selected leafy vegetable fields in Selangor. Ten weed species were recorded. *Cyperus rotundus* and *Ageratum conyzoides* were the most abundant weeds followed by *Eleutheranthera ruderalis*, *Amaranthus spinosus*, and *Cyperus kyllingia*. Selecting allelopathic mulch to inhibit weed seeds germination was studied using extracts of sorghum, paddy straw and empty oil palm fruit bunch (EFB) in a growth chamber. Total germinated seeds were recorded every 24 hour. Shoot and root lengths were measured on germination day 20. Allelopathy residues were detrimental to germination percentage, germination speed, mean germination time and relative hypocotyl and root length of weed species studied. Cucumber, cabbage and sunflower allelochemicals inhibited 100% germination of *C.*

*rotundus*, *A. conyzoides*, *E. ruderalis*, *A. spinosus* and *C. kyllingia*. Sorghum, paddy straw and EFB caused 30-50% germination for *C. kyllingia*, Seed germination of *C. rotundus*, *A. conyzoides*, *E. ruderalis* and *A. spinosus* was 10-30% when treated with extract sorghum, paddy straw and EFB. Germination of *E. ruderalis* was lowest when treated with EFB than other weeds. Final seed germination and seedling growth was reduced 0-50% with sorghum, paddy straw and EFB mulch residues. Effectiveness of sunflower and cucumber mulch on leaf lettuce was studied on vegetable field plot. Types and rates of mulching did not affect soils moisture content and total N, P, K, Mg and Ca. Higher total phenolic content and phenolic soluble concentration occurred in soils with cucumber mulch residue compared to control. Seventeen weed species belonging to six families was observed in Leaf lettuce crop. Unmulched plot had 14 weed species of family Poaceae, three Cyperaceae, two each of Euphorbiaceae and Asteraceae and one each of Amaranthaceae and Portulacaceae. Sum dominance ratio showed five most dominant weed encountered were *C. rotundus*, *C. distans*, *A. spinosus*, *C. kyllingia* and *A. conyzoides*. Sunflower (5 ton/ha) and cucumber (15 ton/ha) mulched plots caused reduction to five weed species. Highest reduction was for *A. spinosus*. Sum dominance ratio of weeds with sunflower mulch residues (10 and 15 ton/ha) was high for *C. rotundus*, and similar to 10 ton/ha cucumber mulch. Cucumber mulch at 5 ton/ha successfully suppressed *C. rotundus* in leaf lettuce plots. Mulching did not affect postharvest quality of Leaf lettuce. All characteristics measured (firmness, pH, TA and AA) were not different from unmulched treatment. Mulch rates did not affect quality characteristics and soil nutrients of leaf lettuce. Cucumber and sunflower mulching controlled weeds effectively and has potential as alternative chemical herbicides. Studies should be conducted for longer period for optimum mulch residues effectiveness.

Abstrak tesis yang dikemukakan kepada Senat of Universiti Putra Malaysia sebagai memenuhi keperluan ijazah Master Sains

**PENGARUH SUNGKUPAN ALLELOPATIK KE ATAS PERTUMBUHAN RUMPAI  
DAN KUALITI LEPAS TUAI SALAD DAUN (*Lactuca sativa* L.)**

By

**BUNGA RAYA KETAREN**

**Januari 2013**

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Pada masa kini, terdapat permintaan yang tinggi bagi bekalan makanan yang bebas daripada bahan toksik termasuk yang digunakan untuk pengeluaran produk mentah. Semasa pengeluaran dan pemprosesan komoditi pertanian, pelbagai jenis bahan kimia dan bahan-bahan asing seperti racun perosak boleh mencemari bahan makanan. Kebanyakan racun perosak adalah toksik secara akut. Walaupun tercemar pada kadar yang rendah/minima namun pendedahan dalam tempoh masa yang panjang boleh mengakibatkan kesan buruk terhadap kesihatan manusia dan juga haiwan. Kajian dijalankan untuk mengenalpasti sungkupan allelopati untuk kawalan rumpai lestari bagi salad daun dan menentukan kesannya ke atas kualiti cirian lepas tuai. Tinjauan kuantitatif dijalankan untuk mengenalpasti rumpai utama di ladang sayur berdaun di Selangor. Sepuluh spesies rumpai dicatatkan. *Cyperus rotundus* dan *Ageratum conyzoides* adalah rumpai terbanyak, diikuti *Eleutheranthera ruderalis*, *Amaranthus spinosus* dan *Cyperus kyllingia*. Kajian memilih sungkupan allelopati penghalang percambahan biji-benih rumpai, rawatan ekstrak sekoi, jerami padi, tandan kosong sawit digunakan. Jumlah benih yang bercambah direkodkan dalam tempoh 24 jam. Panjang hipokotil dan radikal diukur pada hari percambahan ke-20. Sisa allelopati menjejaskan peratusan percambahan biji benih, kecepatan percambahan, min masa

percambahan dan panjang relatif hipokotil dan radikal rumpai yang dikaji. Allelopati timun, kobis dan bunga matahari merencatkan 100% percambahan *C. rotundus* L., *A. conyzoides* L., *E. ruderalis*, *A. spinosus* L., and *C. kyllingia*. Sekoi, jerami padi dan tandan kosong sawit menyebabkan 30-50% percambahan biji benih *C. kyllingia*. Hasil percambahan *C. rotundus*, *A. conyzoides*, *E. ruderalis* and *A. spinosus* adalah 10-30% dengan rawatan sekoi, jerami padi dan tandan kosong sawit. Percambahan *E. ruderalis* paling rendah apabila dirawat dengan tandan kosong sawit berbanding rumpai lain. Percambahan akhir biji benih dan pertumbuhan anak benih berkurangan 0-50% dengan sungkupan sekoi, jerami padi dan tandan kosong sawit. Kajian keberkesanan sungkupan bunga matahari dan timun dilakukan pada salad daun ditanam di ladang. Jenis dan kadar sungkupan tidak memberi kesan kepada % kelembapan tanah dan jumlah N, P, K, Mg dan Ca tanah. Jumlah kandungan fenolik dan kepekatan fenolik terlarut menunjukkan kesan tertinggi dengan sungkupan timun berbanding kawalan. Sejumlah 17 species rumpai daripada 6 keluarga terdapat dalam tanaman salad daun. Dalam plot tanpa sungkupan, terdapat sebanyak 14 species rumpai dari keluarga Poaceae, tiga species Cyperaceae, dua species Euphorbiaceae dan Asteraceae dan satu species Amaranthaceae dan Portulacaceae. Berdasarkan sum dominance ratio, lima species rumpai yang dominan djumpai ialah *C. rotundus*, *C. distans*, *A. spinosus*, *C. kyllingia* dan *A. conyzoides*. Sungkupan bunga matahari (5 tan/ha) dan timun (15 tan/ha) menyebabkan penurunan kepada lima jenis species rumpai. Penurunan tertinggi adalah bagi *A. spinosus*. Jumlah nisbah dorminan rumpai yang disungkup sisa bunga matahari (10 dan 15 tan/ha) adalah tinggi bagi *C. rotundus*, dan sama dengan sungkupan sisa timun pada kadar 10 tan/ha. Sungkupan timun pada kadar 5 tan/ha berjaya menghalang *C. rotundus* dalam kawasan tanaman salad daun. Sungkupan tidak mempengaruhi kualiti lepas tuai salad daun. Kesemua cirian (pH, asid tertitrat dan asid askorbik) salad daun tidak menunjukkan perbezaan berbanding rawatan tanpa sungkupan. Kadar setiap sungkupan juga tidak mempengaruhi kualiti cirian dan nutrient tanah.

Sungkupan sisa timun dan bunga matahari mengawal rumput dengan berkesan dan boleh digunakan sebagai alternatif kepada racun rumput. Akan tetapi, kajian perlu dijalankan pada jangka masa panjang untuk mendapat keberkesanan sisa sungkupan yang optimum.





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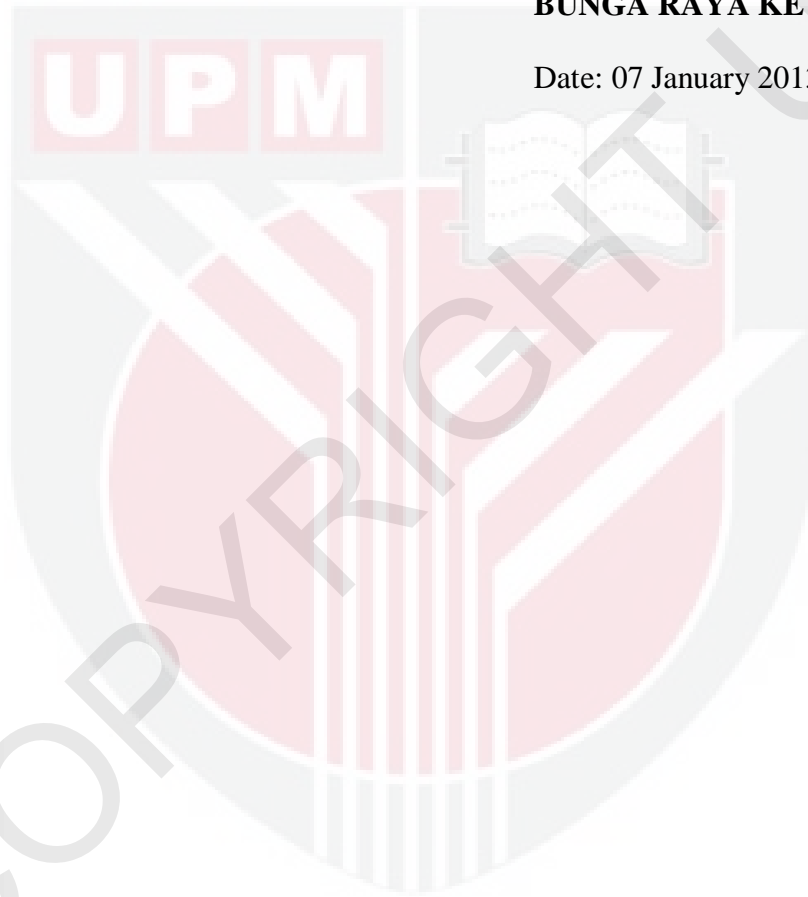
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## DECLARATION

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously, and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institution.

**BUNGA RAYA KETAREN**

Date: 07 January 2013



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## LIST OF ABBREVIATIONS

CEC	: Cation exchange capacity
N	: Nitrogen
P	: Phosphorus
K	: Potassium
USA	: United States of America
HPLC	: High Performance Liquid Chromatography
ATP	: Adenosine Tri-Phosphate
DNA	: Deoxyribonucleic acid
RNA	: Ribonucleic Acid
TA	: Titratable acidity
SSC	: Soluble solids concentration
TPC	: Total phenolic content
R <sup>2</sup>	: Coefficient determination
F	: Frequency
FU	: Field uniformity
D	: Field density
MFD	: Mean field density
RA	: Relative abundance
RFMD	: Relative mean field density
EFB	: Empty fruit bunch
FGP	: Final germination percentage
AS	: Speed of accumulated germination
MGT	: Mean germination time

LSD : Least Significant Difference  
RCBD : Randomized Complete Block Design  
RDW : Relative dry weight  
Mg : Magnesium  
SDR : Summed dominance ratio



## CHAPTER I

### GENERAL INTRODUCTION

The human population has increased rapidly during later half of the 20<sup>th</sup> century and dependence on agrochemicals (fertilizers, herbicides and pesticides) has increased to produce the required agricultural products. These agrochemicals, besides polluting the environment, are also hazardous to human and livestock. Hence, such practices are not sustainable and cannot remain in use forever. Poor yield, by repeated cultivation of certain crops and fruit, due to so called ‘soil sickness’ has been known and investigated since the beginning of horticulture. Use of allelopathy is a safe alternative to overcome these problems and to achieve sustainability in agriculture and maintenance of clean environment for our future generations.

