

UNIVERSITI PUTRA MALAYSIA

EFFECTS OF SELECTED ANIMAL BLOOD AND ARTIFICIAL DIETS ON SURVIVAL, GROWTH, AND CHEMICAL COMPOSITION OF ASIAN BUFFALO LEECH, Hirudinaria manillensis

TEH JUN CHIN

FP 2012 82

EFFECTS OF SELECTED ANIMAL BLOOD AND ARTIFICIAL DIETS ON SURVIVAL, GROWTH, AND CHEMICAL COMPOSITION OF ASIAN BUFFALO LEECH, Hirudinaria manillensis



Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of Master of Science

November 2012

Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

EFFECTS OF SELECTED ANIMAL BLOOD AND ARTIFICIAL DIETS ON SURVIVAL, GROWTH, AND CHEMICAL COMPOSITION OF ASIAN

BUFFALO LEECH, Hirudinaria manillensis

Bv

TEH JUN CHIN

November 2012

Chair: Mohd Salleh Kamarudin, PhD

Faculty: Agriculture

Hirudinaria manillensis is well-known as Asian medicinal leech which have been applied in traditional medicine and blood-letting therapy. H. manillensis which is also known as buffalo leech is a haemophagic fresh water leech that widely distributed in paddy field of Southeast Asia including Philippines, Southern China and Malaysia. This study investigated the effects of various casings as feed carriers (sheep intestine skin, cellulose sausage casings and rubber sacs), two different blood sources (cattle and poultry), feeding frequencies (once every 3, 6, 12, 24 days and no feeding), and some chemical compounds (sodium chloride, L-arginine, glycine, glucose) on growth, survival and body composition of Asian buffalo leech H. manillensis. Among the casings tested, the rubber sac has been found to be more suitable as a feed or blood carrier for H. manillensis. The leeches were able to detect the blood contained in the rubber sacs and successfully imbibed the blood. No blood leakage was observed after the leeches detached the sacs following completion of the

feeding. The leeches fed with cattle blood had a significantly higher weight gain

ii

(88%) than poultry blood fed ones (39%). FCR was more efficient with cattle blood (2.57) compared to that of the poultry blood (4.19). However, no significant difference (P>0.05) was found on survival and body composition of the leeches fed on either blood. The leeches fed every 3 days had the highest growth (116%) followed by those fed every 12 days (92%), every 6 days (87%) and every 24 days (13%). However, there was no significant difference (P>0.05) among the growth of leeches that were fed every 3, 6 or 12 days. Therefore, a feeding regime of every 6 to 12 days interval was recommended for the culture of the leech H. manillensis as it was more cost effective from management view. A solution containing a combination of sodium chloride (150 mM), L-arginine (1 mM), glycine (50 mM) and glucose (1 mg/ml) invoked a feeding approach and food consumption similar to that of the cattle blood. Mortalities up to 40% were observed among all feeding leeches while survivals of the non-feeding leeches were always close to 100%. Overfeeding might be one of the reasons for this problem but a further research has to be carried out to find out the actual cause(s) of such heavy deaths among the feeding leeches.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains.

KESAN DARAH HAIWAN PILIHAN DAN DIET BUATAN TERHADAP KEMANDIRIAN, PERTUMBUHAN DAN KOMPOSISI KIMIA LINTAH

KERBAU, Hirudinaria manillensis

Oleh

TEH JUN CHIN

November 2012

Pengerusi: Mohd Salleh Kamarudin, PhD

Fakulti: Pertanian

Hirudinaria manillensis merupakan lintah perubatan Asia yang digunakan dalam bidang perubatan tradisi dan terapi pembekaman. H. manillensis yang juga dikenali sebagai lintah kerbau, adalah lintah air tawar hemofagik yang boleh dijumpai di sawah padi di sekitar Asia Tenggara termasuk Filipina, Cina Selatan dan Malaysia. Kajian ini telah menyiasat kesesuaian pelbagai bahan karung makanan cecair (kulit usus biri-biri, sarung sosej selulosa dan sarung getah), kesan dua jenis darah haiwan (lembu dan ayam), kesan kekerapan pemakanan (setiap 3, 6, 12, 24 hari dan tidak diberi makan), dan kesan perangsang pemakanan lintah dengan kombinasi sebatian kimia terpilih (natrium klorida, L-arginina, glisina dan glukosa) ke atas pertumbuhan, kemandirian dan komposisi badan lintah kerbau H. manillensis. Di antara bahan karung yang diuji, karung getah didapati paling sesuai digunakan sebagai karung makanan atau darah untuk lintah H. manillensis. Lintah didapati berjaya mengesan darah yang terkandung dalam karung getah dan menghisap darah

itu. Tiada kebocoran darah dari permukaan karung getah dapat dilihat selepas lintah

