



**UNIVERSITI PUTRA MALAYSIA**

***PURIFICATION AND CHARACTERIZATION OF MALAYSIAN  
MAHSEER (TOR TAMBROIDES BLEEKERS) VITELLOGENIN***

**NURUL ASHIKIN BINTI MUHAMMAD**

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MAHSEER (TOR TAMBROIDES BLEEKERS) VITELLOGENIN**

**By**

**NURUL ASHIKIN BINTI MUHAMMAD**

**Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia,  
in Fulfillment of the Requirements for the Degree of Master of Science**

**February 2013**

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## DEDICATIONS

*This work is dedicated to my dearest parents, Muhammad bin Ismail and Hanisah binti Jaafar, and my loving siblings Nooraini Azizah, Nooraini Fazilah, Muhammad Munawwar, Nurul Aishah and Muhammad Shukri. Thank you for the never ending support and undeniable love. Without them, the completion of this study might be impossible.*

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*Thank you,*

***Nurul Ashikin Muhammad***

**UNIVERSITI PUTRA MALAYSIA**

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment  
of the requirement for the degree of Master of Science

**PURIFICATION AND CHARACTERIZATION OF MALAYSIAN  
MAHSEER (TOR TAMBROIDES BLEEKERS) VITELLOGENIN**

By

**NURUL ASHIKIN BINTI MUHAMMAD**

**February 2013**

**Chair: Annie Christianus, PhD**

**Faculty: Agriculture**

Vitellogenin (vtg) is a high molecular weight glycopospholipoprotein synthesized in the liver under stimulation of estrogen. Basically found in sexually mature female, vtg is taken up by developing oocytes during maturation. It functions as a nutrient storage for growing embryo. Vtg has the potential as a maturation indicator for the successful production of fish fry. *Tor tambroides* is one of the most sought after fish in Malaysia, used as a game fish and economically important as cultured species. Main problem in the mass production of *T. tambroides* fry in hatchery is in the availability and selection of matured broodstock. Identification of matured and ready females morphologically can only be done by experience workers. Therefore, establishment of simple technique to identify matured females is necessary. Hence, in this current study, enzyme linked immunosorbent assay (ELISA) has been developed to measure blood plasma vtg as female maturation indicator. The

development of this indicator will definitely contribute to the hatchery production of *T. tambroides* fry.

Estrogen ( $17\beta$ -estradiol) was injected intra-peritoneally into five males ( $1.5 \pm 0.4$ kg). In order to confirm the synthesis of vitellogenin, raw plasma from E2-treated male, vitellogenic female and non-treated male were subjected to SDS-PAGE analysis. The presence of 149kDa protein band in E2-treated male plasma indicated the secretion of vitellogenin. Plasma samples were purified by size exclusion chromatography to separate protein particles according to molecular sizes. Fractions containing the bell-shaped major peaks were collected and subjected to native PAGE. Molecular weight of protein bulk was determined as 700kDa band.

Further reduction of the protein bulk by SDS-PAGE resulted in the appearance of protein bands with similar positions in E2-treated sample and female whereas non-treated male showed no similarity. Specificity of antibody in Western blot revealed that in purified E2-treated male plasma, only three bands (133kDa, 117kDa, 56kDa) were recognized by anti-carp monoclonal antibody and thus identified as vtg. Bands with similar positions were also detected in mature female plasma. Male plasma did not show any cross reactivity against antibody.

Enzyme-linked immunosorbent assay (ELISA) was developed for measurement of vtg concentration at plasma level. Purified vtg (253 ng/ml) was coated on 96-well microplate. Plasma samples were diluted with 1:500 antiserum to a final dilution of

1:1000. Linearization of binding displacement curves by logit transformation revealed that serial dilutions of mature female mahseer plasma slope was not statistically different from purified vtg of mahseer standard ( $F_{0.05}=1.678_{(12,28)}$ ,  $p>0.05$ ). ELISA assay developed for *T. tambroides* vitellogenin was confirmed through inter- and intra-assay validation. At different binding percentages (20, 50 and 80%), the coefficient of variation (CV%) of both precision assays (inter- and intra-assay) were less than 15% which means ELISA developed for the measurement of plasma vtg concentration in *T. tambroides* is sensitive and repeatable.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**PENULENAN DAN PENCIRIAN VITELLOGENIN KELAH MERAH (TOR  
TAMBROIDES BLEEKERS)**