Allelopathic strategies are aimed at reducing environmental pollution and maintaining ecological balance especially soil fauna and flora through reduced use of chemical herbicides or substituting them with natural products (plant and microbial products). Allelochemicals and phytochemicals are eco-friendly and free from the problems associated with the present herbicides. Hence, allelopathy is the priority area of research in the world. Phytotoxicity is a very old component of agriculture and it is described as allelopathy by Molisch (1937). Any direct or indirect and harmful or beneficial effect by one plant (including micro-organisms) on another through production of chemical compounds that escape into the environment is called allelopathy (Rice, 1984).

Allelopathy is an important factor in determining vegetation pattern, species diversity and vegetation dynamics. It plays a significant role in “plant–plant” and “plant–microbe” interactions, which are important in the management of one species by another mainly through biochemical interactions, affecting seed germination and seedling growth of existing species by the intruder species. These chemical compounds (allelochemicals) are released into the environment in appropriate quantities via root exudation, as leachates during litter decomposition and volatilization (Reigosa et al., 2000), are phenolic and allelopathic in nature (Cecchi et al., 2004). Allelopathy inhibition typically results from the combined action of a group of allelochemicals which collectively interfere with several physiological processes of the receiver plants and their growth and development affected. The readily visible effects include inhibited or retarded germination rate, swollen and darkened seeds, reduced root and shoot extension, swelling of root tips (Oliveira et al., 2004), curling of the root axis, discoloration, lack of root hairs and lateral roots (Burgos et al., 2004), increased number of seminal roots, reduced dry weight and lowered reproductive capacity (Rice, 1979).

Allelopathy in plant residue include plant or their parts deliberately left in the field soil for decomposition after the crop has been harvested (Kumar and Goh, 2000). These are considered as natural resource rather than waste because of their usefulness to the fields. Their decomposition by the microbial action improves the physical, chemical and biological properties of the soil and also adds a number of carbon end products and several other nutrients to the soil system. This process of decomposition depends upon the kind of plant residue as several edaphic factors

(Kumar and Goh, 2000). The main concern regarding the crop residues is their allelopathic effect on other or same crop plants (Thorne et al., 1990). The decomposing crop residue releases a variety of allelochemicals, particularly, in the soil causing adverse effects on the other plants (Nelson, 1996). They are known to reduce nodulation and biological nitrogen fixation in legumes (Heckman and Kluchinski, 1995).

Allelopathic effects, strong enough to contribute significantly to weed control in field conditions, have been documented for rye, sorghum and sorghum–sudangrass hybrids, sunflower, cucumber, rapeseed, buckwheat, and subterranean clover (Putnam and Tang, 1986; Rice, 1995; Boydston and Hang, 1995). Hence, there is a need for weed free maintenance during early crop growth. Manual weeding is laborious, time consuming and non-economical. The chemical method of weed control offers pollution and health hazards. As such, onion being vegetable crop use of chemicals has to be discouraged. Allelopathic is an interaction between plant and other organisms that offers alternative uses in agriculture including decreasing dependence on synthetic pesticide for the control of pests, diseases and weeds (Saxena et al., 1996).

Allelopathy offers potential for bio-rational weed control through production and release of allelochemicals from leaves, flowers, seeds, stems and roots of living or decomposing plant materials. Under appropriate conditions, allelochemicals may be released in a quantity which suppress the developing weed seedlings and often exhibit selectivity similar to synthetic herbicides (Weston, 1996). Grundy et al;

(1998) reported that crop plant suppressed weed development physiologically through superior vigor by release of phytotoxins or combinations of both allelopathy may have been a factor in few crops that did reduce weed seedling emergence. The suppression of weeds by crops should be exploited to improve weed control.

Lettuce (*Lactuca sativa* L.) is the leading leafy vegetable worldwide with an estimated production of 23 million tons in 2007 (Food Agriculture Organization of the United Nations, 2007). In Malaysia, the planted acreage for vegetables was 53,057 ha, with a production of 874,602 tons in 2010. For lettuce, acreage was 2,297 ha with a production of 38,790 ton (Ministry of Agriculture and Agro-Based Industry Malaysia, 2011). Lettuce is widely eaten fresh and commonly grown as a hydroponic crop. However, only the local leaf lettuce is commonly grown on soils. Naturally, weeds become the main problem of growing crop in soils, and their control by the application of herbicides cause hazard to the environment. For this reason the use of allelopathic mulch to control weed is an alternative to herbicide control.

The research information is inadequate on allelopathic residue effect with potential in controlling common weeds of vegetables. Such information will help to develop organic herbicides which are environmentally safe and cost effective. Identification of suitable botanicals with herbicidal properties and formulations is gaining a special importance in organic farming. Although many botanicals are reported to have allelopathic properties but information on their compatibility with field crops, effective active ingredient, extraction and utilization technology are lacking.

Postharvest priorities across the globe have evolved considerably over the past four decades, from being exclusively technical in their outlook, to being more responsive to consumer demand. Consumer-driven trends which have contributed to this shift include rising incomes in urban areas, changing dietary habits, more women in the work-place, reduced time for meal preparation and growing demand for safety, quality and convenience. Other factors such as globalization, urbanization and the need to achieve efficiencies and reduce costs have also contributed significantly to this shift in priorities and continue to re-shape and restructure the fresh produce sector.

Due to an increasing concern about possible human health risks and undesirable environmental effects associated with the use of synthetic chemicals in conventional apple production, some growers are interested in organic apple production, which does not involve the use of synthetic mineral fertilizers, pesticides, herbicides, fungicides, and growth regulators. Organic production relies on crop rotation, crop residues, manures, off-farm organic wastes, mechanical cultivation, mineral-bearing rocks, and biological pest control to maintain soil fertility and productivity, supply plant nutrients, control insects and weeds, and other pests (Consumer and Corporate Affairs Canada, 1988).

Much work, however, needs to be done across the region in order to promote quality and safety consciousness, and in particular to assure the use of water of appropriate quality in pre- and postharvest operations, as well as the appropriate use of pesticides, herbicides, fungicides and fertilizers during the production of fruits and



vegetables. The implementation of regulatory frameworks which govern food safety and quality, laboratories and quality assurance services is required in many countries across the region (Rosa, 2006).

The safety of our food supply is important to agribusiness as well as to consumers. Today, there is a high demand for our food supply to be free of potentially toxic substances including those used in the production of raw products. During the production and processing of agricultural commodities, various chemicals and foreign substances may enter food commodities. These chemicals are considered indirect or non-intentional food additives. Pesticides used in the production of raw products are among the most common indirect food additives. Most pesticides are acutely toxic and can produce adverse health effects in humans and animals when ingested at low levels over long periods of time (Smith, 2000). Keeping the above points in view, a study was carried out to identify allelopathic mulch for sustainable weed control of leaf lettuce and determine its effect on growth performance and postharvest quality characteristics. Following specific objectives :

1. To identify common weeds in a vegetable field
2. To select the mulch (plant residue) that inhibit weed seed germination
3. To determine effectiveness of selected mulch in reducing weed competition in a field-cultivated leaf lettuce
4. To determine the allelopathic mulch effect on postharvest quality characteristics of leaf lettuce

## BIBLIOGRAPHY

- Aharoni, N., Barkai-Golan, R., & Karadavia, R. (1979). Reducing rot in stored lettuce with polyethylene line packaging in Scientific Activities: 1974-1977. In I. T. a. S. o. A. Products (Ed.) (pp. 57). Israel.
- Ahmad, S. (1996). Paper presented at the 5th Pakistan Weed Sci. Conf. NARC, Islamabad, Pakistan.
- Akobundu, I. O., & Agyakwa, C. W. (1998). *A Handbook of West African Weeds*. Ibadan, Nigeria: International Institute of Tropical Agriculture.
- Akobundu, I. O., Ekeleme, F., & Chikoye, D. (1999). Influence of fallow management systems and frequency of cropping on weed growth and crop yield. *Weed Research*, 39, 241–256.
- Alex, J. F., & Switzer, C. M. (1976). *Ontario weeds*. Toronto, Ontario, Canada: Ontario Ministry of Agriculture and Food.
- Allaway, W. H. (Ed.). (1975). *The Effect of Soils and Fertilization on Human and Animal Nutrition* (Vol. 378): Agricultural Information Washington, D.C.: USDA
- Ames, B. N., Shigenaga, M. K., & Hagen, T. M. (1993). Oxidants, antioxidants, and the degenerative diseases of aging. *National Academy of Science USA*, 90, 7915-7922.
- Amoo, S. O., Ojo, A. U., & Staden, J. V. (2008). Allelopathic potential of *Tetrapleura tetraptera* leaf extracts on early seedling growth of five agricultural crops. *South African Journal of Botany* 74, 149–152.
- Anjum, T., & Bajwa, R. (2005). Importance of germination indices in interpretation of allelochemical effects. *International Journal of Agriculture and Biology*, 07–3, 417–419.
- Anonymous. (1999). *Annual Reports* Battelle Memorial Institute, USA
- Anonymous. (2000). *Upland Rice Ecosystem: Farmers' Practice of Using Salt for Weed Control in Upland Rice*. Paper presented at the International Rice Research Program Rep.
- Anonymous. (2007). Country report on the state of plant genetic resources for food and agriculture : Food Agriculture Organization of the United Nations Retrieved 7 January 2012, from <http://www.fao.org/docrep/013/i1500e/Lao%20Peoples%20Democratic%20Republic.pdf>

- Anonymous. (2011). Agrofood statistics 2011 : Ministry Of Agriculture And Agro-Based Industry Malaysia. Retrieved 29 December 2011, from [http://www.moa.gov.my/c/document\\_library/get\\_file?uuid=83b5f1e5-21d3-4125-8f6d-59562ee318ab&groupId=10136](http://www.moa.gov.my/c/document_library/get_file?uuid=83b5f1e5-21d3-4125-8f6d-59562ee318ab&groupId=10136)
- AOSA. (Association of Official Seed Analysis) (1983). *Seed Vigour Testing Handbook Contribution No. 32 to the Handbook on Seed Testing*. Springfield, IL, USA.
- Ashrafi, Y. Z., Sadeghi, S., Mashhadi, R. H., & Hassan, A. M. (2008). Allelopathic Effects of sunflower (*Helianthus annuus*) on Germination and Growth of Wild Barley (*Hordeum spontaneum*) *Journal of Agricultural Technology*, 4(1), 219–229.
- Arthey, V. D. (1975). *Quality of Horticultural Products*. Butterworths, London.
- Ayansina, A. D. V., & Osa, B. A. (2006). Effect of two commonly used herbicides on soil microflora at two different concentrations. *African Journal of Biotechnology*, 5(2), 129-132.
- Azania, A. A. P. M., Azania, C. A. M., Alives, P. L. C. A., Palaniraj, R., Kadian, H. S., Sati, S. C., Rawat, L. S., Dahiya, D. S., & Narwal, S. S. (2003). Allelopathic plants. 7. Sunflower (*Helianthus annuus* L.). *Allelopathy Journal*, 11, 1–20.
- Banyikwa, F. F., & Rulangorang, Z. K. (1985). Growth analysis of groundnut (*Arachis hypogea*) in competition with *Ageratum conyzoides*. *Turrialba*, 35, 215–219.
- Barnes, J. P., & Putnam, A. R. (1986). Evidence for allelopathy by residues and aqueous extracts of rye (*Secale cereale*). *Weed Science*, 34:, 384-390.
- Barnes, D. E., & Chan, L. G. (1990). *Common weeds of Malaysia and their control*. Shah Alam, Malaysia.: Ancom.
- Barrett, D. M., Beaulieu, J. C., & Shewfelt, R. (2010). Color, flavor, texture, and nutritional quality of fresh-cut fruits and vegetables: desirable levels, instrumental and sensory measurement, and the effects of processing. *Critical Reviews in Food Science and Nutrition*, 50, 369-389.
- Bartley, I. M., Knee, M., and Casimir, M. A., (1982). Fruit softening. I. Changes in cell wall composition and endopolygalacturonase in ripening pears . *Journal of Experimental Botany* 33,1248-1255.
- Bartley, I., & Knee, M. (1982). The chemistry of textural changes in fruit during storage. *Food Chemistry*, 9, 47-58.

