

# **UNIVERSITI PUTRA MALAYSIA**

GENETIC AND MOLECULAR ANALYSES FOR SEED AND OIL YIELDS IMPROVEMENT IN JATROPHA CURCAS L. POPULATIONS

ALIREZA BIABANIKHANKAHDANI

FP 2012 83

## GENETIC AND MOLECULAR ANALYSES FOR SEED AND OIL YIELDS IMPROVEMENT IN JATROPHA CURCAS L. POPULATIONS



BY

ALIREZA BIABANIKHANKAHDANI

Thesis submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirement for the Degree of Doctor of Philosophy

April 2012

# **DEDICATIONS**

To my beloved parents for

sacrifices, understanding and tremendous encouragement and support throughout my study

> To my sisters for their kindness and taking care

C

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in the Fulfilment of the requirement for the degree of Doctor of Philosophy

# GENETIC AND MOLECULAR ANALYSES FOR SEED AND OIL YIELDS IMPROVEMENT IN *JATROPHA CURCAS* L. POPULATIONS

By

### ALIREZA BIABANIKHANKAHDANI

April 2012

Chairman : Associate Professor Mohd Rafii Yusop, PhD

### Faculty : Agriculture

The current Jatropha improvement program was designed with the main objective to produce superior planting materials with high seed and oil yield production for commercial planting. The specific objectives were to study inter and intra-populations variation using morphological and Inter Simple Sequence Repeat (ISSR) molecular markers, to estimate genetic components, heritability values, general and specific combining abilities of several important characters for identifying the best parental lines and finally to quantify the level of heterosis for  $F_1$  hybrids in diallel crosses. Jatropha populations from Malaysia, Indonesia, India and Philippines were used as a base population in this selection program.

Sixty four plants in each of six Jatropha populations, namely Malaysia1 (My1), Malaysia2 (My2), Indonesia1 (In1), Indonesia2 (In2), India1 (Id1) and Philippines1 (Ph1) were planted for field evaluation in Serdang University Agriculture Park, Universiti Putra Malaysia. Jatropha populations were arranged in a randomized complete block design (RCBD) with four replications at spacing of 4 m × 2 m. Analysis of variance in seed characters revealed that there was significant variation among Jatropha populations for seed length, seed width, seed weight and 100-seed weight at P  $\leq 0.05$ . Population of My2 produced the highest oil content for both years (31.73 and 33.41%) of yield production. The lowest oil content was observed in population In1 for first and second years of harvesting (29.09 and 30.61%, respectively).

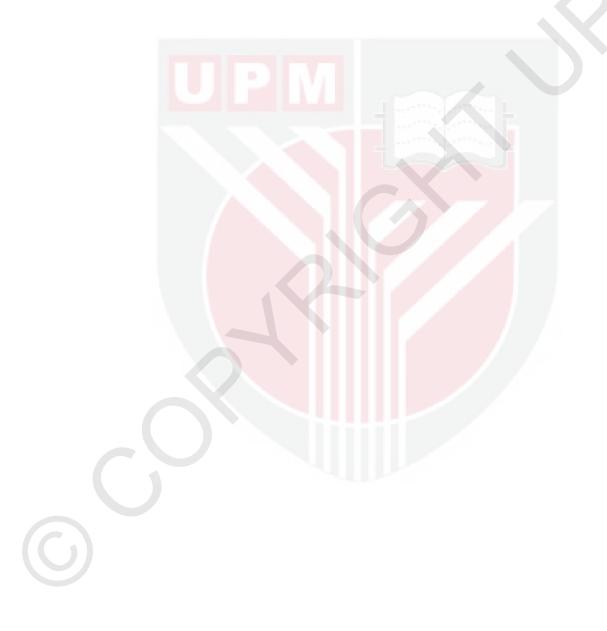
Significant variation ( $P \le 0.05$ ) was observed in plant height, number of secondary branches, days to flowering, number of inflorescences and seed yield. The highest seed yield was recorded in population In1 for first (46.5) and second year (222.8 g) in both years, whereas, the lowest was population In2 for first year and My1 for second year harvesting. Seed yield correlated significantly with days of flowering, number of inflorescences and plant height. Seed yield also correlated significantly with oil content. Days of flowering was found to have a high significantly positive relationship with plant height, number of secondary branches, number of tertiary branches, number of inflorescence, seed length and seed weight. The highest heritability was estimated for 100-seed weight (77.3%). The heritability values for plant height, number of secondary branch, days to flowering, seed weight, seed yield and oil content were 40.8, 47.8, 31.7, 77.2, 52.0 and 49.7% respectively.

ISSR markers were used to detect inter and intra-population variation of the based populations. The percentage of polymorphic bands for each population ranged from

46.15 to 60.84%, with an average of 55.10%. Among populations, the average number of alleles per locus ( $n_a$ ) ranged from 1.46 (My1) to 1.61 (Ph1) with the mean number of 1.55, which the effective number of alleles per locus ( $n_e$ ) ranged from 1.12 (My1) to 1.17 (My2) with the mean number of 1.15. Among the populations, population My1 had the lowest genetic diversity levels, while population Ph1 showed the highest genetic diversity. The partitioning of genetic diversity into within population and between populations based on Shannon's diversity index also revealed more variation within populations (0.81) than variations between populations (0.19).

From the field evaluation of the Jatropha populations of 364 plants, ten superior individual plants (three from Malaysia, three Indonesia, two India and two Philippine population) were selected and were intercrossed in a half-diallel mating design (10×10) including selfed of each selected parental plants. The 45 F<sub>1</sub> hybrids and 10 selfed progenies were evaluated in field using a randomized complete block design (RCBD) with three replications in Puchong University Agriculture Park, Universiti Putra Malaysia. Analyses of variance showed significant (P ≤ 0.01) variations among parents and hybrids for all characters. Analysis of variance for gene effects showed that both general (GCA) and specific combining ability (SCA) were influential for all traits. Analysis of specific combining ability in this study showed that a number of hybrids showed more precise for each character in different individual hybrids, but none showed the best SCA effect for all traits in one particular hybrid.

The percentages of heterosis for traits in some hybrids were considerably high indicating that a high degree of genetic diversity among parents. High broad sense heritability was recorded for all traits. However, the ratios of GCA/SCA indicated that non-additive effects were more important than additive gene effects. The analyses of GCA did not show that any single parent was a high general combiner for all traits simultaneously.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagi memenuhi keperluan untuk ijazah Doktor Falsafah

### ANALISA GENETIK DAN MOLEKULAR UNTUK KEMAJUAN HASIL BIJI DAN MINYAK DI DALAM POPULASI *JATROPHA CURCAS* L.

Oleh

### ALIREZA BIABANIKHANKAHDANI

### April 2012

### Pengerusi: Prof. Madya. Mohd Rafii Bin Yusop, PhD

#### Fakulti: Pertanian

Program memajukan Jatropha mase kimi telah direkabentuk dengan objektif utama adalah untuk menghasilkan bahan tanaman yang unggul, yang memberikan hasil biji dan minyak yang tinggi sebagai bahan tanaman komersial. Objektif khusus adalah untuk mengkaji variasi secara intra dan inter-populasi melalui morfologi dan penanda molekul inter ulangan jujukan ringkas (ISSR), untuk menganggar komponen variasi genetik, nilai keterwarisan, keupayaan bergabung am dan khusus bagi beberapa ciri penting bagi mengenalpasti waris induk yang terbaik dan akhirnya untuk menentukan paras heterosis bagi hibrid  $F_1$  di dalam kacukan dialel. Populasi Jatropha dari Malaysia, Indonesia, India dan Filipina telah digunakan sebagai populasi asas di dalam program pemilihan

ini.