Oleh

**NURUL ASHIKIN BINTI MUHAMMAD**

**Februari 2013**

**Pengerusi: Annie Christianus, PhD**

**Fakulti: Pertanian**

Vitellogenin (vtg) ialah sejenis glikofosfolipoprotein dengan berat jisim molekular yang tinggi, dihasilkan di dalam hepar di bawah rangsangan hormon estrogen. Kebiasaannya ditemui di dalam haiwan betina yang matang, vtg diserap oleh telur yang sedang membesar semasa proses kematangan. Ia berfungsi sebagai penyimpan nutrien bagi tumbesaran embrio. Berpotensi sebagai penanda aras bagi kematangan telur, vtg membantu memperbaiki sistem penghasilan anak ikan. Kelah merah adalah salah satu ikan yang paling dicari di Malaysia, yang digunakan sebagai ikan permainan dan penting dari segi ekonomi sebagai spesies biakan. Masalah utama yang timbul dalam penghasilan benih ikan kelah merah yang banyak di tempat penetasan ialah pada ketersediaan dan pemilihan induk yang matang. Hanya pekerja yang mahir dan pakar sahaja yang berupaya mengenalpasti ikan betina yang matang dan bersedia untuk bertelur daripada aspek morfologi. Oleh itu, suatu kaedah yang



mudah untuk mengenalpasti ikan betina yang sesuai dijadikan sebagai induk adalah diperlukan. Maka, dalam kajian ini, ELISA telah dihasilkan untuk mengukur vtg dalam plasma ikan kelah merah sebagai penanda aras kematangan ikan betina. Penghasilan penanda aras ini pasti akan menyumbang kepada penghasilan pusat penetasan benih ikan kelah merah.

Estrogen ( $17\beta$ -estradiol) telah disuntik secara intra-peritoneal ke dalam lima ekor ikan jantan ( $1.5\pm 0.4$  kg). Untuk memastikan vtg telah terhasil, plasma yang belum melalui proses pengasingan, diambil dari ikan jantan yang telah disuntik, ikan betina yang matang dan ikan yang tidak disuntik telah diuji dan disaring untuk memerhati kehadiran jalur protein dengan jisim molekul yang tinggi menggunakan SDS-PAGE. Kemunculan jalur protein dengan jisim molekul 149 kDa dalam plasma ikan jantan yang telah disuntik dan ikan betina yang matang menandakan vtg telah terhasil dengan kehadiran hormon estrogen. Sampel plasma telah diasingkan menggunakan kromatografi pengasingan berdasarkan saiz molekul. Hasil sampel plasma pengasingan melalui kromatografi (protein bersaiz besar) yang mengandungi puncak simetri berbentuk loceng yang boleh dilihat pada graf yang tercetak bersama data pengasingan melalui kromatografi, telah dikumpulkan untuk analisa seterusnya. Melalui 'native PAGE', saiz protein yang terkumpul dianggarkan mempunyai saiz sebesar 700 kDa.

Pengurangan lanjut saiz protein kepada beberapa pecahan jalur protein oleh SDS-PAGE menunjukkan kehadiran protein jalur dengan kedudukan yang sama dalam sampel ikan jantan yang disuntik dengan estrogen dan ikan betina yang matang,

manakala sampel ikan jantan terkawal (yang tidak disuntik) menunjukkan tiada kesamaan pada kedudukan jalur protein tersebut. Di dalam analisis 'Western blot', plasma ikan jantan yang disuntik dengan estrogen menunjukkan tiga jalur polipeptida (133, 117 dan 56 kDa) dikesan oleh anti-kap antibody monoklon dan dikenalpasti sebagai vtg. Jalur-jalur pada kedudukan yang sama juga dikesan di dalam sampel ikan betina yang matang. Sampel ikan jantan yang tidak disuntik tidak menunjukkan sebarang tindakbalas terhadap antibodi.

ELISA telah pun berjaya dicipta dengan pengkhususan pada spesies ikan kelah merah, untuk pengiraan kepekatan vtg dalam plasma. Sampel vtg yang telah diasingkan (253 ng/ml) dilapis pada permukaan '96-well microplate'. Sampel plasma telah dicairkan dengan antiserum dengan nisbah 1:500 sehingga 1:1000. Pelinearan daripada lengkung anjakan mengikat oleh transformasi logit menunjukkan pencairan bersiri sampel ikan betina kelah merah mempunyai kecerunan yang secara statistiknya sama dengan sampel plasma vtg ikan kelah merah ( $F_{0.05}=1.678_{(12,28)}$ ,  $p>0.05$ ). ELISA yang telah dicipta spesifik untuk *T. tambroides* terbukti boleh digunakan semula, mudah dianalisa dan sensitif sepertimana dibuktikan oleh hasil yang diperoleh melalui satu kaedah bagi mengukur ketepatan esei. Pada peratusan pengikatan yang berlainan (rendah, sederhana, tinggi), pekali variasi (CV %) bagi kedua-dua inter- dan intra-assay yang telah ditentukan adalah kurang daripada 15%. Ini bermakna ELISA yang telah berjaya dihasilkan melalui eksperimen ini yang bertujuan untuk mengukur kepekatan vtg dalam darah ikan kelah merah, adalah merupakan esei yang sangat berguna dan boleh diguna pakai pada masa hadapan.

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I certify that an Examination Committee met on 19<sup>th</sup> February 2013 to conduct the final examination of Nurul Ashikin binti Muhammad on her Master of Science thesis entitled “Purification and Characterization of Malaysian mahseer *Tor tambroides* (Bleeker, 1854) Vitellogenin” in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulation 1981. The Committee recommends that the candidate be awarded the relevant degree.