- Batte, M. T., Forster, D. L., & Hitzhusen, J. (1993). Organic agriculture in Ohioan economic perspective. *Journal of Production Agriculture* 6, 536-542.
- Baziramakenga R, Simard RR, Leroux GD, & P, N. (1997). Allelopathic effects of phenolic acids on nucleic acid and protein levels in soybean seedlings. *Canadian Journal of Botany*, 75, 445-450.
- Bear, F. E., Toth, S. J., & Prince, A. L. (1949). Variation in mineral composition of vegetables. *Proceedings of Soil Science Society of America*, 13, 380-384.
- Begum, M., Juraimi, A. S., Syed Omar, S. R., Rajan, A., & Azmi, M. (2008). Effect of herbicides for the control of *Fimbristylis miliacea* (L.) Vahl. in rice. *Journal of Agronomy*, 7(3), 251-257.
- Bell, D. B. (1974). The influence of osmotic pressure in tests for allelopathy. *Transactions of the Illinois State Academy of Science*, 67, 312-317.
- Bell, D. T., & Koeppe, D. E. (1972). Noncompetitive effects of giant foxtail on the growth of corn. *Agronomy Journal*, 64, 321-325.
- Bernat, W., Gawrońska, H., & Gawroński, S. W. (2004). Physiological effects of allelopathic activity of sunflower on mustard. *Advances of Agricultural Sciences Problem Issue*, 496, 271-283.
- Bhatt, B. P., Tomar, J. M. S., & Misra, L. K. (2001). Allelopathic effects of weeds on germination and growth of legumes and cereal crops of North Eastern Himalayas. *Allelopathy Journal*, 18, 225-232.
- Bhowmik, P. C., & Inderjit. (2003). Challenges and opportunities in implementing allelopathy for natural weed management. *Crop Protection*, 22, 661-671.
- Bhowmik, P. C., & Doll, J. D. (1982). Corn and soybean response to allelopathic effects of weed and crop residues. *Agronomy Journal*, 74, 601-606.
- Bhowmik, P. C., & Doll, J. D. (1984). Allelopathic effects of annual weed residues on growth and nutrient up take of corn and soybeans. *Agronomy Journal*, 76, 383-388.
- Blum, U. (1996). Allelopathic interactions involving phenolic acids. *Journal of Nematol*, 28:, 259-267.
- Bode, H. R. (1958). Beiträge zur Kenntnis allelopathischer Erscheinungen bei einigen Juglandaceen. *Planta*, 51:, 440-480.
- Bodovololona, R., Saidou, S., Philippe, L., Olivier, H., Eliane, R., & Narcisse, M. (2008). Effects of living mulches or residue amendments on soil microbial

- properties in direct seeded cropping systems of Madagascar. *Applied Soil Ecology*, 39, 236-3243.
- Boldt, P. E., Rosenthal, S. S., & Srinivasan, R. (1998). Distribution of field bindweed and hedge bindweed in the USA. *Journal Production Agriculture*, 11, 377-381.
- Borland, J. (1990). Mulch: examining the facts and fallacies behind the uses and benefits of mulch. *American Nurseryman* 172, 132-141.
- Boydston, R. A., & Hang, A. (1995). Rapeseed (*Brassica napus*) green manure suppresses weeds in potato (*Solanumtuberosum*). *Weed Technology*, 9, 669-675.
- Brady, N. C., & Weil, R. R. (Eds.). (2008). *Factors influencing the availability of the trace element cations. In The nature and properties of soils*: Pearson Prentice Hall, Upper Saddle River, NJ.
- Brown, P. D., Morra, M. J., McCafrey, M. P., Ulad, D. L., & William III, L. (1991). Allelochemicals produced during glucosinolate degradation in soil. *Journal of Chemical Ecology*, 17(10), 2012-2034
- Buhler, D. D. (2002). Challenges and opportunities for integrated weed management. *Weed Science* 50, 273-280.
- Buhler, D. D., Liebman, M., & Obrycki, J. J. (2000). Theoretical and practical challenge to an IPM approach to weed management. *Weed Science* 48, 274-280.
- Burgos, N. R., Talbert, R. E., Kim, K. S., & Kuk, Y. I. (2004). Growth inhibition and root ultrastructure of cucumber seedlings exposed to allelochemicals from rye (*Secale cereale*). *Journal of Chemical Ecology*, 30(3), 671-689.
- Cacek, T., & Langner, L. (1986). The economic implications of organic farming. *American Journal Alternative Agriculture* 1, 25-29.
- Cameron, H. J., & Julian, G. R. (1980). Inhibition of protein synthesis in lettuce (*Lactuca sativa* L.) by allelopathic compounds. *Journal of Chemical Ecology*, 6:, 989-995.
- Casini, P., Vecchino, V., & Tamantiti, I. (1998). Allelopathic interference of itchgrass and cogongrass: Germination and early development of rice. *Tropical Agriculture*, 75, 445-451.
- Ceccanti, B., Masciandaro, G., & Macci, C. (2007). Pyrolysis-gas chromatography to evaluate the organic matter quality of a mulched soil. *Soil and Tillage Research*, 97, 71-78.



- Cecchi, A. M., Koshinen, W. C., Cheng, H. H., & Haider, K. (2004). Sorption description of phenolic acids as affected by soil properties. *Biology and Fertility of Soils*, 39(4), 235-242.
- Chan, E. W. C., Lim, Y. Y., & Omar, M. (2007). Antioxidant and antibacterial activity of leaves of *Etlingera* species (Zingiberaceae) in Peninsular Malaysia *Food Chemistry*, 104, 1586-1593.
- Cheema, Z. A. (1988). *Weed control in wheat through sorghum allelochemicals*. Ph.D. Thesis, Dept. of Agronomy, Univ. Of Agriculture, Faisalabad.
- Cheema, Z. A., & Khaliq, A. (2000). Use of sorghum allelopathic properties to control weeds in irrigated wheat in a semi arid region of Punjab *Agriculture, Ecosystems & Environment* 79 105-112.
- Cheema, Z. A., Khaliq, A., & Saeed, S. (2004). Weed control in maize (*Zea mays* L.) through sorghum allelopathy. *Journal of Sustainable Agriculture*, 23(4), 73-86.
- Cheng, H. H. (1989). Assessment of the fate and transport of allelochemicals in the soil. In C. S. C. & G. R. Waller (Ed.), *Phytochemical ecology: Allelochemicals, mycotoxins and insect pheromones and allomones*. Academia Sinica Monograph Ser. 9 (pp. 209-215). Taipei, ROC: Institute of Botany.
- Chivinge, O. A. (1985). Allelopathic effects of purple nutsedge (*Cyperus rotundus*) on the growth and development of cotton (*Gossypium hirsutum*), maize (*Zea mays*) and soybeans (*Glycine max*). *Zimbabwe Agricultural Journal*, 82:, 151-152.
- Chou, C. H. (1998). Adaptive autointoxication mechanisms in rice. In M. Olofsdotter (Ed.), *Allelopathy in rice*. Proc. Workshop on Allelopathy in Rice, Manila, Philippines (pp. 99-115). Manila, Philippines: International Rice Research Institute.
- Chon, S. U., Jang, H. G., Kim, D. K., Kim, Y. M., Boo, H. O., & Kim, Y. J. (2005). Allelopathic potential in lettuce (*Lactuca sativa* L.) plants. *Sci Horticult.* , 106, 309-317.
- Chung, I. M., Kim, K. H., Ahn, J. K., Chun, S. C., Kim, C. S., Kim, J. T., et al. (2002). Screening of allelochemicals on barnyardgrass (*Echinochloa crusgalli*) and identification of potentially allelopathic compounds from rice (*Oryza sativa*) variety hull extracts. *Crop Protection*, 21, 913-920.
- Chung, I. M., Kim, K. H., Ahn, J. K., Lee, S. B., Kim, S. H., & Hahn, S. J. (2003). Comparison of Allelopathic Potential of Rice Leaves, Straw, and Hull Extracts on Barnyardgrass. *Agronomy Journal*, 95, 1063-1070.

- Coble, H. D. (1994). Future directions and need for weed science research. *Weed Technology*, 8, 410-412.
- Cobo, J. G., Barrios, E., Kass, D. C. L., & Thomas, R. J. (2002). Decomposition and nutrient release by green manures in a tropical hillside agroecosystem. *Plant and soil*, 240, 331-342.
- Cole, L. C. (1954). The population consequences of life history phenomena. *Quarterly Review of Biology*, 29, 103-137.
- Cordenunsi, B. R., Genovese, M. I., Olivera do Nascimento, J. R., Hassimotto, N. M. A., José dos Santos, R., & Lajolo, F. M. (2005). Effects of temperature on the chemical composition and antioxidant activity of three strawberry cultivars. *Food Chemistry*, 91, 113-121.
- Correa, J. F., Souza, I. F., Ladeira, A. M., & Young, M. C. M. (2000). Allelopathic potential of *Eupatorium maximiliani* Schrad leaves. *Allelopathy Journal*, 7, 225–234.
- Coseteng, M. Y., & Lee, C. Y. (1987). Changes in apple polyphenoloxidase and polyphenol concentration in relation to degree of browning. *Journal of Food Science*, 52(4), 985-989.
- Consumer and Corporate Affairs Canada (1988). Guide for food manufacturers and advertisers. In Consumer and Corporate Affairs Canada (Ed.), *Hull*. Quebec.
- Czabator, F. J. (1962). Germination value: An index combining speed and completeness of pine seed germination. *Forest Science*, 8, 386 – 395.
- Dadzie, B. K., & Orchard, J. E. (1997). *Routine post-harvest screening of banana/plantation hybrids: criteria and method.*: International Plant Genetic Resources Institute.
- David, L. R., & Geoffrey, D. A. (2010). *Phenolic compounds in NaOH extracts of UK soils and their contribution to antioxidant capacity* Paper presented at the World Congress of Soil Science, Soil Solutions for a Changing World.
- Davis, A. S., & Matt, L. (2001). Nitrogen source influences wild mustard growth and competitive effect on sweet corn. *Weed Science Society of America*, 49(4), 558-566.
- Dayan, F. E., Romagni, J. G., & Duke, S. O. (2000). Investigating the mode of action of natural phytotoxins. *J. Chem. Ecol.*, 26, 2079-2094.
- De Datta, S. K., & Baltazar, A. M. (1996). Integrated weed management in rice in Asia. In R. Naylor (Ed.), *Herbicides in Asian Rice: Transitions in Weed Management* (pp. 145–166. ): Palo Alto (California): Institute for International Studies,

Stanford University and Manila (Philippines): International Rice Research Institute.