Enam puluh empat pokok bagi setiap enam populasi Jatropha, iaitu Malaysia1 (My1), Malaysia2 (My2), Indonesia1 (In1), Indonesia2 (In2), India1 (Id1) dan Filipina1 (Ph1) telah ditanam untuk penilaian ladang di Taman Pertanian Universiti Serdang, Universiti vii Putra Malaysia. Populasi Jatropha tersebut telah disusun menggunakan rekabentuk blok penuh terawak (RCBD) dengan empat replikasi pada jarak tanaman 4 m × 2 m. Analisa varians bagi ciri biji menunjukkan terdapat variasi yang berbeza di kalangan populasi Jatropha yang dikaji untuk panjang biji, lebar biji, berat biji dan berat 100-biji pada P  $\leq 0.05$ . Populasi My2 mengeluarkan kandungan minyak yang tertinggi untuk kedua-dua tahun pengeluaran hasil (31.73 dan 33.41%). Kandungan minyak yang terendah adalah didapati di dalam populasi In1 untuk tahun pertama dan kedua penuaian (masing-masing 29.09 dan 30.61%).

Variasi bererti ( $P \le 0.05$ ) didapati bagi tinggi pokok, bilangan dahan sekunder, bilangan hari berbunga, bilangan jambak bunga dan hasil biji. Hasil biji tertinggi didapati dalam populasi In1 dalam kedua-dua tahun, manakala yang terendah adalah populasi In2 untuk tahun pertama dan My1 untuk tahun kedua penuaian. Hasil biji mempunyai korelasi bererti dengan bilangan hari berbunga, bilangan jambak bunga dan tinggi pokok. Hasil biji juga mempunyai korelasi bererti dengan kandungan minyak. Bilangan hari berbunga didapati mempunyai perhubungan positif bererti dengan tinggi pokok, bilangan dahan sekunder, bilangan dahan tertiari, bilangan jambak bunga, panjang biji dan berat biji. Nilai keterwarisan tertinggi didapati pada berat 100-biji (77.3%). Nilai keterwarisan untuk tinggi pokok, bilangan dahan sekunder, bilangan hari berbunga, berat biji, panjang biji, hasil biji dan kandungan minyak adalah masing-masing 40.8, 47.8, 31.7, 77.2, 52.0 dan 49.7%.

ISSR telah digunakan untuk menentukan variasi inter dan intra-populasi di dalam populasi asas tersebut. Peratusan jalur polimorfik untuk setiap populasi berjulat antara viii

46.15 hingga 60.84%, dengan purata 55.10%. Di kalangan populasi tersebut bilangan purata alel per lokus ( $n_a$ ) berjulat dari 1.46 (My1) hingga 1.61 (Ph1) dengan bilangan purata adalah 1.55, bilangan alel efektif per lokus ( $n_e$ ) berjulat dari 1.12 (My1) hingga 1.17 (My2) dengan bilangan purata adalah 1.15.

Di kalangan populasi tersebut, populasi My1 mempunyai paras kepelbagaian genetik yang terendah, manakala populasi Ph1 menunjukkan kepelbagaian genetik yang tertinggi. Pembahagian kepelbagaian genetik kepada di kalangan dalam populasi dan di antara populasi berdasarkan indeks kepelbagaian Shannon juga membuktikan terdapat lebih variasi di kalangan dalam populasi (0.81) berbanding variasi di antara populasi (0.19).

Dari penilaian di ladang populasi Jatropha tersebut yang terdiri dari 364 pokok, sepuluh individu pokok yang unggul (tiga dari populasi Malaysia, tiga dari Indonesia, dua dari India dan dua dari Filipina) telah dipilih dan dijalankan kacukan dikalangan mereka menggunakan rekabentuk pengawanan separa-dialel (10×10) termasuk penyendirian bagi setiap pokok induk terpilih. Empat puluh lima hybrid F<sub>1</sub> dan 10 progeni penyendirian telah dinilai di ladang mengunakan rekabentuk blok penuh terawak (RCBD) dengan tiga replikasi di Taman Pertanian Universiti Pucong, Universiti Putra Malaysia. Analisa varians menunjukkan terdapat variasi yang bererti ( $p \le 0.01$ ) dikalangan induk dan hibrid bagi semua ciri. Analisa varians bagi kesan gen menunjukkan keupayaan bergabung am (GCA) dan khusus (SCA) mempengaruhi kesemua ciri. Analisis keupayaan bergabung khusus dalam kajian ini menunjukkan beberapa hibrid menberikan kesan SCA yang bererti untuk setiap ciri dalam individu hibrid yang berlainan tetapi tiada satu hibridpun yang memberikan kesan SCA baik bagi semua ciri dalam sesuatu hibrid tertentu.

Peratusan heterosis untuk ciri dalam sebahagian hibrid adalah agak tinggi menunjukkan satu tahap kepelbagaian genetik yang tinggi dikalangan induk. Keterwarisan luas yang tinggi telah direkodkan untuk semua ciri. Walaubagaimanapun, nisbah GCA/SCA menunjukan pengaruh gen bukan-aditif adalah lebih penting berbanding pengaruh gen aditif. Analisa GCA mendapati tiada satu induk pun mempunyai berkeupayaan bergabung am untuk semua ciri secara serentak.

#### ACKNOWLEDGEMENT

My special and sincere appreciation goes to my supervisor of this thesis, Assoc. Prof. Dr. Mohd Rafii Bin Yusop. I am extremely appreciative of his supervision, kindness, constant support and endless patience. Without his well-designed plan and careful review of the draft, this thesis would have never been possible. His kind encouragement, guidance and instruction are invaluable things for me forever. I wish to express my gratitude to my thesis co-supervisors Prof. Dr. Ghizan Saleh and Prof. Dr. Zakaria Wahab for their guidance.

Many thanks are expressed to all teachers and staff of Crop Science Department, Universiti Putra Malaysia, for their help in many ways while I was studying in Malaysia. Special thanks go to Assist. Prof. Dr. Sasan Ghasemi for his encouragement. I would like to thank Mr. Mahmoodreza Shabanimofrad, Mr. Ali Ranjbarfard and Ms Mozhdeh Ebrahimpour and all other Iranian and Malaysian friends here for their help.

My greatest appreciation goes to my sister, Roya Biabani for her kindness and taking care of me during stay in Malaysia. Finally, I would like to express my special thanks to my parents and my sisters for their encouragement, support and understanding, without which the study would have not been successfully completed.

I certify that a Thesis Examination Committee has met on ..... to conduct the final examination of Alireza Biabanikhankahdani on his thesis entitled "Genetic and molecular analyses for seed and oil yields improvement in *Jatropha curcas* L. populations" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Pertanian Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Doctor of Philosophy.

Members of the Thesis Examination Committee were as follows:

### Mihdzar Abdul Kadir, PhD

Associate Professor Faculty of Agriculture Technology Universiti Putra Malaysia (Chairman)

#### Halimi Mohd Saud, PhD

Associate Professor Faculty of Agriculture Technology Universiti Putra Malaysia (Internal Examiner)

### Mohd Ridzwan A. Halim, PhD

Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Internal Examiner)

### Iftikhar Khalil, PhD

Professor Faculty of Agriculture University of Peshawar Pakistan (External Examiner)

#### ZULKARNAIN ZAINAL, PhD

Professor and Deputy Dean School Of Graduate Studies Universiti Putra Malaysia

Date:

xii

This thesis was submitted to the Senate of University Putra Malaysia and has been accepted as fulfillment of the requirement of the Doctor of Philosophy of Agricultural Science. The members of the Supervisory Committee were as follows:

### Mohd Rafii Bin Yusop, PhD Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Chairman)

**Ghizan Saleh, PhD** Professor Faculty of Agriculture Universiti Putra Malaysia (Member)

### Zakaria Wahab, PhD Professor

Faculty of Agriculture Universiti Putra Malaysia (Member)

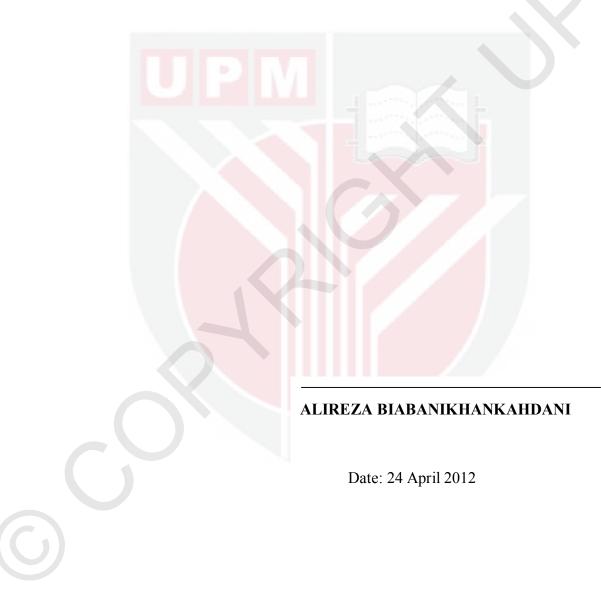
# BUJANG BIN KIM HUAT, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

# DECLARATION

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously, or is not, concurrently, submitted for any other degree at Universiti Putra Malaysia or other institutions.