selesai makan. H. manillensis yang memakan darah lembu mencapai penambahan berat badan (88%) bererti (P<0.05) lebih tinggi daripada lintah yang diberi darah ayam (39%). FCR darah lembu (2.57) adalah lebih cekap (P<0.05) berbanding darah ayam (4.19). Bagaimanapun tiada perbezaan bererti (P>0.05) dari segi kemandirian dan komposisi badan lintah yang memakan darah lembu atau darah ayam. Lintah yang diberi makanan setiap 3 hari mempunyai pertumbuhan (116%) yang paling tinggi, diikuti oleh lintah yang diberi makanan setiap 12 hari (92%), setiap 6 hari (87%) dan setiap 24 hari (13%). Namun tiada perbezaan bererti (P>0.05) antara pertumbuhan lintah yang diberi makanan setiap 3, 6 atau 12 hari. Justeru, pemberian makanan setiap selangan 6 hingga 12 hari disarankan untuk kultur lintah H. manillensis kerana ia lebih kos efektif dari segi pengurusan. Campuran cecair yang mengandungi natrium klorida (150mM), L-arginina (1mM), glisina (50mM) dan glukosa (1mg/ml) didapati berjaya membuatkan lintah menghampiri dan menghisapnya pada kadar yang setanding dengan darah lembu. Kematian lintah sehingga 40% diperhatikan di kalangan lintah yang makan berbanding lintah yang tidak makan yang kerap mencapai hampir 100% kemandirian. Pengambilan makanan yang berlebihan mungkin antara penyebab kemandirian yang rendah ini. Penyelidikan yang selanjutnya harus dijalankan untuk mengenalpasti punca sebenar kematian tinggi di kalangan lintah yang makan.

ACKNOWLEDGEMENTS

I would like to express the deepest appreciation to my committee chair, Professor Dr. Mohd Salleh Kamarudin; and committee members, Assoc. Prof. Dr. Che Roos Saad and Mr. Abdullah Abd Rahim, for their professional advices, assistances and their great interest in the pursuit of this study. Without their guidance and persistent help this dissertation would not have been possible.

I would like to acknowledge the support of Aquaculture Department of Agriculture Faculty, Universiti Putra Malaysia, for providing generous support to conduct the research. I would like to thank the abattoir in Shah Alam and the wet market in Seri Kembangan for providing the animal blood. Besides, I would like to thank my family, friends and peers, particularly Dr. Ehsan Ramezani Fard, Ms. Mardhiah Abdul Majid, Mr. Izharuddin Shah Kamaruddin, Mr. Abes Haghi and Ms. Sairatul Dahlianis Ishak for their enormous helps, encouragements and supports.

I am grateful to Ministry of Science, Technology and Innovation Malaysia (MOSTI) for the offer of National Science Fellowship (NSF). I am also thankful to Malaysian Government E-Science (grant no. 05-01-04-SF0209) for providing the financial support.

I certify that a Thesis Examination Committee has met on 28 November 2012 to conduct the final examination of Teh Jun Chin on her thesis entitled "Effects of Selected Animal Blood and Artificial Diets on Survival, Growth, and Chemical Composition of Asian Buffalo Leech, *Hirudinaria manillensis*" in accordance with the Universities and University Colleges Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U. (A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Fatimah bt Md Yusoff, PhD

Professor Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Loh Teck Chwen, PhD

Professor Faculty of Agriculture Universiti Putra Malaysia (Internal Examiner)

Abdul Razak bin Alimon, PhD

Professor Faculty of Agriculture Universiti Putra Malaysia (Internal Examiner)

Metin Kumlu, PhD

Professor Cukurova University Turkey (External Examiner)

SEOW HENG FONG, PhD

Professor and Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date: 26 February 2013

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master in Science. The members of the Supervisory Committee were as follows:

Mohd Salleh Kamarudin, PhD

Professor Faculty of Agriculture Universiti Putra Malaysia (Chairman)

Che Roos Saad, PhD

Associate Professor Faculty of Agriculture Universiti Putra Malaysia (Member)

Abdullah Abd Rahim

Senior Lecturer
Faculty of Agriculture
Universiti Putra Malaysia
(Member)

BUJANG KIM HUAT, PhD

Professor and Dean School of Graduate Studies Universiti Putra Malaysia

Date:

DECLARATION

I declare that the thesis is my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously, and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institution.