Member of the Examination Committee are follows:

**Aziz Arshad, PhD**

Professor,  
Faculty of Agriculture,  
Universiti Putra Malaysia,  
Malaysia  
(Chairman)

**Hassan Mohd Daud, PhD**

Associate Professor,  
Faculty of Veterinary Medicine,  
Universiti Putra Malaysia,  
Malaysia  
(Internal Examiner)

**Abdul Razak Alimon, PhD**

Professor,  
Faculty of Agriculture,  
Universiti Putra Malaysia,  
Malaysia  
(Internal Examiner)

**Mazlan Abd Ghaffar, PhD**

Associate Professor,  
Faculty of Science and Technology,  
Universiti Kebangsaan Malaysia,  
Malaysia  
(External Examiner)

---

**NORITAH OMAR, PhD**

Associate Professor and Deputy Dean  
School of Graduate Studies  
Universiti Putra Malaysia

Date: 23 May 2013

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirements for the degree of Master of Science. The members of the Supervisory Committee were as follow:

**Annie Christianus, PhD**

Senior Lecturer  
Faculty of Agriculture,  
Universiti Putra Malaysia  
(Chairman)

**Siti Khalijah Daud, PhD**

Associate Professor  
Faculty of Science,  
Universiti Putra Malaysia  
(Member)

**Che Roos Saad, PhD**

Associate Professor  
Faculty of Agriculture,  
Universiti Putra Malaysia  
(Member)

**Sharr Azni Harmin, PhD**

Professor  
Faculty of Science and Biotechnology,  
Universiti Industri Selangor  
(External Member)

-----  
**BUJANG BIN KIM HUAT, PhD**

Professor and Dean  
School of Graduate Studies  
Universiti Putra Malaysia

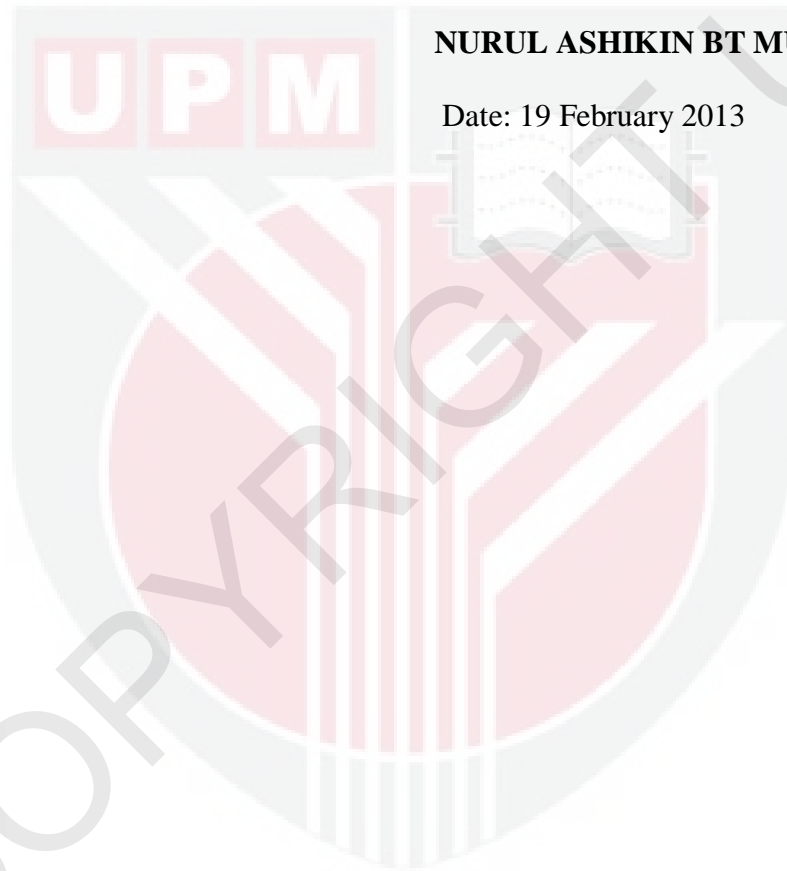
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## DECLARATION

I declare that the thesis is my original work for quotation and citation which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any degree at Universiti Putra Malaysia or other institutions.

