



UNIVERSITI PUTRA MALAYSIA

***PRODUCTION AND PURIFICATION OF NIPAH VIRUS GLYCOPROTEIN IN
Spodoptera frugiperda 9 (J.E. SMITH) INSECT CELL***

RAKSHA SUNHARE

IB 2013 9



**PRODUCTION AND PURIFICATION OF NIPAH
VIRUS GLYCOPROTEIN IN *Spodoptera frugiperda*
9 (J.E. SMITH) INSECT CELL**

RAKSHA SUNHARE

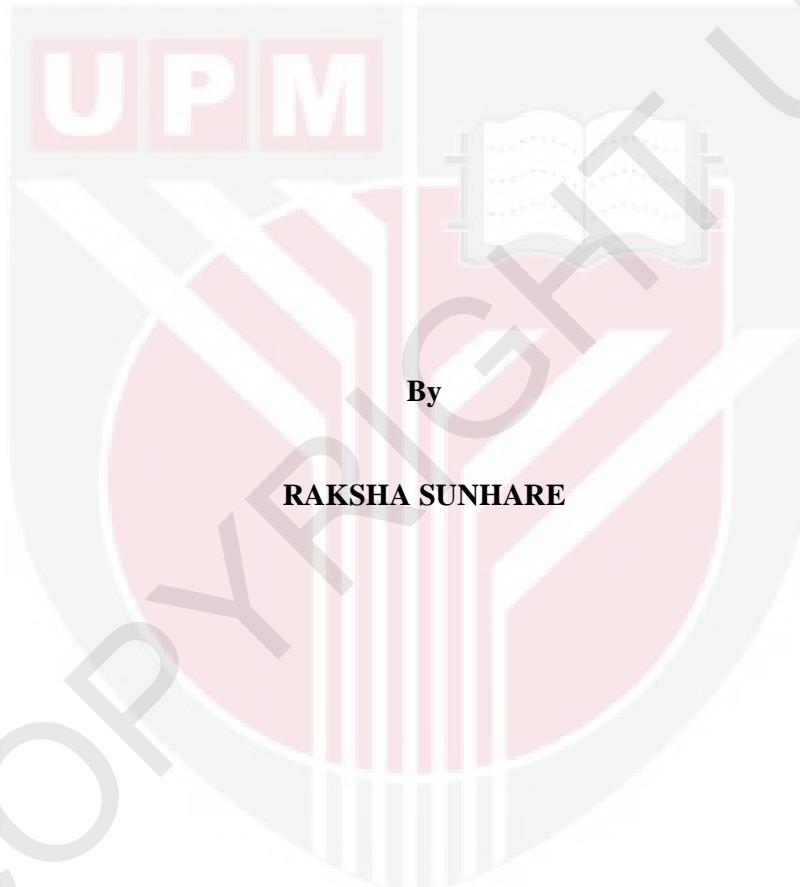


**DOCTOR OF PHILOSOPHY
UNIVERSITI PUTRA MALAYSIA**

2013



**PRODUCTION AND PURIFICATION OF NIPAH VIRUS GLYCOPROTEIN IN
Spodoptera frugiperda 9 (J.E. SMITH) INSECT CELL**



**Thesis Submitted to the School of Graduate Studies,
Universiti Putra Malaysia, in Fulfilment of the Requirements for the Degree of
Doctor of Philosophy**

October 2013

COPYRIGHT

All material contained within the thesis, including without limitation text, logos, icons, photographs and all other artwork, is copyright material of Universiti Putra Malaysia unless otherwise stated. Use may be made of any material contained within the thesis for non-commercial purposes from the copyright holder. Commercial use of material may only be made with the express, prior, written permission of Universiti Putra Malaysia.

Copyright © Universiti Putra Malaysia



Abstract of thesis presented to the Senate of Universiti Putra Malaysia in fulfilment
of the requirement for the degree of Doctor of Philosophy

**PRODUCTION AND PURIFICATION OF NIPAH VIRUS GLYCOPROTEIN IN
Spodoptera frugiperda 9 (J.E. SMITH) INSECT CELL**

By

RAKSHA SUNHARE

October 2013

Chairman: Associate Professor Tey Beng Ti, PhD

Institute: Institute of Bioscience

Nipah virus glycoprotein (NiVG) expressed in *Spodoptera frugiperda* 9 [Sf9 (J.E. Smith)] induces neutralizing antibodies and could be used as an early detection reagent against NiV infection to prevent. Several industrial problems during the development of recombinant truncated glycoprotein of NiV (tNiVG) of baculovirus expression vector system (BEVs) from Sf9 insect cells were investigated in this study that included production, cell disruption techniques and purification.