- Del Moral, R. (1972). On the variability of chlorogenic acid concentration. *Oecologia*, 9, 289-300.
- Devi, L. G., Potty, N. N., Abraham, C. T., & Thomas, G. (1993). The weed flora in sugarcane fields of Palghat District. *J. Trop. Agric.*, 31, 137-139.
- Dhillon, R. S., Singh, S., Kundra, S., & Basra, A.S. (1993). Studies on the chemical composition and biological activity of essential oil from *Cyperus rotundus* Linn. *Plant Growth Regulator*, 13, 89-93.
- Dhole, J. A., Bodke, S. S., & Dhole, N. A. (2011). Allelopathic effect of aqueous leaf extract of *Parthenium hysterophorus* L. on seed germination and seedling emergence of some cultivated crops. *Journal of Research in Biology* 1, 15-18.
- Dielman, A., Hamill, A. S., Fox, G. C., & Swanton, C. J. (1996). Decision rules for postemergence control of pigweed (*Amaranthus* spp.) in soy-bean (*Glycine max*). *Weed Science*, 44, 126-132.
- Diehl, K. C., & Hamann, D. D. (1980). Relationships between sensory profile parameters and fundamental mechanical parameters for raw potatoes, melons and apples. *Journal of Texture Study*, 10, 401-420.
- Dilday, R. H., Nastasi, P., & Smith, R. J. (1989). Allelopathic observation in rice (*Oryza sativa* L.) to duck salad (*Heteranthera limosa*). *Proc. Arkansas Acad. Sci.*, 43, 21-22.
- Di-Matteo, V., & Esposito, E. (2003). Biochemical and therapeutic effects of antioxidants in the treatment of Alzheimer's disease, Parkinson's disease, and amyotrophic lateral sclerosis. *Current Drug Target CNS and Neurological Disorder*, 2, 95-107.
- Drost, D. C., & Doll, J. D. (1980). The allelopathic effect of yellow nutsedge (*Cyperus esculentus*) on corn and soybeans. *Weed Science*, 28, 229-233.
- Duh, P. D., Yen, W. J., Du, P. C., & Yen, G. C. (1997). Antioxidant activity of mung bean hulls. *Journal of the American Oil Chemists' Society*, 74, 1059-1063.
- Duke, S. O. (1985). Biosynthesis of phenolic compounds—Chemical manipulation in higher plants. In A. C. Thompson (Ed.), *The chemistry of allelopathy: Biochemical interactions among plants. American Chemical Society Symposium Series 268* (pp. 113-131). Washington, D.C.: American Chemical Society.



- Duke, S. O., Dayan, F. E., Romagni, J. G., & Rimando, A. M. (2000). Natural products as sources of herbicides: current status and future trends. *Weed Research*, 40:, 99-111.
- Dzyubenko, N. N., & Petrenko, N. I. (1971). On biochemical interaction of cultivated plants and weeds. In A. M. Grodzinsky (Ed.), *Physiological –BioChemical Basis of Plant Interactions in Phytocenoses* (Vol. 2, pp. 60-66). Kiev: Naukova Dumka.
- Einhellig, F. A. (1986). Mechanisms and modes of action of allelochemicals. In A. R. Putnam & T. C-S (Eds.), *The Science of Allelopathy* (pp. 171-188). New York: John Wiley and Sons.
- Einhellig, F. A. (1995). Mechanism of action of allelochemicals in allelopathy. In K. Inderjit, M. M. Dakashini & F. A. Einhellig (Eds.), *Allelopathy: Organisms, Processes, and Applications* (pp. 96-116). Washington, D.C.: American Chemical Society.
- Ekeleme, F., Akobundu, I. O., Isichei, A. O., & Chikoye, D. (2000). Influence of fallow type and land-use intensity on weed seed rain in a forest/savanna transition zone. *Weed Science*, 48, 604–612.
- Ellis, R. A., & Roberts. (1981). The qualification of ageing and survival in orthodox seeds. *Seed Science and Technology*, 9, 373–409.
- Elmore, C. D. (1984). Weed survey-Southern states. *South Weed Sci. Soc. Res. Rep.*, 37, 192-198.
- Erenstein, O. (2002). Crop residue mulching in tropical and semi-tropical countries: An evaluation of residue availability and other technological implications. *Soil & Tillage Research*, 67,115-133.
- F. A. O.(FAO), (2007). *Statistical Database-Agriculture*.
- Fennimore, S., & Smith, R. (2009). *Weed management in lettuce: Monterey County Crop Notes*.
- Fischer, A. J., Ateh, C. M., Bayer, D. E., & Hill, J. E. (2000). Herbicide-resistant *Echinochloa oryzoides* and *E. phyllopogon* in California *Oryza sativa* fields. *Weed Science*, 48, 225-230.
- Forcella, E. J., & Randall, M. (1994). Biology of bull thistle, *Cirsium vulgare* (Savi) Tenore. . *Review of Weed Science*, 6, 29-50.

- Fukumoto, L. R., & Mazza, G. (2000). Assessing antioxidant and prooxidant activities of phenolic compounds. *Journal of Agricultural and Food Chemistry*, 48, 3597-3604.
- Frick, B., & Thomas, A. G. (1992). Weed surveys in different tillage systems in southwestern Ontario field crop. *Canadian Journal of Plant Science*, 72, 1337-1347.
- Garcia, M. A. (1995). Relationship between weed community and seed bank in a tropical agroecosystem. *Agriculture, Ecosystems and Environment*, 55, 139-146.
- García-Orenes, F., Cerdà, A., Mataix-Solera, J., Guerrero, C., Bodí, M. B., Arcenegui, V., et al. (2009). Effects of agricultural management on surface soil properties and soil-water losses in eastern Spain. *Soil and Tillage Research*, 106, 117-123.
- Gerber, M., Boutron-Ruault, M. C., Hereberg, S., Riboli, E., Scalbert, A., & Siess, M. H. (2002). Food and cancer: state of the art about the protective effect of fruits and vegetables. *Bulletin Cancer*, 89, 293-312.
- Gholinejad, B., Asghar, F., Hossein, P., & Hamed, J. J. (2012). Comparative allelopathic effect of *Thymus kotschyanus* on germination and early growth of *Achillea millefolium* under Laboratory and Pot conditions. *Annals of Biological Research* 3 (8), 3978-3983.
- Gleason, M. L., & Iles, J. K. (1998.). Mulch matters. *American Nurseryman* February: 24-31.
- Golueke, C. G. (1972). *Composting - a study of process and its principles*. Penna: Rodale Press Inc.
- Grassmann, J. (2005). Terpenoids as plant antioxidants. In G. Litwack (Ed.), *Plant hormones: vitamins and hormones*. Vol 72. California: Elsevier Academic Press.
- Grichar, W. J., Bessler, B. A., & Brewer, K. D. (2004). Effect of row spacing and herbicide dose on weed control and grain sorghum yield. *Crop Protection*, 23, 263-267.
- Gross, J. (1991). *Pigments in vegetables: chlorophyll and carotenoids*. New York: Van Nostrand Reinhold.
- Grundy, A. C. (1998). The effect of temperature on the viability of weed seeds. *Compost Science and Utilization*, 6(3), 26-33.
- Gupta, K., & Vimala, Y. (2004). Phenolic content of previous crop/crop-weed residues affecting bio-chemical status of wheat seedlings *Abstracts* (pp. 36): IV

International Conference on Allelopathy in Sustainable Terrestrial and Aquatic Ecosystem.

- Gülcin, I., Büyükokuroğlu, M. E., Oktay, M., & Küfrevioğlu, Ö. İ. (2003). Antioxidant and analgesic activities of turpentine of *Pinus nigra* arn.subsp. pallsiana (Lamb.) Holmboe. *Journal of Ethnopharmacology*, 86, 51-58.
- Hader, D. P. (1986). Effects of solar and artificial UV irradiation on motility and phototaxis in the flagellate, *Euglena gracilis*.. *Photochemistry and Photobiology*, 44, 651-656.
- Hagin, R. D. (1989). Isolation and identification of 5-hydroxyindole-3-acetic acid and 5-hydroxytryptophan, major allelopathic aglycones in quackgrass (*Agropyron repens* L. Beauv.). *Journal of Agricultural and Food Chemistry*, 37, 1143-1149.
- Hall, A. B., Blum, U., & Fites, R. C. (1982). Stress modification of allelopathy of *Helianthus annuus* L. debris on seed germination. *American Journal of Botany*, 69, 776-783.
- Halligan, J. P. (1973). Bare areas associated with shrub stands in grasslands: the case of *Artemisia californica*. *Bioscience*, 23, 429-432.
- Halliwell, B., Aeschbach, R., Loliger, J., & Aruoma, O. I. (1995). The characterization of antioxidants. *Food Chemistry Toxicology*, 33, 601-617.
- Hannaway, D. B., Bush, L. P., , & Leggett, J. E. (Eds.). (1980). *Plant Nutrition: Magnesium and Hypomagnesemia in Animals*. (Vol. 716.): Lexington, KY: University of Kentucky, College of Agriculture.
- Haramoto, E. R., & Gallandt, E. R. (2005). Brassica cover cropping: II. Effects on growth and interference of green bean (*Phaseolus vulgaris*) and redroot pigweed (*Amaranthus retroflexus*). *Weed Science*, 53(5), 702-708.
- Hari, O. M., Dhiman, S. D., Kumar, S., & Kumar, H. (2002). Allelopathic response of *Phalaris minor* to crop and weed plants in rice –wheat system. *Crop Protection*, 21, 699-705.
- Harker, F. R., Redgwell, R. J., Hallett, I. C., & Murray, S. H. (1997). Texture of fresh fruit. *Horticultural Review*, 20, 121-224.
- Harmon, R. E., McLaughlin, M. J., Naidu, R., & Correll, R. (1998). Long term changes in cadmium bioavailability in soil. *Environmental Science and Technology*, 32, 3699- 3703.
- Haskin, F. A., & Gorz, H. J. (1985). Dhurrin and p-hydroxybenzaldehyde in seedlings of various sorghum species. *Phytochemistry*, 24, 597-598