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**NURUL ASHIKIN BT MUHAMMAD**

Date: 19 February 2013



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## LIST OF ABBREVIATIONS

ANCOVA	-	Analysis of covariance
APS	-	Ammonium persulfate
CV	-	Coefficient of variation
E2	-	17 $\beta$ -estradiol
EDTA	-	Ethylenediaminetetraacetic acid
ELISA	-	Enzyme-linked immunosorbent assay
MS222	-	Tricaine methanesulfonate
NSB	-	Non-specific binding
OD	-	Optical density
PBST	-	Phosphate buffered saline-Tween 20
PVDF	-	Polyvinylidene fluoride
SDS-PAGE	-	Sodium dodecyl sulphate – polyacrylamide gel electrophoresis
TEMED	-	Tetramethylethylenediamine
TMB	-	Tetramethyl benzidine
vtg	-	vitellogenin

# CHAPTER 1

## INTRODUCTION

Vitellogenin (vtg), an egg yolk precursor, is a high molecular weight glycopospholipoprotein found in sexually mature female vertebrates. It is synthesized in the liver under estrogen stimulation. Plasma vtg level is related to the maturation stage of eggs in female. Vitellogenesis relates to the incorporation of vtg proteins and other molecules such as vitamins and lipids into growing oocyte. At the end of this process, the oocyte containing all molecules important for developing embryo becomes ready for fertilization. Therefore, information acquired on the characteristic and concentration of plasma vtg in female fish is can be used as an indicator for oocytes maturity.

### 1.1 Statement of Problem

*Tor tambroides* or locally known as kelah merah, can be found in pristine rivers.

Their numbers are declining due to overfishing and deforestation (Sungan *et al.*, 2006). Species conservation is crucially critical to avoid extinction of the species.

Documented report suggested that this fish is able to reproduce throughout the year (Ismail *et al.*, 2011). High demand for mahseer fry has urged researchers, farmers, and academicians to come up with reliable technique to enhance breeding and

spawning success. Identification of matured females through physical observation is inadequate to ascertain the maturity stage of the fish. Therefore, a study on vtg characteristics and concentration in mahseer were proposed as indicators for oocytes maturity.

## 1.2 Significance of Study

A number of studies on vitellogenin (vtg) focused on environmental concerns and reproductive physiology of fish. Vtg has been identified in fishes such as Chilean flounder (Leonardi *et al.*, 2009), perch (Hennies *et al.*, 2003), carp (Fukada *et al.*, 2003), greenback flounder, rainbow trout and Atlantic salmon (Watts *et al.*, 2003). Various methods have been developed to purify vertebrates' vtg. Magalhães *et al.* (2004) developed one-step and non-denaturing purification method for *Cyprinus carpio* vitellogenin, while Wunschel *et al.* (2005) described the identification of vtg in fish using High Performance Liquid Chromatography (HPLC) separation coupled to matrix-assisted laser desorption/ionization mass spectrometry (MALDI-MS). During the process, proteins are separated either by size or charge of the molecule. Several methods applied are time-consuming and costly. Numerous purification steps are not preferred due to the probability of loss in protein purity. In this study, a simple flow method for vtg purification was established and described.

Methods for the quantification of vtg in blood plasma of fish species developed include radioimmunoassay (Tyler *et al.*, 1996), immunodiffusion, real time PCR

(Celius *et al.*, 2000) and enzyme-linked immuno sorbent assay (ELISA). ELISA is the most favored quantification assay due to the ease for routine application. Anti-vtg antibodies are species-specific. It requires antibody production in animals such as rabbits and goats. However, the use of commercial antiserum is possible as long as it cross reacts against the target proteins, sensitive and reproducible through ELISA assay.

This study was conducted with the purpose to characterize vtg in blood plasma of mahseer and to develop ELISA for measuring vtg concentration in plasma level. The lack of information available with regards to vtg in *T. tambroides* has led to this study.

### **1.3 OBJECTIVES**

Therefore, the objectives of this study were:-

1. To determine the molecular weight of purified estrogen-induced vtg in mahseer by SDS-PAGE and Western blot
2. To characterize the specificity of commercial antibody against mahseer purified vtg by Western blot
3. To develop an enzyme-linked immunosorbent assay (ELISA) for the quantification of plasma vtg in mahseer

## REFERENCES

- Alwine, J.C., Kemp, D.J., Stark, G.R. (1977). Method for detection of specific RNAs in agarose gels by transfer to diazobenzyloxymethyl-paper and by hybridization with DNA probes. *Proceedings of the National Academy of Sciences of the United States of America* 74: 5350-5354.
- Amano, H., Fujita, T., Hiramatsu, N., Todo, T., Hara, A. (2009). Purification and classification of three lipovitellin subtypes in the marbled sole (*Pleuronectes yokohamae*). *Zoological Science* 26: 510-516.
- Amano, H., Kitamura, M., Fujita, T., Hiramatsu, N., Todo, T., Suyama, S., Hara, A. (2008). Purification and characterization of lipovitellin from Pacific saury *Cololabis saira*. *Fisheries Science* 74: 830-836.
- Amdam, G.V., Norberg, K., Hagen, A., and Omholt, S.W. (2003). Social exploitation of vitellogenin. *Proceedings of the National Academy of Sciences of the United States of America* 100(4): 1799-1802.
- Arukwe, A. and Goksøyr, A. (2003). Eggshell and egg yolk proteins in fish: hepatic proteins for the next generation: oogenetic, population, and evolutionary implications of endocrine disruption. *Comparative Hepatology* 2: 4.
- Azuadi, N.M., Siraj, S.S., Daud, S.K., Christianus, A., Harmin, S.A., Sunagn, S., Britin, R. (2011). Enhancing ovulation of Malaysian mahseer (*Tor tambroides*) in captivity by removal of dopaminergic inhibition. *Journal of Fisheries and Aquatic Science* 6 (7): 740-750.
- Baert, J-L., Britel, M., Slomianny, M-C., Delbart, C., Fournet, B., Sautiere, P., Malecha, J. (1991). Yolk protein in leech. *European Journal of Biochemistry*. 201: 191-198.
- Braathen, M., Mdegela, R.H., Correia, D., Rundberget, T., Myburgh, J., Botha, C., Skaare, J.U. and Sandvik, M. (2009). Vitellogenin in African sharptooth catfish (*Clarias gariepinus*): purification, characterization, and ELISA development. *Journal of Toxicology and Environmental Health Part A* 72: 173-183.
- Bortone, S.A. and Davis, W.P. (1994). Fish intersexuality as indicator of environmental stress. *Bioscience* 44(3): 165-172.
- Burnette, W.N. (1981). "Western blotting": electrophoretic transfer of proteins from sodium dodecyl sulphate-polyacrylamide gels to unmodified nitrocellulose and radiographic detection with antibody and radioiodinated protein A. *Analytical Biochemistry*. 112: 195-203.
- CAMP., (1998). Report of the workshop "Conservation, Assessment and Management Plan for Freshwater fishes of India 1997" organized by Zoo

