



UNIVERSITI PUTRA MALAYSIA

***MOLECULAR CHARACTERIZATION OF GDPMANNOSE
PYROPHOSPHORYLASE, GDPMANNOSE-3', 5'-EPIMERASE AND
GALACTOSE-1-PHOSPHATE URIDYLYLTRANSFERASE
RECOMBINANT PROTEINS FROM GRACILARIA
CHANGII I. A. ABBOTT***

SIOW ROUH SAN

FBSB 2012 48

**MOLECULAR CHARACTERIZATION OF GDP-
MANNOSE PYROPHOSPHORYLASE, GDP-
MANNOSE-3', 5'-EPIMERASE AND GALACTOSE-
1-PHOSPHATE URIDYLYLTRANSFERASE
RECOMBINANT PROTEINS FROM *GRACILARIA*
CHANGII I. A. ABBOTT**

The logo of Universiti Putra Malaysia (UPM) is a shield-shaped emblem. It features a red and white design with a central vertical element and a book-like shape at the top. The letters 'UPM' are prominently displayed in a red box at the top left of the shield.

SIOW ROUH SAN

**MASTER OF SCIENCE
UNIVERSITI PUTRA MALAYSIA**

2012

**MOLECULAR CHARACTERIZATION OF GDP-MANNOSE
PYROPHOSPHORYLASE, GDP-MANNOSE-3', 5'-EPIMERASE AND
GALACTOSE-1-PHOSPHATE URIDYLYLTRANSFERASE RECOMBINANT
PROTEINS FROM *GRACILARIA CHANGII* I. A. ABBOTT**

By

SIOW ROUH SAN

**Thesis Submitted to the School of Graduate Studies, Universiti Putra
Malaysia, in Fulfillment of the Requirements for the Degree of Master of Science**

2012

SIOW ROUH SAN

MASTER OF SCIENCE

2012



Abstract of thesis presented to the senate of Universiti Putra Malaysia in fulfillment of the requirement for the degree of Master of Science

**MOLECULAR CHARACTERIZATION OF GDP-MANNOSE
PYROPHOSPHORYLASE, GDP-MANNOSE-3', 5'-EPIMERASE AND
GALACTOSE-1-PHOSPHATE URIDYLYLTRANSFERASE
RECOMBINANT PROTEINS FROM *GRACILARIA CHANGII* I.A. ABBOTT**

By

SIOW ROUH SAN

2012

Chairman : Assoc. Prof. Dr. Ho Chai Ling, PhD

Faculty : Faculty of Biotechnology and Biomolecular Sciences

Gracilaria changii is a red seaweed which grows in the muddy and silted mangroves fringing the west coast of Peninsular Malaysia such as Morib, Selangor. *Gracilaria* plays an important role in the phycocolloid industry for agar production. GDP-mannose pyrophosphorylase (GMP), GDP-mannose-3', 5'-epimerase (GME) and galactose-1-phosphate uridylyltransferase (GALT) are the enzymes involved in the biosynthesis of D- and L-galactose (basic unit of agar). Although the complete biosynthetic pathway of agar and agarose biosynthesis is not known, the regulating steps of agar and agarose biosynthesis is believed to lie in the intermediate pathways involving the biosynthesis of UDP-D and GDP-L-galactose. The objectives of this study were to express the cDNAs encoding GcGALT, GcGME and GcGMP as recombinant protein in *Escherichia coli* for biochemical assays and to isolate the 5' flanking regions of these three enzymes from *G. changii*. The recombinant proteins of GcGALT and GcGME were successfully expressed as soluble proteins in *E. coli* strain BL21 (DE3) pLysS. The enzyme activity of recombinant GcGALT was

determined in a coupled assay by monitoring the reduction of NAD and NADP. For the forward reaction, the K_m (UDP-glucose) and K_m (galactose-1-phosphate) were 0.134 mM and 0.116 mM, respectively. For the reverse reaction, K_m (glucose-1-phosphate) and K_m (UDP-galactose) were 0.092 mM and 0.051 mM, respectively. The analysis of high performance liquid chromatography (HPLC) showed that the purified recombinant GcGME formed two products, most probably GDP-L-galactose and GDP-L-gulose. The recombinant protein of GcGMP was expressed as inclusion bodies in *E. coli* strain Origami (DE3) pLysS. The inclusion bodies were solubilized and refolded for enzyme assay using HPLC. However, the refolded recombinant GcGMP did not show any activity. The structural gene sequences of GcGALT, GcGME and GcGMP isolated from the genomic DNA of *G. changii* were devoid of introns. Cis-acting regulatory element related to light, methyl jasmonate responses and meristem specific activation/expression were found at the 5' flanking regions of *GcGALT*, *GcGME* and *GcGMP*. The cis-acting regulatory element involved in light response showed the highest frequency in the 5' flanking regions of *GcGALT*, *GcGME* and *GcGMP*. The molecular and biochemical characterization of recombinant GcGALT, GcGME and GcGMP may facilitate the understanding of agar production in *G. changii*.

