

UNIVERSITI PUTRA MALAYSIA

OPTIMIZATION OF DECOLOURISATION OF TEXTILE DYES BY A LOCALLY ISOLATED LIGNINOLYTIC FUNGUS

SIM HAN KOH

FBSB 2007 4



OPTIMIZATION OF DECOLOURISATION OF TEXTILE DYES BY A LOCALLY ISOLATED LIGNINOLYTIC FUNGUS

By

SIM HAN KOH

Thesis Submitted to the School of Graduate Studies, Universiti Putra Malaysia, in Fulfilment of the Requirement for the Degree of Master of Science

January 2007



Dedicated to my father, Sim Wan Chai and mother, Lian Kah Lang, to my elder brother, Sim Han Teck and my two younger sisters, late Sim Hwee Ting and Sim Hwee Min, and to the teachers and lecturers who have taught me everything...



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment of the requirement for the degree of Master of Science

OPTIMIZATION OF DECOLOURISATION OF TEXTILE DYES BY A LOCALLY ISOLATED LIGNINOLYTIC FUNGUS

By

SIM HAN KOH

January 2007

Chairman: Professor Mohd Arif Syed, PhD

Faculty : Biotechnology and Biomolecular Sciences

Water pollution by textile azo dyes is a serious problem worldwide. Local white-rot fungi isolated from soil and wood samples were screened for the ability to degrade textile azo dyes. Seventy one white-rot fungi isolated from various locations in Peninsular Malaysia such as Selangor, Kelantan, Perak and Terengganu were screened for their ability to degrade four textile azo dyes namely Orange G (C.I. 16230), Ponceau 2R (C.I. 16450), Biebrich Scarlet (C.I. 26905) and Direct Blue 71 (C.I. 34140). Forty five isolates gave positive results with varying degrees of degradation. Based on these results, an unidentified white-rot fungus (Isolate S17-UPM) isolated from Universiti Putra Malaysia (UPM) campus in Selangor was selected for further studies due to its ability to completely degrade all four azo dyes in the shortest time. Nutritional studies on defined solid media showed that Isolate



S17-UPM was only able to degrade the four azo dyes under nitrogen-limiting conditions and an additional carbon source in the form of glucose was needed to provide sufficient energy for the degradation to occur. When grown in two-stage liquid culture, Isolate S17-UPM was able to degrade 84 to 99% of 0.2 g/L azo dyes in one to ten days with each dye being degraded at different rates. Orange G was degraded the fastest followed by Ponceau 2R, Direct Blue 71 and Biebrich Scarlet. Generally, azo dye degradation rates were shown to be higher in shake cultures compared to static cultures, with rates almost twice those in static cultures. Isolate S17-UPM degraded the four azo dyes optimally when incubated at temperature between room temperature to 30°C in static cultures. The initial pH of the degradation medium (pH 4.0 to 5.9) had significant effects on the degradation rates, where the highest degradation rate was found to be at pH 4.5. The final pH of all cultures dropped to approximately 4.0. Optimum degradation of the four azo dyes was observed when glucose, sucrose, maltose, lactose and fructose were used separately as additional carbon source. The degradations rates were higher at lower concentrations (0.05 g/L) as compared to higher concentrations (1 g/L) except for Biebrich Scarlet. Assays for lignin-modifying enzymes (LMEs) involved in azo dye degradation showed the presence of laccase (E.C. 1.10.3.2) only while lignin peroxidase (E.C. 1.11.1.14) and manganese peroxidase (E.C. 1.11.1.13) were not detected. Laccase activity profile in static liquid degradation cultures showed correlation to the azo dye degradation profile and was highest in cultures incubated at room temperatures except for Orange G cultures, which was highest at 30 °C. The initial pH of the degradation medium (pH 4.0 to 5.9) did not have any significant