Outreach Organization (ZOO) and National Bureau of Fish Genetic Resources, Lucknow (pp156). September 1997.

- Carnevali, O., Mosconi, G., Angelini, F., Limatola, E., Ciarcia, G. and Polzonetti-Magni, A. (1991). Plasma vitellogenin and 17 $\beta$ -estradiol levels during the annual reproductive cycle of *Podarcis s. sicula* Raf. *General and Comparative Endocrinology* 84: 337-343.
- Celius, T., Matthews, J.B., Giesy, J.P. and Zacharewski, T.R. (2000). Quantification of rainbow trout (*Onchorynchus mykiss*) zona radiata and vitellogenin mRNA levels using real-time PCR after in vivo treatment with estradiol-17 beta or alpha-zearalenol. *Journal of Steroid Biochemistry and Molecular Biology* 75: 109-119.
- Colborn, T., vom Saal, F.S. and Soto, A.M. (1993). Developmental effects of endocrine-disrupting chemicals in wildlife and humans. *Environmental Health Perspectives* 101: 378-384.
- Covens, M., Stynen, D., Ollevier F. and Loof, D. (1988). Concanavalin a reactivity of vitellogenin and yolk proteins of the threespined stickleback *Gasterosteus aculeatus* (Teleostei). *Comparative Biochemistry and Physiology B* 90: 227-233.
- Cutting, J.A. and Roth, T.F. (1973). Staining of phospho-proteins on acrylamide gel electropherograms. *Analytical Biochemistry* 54: 386-394.
- Ernst, O. and Zor, T. (2010). Linearization of the Bradford protein assay. *Journal of Visualized Experiments*. DOI: 10.3791/1918
- Fukada, H., Fujiwara, Y., Takahashi, T., Hiramatsu, N., Sullivan C.V. and Hara, A. (2003). Carp (*Cyprinus carpio*) vitellogenin: purification and development of a simultaneous chemiluminescent immunoassay, *Comparative Biochemistry and Physiology Part A, Molecular and Integrative Physiology* 134: 615–623.
- Garfin, D. (1990) One-dimensional gel electrophoresis. *Methods in Enzymology* 182: 425-441.
- George, A., Morgan, T.J., Alvarez, P., Millan, M., Herod, A.A., Kandiyoti, R. (2010). Fractionation of a coal tar pitch by ultra-filtration, and characterization by size exclusion chromatography, UV-fluorescence and laser desorption-mass spectroscopy. *Fuel* 89: 2953-2970.
- Hennies, M., Wiesmann, M., Allner, B., Sauerwein, H. (2003). Vitellogenin in carp (*Cyprinus carpio*) and perch (*Perca fluviatilis*): purification, characterization and development of an ELISA for the detection of estrogenic effects. *The Science of the Total Environment* 309: 93-103.
- Hong, L., Fujita, T., Wada, T., Amano, H., Hiramatsu, N., Zhang, X., Todo, T., Hara, A. (2009). Choriogenin and vitellogenin in red lip mullet (*Chelon haematocheilus*): purification, characterization, and evaluation as potential



biomarkers for detecting estrogenic activity. *Comparative Biochemistry and Physiology Part C* 149: 9-17.

Ingram, B., Sungan, S., Tinggi, D., Sim, S.Y. and De Silva, S.S. (2007). Breeding performance of Malaysian mahseer, *Tor tambroides* and *Tor douronensis* broodfish in captivity. *Aquaculture Research* 38: 809-818.

Ismail, M.F.S., Siraj, S.S., Daud S.K. and Harmin, S.A. (2011). Association of annual hormonal profile with gonad maturity of mahseer (*Tor tambroides*) in captivity. *General and Comparative Endocrinology* 170: 125-130.