# **TABLE OF CONTENTS**

		Page
D	EDICATIONS	ii
A	iii	
A	vii	
A	CKNOWLEDGEMENT	xi
	ECLARATION	xiv
L	IST OF TABLES	xviii
L	IST OF FIGURES	xxi
	IST OF ABBREVIATIONS HAPTER	xxii
1	GENERAL INTRODUCTION	
	1.1 Introduction	1
	1.2 Statement of problem	6
	1.3 Research Objectives	7
2	LITERATURE REVIEW	
	2.1 Origin, distribution and ecological requirement	8
	2.2 Jatropha naming and species in the world	9
	2.3 Taxonomic status	10
	2.4 Botanical description of Jatropha curcas	10
	2.5 Nature of pollination	13
	2.6 Varieties of Jatropha curcas	14
	2.7 Uses of Jatropha curcas	14
	2.8 Genetic improvement	15
	2.9 Heterosis	17
	2.1 Heritability	19
	2.11 Combining ability (GCA and SCA)	22
	2.12 Genetic diversity assessment methods	23
	2.12.1 Morphological characters variation	24
	2.12.2 Molecular marker based genetic diversity studies	27
	2.13 Inter simple sequence repeat (ISSR)	33
3	INTER AND INTRA POPULATION GENETIC VARIATION SIX JATROPHA POPULATIONS BASED ON QUANTITATI TRAITS	

3.1 Introduction

0

35

3.2	Materials and methods 3		
	3.2.1	Description of the experimental site	37
	3.2.2	Plant materials	37
	3.2.3	Field experimental design	38
	3.2.4	Data collection	40
	3.2.5	Statistical analysis	43
3.3	Resul	ts and discussion	47
	3.3.1	Nursery stage	47
	3.3.2	Field evaluation	53
	3.3.3	Correlation between characters	64
	3.3.4	Heritability and variance components	67
	3.3.5	Selection of superior plants	71
	3.3.6	Cluster analysis	73
	3.3.7	Principal Component Analysis (PCA)	76
3.4	Conc	lusions	78

# 4 INTER AND INTRA-POPULATION GENETIC VARIATION IN SIX JATROPHA CURCAS POPULATIONS REVEALED BY ISSR MARKERS

4.		Introduc	tion	82
		4.2 N	Materials and methods	84
		4.2.1	Study sites and sample collection	84
		4.2.2	DNA extraction techniques	84
		4.2.3	DNA quantification	85
		4.2.4	ISSR primers	86
		4.2.5	PCR amplification	86
		4.2.6	Agarose gel electrophoresis	87
		4.2.7	Comparison between molecular and morphological data	91
	4.3	Results a	and Discussion	91
		4.3.1	Primer Screening and reproducibility	91
		4.3.2	ISSR variation	93
		4.3.3	Genetic diversity within Jatropha populations	95
		4.3.4	Analysis of molecular variance (AMOVA)	98
		4.3.5	Cluster Analysis	99
			Principle component analysis (PCA)	102
		4.3.7	1 1 0	
			marker	105
	4.4	Conclusi	ions	105

# 5 COMBINING ABILITY AND HERITABILITY IN DIALLEL ANALYSIS OF SELECTED GENOTYPES

	5.1	Introduction	107
		5.2 Materials and methods	109
		5.2.1 Description of the experimental site	109
		5.2.2 Parents lines and cross scheme	109
		5.2.3 Field experimental design	111
		5.2.4 Data collection	111
		5.2.5 Statistical analysis	112
	5.3	Results and discussion	119
		5.3.1 Nursery stage	119
		5.3.2 Field evaluation	146
		5.4 Conclusion	181
6		MMARY, GENERAL CONCLUSION AND COMMENDATIONS	
R	EFEI	RENCES	190
A	PPEN	NDIX A	202
B	IODA	ATA OF STUDENT	203

# LIST OF TABLES

Table		Page
3.1	Jatropha populations selected for field evaluation	38
3.2	Outline of the ANOVA table used in analysis of quantitative traits for six Jatropha populations	43
3.3	Mean values and mean square for seed and growth characteristics of six different Jatropha populations in nursery stage	48
3.4	Phenotypic correlation coefficient (r) among the various traits of six Jatropha populations in nursery stage.	50
3.5	Estimation of genetic variables for growth traits in Jatropha in nursery stage	52
3.6	Mean values and mean squares for seed characteristics of six different Jatropha populations	54
3.7	Mean values and mean squares for growth characteristics of six different Jatropha populations	55
3.8	Mean values and mean squares for yield components characteristics of six different Jatropha populations	58
3.9	Mean values and mean squares for yield components and oil yield characteristics of six different Jatropha populations	61
3.10	Correlation coefficients among important quantitative morphological traits of <i>J. curcas</i>	65
3.11	Estimation of heritability and genetic variables for treats in six Jatropha populations	68
3.12	Internet cod and origin of ten superior plants	71

3.13	Jatropha population groupings revealed by cluster analysis	74
3.14	Cluster mean values for morphological characters in six Jatropha populations	75
4.1	List of ISSR primers selected for ISSR analysis in <i>J. curcas</i> populations	86
4.2	ISSR primers, their sequence, the number and size range of bands used among six Jatropha populations	92
4.3	Number of polymorphic bands per population in ten ISSR primers in Jatropha	94
4.4	Genetic diversity estimates of Jatropha populations	96
4.5	Nei's (1973) genetic diversity statistics for six Jatropha populations	97
4.6	Partitioning of the genetic variation into within and between populations based on Shannon's information index	97
4.7	Molecular variation between and within Jatropha populations	99
4.8	Jaccard's similarity coefficients matrix of six Jatropha populations	100
4.9	Component loading of the first three principal components for six Jatropha populations by ISSR markers	104
5.1	Expected mean square (EMS) and genetic interpretation of a half diallel	114
5.2	Mean squares from analysis of variance, general combining ability (GCA) and specific combining ability (SCA) for three characters of diallel cross involving 10 parents of Jatropha	120
5.3	Means of plant height, collar diameter and number of leaves of parents and their hybrids in Jatropha	121
5.4	Estimate of general combining ability effects (GCA) of ten Jatropha parents for measured traits in nursery stage	126
5.5	Estimate of specific combining ability effects (SCA) of 45 Jatropha hybrids for measured traits in nursery stage	127

5.6	Variance components of general and specific combining ability, narrow and broad sense heritability and ratio of GCA/SCA	130
5.7	Heterosis (MP) and heterobeltiosis (BP) for plant height in hybrids of Jatropha in nursery stage	132
5.8	Heterosis (MP) and heterobeltiosis (BP) for collar diameter in hybrids of Jatropha in nursery stage	137
5.9	Heterosis (MP) and heterobeltiosis (BP) for number of leaves in hybrids of Jatropha in nursery stage	142
5.10	Coefficients of phenotypic correlation between traits of Jatropha in diallel crosses involving ten parents in nursery stage	145
5.11	Mean squares from analysis of variance, general combining ability (GCA) and specific combining ability (SCA) for measured traits of diallel cross involving 10 parents of J. curcas in field evaluation	147
5.12	Means of measured traits of parents and their hybrids of Jatropha in field evaluation	148
5.13	Estimate of general combining ability effects (GCA) of ten Jatropha parents for measured traits in field evaluation	156
5.14	Estimate of specific combining ability effects (SCA) of 45 Jatropha hybrids for measured traits in field evaluation	160
5.15	Variance components of general and specific combining ability, narrow and broad sense heritability and ratio of GCA/SCA	167
5.16	Heterosis for measured traits in hybrids of Jatropha in field evaluation	169
5.17	Heterobeltiosis for all measured traits in hybrids of Jatropha in field evaluation	172
5.18	Coefficients of phenotypic correlation between traits of Jatropha in diallel crosses involving ten parents in field evaluation	179