- Hassan, G., & Marwat, K. B. (2001). *Integrated weed management in agricultural crops*. Paper presented at the National Workshop on Technologies for Sustainable Agriculture. NIAB, Faisalabad, Pakistan.
- Haugland, E., & Brandsaeter, L. O. (1996). Experiments on bioassay sensitivity in the study of allelopathy. *Journal of Chemical Ecology*, 22, 1845–1859.
- Heap, I. (2006). The International Survey of Herbicide Resistant Weeds. Retrieved from <http://www.weedscience.com>
- Hegab, M. M., Khodary, S. E. A., Ola Hammouda, & Ghareib, H. R. (2008). Ghareib Autotoxicity of chard and its allelopathic potentiality on germination and some metabolic activities associated with growth of wheat seedlings. *African Journal of Biotechnology* 7(7), 884-892.
- Heekman, J. R., & Kluchinski, D. (1995). Soybean nodulation and nitrogen fixation on soil amended with plant residues. Biology and autotoxicity in alfalfa: characterization and effects of preceding crops and residues incorporation. *Crop Science*, 30, 1255-1259.
- Henis, Y. (Ed.). (1986). *Soil microorganism, soil organic matter and soil fertility*. Y. Chen, Y. Avnimelech (Eds.).
- Hess, F. D., & Duke, S. O. (2000). Genetic engineering in IPM: a case study: herbicide tolerance. In G. G. Kennedy & T. B. Sutton (Eds.), *Concepts, Research and Implementation St. Paul, MN* (pp. 126-140). USA: American Phytopathological Society Press.
- Hierro, J. L., & Callaway, R. M. (2003). Allelopathy and exotic plant invasion. *Plant and soil* 256, 29-39.
- Ho, L. T., Lan, P. T. P., Chin, D. V., & Kato-Noguchi, H. (2008). Allelopathic potential of cucumber (*Cucumis sativus*) on barnyardgrass (*Echinochloa crus-galli*). *Weed Biol. and Management.*, 8(2), 129-132.
- Hoagland, R. E., & Williams, R. D. (1985). The influence of secondary plant compounds on the associations of soil microorganisms and plant roots. In A. C. Thompson (Ed.), *The chemistry of allelopathy: Biochemical interactions among plants. American Chemical Society Symposium Series 268* (pp. 301-325). Washington, D.C.: American Chemical Society.
- Hodges, D. M. (2003). Overview: oxidative stress and postharvest produce. In D. M. Hodges (Ed.), *Postharvest oxidative stress in horticultural crops* (pp. 1-12). New York: Food Products Press.

- Hollapa, L. D., & Blum, U. (1991). Effects of exogenously applied ferulic acid, a potential allelopathic compound on leaf growth, water utilization, and endogenous abscisic acid levels of tomato, cucumber and bean. *Journal of Chemical Ecology*, 17(5), 865-886.
- Huang, P. M., Wang, M. C., & Wang, M. K. (1999). Catalytic transformation of phenolic compounds in the soil. In Inderjit *et al.* (Ed.), *Principles and practices in plant ecology: Allelochemical interactions* (pp. 287-306). Boca Raton, FL: CRC Press.
- Hulugalle, N. R., & Palada, M. C. (1990). Effect of seedbed preparation method and mulch on soil physical properties and yield of cowpea in a rice fallows of an inland valley swamp. *Soil and Tillage Research* 17(1-2), 101-113.
- Hutchinson, C. M., & McGiffen, M. E. (2000). Cowpea cover crop mulch for weed control in desert pepper production. *HortScience*, 35, 196-198.
- Ijani, A. S. M., Mabagala, R. B., & Nchimbi-Msolla, S. (2000). Root-knot nematode species associated with beans and weeds in the Morogoro region, Tanzania. *African Plant Protection*, 6, 37-41.
- Inderjit, A. U. (2001). Environmental effects on allelochemical activity. *Agronomy Journal*, 93, 79-84.
- Inderjit, & Bhowmik, P. C. (2002). The importance of allelochemicals in weed invasiveness and the natural suppression. In Inderjit & A. U. Mallik (Eds.), *Chemical Ecology of Plant: Allelopathy of Aquatic and Terrestrial Ecosystems* (pp. 187-192). Basel: Birkhäuser Verlag AG.
- Inderjit, Cheng, H. H., & Nishimura, H. (1999). Plant phenolics and terpenoids: Transformation, degradation, and potential for allelopathic interactions. In S. Inderjit *et al.* (Ed.), *Principles and practices in plant ecology: Allelochemical interactions* (pp. 255-266). Boca Raton, FL: CRC Press.
- Inderjit, & Dakshini, K. M. M. (1995). On laboratory bioassays in allelopathy. *Botany Review*, 61, 28-44.
- Inderjit, S. (1996). Plant phenolic in allelopathy. *Botanical Review*, 62, 186-202.
- Iqbal, J., & Cheema, Z. a. (2008). Purple nutsedge (*Cyperus rotundus* L.) management in cotton with combined application of sorgaab and s-metolachlor *Pakistan Journal of Botany*, 40(6), 2383-2391.
- Ismail, B. S., & John, B. S. (1993). Allelopathic Effect of Lantana (*Lantana camara*) and Siam Weed (*Chromolaena odorata*) on Selected Crops. *Weed Science*, 41, 303-308.



- Jacobi, K. K., Macrae, E. A., & Hetherington, S. E. (2000). Effects of hot air conditioning of 'Kensington' mango fruit on the response to hot water treatment. *Postharvest Biology and Technology*, 21, 39-49.
- Jalili, A., Abbassi, F., & Bazoobandi, M. (2007). Allelopathic influence of canola on germination of five weeds of canola fields. International Workshop on Allelopathy- current trends and future applications, Univ. of Agri., Faisalabad, Pakistan.
- Jadhav, B. B., & Gaynar, D. G. (1992). Allelopathic effect of *Acacia auriculiformis* A. Cunn. on germination of rice and cowpea. *Indian Journal of Plant Physiology*, 35, 86-89.
- Jarvis, S. C., Stockdale, E. A., Shepherd, M. A., & Powlson, D. S. (1996). Nitrogen mineralization in temperate agricultural soils: Processes and measurements. *Advance Agronomy*, 57, 187-235.
- Janiya, J. D., & Moody, K. (1989). Weed populations in transplanted and wet seeded rice as affected by weed control methods *Tropical Pest Management* 35, 8-11.
- Jayakumar, R., & Jagannathan, R. (2007). *Weed Science Principle* India: Kalyani Publishers
- Jen, J. J. (1989). Quality factors of fruits and vegetables. *American Chemical Society Symposium Series* 405, 1.
- Jordán, A., Zavala, L. M., & Muñoz-Rojas, M. (Eds.). (2011) Encyclopedia of Agrophysics, Chapter: Mulching, effects on soil physical properties. Springer.
- Jordán, A., Zavala, L. M., & Gil, J. (2010). Effects of mulching on soil physical properties and runoff under semi-arid conditions in southern Spain. *Catena*, 81, 77-85.
- Kader, A. A. (2002). *Postharvest Technology of Horticultural Crops, 3rd edition*. California: Cooperative Extension, University of California, Division of Agriculture and Natural Resources.
- Kader, A. A. (2000). Quality of horticultural products. *Acta Horticulturae*, 517, 17-20.
- Kamal, J. (2011). Impact of allelopathy of sunflower (*Helianthus annuus* L.) roots extract on physiology of wheat (*Triticum aestivum* L.). *African Journal of Biotechnology*, 10(65), 14465-14477.
- Karakaya, S., El, S. N., & ve Tas, A. A. (2001). Antioxidant activity of some foods containing phenolic compounds. *International Journal of Food Sciences and Nutrition*, 52, 501-508.

- Kashina, B. D., Mabagala, R. B., & Mpunam, A. A. (2003). First report of *Ageratum conyzoides* L. and *Sida acuta* Burm F. as new weed hosts of tomato yellow leaf curl-Tanzania virus. *Plant Protection Science*, 39, 18–22.
- Kassasion, L. (1971). The place of herbicides and weed research in tropical agriculture. *PANS.* , 17(1), 26-29.
- Kato-Noguchi, H. (2001). Assessment of the allelopathic potential of *Ageratum conyzoides*. *Biologia Plantarum*, 44(2), 309–311.
- Kays, J. K. (1999). Preharvest factors affecting appearance. *Postharvest Biology and Technology*, 15, 233-247.
- Khan, A. H., & Vaishya, R. D. (1992). *Allelopathic effects of different crop residues on germination and growth of weeds*. Paper presented at the Proc. First Natl. Symposium: Allelopathy in agroecosystems (agriculture and forestry). CCS. Haryana Agric. Univ., Hisar (India).
- Khanh, T. D., Chung, I. M., Tawata, S., & Xuan, T. D. (2006). Weed suppression by *Passiflora edulis* and its potential allelochemicals. *Weed Research*.
- Khanh, T. D., Chung, I. M., Xuan, T. D., & Tawata, S. (2005). The exploitation of allelopathy in sustainable agricultural production. *Journal of Agronomy and Crop Science*, 191, 172-184.
- Khosla, S. N., & Sobti, S. N. (1981). Effective control of *Parthenium hysterophorus* L. . *Pesticides*, 15, 18-19.
- Kim, S. C., Park, R. K., & Moody, K. (1983). Changes in the weed flora in transplanted rice as affected by introduction of improve rice cultivars and the relationship between weed communities and soil chemical properties. *Res. Rept. ORD.*, 25, 90–97.
- Kim, S. J., & Ishii, G. (2007). Effect of storage temperature and duration on glucosinolate, total vitamin C and nitrate contents in rocket salad (*Eruca sativa* Mill.). *Journal of Science of Food and Agriculture*, 87, 966-973.
- Kirk, G. J. D., Santos, E. E., & Findenegg, G. R. (1999). Phosphate solubilization by organic anion excretion from rice (*Oryza sativa* L.) growing in aerobic soil. *Plant Soil* 211, 11-18.
- Kirk, G. J. D., Santos, E. E., & Santos, M. B. (1999). Phosphate solubilization by organic anion excretion from rice growing in aerobic soil: Rates of excretion and decomposition, effects on rhizosphere pH, and effects on phosphate solubility and uptake. *New Phytologist*, 142, 185-200.

- Koepe, D. E., Rohrbaugh, L. M., Rice, E. L., & Wender, S. H. (1970). The effect of age and chilling temperatures on the concentration of scopolin and caffeoylquinic acids in tobacco. *Physiology Plant*, 23, 258-266.
- Kohli, R. K., & Singh, D. (1991). Allelopathic impact of volatile components from Eucalyptus on crop plants. *Biologia Plantarum*, 33, 475-483.
- Koitaishi, R., Suzuki, T., Kawazu, T., Sakai, A., Kuroiwa, H., & Kuroiwa, T. (1997). 1,8-Cineole inhibits roots growth and DNA synthesis in the root apical meristem of *Brassica campestris* L. *Journal of Plant Research*, 110, 1-6.
- Komai, K., Sugiwaka, Y., & Sato, S. (1981). Plant growth retardant of extracts obtained from water nutgrass (*Cyperus serotinus* Rottb.). *Mem. Fac. Agric. Kinki Univ*, 14, 57-65.
- Krishnamurthy, R. (1978). *A manual on compost and other organic manures*. New Delhi: Today and tomorrow's Printers and Publishers.
- Kumar, K., & Goh, K. M. (2000). Crop residues and management practices: effects on soil quality, soil nitrogen dynamics, crops yield and nitrogen recovery. *Advances in Agronomy*, 68, 197-319.
- Kumar, J. R., & Jagannathan, R. (2003). *Weed Science Principles*. New Delhi: Kalyani Publisher.
- Lal, R., & Kang, B. T. (1982). *Management of organic matter in the tropics*. Paper presented at the Proceeding 11th International Society of Soil Science, New Delhi.
- Leather, G. R. (1983). Weed control using allelopathic crop plants. *Journal of Chemical Ecology*, 9(8), 983-989.
- Leather, G. R. (1983). Sunflower (*Helianthus annuus* L.) is allelopathic to weeds. *Weeds Science*, 31(1), 37-42.
- Lefsrud, M., Kopsell, D., Wenzel, A., & Sheehan, J. (2007). Changes in kale (*Brassica oleracea* L. var. *acephala*) carotenoid and chlorophyll pigment concentrations during leafy ontogeny. *Scientia Horticulturae*, 112, 136-141.
- Liebman, M., & Davis, A. S. (2000). Integration of soil, crop and weed management in low-external-input farming systems. *Weed Res.*, 40, 27-47.
- Levine, M., Conry-Cantilena, C., & Wang, Y. (1996). Vitamin C pharmacokinetics in healthy volunteers: evidence for a recommended dietary allowance. *Proc. Natl. Acad. Sci. USA*, 93, 3704-3709.