Johnsen, H.K., Tveiten, H., Willassen N.P. and Arnesen, A.M. (1999). Arctic charr (*Salvelinus alpinus*) vitellogenin: development and validation of an enzyme-linked immunosorbent assay. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology* 124: 355-362.

Kang, B.J., Jung, J.-H., Lee, J.M., Lim, S.-G., Saito, H., Kim, M.H., Kim, Y.-J., Saigusa, M. and Han, C.-H. (2007). Structural and expression analyses of two vitellogenin genes in the carp, *Cyprinus carpio*. *Comparative Biochemistry and Physiology Part B* 148: 445-453.

Kwon, H.-C., Hara, A., Mugiya, Y., Yamada, J. (1990). Enzyme Linked-Immunosorbent Assay (ELISA) of vitellogenin in Whitespotted Charr, *Salvelinus leucomaenis* *Bulletin of the Faculty of Fisheries Hokkaido University*, 41(4): 162-180.

Laemmli, U.K. (1970). Cleavage of structural proteins during the assembly of the head of bacteriophage T4. *Nature* 227: 680-685.

Leonardi, M., Vera, J., Tarifeño, E., Puchi M. and Morín, V. (2009). Vitellogenin of the Chilean flounder *Paralichthys adspersus* as a biomarker of endocrine disruption along the marine coast of the South Pacific. Part I: induction, purification, and identification. *Fish Physiology and Biochemistry* 36: 757-765.

Le Menn, F., Cerdà, J., Babin, P.J. (2007). Ultrastructural aspects of the ontogeny and differentiation of ray-finned fish ovarian follicles. In: Babin, P.J. (Ed.), *The Fish Oocyte: From Basic Studies to Biotechnological Application* (pp. 1-37). The Netherlands: Springer.

Liang, F.T., Granstrom, D.E. and Shi, Y.F. (1997). Concentrating protein samples for sodium dodecyl sulphate-polyacrylamide gel electrophoresis and isoelectric focusing using protein-blotting membranes. *Journal of Chromatography A* 764: 143-150.

Lomax, D.P., Roubal, W.T., Moore J.D. and Johnson, L.L. (1998). An enzyme-linked immunosorbent assay (ELISA) for measuring vitellogenin in English sole (*Pleuronectes vetulus*): development, validation and cross-reactivity with other pleuronectids. *Comparative Biochemistry and Physiology B* 121: 425-436.

- Lubzens, E., Young, G., Bobe, J., Cerdà, J. (2009). Oogenesis in teleosts: how fish eggs are formed. *General and Comparative Endocrinology* 165: 367-389.
- Luo, W., Zhou, Q., Jiang, G. (2011). Development of enzyme-linked immunosorbent assays for plasma vitellogenin in Chinese rare minnow (*Gobiocypris rarus*). *Chemosphere* 84: 681-688.
- Mac Donald, A. St. J. (1948). Circumventing the mahseer: and other sporting fish of India and Burma (306p). *Bombay Natural History Society*, Bombay.
- Magalhães, I., Ledrich, M-L., Pihan J-C. and Falla, J. (2004). One-step, non-denaturing purification method of carp (*Cyprinus carpio*) vitellogenin. *Journal of Chromatography B* 799: 87-93.
- Maltais, D., Roy, R.L. (2009). Purification and partial characterization of vitellogenin from shorthead redhorse (*Moxostoma macrolepidotum*) and copper redhorse (*Moxostoma hubbsi*) and detection in plasma and mucus with a heterologous antibody. *Fish Physiology and Biochemistry* 35: 241-254.
- Maltais, D. and Roy, R.L. (2007). A lateral flow immunoassay for rapid evaluation of vitellogenin levels in plasma and surface mucus of the copper redhorse (*Moxostoma hubbsi*). *Environmental Toxicology and Chemistry* 26: 1672-1676.
- Matsubara, T., Wada, T., Hara, A. (1994). Purification and establishment of ELISA for vitellogenin of Japanese sardine (*Sardinops melanostictus*). *Comparative Biochemistry and Physiology Part B: Comparative Biochemistry* 109: 545-555.
- Mohsin, A. K. M. and Ambak, M. A. (1983). *Freshwater Fishes of Peninsular Malaysia*. Selangor, Malaysia: Universiti Pertanian Malaysia Publication.
- Munkittrick, K.R., Portt, C.B., Van der Kraak, G.J., Smith I.R. and Rokosh, D.A. (1991). Impact of bleached kraft mill effluent on population characteristics, liver MFO activity, and serum steroid levels of a Lake Superior white sucker (*Catostomus commersoni*) population. *Canadian Journal of Fisheries and Aquatic Sciences* 48: 1371-1380.
- Nath, P., 1999. Some aspects of teleost vitellogenesis. In: Saksena, I. (Ed.), *Ichthyology: Recent Research Advances*. Science Publishers Inc., Enfield, New Hampshire.
- Ngamniyom, A. and Panyarachun, B. (2011). Expression levels of hormone receptor and vitellogenin mRNAs in livers of Thai medaka, *Oryzias minutillus*, inhabiting the suburbs of Bangkok, Thailand. *Journal of Fisheries and Aquatic Science* 6: 438-446.