effect on laccase activity except in Ponceau 2R and Biebrich Scarlet cultures where it is highest at pH 5.9. Additional carbon sources such as glucose (6C), sucrose (12C), maltose (12C), lactose (12C) and fructose (6C) which were used separately in cultures incubated with Orange G, Ponceau 2R and Direct Blue 71 gave much higher laccase activity compared to other carbon sources used. Dye concentrations ranging from 0.05 to 1.00 g/L have significant effects on the laccase activity especially Ponceau 2R. Staining activities of laccase in non-denaturing sodium dodecyl sulphate- polyacrylamide gel electrophoresis (SDS-PAGE) showed highlighted green bands around 66 kDa. Laccase produced by Isolate S17-UPM during azo dye degradation was partially purified using Macro-Prep High-QTM strong-anion exchanger and SuperoseTM gel filtration column, when 2,2'-azinobis (3ethylbenzothiazoline-6-sulfonate) (ABTS) was used as the substrate, it was shown to have a K_{m (app)} value of 1.6 mM, V_{max (app)} value of 16.5 µmol/min.ml, optimum activity at 55 to 75°C and pH 2.0 to 3.0 while being most stable at room temperature and pH 6.0 to 7.0. Conclusively, an azo dye-degrading fungus was isolated and the decolourisation process was optimized, while the enzyme involve was partially purified and characterized.



Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

PENGOPTIMUMAN PENYAHWARNAAN PEWARNA TEKSTIL OLEH SEJENIS KULAT LIGNINOLITIK TEMPATAN

Oleh

SIM HAN KOH

Januari 2007

Pengerusi: Profesor Mohd Arif Syed, PhD

Fakulti : Bioteknologi dan Sains Biomolekul

Pencemaran air oleh pewarna tekstil azo merupakan satu masalah seluruh dunia. Kulat reput-putih tempatan yang dipencilkan dari sampel tanah dan kayu telah disaring untuk keupayaan mengurai pewarna tekstil azo. Tujuh puluh satu kultur kulat reput-putih telah dipencilkan dari beberapa lokasi di Selangor, Kelantan, Perak dan Terengganu dan disaring untuk keupayaan mengurai empat pewarna tekstil azo; Orange G (C.I. 16230), Ponceau 2R (C.I. 16450), Biebrich Scarlet (C.I. 26905) dan Direct Blue 71 (C.I. 34140). Empat puluh lima kultur pencilan telah memberikan keputusan positif yang berbeza-beza tahap penguraiannya. Berdasarkan keputusan ini, satu kultur kulat reput-putih yang tidak dikenalpasti (Isolat S17-UPM) yang telah dipencilkan dari sampel di kampus Universiti Putra Malaysia (UPM) Selangor telah dipilih untuk kajian seterusnya kerana keupayaanya mengurai keempat-empat pewarna azo yang digunakan dalam masa yang tersingkat. Kajian nutrisi menggunakan media kultur pejal terperinci menunjukkan Isolat S17-UPM hanya