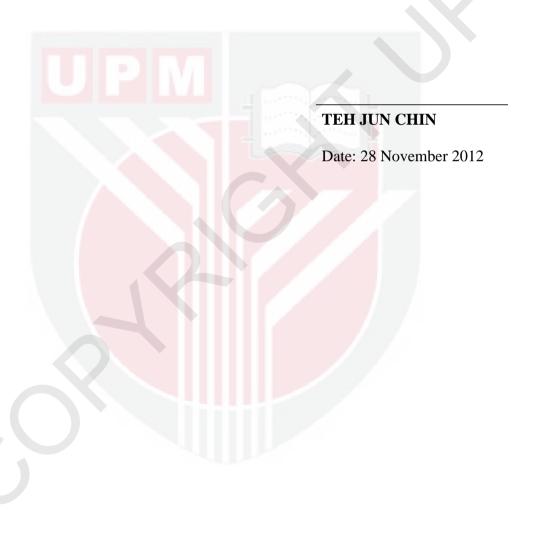


TABLE OF CONTENTS

		Pag		
ABST	RACT	ii		
ABSTRAK ACKNOLEDGEMENTS				
DECL	DECLARATION			
LIST (OF TABLES	xiii		
LIST (OF FIGURES	xvi		
LIST (OF ABBREVIATIONS	xv		
СНАР	TER UP M			
1.	INTRODUCTION	1		
	1.1. Background of Leech	1		
	1.2. Objectives of the Research	3		
2.	LITERATURE REVIEW	5		
	2.1. Importance of Leech	5		
	2.2. Distribution and Identification of Leech	6		
	2.3. Diet and Foraging Behavior	10		
	2.3.1. Chemical Stimuli and Chemosensory	11		
	2.3.2. Leech Digestive Physiology	13		
	2.4. Growth and Reproduction	15		
	2.5. Physiology Behavior and Rearing Condition	16		
	2.5.1. Survivorship, Injury and Disease	18		
	2.5.2. Commercial Culture Practice and Quality Issue	19		
	CENEDAL METHODOLOGY	22		
3.	GENERAL METHODOLOGY 2.1. Collection Standard Proposition of Animal Place	22		
	3.1. Collection, Storage and Preparation of Animal Blood 3.2. Brood Stock Maintenance	22		
		23		
	3.3. Proximate Analysis	23		
4.	PERFORMANCE OF SELECTED CASINGS AS FEED			
	CARRIER FOR FEEDING OF ASIAN BUFFALO LEECH			
	Hirudinaria manillensis	25		
	4.1. Introduction	25		
	4.2. Materials and Methods	27		
	4.2.1. Preparation of Carrier	27		

	4.2.2. Experiment Design	28
	4.2.3. Statistical Analysis	29
	4.3. Results	29
	4.4. Discussion	31
	4.5. Conclusion	35
5.	EFFECTS OF SELECTED ANIMAL BLOOD ON THE	
	SURVIVAL, GROWTH AND BODY COMPOSITION OF ASIAN	
	BUFFALO LEECH Hirudinaria manillensis	35
	5.1. Introduction	35
	5.2. Methodology	36
	5.2.1. Composition of Animal Blood	36
	5.2.2. Feeding with Cattle and Poultry Blood	36
	5.3. Results	38
	5.3.1. Proximate Analysis of Cattle Blood and Poultry Blood	38
	5.3.2. Effect of Feeding with Cattle Blood and Poultry Blood	38
	5.4. Discussion	40
	5.5. Conclusion	41
6.	EFFECT OF FEEDING FREQUENCY ON THE SURVIVAL,	
	GROWTH AND BODY COMPOSITION OF ASIAN BUFFALO	
	LEECH Hirudinaria manillensis	43
	6.1. Introduction	43
	6.2. Methodology	45
	6.2.1. Culture Condition	45
	6.2.2. Experiment Design	45
	6.2.3. Statistical Analysis	46
	6.3. Results	47
	6.4. Discussion	49
	6.5. Conclusion	50
7.	PERFORMANCE OF SELECTED CHEMICAL COMPOUNDS	
	IN ELICITING FEEDING OF ASIAN BUFFALO LEECH	
	Hirudinaria manillensis	52
	7.1. Introduction	52
	7.2. Methodology	54
	7.3. Results	56
	7.4. Discussion	57
	7.5. Conclusion	60

REFERENCES 67



LIST OF TABLES

Table		Page
2.1	Scientific classification of <i>Hirudinaria manillensis</i> (The UniProt Consortium, 2011).	8
4.1	Performance of selected materials as feed carrier.	30
5.1	Chemical composition (% dry matter) of blood.	38
5.2	Effect of feeding with cattle blood and poultry blood on growth performance of <i>H. manillensis</i> in 30 days.	39
5.3	Body composition (% dry matter) of leech after 30 days feeding with cattle blood and poultry blood.	39
6.1	Schedule of feeding for leech <i>H. manillensis</i> .	46
6.2	Effect of different feeding frequencies on growth performance and survival of the buffalo leech at day 30 and day 60.	48
6.3	Proximate composition (% dry matter) of the leeches following different feeding frequencies.	48
7.1	Chemical compounds used in the experiment for the Asian buffalo leech <i>H. manillensis</i> .	55
7.2	The performances of selected chemicals and combinations in stimulating feeding of <i>H. manillensis</i> .	57

LIST OF FIGURES

Figure		Page
2.1	Phylogenetic relationships of seven annelid species (Elliott & Kutschera, 2011).	9
2.2	Picture of <i>Hirudinaria manillensis</i> (Philips & Siddall, 2009).	9
4.1	Red fluid was found in the body of a satiated leech.	30
4.2	No red fluid was observed in a starved leech.	31
8.1	Regurgitation after feeding arisen from leeches.	63
8.2	A deceased leech with hardening nodules on body.	64
8.3	Dead leech probably due to digestion difficulty.	64
8.4	Body of a living leech was partially rotted.	65