- Nilsen, B.M., Berg, K., Eidem, J.K., Kristiansen, S-I., Brion, F., Porcher, J-M., Goksøyr, A. (2004). Development of quantitative vitellogenin-ELISAs for fish test species used in endocrine disruptor screening. *Analytical and Bioanalytical Chemistry* 378: 621-633.
- O'Brien, E.D., Salicioni, A.M., Cabada, M.O. and Arranz, S.E. (2010). Vitellogenesis in *Bufo arenarum*: Identification, characterization and immunolocalization of high molecular mass lipovitellin during oogenesis. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology* 155: 256-265.
- Ogale, S.N. (2002). Mahseer ranching (pp. 225-229). In: *Riverine and Reservoir Fisheries of India* (Boopendranath, M.R., Meenakumari, B., Joseph, J., Sankar, T.V., Pravin, P. and Edwin, L. Eds.). 458 p.
- Palumbo, A.J., Koivunen, M., Tjeerdema, R.S. (2009). Optimization and validation of a California halibut environmental estrogen bioassay using heterologous ELISA. *Science of the Total Environment* 407: 953-961.
- Pan, M.L., Bell, W.J., Telfer, W.H. (1969). Vitellogenic blood protein synthesis by insect fat body. *Science* 165: 393-394.
- Parks, L.G., Cheek, A.O., Denslow, N.D., Heppel, S.A., Mclachlan J.A. and LeBlanc, G.A. (1999). Fathead minnow (*Pimephales promelas*) vitellogenin: purification, characterization and quantitative immunoassay for the detection of estrogenic compounds. *Comparative Biochemistry and Physiology Part C* 123: 113-125.
- Prakash, O., Goswami, S.V., Sehgal, N. (2007). Establishment of ELISA for murrel vitellogenin and choriogenin, as biomarkers of potential endocrine disruption. *Comparative Biochemistry and Physiology Part C, Toxicology and Pharmacology* 146(4): 540-551.
- Prat, J.P., Lamy J.N. and Weill, J.D. (1969). Staining of lipoproteins after electrophoresis in polyacrylamide gel. *Bulletin de la Société de Chimie Biologique* 51(9): 1367.
- Rainwater, T.R., Selcer, K.W., Nespoli, L.M., Finger, A.G., Ray, D.A., Platt, S.G., Smith, P.N., Densmore, L.D., Anderson T.A. and McMurry, S.T. (2008). Plasma vitellogenin in Morelet's crocodiles from contaminated habitats in northern Belize. *Environmental Pollution* 153: 101-109.
- Rodbard, D. and Lewald, J.E. (1970). Computer analysis of radioligand assay and radioimmunoassay data. *Acta Endocrinologica Supplementum* 147: 79-103.
- Ross, L.G. and Ross, B.R. (1999). *Anaesthetic and Sedative Techniques for Aquatic Animals*. 2<sup>nd</sup> Ed. Malden, MA: Blackwell Science, 1999.
- Roy, R.L., Morin, Y., Courtenay, S.C., Robichaud, P. (2004). Purification of vitellogenin from smooth flounder (*Pleuronectes putnami*) and measurement

in plasma by homologous ELISA. *Comparative Biochemistry and Physiology Part B* 139: 235-244.

- Saka, M., Tada, N. and Kamata, Y. (2008). Cross-reactivity of a polyclonal antibody against *Chinemys reevesii* vitellogenin with the vitellogenins of other turtle species: *Chelydra serpentina*, *Macrochelys temminckii* and *Pelodiscus sinensis*. *Zoological Science* 25: 907-911.
- Shao, J., Shi, G., Song, M., Jiang, G. (2005). Development and validation of an enzyme-linked immunosorbent assay for vitellogenin in Chinese loach (*Misgurnus anguillicaudatus*). *Environment International* 31(5): 763-770.
- Shapiro, A. L., Vinuela, E. and Maizee, J. V. (1967). Molecular weight estimation of polypeptide chains by electrophoresis on SDS-polyacrylamide gels. *Biochemical and Biophysical Research Communications* 28: 815-820.
- Sherry, J., Gamble, A., Fielden, M., Hodson, P., Burnison, B., Solomon, K. (1999). An ELISA for brown trout (*Salmo trutta*) vitellogenin and its use in bioassays for environmental estrogens. *The Science of the Total Environment* 225: 13-31.
- Shreck, C.B., Contreras-Sanchez, W., Fitzpatrick, M.S. (2001). Effects of stress on fish reproduction, gamete quality, and progeny. *Aquaculture* 197: 3-24.
- Siraj, S.S., Daud, S.K., Keong, R.B.P. and Ng, C.K. (2007). Characterisation of the Malaysian mahseer (kelah), *Tor tambroides*. Mahseer: The biology, culture and conservation. *Proceedings of the International Symposium on the Mahseer* (pp. 179-201). Malaysian Fisheries Society Occasional Publication No. 14, Kuala Lumpur 2007, 236p.
- Southern, E.M. (1975). Detection of specific sequences among DNA fragments separated by gel electrophoresis. *Journal of Molecular Biology* 98: 503-517.
- Specker, J.L., Sullivan, C.V. (1994). Vitellogenesis in fishes: status and perspectives. In: Davey, K.G., Peter, R.E., Tobe, S.S. (Eds.), *Perspectives in Comparative Endocrinology* (pp. 304-315). Ottawa: National Research Council Canada.
- Stellwagen, E. (1990). Gel filtration. *Methods in Enzymology* 182: 317-328.
- Sun, B., Pankhurst, N.W., Watts, M. (2003). Development of an enzyme-linked immunosorbent assay (ELISA) for vitellogenin measurement in greenback flounder *Rhombosolea tapirina*. *Fish Physiology and Biochemistry* 29: 13-21.
- Sungan, S., Tinggi, D., Salam N. and Sadi, C. (2006). Aspects of the biology and ecology of empurau (*Tor tambroides*) and semah (*T. duoronensis*) in Sarawak, Malaysia. Paper presented at the International Symposium on the Mahseer, Kuala Lumpur, Malaysia. March 2006.