mampu mengurai keempat-empat pewarna azo tersebut ketika berada di dalam keadaan kekurangan nitrogen dan sumber karbon tambahan seperti glukosa diperlukan untuk membekalkan tenaga yang cukup bagi proses penguraian untuk berlaku. Apabila ditumbuhkan di dalam kultur cecair dua peringkat, Isolat S17-UPM mampu mengurai 84 hingga 99% 0.2 g/L pewarna azo dalam satu hingga sepuluh hari dengan kadar penguraian yang berbeza-beza. Orange G telah diurai terpantas, diikuti oleh Ponceau 2R, Direct Blue 71 dan Biebrich Scarlet. Secara amnya, kadar penguraian pewarna azo adalah lebih tinggi di dalam kultur goncang berbanding di dalam kultur pegun, dengan kadarnya hampir dua kali ganda di dalam kultur pegun. Isolat S17-UPM mengurai pewarna-pewarna azo tersebut secara optimum apabila dieramkan pada suhu bilik hingga 30°C di dalam kultur pegun manakala pH awal media penguraian (pH 4.5 hingga 5.9) mempunyai kesan yang bermakna ke atas kadar penguraian di dalam semua kultur di mana kadar penguraian yang tertinggi berlaku pada pH 4.5. Walaubagaimanapun, pH akhir kesemua kultur telah menurun ke sekitar pH 4.0. Degradasi optimum dapat diperhatikan apabila glukosa, sukrosa, maltosa, laktosa dan fruktosa digunakan secara berasingan sebagai sumber karbon tambahan. Degradasi untuk pewarna yang diuji adalah lebih tiggi pada kepekatan rendah (0.05 g/L) berbanding dengan kepekatan tinggi (1.00 g/L), kecuali Biebrich Scarlet. Pencerakinan untuk enzim-enzim pengubah lignin yang terlibat dengan penguraian pewarna azo hanya menunjukkan kehadiran lakase (E.C. 1.10.3.2) manakala lignin peroksidase (E.C. 1.11.1.14) dan mangan peroksidase (E.C. 1.11.1.13) tidak dapat dikesan. Profil aktiviti laccase di dalam kultur cecair pegun menunjukkan korelasi dengan profil penguraian pewarna azo dan adalah tertinggi di



dalam kultur yang dieram pada suhu bilik kecuali kultur Orange G (30 °C). pH awal media penguraian (pH 4.5 hingga 5.9) tidak mempunyai kesan yang bermakna ke atas aktiviti lakase kecuali di dalam kultur Ponceau 2R dan Biebrich Scarlet di mana ia adalah tertinggi pada pH 5.9. Sumber karbon tambahan seperti glukosa, sukrosa, maltosa, laktosa dan fruktosa yang digunakan secara berasingan dalam kultur Orange G, kultur Ponceau 2R dan kultur Direct Blue 71 menghasilkan aktiviti lakase yang lebih tinggi berbanding dengan kultur yang menggunakan sumber karbon yang lain. Kepekatan pewarna yang digunakan (0.05 hingga 1.00 g/L) mempunyai kesan yang bermakna kepada aktiviti lakase terutamanya dalam kultur Ponceau 2R. Pewarnaan aktiviti laccase menerusi gel elektroforisis sodium dodesil sulfatpoliakrilamida tanpa urai menunjukkan garisan berwarna hijau di sekitar 66 kDa. Lakase yang telah dihasilkan oleh Isolat S17-UPM semasa penguraian pewarna azo telah ditulenkan separa menggunakan kolum penukar anion kuat Macro-Prep Highdan kolum penurasan gel SuperoseTM. Apabila 2,2'-azinobis (3- O^{TM} etilbenzothiazolin-6-sulfonat) (ABTS) digunakan sebagai substrat, ia didapati mempunyai nilai K_m 1.6 mM, nilai V_{max} 16.5 µmol/min.ml, aktiviti optimum pada 55 hingga 75°C dan pada pH 2.0 hingga 3.0 manakala ia adalah paling stabil pada suhu bilik atau ke bawah dan pada pH 6.0 dan 7.0. Kesimpulannya, sejenis kulat pengurai pewarna azo telah dipencilkan dan proses penyahwarnaan telah dioptimumkan, manakala enzim yang terlibat telah ditulenkan separa dan dicirikan.



ACKNOWLEDGEMENTS

First and foremost, I would like to extend my greatest and deepest gratitude to my supervisor, Prof. Dr Mohd Arif Syed for his invaluable guidance throughout the completion of this project. My warmest gratitude also goes to Dr. Mohd Yunus Abdul. Shukor and Dr. Shuhaimi Mustafa for their supervision, patience guidance, and critical review of my work during the course of this research.