LIST OF ABBREVIATIONS

USA United States of America

UK United Kingdom

ANOVA Analysis of Variance

S.E. Standard Error

FDA Food and Drugs Administration, USA

NFE Nitrogen-free Extract

DO Dissolved Oxygen

NaCl Sodium Chloride

Arg Arginine

Glycine

Glc Glucose

DNA Deoxyribonucleic Acid

FCR Food Conversion Ratio

WG Weight Gain

SGR Specific Growth Rate

CHAPTER 1

INTRODUCTION

1.1. Background of Leech

Leeches are widely used as a model animal in toxicology, physiology, neurobiology, biochemist, histology and many other fields of study. The history of clinical and medical use of leech can be traced back to 5th century BC. It was then considerably used in 19th century while the usage went into a decline in the early 20th century (Whitaker *et al.*, 2004; Papavramidou & Christopoulou-Aletra, 2009).

Leech salivary gland secretion contains unique bioactive compounds such as hirudin, destabilase, bdellins, eglins and hyaluronidase. These bioactive components are found very useful for prevention and treatment of various cardiovascular and dermatological diseases (Nikonov *et al.*, 1999; Baskova & Zavalova, 2001; Baskova *et al.*, 2001). Following the recognition by US Food and Drug Administration (FDA), *Hirudo medicinalis* has been used as "medical device" in plastic surgical and hirudo therapy in United States of America (Rados, 2004). Leeches of other species such as *Hirudinaria manillensis* are also getting into the spotlight for their potential value in health care and pharmaceutical products.

H. manillensis is haemophagic fresh water leech which is widely distributed in paddy field of Southeast Asia including the Philippines, Southern China and Malaysia (Harding & Moore, 1927; Enguang, 2008). It is also commonly known as

buffalo leech and being traditionally used for leech therapy by local people. Indeed, the huge interest in *H. manillensis* is emerging as a result of discovery and development of its unique protein extracts that are shown to have important thrombolytic effect and can potentially be the best drug for treatment of cardiovascular and cerebrovascular diseases (Steiner *et al.*, 1990; Latif, 1994; Hayson, 2005).

Not as the perception of most people about leeches, blood is not the only food for them. *H. manillensis* is predacious leech which is equipped with jaws and proboscis at their mouth (Pfeiffer *et al.*, 2005). Leeches can be fed a variety of different invertebrates such as insects (water bugs, gnats and mosquito larvae,), oligochaetes (both aquatic black worms and earthworms), amphipods and different kinds of mollusks including pond snails and freshwater clams (Galun & Kindler, 1968; Govedich & Bain, 2005). The predacious leech can be either engulfs the whole prey or uses its jaws to intrude and suck up the juice of its prey.

Leeches are bred commercially with blood of animals such as eel, catfish, bullfrog, cattle, goat, poultry and swine (Michalsen *et al.*, 2007; Hassan, 2008). Leeches are fed either the live animals or the blood packed in sausage casing (Kornreich & Kleinhaus, 1999; Spencer & Jones, 2007). In Malaysia, live eel or catfish held in the rat traps are put into the leech ponds or tanks. The leeches suck the fish blood to death (Hassan, 2008). This process may takes up to a few hours and thousands of leeches are fed. This method is rather torturing and the welfare of the fishes is seriously neglected.

In fact, the quality of leeches especially for medicinal use must be of highest standard and they must be free from possible contaminants and diseases. Feeding with fresh blood can expose the leeches to potential risk of various diseases. For instance, the fresh cattle blood may carry virus of mad cow disease and has been banned from use in livestock feed in USA and Europe (Phuong, 2004; Fumiere *et al.*, 2009). Besides, some animals are prohibited for certain religions. For example, leeches that feed on swine blood are considered "najis muhalazah" and cannot be used by Muslims.

Therefore, the understanding of appetitive and foraging behavior of medicinal leeches is important for successful breeding and culture of healthy leeches. In this research, suitability of different casings as feed carrier, blood sources from selected live stocks and different feeding frequencies were studied for their effects on survival, growth and body composition of leech. The response of leech *H. manillensis* towards combinations of selected chemical compounds in blood was also evaluated.

1.2. Objectives of the Research

The general purpose of this research was to improve the feeding of leech *H. manillensis* by study on the suitability of different materials as feed carrier and then the effects of selected animal blood, feeding frequency and performance of selected chemical compounds on leech feeding responses. The specific objectives were:

1. To study suitability of different materials (sheep intestine skin, cellulose

- sausage casing and rubber sac) as feed carrier for the Asian Buffalo leech.
- 2. To study the effects of selected animal blood (cattle and poultry) on the survival, growth and chemical composition of leech.
- 3. To study the effects of feeding frequency on survival, growth and chemical composition of leech.
- 4. To study the performance of selected chemical compounds in eliciting feeding responses of leech.