The production of recombinant tNiVG was enhanced by investigating the preferred medium and fermentation conditions of Sf9 cells. The maximum production of tNiVG was achieved by infecting Sf900 III serum free medium (SFM) developed middle exponential 5.0×10^6 cells/mL with quaternary amplified recombinant baculovirus at a multiplicity of infection (MOI) of 5. At the preferred condition, about 1.43 mg of tNiVG per 5.0×10^6 cells was obtained after 3 days of post-infection. The amount of tNiVG produced was equivalent to 40% of total cellular protein and thus maximum production was achieved from the insect cell-baculovirus

expression vector system. Subsequent replacement of fresh *Sf*900 III SFM (every 3 days) reduced the doubling time (22 h) and achieved the maximum density (15.7×10^6 cells/mL) of *Sf*9 cells after 4 days. Hence, fresh *Sf*900 III SFM was used to study the effect of amplification of recombinant baculovirus, MOI and time of infection (TOI) on tNiVG production.

The chemical lysis, freeze–thawing, high–pressure homogenisation and ultrasonication cell disruption methods were investigated to release the recombinant tNiVG from *Sf*9 cells. Among the investigated methods, high–pressure homogenisation with a single–pass successfully released 1.64 mg/mL tNiVG per 5.0×10^6 insect cells with a purity of 45% at high sample volume.

Comparative evaluations of three immobilised affinity chromatography methods HisTrapTM FF 1 mL prepacked column, Ni SepFastTM MAG 1 mL adsorbent and conventional method for the recovery of tNiVG from *Sf*9 cells homogenate were investigated. The adsorption efficiency of applied clarified and unclarified feedstock onto HisTrapTM FF 1 mL prepacked column and Ni SepFastTM 1 mL MAG adsorbent was performed at 20 sodium phosphate mM, 500 mM sodium chloride, 20 mM imidazole and 5% glycerol containing pH 8 buffer. A single–step elution of bound tNiVG was performed with 20 mM sodium phosphate containing 250 mM sodium chloride, pH 7 buffer in the presence of 200 and 300 mM imidazole from HisTrapTM prepacked column and SepFastTM MAG adsorbent, respectively. Both IMAC operations achieved 94% purity and 92% recovery yield of tNiVG. Additionally, the unclarified feedstock application onto IMAC shortened the total processing time by about 13–fold as compared to the conventional method.

Followed by a single-step purification strategy, SepFastTM Supor Q column of strong anion-exchange chromatography (AEC) was used to purify tNiVG produced in *Sf9* insect cells. The preferred conditions of buffer to bind and to elute tNiVG were 50 mM sodium carbonate, pH 9 and 50 mM sodium citrate, pH 5. The use of elution buffer without sodium chloride separated the loosely bound tNiVG from the tightly bound major host proteins and subsequently avoided the desalting step as one of the further downstream processes. The developed method has recovered 89% tNiVG from the original supernatant with a protein purity of 90%. SDS-PAGE, Western blot and ELISA conformed purity and immunogenicity of single-step salt-free tNiVG (57 kDa). Further, the results of mass spectrometry confirmed the identity of tNiVG.

Information obtained from these studies was useful for development of efficient production and single-step purification of recombinant tNiVG from *Sf9* cells. The overall recovery yield and purity throughout the studies proposed unit operations as simple, economic and fast method for the development of tNiVG. Along with these, the single-step purified tNiVG could be used as a potential agent for the development of an immunoassay for NiV antibodies.