I would also like to take this opportunity to thank all members, postgraduates as well as undergraduates from Faculty of Biotechnology and Biomolecular Sciences especially members from Enzymology and Bioremediation Lab (115 and 204) which include Mr Ariff, Mrs Sukirah, Mrs Fazilah, Ms Fazuriana, Ms Surini, Mr Natarajan, Mrs Neni, Ms Farrah, Ms Alia, Mr Fadhil, Mr Tham, Ms Aqlima, and Mr Sharizal for sharing their experiences and knowledge, directly or indirectly. Not forgetting my fellow coursemates, Mr Tang Kah Fai, Mr Lee Lin Keat, Mr Mokrish, Ms Siti Salwa, Ms Ching Mai Keow, Mr Shukuri, Mr Mas Jaffri, Ms Sarah and every staffs from Faculty of Biotechnology and Biomolecular Sciences for their encouragement in making this project a successful one for me and companionship through thick and thin.

Last but not least, I would like to thank my parents Sim Wan Chai and Lian Kah Lang, my brother Sim Han Teck, my sisters Sim Hwee Ting and Sim Hwee Min for their unconditional sacrifices, love and undying support.



I certify that an Examination Committee met on 9th January 2007 to conduct the final examination of Sim Han Koh on his Master of Science thesis entitled "Optimization of Decolourisation of Textile Dyes by a Locally Isolated Ligninolytic Fungus" in accordance with Universiti Pertanian Malaysia (Higher Degree) Act 1980 and Universiti Pertanian Malaysia (Higher Degree) Regulations 1981. The Committee recommends that the candidate be awarded the relevant degree. Members of the Examination Committee are as follows:

Raha Abdul Rahim, PhD

Associate Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Chairman)

Faridah Abdullah, PhD

Associate Professor Faculty of Sciences Universiti Putra Malaysia (Internal Examiner)

Tong Chow Chin, PhD

Associate Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Internal Examiner)

Vikineswary Sabaratnam, PhD

Professor Institute of Biological Sciences Universiti of Malaya (External Examiner)

HASANAH MOHD GHAZALI, PhD

Professor/Deputy Dean School of Graduate Studies Universiti Putra Malaysia

Date :



This thesis submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Master of Science. The members of the Supervisory Committee are as follows:

Mohd. Arif Syed, PhD

Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Chairman)

Mohd. Yunus Abdul Shukor, PhD

Lecturer Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Member)

Shuhaimi Mustafa, PhD

Associate Professor Faculty of Biotechnology and Biomolecular Sciences Universiti Putra Malaysia (Member)

AINI IDERIS, PhD

Professor/Dean School of Graduate Studies Universiti Putra Malaysia

Date : 10 MAY 2007



DECLARATION

I hereby declare that the thesis is based on my original work except for quotations and citations which have been duly acknowledged. I also declare that it has not been previously or concurrently submitted for any other degree at UPM or other institutions.

SIM HAN KOH

Date: 23 APRIL 2007



TABLE OF CONTENTS

iii
vi
ix

ABSTRACT	
ABSTRAK	
ACKNOWLEDGEMENTS	
APPROVAL	
DECLARATION	
LIST OF TABLES	
LIST OF FIGURES	
LIST OF ABBREVIATIONS	

CHAPTER

1	INTI	RODUCTION	1
2	LITH	ERATURE REVIEW	5
	2.1	Industrial Production of Synthetic and Azo Dyes	5
	2.2	Structure and Synthesis of Azo Dyes	6
	2.3	Nomenclature of Azo Dye	9
	2.4	Recalcitrance of Azo Dye in Environments	11
	2.5	Effects of the Contamination caused by Azo Dye from Industrial Effluents	11
	2.6	Partial Degradation of Azo Dyes and the Origin of Aromatic Amines	14
		2.6.1 Definition, Structure and Sources of Aromatic Amines	14
		2.6.2 Carcinogenecity and Toxicity of Aromatic Amines	15
		2.6.3 Formation of Aromatic Amines from Azo Dyes:	17
		Conditions and Mechanisms	
	2.7	Legislations Involving the Use of Azo Dyes Worldwide	19
	2.8	Azo Dye Removal from Wastewater - Existing Examples of	20
		Conventional Technologies	
	2.9	Bioremediation as a Viable Alternative Technology for	23
		Pollution Treatment	
		2.9.1 Bioremediation - Definition and Basic Concepts	23
		2.9.2 Fundamental Approaches in Bioremediation	24
		2.9.3 Factors Influencing Bioremediation	25
	2.10	Bioremediation of Azo Dye by White-rot Fungi	29
		2.10.1 White-rot Fungi and Basidiomycetes	29
		2.10.2 Azo Dye Biodegradation by White-rot Fungi – Before,	31
		Now and Future	
		2.10.3 Advantages of White-rot Fungi Over Other	33
		Microorganisms in Azo Dye Biodegradation	