- Swart, J.C., Pool, E.J. (2009). The development and validation of a quantitative ELISA for in vivo and in vitro synthesized vitellogenin from Mossambicus tilapia (*Oreochromis mossambicus*). *Journal of Immunoassay and Immunochemistry* 30: 208-223.
- Taki, T., Handa, S., Ishikawa, D. (1993). Blotting of glycolipids and phospholipids from a high-perforamnce thin-layer chromatogram to a polyvinylidene difluoride membrane. *Analytical Biochemistry* 221: 312-316.
- Tan E.S.P. (1980). Some aspects of the biology of Malaysian riverine cyprinids. *Aquaculture* 20: 218-289.
- Towbin, H., Staehelin, T., Gordon, J. (1979). Electrophoretic transfer of proteins from polyacrylamide gels to nitrocellulose sheets: procedure and some applications. *Proceedings of the National Academy of Sciences of the United States of America* 76: 4350-4354.
- Trathnigg B. (2000). Size-exclusion chromatography of polymers. In: Meyers R, ed. *Encyclopedia of Analytical Chemistry*. Chichester, UK: Wiley & Sons. p 8008–8034.
- Tyler, C.R., Santos, E.M., Prat, F. (2000). Unscrambling the egg - cellular biochemical molecular and endocrine advances in oogenesis. In: Norberg, B., Kjesbu, O.S., Taranger, G.L., Andersson, E., Stefansson, S.O. (Eds.), *Proceedings of the 6th International Symposium of Reproductive Physiology of Fish* (pp273-280). Bergen, Norway. July 1999.
- Tyler, C.R., van der Eerden, B., Jobling, S., Panter G. and Sumpter, J.P. (1996). Measurement of vitellogenin, a biomarker for exposure to oestrogenic chemicals, in a wide variety of cyprinid fish. *Journal of Comparative Physiology B* 166: 418-426.
- Tyler, C.R. and Sumpter, J.P. (1990). The purification and partial characterization of carp, *Cyprinus carpio* vitellogenin. *Fish Physiology and Biochemistry* 8(2): 111-120.
- Wagner, U., Hädige, D., Gudmundsdóttir, B.K, Nold, K., Drössler, K. (2001). Antibody response in salmonids against the 70 kDa serine protease of *Aeromonas salmonicida* studied by a monoclonal antibody-based ELISA. *Veterinary Immunology and Immunopathology* 82: 121-135.
- Wang, H., Tan, J.T.T., Emelyanov, A., Korzh, V., Gong, Z. (2005). Hepatic and extrahepatic expression of vitellogenin genes in the zebrafish, *Danio rerio*. *Gene* 356: 91-100.
- Watts, M., Pankhurst, N.W., Pryce A. and Sun, B. (2003). Vitellogenin isolation, purification and antigenic cross-reactivity in three teleost species. *Comparative Biochemistry and Physiology Part B* 134: 467-476.

Williams, R.J., Keller, V.D., Johnson, A.C., Young A.R. and Holmes, M.G. (2009). A national risk assessment for intersex in fish arising from steroid estrogens. *Environmental Toxicology and Chemistry* 28: 220-230.

Wunschel, D., Schultz, I., Skillman A. and Wahl, K. (2005). Method for detection and quantitation of fathead minnow vitellogenin (vtg) by liquid chromatography and matrix-assisted laser desorption/ionization mass spectrometry. *Aquatic Toxicology* 73: 256-267.

Zohar, Y. and Mylonas, C. C. (2001). Endocrine manipulations of spawning in cultured fish: from hormones to genes. *Aquaculture* 197: 99-136.