Abstrak tesis yang dikemukakan kepada senat Universiti Putra Malaysia sebagai memenuhi keperluan untuk ijazah Doktor Falsafah

**PENGHASILAN DAN PENULENAN GLIKOPROTEIN VIRUS NIPAH
YANG TERDAPAT DALAM SEL SERANGGA *Spodoptera frugiperda 9*
(J.E.SMITH)**

Oleh

RAKSHA SUNHARE

October 2013

Pengerusi: Profesor Tey Beng Ti, PhD

Institut: Institut Biosains

Glikoprotein virus Nipah (NiVG) ungkapan dalam *Spodoptera frugiperda 9* [Sf9 (J.E. Smith)] mengaruhi antibodi peneutralan dan boleh digunakan sebagai pengesanan awal bagi melawan jangkitan NiV. Beberapa masalah industry semasa membangunkan glikoprotein rekombinan terpangkas NiV (tNiVG) daripada baculovirus ungkapan sistem vektor (BEVs) daripada sel-sel serangga Sf9 telah disiasat dalam kajian ini yang dimasukkan penghasilan, teknik-teknik gangguan sel dan penulenan.

Penghasilan protein rekombinan tNiVG telah dipertingkatkan dengan mengoptimalkan medium pilihan dan keadaan-keadaan penapaian sel-sel Sf9. Penghasilan maksimum protein tNiVG telah dicapai dengan menjangkitkan medium bebas serum (MBS) Sf900 II dibangunkan eksponen pertengahan 5.0×10^6 sel/mL dengan bakulovirus diperkuatkan kuarerner pada kegandaan jangkitan (MOI) 5. Pada keadaan pilihan, kira-kira 1.43 mg protein tNiVG per 5.0×10^6 sel telah diperolehi setiap 3 hari selepas pasca-jangkitan. Jumlah protein tNiVG yang telah dihasilkan adalah bersamaan 40% daripada jumlah protein selular dan dengan itu

penghasilan maksimum telah dicapai daripada ungkapan sistem vektor sel serangga–bakulovirus. Penggantian *Sf*900 III SFM segar berikutnya (setiap 3 hari) telah mengurangkan waktu penggandaan (22 j) dan telah mencapai ketumpatan maksimum (15.7×10^6 sel/mL) bagi sel–sel *Sf*9 selepas 4 hari. Oleh yang demikian, *Sf*900 III SFM segar telah digunakan untuk mengkaji kesan amplifikasi bakulovirus rekombinan, MOI dan waktu jangkitan terhadap penghasilan protein tNiVG. Lisis kimia, pencairan pembekuan, penghomogenan tekanan tinggi dan kaedah ultrasonikasi gangguan sel telah disiasat untuk membebaskan protein rekombinan tNiVG daripada sel–sel *Sf*9. Antara kaedah–kaedah yang telah dikaji adalah penghomogenan tekanan tinggi dengan dengan laluan tunggal telah berjaya membebaskan 1.64 mg/mL protein tNiVG per 5.0×10^6 sel–sel serangga dengan ketulenan 45% pada jumlah sampel yang tinggi.

Penilaian perbandingan terhadap tiga kaedah kromatografi keafinan pegun turus siap dibungkus HisTrapTM FF 1 mL, penjerap Ni SepFastTM MAG dan kaedah konvensional bagi pemulihan protein tNiVG dari homogenate sel–sel telah diselidik. Kecekapan pengikatan bagi stok suapan dijernihkan dan tidak dijernihkan yang digunakan ke atas turus siap dibungkus HisTrapTM FF 1 mL dan penjerap Ni SepFastTM MAG telah dijalankan pada 20 natrium foSfat mM, 500 mM natrium klorida, 20 mM imidazole dan 5 % gliserol yang mengandungi penimbal pH 8. Elusi langkah tunggal telah dijalankan ke atas protein tNiVG yang terikat dengan 20 mM natrium foSfat yang mengandungi 250 mM natrium klorida, penimbal pH 7 dengan kehadiran 200 dan 300 mM imidazola daripada turus siap dibungkus HisTrapTM dan penjerap SepFastTM MAG masing–masingnya. Kedua–dua operasi IMAC telah mencapai 94% ketulenan dan 92% hasil pemulihan protein tNiVG. Selain itu,

penggunaan stok suapan tidak dijernihkan ke atas IMAC telah memendekkan jumlah masa pemprosesan sehingga 13-kali ganda jika dibandingkan dengan kaedah konvensional.