х

xii

xvi xvii xxi

		2.10.4	Lignin-Modifying Enzymes	35
3	MAT	ERIAL	S AND METHODS	44
	3.1	Overvie	2W	44
	3.2	Chemic	als and Materials	44
	3.3	Isolatio	n and Screening of Fungi	45
		3.3.1	Fungi Sampling	45
		3.3.2	Cleaning and Disinfection of Fungi Samples	46
		3.3.3	Fungi Growth and Isolation	47
		3.3.4	Maintenance of White-Rot Fungi Isolates	49
		3.3.5	Screening of White-Rot Cultures for Azo Dye	50
			Degrading Ability	
	3.4	Azo Dy	re Degradation Studies	53
		3.4.1	Effects of Carbon and Nitrogen Limitation on Azo Dye	53
			Degradation by Isolate S17-UPM	
		3.4.2	Profiles of Static and Shake Liquid Cultures in Azo	55
			Dyes Degradation	
		3.4.3	Effects of Physico-Chemical and Nutritional Factors	61
		_	on Azo Dye Degradation in Static Liquid Cultures	
-	3.5	Enzyma	atic Studies during Azo Dye Degradation	65
		3.5.1	Detection of LMEs Produced by Isolate S17-UPM in	65
			Degradation Cultures and the Effects of Different	
			Environmental Conditions on LMEs Activity	60
		3.5.2	Profile of Azo Dye Degradation and LMEs Production	68
		2 5 2	in I L Stirred Degradation Cultures	-
		3.5.3	Fungal Growth and Laccase Study from 1 L Stirred	/0
		251	Cultures Dartial Durification of Lacassa	72
		3.5.4	Laccase Characterization Studies	74 76
		5.5.5	Laccase Characterization Studies	70
4	RESU	ULTS A	ND DISCUSSIONS	83
	4.1	Ligninc	olytic Fungi Isolated from Peninsular Malaysia	83
	4.2	Screeni	ng of White-Rot Cultures for Azo Dye Degrading	86
		Ability		
	4.3	Azo Dy	e Degradation Studies	91
		4.3.1	Effects of Carbon Limitation and Nitrogen Limitation	91
			on Azo Dye Degradation by Isolate S17-UPM	
		4.3.2	Azo Dyes Degradation Profiles in Static and Shake	94
			Liquid Cultures	
		4.3.3	Effects of Different Environmental Conditions on Azo	101
			Dye Degradation	
	4.4	Enzyma	atic Studies on Azo Dye Degradation	124
		4.4.1	Detection and Studies on the Effects of Environmental	124
			Conditions on LMEs Activity by Isolate S17-UPM in	
			Degradation Cultures	
		4.4.2	Azo Dye Degradation and LMEs Profile in One-Litre	146



			Stirred Degradation Cultures	
		4.4.3	Partial Purification of Laccase	148
		4.4.4	Laccase Characterization Studies	154
	4.5	Identif	ication of Isolate S17-UPM	165
		4.5.1	Fungus identification through morphology	165
5	CON	NCLUSI	ONS	167
RE	FEREN	NCES		170
AP	PENDI	CES		178
BIC	DDATA	OF TH	E AUTHOR	215