# LIST OF FIGURES

Figure		Page
3.1	Jatropha experimental field at agriculture park, Universiti Putra Malaysia	40
3.2	Seed yield and oil content of ten superior plants form six Jatropha populations	72
3.3	The dendrogram of morphological traits derived from six Jatropha populations constructed using the UPGMA method. The scale is based on Jaccard's similarity coefficient	73
3.4	Morphological data three-dimensional PCA indicating relationships among six Jatropha populations	77
4.1	Amplified products of <i>Jatropha curcas</i> population (In1), using primer ISSR1, showing polymorphism for DNA banding pattern	88
4.2	Dendrogram of six Jatropha populations using Jacard similarity matrix based on ISSR data	101
4.3	Three-dimensional plot of the principle components from results of ISSR marker data among the six populations of <i>J. curcas</i> .	103
5.1	Jatropha in nursery stage	110

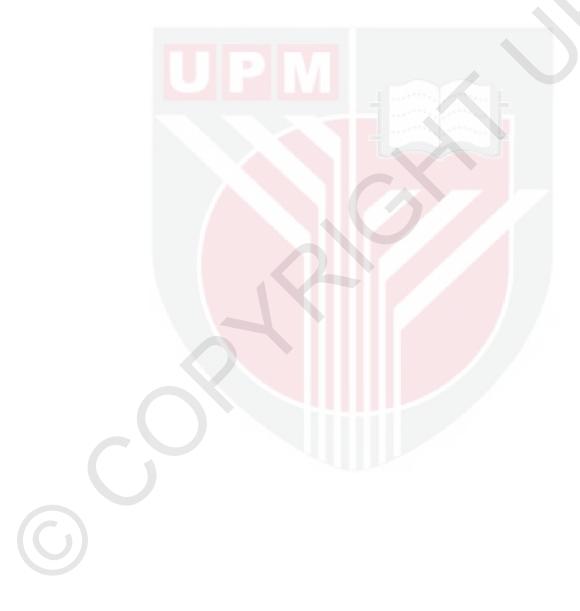
C

# LIST OF ABBREVIATIONS

	AFLP	Amplified fragment length polymorphism
	ANOVA	Analysis of variance
	bp	Base pairs
	$h^2_B$	Broad sense heritability
	CTAB	Cetyltrimethylammonium bromide
	G <sub>ST</sub>	Coefficient of gene differentiation
	CV	Coefficient of variation
	r	Correlation coefficient
	DNA	Deoxyribonucleic acid
	n <sub>e</sub>	Effective number of alleles
	Nm	Estimate of gene flow
	FAO	Food and Agriculture Organization of the United Nations
	Hs	Gene diversity within populations
	GA	Genetic advance
	GCV	Genotypic coefficient of variation
	GV	Genotypic variance
	ha	Hectare
	ISSR	Inter simple sequence repeat
	IPGRI	International Plant Genetic Resource Institute
	$MS_E$	Mean square of error

$MS_G$	Mean square of genotype
masl	Metre above sea level
h <sup>2</sup> <sub>B</sub>	Narrow sense heritability
h	Nei's (1973) gene diversity
Ν	North
NPB	Number of polymorphism bands
NTSYS	Numerical taxonomy multivariate analysis system
n <sub>a</sub>	Observed number of alleles
ppm	Parts per million
PPB	Percent of polymorphic bands
PCV	Phenotypic coefficient of variation
PV	Phenotypic variance
PCR	Polymerase chain reaction
PC	Principal component
PCA	Principle component analysis
RAPD	Random amplified polymorphic DNA
RFLP	Restriction fragment length polymorphism
rpm	Revolution per minute
Ι	Shannon's information index
SSR	Simple sequence repeat
SE	Standard error
Taq	Thermus aquaticus

- H<sub>t</sub> Total gene diversity
- TE Tris EDTA buffer
- TBE Tris-borate/EDTA
- UV Ultraviolet
- UPGMA Unweighted pair group method using arithmetic averages



### **CHAPTER 1**

#### **GENERAL INTRODUCTION**

#### 1.1 Introduction

The employment of energy crops as a source of renewable fuels is a concept with great relevance to current ecological and economic issues at both national and global scales. Biodiesel, an alternative fossil fuel, is produced from renewable biological sources such as vegetable oils and animal fats (Fangrui and Milford, 1999).

A hundred years ago, Rudolf Diesel tried vegetable oil as fuel for his engine (Shay, 1993). In the 1930s and 1940s vegetable oils were utilized as diesel fuels from time to time, but usually only in emergency conditions. In recent times, because of increase in crude oil prices, limited resources of fossil oil and environmental concerns there has been a renewed focus on vegetable oils to make biodiesel fuels (Shay, 1993; Fangrui and Milford, 1999). Recently, biofuels extracted from plant species such as *Jatropha curcas* L. has been a major renewable source of energy (Mukherjee *et al.*, 2011).

Biomass as a source of renewable energy is essential for the development and sustenance of civilization. Because of increasing attention in renewable energy sources, liquid bio-energy production from vegetable oils has been seen as one of the possible options to decrease greenhouse gas emissions (Mukherjee *et al.*, 2011). There is a growing interest in technologies to produce biofuels and make them cost competitive with fossil petroleum extracted fuels. Global consciousness of the climatic changes linked to a gradual increase in  $CO_2$  emission in the atmosphere is producing a relative impact on the societies of developed and developing countries (Fangrui and Milford, 1999).

Regarding the abundant benefits of this technology, many countries including Malaysia are interested to invest in it. During 2004–2005, the biodiesel production from vegetable oils was appraised at 2.36 million tonnes globally. Of this, EU and USA accounted for about 82 and 6%, respectively. Global biodiesel production is set to reach some 24 billion liters by 2017 (Divakara *et al.*, 2010).

Jatropha is one of the most promising biofuel sources today. About 30% of the Jatropha seed is comprised of oil. This oil can be easily processed into fuel that can replace or mixed with petroleum based diesel to save on imported oil and most importantly to raise the utilization of green energy (Openshaw, 2000). During the recent years, *J. curcas* has received much attention as a good substitute for fossil fuel in Malaysia. Since Jatropha is a species which can be cultivated by farmers on marginal agricultural farms, it is very logical to invest in Jatropha besides the palm industry.

Regarding large semi wild distribution of Jatropha in different parts of Malaysia, it would be expected to have substantial genetic variation. Environmental factors in addition togenetic and physiological factors play important role in determination of plant potential for seed quality. These characters seem to be under strong genetic control (Ginwal *et al.*, 2005). Depending on the species, germination responses of seed alter in accord with geographical and environmental factors, for example: latitude, elevation, soil nutrient, soil moisture, temperature, density and kind of plant cover (Heller, 1996; Ginwal *et al.*, 2004).

Substantial genetic variation in growth, chemical composition of seed and seed traits at the level of provenance, variety, accession and progeny can be assumed specifically in out crossing species such as many species of Jatropha, *Acacia* and *Prosopis*, which are widely employed in agro-forestry systems overall the world wide landscapes. The variation would be practical as a source for future genetic selection of desired ideo types (Von Carlowitz, 1986). Heavy and bold seeds make possible the seedling to grow vigorously (Kaushik *et al.*, 2003) and better seedling vigor will to lower nursery management time and the maintenance cost. Genetic variation in seed morphology and oil content of Jatropha can be of extensive potential in plant improvement programs, especially selection of genotypes having more seed yield and oil content (Kaushik *et al.*, 2003).

The screening of existing variation in populations for growth and oil yield could be utilized profitably for selection of best genotypes for production of oil. There is a good reason for developing Jatropha as a new energy crop as it does not compete with conventional food crops for land, water and manpower resources and also it has the ability to make a significant contribution to the nation's growing needs for energy though large scale cultivation with ease (Ginwal *et al.*, 2004).