Diikuti dengan satu langkah strategi penulenan, turus kromatografi pertukaran anion (AEC) SepFastTM Supor Q yang kuat telah digunakan untuk menulenkan tNiVG yang telah dihasilkan oleh sel-sel serangga *Sf9*. Keadaan penimbal yang dipilih untuk mengikat dan menanggalkan protein tNiVG adalah 50 mM natrium karbonat, pH 9 dan 50 mM natrium sitrat, pH 5. Penggunaan penimbal elusi tanpa natrium klorida mengasingkan protein tNiVG yang terikat secara longgar daripada protein hos utama yang terikat dengan kuat dan seterusnya mengelakkan langkah penyahgaraman sebagai salah satu proses hiliran berikutnya. Kaedah yang telah dibangunkan telah mendapatkan kembali 89% protein tNiVG daripada supernatant dengan ketulenan protein 90%. SDS-PAGE, ujian pemendapan Western dan ELISA mengesahkan ketulenan dan keimunogenan bagi langkah-tunggal protein tNiVG bebas-garam (57 kDa). Tambahan lagi, keputusan spektrometri jisim mengesahkan identiti protein tNiVG.

Maklumat yang diperoleh daripada kajian ini berguna untuk pembangunan pengeluaran yang cekap dan penulenan langkah tunggal bagi rekombinan protein tNiVG dari sel-sel *Sf9*. Hasil pemulihan dan ketulenan keseluruhan di sepanjang kajian mencadangkan bahawa operasi unit adalah mudah, ekonomi dan merupakan kaedah pantas bagi pembangunan protein tNiVG. Pada masa yang sama, penulenan protein tNiVG satu-langkah boleh digunakan sebagai agen yang berpotensi untuk pembangunan satu imuocerakinan bagi antibodi-antibodi NiV.

ACKNOWLEDGEMENTS

“Failures are part of life. If don’t fail, don’t learn. If don’t learn, we’ll never change”

(Source: Unknown Quotes)

Thanks to my parents for giving me the gift of life and the positive values, they taught me as a child.

My most sincere gratitude to my supervisors, Professor Dr. Tey Beng Ti, Professor Dr. Tan Wen Siang and Associate Professor Dr. Muhajir Hamid for the opportunity they gave me, their invaluable guidance, careful supervision and trust on my abilities throughout the study. Their motivational guidance made me understand the ways to apply technology effectively. My special thanks to Dr. Ramakrishnan Nagasundara Ramanan of Monash University for his generous help to accomplish this thesis.

I am heartily thankful to UPM, providing me such a great opportunity and a RUGS grant to carry out this project. I would like to acknowledge the Ministry of Higher Education, Malaysia for their financial assistance (MTCP–MOHE). I am especially thankful to all Lab Fellows for having a healthy relationship among scientists and research scholars, which lays the foundation of a united world.

Last but not least, I express my deep gratitude to my family for their patience, sacrifices and moral support without which I could not have been successful! I am sure someone will continue research with this work; my best-wishes will always be with that person.

Thanks to Everyone!

Raksha Sunhare
<http://www.studymalaysia.com/student/views.php?code=UPM>

This thesis was submitted to the Senate of Universiti Putra Malaysia and has been accepted as fulfilment of the requirement for the degree of Doctor of Philosophy. The members of the Supervisory Committee were as follows;

Tey Beng Ti, PhD

Associate Professor

Faculty of Engineering

Universiti Putra Malaysia

(Chairman)

Tan Wen Siang, PhD

Professor

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Member)

Muhajir Hamid, PhD

Associate Professor

Faculty of Biotechnology and Biomolecular Sciences

Universiti Putra Malaysia

(Member)

BUJANG BIN KIM HUAT, PhD

Professor and Dean

School of Graduate Studies

Universiti Putra Malaysia

Date:



TABLE OF CONTENTS

	Page
ABSTRACT	i
ABSTRAK	iv
ACKNOWLEDGEMENTS	vii
APPROVAL	viii
DECLARATION	x
LIST OF TABLES	xiv
LIST OF FIGURES	xv
LIST OF APPENDIX	xvii
LIST OF ABBREVIATIONS	xviii
CHAPTER	
1 INTRODUCTION	1
2 LITERATURE REVIEW	6
2.1 Nipah virus (NiV)	8
2.1.1 Nipah virus glycoprotein (NiVG)	9
2.1.2 Studies on the NiVG	10
2.2 Studies on recombinant protein expression and production	11
2.2.1 Glycoprotein expression technologies	12
2.2.2 Insect cell–Baculovirus expression vector system	15
2.2.3 Effects of factor affecting protein production	17
2.3 Studies on recombinant protein purification	18
2.3.1 Cell disruption method	20
2.3.2 Immobilised metal affinity chromatography (IMAC)	24
2.3.3 Ion–exchange chromatography (IEC)	29
2.4 Concluding remarks	33
3 GENERAL MATERIALS AND METHODS	35
3.1 Maintenance of cell lines	35
3.1.1 Cell thawing and freezing	35
3.1.2 Subculture and counting of cells	36
3.1.3 Adapting monolayer to suspension culture	37
3.1.4 Sequential development of serum free Sf9 cells	37
3.2 Generation of recombinant baculovirus	38
3.2.1 Competent DH10 Bac cell preparation	38
3.2.2 Transposition	38
3.2.3 Transfection	39
3.2.4 Recombinant virus propagation	40
3.3 Analytical methods	40
3.3.1 PCR analysis of bacmid DNA	40
3.3.2 Virus titration by plaque assay	41