Abstrak tesis yang dikemukakan kepada Senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Master Sains

**PENCIRIAN MOLEKUL GDP-MANNOSE PYROPHOSPHORYLASE,
GDP-MANNOSE-3', 5'-EPIMERASE DAN GALACTOSE-1-PHOSPHATE
URIDYLYLTRANSFERASE PROTEIN REKOMBINAN DARIPADA
GRACILARIA CHANGII I.A. ABBOTT**

Oleh

SIOW ROUH SAN

2012

Pengerusi : Prof. Madya Dr. Ho Chai Ling

Fakulti : Fakulti Bioteknologi dan Sains Biomolekul

Gracilaria changii merupakan sejenis rumpai laut merah yang tumbuh di paya bakau berlumpur di sekitar pantai barat Semenanjung Malaysia, seperti Morib di Selangor dan Port Dickson di Negeri Sembilan. *Gracilaria* memainkan peranan penting dalam industri fikokoloid untuk penghasilan agar. GDP-mannose pyrophosphorylase (GMP), GDP-mannose-3', 5'-epimerase (GME) dan galactose-1-phosphate uridylyltransferase (GALT) merupakan enzim yang terlibat dalam biosintesis D- dan L-galaktosa iaitu unit asas untuk pembinaan agar. Walaupun pengetahuan mengenai tapak jalan biosintesis agar dan agarosa yang lengkap tidak diketahui, biosintesis agar dan agarosa dipercayai terletak di laluan perantaraan yang melibatkan biosintesis UDP-D dan GDP-L-galaktosa. Objektif kajian ini adalah untuk menghasilkan protein rekombinan dengan mengekspres cDNA yang mengekodkan GcGALT, GcGME dan GcGMP di dalam *Escherichia coli* untuk asai biokimia dan untuk mengasingkan bahagian terapan 5' *GcGALT*, *GcGME* dan

GcGMP daripada *G. changii*. Protein rekombinan GcGALT dan GcGME dihasilkan sebagai protein yang larut di dalam *E. coli* BL21 (DE3) pLysS. Aktiviti enzim bagi rekombinan protein GcGALT ditentukan oleh asai berpasang dengan memantau reduksi NADP dan NAD. Bagi tindak balas ke depan, K_m (UDP-glucose) dan K_m (galactose-1-phosphate) adalah 0.134 mM dan 0.116 mM masing-masing. Bagi tindak balas ke belakang, K_m (glucose-1-phosphate) dan K_m (UDP-galactose) adalah 0.092 mM and 0.051 mM masing-masing. Analisis kromatografi cecair berprestasi tinggi menunjukkan bahawa protein rekombinan GcGME yang tulen membentuk dua produk, yang berkemungkinan iaitu GDP-L-galactose dan GDP-L-gulose. Protein rekombinan GcGMP dihasilkan sebagai protein yang tidak larut di dalam *E. coli* Origami (DE3) pLysS. Protein rekombinan GcGMP yang tidak larut telah dilarutkan untuk penglipatan semula dan menjalani asai enzim dengan menggunakan kromatografi cecair berprestasi tinggi. Namun, protein rekombinan GcGMP tidak menunjukkan sebarang aktiviti. Jujukan gen struktur *GcGALT*, *GcGME* dan *GcGMP* daripada DNA genomik *G. changii* adalah tanpa intron. Elemen pengawalaturan cis yang berkaitan dengan tindak balas cahaya, metil jasmonat dan pengaktifan/ekspresi meristem dijumpai di bahagian terapan 5' *GcGALT*, *GcGME* dan *GcGMP*. Elemen pengawalaturan cis yang berkaitan dengan tindak balas cahaya menunjukkan kekerapan tertinggi dalam bahagian terapan 5' *GcGALT*, *GcGME* dan *GcGMP*. Pencirian molekul dan biokimia *GcGALT*, *GcGME* and *GcGMP* dapat meningkatkan pemahaman tentang penghasilan agar daripada *G. changii*.