- Lincoln, C., & Peirce, J. W. (1987). Vegetable characteristic, production and marketing in New York. *Scientia Horticulturae*, 36, 153-154.
- Lincoln, R., Boxshall, G., & Clark, P. (1998). *A dictionary of ecology, evolution and systematic* (2nd ed.). UK: Cambridge University Press.
- Loux, M. M., & Berry, M. A. (1991). Use of a grower survey for estimating weed problems. *Weed Technology*, 5, 460-466.
- Lovelace, M. L., Talbert, R. E., Dilday, R. H., Scherder, E. F., & Buehring, N. W. (2001). Use of allelopathic rice with reduced herbicide rates for control of barnyardgrass (*Echinochloa crus-galli*). *Arkansas Agricultural Experiment Station Research Series*, 485, 75-79.
- Ludwig, B., Khanna, B. K., Anurugsa, B., & Folster, H. (2001). Assessment of cation exchange capacity and pH buffering in an Amazonian Ultisol. *Geoderma*, 102, 27-40.
- Lurie, S. (2003). Antioxidants. In M. Hodges (Ed.), *Postharvest oxidative stress in horticultural crops* (pp. 131-150). New York: Food Products Press.
- Macias, F. A. (1995). Allelopathy in the search for natural herbicides models. In Inderjit, D. M. M. Dashini & F. A. Einhellig (Eds.), *Allelopathy: organisms, processes, and applications*. (pp. 310-329). Washington (D.C., USA): ACS Symp. Ser. 582.
- Macias, F. A., Molinillo, J. M. G., Varela, R. M., & Torres, A. (1994). Structural elucidation and chemistry of a novel family of bioactive sesquiterpenes: Heliannuols. *Journal of Organic Chemistry*, 59(26), 8261-8266.
- Macias, F. A., Oliva, A. M., Simonet, & Galindo, J. C. G. (1998). What are allelochemicals? In M. Olofsdotter (Ed.), *Allelopathy in Rice* (pp. 69-77). Manila, Phillippines: International Rice Research Institute.
- Macias, F. A., Varela, R. M., Torres, A., & Molinillo, J. M. G. (1993). Potential allelopathic guaianolides from cultivar sunflower leaves, var. SH-222. *Phytochemistry*, 34(3), 669-674.
- Macias, F. A., Varela, R. M. V., Torres, A., Molinillo, J. M. G., & Fronczek, F. (1993). Novel sesquiterpene from bioactive fractions of cultivars sunflowers. *Tetrahedron Letters*, 34, 1999.
- Maciel, C. D. G., Poletine, C. J. R., Aquino Ferreira, D. M., & Maio, R. M. D. (2008). Floristic composition of the weed community in *Paspalum notatum* flügge turf grasses in Assis, sp. *Planta Daninha Viçosa-MG*, 26, 57-64.

- Mahmood, A. (2003). *Utilization of allelopathic properties of sorghum for controlling purple nutsedge (Cyperus rotundus L.) in maize*. University of Agriculture, Faisalabad, Pakistan.
- Malik, C. P., & Singh, M. B. (Eds.). (1980). *Plant enzymology and histo-enzymology*. : Kalyani Publishers.
- Maness, N., & Perkins-Veazie, P. (2003). Soluble and storage carbohydrates. In J. A. Bartz & J. K. Brecht (Eds.), *Postharvest Physiology and Pathology of Vegetables* (pp. 361-382). New York: Marcel Dekker, Inc.
- Mann, J. (1987). *Secondary Metabolism* (2nd ed.). Oxford: Clarendon Press.
- Martens, D. A. (2002). Identification of Phenolic Acid Composition of Alkali-extracted Plants and Soils. *Soil Science Society of American Journal*, 66, 1240-1248.
- Martens, D. A. (2000a). Management and crop residue influence soil aggregate stability. *Journal of Environmental Quality*, 29, 723-727.
- Masia, A. (2003). Physiological effects of oxidative stress in relation to ethylene in post-harvest produce. In D. M. Hodger (Ed.), *Postharvest Oxidative Stress in Horticultural Crops* (pp. 165-197). New York: Food Products Press.
- McClosky, W. B., Baker, P.B. and Sherman, W. (1998). *Survey of cotton weeds and weed control practices in Arizona upland cotton fields*. College of Agriculture, University of Arizona: Publication AZ1006 cotton.
- Meentemeyer, V. (1978). Macroclimate and lignin control of litter decomposition. *Ecology*, 59, 405-472.
- Miliauskas, G., Venskutonis, P. R. & van Beek, T. A. (2004). Screening of radical scavenging activity of some medicinal and aromatic plant extracts. *Food Chemistry*, 85, 231-237.
- Minorsky, P. V. (2002). Allelopathy and grain crop production. *Plant Physiology*, 130, 1745-1746.
- Mokhtar, I. M. (2008). *Agriculture Statistical Handbook: Kementrian Pertanian dan Industri Asas Tani Malaysia*.
- Molisch, H. (1937). *Der Enfusslinier pflanze aufdie andere-Allelopathic Fischer*. Jena, GDR.
- Moosavi A., Afshari R. T., & Asadi A. (2011). Allelopathic effects of aqueous extract of leaf stem and root of *Sorghum bicolor* on seed germination and seedling growth of *Vigna radiata* L. *Notulae Scientia Biologicae*, 3(2), 114-118.

- Morohashi, Y., & Suguimoto, M. (1988). ATP synthesis in cotyledons of cucumber and mung bean seeds during the first hours of inhibition. *Plant Cell Physiology*, 29, 893-896.
- Mozafar, A. (1993). Nitrogen fertilizers and the amount of vitamins in plants *Journal of Plant Nutrition*, 16(12), 2479-2506.
- Muller, C. H. (1969). Allelopathy as a factor in ecological process. *Vegetatio*, 18, 348.
- Narwal, S. S. (2003). Allelopathic plants. 7. Sunflower (*Helianthus annuus* L.). *Allelopathy Journal*, 11, 1-20.
- Naseem, M. (1997). *Allelopathic effects of autumn sunflower residues on wheat productivity and wheat-weeds*. Ph.D. Thesis, Dept. of Agronomy Univ. of Agriculture, Faisalabad, Pakistan.
- Naseem, M., Aslam, M., Ansar, M., & Azhar, M. (2009). Allelopathic effects of sunflower water extract on weed control and wheat productivity. *Pakistan Journal of Weed Science Research*, 15(1), 107-116.
- Naseem, M., Cheema, Z. A., & Bazmi, S. A. (2003). Allelopathic effects of sunflower aqueous extracts on germination of wheat and some important wheat weeds. *Pakistan Journal of Scientific Research*, 55 (3-4), 71-75.
- Navarez, D., & Olofsdotter, M. (1996). *Relay seeding technique for screening allelopathic rice (Oryza sativa)*. Paper presented at the Proceeding 2nd International Weed Control Congress, Copenhagen.
- Nelson, C. J. (1996). Allelopathy in cropping system. *Agronomy Journal* 88, 991-996.
- Nerd, A., & Mizrahi, Y. (1997). Reproductive biology of cactus fruit crops. *Horticulture Review*, 18, 321-346.
- Nielsen, K. A., Tattersall, D. B., Jones, P. R., & Møller, B. L. (2008). Metabolon formation in dhurrin biosynthesis. *Phytochemistry*, 69, 88-98.
- Nsolomo, V. R., Mrecha, M. S., & Maghembe, J. A. (1995). Effect of *Acacia xanthopholea* leachates on seed germination of some agriculture and multipurpose tree crops. *Journal of Tropical Forest Science*, 7:, 398-404.
- O' Donovan, J. T., Harker, K. N., Clayton, G. W., Newman, J. C., Robinson, D., & Hall, L. M. (2001). Barley seedin rate influence the eeffects of variable herbicide rates. *Weed Science*, 49, 746-754.

- Oerke, E. C., & Steiner, U. (1996). *pflanzenschutz, Schriftenreihe der Deutschen Phytomedizinischen Gesseschaft*. Stuttgart: Euge Ulmer Verlag.
- Ogbodo, E. N. (2011). Effect of tillage methods on some soil properties and rice yields on an acid utisol of Abakaliki, southern Nigeria. *Nigerian Journal of Soil Science* 21.
- Oktay, M., Gülcin, I., & Küfrevioğlu, Ö. İ. (2003). Determination of in vitro antioxidant activity of fennel (*Foeniculum vulgare*) seed extracts. *Lebensmittel Wissenschaft and Technologie*, 36, 263-271.
- Olasantan, F. O. (1999). Effect of time mulching on soil temperature and moisture regime and emergence, growth and yield of white yam in western Nigeria. *Soil and Tillage*, 50(3-4), 215-221.
- Oleszek, W., & Jurzysta, M. (1987). The allelopathic potential of alfalfa root medicagenic acid glycosides and their fate in soil environments. *Plant and soil*, 98(67-80).
- Oliveria, D. S., Oliveira, R. D. I., Freitas, L. G., & Silva, R. V. (2004). Variability of *Meloidogyne exigua* on coffee in the Zona da Mata of Minas Gerais State, Brazil. *Journal of Nematology*, 37(3), 323-327.
- Olofsdotter, M., Navarez, D., & Rebulanan, M. (1997). Rice allelopathy - where are we and how far can we get? . *Brighton Crop Protect. Conf. Weeds*, 1, 99-104.
- Om, H., Dhiman, S. D., Kumar, S., & Kumar, H. (2002). Allelopathic response of *Phalaris minor* to crop and weed plants in rice – wheat system. *Crop Protection* 21, 699–705.
- Opara-Nadi, O. A., & Lal, R. (1987). Influence of method of mulch application on growth and yield of tropical root crops in southeastern Nigeria. *Soil Tillage Research*, 3, 217-230.
- Pandey, D. K. (1994). Inhibition of *Salvinia* (*Salvinia molesta* Mitchell) by parthenium (*Parthenium hysterophorus* L.). I. Effect of leaf residue. *Journal of Chemical Ecology*, 20, 3111-3122.
- Pantastico, E. B., DeFossard, R. A., & Safran, H. (1975). *Physiological disorders other than chilling injury*. Westport, CT: The AVI Publishing Co., Inc.
- Pantastico, E. B., Subramanyam, H., Bhatti, M. B., Ali, N., & Akamine, E. K. (1975). Postharvest physiology: harvest indices. In E. B. Pantastico (Ed.), *Postharvest physiology, Handling and Utilization of Tropical and Sub-tropical Fruits and Vegetables* (pp. 56). Westport: AVI Publishing.