REFERENCES

Abiola, S. S., Sangodoyin, O. S. & Okewole, E. A., 2003. Assessment of quality characteristics of cellulose, sheep and goat casings as sausage containers. *Tropical Vererinarian*, 21(2), pp. 49-53.

AOAC, 1997. Official Methods of Analysis. 16th ed. Maryland: AOAC International.

Barnes, B. M. & Bick, K. L., 1983. Histochemistry of the hemocoel tissues of the leech *Haemopis marmorata*. *Transactions of the American Microscopical Society*, 103(3), pp. 194-212.

Baskova, I. P. & Zavalova, L. L., 2001. Proteinase inhibitors from the medicinal leech *Hirudo medicinalis*. *Biochemistry* (*Moscow*), 66(7), pp. 869-883.

Baskova, I. P., Zavalova, L. L., Basanoval, A. V. & Sass, A. V., 2001. Separation of monomerizing and lysozyme activities of destabilase from medicinal leech salivary gland secretion. *Biochemistry (Moscow)*, 66(12), pp. 1368-1373.

Bauters, T. G. M., Buyle, F. M. A., Verschraegen, G., Vermis, K., Vogelaers, D., Claeys, G. & Robays, H., 2007. Infection risk related to the use of medical leeches. *Pharm World Sci*, Volume 29, pp. 122-125.

Blood Bank Malaysia, 2009. *Penyimpanan dan Kegunaan Darah*. [Online] Available at: http://www.pdn.org.my [Accessed 05 February 2010].

Bradley, R., 2002. Report on the safety of sheep intestine and natural casings derived therefrom in regard to risks from animal TSE and BSE in particular, s.l.: European Commission.

Brodfuehrer, P. D., Tapyrik, L., Pietras, N., Zekavat, G. & Convery, M., 2006. Modification of leech behaviour following foraging for artificial blood. *Journal of Comparative Physiology A.*, Volume 192, pp. 817-825.

Claflin, S. B., Pien, C. L., Rangel, E. N., Utz, K. E., Walther, H. V., Wright, A. N. & Ellerby, D. J., 2009. Effects of feeding on medicinal leech swimming performance. *Journal of Zoology*, Volume 277, pp. 241-247.

Cruz-Lacierda, E. R., Toledo, J. D., Tan-Fermin, J. D. & Burreson, E. M., 2000. Marine leech (*Zeylanicobdella arugamensis*) infestation incultured orange-spotted grouper, *Epinephelus coioides*. *Aquaculture*, Volume 185, pp. 191-196.

Davies, R. W., Dratnal, E. & Linton, L. R., 1996. Activity and foraging behaviour in the predatory freshwater leech *Nephelopsis obscura* (Erpobdellidae). *Funtional Ecology*, 10(1), pp. 51-54.

Davies, R. W., Linton, L. R., Parsons, W. & Edgington, E. S., 1982. Chemosensory detection of prey by *Nephelopsis obscura* (Hirudinoidea: Erpobdellidae). *Hydrobiologia*, Volume 97, pp. 157-161.

Dickinson, M. H. & Lent, C. M., 1984. Feeding behaviour of the medicinal leech, *Hirudo medicinalis* L.. *J. Comp. Physiol. A.*, Volume 154, pp. 449-455.

Dratnal, E., Dratnal, P. A. & Davies, R. W., 1992. The effects of food availability and foraging constraint on the life history of a predatory leech, *Nephelopsis obscura*. *Journal of Animal Ecology*, 61(2), pp. 373-379.

Ducharme, P. E. J., Kajiwara, E. M. & Portnoy, N. A., 1994. *Cellulose food casing manufacturing method*. Illinois, Patent No. 0662283.

Electricwala, A., Hartwell, R., Scawen, M. D. & Atkinson, T., 1993. The complete amino acid sequence of a hirudin variant from the leech *Hirudinaria manillensis*. *Journal of Protein Chemistry*, 12(3), pp. 365-370.

Electricwala, A., Sawyer, R. T., Jones, C. P. & Atkinson, T., 1991. Isolation of thrombin inhibitor from the leech *Hirudinaria manillensis*. *Blood Coagulation Fibrinolysis*, Volume 2, pp. 83-89.

Elliott, E. J., 1986. Chemosensory stimuli in feeding behaviour of the leech *Hirudo* medicinalis. J. Comp. Physiol. A., Volume 159, pp. 391-401.

Elliott, J. M. & Kutschera, U., 2011. Medicinal leeches: historical use, ecology, genetics and conservation. *Freshwater Reviews*, 4(1), pp. 21-41.

El-Shimy, N. A. & Davies, R. W., 1991. The life-cycle, ecology and host specificity of the freshwater leech *Alboglossiphonia polypompholyx* (Glossiphoniidae) in Egypt. *Hydrobiologia*, Volume 222, pp. 173-178.

ENERSOL, 2012. *Interlaboratory Proficiency Trial of Condom Testing Laboratories*, Australia: ENERSOL.