The superior genotype in a plant breeding program is with high yielding, fastest growing and most resistant to diseases and pests. There is no genotype, which has all these characteristics (Gadow and Bredenkamp, 1992). The superior genotype always represents stability. It performs better than other plant when all the criteria together are considered; anyway it may be out performed in any one of the criteria. However, in case of Jatropha, number of branches, number of inflorescences, number of fruits, number of seeds and oil content may be more important traits compared to other traits. The superior plant selection is the first and most important criteria in any plant improvement program. The selection of the superior genotype is based upon various important traits associated with the species and their relative ranking (Mishra, 2009). Genetic diversity underlies the improvement of crops in plant breeding (Pervaiz *et al.*, 2010) and plant breeders lean on the availability of genetic diversity during selection in cultivar development (Ahmed *et al.*, 2010).

Evaluation of the genetic diversity can be computed from utilizing molecular markers (Ahmed *et al.*, 2010). Recently, PCR-based methods for analyzing diversity of the collected populations are usual procedures for revealing and classifying the populations. Molecular marker methods are supporting the classic methods such as physiological and morphological characteristics (Solouki *et al.*, 2008) but the advantage of these techniques is their capacity to discover genetic diversity a more powerful level of resolution than other methods (Sergio and Gianni, 2005).

Therefore, development of diversity studies with molecular genetic analysis for the available populations is significant for Jatropha improvement. Several DNA marker techniques are available to study genetic diversity. But among these, ISSR is an inexpensive and rapid method not requiring any information regarding the genome of the plant, and has been widely used to ascertain the genetic diversity in several plants (Ganesh Ram *et al.*, 2008). In the present study, ISSR markers were utilized to investigate the level and distribution of genetic diversity in different population of Jatropha.

Several researchers have reported crossing and intensive selection between species with desirable traits of Jatropha. Although hybrids have been made in Jatropha diallel mating were not employed to determine general and specific combining ability, heterosis and compatibility. Stability of varieties has not been assayed over different environments. Therefore, genetic diversity studies using molecular markers (ISSR), morphological markers, seed yield and oil content are important to determine genetic diversity in *Jatropha curcas*. In combination with diallel crosses, heterotic groups and performances can be estimated and stable varieties recognized. Molecular marker knowledge will increase efficiency and effectiveness of marker assisted breeding and in conservation of plant genetic resources. Improved Jatropha varieties are urgently needed to improve the Jatropha market through seed yield and oil content as these factors affect oil price on the world market.

### 1.2 Statement of problem

Jatropha is a new crop for Malaysia but the government and private sector is very highly interested in Jatropha oil as a new source of biodiesel. However there is no planting material (seeds) produced locally and they are importing seeds from abroad. In addition, there is lack of published information with regards to genetics and breeding and agronomic performance of Jatropha in Malaysia. It is important to produce local varieties with genetic superiority in yield. From this study, comprehensive information on genetics and agronomic performance of Jatropha evaluated in Malaysia will be published.

### 1.3 Research objectives

The present study was conducted with the following objectives

### Main objective

To identify and select parent lines with high seed and oil yields for commercial

planting

# **Specific objectives**

- 1. To determine inter and intra-population variation using morphological and ISSR molecular markers.
- 2. To estimate genetic components and heritability values of the more important characters in the Jatropha populations.
- 3. To estimate general and specific combining abilities for identifying the best parental lines and the best hybrids.
- 4. To quantify the level of heterosis for  $F_1$  hybrids in diallel crosses.

#### REFERENCES

- Achten, W., Nielsen, L., Aerts, R., Lengkeek, A., Kjær, E., Trabucco, A., *et al.* (2010). Towards domestication of *Jatropha curcas*. *Biofuels*, 1(1), 91-107.
- Achten, W., Verchot, L., Franken, Y., Mathijs, E., Singh, V., Aerts, R., et al. (2008). Jatropha bio-diesel production and use. *Biomass and Bioenergy*, 32 (12), 1063-1084.
- Agarwal, D., and Agarwal, A. (2007). Performance and emissions characteristics of Jatropha oil (preheated and blends) in a direct injection compression ignition engine. *Applied thermal engineering*, 27(13), 2314-2323.
- Agrawal, R. L. (1998). *Fundamentals of plant breeding and hybrid seed production*: Science Publishers, Inc.
- Ahmed, M., Iqbal, M., Masood, M., Rabbani, M., and Munir, M. (2010). Assessment of genetic diversity among Pakistani wheat (*Triticum aestivum* L.) advanced breeding lines using RAPD and SDS-PAGE. *Electronic Journal of Biotechnology*, 13, 1-2.
- Akbar, E., Yaakob, Z., Kamarudin, S., Ismail, M., and Salimon, J. (2009). Characteristic and Composition of *Jatropha Curcas* Oil Seed from Malaysia and its Potential as Biodiesel Feedstock. *European Journal of Scientific Research*, 29(3), 396-403.
- Ali, M., Seyal, M., Awan, S., Niaz, S., Ali, S., and Abbas, A. (2008). Hybrid authentication in upland cotton through RAPD analysis. *Australian Journal of Crop Science*, 2(3), 141-149.
- Arshad, M., Ghafoor, A., and Qureshi, A. (2005). Inheritance of qualitative traits and their linkage in blackgram [*Vigna mungo* (L.) Hepper]. *Pak. J. Bot*, 37(1), 41-46.
- Assefa, K., Ketema, S., Tefera, H., Nguyen, H., Blum, A., Ayele, M., et al. (1999). Diversity among germplasm lines of the Ethiopian cereal tef. *Euphytica*, 106(1), 87-97.
- Bains, N. S., Singh, K., and Basra, A. S. (1999). Genetic basis of heterosis: search for a new paradigm. *Heterosis and hybrid seed production in agronomic crops*, 1-24.
- Basha, S., and Sujatha, M. (2007). Inter and intra-population variability of *Jatropha curcas* (L.) characterized by RAPD and ISSR markers and development of population-specific SCAR markers. *Euphytica*, 156(3), 375-386.

Becker, W. A. (1984). Manual of quantitative genetics: Academic Enterprises.

- Bhattacharya, A., Datta, K., and Datta, S. (2005). Floral biology, floral resource constraints and pollination limitation in Jatropha curcas L. *Pakistan Journal of Biological Sciences*, 8(3), 456-460.
- Bolanos-Aguilar, E., Huyghe, C., Djukic, D., Julier, B., and Ecalle, C. (2001). Genetic control of alfalfa seed yield and its components. *Plant breeding*, 120(1), 67-72.
- Cai, Y., Sun, D., Wu, G., and Peng, J. (2010). ISSR-based genetic diversity of *Jatropha curcas* germplasm in China. *Biomass and Bioenergy*, 34, 1739-1750.
- Chang-wei, L., Kun, L., You, C., Yong-yu, S., and Wen-yun, Y. (2007). Pollen Viability, Stigma Receptivity and Reproductive Features of *Jatropha curcas* L.(Euphorbiaceae)[J]. Acta Botanica Boreali-Occidentalia Sinica, 27. 1994-1996.
- Cilas, C., Bouharmont, P., Boccara, M., Eskes, A., and Baradat, P. (1998). Prediction of genetic value for coffee production in Coffea arabica from a half-diallel with lines and hybrids. *Euphytica*, 104(1), 49-59.
- Daniel, J., and Hegde, N. (2007). *Tree-Borne Oilseeds in Agrof*orestry. Proceedings of Conservation and Management of Agro-resources in Accelerating the Food Production for 21st century held at ICAU, Raipur on 14-15 December, 2006. pp 263-276.
- Das, S., Misra, R., Mahapatra, A., Gantayat, B., and Pattnaik, R. (2010). Genetic Variability, Character Association and Path Analysis in *Jatropha curcas*. *World Applied Sciences Journal*, 8(11), 1304-1308.
- Davis, J., Kay, D., and Clark, V. (1983). Plants tolerant of arid, or semi-arid, conditions with non-food constituents of potential use: Tropical Products Institute, Overseas Development Administration.
- Delannay, I. Y. (2010). Use of molecular markers to increase genetic diversity of Beit Alpha, European Long, and US processing market classes of cucumber (*Cucumis sativus* L.) through marker-assisted selection. The University of Wisconsin-madison.
- Dhillon, R., Hooda, M., Handa, A., Ahlawat, K., and Kumar, Y. (2006). Clonal propagation and reproductive biology in *Jatropha curcas* L. *Indian Journal Agroforest*, 8(2), 18-27.