- Pariasca, J. A. T., Miyazaki, T., Hisaka, H., Nakagawa, H., & Sato, T. (2000). Effect of modified atmosphere packaging (MAP) and controlled atmosphere (CA) storage on the quality of snow pea pods (*Pisum sativum* L. var. *saccharatum*). *Postharvest Biology and Technology*, 21, 213-223.
- Pejman, N., Hassan, K., Morteza, M., & Nayereh, A. S. H. (2011). Allelopathic potential of sunflower on weed management in safflower and wheat *Australian journal* 5(11), 1434-1440.
- Phuong, L. T., Denich, M., Vlek, P. L. G., & Balasubramaniam, V. (2005). Suppressing weeds in direct seeded lowland rice: effects of methods and rates of seeding. *Journal of Agronomy and Crop Sciences*, 191, 185-194.
- Pitt, R. E., & Chen, H. L. (1983). Time-dependent aspects of the strength and rheology of vegetative tissue. *Trans. ASAE*, 26, 1275-1280.
- Promila, R., Lin, Y., & Shetty, K. (2004). Stimulation of phenolics, antioxidant and antimicrobial activities in dark germinated mungbean sprouts in response to peptide and phytochemical elicitors. *Process Biochemistry*, 30, 637-646.
- Putnam, A. R., & Duke, S. O. (1974). Biological suppression of weeds: evidence for allelopathy in accessions of cucumber. *Science*, 185, 370-372.
- Putnam, A. R., & DeFrank, J. (1979). Use of cover crops to inhibit weeds. *In Proceeding IX International Congress Plant Protection*, 580-582.
- Putnam, A. R., & Tang, C. S. (1986). *The Science of allelopathy*. New York, NY.: John Wiley & Sons.
- Putnam, A. R. (1988). Allelochemical from plants as herbicides. *Weed Technology*, 2, 510-518.
- Qasem, J. R. (1995). The allelopathic effect of three *Amaranthus spp.* (Pigweeds) on wheat (*Triticum durum*). *Weed Research*, 35, 41-49.
- Qasem, J. R., & Foy, C. L. (2001). Weed Allelopathy, its ecological impacts and future prospects: a review. *Journal of Crop Production*, 4, 43-119.
- Rahman, M. M., Sahid, I. B., & Juraimi, A. S. (2010). Study of resistant biotypes of *Echinochloa crus-galli* in Malaysia. *Australian Journal of Crop Science*, 4(2), 107-115.
- Ranade, S. D., & Burns, W. (1925). The eradication of *Cyperus rotundus* L. *Memoirs of Indian Department of Agriculture. Botanical Series*, 13, 99-192.



- Randhawa M. A., Cheema Z. Ali M. A. (2002). Allelopathic effect of sorghum water extract on germination and seedling growth of *Trianthema portulacastrum*. *International Journal of Agriculture Biology*, 4, 383-384.
- Ranganna, S. (Ed.). (1977). *Manual of analysis of fruits and vegetables products*. New Delhi: McGraw Hill Publication & Co.
- Rasmussen, J., & Accard, J. (1995). Weed control in organic farming systems. In D. M. Glen, M. P. Greaves & H. M. Anderson (Eds.), *Ecology and Integrated Farming Systems* (pp. 49-67). Chichester, U.K: Wiley.
- Reigosa, M. S., Gonzales, L., Souto, X. C., & Pastoriza, J. E. (2000). Allelopathy in forest ecosystem. In S. S. Narwal (Ed.), *Allelopathy in Ecological Agriculture and Forestry* Kluwer Academic Publishers.
- Reuss, J., Dooley, H. L., Griffis, W., & (1976). *Plant uptake of cadmium from phosphate fertilizer*: Corvallis or Ecological Research Service.
- Reynolds, M. P., & Borlaug, N. E. (2006). Applying innovations and new technologies for international collaborative wheat improvement . *J. Agric Sci. , 144*, 95-110.
- Rice, E. L. (1974). New York: Allelopathy. Academic Press Inc
- Rice, E. L. (1979). Allelopathy: an up to date. *Botanical review*, 45, 15-109.
- Rice, E. L. (1984). *Allelopathy* (2nd ed.). New York: Academic Press.
- Rice, E. L. (1995). *Biological control of weeds and plant diseases: Advances in applied allelopathy*. Norman, OK.: University of Oklahoma Press.
- Richardson, P. J., Webster, A. D., & Quinlan, J. D. The effect of paclobutrazol sprays with or without the addition of surfactants on the shoot growth, yield and fruit quality of the cultivars Cox and Suntan. *Journal of Horticultural Science*, 61, 439-446.
- Ridenour, W. M., & Callaway, R. M. (2001). The relative importance of allelopathy in interference : the effects of invasive weed of native bunchgrass. *Oecologica*, 126, 444-450.
- Rimmer, D. L., & Abbott, G. D. (2011). Phenolic compounds in NaOH extracts of UK soils and their contribution to antioxidant capacity *European Journal of Soil Science*, 62(2), 285-294.
- Robards, K., Prenzler, P. D., Tucker, G., Swatsitang, P., & Glover, W. (1999). Phenolic compounds and their role in oxidative processes in fruits. *Food Chemistry*, 66, 401-436.

- Robert, M. M. (1988). Allelopathic Effects of Palmer Amaranth (*Amaranthus palmeri*) on Seedling Growth. *Weed Science*, 36, 325-328.
- Roe, N. E., Stoffella, J., & Bryan, H. H. (1993). Municipal solid waste compost suppresses weeds in vegetable crop alleys. *HortScience*, 28, 1171-1172.
- Romeo, J. T., & Weidenhamer, J. D. (1999). Bioassays for allelopathy in terrestrial plants. In K. F. Haynes & J. G. Millar (Eds.), *Methods in chemical ecology* (pp. 179-211). Boston: Kluwer Academic Publishing.
- Rosa, S. R. (2006). *Improving postharvest management and marketing in the asia-pacific region: issues and challenges*. India: APO.
- Roy, S. K., & Chakrabarti, A. K. (2003). Vegetables of temperate climates: commercial and dietary importance. In B. Caballerom, L. C. Trugo and P. M. Finglas (Ed.), *Encyclopedia of Food Sciences and Nutrition* (2nd ed., Vol. Nine, pp. 5925-5932): Academic Press.
- Rubatzky, V. E., & Yamaguchi, M. (1997). *World Vegetables*. New York: Chapman & Hall.
- Ryall, A. L., & Lipton, W. J. (1972). *Handling, transportation and storage of fruits and vegetables* (Vol. 1). Westport: AVI Publishing.
- Ryder, E. J. (1999). Genetics in lettuce breeding: past, present and future. In A. Lebeda & E. Křístkov (Eds.), *Eucarpia Leafy Vegetables '99*. (pp. 225-231). Olomouc: Palacky University.
- Sanchez, P. A. (1976). *Properties and management of soils in the tropics*. New York: Wiley and Sons.
- Saghir, A. R., & Bhatti, S. M. (1970). *The influence of herbicides on the chemical composition of soybean seeds*. Paper presented at the Proc. 10th British Weed Control Conf. 1.
- Salunkhe, D. K., & Desai, B. B. (Eds.). (1988). *Effects of agricultural practices, handling, processing, and storage on vegetables*. In *Nutritional Evaluation of Food Processing*. : Avi Book, Van Nostrand Reinhold Co., New York.
- Saxena, A., Singh, D. V., & Joshi, N. I. (1996). Allelopathy in agroecosystem. *Field Crop Abstracts*, 49(10), 891-899.
- Schuab, S. R. P., Braccini, A. L., Ferrares-Filho, O. F., Scapim, C. A., & Braccini, M. C. L. (2001). Physiological seed quality evaluation and seedling lipid and protein

content of soybean (*Glycine max* L.) in the presence of pcoumaric acid. *Seed Science and Technology*, 29, 151-162

- Shad, R. A. (1987). Status of weed science activities in Pakistan. *Program Farm*, 7(1), 10-16.
- Shahid, M., B. Ahmad, Khattak, R. A., Hassan, G., & Khan, H. (2006). Response of wheat and its weeds to different allelopathic plant water extract. *Pakistan Journal of Weed Science Research*, 12(1-2), 61-68.
- Shamshuddin, J., & Che Fauziah, I. (2010). Alleviating Acid Soil Infertility Constraints Using Basalt, Ground Magnesium Limestone and Gypsum in a Tropical Environment. *Malaysian Journal of Soil Science*, 14, 1-13.
- Serafini, M., Bellocco, R., Wolk, A., & Ekstrom, A. M. (2002). Total antioxidant potential of fruit and vegetables and risk of gastric cancer. *Gastroenterology* 123, 985-991.
- Seigler, D. S. (1996). Chemistry and mechanisms of allelopathic interactions. *Agronomy Journal*, 88, 876-885.
- Silva, E. M., Souza, J. N. S., Rogez, H., Rees, J. F., & Larondelle, Y. (2007). Antioxidant activities and polyphenolic contents of fifteen selected plant species from Amazonian region. *Food Chemistry*, 101, 1012-1018.
- Silvertown, J., & Dodd, M. (1997). Comparing plants and connecting traits. In J. Silvertown, M. Franco & J. L. Harper (Eds.), *Plant life histories: ecology, phylogeny and evolution*. (pp. 3-16). Cambridge: Cambridge University Press.
- Silvertown, J., & Charlesworth, D. (2001). *Introduction to plant population biology* (4th ed.). Ames: Blackwell Science, Iowa State Press.
- Singh, S. K., & Saha, G. P. (2001). Effect of weed management practices on performance of prominent wet-season crops. *Indian Journal of Agronomy*, 46, 489-495
- Singh., H. P., Batish, D. R., & Kohli, R. K. (2002). Allelopathic effect of two volatile monoterpenes against bill goat weed (*Ageratum conyzoides* L.). *Crop Protection*, 21, 347-350.
- Singh, G. (1994). Management and utilization of palm oil by-products. In Koh M.P et al. (Ed.), *Proceedings of Third National Seminar on Utilisation of Oil Palm Tree and Other Palms* (pp. 19-48): Oil Palm Trunk Utilisation Committee.
- Smirnoff, N. (1996). The function and metabolism of ascorbic acid in plants. *Annual Botany*, 78, 661-669.



- Smith, J. S. (2000). Contaminants and indirect additives. In J. S. Smith (Ed.), *Food Chemistry: Principles and Applications*. California: Science Technology System.
- Smith Jr, R. J. (1992). *Biological control as components of integrated weed management for rice in the U. S.* Paper presented at the International Symposium on Biological Control and Integrated Management of Paddy and Aquatic Weeds in Asia, Sukuba, Japan.
- Smith, R., & Fennimore, S. (2009, August). Spinach weed control update. *Monterey County Crop Notes*.
- Srivastava, J. P., Tamboli, P. M., English, J. C., Lal, R., & Stewart, B. A. (1993). Conserving soil moisture and fertility in the warm seasonally dry tropics. *World Bank Technical Paper Number*, 221.
- Stall, W. M., & MacRae, A. W. (2009). *Weed Management*: University of Florida
- Hollapa, L. D., & Blum, U. (1991). Effects of exogenously applied ferulic acid, a potential allelopathic compound on leaf growth, water utilization, and endogenous abscisic acid levels of tomato, cucumber and bean. *Journal of Chemical Ecology*, 17(5), 865-886.
- Stevenson, F. J., & Ardakani, M. S. (Eds.). (1972). *Organic matter reactions involving micronutrients in soils* J.J. Mortvedt et al. (ed.) *Micronutrients in agriculture*. SSSA, Madison, WI.
- Stinson, J. M., Brinen, G. H., McConnell, D. B., & Black., R. J. (1990). Evaluation of landscape mulches. . *Proceedings of Florida State Horticultural Society*, 103, 372-377.
- Swanton, C. J., & Weise, S. F. (1991). Integrated weed management: the rationale and approach. *Weed Technology*, 5, 657-663.
- Tang, C., Sparling, G. P., Mc Lay, C. D. A., & Raphael, C. (1999). The effect of short-term legume residue decomposition on soil activity. *Analytical Soil Research*, 37, 561-573.
- Tehmina, A., & Rukhsana, B. (2005). Importance of Germination Indices in Interpretation of Allelochemical Effects *Int. J. Agri. Biol.*, 7(3), 417-419.
- Teo, C. K. H., & Nishimoto, R. K. (1973). Cytokinin - enhanced sprouting of purple nutsedge as a basis for control. *Weed Research*, 13, 118-121.