Enguang, T., 2008. Progress in the study of ecology, zoogeography, group, control repellent and medical usage of Hirudinea in China. *Acta Ecologica Sinica*, 28(12), pp. 6272-6281.

Fumiere, O., Veys, P., Boix, A., Holst, C. V., Baeten, V. & Berben, G., 2009. Methods of detection, species identification and quantification of processed animal proteins in feedingstuffs. *Biotechnol. Agron. Soc. Environ.*, 13(S), pp. 59-70.

Galun, R. & Kindler, S. H., 1966. Chemical specificity of the feeding response in *Hirudo medicinalis* (L.). *Comp. Biochem. Physiol.*, Volume 17, pp. 69-73.

Galun, R. & Kindler, S. H., 1968. Regulation of feeding in leech. *Experientia*, 24(11), p. 1140.

Ghawi, A. M., Abdulkader, A. M., Merzouk, A. & Alaama, M., 2012. Season variation and starvation period influence on the antithrombotic activity of leech saliva extract from the medicinal Malaysian leech, *Hirudinaria manillensis*. *J. Bioequiv Availab.*, 4(3), p. 80.

Gouda, H. A., 2006. The effect of peritrich ciliates on some freshwater leeches from Assiut, Egypt. *Journal of Invertebrate Pathology*, Volume 93, pp. 143-149.

Govedich, F. R. & Bain, B. A., 2005. *All About Leeches*, United States of America: Northem Arizona University.

Govedich, F. R., Bain, B. A., Burd, M. & Davies, R. W., 2003. Reproductive biology of the invasive Asian freshwater leech *Barbronia weberi* (Blanchard, 1987). *Hydrobiologia*, Volume 510, pp. 125-129.

Graf, J., Kikuchi, Y. & Rio, R. V. M., 2006. Leeches and their microbiota: naturally simple simbiosis models. *TRENDS in Microbiology*, 14(8), pp. 365-371.

Harding, W. A. & Moore, J. P., 1927. *Hirudinea: The Fauna of British India Including Ceylon and Burma*. London: Thacker, Spink & Co..

Hassan, H. R., 2008. Kertas Makluman Lintah: Biologi, Kegunaan, Ternakan dan Prospek. [Online]

Available at: http://www.pppat.gov.my/culture/lintah/lintah.htm [Accessed 16 December 2009].

Haycox, C. L., Odland, P. B., Coltrera, M. D. & Raugi, G. J., 1995. Indications and complications of medicinal leech therapy. *Journal of the American Academy of Dermatology*, 33(6), pp. 1053-1055.

Hayson, J. M., 2005. Leech therapy: a history. *Journal of the History of Dentistry*, 53(1), pp. 25-27.

Hildebrandt, J. & Lemke, S., 2011. Small bite, large impact - saliva and salivary molecules in the medicinal Leech, *Hirudo medicinalis*. *Naturwissenschaften*, 98(12), pp. 995-1008.

Hovingh, P. & Linker, A., 1999. Hyaluronidase activity in leeches (Hirudinea). *Comparative Biochemistry and Physiology Part B*, Volume 124, pp. 319-326.

Hungtingford, F., 2008. Aquaculture, Innovation and Social Transformation. In: K. Culver & D. Castle, eds. *Chapter 1: Animal Welfare in Aquaculture*. United Kingdom: Springer Science and Business Media B.V., pp. 21-33.

Kalarani, V., Reddy, D. C., Blinn, D. W. & Davies, R. W., 1993. Interspecific differences in respiration and energy storage reserves in two freshwater predatory leeches from ecosystems of constrasting stability. *Comp. Biochem. Physiol. A*, 104(2), pp. 239-242.

Kasparek, M., Demirsoy, A., Akbulut, A., Akbuluk, N. E., Caliskan, M. & Durmus, Y., 2000. Distribution and status of the medicinal leech (*Hirudo medicinalis* L.) in Turkey. *Hydrobiologia*, Volume 441, pp. 37-44.

Kornreich, L. & Kleinhaus, A. L., 1999. Postingestive chemosensation and feeding by leeches. *Physiology & Behavior*, 67(5), pp. 635-641.

Kristan-Jr., W. B., Calabrese, R. L. & Friesen, W. O., 2005. Neuronal control of leech behavior. *Progress in Neurobiology*, Volume 76, pp. 279-327.

Kutchera, U. & Roth, M., 2005. Cannibalism in a population of the medicinal leech (*Hirudo medicinalis* L.). *Biology Bulletin*, 32(6), pp. 626-628.

Lai, Y., Chen, J. & Lee, L., 2011. The chemosensory ability of the predatory leech *Whitmania laevis* (Arhynchobdellida: Haemopidae) for prey searching. *Chemoecology*, Volume 21, pp. 67-74.

Latif, F. A., 1994. *Isolation of antithrombin protein from local leeches (Hirudinaria manillensis)*, Malaysia: Universiti Teknologi Malaysia Institutional Repository.