- Divakara, B., Upadhyaya, H., Wani, S., and Gowda, C. (2010). Biology and genetic improvement of *Jatropha curcas* L.: A review. *Applied Energy*, 87(3), 732-742.
- Dudley, J. (1969). Interpretation and Use of Estimates of Heritability and Genetic Variances in Plant Breeding1. *Crop Science*, 9(3), 257.
- Falconer, D. S. (1989). Introduction to Quantitative Genetics. Third Edition Longman Group Ltd. New York
- Falconer, D. S., and Mackay, T. F. C. (1996). *Introduction to quantitative genetics*. Fourth Edition. Longman Group Ltd., England.
- Fangrui, M., and Milford, A. H. (1999). Biodiesel production: a review. *Bioresource Technology*, 70(1), 1-15.
- Fehr, W. R. (1987). *Principles of cultivar development*. Volume 2. Crop species: Macmillan Publishing Company.
- Foundation, F. (2006). Jatropha Handbook. First draft. www. fact-fuels. org.
- Franco, J., Crossa, J., Ribaut, J., Bertran, J., Warburton, M., and Khairallah, M. (2001). A method for combining molecular markers and phenotypic attributes for classifying plant genotypes. *Theoretical and Applied Genetics*, 103(6), 944-952.
- Freeman, G. (1919). The heredity of quantitative characters in wheat. *Genetics*, 4(1), 1.
- Freitas, R., Missio, R., Matos, F., Resende, M., and Dias, L. (2011). Genetic evaluation of *Jatropha curcas*: an important oilseed for biodiesel production. *Genetics and Molecular Research*, 10(3), 1490-1498.
- Gadow, K., and Bredenkamp, B. (1992). Forest management. Academica, Pretoria, 151.
- Ganesh Ram, S., Parthiban, K., Senthil Kumar, R., Thiruvengadam, V., and Paramathma, M. (2008). Genetic diversity among Jatropha species as revealed by RAPD markers. *Genetic Resources and Crop Evolution*, 55(6), 803-809.
- Garnayak, D., Pradhan, R., Naik, S., and Bhatnagar, N. (2008). Moisture-dependent physical properties of Jatropha seed (*Jatropha curcas* L.). *Industrial crops and products*, 27(1), 123-129.

- Ghosh, A. and Singh, L. (2011). Variation in seed and seedling characters of *Jatropha curcas* L. with varying zones and provenances. *Tropical Ecology*, 52(1), 113-122.
- Ginwal, H., Phartyal, S., Rawat, P., and Srivastava, R. (2005). Seed source variation in morphology, germination and seedling growth of *Jatropha curcas* Linn. in central India. *Silvae Genetica*, 54(2), 76-79.
- Ginwal, H., Rawat, P., and Srivastava, R. (2004). Seed source variation in growth performance and oil yield of *Jatropha curcas* Linn. in central India. *Silvae Genética*, 53(4), 186-191.
- Glaszmann, J., Kilian, B., Upadhyaya, H., and Varshney, R. (2010). Accessing genetic diversity for crop improvement. *Current Opinion in Plant Biology*. 13, 167-173.
- Gohil, R., and Pandya, J. (2008). Genetic diversity assessment in physic nut (*Jatropha curcas* L.). *International Journal of Plant Production*, 2, 321-326.
- Gohil, R., and Pandya, J. (2009). Genetic evaluation of Jatropha (*Jatropha curcas* Linn.) genotypes. *Journal of Agricultural Research (Lahore)*, 47(3), 221-228.
- Grativol, C., da Fonseca Lira-Medeiros, C., Hemerly, A. S., and Ferreira, P. C. G. (2010). High efficiency and reliability of inter-simple sequence repeats (ISSR) markers for evaluation of genetic diversity in Brazilian cultivated *Jatropha curcas* L. accessions. *Molecular biology reports*, 1-12.
- Griffing, B. (1956). Concept of general and specific combining ability in relation to diallel crossing systems. *Australian Journal of Biological Sciences*, 9(4), 463-493.
- Griffiths, A. J. F. (2005). Introduction to genetic analysis: WH Freeman.
- Gübitz, G., Mittelbach, M., & Trabi, M. (1999). Exploitation of the tropical oil seed plant *Jatropha curcas* L. *Bioresource Technology*, 67(1), 73-82.
- Gupta, S., Srivastava, M., Mishra, G., Naik, P., Chauhan, R., Tiwari, S., *et al.* (2008). Analogy of ISSR and RAPD markers for comparative analysis of genetic diversity among different *Jatropha curcas* genotypes. *African Journal of Biotechnology*, 7(23), 4230-4243.
- Hallauer, A. R., and Miranda, J. (1988). *Quantitative genetics in maize breeding*: Iowa State University.
- Hayman, B. (1954). The analysis of variance of diallel tables. *Biometrics*, 10(2), 235-244.

- Heller, J. (1996). Physic nut *Jatropha curcas* L. Promoting the conservation and use of underutilized and neglected crops, 1. *Rome: Institute of Plant Genetics and Crop Plant Research*.
- Ikbal, K., and Dhillon, R. (2010). Evaluation of genetic diversity in *Jatropha curcas* L. using RAPD markers. *Indian Journal of Biotechnology*, 9, 50-57.
- Johnson, G., and King, J. (1998). Analysis of half diallel mating designs. Silvae Genet, 47, 74-79.
- Jongschaap, R., Corré, W., Bindraban, P., and Brandenburg, W. (2007). Claims and Facts on Jatropha curcas L. *Wageningen: Plant Research International*.
- Jubera, M. A., Janagoudar, B., Biradar, D., Ravikumar, R., Koti, R., and Patil, S. (2010). Genetic diversity analysis of elite *Jatropha curcas* (1.) genotypes using randomly amplified polymorphic DNA markers. *Karnataka Journal of Agricultural Sciences*, 22. 1378-1383.
- Katwal, R., and Soni, P. (2003). Biofuels: An opportunity for socio-economic development and cleaner environment. *Indian forester*, 129(8), 939-949.
- Kaushik, N., and Kumar, S. (2004). *Jatropha curcas* L. Silviculture and Uses. *Agrobios (India), Jodhpur*.
- Kaushik, N., Kaushik, J., and Kumar, S. (2003). Response of Jatropha seedlings to seed size and growing medium. *Journal of Non-Timber Forest Products*, 10(1/2), 40-42.
- Kaushik, N., Kumar, K., Kumar, S., and Roy, S. (2007). Genetic variability and divergence studies in seed traits and oil content of Jatropha (*Jatropha curcas* L.) accessions. *Biomass and Bioenergy*, 31(7), 497-502.
- King, A., He, W., Cuevas, J., Freudenberger, M., Ramiaramanana, D., and Graham, I. (2009). Potential of *Jatropha curcas* as a source of renewable oil and animal feed. *Journal of Experimental Botany*, 60. 2897-2901.
- Kobilke, H. (1989). Untersuchungen zur Bestandesbegründung von Purgiernuß (Jatropha curcas L.). Diploma thesis. University Hohenheim, Stuttgart.
- Kohil, A., Raorane, M., Popluechai, S., Kannan, U., Syers, J., and O'donnell, A. (2009). Biofuels: *Jatropha curcas* as a Novel, Non-edible Oilseed Plant for Biodiesel. *Environmental Impact of Genetically Modified Crops*, 296.
- Kojima, T., Nagaoka, T., Noda, K., and Ogihara, Y. (1998). Genetic linkage map of ISSR and RAPD markers in Einkorn wheat in relation to that of RFLP markers. *Theoretical and Applied Genetics*, 96(1), 37-45.