LIST OF TABLES

Table		Page
1	Colour Index (C.I.) classification of azo dyes (Ollgaard <i>et al.</i> , 1998)	9
2	Azo dye classification according to method of application	10
3	Non-exhaustive list of other white-rot fungi capable of azo dye biodegradation (Maximo <i>et al.</i> , 2003; Toh <i>et al.</i> , 2003; Wesenberg <i>et al.</i> , 2003; Levin <i>et al.</i> , 2004)	34
4	Effects of nitrogen and carbon limitation on azo dye degradation by Isolate S17-UPM on modified Kirk's basal medium (Tien and Kirk, 1988; Shin <i>et al.</i> , 2002)	54
5	Composition of the growth and degradation media	57
6	Absorption maxima of the four azo dyes used	60
7	Buffers used and their respective pH	78
8	List of successfully isolated ligninolytic fungi	84
9	Time (days) required for white-rot fungi to fully decolourise azo dyes on solid screening medium	87
10	Degradation by Isolate S17-UPM in carbon-limiting conditions	92
11	Degradation by Isolate S17-UPM in nitrogen-limiting conditions	92
12	Percentage of initial degradation, initial adsorption, final adsorption, final degradation and time (days) required for azo dyes degradation in static and shake cultures	97
13	Initial and final pH of the degradation medium	108
14	Laccase purification table	152



LIST OF FIGURES

Figure		Page
1	Structural representation of a trans- monoazo dye.	7
2	Azo dye synthesis via a two-reaction process (Wang et al., 2003)	8
3	Chemical structure of Aniline	14
4	Reduction of azo dyes and formation of aromatic amines	17
5	Proposed mechanism for laccase degradation of azo dyes (Chivukula and Renganathan, 1995)	42
6	Method to obtain dikaryotic tissue by cutting the stipe tissue	48
7	Chemical structures of the sulphonated azo dyes used throughout the study. Depending on the number of azo bond(s), the dyes are classified as monoazo (OG and P2R), disazo (BS) or trisazo (DB71)	52
8	Apparatus setup for 1 L stirred cultures	71
9	Degradation of dyes (decolourised zones) by Isolate S17-UPM on day 7	89
10	Azo dyes (0.2 g/L) degradation profiles by Isolate S17-UPM in 40 ml static liquid cultures.	95
11	Azo dyes (0.2 g/L) degradation profiles by Isolate S17-UPM in 40 ml shake liquid cultures incubated in orbital shakers (115 rpm).	96
12	Effects of incubation temperatures on Orange G (0.2 g/L) degradation in static cultures.	102
13	Effects of incubation temperatures on Ponceau 2R (0.2 g/L) degradation in static cultures.	103
14	Effects of incubation temperatures on Biebrich Scarlet (0.2 g/L) degradation in static cultures.	104
15	Effects of incubation temperatures on Direct Blue 71 (0.2 g/L) degradation in static cultures.	105



16	Effects of initial pH of degradation medium on Orange G (0.2 g/L) degradation.	109
17	Effects of initial pH of degradation medium on Ponceau 2R (0.2 g/L) degradation.	110
18	Effects of initial pH of degradation medium on Biebrich Scarlet (0.2 g/L) degradation.	111
19	Effects of initial pH of degradation medium on Direct Blue 71 (0.2 g/L) degradation.	112
20	Effects of various types of carbon sources on Orange G (0.2 g/L) degradation.	114
21	Effects of various types of carbon sources on Ponceau 2R (0.2 g/L) degradation.	115
22	Effects of various types of carbon sources on Biebrich Scarlet (0.2 g/L) degradation.	116
23	Effects of various types of carbon sources on Direct Blue 71 (0.2 g/L) degradation.	117
24	Effects of azo dye concentrations on Orange G degradation.	119
25	Effects of azo dye concentrations on Ponceau 2R degradation.	120
26	Effects of azo dye concentrations on Biebrich Scarlet degradation.	121
27	Effects of azo dye concentrations on Direct Blue 71 degradation.	122
28	Effects of incubation temperatures on laccase activity in Orange G (0.2 g/L) degradation cultures.	126
29	Effects of incubation temperatures on laccase activity in Ponceau 2R (0.2 g/L) degradation cultures.	127
30	Effects of incubation temperatures on laccase activity in Biebrich Scarlet (0.2 g/L) degradation cultures.	128
31	Effects of incubation temperatures on laccase activity in Direct Blue 71 (0.2 g/L) degradation cultures.	129
32	Effects of initial degradation medium pH on laccase activity in	132