- Thi, H. L., Lan, P. T. P., Chin, D. V., & Kato-Noghuci, H. (2008). Allelopathic potential of cucumber (*Cucumis sativus*) on barnyardgrass (*Echinochloa crus-galli*). *Weed Biology and Management*, 8, 129–132
- Tian, G., Olimah, J. A., Adeoye, G. O., & Kang, B. T. (2000). Regeneration of Earthworm Populations in a Degraded Soil by Natural and Planted Fallows under Humid Tropical Conditions. *Soil Science Society of American Journal*, 64, 222-228.
- Tian, G., Bussard, L., & Kang, B. T. (1992). Mulching effect of plant residues with chemically contrasting composition of maize growth and nutrients accumulation. *Plant and Soil*, 153, 179-187.
- Tiquia, S. M., Lloyd, J., Herms, D. A., Hoitink, H. A. J., & Michel Jr, F. C. (2002). Effects of mulching and fertilization on soil nutrients, microbial activity and rhizosphere bacterial community structure determined by analysis of TRFLPs of PCR-amplified 16S rRNA genes. [doi: 10.1016/S0929-1393(02)00040-9]. *Applied Soil Ecology*, 21(1), 31-48.
- Thorne, R. L. Z., Walker, G. R., McPherson, J., K., Krenzer, E. G. J., & Young, C. C. (1990). Autotoxic effects of old and new wheat straw in conventional-tillage wheat soil. *Botanical Bulletin of Academia Sinica*, 31, 35-39.
- Thomas, A. G. (1985). Weed survey system used in Saskatchewan for cereal and oilseed crops. *Weed Science*, 33, 34–43.
- Thomas, A. G., & Wise, R. F. (1987). *Weed Survey of Saskatchewan for Cereal and Oilseed Crops*. Saskatchewan: Publication 87-1. Agriculture Can. Regina.
- Thompson, H. C., & Kelly, W. C. (1957). *Vegetable Crop* (5th ed.). New York: McGraw-Hill.
- Tooke, H. L., Van Etten, C. H., & Daxenbichler, M. E. (1980). Glucosinates. In I. E. Liener (Ed.), *Toxic Constituents of Plant Foodstuffs* (2nd ed., pp. 103-142). New York: Academic Press.
- Tovar, B., Garcia, H. S., & Mata, M. (2001). Physiology of pre-cut mango II. Evolution of organic acids. *Food Review International*, 34, 705-714.
- Troeh, F. R., & Thompson, L. M. (Eds.). (2005). *Soils and soil fertility*: 6th ed. Blackwell Publ. Ames, IA.
- Tuba, A., & Gülçin, I. (2008). Antioxidant and radical scavenging properties of curcumin. *Chemico-Biological Interactions*, 174, 27-37.

- Tuffi, S. L. D. (2004). Survey fitossociológico in conditions of degraded grassland plains. *Planta Daninha* 22, 343–349.
- Tukey, H. B. J. (1969). Implications of allelopathy in agricultural plant science. *Bot. Rev.* 35, 1-16.
- Tukey, H. B. J. (1971). Leaching of substances from plants *Biochemical Interactions Among Plants* (pp. 25-32). Washington, D.C.: National Academy of Sciences.
- Uddin, M. K., Juraimi, A. S., Begum, M., Ismail, M. R., Rahim, A. A., & Othman, R. (2009). Floristic composition of weed community in turf grass area of west peninsular Malaysia. *International Journal Agriculture Biology*, 11, 13–20.
- Uddin, M. K., Juraimi, A. S., Ismail, M. R., & Brosnan, J. B. (2010). Characterizing Weed Populations in Different Turfgrass Sites throughout the Klang Valley of Western Peninsular Malaysia. *Weed Technology*, 24, 173-181.
- Uygun, F. N., & Skendero, N. (1995, 26-29 September 1995). *Allelopathic and Bioherbicidal Effect of the Parts of Plant Residues on the Growth of Both Weeds and Corn, VII*. Paper presented at the Turkish Phytopathology Congress, Adana-Turkey.
- Valverde, B. E., Riches, C. R., & Caseley, J. C. (2000). Prevention and management of herbicide-resistant weeds in rice: Experiences from Central America with *Echinochloa colona* (pp. 123). Costa Rica: Cámara de Insumos Agropecuarios.
- Van Etten, C. H., & Tookey, H. L. (1983). Glucosinates. In M. R. Jr (Ed.), *CRC Handbook of Naturally Occurring Food Toxicants* (pp. 15-30). Boca Raton: CRC Press.
- Vaughn, S. F., & Spencer, G. F. (1993). Volatile monoterpenes as potential parent structures for new herbicides. *Weed Science*, 41, 114-119.
- Vaughn, K. C., Hoffman, J. C., Hahn, M. G., & Staehelin, L. A. (1996). The herbicide dichlobenil disrupts cell plate formation: immunogold characterization. *Protoplasma*, 194, 117–132.
- Velimirov, A., Plochberger, K., Huspeka, U., & Schott, W. (1992). The influence of biologically and conventionally cultivated food on the fertility of rats. *Biological Agriculture and Horticulture*, 8, 325-337.
- Verhulst, N., Bram, G., Els, V., Fabian, K., Limon-Ortega, A., & Jozef, D. (2009). *The Importance of Crop Residue Management in Maintaining Soil Quality in Zero Tillage Systems; A Comparison between Long-term Trials in Rainfed and Irrigated Wheat Systems*. Paper presented at the 4th World Congress on Conservation Agriculture.

- Vries, I. M. d. (1997). Origin and domestication of *Lactuca sativa* L. . *Genetic Resources and Crop Evolution*, 44, 165-174.
- Wan Asma, I., Wan Rasidah, K., Rosenani, A. B., & Aminuddin, H. (2004). *The application of EFB mulch mat from oil palm empty fruit bunches to sentang seedling grown on sandy soil*. Paper presented at the Proceedings of the conference on forestry and forest Product Research.
- Wardle, D. A., Ahmad, M., & Nicholson, K. S. (1991). Allelopathic influence of nodding thistle (*Carduus nutans* L.) seeds on germination and redicle growth of pasture plants. *New Zealand Journal of Agricultural Research*, 34, 185–191.
- Watanabe, K., Ohno, N., Yoshioka, H., Gershenzon, J., & Mabry, T. J. (1982). Sesquiterpene lactones and terpenoids from *Helianthus argophyllus*. *Phytochemistry*, 21, 709-713
- Watkins, C. B., & Ekman, J. H. (2005). How postharvest technologies affect quality. In S. Ben-Yehoshua (Ed.), *Environmentally Friendly Technologies for Agricultural Produce Quality* (pp. 447-491). Bota Raton: Taylor & Francis Group.
- Watson, G. W. (1988.). Organic mulch and grass competition influence tree root development. *Journal of Arboriculture*, 14(8), 200-203.
- Watts, R. L., & Watts, G. S. (1954). *The Vegetable Growing Business*. New York: Orange Judd Publishing.
- Webster, T. M., & Coble, H. D. (1997). Changes in the weed species composition of the southern United States: 1974–1995. *Weed Technology*, 11, 308–317.
- Webster, T.M. & MacDonald G. E. (2001). A Survey of Weeds in Various Crops in Georgia. *Weed Technology*, 15, 771-790.
- Weston, L. A., & Duke, S. O. (2003). Weed and crop allelopathy. *Critical Review in Plant Science*, 22, 367-389.
- Weston, L. A. (1996). Utilization of Allelopathy for Weed Management in Agroecosystems. *Agronomy Journal*, 88, 860-866.
- Wezel, A. (2000). Weed vegetation and land use of upland maize fields in northwest Vietnam. *GeoJournal*, 50, 337–349.
- Whittaker, T. W., Ryder, E. J., Rubatzky, V. E., & Vail, P. E. (1974). *Lettuce production in the Unite States* (Vol. 221). Washington D. C.: U. S. Department of Agriculture.

- Whittaker, R. H. (1970). The biochemical ecology of higher plants. In E. Sondheimer & J. B. Simeone (Eds.), *Chemical Ecology*. New York: Academic Press.
- Wilcox, M. (1994). Cultural practices in the production of iceberg lettuce in southwestern Arizona. *Arizona Cooperative Extension*.
- Wills, R. B. H., McGlasson, W. B., Graham, D., Lee, T. H., & Hall, E. G. (2004). *Postharvest; An introduction to the physiology and handling of fruit and vegetables*. Australia: New South Wales Univ. Press.
- Wills, R. B. H., McGlasson, W. B., Graham, D., & Joyce, D. C. (2007). *Postharvest : An introduction to the physiology and handling of fruit, vegetables and ornamentals 5th Edition* (Vol. 5). Sydney: University of New South Wales Press.
- Wembeke, V. A. (Ed.). (1992). *Soil of the tropics: Properties and appraisal*. New York: McGraw-Hill
- Weston, L. A. (1996). Utilization of allelopathy for weed management in agro-ecosystems. *Agronomy Journal*, 88, 860-866.
- Wood, L. G., Gibson, P. G., & Garg, M. L. (2006). A review of the methodology for assessing in vivo antioxidant capacity. *Journal of the Science of Food and Agriculture*, 86, 2057-2066.
- Xing, A. C., Qiang, W., Ping, Z. A., Fen, D., & Ming, D. (2000). Survey of weeds in turf in Hangzhou. *Acta Agriculturae Zhejiangensis*, 12, 360-362.
- Yamaguchi, M. (1983). Composites *World Vegetables* (pp. 207 -213). California: The Avi Publisher, Inc.
- Yongqing, M. A. (2005). Allelopathic studies of common wheat (*Triticum aestivum* L). *Weed Biology and Management*, 5, 93-104.
- Yu, J. Q., & Matsui, Y. (1993). p-Thiocyanatophenol, a novel allelopathic compound in exudates from root of cucumber. *Chem. Express*, 8, 577-580.
- Yu, J. Q., & Matsui, Y. (1994). Phytotoxic substances in the root exudates of *Cucumis sativus* L. *Journal of Chemical Ecology*, 20, 21-31.
- Zink, F. W., & Yamaguchi, M. (1962). Studies on the growth rate and nutrientabsorption of head lettuce. *Hilgardia*, 32, 471-500.
- Zhou, Y., Han, Y. J., Li, Z. G., Fu, Y., Fu, Z. Y., Xu, S. T., et al. (2012). ZmcrtrB3 encodes a hydroxylase that effects the accumulation of  $\alpha$ -carotene in maize kernel. *Journal of Integrative Plant Biology*, 54(4), 210-214.