Marquez, E., Bracho, M., Archile, A., Rangel, L. & Benitez, B., 2005. Proteins, isoleucine, lysine and methionine content of bovine, porcine and poultry blood and their fractions. *Food Chemistry*, Volume 93, pp. 503-505.

Mason, T. A., Sayers, C. W., Paulson, T. L., Coleman, J. & Shain, D. H., 2005. Cocoon deposition and hatching in the aquatic leech, *Theromyzon tessulatum* (Annelida, Hirudinea, Glossiphoniidae). *The American Midland Naturalist*, 154(1), pp. 78-87.

McCue, M., 2010. Starvation physiology: reviewing the different strategies animals use to survive a common challenge. *Comparative Biochemistry and Physiology Part A*, Volume 156, pp. 1-18.

Merila, J. & Sterner, M., 2002. Medicinal leech (*Hirudo medicinalis*) attacking and killing adult amphibians. *Ann. Zool. Fennici*, Volume 39, pp. 343-346.

Michalsen, A., Roth, M. & Dobos, G., 2007. *Medicinal Leech Therapy*. New York: Thieme.

Moore, J. P., 1929. Leeches from Borneo with description of new species. *Proceedings of the Academy of Natural Sciences of Philadelphia*, Volume 81, pp. 267-295.

Mumcuoglu, K. Y., Pidhorz, C., Cohen, R., Ofek, A. & Lipton, H. A., 2007. The use of the medicinal leech, *Hirudo medicinalis*, in the reconstructive plastic surgery. *The Internet Journal of Plastic Surgery*, 4(2), pp. 1-17.

National Research Council Subcommittee on Feed Composition, 1982. *United States-Canadian Tables of Feed Composition: Nutritional Data for United States and Canadian Feeds.* 3rd ed. USA: The National Academic Press.

Nikonov, G. I., Titova, E. A. & Seleznev, K. G., 1999. Biological activity and pharmacological properties of anticoagulant complex (hirudin, plasma kallikrein inhibitor, prostaglandin) from *Hirudo medicinalis*. *Bulletin of Experimental Biology and Medicine*, Volume 12, pp. 1244-1247.

Ooi, C. P., Ahmad, R., Ismail, Z. & Chua, W. S., 2005. Research note: effectiveness of several locally available membranes used for artificial feeding of *Aedes albopictus* Skuse. *Tropical Biomedicine*, 22(1), pp. 69-71.

Orevi, M., Eldor, A., Giguzin, I. & Rigbi, M., 2000. Jaw anatomy of the blood-sucking leeches, Hirudinea *Limnatis nilotica* and *Hirudo medicinalis*, and its relationship to their feeding habits. *J. Zool. Lond.*, Volume 250, pp. 121-127.

Papavramidou, N. & Christopoulou-Aletra, H., 2009. Medicinal use of leeches in the texts of ancient Greek, Roman and early Byzantine writers. *Internal Medicine Journal*, Volume 39, pp. 624-627.

Petrauskiene, L., 2004. The medicinal leech as a convenient tool for water toxicity assessment. *Environ. Toxicol.*, Volume 19, pp. 336-341.

Pfeiffer, I., Brenig, B. & Kutschera, U., 2005. Molecular phylogeny of selected predaceous leeches with reference to the evolution of body size and terrestrialism. *Theory in Biosciences*, Volume 124, pp. 55-64.

Philips, A. J. & Siddall, M. E., 2009. Poly-paraphyly of Hirudinidae: many lineages of medicinal leeches. *BMC Evolutionary Biology*, Volume 9, p. 246.

Phuong, C. L., 2004. Cattle Blood Banned from Feed, USA: Hearst Seattle Media.

Pothikasikorn, J., Bangs, M. J., Chareonviriyaphap, T., Roongruangchai, K. & Roongruangchai, J., 2007. Comparison of blood feeding response and infection of *Aedes aegypti* to *Wuchereria ancrofti* using animal membranes and direct host contact. *Journal of the American Mosquito Control Association*, 23(3), pp. 294-298.

Rados, C., 2004. *Beyond Bloodletting: FDA Gives Leeches a Medical Makeover*, USA: FDA Office of Public Affairs.

Rost-Roszkowska, M. M., Swiatek, P., Kszuk, M., Glowczyk, K. & Bielecki, A., 2011. Morphology and ultrastructure of the midgut in *Piscicola geometra* (Annelida, Hirudinea). *Protoplasma*, pp. 1-11.

Sartor, C., Limouzin-Perotti, F., Legre, R., Casanova, D., Bongrand, M., Sambuc, R. & Drancourt, M., 2002. Nosocomial infections with *Aeromonas hydrophila* from leeches. *Clinical Infectious Diseases*, Volume 35, pp. e1-5.

Sawyer, R. T., 1986. *Leech Biology and Behaviour Volume II: Feeding, Biology, Ecology and Systematics*. Oxford: Oxford Scientific Publications.