Acknowledgements

I would like to express my sincere gratitude and whole-hearted appreciation to my supervisor Assoc. Prof. Dr. Ho Chai Ling and co-supervisor Assoc. Prof. Dr. Mohd. Yunus Abd. Shukor for their advice and guidance throughout my master study.

I would like to thank the Ministry of Science, Technology and Innovation (MOSTI) of Malaysia for giving the Intensified Research Grant for Priority Area (IRPA) No. 06-02-02-003 BTK/ER/01 and eSciencefund grant number 02-01-04-SF0018. I would also like to thank Universiti Putra Malaysia for giving me the Graduate Research Fellowship.

My highest appreciation to Assoc. Prof. Dr. Mohd. Puad Abdullah and Assoc. Prof. Dr. Norihan Mohd. Saleh for giving me the permission to use the equipment in Proteomic Lab and Plant Transformation Lab, respectively. I gratefully acknowledge Encik Ibrahim Yunus and Encik Mohomad Cyrill Kamal from the Department of Biochemistry, Faculty of Biotechnology and Biomolecular Sciences for their assistance in the HPLC work. Also I would like to forward my many thanks to Miss Nur Anisza Hanoum bt Naseron for the preparation of genomic DNA and Genome Walker Library from *Gracilaria changii*. Finally, I wish to extend my gratitude to all members from Cell and Molecular Biology Lab for their concern, assistance and constructive discussion throughout this project.

Of course, I would not forget to express my deepest appreciation to my family for their love and support throughout these years. It would not have been anything without all of you. I sincerely thank you all for your care and support.

Approval Sheet 1

I certify that a Thesis Examination Committee has met on (4th September 2012) to conduct the final examination of Siow Rouh San on her thesis entitled “Molecular characterization of GDP-mannose pyrophosphorylase, GDP-mannose-3’, 5’-epimerase and galactose-1-phosphate uridylyltransferase recombinant proteins from *Gracilaria changii* I.A. Abbott” in accordance with the Universities and University College Act 1971 and the Constitution of the Universiti Putra Malaysia [P.U.(A) 106] 15 March 1998. The Committee recommends that the student be awarded the Master of Science.

Members of the Thesis Examination Committee were as follows:

Noorjahan Banu bt Mohamed Alitheen, PhD

Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Chairman)

Sieo Chin Chin, PhD

Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Internal examiner)

Adam Leow Thean Chor, PhD

Faculty of Biotechnology and Biomolecular Sciences
Universiti Putra Malaysia
(Internal examiner)

K. Sudesh Kumar C. Kanapathi Pillai, PhD

School of Biological Sciences
Universiti Sains Malaysia
(External examiner)

SEOW HENG FONG, PhD
Professor and Deputy Dean
School of Graduate Studies
Universiti Putra Malaysia

Date:

Approval Sheet 2

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfillment of the requirement for the degree of Master of Science. The members of the Supervisory Committee were as follows:

Ho Chai Ling, PhD

Associate Professor

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Chairman)

Mohd. Yunus Abd. Shukor, PhD

Associate Professor

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Member)