- Kumar, A., and Sharma, S. (2008). An evaluation of multipurpose oil seed crop for industrial uses (*Jatropha curcas* L.): A review. *Industrial crops and products*, 28(1), 1-10.
- Kumar, S., Kumaria, S., Sharma, S. K., Rao, S. R., and Tandon, P. (2011). Genetic diversity assessment of *Jatropha curcas* L. germplasm from Northeast India. *Biomass and Bioenergy*, 35 (7), 3063-3070.
- Lacaze, J. (1979). Advances in species and provenance selection. *Third FAO/IUFRO World*.
- Libby, W., Abbott, A., and Atkin, R. (1987). Genetic resources and variation in forest trees. *Improving vegetatively propagated crops*, 200-209.
- Lörz, H., and Wenzel, G. (2005). *Molecular marker systems in plant breeding and crop improvement*: Springer Verlag.
- Machikowa, T., Saetang, C., and Funpeng, K. (2011). General and Specific Combining Ability for Quantitative Characters in Sunflower. *Journal of Agricultural Science*, 3(1), p91.
- Mantel, N. (1967). The detection of disease clustering and a generalized regression approach. *Cancer research*, 27, 209.
- Mayo, O. (1987). The theory of plant breeding: Clarendon press.
- Meyer, W., Mitchell, T. G., Freedman, E., and Vilgalys, R. (1993). Hybridization probes for conventional DNA fingerprinting used as single primers in the polymerase chain reaction to distinguish strains of Cryptococcus neoformans. *Journal of Clinical Microbiology*, 31 (9), 2274.
- Milbourne, D., Meyer, R., Bradshaw, J., Baird, E., Bonar, N., Provan, J., *et al.* (1997). Comparison of PCR-based marker systems for the analysis of genetic relationships in cultivated potato. *Molecular Breeding*, 3 (2), 127-136.
- Mishra, D. (2009). Selection of candidate plus phenotypes of *Jatropha curcas* L. using method of paired comparisons. *Biomass and Bioenergy*, 33(3), 542-545.
- Mndeme, S. (2008). Sustainable Energy use in rural areas Socio-technical analysis of Jatropha in Same district-Tanzania. University of Wageningen. 105p.
- Mohapatra, S., and Panda, P. K. (2010). Genetic Variability on Growth, Phenological and Seed Characteristics of *Jatropha curcas* L. *Notulae Scientia Biologicae*, 2 (2), 127-132.

- Moyano, C., Alfonso, C., Gallego, J., Raposo, R., and Melgarejo, P. (2003). Comparison of RAPD and AFLP marker analysis as a means to study the genetic structure of Botrytis cinerea populations. *European journal of plant pathology*, 109 (5), 515-522.
- Muchugi, A., Kadu, C., Kindt, R., Kipruto, H., Lemurt, S., Olale, K., et al. (2008). Molecular markers for tropical trees. A practical guide to principles and procedures. ICRAF Technical Manual.
- Mukherjee, P., Varshney, A., Johnson, T. S., and Jha, T. B. (2011). Jatropha curcas: a review on biotechnological status and challenges. *Plant Biotechnology Reports*, 1-19.
- Müller, J., Kratzeisen, M., Weis, K., Stumpf, E., and Mühlbauer, W. (2006). Jatropha curcas derivatives as alternative energy source for households. Proceedings of the Workshop II: Physic nut technological status of *Jatropha curcas* "Farming technology and improved seed of Physic nut in Indonesia", Bogor, Indonesia.
- Nei, M. (1973). Analysis of gene diversity in subdivided populations. Proceedings of the National Academy of Sciences of the United States of America, 70 (12), 3321.
- Nei, M. (1987). Molecular evolutionary genetics: Columbia Univ Pr.
- Nybom, H., and Bartish, I. (2000). Effects of life history traits and sampling strategies on genetic diversity estimates obtained with RAPD markers in plants. *Perspectives in Plant Ecology, Evolution and Systematics*, 3 (2), 93-114.
- Openshaw, K. (2000). A review of *Jatropha curcas:* an oil plant of unfulfilled promise. *Biomass and Bioenergy*, 19 (1), 1-15.
- Ovando-Medina, I., Espinosa-García, F., Núñez-Farfán, J., and Salvador-Figueroa, M. (2011). State of the art of genetic diversity research in *Jatropha curcas*. *Scientific Research and Essays*, 6 (8), 1709-1719.
- Pahlavani, M., Saeidi, G., and Mirlohi, A. (2007). Genetic analysis of seed yield and oil content in safflower using F1 and F2 progenies of diallel crosses. *International Journal of Plant Production*. 1, 129-140.
- Paramathma, M., Reeja, S., Parthiban, K., and Malarvizhi, D. (2006). Development of interspecific hybrids in jatropha. Proceedings of the biodiesel conference toward energy independence—focus on Jatropha, June 9–10, Rashtrapati Bhawan, Hyderabad, India, pp 136-142.

- Pecina-Quintero, V., Anaya-Lopez, J. L., Colmenero, A. Z., Garcia, N. M., Nunez Colin, C. A., Solis Bonilla, J. L., et al. (2011). Molecular characterisation of Jatropha curcas L. genetic resources from Chiapas, México through AFLP markers. *Biomass and Bioenergy*. 35(5):1897-1905.
- Pandey, A., Bhargava, P., Gupta, N., and Sharma, D. (2010). Performance of *Jatropha curcas*: A biofuel crop in wasteland of Madhya Pradesh, India. *Journal homepage: www. IJEE. IEEFoundation. org, 1*(6), 1017-1026.
- Pervaiz, Z., Rabbani, M., Khaliq, I., Pearce, S., and Malik, S. (2010). Genetic diversity associated with agronomic traits using microsatellite markers in Pakistani rice landraces. *Electronic Journal of Biotechnology*, 13, 4-5.
- Popluechai, S., Breviario, D., Mulpuri, S., Makkar, H., Raorane, M., Reddy, A., *et al.* (2009). Narrow genetic and apparent phenetic diversity in Jatropha curcas: initial success with generating low phorbol ester interspecific hybrids. Nature Preceding, pp. 1-28.
- Powers, L. (1944). An expansion of JONES' theory for the exploitation of heterosis. *American Naturalist*, 78, 275-287.
- Raju, A., and Ezradanam, V. (2002). Pollination ecology and fruiting behaviour in a monoecious species, *Jatropha curcas* L.(Euphorbiaceae). *Current Science*, 83(11), 1395-1398.
- Ramawat, K. (2010). Desert Plants: Biology and Biotechnology: Springer Berlin.
- Ramezanpour, S. S., Bastam, S. V., Soltanloo, H., Kia, S., and Kalate, M. (2010). Estimation of Combining Abilities and Heterosis of Septoria tritici'Blotch Resistance in Wheat Genotypes. *Australian Journal of Crop Science*, 4(7), 480.
- Ranade, S., Srivastava, A., Rana, T., Srivastava, J., and Tuli, R. (2008). Easy assessment of diversity in *Jatropha curcas* L. plants using two single-primer amplification reaction (SPAR) methods. *Biomass and Bioenergy*, 32(6), 533-540.
- Rao, G., Korwar, G., Shanker, A., and Ramakrishna, Y. (2008). Genetic associations, variability and diversity in seed characters, growth, reproductive phenology and yield in *Jatropha curcas* (L.) accessions. *Trees-Structure and Function*, 22(5), 697-709.
- Rêgo, E. R., do Rêgo, M. M., Finger, F. L., Cruz, C. D., and Casali, V. W. D. (2009). A diallel study of yield components and fruit quality in chilli pepper (*Capsicum baccatum*). *Euphytica*, 168 (2), 275-287.