Sawyer, R. T., Lepont, F., Stuart, D. K. & Kramer, A. P., 1981. Growth and reproduction of the giant Glossiphoniid leech *Haementeria ghilianii*. *Biol. Bull.*, Volume 160, pp. 322-331.

Scacheri, E., Nitti, G., Valsasina, B., Orsini, G., Visco, C., Ferrera, M., Sawyer, R. T. & Sarmientos, P., 1993. Novel hirudin variants from the leech *Hirudinaria manillensis*. Amino acid sequence, cDNA cloning and genomic organization. *Eur. J. Biochem.*, Volume 214, pp. 295-304.

SCMPMD, 2003. The protection offered by natural rubber latex medical devices (medical gloves and condoms) against transmissible diseases. Brussels, European Commission.

Shimizu, T. & Nakamoto, A., 2001. Segmentation in Annelids: Cellular and Molecular Basis for Metameric Body Plan. *Zoological Science*, 18(3), pp. 285-298.

Sket, B. & Trontelj, P., 2008. Global diversity of leeches (Hirudinea) in freshwater. *Hydrobiologia*, Volume 595, pp. 129-137.

Smith, D. E. & Davies, R. W., 1995. Effects of feeding frequencies on energy partitioning and life history of the leech *Nephelopsis obscura*. *Journal of the North American Benthological Society*, 14(4), pp. 563-576.

Spencer, W. & Jones, G., 2007. The captive breeding and educational display of the medicinal leech *Hirudo medicinalis* (Linnaeus 1758) at Bristol Zoo Gardens. *Int. Zoo Yb.*, Volume 41, pp. 138-144.

Steiner, V., Knecht, R. & Gruetter, M., 1990. Isolation and purification of novel hirudins from the leech *Hirudinaria manillensis* by high-performance liquid chromatography. *Journal of Chromatography*, Volume 530, pp. 273-282.

The UniProt Consortium, 2011. *The UniProt Consortium*. [Online] Available at: http://www.uniprot.org/taxonomy/6419 [Accessed 17 April 2011].

Thear, J. M., 1998. Leeches in medicine. *Ausr. N. Z. J. Surg.*, Volume 68, pp. 292-295.

Tschoerner, P. & Zebe, E., 1989. Ammonia formation in the medicinal leech, *Hirudo medicinalis* - in vivo and in vitro investigation. *Comp. Biochem. Physiol. A.*, 94(2), pp. 187-194.

Ustunol, Z., 2009. Edible Films and Coatings for Meat and Poultry. In: K. C. Huber & M. E. Embuscado, eds. *Edible Films and Coatings for Food Applications*. New York: Springer, pp. 245-268.

Wenning, A., 1996. Managing high salt loads: from neuron to urine in the leech. *Physiological Zoology*, 69(4), pp. 719-745.

Whitaker, I. S., Josty, I. C., Hawkins, S., Azzopardi, E., Naderi, N., Graf, J., Damaris, L., Lineaweaver, W. & Kon, M., 2011. Medicinal leeches and the microsurgeon: a four-year study, clinical series and risk benefit review. *Microsurgery*, 31(4), pp. 281-287.

Whitaker, I. S., Rao, J., Izadi, D. & Butler, P. E., 2004. Historical article: *Hirudo medicinalis*: ancient origins of, and trends in the use of medicinal leeches throughout history. *British Journal of Oral and Maxillofacial Surgery*, Volume 42, pp. 133-137.

Worthen, P. L., Gode, C. J. & Graf, J., 2006. Culture independent characterization of the digestive tract microbiota of the medicinal leech reveals a tripartite symbiosis. *Applied and Environmental Microbiology*, 72(7), pp. 4475-4781.

Wrona, F. J. & Calow, P., 1988. Optimal feeding in a freshwater sit-and-wait predator, *Aloglossiphonia heteroclite* (L.) (Hirudinoidea: Glossiphoniidae). *Functional Ecology*, 2(2), pp. 171-175.

Wurts, W. A., 2003. Daily pH cycle and ammonia toxicity. *World Aquaculture*, 34(2), pp. 20-21.

Yantis, M. A., O'Toole, K. N. & Ring, P., 2009. Leech therapy: *Hirudo medicinalis* has made a comeback. *AJN*, 109(4), pp. 36-43.

Zaidi, S. M., Jameel, S. S., Zaman, F., Jilani, S., Sultana, A. & Khan, S., 2011. A systematic overview of the medicinal importance of sanguivorous leeches. *Alternative Medicine Review*, Volume 16, pp. 59-65.

Zhang, B., Lin, Q., Lin, J., Chu, X. & Lu, J., 2008. Effects of broodstock density and diet on reproduction and juvenile culture of the leech, *Hirudinaria manillensis* Lesson, 1842. *Aquaculture*, Volume 276, pp. 198-204.

Zulhisyam, A. K., Ismail, A. A. & Omar, I. C., 2011. Optimization of growth conditions of Hirudinea sp.. *Australian Journal of Basic and Applied Sciences*, 5(3), pp. 268-275.