BUJANG KIM HUAT, PhD

Professor and Dean

School of Graduate Studies

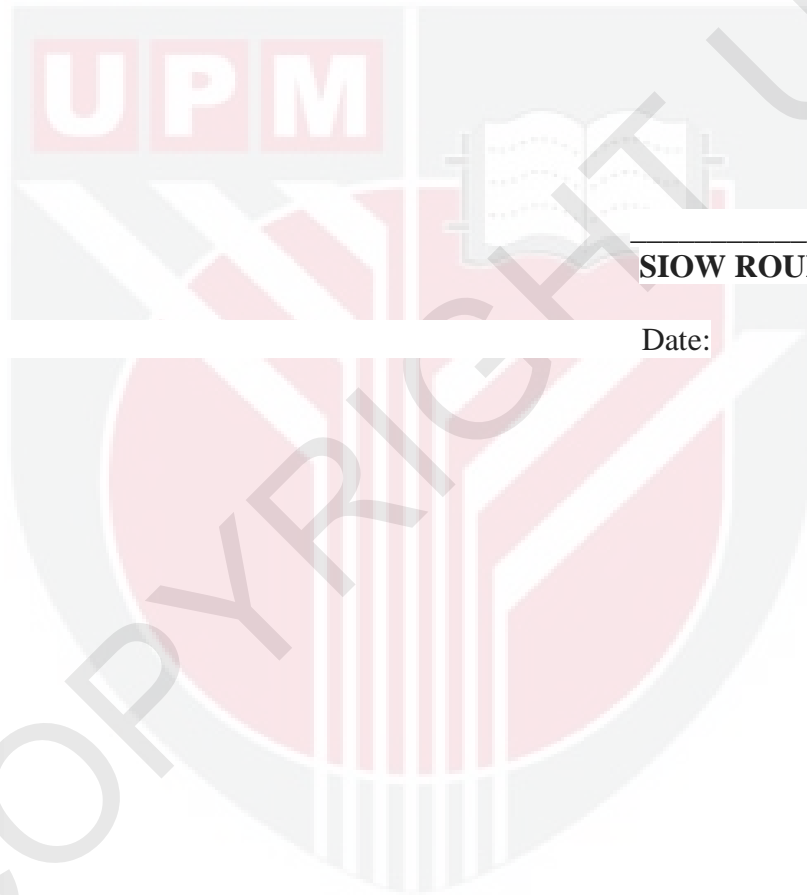
Universiti Putra Malaysia

Date:

Declaration Form

DECLARATION

I declare that the thesis is my original work except for the quotations and citations which have been duly acknowledged. I also declare that it has not been previously, and is not concurrently, submitted for any other degree at Universiti Putra Malaysia or at any other institution.



SIOW ROUH SAN

Date:



TABLE OF CONTENTS

ABSTRACT	i
ABSTRAK	iii
ACKNOWLEDGEMENTS	v
APPROVAL	vi
DECLARATION	viii
LIST OF TABLES	xiii
LIST OF FIGURES	xiv
LIST OF APPENDICES	xvii
LIST OF ABBREVIATION	xviii
LIST OF SYMBOLS AND UNITS	xx

CHAPTER

1	INTRODUCTION	1
2	LITERATURE REVIEW	
	2.1 Seaweeds	4
	2.1.1 Classification and biology of seaweeds	4
	2.1.2 The uses of seaweeds	5
	2.1.3 <i>Gracilaria</i>	7
	2.2 Agar Biosynthesis	12
	2.2.1 The structure of agar	12
	2.2.2 The biosynthesis of agar	13
	2.2.3 Enzymes involved in the biosynthesis of galactose	15
	2.2.3.1 Galactose-1-phosphate uridylyltransferase	15
	2.2.3.2 GDP-mannose-3',5'-epimerase	18
	2.2.3.3 GDP-mannose pyrophosphorylase	22
	2.2.4 Factors affecting the agar yield and quality	24
	2.2.5 The uses and market value of agar	26
	2.3 The regulatory elements involved in the regulation of gene expression of <i>GALT</i> and <i>GMP</i>	27
	2.4 The biochemical characterization of <i>GALT</i> , <i>GME</i> and <i>GMP</i>	28
3	METHODOLOGY	
	3.1 Sequences analysis and homology modeling	31
	3.2 Transformation of the expression vector into expression host	31
	3.2.1 Isolation of the expression vectors harboring <i>GcGALT</i> , <i>GcGME</i> and <i>GcGMP</i>	31
	3.2.2 Transformation of expression vector into <i>E. coli</i> strain BL21 (DE3) pLysS	33
	3.2.3 Transformation of expression vector into <i>E. coli</i> Origami (DE3) pLysS	34
	3.2.4 Confirmation of positive transformants	34
	3.2.4.1 Bacterial culture and plasmid preparation	34
	3.2.4.2 Restriction enzymes (RE) analysis	35