Orange G (0.2 g/L) static cultures.

33	Effects of initial degradation medium pH on laccase activity in Ponceau 2R (0.2 g/L) static cultures.	133
34	Effects of initial degradation medium pH on laccase activity in Biebrich Scarlet (0.2 g/L) static cultures.	134
35	Effects of initial degradation medium pH on laccase activity in Direct Blue 71 (0.2 g/L) static culture.	135
36	Effects of various types of carbon sources on laccase activity in Orange G (0.2 g/L) static culture.	137
37	Effects of various types of carbon sources on laccase activity in Ponceau 2R (0.2 g/L) static culture.	138
38	Effects of various types of carbon sources on laccase activity in Biebrich Scarlet (0.2 g/L) static culture.	139
39	Effects of various types of carbon sources on laccase activity in Direct Blue 71 (0.2 g/L) static culture.	140
40	Effects of azo dye concentrations on laccase activity in Orange G static culture.	142
41	Effects of azo dye concentrations on laccase activity in Ponceau 2R static culture.	143
42	Effects of azo dye concentrations on laccase activity in Biebrich Scarlet static culture.	144
43	Effects of azo dye concentrations on laccase activity in Direct Blue 71 static culture.	145
44	Orange G degradation and laccase activity profile in 1 L stirred cultures.	147
45	Laccase elution profile using Macro-Prep High- Q^{TM} anion exchanger column	149
46	Laccase elution profile using Superose TM gel filtration column.	151
47	Coomasie Brilliant Blue staining of non-denaturing SDS-PAGE of Isolate S17-UPM azo dye degradation cultures fluid.	153



48	Laccase activity staining of non-denaturing SDS-PAGE of Isolate S17-UPM azo dye degradation cultures fluid.	155
49	Lineweaver-Burke graph for laccase with ABTS as the substrate	156
50	Michaelis-Menten curve for laccase with ABTS as the substrate	157
51	Effects of assay temperatures on laccase activity with ABTS as the substrate	159
52	Effects of incubation pH and buffers on laccase activity with ABTS as the substrate	161
53	Effects of different pre-incubation temperatures on laccase stability	162
54	Effects of different pre-incubation pH and buffers on laccase stability	164
55	Isolate S17-UPM used in this study	166



LIST OF ABBREVIATIONS

2,6-DMP	2,6-dimethoxyphenol
ABTS	2,2'- azinobis (3-ethylbenzothiazoline-6-sulfonate)
BS	Biebrich Scarlet
C.I.	Colour Index
CAS	Chemical Abstracts Service
COD	Chemical Oxygen Demand
DB71	Direct Blue 71
DNA	Deoxyribonucleic acid
E.C.	Enzyme Commission
EDTA	Ethylenediaminetetraacetic acid
GPS	Global Positioning System
HPLC	High Performance Liquid Chromatography
IUPAC	International Union of Pure and Applied Chemistry
K _m	Michaelis-Menten Constant
LME	Lignin Modifying Enzyme
OG	Orange G
PDA	Potato Dextrose Agar
P2R	Ponceau 2R
SAAA	Sodium acetate-acetic acid buffer
TE	Trace Elements
V _{max}	Maximum enzyme velocity



CHAPTER 1

INTRODUCTION

The use of synthetic dyes makes this modern world an interesting and colourful place for mankind. Synthetic dyes have mostly replaced natural dyes, especially in the textile industry as a result of their generally superior qualities such as range of colours, colour intensity, ease of manufacture, fastness and resistance to fading by physical, chemical and microbial agents (Wesenberg *et al.*, 2003).