3.3.3 Bradford Assay	42
3.3.4 Sodium dodecyl sulphate–polyacrylamide gel electrophoresis (SDS–PAGE)	42
3.3.5 Western blotting	43
3.3.6 Quantitative analysis of tNiVG	44
3.3.7 Enzyme–linked immunosorbent assay (ELISA)	44
3.4 Calculations	45
3.4.1 Calculations for inoculum volume	45
3.4.2 Subculture and growth related calculations	46
3.4.3 Calculations for adsorption analysis	47
3.4.4 Calculations for purification profile analysis	48
4 EFFECT OF MEDIUM AND INFECTION CONDITION OF <i>Spodoptera frugiperda</i> 9 (J.E. Smith) FOR THE PRODUCTION OF NIPAH VIRUS GLYCOPROTEIN	50
4.1 Introduction	50
4.2 Materials and methods	52
4.2.1 Effect of media over Sf9 cell densities	52
4.2.2 Identification and amplification of tNiVG	53
4.2.3 Effect of MOI on production of tNiVG	54
4.2.4 Effect of TOI on production of tNiVG	54
4.2.5 Protein qualitative and quantitation analysis	56
4.3 Results and discussion	56
4.3.1 Effect of media over Sf9 cell densities	56
4.3.2 Identification and amplification of tNiVG	58
4.3.3 Effect of MOI on production of tNiVG	59
4.3.4 Effect of TOI on production of tNiVG	61
4.4 Summary	63
5 COMPARISON OF THE PERFORMANCE OF TWO IMMOBILISED METAL AFFINITY CHROMATOGRAPHY SYSTEMS FOR THE PURIFICATION OF NIPAH VIRUS GLYCOPROTEIN	64
5.1 Introduction	64
5.2 Materials and methods	66
5.2.1 Materials	66
5.2.2 Feedstock preparation	67
5.2.3 Cell disruption	67
5.2.4 Investigation of binding and elution of tNiVG	68
5.2.5 Dynamic binding capacity and batch adsorption	69
5.2.6 Protein qualitative and quantitation	70
5.3 Results and discussion	71
5.3.1 Cell disruption	71
5.3.2 Protein adsorption condition of tNiVG	74

5.3.3	Elution condition of tNiVG	76
5.3.4	Determination of loading condition	79
5.3.5	Purification of tNiVG and analysis	81
5.4	Summary	89
6	A SINGLE STEP PURIFICATION OF GLYCOPROTEIN OF NIPAH VIRUS IN INSECT CELLS USING AN ANION-EXCHANGE CHROMATOGRAPHY METHOD	90
6.1	Introduction	90
6.2	Materials and methods	92
6.2.1	Materials	92
6.2.2	Feedstock preparation	92
6.2.3	Investigation of binding and elution of tNiVG	93
6.2.4	Determination of dynamic binding capacity and protein adsorption efficiency	94
6.2.5	Protein qualitative and quantitation analysis	94
6.2.6	Identification of tNiVG by Mass spectrometry	94
6.3	Results and discussion	95
6.3.1	Protein adsorption condition of tNiVG	95
6.3.2	Elution condition of tNiVG	96
6.3.3	Determination of loading condition	98
6.3.4	Purification of tNiVG and analysis	100
6.4	Summary	105
7	GENERAL DISCUSSION, CONCLUSION AND FUTURE PERSPECTIVES	106
7.1	General discussion	106
7.2	General conclusion	110
7.3	Future prospects	111
REFERENCES		113
APPENDIX		133
BIODATA OF STUDENT		139
LIST OF PUBLICATIONS		140