3.3 Recombinant protein expression	36
3.3.1 Recombinant protein induction	36
3.3.2 Cell lysis	37
3.3.3 Purification of recombinant proteins	37
3.3.3.1 Purification of inclusion bodies	37
3.3.3.2 Refolding of inclusion bodies	38
3.3.3.3 Purification of refolded recombinant GcGMP	39
3.3.4 Protein quantification using Bradford assay	39
3.3.5 Sodium dodecyl sulfate-polyacrylamide gel electrophoresis(SDS-PAGE)	39
3.3.6 Western blot	40
3.4 Enzyme assays	41
3.4.1 Enzyme assay of recombinant GcGALT	41
3.4.1.1 Standard curve of NADH and NADPH	41
3.4.1.2 Enzyme assay of recombinant protein GcGALT	41
3.4.1.3 Kinetic analysis of recombinant protein GcGALT	42
3.4.1.4 Extraction of total proteins from seaweeds and enzyme assay of seaweed GALT	43
3.4.2 Enzyme assay of recombinant GcGME	44
3.4.2.1 Thin layer chromatography (TLC)	44
3.4.2.2 High performance liquid chromatography (HPLC)	44
3.4.3 Enzyme assay of recombinant GcGMP	45
3.4.3.1 Enzyme assay for crude extract of recombinant GcGMP(soluble fraction)	45
3.4.3.2. Enzyme assay for refolded recombinant GcGMP	45
3.5 Isolation of genomic DNA sequences	46
3.5.1 Cloning of structural gene sequences	46
3.5.1.1 Primer design	46
3.5.1.2 PCR amplification of structural gene sequences	47
3.5.1.3 Purification and ligation of PCR products	48
3.5.1.4 Transformation	49
3.5.2 Confirmation of putative transformants (structural gene sequences)	49
3.5.2.1 Colony PCR	49
3.5.2.2 Restriction enzymes (RE) analysis	50
3.5.2.3 Analysis of structural gene sequences	50
3.5.3 Cloning of 5' flanking region of <i>GcGALT</i> , <i>GcGME1</i> and <i>GcGMP</i>	50
3.5.3.1 Primer design	50
3.5.3.2 Primary and secondary PCR	51
3.5.3.3 Purification and ligation of secondary PCR products	52
3.5.3.4 Transformation	53
3.5.4 Confirmation of putative transformants (5' flanking regions)	53
3.5.4.1 Colony PCR	53
3.5.4.2 Analysis of 5' flanking regions	53

3.6	Inverse PCR to isolate the 5' flanking region of <i>GcGME2</i>	53
3.6.1	Genomic DNA extraction from <i>G. changii</i>	53
3.6.2	RE digestion of genomic DNA	54
3.6.3	Inverse PCR for GcGME	55
3.6.4	Cloning of the GcGME inverse PCR products	55
3.6.5	Analysis of the 5' flanking region of <i>GcGME 2</i>	56
4	RESULTS	
4.1	Galactose-1-phosphate uridylyltransferase (GcGALT)	57
4.1.1	Sequence analysis and homology modeling	57
4.1.2	Induction and detection of recombinant GcGALT	61
4.1.3	Enzyme assay of recombinant GcGALT	63
4.1.3.1	Enzyme assay and kinetic analyses of recombinant GcGALT	63
4.1.3.2	Enzyme activity of GcGALT in the seaweeds crude extract	70
4.1.4	Isolation of genomic sequence of GcGALT	71
4.2	GDP-mannose-3',5'-epimerase (GcGME)	77
4.2.1	Sequence analysis and homology modelling	77
4.2.2	Induction and detection of recombinant GcGME	81
4.2.3	Enzyme assay for recombinant GcGME	83
4.2.4	Isolation of genomic sequence of GcGME	88
4.3	GDP-mannose pyrophosphorylase (GcGMP)	95
4.3.1	Sequence analysis and homology modelling	95
4.3.2	Induction and detection of recombinant GcGMP	99
4.3.3	Enzyme assay for crude extract of recombinant GcGMP	101
4.3.4	Refolding of recombinant GcGMP	102
4.3.4.1	Enzyme assay for refolded recombinant GcGMP	103
4.3.5	Isolation of genomic sequence of GcGMP	104
5	DISCUSSION	
5.1	Galactose-1-phosphate uridylyltransferase (GcGALT)	110
5.1.1	Sequence analysis and homology modelling	110
5.1.2	Induction and enzyme assay of recombinant GcGALT	111
5.1.3	Isolation of genomic sequence of GcGALT	113
5.2	GDP-mannose-3',5'-epimerase (GcGME)	115
5.2.1	Sequence analysis and homology modelling	115
5.2.2	Induction and enzyme assay of recombinant GcGME	117
5.2.3	Isolation of genomic sequence of GcGME	119
5.3	GDP-mannose pyrophosphorylase (GcGMP)	120
5.3.1	Sequence analysis and homology modelling	120
5.3.2	Induction and enzyme assay of recombinant GcGMP	121
5.3.3	Isolation of genomic sequence of GcGMP	123
5.4	Analysis of cis-elements in <i>GcGALT</i> , <i>GcGME</i> and <i>GcGMP</i>	125

6 SUMMARY, CONCLUSIONS AND RECOMMENDATION FOR FUTURE STUDIES	
6.1 Summary and Conclusions	128
6.2 Recommendations for future studies	129
REFERENCES	131
APPENDICES	147
BIODATA OF STUDENT	159
LIST OF PUBLICATIONS	160