- Rohlf, F. (2002). NTSYS-pc: Numerical Taxonomy System, version 2.1 Exeter Publishing. Ltd., Setauket, New York, USA.
- Sabaghnia, N., Dehghani, H., Alizadeh, B., and Mohghaddam, M. (2010). Heterosis and combining ability analysis for oil yield and its components in rapeseed. *Australian Journal of Crop Science*, 4 (6), 390.
- Saikia, S., Bhau, B., Rabha, A., Dutta, S., Choudhari, R., Chetia, M., et al. (2009). Study of accession source variation in morpho-physiological parameters and growth performance of *Jatropha curcas* Linn. *Current Science*, 96 (12), 1631.
- Salimon, J., and Abdullah, R. (2008). Physicochemical Properties of Malaysian Jatropha curcas Seed Oil. Sains Malaysiana, 37 (4), 379-382.
- SAS Institute Inc., (2005). SAS/STAT<sup>®</sup> Users Guide. Version 9.1. SAS Institute Inc. Cary, NC.
- Sayyar, S., Abidin, Z., Yunus, R., and Muhammad, A. (2009). Extraction of Oil from Jatropha Seeds-Optimization and Kinetics. *American Journal of Applied Sciences*, 6 (7), 1390-1395.
- Semagn, K., Bjornstad, Å., and Ndjiondjop, M. (2010). An overview of molecular marker methods for plants. *African Journal of Biotechnology*, 5 (25). 2540-2568.
- Senthil Kumar, R., Parthiban, K., and Govinda Rao, M. (2009). Molecular characterization of Jatropha genetic resources through inter-simple sequence repeat (ISSR) markers. *Molecular biology reports*, 36 (7), 1951-1956.
- Sergio, L., and Gianni, B. (2005). Molecular markers based analysis for crop germplasm preservation. In Proceedings of the International Workshop on the Role of Biotechnology for the Characterisation and Conservation of Crop, Forestry, Animal and Fishery Genetic Resources. pp. 55-65.
- Shay, E. G. (1993). Diesel fuel from vegetable oils: status and opportunities. *Biomass and Bioenergy*, 4 (4), 227-242.
- Shuit, S., Lee, K., Kamaruddin, A., and Yusup, S. (2010). Reactive extraction and in situ esterification of Jatropha curcas L. seeds for the production of biodiesel. *Fuel*, 89 (2), 527-530.
- Shull, G. H. (1914). Duplicate genes for capsule-form inBursa bursa-pastoris. *Molecular and General Genetics MGG*, 12 (1), 97-149.

- Singh, A., Singh, M., Singh, R., Kumar, S., and Kalloo, G. (2006). Genetic diversity within the genus Solanum (Solanaceae) as revealed by RAPD markers. *Current Science*, 90 (5), 711-716.
- Singh, R. K., and Chaudhary, B. D. (1979). *Biometrical methods in quantitative genetic analysis*. Kalyani Publishers, India.
- Singh, Y., Mittal, P., and Katoch, V. (2003). Genetic variability and heritability in turmeric (Curcuma longa L.). *Himachal Journal of Agricultural Research*, 29 (1-2), 31-34.
- Solouki, M., Mehdikhani, H., Zeinali, H., and Emamjomeh, A. (2008). Study of genetic diversity in Chamomile (*Matricaria chamomilla*) based on morphological traits and molecular markers. *Scientia Horticulturae*, 117 (3), 281-287.
- Song, Z., Guan, Y., Rong, J., Xu, X., and Lu, B. (2006). Inter-simple sequence repeat (ISSR) variation in populations of the cutgrass Leersia hexandra. *Aquatic Botany*, 84 (4), 359-362.
- Soontornchainaksaeng, P., and Jenjittikul, T. (2003). Karyology of Jatropha (Euphorbiaceae) in Thailand. *Thai Forest Bulletin*, 31, 105.
- Spooner, D. M., van Treuren, R., and De Vicente, M. (2005). Molecular markers for genebank management: Bioversity International.
- Sprague, G., and Tatum, L. A. (1942). General vs. specific combining ability in single crosses of corn. J. Am. Soc. Agron, 34 (10), 923-932.
- Subramanyam, K., Muralidhararao, D., and Devanna, N. (2010). Genetic diversity assessment of wild and cultivated varieties of *Jatropha curcas* (L.) in India by RAPD analysis. *African Journal of Biotechnology*, 8 (9).1900-1910.
- Sudheer Pamidiamarri, D., Pandya, N., Reddy, M., and Radhakrishnan, T. (2009). Comparative study of interspecific genetic divergence and phylogenic analysis of genus Jatropha by RAPD and AFLP. *Molecular biology reports*, 36 (5), 901-907.
- Sudheer Pamidimarri, D., Singh, S., Mastan, S., Patel, J., and Reddy, M. (2009). Molecular characterization and identification of markers for toxic and nontoxic varieties of *Jatropha curcas* L. using RAPD, AFLP and SSR markers. *Molecular biology reports*, 36 (6), 1357-1364.
- Sujatha, M., Reddy, T., and Mahasi, M. (2008). Role of biotechnological interventions in the improvement of castor (*Ricinus communis* L.) and *Jatropha curcas* L. *Biotechnology advances*, 26 (5), 424-435.

- Sukarin, W., Yamada, Y., and Sakaguchi, S. (1987). Characteristics of physic nut, *Jatropha curcas* L. as a new biomass crop in the Tropics. *JARQ (Japan)*.
- Sultana, T., Ghafoor, A., and Ashraf, M. (2005). Genetic divergence in lentil germplasm for botanical descriptors in relation with geographic origin. *Pak. J. Bot*, 37 (1), 61-69.
- Sun, Q., Li, L., Li, Y., Wu, G., and Ge, X. (2008). SSR and AFLP markers reveal low genetic diversity in the biofuel plant *Jatropha curcas* in China. *Crop Science*, 48 (5), 1865.
- Sunil, N., Sivaraj, N., Anitha, K., Abraham, B., Kumar, V., Sudhir, E., et al. (2009). Analysis of diversity and distribution of *Jatropha curcas* L. germplasm using Geographic Information System (DIVA-GIS). *Genetic Resources and Crop Evolution*, 56 (1), 115-119.
- Takahata, N., and Nei, M. (1984). FST and GST statistics in the finite island model. *Genetics*, 107 (3), 501.
- Tar, M. M., Tanya, P., and Srinives, P. (2011). Heterosis of Agronomic Characters in Jatropha (Jatropha curcas L.). Kasetsart J., 45, 583 - 593.
- Tatikonda, L., Wani, S., Kannan, S., Beerelli, N., Sreedevi, T., Hoisington, D., et al. (2009). AFLP-based molecular characterization of an elite germplasm collection of *Jatropha curcas* L., a biofuel plant. *Plant Science*, 176 (4), 505-513.
- Tchiagam, J. B. N., Bell, J. M., Nassourou, A. M., Njintang, N. Y., and Youmbi, E. (2011). Genetic analysis of seed proteins contents in cowpea (Vigna unguiculata L. Walp.). African Journal of Biotechnology, 10 (16), 3077-3086.
- Vaghela, P., Thakkar, D., Bhadauria, H., Sutariya, D., Parmar, S., and Prajapati, D. (2011). Heterosis and combining ability for yield and its component traits in Indian mustard [Brassica juncea (L.)]. *Journal of Oilseed Brassica*, 2 (1), 39-43.
- Von Carlowitz, P. (1986). Defining ideotypes of multipurpose trees for their phenotypic selection and subsequent breeding. International Workshop on Biological Diversity and Genetic Resources of Underexploited Plants. 20-34 October. Kew Gardens.
- Weiss, E. (1989). Guide to plants tolerant of arid and semi-arid conditions. Nomenclature and potential uses. Margraf Scientific Publishers. Germany. 543pp.

- Wells, O. O., and Wakeley, P. C. (1970). Variation in longleaf pine from several geographic sources. *Forest Science*, 16 (1), 28-42.
- Werlemark, G., Uggla, M., and Nybom, H. (1999). Morphological and RAPD markers show a highly skewed distribution in a pair of reciprocal crosses between hemisexual dogrose species, Rosa sect. Caninae. *TAG Theoretical* and Applied Genetics, 98 (3), 557-563.
- Yeh, F., Yang, R., and Boyle, T. (1999). POPGENE Microsoft Windows-Based Freeware for Population Genetic Analysis Release 1.31. *Alberta, Canada University of Alberta*.
- Zhang, Y., Kang, M. S., and Lamkey, K. R. (2005). Diallel-SAS05: a comprehensive program for griffing's and gardner-eberhart analyses. *Agronomy Journal*, 97 (4), 1097.
- Zhou, Z., Bebeli, P., Somers, D., and Gustafson, J. (1997). Direct amplification of minisatellite-region DNA with VNTR core sequences in the genus Oryza. *Theoretical and Applied Genetics*, 95 (5), 942-949.
- Zietkiewicz, E., Rafalski, A., and Labuda, D. (1994). Genome fingerprinting by simple sequence repeat (SSR)-anchored polymerase chain reaction amplification. *Genomics*, 20 (2), 176-183.