Regardless of the advantages of synthetic dyes over natural dyes, synthetic dyes present their own new set of problems. The most noticeable is the aesthetic pollution of waterways caused by the presence of dyes leached from textile factories since they are visible even in minute amounts (Banat *et al.*, 1996). Not only that, the presence of dyes could also potentially reduce the amount of sunlight reaching the bottom of rivers and lakes and thus affects the ability of water plants to carry out photosynthesis (Banat *et al.*, 1996; Torres *et al.*, 2003; Wesenberg *et al.*, 2003). This will have the net effect of reducing the availability of oxygen in the water to other aquatic animals (Yesilada *et al.*, 2003). Another more dangerous problem is the production of potentially carcinogenic aromatic amine compounds from the partial cleavage of synthetic dyes by anaerobic bacteria found in wastewater treatment plants (Pinheiro *et al.*, 2004), especially from the azoic dye group.



Existing azo dye removal methods usually involve physical and/or chemical treatments. Conventional wastewater treatment such as activated sludge and trickling filters generally fail to decolourise these dye effluents (Kasinath *et al.*, 2003; Wesenberg *et al.*, 2003) and as stated above, they might actually worsen the problem. These methods have many shortcomings. Chemical treatments produce large amounts of chemical sludge with the attendant disposal problems while production of ozone is very costly (Supaka *et al.*, 2003). Physical treatments are also very expensive due to the high operating expenses to produce and regenerate activated carbon (Shen *et al.*, 1992). For these reasons, biological treatments such as utilizing the biodegradative ability of bacteria and ligninolytic fungi are being investigated as a viable and cost effective alternative.

Research into bioremediation, or the use of microorganisms or their enzymes to biotransform the contaminated environments to their original state (Thassitou and Arvanitoyannis, 2001) are currently still in the early stages. Many investigators have isolated fungi from the environment for the biodegradation of textile dyes for the past 20 years or so. Fungi, such as *Phanerochaete chrysoporium* and *Tinctporia* sp., both belonging to the ligninolytic white-rot fungi, are among the first to have been shown to have the ability to degrade azo dyes (Awaluddin *et al.*, 2001). However, until recently, most published research, including those that have been carried out in Malaysia have focused on these temperate species (Awaluddin *et al.*, 2001; Levin *et al.*, 2004) while ignoring the rich biodiversity available in our tropical country.



Despite good degradation, microbes from foreign locations may cause ecological diseases. Thus, local isolates as the best candidates for bioremediation.

Azo dyes, which were designed to be very resistant to physical and biological degradation, are widely used colorants in various industries especially in textile industry. Its ubiquity arises due to its ease of manufacture, low production costs and excellent colours. However, it is now realized that contamination of waterways by azo dyes could lead to some potentially serious ecological and health problems. Current conventional water treatment methods are unable to remove them efficiently or are too expensive to apply in large-scale plants. To biodegrade the azo dyes efficiently, economically and at the same time being environmentally friendly, bioremediation offers an attractive solution.

Biodegradation of azo dyes by white-rot fungi presents a great potential for largescale applications after many bioremediation processes being investigated for this purpose. Its biodegradation system, comprising of lignin modifying enzymes are not only efficient but also have a wide substrate range. At this time, most research are focused on a narrow range of well-known white-rot fungi while the rich biodiversity of fungi found in tropical forests such as in Malaysia is ignored most of the time. There are reasons to believe that these undiscovered species might have greater azo dye degrading abilities compared to the ones that are being studied now. Hence, the processes to isolate and screen new white-rot fungi for the biodegradation of azo dyes have to be done intensively.

